

Review **Molecular Mechanisms and Therapeutic Implications of Endothelial Dysfunction in Patients with Heart Failure**

Vasiliki Tsigkou ¹ , Evangelos Oikonomou 1,* [,](https://orcid.org/0000-0001-8079-0599) Artemis Anastasiou ¹ , Stamatios Lampsas ¹ [,](https://orcid.org/0000-0001-6481-0000) \bf{G} eorge E. Zakynthinos ¹[,](https://orcid.org/0000-0001-8289-4869) Konstantinos Kalogeras ¹D, Maria Katsioupa ¹D, Maria Kapsali ¹, Islam Kourampi ¹D, **Theodoros Pesiridis ¹ , Georgios Marinos ¹ , Michael-Andrew Vavuranakis ¹ [,](https://orcid.org/0000-0002-1385-1811) Dimitris Tousoulis ² , Manolis Vavuranakis ¹ and Gerasimos Siasos 1,3**

- ¹ 3rd Department of Cardiology, Medical School, National and Kapodistrian University of Athens, Sotiria Chest Disease Hospital, 11527 Athens, Greece
- ² 1st Department of Cardiology, Medical School, National and Kapodistrian University of Athens, Hippokration General Hospital, 11527 Athens, Greece
- ³ Cardiovascular Division, Brigham and Women's Hospital, Harvard Medical School, Boston, MA 02115, USA
- ***** Correspondence: boikono@gmail.com; Tel.: +30-69-4770-1299

Abstract: Heart failure is a complex medical syndrome that is attributed to a number of risk factors; nevertheless, its clinical presentation is quite similar among the different etiologies. Heart failure displays a rapidly increasing prevalence due to the aging of the population and the success of medical treatment and devices. The pathophysiology of heart failure comprises several mechanisms, such as activation of neurohormonal systems, oxidative stress, dysfunctional calcium handling, impaired energy utilization, mitochondrial dysfunction, and inflammation, which are also implicated in the development of endothelial dysfunction. Heart failure with reduced ejection fraction is usually the result of myocardial loss, which progressively ends in myocardial remodeling. On the other hand, heart failure with preserved ejection fraction is common in patients with comorbidities such as diabetes mellitus, obesity, and hypertension, which trigger the creation of a micro-environment of chronic, ongoing inflammation. Interestingly, endothelial dysfunction of both peripheral vessels and coronary epicardial vessels and microcirculation is a common characteristic of both categories of heart failure and has been associated with worse cardiovascular outcomes. Indeed, exercise training and several heart failure drug categories display favorable effects against endothelial dysfunction apart from their established direct myocardial benefit.

Keywords: heart failure; endothelial dysfunction; pathophysiology; molecular mechanisms

1. Introduction

Heart failure (HF) is a heterogenous clinical syndrome with a broad range of symptoms and signs, which are attributed to divergent underlying etiologies that induce structural or functional abnormality of the heart [\[1\]](#page-19-0). HF affects more than 60 million adults globally and is characterized by severe morbidity, mortality, and poor quality of life [\[1\]](#page-19-0). Despite the decreased age and sex-adjusted incidence for both heart failure with reduced ejection fraction (HFrEF) and heart failure with preserved ejection fraction (HFpEF), the prevalence remains high and is projected to increase worldwide due to the aging process, improved therapeutic options for ischemic heart disease, and the availability of effective evidencebased therapies [\[2\]](#page-19-1). Although the prognosis of HF has, in general, been improved, it should be emphasized that mortality remains high according to recent studies, irrespectively of left ventricular ejection fraction (LVEF) and type of HF (acute or chronic) [\[2\]](#page-19-1).

HFrEF is mainly attributed to the loss of cardiomyocytes due to ischemia, myocarditis, or genetic mutations, which trigger the mechanisms of cardiovascular remodeling [\[3\]](#page-19-2). Patients with HFrEF are predominantly men, usually post-myocardial infarction (MI), and develop more frequently a profile of eccentric hypertrophy of the left ventricle (LV) due to

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pressure overload [\[3\]](#page-19-2). On the other hand, HFpEF, which is now considered to be the most common category of HF, is associated with the presence of comorbidities, such as arterial hypertension, diabetes mellitus, renal dysfunction, obesity, and increased age [\[4\]](#page-19-3).

Endothelial dysfunction (ED) has an important role in the pathophysiology of HF since repeated episodes of microvascular dysfunction might precipitate myocardial stunning and ventricular remodeling and might be associated with HF hospitalizations [\[5\]](#page-19-4). Moreover, ED has been linked to the presence of a procoagulant state, the expression of adhesion molecules, and a pro-inflammatory status [\[6,](#page-19-5)[7\]](#page-19-6). Additionally, the pathophysiology of HFpEF is quite different from HFrEF according to the literature since it is largely attributed to the existence of coronary microvascular ED due to the presence of cardiovascular risk factors, which trigger a state of systemic inflammation [\[8\]](#page-19-7). However, HFpEF exhibits a profile of concentric hypertrophy [\[3\]](#page-19-2), and these differences in risk factors, as well as in the phenotypic presentation of different HF categories, may be caused by differences in molecellular mechanisms and expression [\[9\]](#page-19-8).

Therefore, in this article, we review the molecular mechanisms underlying the association of ED with the development and progression of HF as well as the therapeutic implications of ED improvement in patients with HF.

2. Etiopathogenesis and Molecular Mechanisms of Heart Failure

2.1. Neurohormonal System Activation

Activation of neurohormonal systems is a key component of HF [\[10\]](#page-19-9). The sympathetic nervous system (SNS) and the renin–angiotensin–aldosterone system (RAAS) are the most important neurohormonal systems in the pathophysiology of HF and an important target of several therapeutic regimens [\[11\]](#page-19-10). Persistent overstimulation of SNS has been linked to the development of cardiomyocyte hypertrophy, interstitial fibrosis, inflammation, and oxidative stress, which progressively result in loss of myocardial contractility and LVEF deterioration [\[12\]](#page-19-11). Moreover, there is an interrelation between the RAAS system and SNS, which exacerbates cardiovascular damage [\[13\]](#page-19-12). It should be highlighted that aldosterone is associated with the development of ED, inflammation, and production of reactive oxygen species (ROS), which further deteriorate cardiovascular function [\[14](#page-19-13)[,15\]](#page-19-14). All these effects are mainly driven by angiotensin (AT) receptors AT-1 since AT-2 receptors have displayed antifibrotic, anti-inflammatory, and anti-apoptotic actions due to activation of bradykinin and nitric oxide (NO) synthesis [\[16\]](#page-19-15).

2.2. Oxidative Stress

Oxidative stress results as an imbalance between ROS production and their reversal by the antioxidant systems of the body; normally, a small amount of ROS is formed during mitochondrial respiration, which is detoxified by cells' antioxidant enzymes [\[17\]](#page-19-16). Nevertheless, excessive ROS production at mitochondria and the presence of dysfunctional antioxidant systems are the key contributors to oxidative stress in HF [\[17\]](#page-19-16). ROS overproduction induces ED due to NOS uncoupling and concomitant superoxide anion and peroxynitrite release, which decreases nitric oxide (NO) availability and further causes vasoconstriction [\[18\]](#page-19-17). Ischemia or hypoxia further exacerbates ROS production by mitochondria which accelerates myocardial damage, both at the stages of ischemia–reperfusion injury as well as in chronic ischemic conditions [\[19\]](#page-19-18). ROS induces post-translational modification of cellular compartments, reversibly or irreversibly, in a process that ends in cardiac hypertrophy [\[20\]](#page-19-19). Indeed, protein kinase C and mitogen-activated protein kinases (MAPK) induce myocardial remodeling and hypertrophy [\[21\]](#page-19-20). Moreover, excessive ROS production might damage mitochondrial DNA, which has a low capacity for repair; therefore, a vicious cycle of ROS overproduction and subsequent myocardial damage occurs [\[22\]](#page-20-0). Indeed, oxidative stress upon neurohormonal activation deteriorates the proper mitochondrial function of cardiomyocytes; mitochondrial dysfunction causes further deficiency of mitochondrial energetics [\[23\]](#page-20-1). Contrarily, cardiovascular risk factors in HFpEF create an environment of oxidative stress, inflammation, and microvascular ED; nonetheless, less is

understood so far about the implication of mitochondrial dysfunction and cardiac energetics and should be further investigated [\[23–](#page-20-1)[25\]](#page-20-2). Importantly, oxidative stress and chronic, low-grade inflammation are important contributors to the poor regenerative properties of cardiomyocytes [\[26\]](#page-20-3).

2.3. Calcium Regulation

High cytosolic calcium input in HF is particularly attributed to the dysfunction of the L-type calcium channel (LTCC); indeed, increased phosphorylation of LTCC is evident in HF and results in a compensatory leak of sarcoplasmic reticulum (SR) calcium through ryanodine receptor 2 (RyR2), which is in the uncoupled form due to chronic SNS stimulation and oxidative stress [\[27,](#page-20-4)[28\]](#page-20-5). On the other hand, dysfunction of the sodium–calcium exchanger (NCX), which controls cytosolic calcium outflux due to the diminished transmembrane sodium gradient, results in sarcolemmal depolarization [\[29–](#page-20-6)[31\]](#page-20-7). It should be mentioned that myocardial hypertrophy per se is associated with prolonged duration of isometric contraction and relaxation and molecular changes due to fibrosis and dysregulated creatine kinase system [\[32\]](#page-20-8). Furthermore, the transverse tubular system and proteins of the excitation–contraction coupling system display dysfunction in the aged cardiomyocytes, especially in the HF condition [\[33\]](#page-20-9). On the other hand, improper function and diminuted expression of cardiac SR calcium ATPase (SERCA2 a) end in reduced calcium transfer to SR [\[34\]](#page-20-10). Additionally, dephosphorylazation of phospholamban from protein phosphatase-1 has also been identified in HF [\[35\]](#page-20-11). SERCAa is controlled by hormones, microRNAs (miR), and endogenous protein inhibitors and undergoes post-translational modifications [\[36\]](#page-20-12). Lastly, recent data indicate that inflammation and, specifically, damageassociated molecular patterns (DAMPs, i.e., destroyed or stressed cardiomyocytes) release mediators that trigger the immune response; as a result, dysfunctional cardiac contraction and electromechanical uncoupling occur due to the unfavorable calcium homeostasis [\[36\]](#page-20-12).

2.4. Impaired Metabolism and Mitochondrial Energetics

Myocardial contraction and relaxation depend on the proper energy production by cardiomyocytes, which is orchestrated by cardiac mitochondria during oxidative metabolism in the mitochondrial matrix in a process known as mechano-energetic coupling [\[37\]](#page-20-13). Normally, cardiac mitochondria, which are the main source of energy, ROS production, and calcium ion control, utilize free fatty acids in order to produce energy in the form of adenosine triphosphate (ATP) [\[37\]](#page-20-13). As a matter of fact, utilization of free fatty acids results in more efficient energy production than utilization of glucose in terms of ATP production at the expense of more oxygen consumption, though [\[38\]](#page-20-14). According to the literature, HF is characterized by dysfunctional energy metabolism [\[37\]](#page-20-13). HF is perceived to be a state of 'energy deprivation' due to mitochondrial dysfunction, which is attributed to the altered mitochondrial structure and function [\[39\]](#page-20-15). Indeed, evidence from animal studies of HF has revealed that mitochondria display hyperplasia, decreased size or fragmentation, and disruption of their inner and outer membranes [\[39\]](#page-20-15). Several mitochondrial proteins in HF undergo post-translational modification (such as phosphorylation, acetylation, ubiquitination, conjugation of small ubiquitin-like modifier proteins, O-linked-N-acetyl-glucosamine glycosylation, proteolysis), which results in fission and fusion of mitochondria [\[40\]](#page-20-16). Furthermore, HF is associated with impaired mitochondrial biogenesis, dysfunctional mitochondrial DNA replication as well as mitochondrial DNA depletion [\[41\]](#page-20-17). Lastly, mitochondria-related genes (such as *IFIT3, XAF1, RSAD2,* and *MX1*) have displayed associations with HF and the biological processes of oxidative stress, amino-acid metabolism, and aging [\[42\]](#page-20-18).

2.5. Inflammation

Inflammation and HF display a strong interrelationship; as a matter of fact, inflammation is associated with the development of molecular, cellular, and functional changes in the heart [\[43](#page-20-19)[–45\]](#page-20-20). Inflammation in HFrEF is most commonly the result of cardiomyocyte injury or loss; on the other hand, cardiovascular risk factors in HFpEF create an environment of

chronic, low-grade systemic, and local ongoing myocardial inflammation, which trigger myocardial damage [\[46\]](#page-20-21). Moreover, activation of neurohormonal mechanisms in HF, such as the RAAS system, strengthens the inflammatory response and induces immune system activation [\[47\]](#page-20-22). Additionally, the presence of a pro-inflammatory milieu in HF has been associated with the tendency for thromboses [\[48\]](#page-20-23). Furthermore, acute HF with peripheral hypoperfusion and systemic congestion favor damage to endothelial glycocalyx and cause ED [\[49\]](#page-21-0). Interestingly, inflammation has been linked to worse cardiovascular prognosis in patients with HF, cardiac cachexia due to muscle wasting, higher vascular resistance, and decreased functional capacity [\[50\]](#page-21-1).

Indeed, patients with HFpEF display higher levels of inflammatory markers such as C-reactive protein (CRP), interleukin-6 (IL-6), and tumor necrosis factor-alpha (TNF-α); furthermore, CRP levels are proportional to the number of associated comorbidities [\[51\]](#page-21-2). Other circulating biomarkers, such as miRs, have been involved in the pathophysiology of inflammation in HF; specifically, miR-21 is highly expressed during the course of chronic HF [\[52\]](#page-21-3). Indeed, miR-21 has been implicated in the development of ischemia/reperfusion injury and LV remodeling [\[53\]](#page-21-4). As a matter of fact, several miRs are implicated in the pathophysiology of coronary artery disease, acute coronary syndromes, post-MI myocardial remodeling, and fibrosis, and they could serve as possible biomarkers for diagnosis, prognosis, and treatment [\[53\]](#page-21-4). Most importantly, some mechanisms of their action involve detrimental effects on endothelial function, inflammation, and oxidative stress [\[53\]](#page-21-4). Last but not least, other non-coding RNAs have been studied intensively as possible pathophysiologic mediators not only for HF but also for several cardiovascular diseases [\[45\]](#page-20-20).

On the other hand, in circumstances of myocardial damage, DAMPs activate the pathogen recognition receptor (RPR) pathway, which is responsible for the release of intracellular cytokines such as the receptor for advanced glycation end-products (RAGE) and cluster of differentiation-36 (CD-36) [\[54\]](#page-21-5). Dying cells release pro-oxidant mediators, ROS, IL-1β, myeloperoxidase (MPO), and matrix metalloproteinases (MMPs), which break down extracellular matrix (ECM) [\[54–](#page-21-5)[56\]](#page-21-6). ROS provoke irreversible damage to cellular compartments and death of cardiomyocytes; also, oxidative stress is highly linked to the inflammatory response [\[57\]](#page-21-7). B-and T-cells participate in the inflammatory processes and cardiac remodeling; although T-lymphocytes have a protective role in cardiac remodeling post-MI, the balance between destroyed cardiomyocytes and the adaptive immune response is insufficient to prevent cardiac damage in HF [\[58–](#page-21-8)[60\]](#page-21-9).

Infections (i.e., viral infections) might result in the development of myocarditis and HF due to non-sterile inflammatory damage; as a matter of fact, in the acute phase, there is a loss of cardiomyocytes and myocardial infiltration with mononuclear cells [\[61\]](#page-21-10). Afterward, innate immune cells such as natural killer (NK) cells and macrophages hinder viral propagation until the adaptive immunity response begins; finally, antigen-specific T cells are activated, and in the subacute phase, the specific CD8+ cytotoxic T cells destroy the affected cells [\[61\]](#page-21-10). Last but not least, in the chronic phase, low-grade inflammation (due to the presence of infection or an inflammatory disease in general), which might last for years, triggers cardiac remodeling and augments the risk for dilated cardiomyopathy [\[62](#page-21-11)[–64\]](#page-21-12).

3. The Endothelium in Heart Failure

3.1. Endothelial Function in Patients with Heart Failure

The endothelium is a monolayer of cells that cover the inner surface of the vascular wall; apart from an anatomic barrier that separates blood flow from the vascular wall, endothelium serves as an important endocrine organ, which controls vascular tone, inflammation, and oxidative stress, cellular proliferation and coagulation [\[65\]](#page-21-13). Healthy endothelium produces NO from the enzyme endothelial nitric oxide synthase (eNOS); production of NO by eNOS is one of the most important mediators of endothelial-dependent relaxation as a response to mechanical stimuli (i.e., shear stress) or chemical substances (i.e., acetylcholine, arachidonic acid) [\[66\]](#page-21-14). Interestingly, eNOS is calcium-calmodulin dependent, and its activation depends on the intracellular concentration of calcium [\[66\]](#page-21-14). Sufficient

NO production is responsible for anti-inflammatory, antioxidant, anti-coagulant, and vasodilative effects [\[67\]](#page-21-15). Moreover, normal endothelial function affects almost every vascular bed, such as myocardial and coronary circulation and renal, systemic, and pulmonary circulation [\[68\]](#page-21-16).

Reduced NO production due to ED might contribute to hemodynamic compromise in the settings of acute HF [\[69\]](#page-21-17). According to the literature, decreased production of NO by eNOS has been linked to augmented vascular tone, cellular proliferation, and myocardial remodeling [\[69\]](#page-21-17). Furthermore, it has not been determined if venous congestion and fluid accumulation are the initial trigger or the result of ED in patients with acute HF [\[70\]](#page-21-18). Moreover, ED is also implicated in the pathophysiology of cardiorenal syndrome, which is characterized by the bidirectional dysfunction of the heart and kidney due to acute or chronic dysfunction of either organ or due to another systemic disease [\[48\]](#page-20-23).

On the other hand, acute endotheliitis might present in acute HF and has been associated with ED and impaired NO production due to excessive oxidative stress, inflammation, and vasospasm upon an initial myocardial insult [\[68\]](#page-21-16). As a matter of fact, acute endothelial injury is a key characteristic of Takotsubo cardiomyopathy and is related to the presence of an excessive inflammatory response, oxidative stress, and SNS activation [\[71\]](#page-21-19). Experimental data in hypertensive rats demonstrate that apart from the pressure-dependent uncoupling of eNOS during pulmonary edema, disrupted endothelial cell integrity and impaired endothelial mechanotransduction favor eNOS uncoupling and excessive ROS production during episodes of acute HF [\[72\]](#page-21-20).

Chronic HF is characterized by disturbance of normal endothelial function, which is associated with poor cardiovascular prognosis irrespectively of the cause or severity of HF [\[69,](#page-21-17)[73,](#page-21-21)[74\]](#page-22-0). Patients with chronic HF display vasoconstriction and deteriorated peripheral tissue perfusion due to ED; as a result, myocardial injury occurs, whereas poor tissue perfusion exacerbates the pre-existing vasoconstriction of both renal and coronary vascular beds [\[69\]](#page-21-17). Excessive oxidative stress in chronic HF is attributed to the dysfunctional antioxidant cellular defense systems due to eNOS uncoupling and impaired NO production [\[69\]](#page-21-17). Additionally, oxidative stress has been implicated in the dysfunctional regulation of calcium ions during systole and impaired cardiac relaxation [\[75\]](#page-22-1). Furthermore, oxidative modification and damage of cellular phospholipids result in high endothelial permeability and loss of endothelial integrity [\[76\]](#page-22-2). Decreased NO/ endothelin-1 (ET-1) ratio in chronic HF is linked to the continuous impairment of cardiac function, which is reflected by echocardiographic indices such as LVEF and LV short-axis shortening rate [\[77\]](#page-22-3). Lastly, increased expression of serum soluble angiotensin converting enzyme-2 (ACE) is related to poor exercise tolerance and raised asymmetric dimethyl-arginine (ADMA) levels, implying its association with the development of oxidative stress-induced ED [\[78\]](#page-22-4).

Regarding myocardial fibrosis, the transition of endothelial cells into fibroblast-like cells in chronic HF, which is termed endothelial-to-mesenchymal transition, has been intensively investigated recently [\[79\]](#page-22-5). There is evidence that epigenetic control of gene transcription and translation, including DNA methylation, histone modifications, and the actions of non-coding RNAs (miR, long non-coding RNAs, and circular RNAs), are implicated in these cellular processes in chronic HF [\[79\]](#page-22-5). As for miR, their role in the mechanisms of vascular repair has already been established for HF [\[77\]](#page-22-3). Moreover, RhoA/Rho kinase overexpression is another pathway of myocardial fibrosis and LV remodeling in chronic HF, associated with impaired NO bioavailability [\[80\]](#page-22-6). Lastly, according to a recent study in patients with chronic HF, apoptotic endothelial cell-derived micro-vesicles such as cluster of differentiation 31 (CD31)+/annexin V+ might discriminate different HF categories along with the measurement of classic biomarkers of fibrosis such as galectin-3 [\[81\]](#page-22-7).

On the other hand, another important pathophysiologic aspect of chronic HF is the dysfunction of endothelial circulating progenitor cells (EPCs); as a matter of fact, the pathway of AMP-activated protein kinase is involved in the regulation of EPCs number and expression and could be a possible therapeutic target for HF [\[73,](#page-21-21)[82\]](#page-22-8). Additionally, deteriorated NO production in chronic HF exhibits detrimental effects in the expression

of Vascular endothelial growth factor (VEGF), which normally mediates angiogenesis; as a result, diminuted capillary density occurs, which along with the dysfunctional energy metabolism of cardiomyocytes progress to the development of cardiomyopathy [\[83,](#page-22-9)[84\]](#page-22-10). Moreover, high expression of von Willebrand factor (vWF), which is a biomarker of endothelial damage, results in worse endothelial function reflected by deteriorated flow-mediated dilatation (FMD) [\[85](#page-22-11)[,86\]](#page-22-12).

3.2. Molecular Mechanisms of Endothelial Dysfunction in Patients with Heart Failure 3.2.1. Heart Failure with Reduced Ejection Fraction

Coronary microvascular ED has been linked to asymptomatic LV dysfunction and could be an early step in the pathophysiology of systolic HF [\[87\]](#page-22-13). On the other hand, peripheral ED is a risk factor for the development of stage B HF, which is defined as the asymptomatic stage of HF with impaired systolic function [\[88\]](#page-22-14). Interestingly, even low-risk individuals of both sexes but with ED have a greater risk for the development of stage B HF, although the exact pathophysiologic mechanisms of cardiac remodeling have not been determined [\[88\]](#page-22-14). Possibly, the diminuted NO production could result in LV systolic dysfunction and the progression of HF syndrome [\[88\]](#page-22-14). The association between endothelial function and HF is thought to be more complex than the already known NO-mediated effects in the cardiovascular system; as a matter of fact, ED is a key characteristic of several circulatory beds in HF, irrespectively of LVEF [\[89\]](#page-22-15). Moreover, biomarkers of endothelial glycocalyx degradation, such as elevated heparin sulfate, have also been linked to HFrEF and worse prognosis [\[90,](#page-22-16)[91\]](#page-22-17). What is more, ED occurs at a late stage in patients with HFrEF in contrast to HFpEF [\[4\]](#page-19-3). Lastly, according to the Multi-Ethnic Study of Atherosclerosis, impaired FMD of the brachial artery has been related to the risk of developing HF, and especially HFrEF, independently of the classic risk factors and natriuretic peptides levels [\[92\]](#page-22-18).

Interestingly, polymorphisms of eNOS might be implicated in the risk of systolic HF in certain populations, whereas ethnic differences between microvascular and macrovascular ED have also been recorded [\[93](#page-22-19)[–95\]](#page-22-20). Moreover, depletion of inducible nitric oxide synthase (iNOS) in an experimental study of wild-type mice with chronic transverse aortic constriction beneficially affect cardiac hypertrophy, dilation, fibrosis, and dysfunction, implying the detrimental effects of iNOS dysregulation in the maladaptive response to systolic overload of the heart [\[96\]](#page-23-0).

On the other hand, oxidative stress plays an important role in the pathophysiology of ED in both systolic and diastolic HF; while low concentrations of ROS are normal during cellular function, ROS overexpression damages cellular gene expression and signaling pathways, which affects cardiac mechanics and energy utilization [\[97\]](#page-23-1). Additionally, ROS overproduction is implicated in the development of myocardial hypertrophy and dilation [\[98\]](#page-23-2). HFrEF is characterized by dysfunction of the NO-sCG-cGMP pathway, which implies the presence of ED; oxidative stress induces deleterious effects in the enzymes of this system, which end in deteriorated NO production and development of myocardial injury according to preclinical and clinical data [\[99,](#page-23-3)[100\]](#page-23-4). As a matter of fact, treatment with agents with established antioxidant properties, such as allopurinol, has displayed beneficial effects in markers of systolic dysfunction of patients with HF, such as global longitudinal peak strain [\[98\]](#page-23-2). Moreover, activation of the nuclear factor kappa B (NFκB) pathway and release of pro-inflammatory molecules such as intracellular adhesion molecule-1 (ICAM-1) and vascular adhesion molecule-1 (VCAM-1) is evident in systolic HF according to studies [\[101](#page-23-5)[,102\]](#page-23-6). Additionally, other data indicate that decreased adiponectin levels are related to diminuted NO production in patients with systolic HF and the severity of HF [\[103\]](#page-23-7).

Patients with chronic systolic HF display not only ED but also abnormal ventriculararterial uncoupling, which is the pressure–volume interaction between LV and the vascular system [\[104\]](#page-23-8). Ventricular–arterial uncoupling has been associated with dysfunction of the mechanisms of cardiac energetics, pump efficiency, and poorer clinical outcomes [\[105\]](#page-23-9). Lastly, mineralocorticoid receptors of the endothelial cells have been related to the transition of cardiac hypertrophy to systolic dysfunction of the heart independently of pressureinduced overload damage and LV remodeling [\[106\]](#page-23-10).

Finally, patients with systolic HF and pulmonary arterial hypertension display low numbers of EPCs and increased levels of osteoprotegerin, implying the deleterious effects of osteoprotegerin in the development of pulmonary ED and worsening the prognosis of systolic HF [\[107\]](#page-23-11). Similarly, according to another study, high levels of osteoprotegerin, diminuted EPCs, and increased mean pulmonary artery pressure have been associated with the damage induced by hypoxemia in patients with sleep-disordered breathing; as a result, it is speculated that ED and vascular remodeling of pulmonary vasculature orchestrate the deterioration of systolic function in HF [\[108\]](#page-23-12).

3.2.2. Heart Failure with Preserved Ejection Fraction

ED induces LV diastolic dysfunction, which is a key characteristic of both HF (and, in particular, HFpEF) and coronary artery disease [\[109](#page-23-13)[,110\]](#page-23-14). Decreased microvascular reactivity in patients with diastolic HF implies that microvascular ED is responsible for the progression of subclinical heart remodeling [\[111\]](#page-23-15). Furthermore, ED is an important contributor to the development of LV diastolic and right ventricular dysfunction in patients with end-stage renal disease, whose volume status is normal [\[112\]](#page-23-16). Interestingly, a study in mice revealed that the induction of endothelial permeability was associated with the provocation of diastolic dysfunction and deterioration of cardiac function due to the disruption of endothelial cell–cardiomyocyte interactions and decreased ECM protein synthesis [\[113\]](#page-23-17). Moreover, overexpression of the human β3 adrenergic receptor in a transgenic rat model was linked to diminuted NOS3 mRNA expression, diastolic dysfunction of the aging heart, and reduced aortic flow upon diastolic stress [\[114\]](#page-23-18). Additionally, deterioration of the NOcGMP pathway is implicated in the pathophysiology of diastolic HF since a dysfunctional endothelium is related to repeated episodes of ischemia/reperfusion and the development of a chronically stunned myocardium with systolic dysfunction and increased diastolic stiffness [\[68\]](#page-21-16). Nonetheless, a study in knockout mice for nuclear factor (erythroid-derived 2)-like 2 demonstrated that the development of LV diastolic dysfunction is attributed to SERCA2 a downregulation and not to the changes in coronary vascular function or systemic hemodynamics, which were preserved by compensatory upregulation of eNOS expression in the aorta and the heart [\[115\]](#page-23-19).

Another important pathophysiologic aspect to be mentioned is the impact of diabetic metabolic derangement on LV function; interestingly, according to an experimental study, the effects of metabolic syndrome (hyperglycemia, hypercholesterolemia, and hypertriglyceridemia) were related to eNOS uncoupling, excessive nitroso-redox balance, alteration in genes of glucose and fatty acid metabolism as well as mitochondrial dysfunction [\[116\]](#page-23-20). Similarly, in another study of prediabetes, poor coronary endothelial function was linked to increased protein kinase C activity, mitochondrial oxidative stress, as well as rho-kinaseimpairment of myosin head extension to actin filaments [\[117\]](#page-24-0). Additionally, a decrease in soluble guanylyl cyclase/PKG activity and stiffness of myocardial titin was evident, too [\[117\]](#page-24-0). Interestingly, epicardial adipose tissue demonstrates a positive association with cardiac structural and protein alterations, ED, reduced insulin sensitivity, and inflammation; possibly, local mechanic or paracrinic effects of epicardial fat could justify these results [\[118\]](#page-24-1). Furthermore, impaired expression of endothelial sirtuin-6 in diabetes mellitus has been linked to the dysregulated fatty acid transportation across the endothelium, which might contribute to the pathophysiology of HFpEF [\[51\]](#page-21-2). Finally, according to recent data, miR-30 d/e has been associated with the presence of diastolic dysfunction, impaired free fatty acid metabolism, and microvascular dysfunction in diabetes [\[119\]](#page-24-2).

As for the role of inflammation, it should be mentioned that several risk factors induce a chronic, pro-inflammatory environment, which is responsible for the stimulation of the immune response, the perpetuation of hypoxemia, and the activation of neurohormonal systems; as a result, coronary microvascular ED develops and, subsequently, diastolic dysfunction of the heart [\[112](#page-23-16)[,120\]](#page-24-3). As a matter of fact, circulating pro-inflammatory biomarkers such as IL-6 and CRP have displayed an association with echocardiographic parameters of diastolic dysfunction in HF [\[121\]](#page-24-4). Interestingly, according to a recent study in female mice, a deficiency of endothelial sirtuin-3 is responsible for the induction of diastolic dysfunction in the aged heart as well as for the elevation of blood pressure [\[122\]](#page-24-5).

Evidence from basic science and clinical studies has revealed that systemic and myocardial oxidative stress derived by the enzyme nicotinamide adenine dinucleotide phosphate (NADPH) oxidase is an important contributor to LV diastolic dysfunction [\[123\]](#page-24-6). Moreover, according to an experimental study of pulmonary hypertension, diastolic HF has been associated with the lack of normal endothelial function as well as the development of increased pulmonary vascular resistance, vascular thickness, and biventricular cardiac hypertrophy [\[124\]](#page-24-7). Finally, according to other data, angiotensin II has been linked to the dysfunctional mechanisms of angiogenesis through effects in the Akt pathway, which ultimately result in the development of diastolic HF [\[125\]](#page-24-8).

3.2.3. Heart Failure of Ischemic Etiology

Acute myocardial ischemia (i.e., post MI) may result in scar formation, triggering cardiac remodeling and the development of HF [\[126\]](#page-24-9). Ischemic and non-ischemic HF display differences regarding their relation to ED [\[127\]](#page-24-10). Specifically, patients with ischemic HF usually have systemic ED, which involves arteries and veins, microcirculation, as well as coronary, pulmonary, and peripheral vessels [\[128\]](#page-24-11). Interestingly, according to a study, peripheral endothelium-dependent and endothelium-independent function was more deteriorated in patients with ischemic HF than in patients with non-ischemic HF [\[129\]](#page-24-12). Furthermore, a study by our research team has revealed that patients with ischemic HF have impaired FMD, and there is a linear improvement of FMD according to LVEF, while impaired endothelial function was associated with a worse cardiovascular prognosis [\[130\]](#page-24-13).

Regarding the pathophysiology, according to a protein network analysis from patients with ischemic and non-ischemic HF, upregulation of 18 proteins related to the pathways of inflammation, ED due to superoxide production, coagulation, and atherosclerosis was evident in ischemic HF [\[126\]](#page-24-9). Surprisingly, five key network proteins such as acid phosphatase 5, epidermal growth factor receptor, insulin-like growth factor binding protein-1, plasminogen activator urokinase receptor, and secreted phosphoprotein-1 could discriminate ischemic HF from non-ischemic HF [\[126\]](#page-24-9). The involvement of inflammation in the development of post-MI HF has already been established; according to an experimental study in mice, angiotensin II exerts myocardial damage post-MI through the attachment of the pro-inflammatory Nox2+ myelomonocytic cells, macrophages, and monocytes at the vessel wall along with stimulation of oxidative stress and the ED [\[131\]](#page-24-14). Other important pro-inflammatory mediators in the pathophysiology of ischemic HF are CRP, pentraxin-3, osteoprotegerin, BNP, neopterin, and soluble suppression of tumorigenesis-2 (sST2) [\[132\]](#page-24-15). As a matter of fact, sST2, which is a biomarker of fibrosis, has exhibited higher expression in patients with ischemic HF and has been linked to the functional capacity of the patients; moreover, this biomarker is inversely associated with FMD of the brachial artery [\[133\]](#page-24-16). Finally, according to another study, endothelial function in patients with ischemic HF is further impaired compared to patients with dilated cardiomyopathy, implying the involvement of underlying atherosclerosis in the pathophysiology; indeed, patients with ischemic HF displayed higher levels of IL-6 and TNF- α [\[134\]](#page-24-17).

As for oxidative stress, increased ADMA levels, which antagonize NO production and are a key characteristic of ED, were associated with poor cardiovascular prognosis in patients with ischemic HF [\[135\]](#page-24-18). Moreover, increased thrombogenicity, ED, and oxidative stress have been implicated in the development of atrial fibrillation in ischemic HF; specifically, evidence from a study in mice exhibited decreased expression of atrial eNOS, SERCAa, thrombomodulin, tissue factor pathway inhibitor, and tissue plasminogen activator [\[135\]](#page-24-18). Additionally, at the molecular level, exosomes (which are vectors for intracellular communication) are associated with ischemic heart disease and its evolution to HF

through the mechanisms of ED, lipid accumulation, atherosclerotic plaque development, and ischemia–reperfusion injury [\[136,](#page-24-19)[137\]](#page-24-20).

Post-MI remodeling is a detrimental consequence of MI; interestingly, in a study of patients with ischemic HF, increased mRNA levels of adrenomedullin exhibited a relation with post-ischemic myocardial remodeling [\[138\]](#page-24-21). Similarly, dysfunction of the T-regulatory cells has also been implicated in the development of chronic ischemic HF through immune system activation and LV remodeling [\[139\]](#page-25-0). What is more, mineralocorticoid receptors and RAAS activation are important determinants of post-MI LV dysfunction; specifically, according to a study in mice, deletion of mineralocorticoid receptors in Vascular smooth muscle cells (VSMCs) could ameliorate LV dysfunction post-MI through control of coronary flow reserve and improvement of endothelial function [\[140\]](#page-25-1).

As for EPCs, increased circulating levels of EPCs along with FMD could predict LV remodeling post-MI and the occurrence of major adverse cardiovascular events [\[141\]](#page-25-2). Interestingly, insulin resistance in diabetic patients with ischemic HF has been linked to decreased circulating numbers of proangiogenic EPCs [\[141\]](#page-25-2). Additionally, the reduced amount of CD14(+)CD309(+)-and CD14(+)CD309(+)Tie2(+) circulating EPC was related to the severity of LV dysfunction in patients with ischemic HF, whereas CD45(+) CD34(+) and CD45(-) CD34(+) mononuclear cell counts were associated with the severity of coronary artery lesion [\[142\]](#page-25-3). Last but not least, increased endothelial-derived apoptotic microparticles in patients with ischemic HF are associated with ED and poor prognosis [\[143\]](#page-25-4).

3.2.4. Non-Ischemic Heart Failure

Non-ischemic HF is a primary disease of cardiomyocytes and interstitial space [\[144\]](#page-25-5). The use of multiple circulating biomarkers might be used to reveal possible pathophysio-logic pathways that are linked to a certain phenotype of HF (ischemic vs. non-ischemic) [\[126\]](#page-24-9). As for ED, there is evidence that peripheral endothelial function is not impaired in patients with non-ischemic HF [\[145\]](#page-25-6). Indeed, the pattern of ED in patients with non-ischemic HF is more heterogenous and exhibits fewer systemic abnormalities, whereas ED of coronary circulation occurs more frequently [\[128\]](#page-24-11). Abnormal coronary microvascular flow has been associated with deteriorated myocardial perfusion and consequent metabolic changes in cardiomyocytes, which trigger local myocardial ischemia [\[146](#page-25-7)[,147\]](#page-25-8). Other studies have also confirmed the heterogenous nature of microvascular ED in patients with LV dysfunction of unknown cause [\[146\]](#page-25-7). Coronary endothelial-independent microvascular dysfunction has been related to higher brain natriuretic peptide (BNP) levels and ventricular wall tension in patients with non-ischemic HF, especially in those with cardiac fibrosis [\[148\]](#page-25-9).

As for the pathophysiology, diabetes mellitus is associated with the development of diabetic cardiomyopathy irrespectively of the ischemic damage through complex effects on vascular endothelial function; specifically, hyperglycemia increased free fatty oxidation, decreased NO production, oxidative stress, inflammation, and dysfunctional endothelial permeability contribute to the pathophysiology of diabetic cardiomyopathy [\[149\]](#page-25-10). Additionally, according to the literature, patients with non-ischemic HF display increased myocardial expression of vWF (which is a glycoprotein produced by endothelial cells that control platelet aggregation and thrombus formation at the sites of vascular injury), implying the impact of ED-derived vWF release in the development of non-ischemic HF [\[150\]](#page-25-11).

On the other hand, in patients with non-ischemic HF, NO and a secondary endotheliumderived relaxing factor sensitive to high K+ have demonstrated vasodilative properties [\[151\]](#page-25-12). In general, ED impairs LV systolic function due to the increase in systemic vascular resistance; then, LV dysfunction further deteriorates endothelial function through decrease in shear stress and NO bioavailability [\[152\]](#page-25-13). Interestingly, in a study of patients with systolic, non-ischemic HF, LVEF was related to ED, implying the importance of the management of ED in these patients [\[152\]](#page-25-13). Lastly, NO inhibition of the peripheral vasculature in patients with non-ischemic HF resulted in higher basal vascular tone and the progression of the disease [\[153\]](#page-25-14).

Therefore, the involvement of ED in the pathophysiology of non-ischemic HF is not consistent among the studies suggesting that further research should be performed in order to elucidate the exact impact of ED on non-ischemic HF.

3.2.5. Right Heart Failure Complicating Pulmonary Arterial Hypertension

In pulmonary arterial hypertension (PAH), the increase in pulmonary vascular resistance is responsible for the rise in right ventricular afterload and the progression to right HF [\[154\]](#page-25-15). The abnormal hypertrophy of small pulmonary arteries ranges from hypertrophy and hyperplasia of the media layer to the excessive apoptosis and proliferation of pulmonary arterial smooth muscle cells, which end in the formation of plexogenic lesions that obstruct artery lumen and decrease pulmonary blood flow [\[155\]](#page-25-16). Importantly the progressive occlusive arterial remodeling of pulmonary arterioles is characterized by the presence of significant ED [\[156,](#page-25-17)[157\]](#page-25-18).

Interestingly, it has been proposed that ED of pulmonary circulation displays an association with ED of the systemic circulation, implying that PAH is a situation of global vasculopathy [\[158\]](#page-25-19). Pulmonary artery endothelial cells are vulnerable to several insults (such as toxins, hypoxia, pro-inflammatory cytokines, and shear stress), which, along with the presence of genetic susceptibility, are responsible for the pathophysiology of disease; increased shear stress, in particular, develops an environment of raised arterial pressure and fluid dynamics which end in endothelial cell injury [\[159](#page-25-20)[,160\]](#page-25-21). As a result of pulmonary artery endothelial cell damage, excessive oxidative stress, hyperproliferation, and coagulation occur [\[159\]](#page-25-20). Moreover, at the later stages of endothelial cell damage, apoptosis-resistant endothelial cells develop along with excessive angiogenesis [\[161,](#page-26-0)[162\]](#page-26-1). Interestingly, research has revealed that pulmonary microvascular endothelial cells have intrinsic deficits that hinder proper response to Vascular endothelial growth factor A stimulation [\[155\]](#page-25-16). Another pathophysiologic characteristic is the poor tolerance of pulmonary microvascular endothelial cells to hypoxic injury [\[155\]](#page-25-16). Moreover, evidence from experiments in mice has revealed that inhibition of Hypoxia-inducible factor 1α could decrease right ventricular systolic pressure and hypertrophy as well as the amount of fibrosis and obliterative pulmonary vascular remodeling [\[163\]](#page-26-2).

Endothelial cell permeability is compromised in PAH and, under the expression of growth factors such as VEGF, activation of pro-inflammatory mediators and cytokines occurs [\[164\]](#page-26-3). As a result, inflammation perpetuates vascular damage and endothelial barrier permeability, which is responsible for the distorted gas exchange and coagulation between lung and blood tissue [\[165\]](#page-26-4). Moreover, crosstalk between endothelial cells and VSMCs, as well as with non-smooth muscle cells, results in the chemoattraction of immune cells at the sites of vascular damage such as myofibroblasts and pro-inflammatory leucocytes [\[166\]](#page-26-5). Another important pathophysiologic aspect is endothelial-to-mesenchymal transition in which endothelial cells transform into a profile that resembles myofibroblast or mesenchymal cells; as a matter of fact, endothelial cells lose the expression of their typical markers such as CD31 and cadherins and exhibit proliferation of α -smooth muscle actin and vimentin [\[167\]](#page-26-6). According to the studies, the transforming growth factor-β (TGF-β) signaling pathway mediates the expression of Smooth muscle alpha-actin (α SMA) and type I collagen (and not VE-cadherin) in pulmonary arterial endothelial cells [\[168,](#page-26-7)[169\]](#page-26-8). Endothelial-to-mesenchymal transition is controlled epigenetically by several miR such as miR-21; for instance, TGF-β augments miR-21 expression in endothelial cells through AKT-dependent pathway [\[170\]](#page-26-9). Also, there is evidence that miR affect ion expression, mitochondrial function and are implicated in the angiogenic impairment in PAH [\[171\]](#page-26-10).

A genetic involvement is present in 6–10% of patients with PAH and most commonly involves the heterozygous germline mutation of the *Bone Morphogenetic Protein Receptor* (*BMPR)* gene, which encodes type 2 bone morphogenetic protein receptor (BMPR-2) [\[172\]](#page-26-11). There is evidence that autophagy in the lysosomes of human pulmonary artery endothelial cells might contribute to BMPR-2 deletion in PAH [\[173\]](#page-26-12). Additionally, control of gene expression at the transcriptional, post-translational, and post-transcriptional level by long

non-coding RNAs is implicated according to recent studies in the development of pulmonary vascular remodeling through effects in endothelial function, cell proliferation, angiogenesis, endothelial-to-mesenchymal transition and cellular metabolism [\[174\]](#page-26-13). Last but not least, improper NO release due to ED induces DNA damage and metabolic dysregulation [\[175\]](#page-26-14). Indeed, it has been described that pulmonary artery endothelial cells and adventitial fibroblasts display a shift from glucose oxidation towards uncoupled aerobic glycolysis, which hinders the contractility of cardiomyocytes [\[176\]](#page-26-15). Moreover, metabolic abnormalities in PAH include irregular polyamine and sphingosine metabolism, impaired insulin sensitivity, and poor iron handling [\[177\]](#page-26-16). Gene expression studies indicate that human pulmonary artery smooth muscle cells exhibit increased fatty acid metabolism, formation of unsaturated fatty acids as well as a phenotype of the energy-driven proliferative profile along with decreased expression of the genes of the tricarboxylic acid cycle [\[178](#page-26-17)[,179\]](#page-26-18). For instance, the deletion of peroxisome proliferator-activated receptor γ (PPAR γ) results in systolic dysfunction of both ventricles and lipid accumulation inside cardiomyocytes [\[180\]](#page-26-19). Finally, a disintegrin and metalloproteinase with thrombospondin motifs 8 (ADAMTS8) (disintegrin and metalloproteinase with thrombospondin motifs 8) is implicated, according to experimental evidence, in mitochondrial fragmentation under hypoxia as well as with the proliferation of pulmonary artery smooth muscle cells [\[181\]](#page-26-20).

3.3. Endothelial Function in the Course of Heart Failure

Impaired endothelial function in patients with HF is associated with poorer cardiovascular outcomes, implying proper therapeutic management of ED [\[182\]](#page-26-21). ED is more severe in HFpEF and appears earlier during the progression of HFpEF; contrarily, ED is evident at a later stage in HFrEF [\[4\]](#page-19-3). Moreover, ED of the peripheral vasculature is maximal during the early stages of HF in contrast to more severe stages of this syndrome [\[183\]](#page-26-22). Patients with acute HF are characterized by the presence of a procoagulant state due to ED, which is linked to poorer cardiovascular prognosis [\[184\]](#page-27-0). Deteriorated peripheral endothelial function may also predict long-term cardiovascular events in patients with end-stage HF and could facilitate risk stratification strategies in these patients [\[185\]](#page-27-1). Similarly, impaired peripheral endothelial function and decreased exhaled NO (as a marker of pulmonary circulation) during submaximal exercise in patients with chronic HF have been linked with higher mortality after adjustments for clinical factors [\[186\]](#page-27-2). Finally, according to another study, evaluation of ED in patients who receive treatment with cardiac resynchronization therapy (CRT) might also indicate the patients with better response [\[187\]](#page-27-3).

On the other hand, there is evidence that not only peripheral but also coronary microvascular and epicardial dysfunction are associated with the clinical outcomes in HF; as a matter of fact, preservation of endothelial function might improve LVEF in patients with HF [\[188\]](#page-27-4). Coronary microvascular ED drives the development of HFpEF due to the existence of an environment of chronic, subclinical inflammation [\[188\]](#page-27-4). Interestingly, circulating inflammatory markers have been related to coronary microvascular ED (assessed by transthoracic Doppler echocardiography), as well as with markers of diastolic dysfunction, such as the increased E/e' ratio [\[188\]](#page-27-4). Similarly, in a study of patients with HFpEF, oxidative stress reflected by the levels of increased MPO, uric acid, calprotectin, and symmetric dimethyl arginine, are associated with diastolic dysfunction; as a matter of fact, microvascular ED was linked to worse cardiovascular prognosis [\[189\]](#page-27-5).

Patients with acute and chronic HF exhibit various degrees of ED and circulating biomarkers of endothelial activation [\[190\]](#page-27-6). Interestingly, a study in hypertensive patients revealed that impaired endothelial function and high CRP levels are associated with the development of new-onset HF [\[191\]](#page-27-7). Furthermore, according to recent data, high circulating levels of biomarkers of endothelial glycocalyx impairment are linked to increased mortality in patients with decompensated HFrHF, implying the role of dysfunctional endothelium for poor prognosis [\[90\]](#page-22-16). Moreover, ED could predict the incidence of adverse events in acute HF as well as HF progression [\[68\]](#page-21-16). Moreover, the identification of ED might reveal individuals at risk for developing HF and facilitate the therapeutic monitoring of those

who already receive cardiotoxic agents [\[88\]](#page-22-14). Interestingly, according to another study by our research team, FMD could serve as a risk-stratification tool in order to evaluate anthracycline-induced cardiotoxicity in patients who receive chemotherapy [\[192\]](#page-27-8) (Figure [1\)](#page-11-0). (Figure 1).

Figure 1. Role of Endothelial dysfunction on Heart failure. Endothelial dysfunction induces im-**Figure 1.** Role of Endothelial dysfunction on Heart failure. Endothelial dysfunction induces impaired NO production and ROS overproduction. Excessive ROS increases ADMA levels resulting in decreased exercise tolerance. Moreover, ROS production at mitochondria and the presence of dysfunctional antioxidant system cause hypoxia and accelerate myocardial damage. Deterioration of NO-cGMP-PKG pathway increases diastolic dysfunction, myocardial fibrosis, and myocardial hypertrophy. NO: nitric oxide; ROS: reactive oxygen species; eNOS: Endothelial nitric oxide synthase; tension III receptor to charge 2; New York Catalogue 2; New York Charge and the charge symmetric Dimethylarginine; ATR1: Angiotensin II receptor type 1; ATR2: Angiotensin II receptor type 2; NF-Kβ: Nuclear factor kappa beta; ICAM-1: Intercellular Adhesion Molecule 1; VCAM-1: vascular cell adhesion molecule 1; cGMP: Cyclic guanosine monophosphate; sGC: soluble guanylate cyclase; PKG: protein kinase G. Parts of the figure were drawn by using pictures from Servier Medical Art. Servier Medical Art by Servier is licensed under a Creative Commons Attribution 3.0 Unported License [\(https://creativecommons.org/licenses/by/3.0/](https://creativecommons.org/licenses/by/3.0/) accessed on 23 of December 2022).

4. Modification of Endothelial Dysfunction in Patients with Heart Failure

4.1. Effects of Exercise Training

Exercise and cardiac rehabilitation have demonstrated beneficial effects in patients with HF and especially in HFrEF; as a matter of fact, HFrEF is characterized by exercise intolerance, and several assessments could evaluate exercise capacity in these patients, such as a 6-minute walk test and peak oxygen uptake (VO2) [\[193\]](#page-27-9). It should be pointed out, though, that not all patients are capable of cardiac rehabilitation, and those who are appropriate candidates achieve an improved quality of life, better exercise capacity, and fewer cardiovascular events [\[193\]](#page-27-9). The benefits of structured exercise training (ET) have been well established for HF and have been given a class IA recommendation for patients with stable HF [\[194](#page-27-10)[,195\]](#page-27-11). Three modalities of ET have been proposed: enduranceaerobic (continuous or interval training, with superior effects of endurance on LV function), strength/resistance training (individually tailored to each patient's needs), and respiratory training (preferred in circumstances of inspiratory muscle weakness) [\[196\]](#page-27-12).

According to the literature, ET improves FMD through effects on shear stress as well as arterial compliance [\[197\]](#page-27-13). Specifically, evidence from a meta-analysis of 16 studies indicated that ET enhances NO bioavailability by assisting eNOS function and antioxidant enzymes expression, mobilization of EPCs, and decrease in TNF- α , IL-10, and IL-6 expression [\[197\]](#page-27-13). Interestingly, in the Leipzig Exercise Intervention in Chronic heart failure and Aging (LE-ICA) study, 4-week ET in patients with stable congestive HF improved FMD regardless of age; a rise in EPCs number and function was also documented [\[198\]](#page-27-14). Moreover, maximal cardiopulmonary exercise testing (CPET) in patients with chronic HF of different severity has displayed enhanced mobilization and circulation of EPCs, although the effect was irrelevant to the disease severity [\[60\]](#page-21-9). ET, according to another study, demonstrated favorable effects in LVEF of elderly patients with chronic HF, which was mediated by mobilization of EPCs, stimulation of NO and VEGF expression, as well as of PI3 K/AKT pathway of angiogenesis [\[199\]](#page-27-15). Interestingly, 12 weeks of high-intensity interval exercise (HIIT) in patients with HFrEF resulted in decreased SNS activity and better peripheral vascular function reflected by brachial artery FMD in contrast to moderate-intensity continuous training [\[200\]](#page-27-16).

As for HFpEF, ET has indicated favorable effects and is considered to be an important non-pharmaceutic option, which improves the quality of life and exercise capacity through mechanisms that involve endothelial function, such as the regulation of inflammation [\[201\]](#page-27-17). In elderly patients with HFpEF, 16 weeks of ET had a beneficial effect on VO2 and quality of life without effects on FMD or arterial stiffness, and the possible mechanism could be better skeletal muscle perfusion or oxygen utilization [\[202\]](#page-27-18). According to a systematic review of 9 studies of patients with HFpEF, ET resulted in higher VO2 uptake, 6-minute walking distance, and improved ventilation threshold, although there was no significant effect on endothelial function and arterial stiffness; interestingly, only in some of the studies echocardiographic parameters and quality of life displayed improvement [\[195\]](#page-27-11).

In conclusion, ET exerts beneficial effects in the appropriate candidates with HF, and the possible mechanisms involve the improvement of endothelial function and SNS activity, which result in enhanced exercise tolerance, possibly due to better diffusion and utilization of oxygen in skeletal muscles and oxygen transport to tissues [\[193\]](#page-27-9).

4.2. Drugs for the Treatment of Endothelial Dysfunction in Patients with Heart Failure 4.2.1. Statins

Statins, known as 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductases, display not only lipid-lowering actions against LDL levels but also pleiotropic actions such as anti-inflammatory, anti-atherosclerotic, and antioxidant actions [\[203\]](#page-27-19). In fact, these cardiovascular effects are independent of lipid-lowering actions and involve the mobilization of EPCs in circulation and enhancement of endothelial function [\[203\]](#page-27-19). According to the literature, statins improve LV remodeling and diastolic dysfunction of the heart as well as natriuretic peptide expression, possibly through the control of inflammation and restoration of endothelial function [\[204\]](#page-27-20).

Patients with HFrEF statins displayed increased mobilization of EPCs, which further improved the exercise capacity and morbidity of the patients [\[205\]](#page-28-0). Additionally, statins decrease TNF- α levels and augment NO bioavailability through the stabilization of the mRNA of eNOs synthase [\[205\]](#page-28-0). Similarly, in another study, short-term administration of rosuvastatin, but not allopurinol, increased the number of circulating EPCs in patients with systolic HF, although EPCs did not display associations with biomarkers of oxidative stress and inflammation [\[206\]](#page-28-1). Furthermore, in a study of patients with congestive HF administration of statins activated circulating CD34+ EPCs and enhanced the neovascularization process, which was reflected by higher expression of VEGF, improved endothelial function, and LVEF [\[207\]](#page-28-2). In patients with congestive HF, atorvastatin restored peripheral endothelial function assessed by gauge-strain plethysmography and decreased the expression of TNF-α, IL-6, and VCAM-1 [\[208\]](#page-28-3). Higher doses of atorvastatin, in contrast to the lower doses, induced favorable effects on endothelial function in patients with ischemic HF and

improved FMD, augmentation index, as well as MMP-9 and ICAM-1 levels [\[209\]](#page-28-4). Last but not least, the administration of rosuvastatin in patients with congestive HF raised FMD in contrast to the administration of ezetimibe, although there were no differences in lipid levels between study arms [\[210\]](#page-28-5).

4.2.2. Beta Blockers

Beta-blockers (β blockers) belong to the antagonists of beta-adrenergic receptors, which are normally expressed in cardiomyocytes and mediate the actions of SNS [\[211\]](#page-28-6). Beta-blockers bind to $β1$, $β2$, and $β3$ receptors of the G-protein-coupled receptors family; the first generation of β blockers is non-selective for β 1 receptors, the second-generation of beta blockers is more cardio-selective, and the third generation is highly selective for β 1 receptors [\[211\]](#page-28-6). As a matter of fact, the third generation of β blockers exhibits vasodilatory properties along with antioxidant, anti-proliferative, anti-hypertrophic, angiogenic, and anti-apoptotic properties, which are under investigation [\[211\]](#page-28-6). Therefore, β blockers with NO-mediated vasodilatory properties could be a promising treatment for the restoration of ED [\[212\]](#page-28-7). Moreover, according to clinical trials, β blockers display a survival benefit for patients with chronic HF, too [\[213\]](#page-28-8).

Interestingly, in the study of Chin BSP et al., a three-month scheme of β blockers in patients with chronic HF resulted in marked improvement of biomarkers of lipid peroxidation without changes in total antioxidant capacity and vWF levels [\[214\]](#page-28-9). In another study in patients with congestive HF and NYHA II-III, administration of bisoprolol for 20 ± 10 weeks could also restore ED [\[215\]](#page-28-10). Similarly, a 3-week regimen of carvedilol in patients with HF and NYHA II-III improved L-arginine and L-citrulline levels and decreased the expression of VCAM-1, implying a beneficial effect in endothelium-dependent dilatation, fibrinolysis and hemorheological profile of patients [\[216\]](#page-28-11). Furthermore, Poelzl G. et al. displayed that short-term administration of β blockers and ACE inhibitors in patients with chronic HF improved FMD and submaximal exercise capacity [\[217\]](#page-28-12). Additionally, treatment with carvedilol for six months could improve HF functional class, LVEF, 6-minute walk distance in patients with chronic HF (without effects in peak VO2 max), and plasma malondialdehyde levels [\[218\]](#page-28-13). What is more, in a study of patients with HF and NYHA II-IV, administration of carvedilol for 40 ± 14 months ameliorated HF symptoms and the expression of pro-inflammatory biomarkers irrespectively of LVEF; interestingly, patients who exhibited an improvement of LVEF had also decreased ADMA levels [\[219\]](#page-28-14). As for the effects of specific β blockers, the switch from carvedilol to either metoprolol tartrate or succinate in patients with mild HF did not affect endothelium-dependent vasodilation, blood pressure, or heart rate [\[220\]](#page-28-15). Last but not least, combination of sacubitril/valsartan and metoprolol in patients with congestive HF ameliorated LV end-systolic and end-diastolic dimensions, LVEF, biomarkers of oxidative stress and coagulation parameters, implying possible effects in cardiac remodeling [\[221\]](#page-28-16). In conclusion, the majority of studies underline that β blockers might restore ED in HF, and to our knowledge, only one study in patients with chronic HF has demonstrated neutral effects of $β$ blockers on markers of endothelial, platelet, or hemorheological function [\[222\]](#page-28-17).

4.2.3. ACE Inhibitors/ARBs

Angiotensin-converting enzyme inhibitors (ACEi) are the mainstay drug category of patients with HF due to their antihypertensive and anti-atherosclerotic properties, including the ability to delay the progression of LV remodeling [\[223\]](#page-28-18). Concerning their effect on endothelial function, chronic administration of ACEi in patients with congestive HF has proven beneficial through the improvement of FMD, compliance, and distensibility of the brachial artery, possibly due to their blood-pressure-lowering actions [\[224\]](#page-28-19). According to another study, administration of ACEi but not beta-blockers in patients with chronic HF resulted in a more controlled hypercoagulable state, which was reflected by the decreased levels of soluble P-selectin, vWF, and fibrinogen, especially in females and in patients with more progressed NYHA status [\[222\]](#page-28-17). Interestingly, treatment of patients with chronic HF with ramipril and sildenafil (both solely and in combination) improved FMD, and this effect remained significant at 4-hour post intervention [\[225\]](#page-28-20). Lastly, a recent study that investigated patients with HFmrEF and HFpEF demonstrated that therapy with perindopril for 12 months ameliorated endothelial function of large blood vessels and microvessels (assessed by the method of photoplethysmography); interestingly, both categories of HF exhibited decreased expression of E-selectin, whereas ET-1 had the maximal improvement in patients with HFpEF [\[226\]](#page-28-21).

Angiotensin Receptor Inhibitors (ARBIs) selectively block the AT-1 receptor pathway, which mediates their antihypertensive functions; as a result, angiotensin-II binds to the AT-2 receptor, which possesses atheroprotective functions [\[227\]](#page-29-0). ARBIs do not affect the bradykinin pathway, which is characteristic of ACEi effects on endothelial continuity [\[227\]](#page-29-0). Angiotensin-II induces deleterious effects on endothelial function, which involve the senescence of EPCs due to oxidative stress and telomerase inactivation [\[228\]](#page-29-1). ARBIs, in general, have beneficial actions against several atherosclerotic diseases, including HF [\[33\]](#page-20-9). Nevertheless, their exact effects on endothelial function are less well understood [\[227\]](#page-29-0). For instance, according to a study of patients with congestive HF, ACEi or AT-II antagonists improved flow-dependent vasodilatation, shear stress, as well as compliance and distensibility of the radial artery [\[229\]](#page-29-2).

4.2.4. Mineralocorticoid Receptor Antagonists (MRA)

Aldosterone is a mineralocorticoid hormone with detrimental effects on endothelial cells and cardiomyocytes; specifically, aldosterone is implicated in cardiac hypertrophy and fibrosis in HF in addition to direct vascular injury [\[230\]](#page-29-3). Aldosterone also hinders the function, growth, and mobilization of EPCs in a concentration-dependent manner through VEGF-mediated phosphorylation of the Akt pathway [\[230](#page-29-3)[,231\]](#page-29-4). MRAs, such as eplerenone and spironolactone, neutralize the harmful effects of aldosterone on the cardiovascular system [\[231\]](#page-29-4). Indeed, treatment with spironolactone restores endothelial function through augmentation of NO bioavailability and endothelium-dependent vasodilation in patients with NYHA class II-III chronic HF under standard diuretic/ACEI therapy [\[232\]](#page-29-5). Similarly, in another study, the administration of spironolactone in patients with congestive HF improves FMD at 4 weeks, and these effects remain at 8 weeks, possibly due to the attenuation of aldosterone actions in endothelial function [\[233\]](#page-29-6). Finally, in a recent study, Levi et al. displayed that 8-week treatment with eplerenone or spironolactone in patients with congestive HF raised VEGFR2+/CD34+ and VEGFR-2+/CD133+ levels of circulating EPCs, implying the beneficial effects of MRA antagonism for maintenance of endothelial function [\[230\]](#page-29-3).

4.2.5. SGLT2 Inhibitors

The novel drug category of sodium-glucose cotransporter 2 (SGLT2) inhibitors has cardioprotective effects in patients with HF through various mechanisms, including improvement of endothelial function, aside from their established effectiveness in the treatment of diabetes mellitus [\[234\]](#page-29-7). SGLT2 inhibitors act on the renal proximal tubule, reduce glucose reuptake, and promote sodium excretion leading to glycosuria, natriuresis, and diuresis [\[235\]](#page-29-8). According to the literature, the intracellular decrease in sodium levels is responsible for the cardioprotective actions of SGLT2 inhibitors in HF through amelioration of calcium ion handling in cardiomyocytes; as a result, there is an enhancement of the electromechanical function of the heart [\[235\]](#page-29-8). Other mechanisms involve control of oxidative stress, fibrosis, autophagy, and inflammation [\[235\]](#page-29-8). Evidence from experimental studies indicates that SGLT2 inhibitors counteract mitochondrial dysfunction (due to energy starvation in HF), activate the sirtuin-1 pathway and stimulate ketogenesis [\[236,](#page-29-9)[237\]](#page-29-10). Apart from the direct effects of SGLT2 inhibitors in cardiomyocytes, many of their actions involve the regulation of ED, diastolic dysfunction, cardiac stiffness, and reduction of epicardial tissue [\[238](#page-29-11)[,239\]](#page-29-12).

Empagliflozin improves eNOS-dependent PKGIα oxidation and decreases the expression of ICAM-1, VCAM-1, IL-6, and TNF- α in myocardial tissues of patients with

HFpEF [\[240\]](#page-29-13). Additionally, empagliflozin enhances the phosphorylation of myofilament proteins and the NO-cGMP pathway, which reflects its antioxidant effects [\[240\]](#page-29-13). Cardiomyocytes and macrophages, when treated with empagliflozin, present activation of the AMK kinase pathway as well as inhibition of iNOS function [\[241\]](#page-29-14). Data from clinical studies are yet scarce; nevertheless, according to an observational, nonrandomized study, the administration of empagliflozin in diabetic patients with chronic HF improves FMD [\[182\]](#page-26-21).

4.2.6. Sacubitril-Valsartan (ARNIs)

Sacubitril-valsartan, an angiotensin-receptor/neprilysin inhibitor (ARNI), is a drug category for chronic symptomatic HFrEF and HFpEF according to the recent guidelines for HF [\[242\]](#page-29-15). Sacubitril is a pro-drug, and its activated metabolite inhibits neprilysin from breaking down natriuretic peptides; as a result, vasodilation, natriuresis, and diuresis occur, which are associated with enhanced endothelial function [\[242\]](#page-29-15). Blockage of neprilysin increases bradykinin, which possesses endothelium-dependent vasodilatory actions, too [\[242\]](#page-29-15). On the other hand, valsartan is an ARB that blocks the RAAS system and protects against vasoconstriction, hypertension, and cardiac remodeling in HF [\[242,](#page-29-15)[243\]](#page-29-16). Nevertheless, neprilysin disintegrates angiotensin II; therefore, sacubitril should be used along with an ARB in order to counteract high levels of angiotensin II [\[242](#page-29-15)[,243\]](#page-29-16).

Considering the effects of sacubitril-valsartan on endothelial function, Amore et al. indicated that treatment with sacubitril-valsartan for six months in patients with dilated cardiomyopathy and reduced LVEF restore endothelial function, LVEF, diastolic dysfunction, and mitral regurgitation, without any significant effects in arterial stiffness [\[244\]](#page-29-17). Interestingly, a study in 80 patients with HFrEF proves that sacubitril-valsartan added on standard-of-care regimens for a period of 12 weeks augment FMD; furthermore, an increase in NO and NOS levels, LVEF and calcitonin gene-related peptide was found as well as decreased expression of ET-1 [\[245\]](#page-29-18). Lastly, sacubitril-valsartan administration for 12 weeks on top of conventional treatment improved FMD as well as arterial stiffness of patients with HF, too [\[246\]](#page-29-19) (Table [1\)](#page-15-0).

Table 1. Therapeutic options for the treatment of endothelial dysfunction in patients with heart failure.

Table 1. *Cont.*

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Table 1. *Cont.*

ADMA: asymmetric dimethylarginine; ACEi: Angiotensin-converting enzyme inhibitor; Alx: Augmentation index; ARBs: Angiotensin Receptor Inhibitors; BNP: Brain natriuretic peptide; EID: endothelial independent dilation; EPCs: endothelial progenitor cells; ERAs: Endothelin receptor antagonists; ET: Exercise training; ET-1: Endothelin-1; FMD: flow-mediated dilatation; HFrEF: heart failure with reduced ejection fraction; HFmrEF: heart failure with mildly reduced ejection fraction; HFpEF: heart failure with preserved ejection fraction; ICD: Implantable cardioverter defibrillator; IL: interleukin; IVUS: Intravascular ultrasound; LFA-1: Lymphocyte function-associated antigen-1; LVEF: left ventricular ejection fraction; LVFS: left ventricular short-axis shortening rate; LVEDD: left ventricular end-diastolic diameter; LVESD: left ventricular end-systolic diameter; MRA: Mineralocorticoid Receptor Antagonists; MMP: matrix metalloproteinase; MPO: myeloperoxidase; NT-proBNP: N-terminal probrain natriuretic peptide; oxLDL: oxidized low-density lipoprotein; PAH: Pulmonary arterial hypertension; PAP: Pulmonary artery pressure; PECAM-1: platelet/endothelial cell adhesion molecule; PerOx: total lipid peroxides; PVR: pulmonary vascular resistance; PWV: Pulse wave velocity; RCT: Randomized controlled trial; RHI: reactive hyperemia index; RH-PAT; reactive hyperemia-peripheral arterial tonometry; SGLT2 i: Sodiumglucose-cotransporter-type-2 inhibitors; sVCAM-1: soluble vascular cell adhesion molecule; sICAM-1: soluble intercellular adhesion molecule; SDF-1: stromal-derived factor 1; SMD: Standardized mean difference; TNF-α: tumor necrosis factor alpha; VEGF: vascular endothelial growth factor; vWF: von Willebrand factor.

4.2.7. Endothelin Receptor Antagonists (ERAs)

ET-1 is a powerful vasoconstrictor involved in several cardiovascular diseases, including PAH and congestive HF, and participates in the processes of cardiac hypertrophy, inflammation, and atherosclerosis [\[259\]](#page-30-10). ET-1 exerts vasoconstrictive and pro-inflammatory actions upon binding to ETA receptors of smooth muscle cells and contradictory actions upon binding to ETB receptors of pulmonary artery endothelial cells, which trigger the clearance of ET-1 from circulation and the release of endogenous NO and prostacyclin [\[260](#page-30-11)[,261\]](#page-30-12). Therefore, blockage of ETA receptors could be a promising therapeutic option against PAH, and currently three endothelin receptor antagonists (ERAs)—bosentan, ambrisentan, and macitentan—have been evaluated in clinical trials and have displayed favorable effects in PAH [\[259,](#page-30-10)[262\]](#page-30-13). According to evidence from in vitro studies, bosentan improves endothelial function and decreases neointimal and smooth muscle cell proliferation in PAH [\[263\]](#page-30-14). Furthermore, evidence from in vivo studies in pigs have demonstrated that bosentan might partially improve hypoxia-related decrease in NO production [\[264\]](#page-30-15). Interestingly, administration of bosentan in humans with PAH for a period of six months resulted in ameliorated endothelial function of pulmonary microcirculation upon invasive assessment of endothelial function during right heart catheterization [\[263\]](#page-30-14). Similarly, in a study of patients with systemic sclerosis and no-PAH-related symptoms, bosentan reduced exercise-induced PAH; as a matter of fact, bosentan improved 6-minute walk indices, FMD of the brachial artery, and peripheral vasodilation [\[265\]](#page-30-16). Another study in patients with moderate to severe idiopathic PAH indicated that treatment with bosentan diminuted the expression of proinflammatory biomarkers such as ICAM-1, VCAM-1, IL-6 along with BNP and ameliorated the clinical status of the patients [\[256\]](#page-30-7). What is more, evidence from a study of patients with systemic sclerosis and PAH proved that a regimen of bosentan for 12 months could normalize the expression of ICAM-1, VCAM-1, P-selectin, and platelet/endothelial cell adhesion molecule (PECAM-1) and restore T-cell function [\[257\]](#page-30-8). Additionally, in patients

with connective tissue diseases and PAH, a 3-month scheme of bosentan could ameliorate several biomarkers of endothelial function such as NO and sCD40 L and clinical status; interestingly, responders to the treatment demonstrated a decrease in P-selectin levels [\[266\]](#page-30-17). Last but not least, a study by Sfikakis PP et al. demonstrated that a 4-week scheme of bosentan in patients with systemic sclerosis could improve FMD of the brachial artery but not endothelium-independent vascular function; therefore, it has been speculated that this drug protects against systemic scleroderma-associated endothelial injury [\[258\]](#page-30-9) (Table [2\)](#page-18-0).

Table 2. Ongoing clinical trials evaluating the therapeutic modalities against endothelial dysfunction in heart failure.

6 MWT: six-minute walk test; CPET: Cardiopulmonary Exercise Test; EF: Ejection fraction; HF: Heart failure; HGS: Handgrip strength; HRV: Heart rate variability; FMD: Flow mediated dilatation; ICU: Intensive care unit; KCCQ: Kansas City Cardiomyopathy Questionnaire; LVAD: left ventricular assist devices; NO: Nitric oxide.

5. Conclusions

HF is a clinical syndrome with a diverse etiopathology but common pathophysiologic background. Several mechanisms are implicated in the course of HF, progression, and prognosis, with neurohormonal activation considered the key determinant of the syndrome. Endothelial dysfunction is a significant characteristic that accompanies HF of either etiology or type. Moreover, HF per se may worsen endothelial health through decrease in shear stress, stimulation of inflammation, inactivation of eNOS, and increased oxidative stress. Indeed, impaired endothelial function has been linked to deteriorated functional status and ventricular function in patients with HF. Importantly, most of the therapeutic options with established benefits in patients with HF have a parallel beneficial effect in the restoration of endothelial function.

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References

- 1. Bozkurt, B.; Coats, A.J.; Tsutsui, H.; Abdelhamid, M.; Adamopoulos, S.; Albert, N.; Anker, S.D.; Atherton, J.; Bohm, M.; Butler, J.; et al. Universal Definition and Classification of Heart Failure: A Report of the Heart Failure Society of America, Heart Failure Association of the European Society of Cardiology, Japanese Heart Failure Society and Writing Committee of the Universal Definition of Heart Failure. *J. Card. Fail.* **2021**, *27*, 387–413. [\[CrossRef\]](http://doi.org/10.1016/j.cardfail.2021.01.022)
- 2. Becher, P.M.; Lund, L.H.; Coats, A.J.S.; Savarese, G. An update on global epidemiology in heart failure. *Eur. Heart J.* **2022**, *43*, 3005–3007. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehac248) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35578978)
- 3. Ho, J.E.; Lyass, A.; Lee, D.S.; Vasan, R.S.; Kannel, W.B.; Larson, M.G.; Levy, D. Predictors of new-onset heart failure: Differences in preserved versus reduced ejection fraction. *Circ. Heart Fail.* **2013**, *6*, 279–286. [\[CrossRef\]](http://doi.org/10.1161/CIRCHEARTFAILURE.112.972828) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23271790)
- 4. Paulus, W.J.; Tschope, C. A novel paradigm for heart failure with preserved ejection fraction: Comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J. Am. Coll Cardiol.* **2013**, *62*, 263–271. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2013.02.092)
- 5. Taqueti, V.R.; Solomon, S.D.; Shah, A.M.; Desai, A.S.; Groarke, J.D.; Osborne, M.T.; Hainer, J.; Bibbo, C.F.; Dorbala, S.; Blankstein, R.; et al. Coronary microvascular dysfunction and future risk of heart failure with preserved ejection fraction. *Eur. Heart J.* **2018**, *39*, 840–849. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehx721)
- 6. Daiber, A.; Xia, N.; Steven, S.; Oelze, M.; Hanf, A.; Kroller-Schon, S.; Munzel, T.; Li, H. New Therapeutic Implications of Endothelial Nitric Oxide Synthase (eNOS) Function/Dysfunction in Cardiovascular Disease. *Int. J. Mol. Sci.* **2019**, *20*, 187. [\[CrossRef\]](http://doi.org/10.3390/ijms20010187)
- 7. Breitenstein, S.; Roessig, L.; Sandner, P.; Lewis, K.S. Novel sGC Stimulators and sGC Activators for the Treatment of Heart Failure. *Handb. Exp. Pharmacol.* **2017**, *243*, 225–247. [\[CrossRef\]](http://doi.org/10.1007/164_2016_100)
- 8. Cornuault, L.; Rouault, P.; Duplaa, C.; Couffinhal, T.; Renault, M.A. Endothelial Dysfunction in Heart Failure With Preserved Ejection Fraction: What are the Experimental Proofs? *Front. Physiol.* **2022**, *13*, 906272. [\[CrossRef\]](http://doi.org/10.3389/fphys.2022.906272)
- 9. Lee, D.S.; Gona, P.; Vasan, R.S.; Larson, M.G.; Benjamin, E.J.; Wang, T.J.; Tu, J.V.; Levy, D. Relation of disease pathogenesis and risk factors to heart failure with preserved or reduced ejection fraction: Insights from the framingham heart study of the national heart, lung, and blood institute. *Circulation* **2009**, *119*, 3070–3077. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.108.815944)
- 10. Tota, B.; Cerra, M.C.; Gattuso, A. Catecholamines, cardiac natriuretic peptides and chromogranin A: Evolution and physiopathology of a 'whip-brake' system of the endocrine heart. *J. Exp. Biol.* **2010**, *213*, 3081–3103. [\[CrossRef\]](http://doi.org/10.1242/jeb.027391)
- 11. Krum, H. New and emerging pharmacological strategies in the management of chronic heart failure. *Curr. Opin. Pharmacol.* **2001**, *1*, 126–133. [\[CrossRef\]](http://doi.org/10.1016/S1471-4892(01)00025-X) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11714086)
- 12. Bonnefont-Rousselot, D.; Mahmoudi, A.; Mougenot, N.; Varoquaux, O.; Le Nahour, G.; Fouret, P.; Lechat, P. Catecholamine effects on cardiac remodelling, oxidative stress and fibrosis in experimental heart failure. *Redox. Rep.* **2002**, *7*, 145–151. [\[CrossRef\]](http://doi.org/10.1179/135100002125000389) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12189044)
- 13. von Lueder, T.G.; Kotecha, D.; Atar, D.; Hopper, I. Neurohormonal Blockade in Heart Failure. *Card. Fail. Rev.* **2017**, *3*, 19–24. [\[CrossRef\]](http://doi.org/10.15420/cfr.2016:22:2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28785471)
- 14. Andreozzi, F.; Laratta, E.; Sciacqua, A.; Perticone, F.; Sesti, G. Angiotensin II impairs the insulin signaling pathway promoting production of nitric oxide by inducing phosphorylation of insulin receptor substrate-1 on Ser312 and Ser616 in human umbilical vein endothelial cells. *Circ. Res.* **2004**, *94*, 1211–1218. [\[CrossRef\]](http://doi.org/10.1161/01.RES.0000126501.34994.96) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15044323)
- 15. Reina-Couto, M.; Afonso, J.; Carvalho, J.; Morgado, L.; Ronchi, F.A.; de Oliveira Leite, A.P.; Dias, C.C.; Casarini, D.E.; Bettencourt, P.; Albino-Teixeira, A.; et al. Interrelationship between renin-angiotensin-aldosterone system and oxidative stress in chronic heart failure patients with or without renal impairment. *Biomed. Pharmacother.* **2021**, *133*, 110938. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2020.110938)
- 16. Kaschina, E.; Namsolleck, P.; Unger, T. AT2 receptors in cardiovascular and renal diseases. *Pharmacol. Res.* **2017**, *125*, 39–47. [\[CrossRef\]](http://doi.org/10.1016/j.phrs.2017.07.008) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28694144)
- 17. Ide, T.; Tsutsui, H.; Kinugawa, S.; Utsumi, H.; Kang, D.; Hattori, N.; Uchida, K.; Arimura, K.; Egashira, K.; Takeshita, A. Mitochondrial electron transport complex I is a potential source of oxygen free radicals in the failing myocardium. *Circ. Res.* **1999**, *85*, 357–363. [\[CrossRef\]](http://doi.org/10.1161/01.RES.85.4.357)
- 18. Dubois-Deruy, E.; Peugnet, V.; Turkieh, A.; Pinet, F. Oxidative Stress in Cardiovascular Diseases. *Antioxidants* **2020**, *9*, 864. [\[CrossRef\]](http://doi.org/10.3390/antiox9090864)
- 19. Bugger, H.; Pfeil, K. Mitochondrial ROS in myocardial ischemia reperfusion and remodeling. *Biochim. Biophys. Acta Mol. Basis Dis.* **2020**, *1866*, 165768. [\[CrossRef\]](http://doi.org/10.1016/j.bbadis.2020.165768)
- 20. Ho, E.; Karimi Galougahi, K.; Liu, C.C.; Bhindi, R.; Figtree, G.A. Biological markers of oxidative stress: Applications to cardiovascular research and practice. *Redox. Biol.* **2013**, *1*, 483–491. [\[CrossRef\]](http://doi.org/10.1016/j.redox.2013.07.006)
- 21. Cesselli, D.; Jakoniuk, I.; Barlucchi, L.; Beltrami, A.P.; Hintze, T.H.; Nadal-Ginard, B.; Kajstura, J.; Leri, A.; Anversa, P. Oxidative stress-mediated cardiac cell death is a major determinant of ventricular dysfunction and failure in dog dilated cardiomyopathy. *Circ. Res.* **2001**, *89*, 279–286. [\[CrossRef\]](http://doi.org/10.1161/hh1501.094115)
- 22. Ide, T.; Tsutsui, H.; Hayashidani, S.; Kang, D.; Suematsu, N.; Nakamura, K.; Utsumi, H.; Hamasaki, N.; Takeshita, A. Mitochondrial DNA damage and dysfunction associated with oxidative stress in failing hearts after myocardial infarction. *Circ. Res.* **2001**, *88*, 529–535. [\[CrossRef\]](http://doi.org/10.1161/01.RES.88.5.529)
- 23. Kiyuna, L.A.; Albuquerque, R.P.E.; Chen, C.H.; Mochly-Rosen, D.; Ferreira, J.C.B. Targeting mitochondrial dysfunction and oxidative stress in heart failure: Challenges and opportunities. *Free Radic. Biol. Med.* **2018**, *129*, 155–168. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2018.09.019) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30227272)
- 24. Piquereau, J.; Ventura-Clapier, R. Maturation of Cardiac Energy Metabolism During Perinatal Development. *Front. Physiol.* **2018**, *9*, 959. [\[CrossRef\]](http://doi.org/10.3389/fphys.2018.00959) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30072919)
- 25. Miranda-Silva, D.; Lima, T.; Rodrigues, P.; Leite-Moreira, A.; Falcao-Pires, I. Mechanisms underlying the pathophysiology of heart failure with preserved ejection fraction: The tip of the iceberg. *Heart Fail. Rev.* **2021**, *26*, 453–478. [\[CrossRef\]](http://doi.org/10.1007/s10741-020-10042-0) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33411091)
- 26. Triposkiadis, F.; Xanthopoulos, A.; Butler, J. Cardiovascular Aging and Heart Failure: JACC Review Topic of the Week. *J. Am. Coll Cardiol.* **2019**, *74*, 804–813. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2019.06.053)
- 27. Schroder, F.; Handrock, R.; Beuckelmann, D.J.; Hirt, S.; Hullin, R.; Priebe, L.; Schwinger, R.H.; Weil, J.; Herzig, S. Increased availability and open probability of single L-type calcium channels from failing compared with nonfailing human ventricle. *Circulation* **1998**, *98*, 969–976. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.98.10.969)
- Gomes, K.P.; Jadli, A.S.; de Almeida, L.G.N.; Ballasy, N.N.; Edalat, P.; Shandilya, R.; Young, D.; Belke, D.; Shearer, J.; Dufour, A.; et al. Proteomic Analysis Suggests Altered Mitochondrial Metabolic Profile Associated With Diabetic Cardiomyopathy. *Front. Cardiovasc. Med.* **2022**, *9*, 791700. [\[CrossRef\]](http://doi.org/10.3389/fcvm.2022.791700)
- 29. Dashwood, A.; Cheesman, E.; Beard, N.; Haqqani, H.; Wong, Y.W.; Molenaar, P. Understanding How Phosphorylation and Redox Modifications Regulate Cardiac Ryanodine Receptor Type 2 Activity to Produce an Arrhythmogenic Phenotype in Advanced Heart Failure. *ACS Pharmacol. Transl. Sci.* **2020**, *3*, 563–582. [\[CrossRef\]](http://doi.org/10.1021/acsptsci.0c00003)
- 30. Rupee, S.; Rupee, K.; Singh, R.B.; Hanoman, C.; Ismail, A.M.A.; Smail, M.; Singh, J. Diabetes-induced chronic heart failure is due to defects in calcium transporting and regulatory contractile proteins: Cellular and molecular evidence. *Heart Fail. Rev.* **2022**. [\[CrossRef\]](http://doi.org/10.1007/s10741-022-10271-5)
- 31. Sipido, K.R.; Volders, P.G.; Vos, M.A.; Verdonck, F. Altered Na/Ca exchange activity in cardiac hypertrophy and heart failure: A new target for therapy? *Cardiovasc. Res.* **2002**, *53*, 782–805. [\[CrossRef\]](http://doi.org/10.1016/S0008-6363(01)00470-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11922890)
- 32. Gwathmey, J.K.; Liao, R.; Ingwall, J.S. Comparison of twitch force and calcium handling in papillary muscles from right ventricular pressure overload hypertrophy in weanling and juvenile ferrets. *Cardiovasc. Res.* **1995**, *29*, 475–481. [\[CrossRef\]](http://doi.org/10.1016/S0008-6363(96)88522-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/7796440)
- 33. Werner, C.; Baumhakel, M.; Teo, K.K.; Schmieder, R.; Mann, J.; Unger, T.; Yusuf, S.; Bohm, M. RAS blockade with ARB and ACE inhibitors: Current perspective on rationale and patient selection. *Clin. Res. Cardiol.* **2008**, *97*, 418–431. [\[CrossRef\]](http://doi.org/10.1007/s00392-008-0668-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18454336)
- 34. Studer, R.; Reinecke, H.; Bilger, J.; Eschenhagen, T.; Bohm, M.; Hasenfuss, G.; Just, H.; Holtz, J.; Drexler, H. Gene expression of the cardiac Na(+)-Ca2+ exchanger in end-stage human heart failure. *Circ. Res.* **1994**, *75*, 443–453. [\[CrossRef\]](http://doi.org/10.1161/01.RES.75.3.443) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/8062418)
- 35. Schwinger, R.H.; Munch, G.; Bolck, B.; Karczewski, P.; Krause, E.G.; Erdmann, E. Reduced Ca(2+)-sensitivity of SERCA 2a in failing human myocardium due to reduced serin-16 phospholamban phosphorylation. *J. Mol. Cell Cardiol.* **1999**, *31*, 479–491. [\[CrossRef\]](http://doi.org/10.1006/jmcc.1998.0897)
- 36. Liu, G.; Li, S.Q.; Hu, P.P.; Tong, X.Y. Altered sarco(endo)plasmic reticulum calcium adenosine triphosphatase 2a content: Targets for heart failure therapy. *Diab. Vasc. Dis. Res.* **2018**, *15*, 322–335. [\[CrossRef\]](http://doi.org/10.1177/1479164118774313)
- 37. Bertero, E.; Maack, C. Calcium Signaling and Reactive Oxygen Species in Mitochondria. *Circ. Res.* **2018**, *122*, 1460–1478. [\[CrossRef\]](http://doi.org/10.1161/CIRCRESAHA.118.310082)
- 38. Jiang, M.; Xie, X.; Cao, F.; Wang, Y. Mitochondrial Metabolism in Myocardial Remodeling and Mechanical Unloading: Implications for Ischemic Heart Disease. *Front. Cardiovasc. Med.* **2021**, *8*, 789267. [\[CrossRef\]](http://doi.org/10.3389/fcvm.2021.789267)
- 39. Sabbah, H.N. Targeting the Mitochondria in Heart Failure: A Translational Perspective. *JACC Basic Transl. Sci.* **2020**, *5*, 88–106. [\[CrossRef\]](http://doi.org/10.1016/j.jacbts.2019.07.009)
- 40. Adaniya, S.M.; J, O.U.; Cypress, M.W.; Kusakari, Y.; Jhun, B.S. Posttranslational modifications of mitochondrial fission and fusion proteins in cardiac physiology and pathophysiology. *Am. J. Physiol. Cell Physiol.* **2019**, *316*, C583–C604. [\[CrossRef\]](http://doi.org/10.1152/ajpcell.00523.2018)
- 41. Karamanlidis, G.; Nascimben, L.; Couper, G.S.; Shekar, P.S.; del Monte, F.; Tian, R. Defective DNA replication impairs mitochondrial biogenesis in human failing hearts. *Circ Res* **2010**, *106*, 1541–1548. [\[CrossRef\]](http://doi.org/10.1161/CIRCRESAHA.109.212753) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20339121)
- 42. Yu, H.; Yu, M.; Li, Z.; Zhang, E.; Ma, H. Identification and analysis of mitochondria-related key genes of heart failure. *J. Transl. Med.* **2022**, *20*, 410. [\[CrossRef\]](http://doi.org/10.1186/s12967-022-03605-2)
- 43. Van Linthout, S.; Tschope, C. Inflammation—Cause or Consequence of Heart Failure or Both? *Curr. Heart Fail. Rep.* **2017**, *14*, 251–265. [\[CrossRef\]](http://doi.org/10.1007/s11897-017-0337-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28667492)
- 44. Murphy, S.P.; Kakkar, R.; McCarthy, C.P.; Januzzi, J.L., Jr. Inflammation in Heart Failure: JACC State-of-the-Art Review. *J. Am. Coll. Cardiol.* **2020**, *75*, 1324–1340. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2020.01.014) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32192660)
- 45. Sansonetti, M.; De Windt, L.J. Non-coding RNAs in cardiac inflammation: Key drivers in the pathophysiology of heart failure. *Cardiovasc. Res.* **2022**, *118*, 2058–2073. [\[CrossRef\]](http://doi.org/10.1093/cvr/cvab192)
- 46. Chen, G.Y.; Nunez, G. Sterile inflammation: Sensing and reacting to damage. *Nat. Rev. Immunol.* **2010**, *10*, 826–837. [\[CrossRef\]](http://doi.org/10.1038/nri2873)
- 47. Sekiguchi, K.; Li, X.; Coker, M.; Flesch, M.; Barger, P.M.; Sivasubramanian, N.; Mann, D.L. Cross-regulation between the renin-angiotensin system and inflammatory mediators in cardiac hypertrophy and failure. *Cardiovasc. Res.* **2004**, *63*, 433–442. [\[CrossRef\]](http://doi.org/10.1016/j.cardiores.2004.02.005)
- 48. Mentz, R.J.; O'Connor, C.M. Pathophysiology and clinical evaluation of acute heart failure. *Nat. Rev. Cardiol.* **2016**, *13*, 28–35. [\[CrossRef\]](http://doi.org/10.1038/nrcardio.2015.134)
- 49. Harjola, V.P.; Mullens, W.; Banaszewski, M.; Bauersachs, J.; Brunner-La Rocca, H.P.; Chioncel, O.; Collins, S.P.; Doehner, W.; Filippatos, G.S.; Flammer, A.J.; et al. Organ dysfunction, injury and failure in acute heart failure: From pathophysiology to diagnosis and management. A review on behalf of the Acute Heart Failure Committee of the Heart Failure Association (HFA) of the European Society of Cardiology (ESC). *Eur. J. Heart. Fail.* **2017**, *19*, 821–836. [\[CrossRef\]](http://doi.org/10.1002/ejhf.872)
- 50. Levine, B.; Kalman, J.; Mayer, L.; Fillit, H.M.; Packer, M. Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. *N. Engl. J. Med.* **1990**, *323*, 236–241. [\[CrossRef\]](http://doi.org/10.1056/NEJM199007263230405)
- 51. Tromp, J.; Khan, M.A.; Klip, I.T.; Meyer, S.; de Boer, R.A.; Jaarsma, T.; Hillege, H.; van Veldhuisen, D.J.; van der Meer, P.; Voors, A.A. Biomarker Profiles in Heart Failure Patients With Preserved and Reduced Ejection Fraction. *J. Am. Heart Assoc.* **2017**, *6*, e003989. [\[CrossRef\]](http://doi.org/10.1161/JAHA.116.003989) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28360225)
- 52. Li, W.; Li, Y.; Jiang, F.; Liu, H. Correlation between serum levels of microRNA-21 and inflammatory factors in patients with chronic heart failure. *Medicine (Baltimore)* **2022**, *101*, e30596. [\[CrossRef\]](http://doi.org/10.1097/MD.0000000000030596) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36197244)
- 53. Kablak-Ziembicka, A.; Badacz, R.; Przewlocki, T. Clinical Application of Serum microRNAs in Atherosclerotic Coronary Artery Disease. *J. Clin. Med.* **2022**, *11*, 6849. [\[CrossRef\]](http://doi.org/10.3390/jcm11226849) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36431326)
- 54. Nahrendorf, M.; Swirski, F.K. Innate immune cells in ischaemic heart disease: Does myocardial infarction beget myocardial infarction? *Eur. Heart J.* **2016**, *37*, 868–872. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehv453) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26351395)
- 55. Puhl, S.L.; Steffens, S. Neutrophils in Post-myocardial Infarction Inflammation: Damage vs. Resolution? *Front. Cardiovasc. Med.* **2019**, *6*, 25. [\[CrossRef\]](http://doi.org/10.3389/fcvm.2019.00025) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30937305)
- 56. Oikonomou, E.; Tsaplaris, P.; Anastasiou, A.; Xenou, M.; Lampsas, S.; Siasos, G.; Pantelidis, P.; Theofilis, P.; Tsatsaragkou, A.; Katsarou, O.; et al. Interleukin-1 in Coronary Artery Disease. *Curr. Top. Med. Chem.* **2022**, *22*, 2368–2389. [\[CrossRef\]](http://doi.org/10.2174/1568026623666221017144734) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36263481)
- 57. Takimoto, E.; Kass, D.A. Role of oxidative stress in cardiac hypertrophy and remodeling. *Hypertension* **2007**, *49*, 241–248. [\[CrossRef\]](http://doi.org/10.1161/01.HYP.0000254415.31362.a7) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17190878)
- 58. Tang, T.T.; Yuan, J.; Zhu, Z.F.; Zhang, W.C.; Xiao, H.; Xia, N.; Yan, X.X.; Nie, S.F.; Liu, J.; Zhou, S.F.; et al. Regulatory T cells ameliorate cardiac remodeling after myocardial infarction. *Basic Res. Cardiol.* **2012**, *107*, 232. [\[CrossRef\]](http://doi.org/10.1007/s00395-011-0232-6)
- 59. Zouggari, Y.; Ait-Oufella, H.; Bonnin, P.; Simon, T.; Sage, A.P.; Guerin, C.; Vilar, J.; Caligiuri, G.; Tsiantoulas, D.; Laurans, L.; et al. B lymphocytes trigger monocyte mobilization and impair heart function after acute myocardial infarction. *Nat. Med.* **2013**, *19*, 1273–1280. [\[CrossRef\]](http://doi.org/10.1038/nm.3284)
- 60. Kourek, C.; Karatzanos, E.; Psarra, K.; Georgiopoulos, G.; Delis, D.; Linardatou, V.; Gavrielatos, G.; Papadopoulos, C.; Nanas, S.; Dimopoulos, S. Endothelial progenitor cells mobilization after maximal exercise according to heart failure severity. *World J. Cardiol.* **2020**, *12*, 526–539. [\[CrossRef\]](http://doi.org/10.4330/wjc.v12.i11.526)
- 61. Kawai, C. From myocarditis to cardiomyopathy: Mechanisms of inflammation and cell death: Learning from the past for the future. *Circulation* **1999**, *99*, 1091–1100. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.99.8.1091) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10051305)
- 62. Kuhl, U.; Pauschinger, M.; Seeberg, B.; Lassner, D.; Noutsias, M.; Poller, W.; Schultheiss, H.P. Viral persistence in the myocardium is associated with progressive cardiac dysfunction. *Circulation* **2005**, *112*, 1965–1970. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.105.548156) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16172268)
- 63. Beydoun, N.; Feinstein, M.J. Heart Failure in Chronic Infectious and Inflammatory Conditions: Mechanistic Insights from Clinical Heterogeneity. *Curr. Heart Fail. Rep.* **2022**, *19*, 267–278. [\[CrossRef\]](http://doi.org/10.1007/s11897-022-00560-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35838874)
- 64. Oikonomou, E.; Lampsas, S.; Theofilis, P.; Souvaliotis, N.; Papamikroulis, G.A.; Katsarou, O.; Kalogeras, K.; Pantelidis, P.; Papaioannou, T.G.; Tsatsaragkou, A.; et al. Impaired left ventricular deformation and ventricular-arterial coupling in post-COVID-19: Association with autonomic dysregulation. *Heart Vessels* **2023**, *38*, 381–393. [\[CrossRef\]](http://doi.org/10.1007/s00380-022-02180-2)
- 65. Rajendran, P.; Rengarajan, T.; Thangavel, J.; Nishigaki, Y.; Sakthisekaran, D.; Sethi, G.; Nishigaki, I. The vascular endothelium and human diseases. *Int. J. Biol. Sci.* **2013**, *9*, 1057–1069. [\[CrossRef\]](http://doi.org/10.7150/ijbs.7502)
- 66. Forstermann, U.; Munzel, T. Endothelial nitric oxide synthase in vascular disease: From marvel to menace. *Circulation* **2006**, *113*, 1708–1714. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.105.602532)
- 67. Premer, C.; Kanelidis, A.J.; Hare, J.M.; Schulman, I.H. Rethinking Endothelial Dysfunction as a Crucial Target in Fighting Heart Failure. *Mayo Clin. Proc. Innov. Qual. Outcomes* **2019**, *3*, 1–13. [\[CrossRef\]](http://doi.org/10.1016/j.mayocpiqo.2018.12.006)
- 68. Marti, C.N.; Gheorghiade, M.; Kalogeropoulos, A.P.; Georgiopoulou, V.V.; Quyyumi, A.A.; Butler, J. Endothelial dysfunction, arterial stiffness, and heart failure. *J. Am. Coll. Cardiol.* **2012**, *60*, 1455–1469. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2011.11.082)
- 69. Giannitsi, S.; Bougiakli, M.; Bechlioulis, A.; Naka, K. Endothelial dysfunction and heart failure: A review of the existing bibliography with emphasis on flow mediated dilation. *JRSM Cardiovasc. Dis.* **2019**, *8*, 2048004019843047. [\[CrossRef\]](http://doi.org/10.1177/2048004019843047)
- 70. Colombo, P.C.; Doran, A.C.; Onat, D.; Wong, K.Y.; Ahmad, M.; Sabbah, H.N.; Demmer, R.T. Venous congestion, endothelial and neurohormonal activation in acute decompensated heart failure: Cause or effect? *Curr. Heart Fail. Rep.* **2015**, *12*, 215–222. [\[CrossRef\]](http://doi.org/10.1007/s11897-015-0254-8)
- 71. Al Houri, H.N.; Jomaa, S.; Jabra, M.; Alhouri, A.N.; Latifeh, Y. Pathophysiology of stress cardiomyopathy: A comprehensive literature review. *Ann. Med. Surg.* **2022**, *82*, 104671. [\[CrossRef\]](http://doi.org/10.1016/j.amsu.2022.104671)
- 72. Chignalia, A.Z.; Isbatan, A.; Patel, M.; Ripper, R.; Sharlin, J.; Shosfy, J.; Borlaug, B.A.; Dull, R.O. Pressure-dependent NOS activation contributes to endothelial hyperpermeability in a model of acute heart failure. *Biosci. Rep.* **2018**, *38*, BSR20181239. [\[CrossRef\]](http://doi.org/10.1042/BSR20181239) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30355657)
- 73. Alem, M.M. Endothelial Dysfunction in Chronic Heart Failure: Assessment, Findings, Significance, and Potential Therapeutic Targets. *Int. J. Mol. Sci.* **2019**, *20*, 3198. [\[CrossRef\]](http://doi.org/10.3390/ijms20133198) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31261886)
- 74. Oikonomou, E.; Souvaliotis, N.; Lampsas, S.; Siasos, G.; Poulakou, G.; Theofilis, P.; Papaioannou, T.G.; Haidich, A.B.; Tsaousi, G.; Ntousopoulos, V.; et al. Endothelial dysfunction in acute and long standing COVID-19: A prospective cohort study. *Vascul. Pharmacol.* **2022**, *144*, 106975. [\[CrossRef\]](http://doi.org/10.1016/j.vph.2022.106975) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35248780)
- 75. Kaneko, M.; Matsumoto, Y.; Hayashi, H.; Kobayashi, A.; Yamazaki, N. Oxygen free radicals and calcium homeostasis in the heart. *Mol. Cell Biochem.* **1994**, *139*, 91–100. [\[CrossRef\]](http://doi.org/10.1007/BF00944207)
- 76. Meerson, F.Z.; Kagan, V.E.; Kozlov Yu, P.; Belkina, L.M.; Arkhipenko Yu, V. The role of lipid peroxidation in pathogenesis of ischemic damage and the antioxidant protection of the heart. *Basic Res. Cardiol.* **1982**, *77*, 465–485. [\[CrossRef\]](http://doi.org/10.1007/BF01907940)
- 77. Dai, M.; Wang, Y.; Peng, L.; Liu, X.; Lu, J.; He, L.; Li, K.; Qian, D.; Wang, R. Relationship between Quantitative Parameters of Echocardiography and Vascular Endothelial Function in Patients with Chronic Heart Failure and Its Predictive Value for Short-Term MACE Risk. *J. Healthc. Eng.* **2021**, *2021*, 1985962. [\[CrossRef\]](http://doi.org/10.1155/2021/1985962)
- 78. Shao, Z.; Schuster, A.; Borowski, A.G.; Thakur, A.; Li, L.; Wilson Tang, W.H. Soluble angiotensin converting enzyme 2 levels in chronic heart failure is associated with decreased exercise capacity and increased oxidative stress-mediated endothelial dysfunction. *Transl. Res.* **2019**, *212*, 80–88. [\[CrossRef\]](http://doi.org/10.1016/j.trsl.2019.06.004)
- 79. Hulshoff, M.S.; Xu, X.; Krenning, G.; Zeisberg, E.M. Epigenetic Regulation of Endothelial-to-Mesenchymal Transition in Chronic Heart Disease. *Arterioscler. Thromb. Vasc. Biol.* **2018**, *38*, 1986–1996. [\[CrossRef\]](http://doi.org/10.1161/ATVBAHA.118.311276)
- 80. An, L.; An, S.; Jia, Z.; Wang, H.; Yang, Z.; Xu, C.; Teng, X.; Wang, J.; Liu, X.; Cao, Q.; et al. Atorvastatin improves left ventricular remodeling and cardiac function in rats with congestive heart failure by inhibiting RhoA/Rho kinase-mediated endothelial nitric oxide synthase. *Exp. Ther. Med.* **2019**, *17*, 960–966. [\[CrossRef\]](http://doi.org/10.3892/etm.2018.6976)
- 81. Berezin, A.E.; Kremzer, A.A.; Samura, T.A.; Berezina, T.A. Altered signature of apoptotic endothelial cell-derived microvesicles predicts chronic heart failure phenotypes. *Biomark. Med.* **2019**, *13*, 737–750. [\[CrossRef\]](http://doi.org/10.2217/bmm-2018-0449) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31157550)
- 82. Huang, W.P.; Yin, W.H.; Chen, J.S.; Huang, P.H.; Chen, J.W.; Lin, S.J. Fenofibrate Reverses Dysfunction of EPCs Caused by Chronic Heart Failure. *J. Cardiovasc. Transl. Res.* **2020**, *13*, 158–170. [\[CrossRef\]](http://doi.org/10.1007/s12265-019-09889-y) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31701352)
- 83. Murohara, T.; Asahara, T.; Silver, M.; Bauters, C.; Masuda, H.; Kalka, C.; Kearney, M.; Chen, D.; Symes, J.F.; Fishman, M.C.; et al. Nitric oxide synthase modulates angiogenesis in response to tissue ischemia. *J. Clin. Invest.* **1998**, *101*, 2567–2578. [\[CrossRef\]](http://doi.org/10.1172/JCI1560) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/9616228)
- 84. Giordano, F.J.; Gerber, H.P.; Williams, S.P.; VanBruggen, N.; Bunting, S.; Ruiz-Lozano, P.; Gu, Y.; Nath, A.K.; Huang, Y.; Hickey, R.; et al. A cardiac myocyte vascular endothelial growth factor paracrine pathway is required to maintain cardiac function. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 5780–5785. [\[CrossRef\]](http://doi.org/10.1073/pnas.091415198) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11331753)
- 85. Chong, A.Y.; Blann, A.D.; Patel, J.; Freestone, B.; Hughes, E.; Lip, G.Y. Endothelial dysfunction and damage in congestive heart failure: Relation of flow-mediated dilation to circulating endothelial cells, plasma indexes of endothelial damage, and brain natriuretic peptide. *Circulation* **2004**, *110*, 1794–1798. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.0000143073.60937.50)
- 86. Lampsas, S.; Tsaplaris, P.; Pantelidis, P.; Oikonomou, E.; Marinos, G.; Charalambous, G.; Souvaliotis, N.; Mystakidi, V.C.; Goliopoulou, A.; Katsianos, E.; et al. The Role of Endothelial Related Circulating Biomarkers in COVID-19. A Systematic Review and Meta-analysis. *Curr. Med. Chem.* **2022**, *29*, 3790–3805. [\[CrossRef\]](http://doi.org/10.2174/0929867328666211026124033)
- 87. Prasad, A.; Higano, S.T.; Al Suwaidi, J.; Holmes, D.R., Jr.; Mathew, V.; Pumper, G.; Lennon, R.J.; Lerman, A. Abnormal coronary microvascular endothelial function in humans with asymptomatic left ventricular dysfunction. *Am. Heart J.* **2003**, *146*, 549–554. [\[CrossRef\]](http://doi.org/10.1016/S0002-8703(03)00364-8)
- 88. Taher, R.; Sara, J.D.; Toya, T.; Borlaug, B.A.; Lerman, L.O.; Lerman, A. Peripheral endothelial dysfunction is a novel risk factor for systolic dysfunction and heart failure progression. *Int. J. Cardiol. Heart Vasc.* **2020**, *30*, 100584. [\[CrossRef\]](http://doi.org/10.1016/j.ijcha.2020.100584)
- 89. Triposkiadis, F.; Butler, J.; Abboud, F.M.; Armstrong, P.W.; Adamopoulos, S.; Atherton, J.J.; Backs, J.; Bauersachs, J.; Burkhoff, D.; Bonow, R.O.; et al. The continuous heart failure spectrum: Moving beyond an ejection fraction classification. *Eur. Heart J.* **2019**, *40*, 2155–2163. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehz158)
- 90. Kim, Y.H.; Kitai, T.; Morales, R.; Kiefer, K.; Chaikijurajai, T.; Tang, W.H.W. Usefulness of Serum Biomarkers of Endothelial Glycocalyx Damage in Prognosis of Decompensated Patients with Heart Failure with Reduced Ejection Fraction. *Am. J. Cardiol.* **2022**, *176*, 73–78. [\[CrossRef\]](http://doi.org/10.1016/j.amjcard.2022.04.036)
- 91. Nijst, P.; Cops, J.; Martens, P.; Swennen, Q.; Dupont, M.; Tang, W.H.W.; Mullens, W. Endovascular shedding markers in patients with heart failure with reduced ejection fraction: Results from a single-center exploratory study. *Microcirculation* **2018**, *25*, e12432. [\[CrossRef\]](http://doi.org/10.1111/micc.12432)
- 92. Arnlov, J.; Sang, Y.; Ballew, S.H.; Vaidya, D.; Michos, E.D.; Jacobs, D.R., Jr.; Lima, J.; Shlipak, M.G.; Bertoni, A.G.; Coresh, J.; et al. Endothelial dysfunction and the risk of heart failure in a community-based study: The Multi-Ethnic Study of Atherosclerosis. *ESC Heart Fail.* **2020**, *7*, 4231–4240. [\[CrossRef\]](http://doi.org/10.1002/ehf2.13054) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33084248)
- 93. Martinelli, N.C.; Santos, K.G.; Biolo, A.; La Porta, V.L.; Cohen, C.R.; Silvello, D.; Andrades, M.E.; Clausell, N.; Rohde, L.E. Polymorphisms of endothelial nitric oxide synthase gene in systolic heart failure: An haplotype analysis. *Nitric. Oxide* **2012**, *26*, 141–147. [\[CrossRef\]](http://doi.org/10.1016/j.niox.2012.01.003) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22290017)
- 94. Shantsila, E.; Wrigley, B.; Shantsila, A.; Tapp, L.D.; Blann, A.D.; Gill, P.S.; Lip, G.Y. Ethnic differences in macrovascular and microvascular function in systolic heart failure. *Circ. Heart Fail.* **2011**, *4*, 754–762. [\[CrossRef\]](http://doi.org/10.1161/CIRCHEARTFAILURE.111.962365) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21914813)
- 95. Lampsas, S.; Oikonomou, E.; Pantelidis, P.; Theofilis, P.; Grammatopoulos, K.; Marathonitis, A.; Vavuranakis, M.A.; Siasos, G.; Tousoulis, D.; Vavuranakis, M. Lipoprotein (a) Levels and Abdominal Aortic Aneurysm. A Systematic Review and Meta-analysis. *Curr. Pharm. Des.* **2022**, *28*, 3492–3499. [\[CrossRef\]](http://doi.org/10.2174/1381612829666221124110920)
- 96. Zhang, P.; Xu, X.; Hu, X.; van Deel, E.D.; Zhu, G.; Chen, Y. Inducible nitric oxide synthase deficiency protects the heart from systolic overload-induced ventricular hypertrophy and congestive heart failure. *Circ. Res.* **2007**, *100*, 1089–1098. [\[CrossRef\]](http://doi.org/10.1161/01.RES.0000264081.78659.45)
- 97. Munzel, T.; Gori, T.; Keaney, J.F., Jr.; Maack, C.; Daiber, A. Pathophysiological role of oxidative stress in systolic and diastolic heart failure and its therapeutic implications. *Eur. Heart J.* **2015**, *36*, 2555–2564. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehv305)
- 98. Ansari-Ramandi, M.M.; Maleki, M.; Alizadehasl, A.; Amin, A.; Taghavi, S.; Alemzadeh-Ansari, M.J.; Kazem Moussavi, A.; Naderi, N. Safety and effect of high dose allopurinol in patients with severe left ventricular systolic dysfunction. *J. Cardiovasc. Thorac. Res.* **2017**, *9*, 102–107. [\[CrossRef\]](http://doi.org/10.15171/jcvtr.2017.17)
- 99. Cordwin, D.J.; Berei, T.J.; Pogue, K.T. The Role of sGC Stimulators and Activators in Heart Failure With Reduced Ejection Fraction. *J. Cardiovasc. Pharmacol. Ther.* **2021**, *26*, 593–600. [\[CrossRef\]](http://doi.org/10.1177/10742484211042706)
- 100. Aimo, A.; Castiglione, V.; Vergaro, G.; Panichella, G.; Senni, M.; Lombardi, C.M.; Emdin, M. The place of vericiguat in the landscape of treatment for heart failure with reduced ejection fraction. *Heart Fail. Rev.* **2022**, *27*, 1165–1171. [\[CrossRef\]](http://doi.org/10.1007/s10741-021-10146-1)
- 101. Yin, W.H.; Chen, J.W.; Chen, Y.H.; Lin, S.J. Fenofibrate Modulates HO-1 and Ameliorates Endothelial Expression of Cell Adhesion Molecules in Systolic Heart Failure. *Acta Cardiol. Sin.* **2013**, *29*, 251–260. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27122714)
- 102. Theofilis, P.; Lampsas, S.; Oikonomou, E.; Siasos, G.; Vavuranakis, M.A.; Marinos, G.; Tsioufis, K.; Vavuranakis, M.; Tousoulis, D. Endothelial dysfunction in convalescent COVID-19 patients: Systematic review and meta-analysis. *Arch. Cardiovasc. Dis.* **2022**, *115*, 675–677. [\[CrossRef\]](http://doi.org/10.1016/j.acvd.2022.09.001) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36335024)
- 103. Tang, W.H.; Shrestha, K.; Tong, W.; Wang, Z.; Troughton, R.W.; Borowski, A.G.; Klein, A.L.; Hazen, S.L. Nitric oxide bioavailability and adiponectin production in chronic systolic heart failure: Relation to severity of cardiac dysfunction. *Transl. Res.* **2013**, *162*, 26–33. [\[CrossRef\]](http://doi.org/10.1016/j.trsl.2013.02.005) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23499315)
- 104. Bonadei, I.; Sciatti, E.; Vizzardi, E.; Fabbricatore, D.; Pagnoni, M.; Rossi, L.; Carubelli, V.; Lombardi, C.M.; Metra, M. Effects of ivabradine on endothelial function, aortic properties and ventricular-arterial coupling in chronic systolic heart failure patients. *Cardiovasc. Ther.* **2018**, *36*, e12323. [\[CrossRef\]](http://doi.org/10.1111/1755-5922.12323) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29460403)
- 105. Andrei, S.; Nguyen, M.; Longrois, D.; Popescu, B.A.; Bouhemad, B.; Guinot, P.G. Ventriculo-Arterial Coupling Is Associated With Oxygen Consumption and Tissue Perfusion in Acute Circulatory Failure. *Front. Cardiovasc. Med.* **2022**, *9*, 842554. [\[CrossRef\]](http://doi.org/10.3389/fcvm.2022.842554)
- 106. Salvador, A.M.; Moss, M.E.; Aronovitz, M.; Mueller, K.B.; Blanton, R.M.; Jaffe, I.Z.; Alcaide, P. Endothelial mineralocorticoid receptor contributes to systolic dysfunction induced by pressure overload without modulating cardiac hypertrophy or inflammation. *Physiol. Rep.* **2017**, *5*, e13313. [\[CrossRef\]](http://doi.org/10.14814/phy2.13313)
- 107. Zhang, H.; Feng, L.; Wan, Q.; Hong, Y.; Li, Y.; Cheng, G.; Han, X. Osteoprotegerin is associated with depletion of circulating endothelial progenitor cells and elevation in pulmonary arterial pressure in patients with systolic heart failure. *Acta Cardiol.* **2015**, *70*, 435–441. [\[CrossRef\]](http://doi.org/10.1080/AC.70.4.3094653)
- 108. Zhang, H.; Feng, L.; Wan, Q.L.; Hong, Y.; Li, Y.M.; Cheng, G.C.; Han, X.Q. Sleep-disordered breathing is associated with depletion of circulating endothelial progenitor cells and elevation in pulmonary arterial pressure in patients with decompensated systolic heart failure. *J. Geriatr. Cardiol.* **2015**, *12*, 424–430. [\[CrossRef\]](http://doi.org/10.11909/j.issn.1671-5411.2015.04.015)
- 109. Susic, L.; Maricic, L.; Vincelj, J.; Vadoci, M.; Susic, T. Understanding the association between endothelial dysfunction and left ventricle diastolic dysfunction in development of coronary artery disease and heart failure. *Acta Biomed.* **2021**, *92*, e2021204. [\[CrossRef\]](http://doi.org/10.23750/abm.v92i3.11495)
- 110. Gamrat, A.; Surdacki, M.A.; Chyrchel, B.; Surdacki, A. Endothelial Dysfunction: A Contributor to Adverse Cardiovascular Remodeling and Heart Failure Development in Type 2 Diabetes beyond Accelerated Atherogenesis. *J. Clin. Med.* **2020**, *9*, 2090. [\[CrossRef\]](http://doi.org/10.3390/jcm9072090)
- 111. Cauwenberghs, N.; Godderis, S.; Sabovcik, F.; Cornelissen, V.; Kuznetsova, T. Subclinical heart remodeling and dysfunction in relation to peripheral endothelial dysfunction: A general population study. *Microcirculation* **2021**, *28*, e12731. [\[CrossRef\]](http://doi.org/10.1111/micc.12731) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34569675)
- 112. Sorop, O.; Heinonen, I.; van Kranenburg, M.; van de Wouw, J.; de Beer, V.J.; Nguyen, I.T.N.; Octavia, Y.; van Duin, R.W.B.; Stam, K.; van Geuns, R.J.; et al. Multiple common comorbidities produce left ventricular diastolic dysfunction associated with coronary microvascular dysfunction, oxidative stress, and myocardial stiffening. *Cardiovasc. Res.* **2018**, *114*, 954–964. [\[CrossRef\]](http://doi.org/10.1093/cvr/cvy038) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29432575)
- 113. Abelanet, A.; Camoin, M.; Rubin, S.; Bougaran, P.; Delobel, V.; Pernot, M.; Forfar, I.; Guilbeau-Frugier, C.; Gales, C.; Bats, M.L.; et al. Increased Capillary Permeability in Heart Induces Diastolic Dysfunction Independently of Inflammation, Fibrosis, or Cardiomyocyte Dysfunction. *Arterioscler. Thromb. Vasc. Biol.* **2022**, *42*, 745–763. [\[CrossRef\]](http://doi.org/10.1161/ATVBAHA.121.317319) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35510550)
- 114. Dhot, J.; Ferron, M.; Prat, V.; Persello, A.; Roul, D.; Stevant, D.; Guijarro, D.; Piriou, N.; Aillerie, V.; Erraud, A.; et al. Overexpression of endothelial beta(3) -adrenergic receptor induces diastolic dysfunction in rats. *ESC Heart Fail.* **2020**, *7*, 4159–4171. [\[CrossRef\]](http://doi.org/10.1002/ehf2.13040) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33034410)
- 115. Erkens, R.; Kramer, C.M.; Luckstadt, W.; Panknin, C.; Krause, L.; Weidenbach, M.; Dirzka, J.; Krenz, T.; Mergia, E.; Suvorava, T.; et al. Left ventricular diastolic dysfunction in Nrf2 knock out mice is associated with cardiac hypertrophy, decreased expression of SERCA2a, and preserved endothelial function. *Free Radic. Biol. Med.* **2015**, *89*, 906–917. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2015.10.409)
- 116. Heinonen, I.; Sorop, O.; van Dalen, B.M.; Wust, R.C.I.; van de Wouw, J.; de Beer, V.J.; Octavia, Y.; van Duin, R.W.B.; Hoogstrate, Y.; Blonden, L.; et al. Cellular, mitochondrial and molecular alterations associate with early left ventricular diastolic dysfunction in a porcine model of diabetic metabolic derangement. *Sci. Rep.* **2020**, *10*, 13173. [\[CrossRef\]](http://doi.org/10.1038/s41598-020-68637-4)
- 117. Waddingham, M.T.; Sonobe, T.; Tsuchimochi, H.; Edgley, A.J.; Sukumaran, V.; Chen, Y.C.; Hansra, S.S.; Schwenke, D.O.; Umetani, K.; Aoyama, K.; et al. Diastolic dysfunction is initiated by cardiomyocyte impairment ahead of endothelial dysfunction due to increased oxidative stress and inflammation in an experimental prediabetes model. *J. Mol. Cell Cardiol.* **2019**, *137*, 119–131. [\[CrossRef\]](http://doi.org/10.1016/j.yjmcc.2019.10.005)
- 118. Gevaert, A.B.; Lemmens, K.; Vrints, C.J.; Van Craenenbroeck, E.M. Targeting Endothelial Function to Treat Heart Failure with Preserved Ejection Fraction: The Promise of Exercise Training. *Oxid. Med. Cell Longev.* **2017**, *2017*, 4865756. [\[CrossRef\]](http://doi.org/10.1155/2017/4865756)
- 119. Hirai, D.M.; Zelt, J.T.; Jones, J.H.; Castanhas, L.G.; Bentley, R.F.; Earle, W.; Staples, P.; Tschakovsky, M.E.; McCans, J.; O'Donnell, D.E.; et al. Dietary nitrate supplementation and exercise tolerance in patients with heart failure with reduced ejection fraction. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2017**, *312*, R13–R22. [\[CrossRef\]](http://doi.org/10.1152/ajpregu.00263.2016)
- 120. Mocan, M.; Mocan Hognogi, L.D.; Anton, F.P.; Chiorescu, R.M.; Goidescu, C.M.; Stoia, M.A.; Farcas, A.D. Biomarkers of Inflammation in Left Ventricular Diastolic Dysfunction. *Dis. Markers* **2019**, *2019*, 7583690. [\[CrossRef\]](http://doi.org/10.1155/2019/7583690) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31275453)
- 121. Kloch, M.; Stolarz-Skrzypek, K.; Olszanecka, A.; Wojciechowska, W.; Bednarski, A.; Stefaniak, J.; Czarnecka, D. Inflammatory markers and left ventricular diastolic dysfunction in a family-based population study. *Kardiol. Pol.* **2019**, *77*, 33–39. [\[CrossRef\]](http://doi.org/10.5603/KP.a2018.0214) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30406940)
- 122. Zeng, H.; He, X.; Chen, J.X. A Sex-Specific Role of Endothelial Sirtuin 3 on Blood Pressure and Diastolic Dysfunction in Female Mice. *Int. J. Mol. Sci.* **2020**, *21*, 9744. [\[CrossRef\]](http://doi.org/10.3390/ijms21249744) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33371209)
- 123. Teuber, J.P.; Essandoh, K.; Hummel, S.L.; Madamanchi, N.R.; Brody, M.J. NADPH Oxidases in Diastolic Dysfunction and Heart Failure with Preserved Ejection Fraction. *Antioxidants* **2022**, *11*, 1822. [\[CrossRef\]](http://doi.org/10.3390/antiox11091822) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36139898)
- 124. Yin, J.; Kukucka, M.; Hoffmann, J.; Sterner-Kock, A.; Burhenne, J.; Haefeli, W.E.; Kuppe, H.; Kuebler, W.M. Sildenafil preserves lung endothelial function and prevents pulmonary vascular remodeling in a rat model of diastolic heart failure. *Circ. Heart Fail.* **2011**, *4*, 198–206. [\[CrossRef\]](http://doi.org/10.1161/CIRCHEARTFAILURE.110.957050)
- 125. Serpi, R.; Tolonen, A.M.; Huusko, J.; Rysa, J.; Tenhunen, O.; Yla-Herttuala, S.; Ruskoaho, H. Vascular endothelial growth factor-B gene transfer prevents angiotensin II-induced diastolic dysfunction via proliferation and capillary dilatation in rats. *Cardiovasc. Res.* **2011**, *89*, 204–213. [\[CrossRef\]](http://doi.org/10.1093/cvr/cvq267)
- 126. Sama, I.E.; Woolley, R.J.; Nauta, J.F.; Romaine, S.P.R.; Tromp, J.; Ter Maaten, J.M.; van der Meer, P.; Lam, C.S.P.; Samani, N.J.; Ng, L.L.; et al. A network analysis to identify pathophysiological pathways distinguishing ischaemic from non-ischaemic heart failure. *Eur. J. Heart Fail.* **2020**, *22*, 821–833. [\[CrossRef\]](http://doi.org/10.1002/ejhf.1811)
- 127. Rubattu, S.; Bigatti, G.; Evangelista, A.; Lanzani, C.; Stanzione, R.; Zagato, L.; Manunta, P.; Marchitti, S.; Venturelli, V.; Bianchi, G.; et al. Association of atrial natriuretic peptide and type a natriuretic peptide receptor gene polymorphisms with left ventricular mass in human essential hypertension. *J. Am. Coll Cardiol.* **2006**, *48*, 499–505. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2005.12.081)
- 128. Shantsila, E.; Wrigley, B.J.; Blann, A.D.; Gill, P.S.; Lip, G.Y. A contemporary view on endothelial function in heart failure. *Eur. J. Heart Fail.* **2012**, *14*, 873–881. [\[CrossRef\]](http://doi.org/10.1093/eurjhf/hfs066)
- 129. Klosinska, M.; Rudzinski, T.; Grzelak, P.; Stefanczyk, L.; Drozdz, J.; Krzeminska-Pakula, M. Endothelium-dependent and -independent vasodilation is more attenuated in ischaemic than in non-ischaemic heart failure. *Eur. J. Heart Fail.* **2009**, *11*, 765–770. [\[CrossRef\]](http://doi.org/10.1093/eurjhf/hfp091)
- 130. Tsigkou, V.; Siasos, G.; Oikonomou, E.; Zaromitidou, M.; Mourouzis, K.; Dimitropoulos, S.; Bletsa, E.; Gouliopoulos, N.; Stampouloglou, P.K.; Panoilia, M.E.; et al. The prognostic role of galectin-3 and endothelial function in patients with heart failure. *Cardiol. J.* **2022**. [\[CrossRef\]](http://doi.org/10.5603/CJ.a2022.0074)
- 131. Molitor, M.; Rudi, W.S.; Garlapati, V.; Finger, S.; Schuler, R.; Kossmann, S.; Lagrange, J.; Nguyen, T.S.; Wild, J.; Knopp, T.; et al. Nox2+ myeloid cells drive vascular inflammation and endothelial dysfunction in heart failure after myocardial infarction via angiotensin II receptor type 1. *Cardiovasc. Res.* **2021**, *117*, 162–177. [\[CrossRef\]](http://doi.org/10.1093/cvr/cvaa042) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32077922)
- 132. Isaksen, K.; Halvorsen, B.; Munk, P.S.; Aukrust, P.; Larsen, A.I. Effects of interval training on inflammatory biomarkers in patients with ischemic heart failure. *Scand. Cardiovasc. J.* **2019**, *53*, 213–219. [\[CrossRef\]](http://doi.org/10.1080/14017431.2019.1629004)
- 133. Dimitropoulos, S.; Mystakidi, V.C.; Oikonomou, E.; Siasos, G.; Tsigkou, V.; Athanasiou, D.; Gouliopoulos, N.; Bletsa, E.; Kalampogias, A.; Charalambous, G.; et al. Association of Soluble Suppression of Tumorigenesis-2 (ST2) with Endothelial Function in Patients with Ischemic Heart Failure. *Int. J. Mol. Sci.* **2020**, *21*, 9385. [\[CrossRef\]](http://doi.org/10.3390/ijms21249385) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33317161)
- 134. Tentolouris, C.; Tousoulis, D.; Antoniades, C.; Bosinakou, E.; Kotsopoulou, M.; Trikas, A.; Toutouzas, P.; Stefanadis, C. Endothelial function and proinflammatory cytokines in patients with ischemic heart disease and dilated cardiomyopathy. *Int. J. Cardiol.* **2004**, *94*, 301–305. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2003.08.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15093997)
- 135. Hsu, C.P.; Lin, S.J.; Chung, M.Y.; Lu, T.M. Asymmetric dimethylarginine predicts clinical outcomes in ischemic chronic heart failure. *Atherosclerosis* **2012**, *225*, 504–510. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2012.09.040) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23092827)
- 136. Zhou, H.; Wang, B.; Yang, Y.; Jia, Q.; Qi, Z.; Zhang, A.; Lv, S.; Zhang, J. Exosomes in ischemic heart disease: Novel carriers for bioinformation. *Biomed. Pharmacother.* **2019**, *120*, 109451. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2019.109451)
- 137. Oikonomou, E.; Theofilis, P.; Lampsas, S.; Katsarou, O.; Kalogeras, K.; Marinos, G.; Tsatsaragkou, A.; Anastasiou, A.; Lysandrou, A.; Gounaridi, M.I.; et al. Current Concepts and Future Applications of Non-Invasive Functional and Anatomical Evaluation of Coronary Artery Disease. *Life* **2022**, *12*, 1803. [\[CrossRef\]](http://doi.org/10.3390/life12111803)
- 138. Oie, E.; Vinge, L.E.; Yndestad, A.; Sandberg, C.; Grogaard, H.K.; Attramadal, H. Induction of a myocardial adrenomedullin signaling system during ischemic heart failure in rats. *Circulation* **2000**, *101*, 415–422. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.101.4.415)
- 139. Bansal, S.S.; Ismahil, M.A.; Goel, M.; Zhou, G.; Rokosh, G.; Hamid, T.; Prabhu, S.D. Dysfunctional and Proinflammatory Regulatory T-Lymphocytes Are Essential for Adverse Cardiac Remodeling in Ischemic Cardiomyopathy. *Circulation* **2019**, *139*, 206–221. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.118.036065)
- 140. Gueret, A.; Harouki, N.; Favre, J.; Galmiche, G.; Nicol, L.; Henry, J.P.; Besnier, M.; Thuillez, C.; Richard, V.; Kolkhof, P.; et al. Vascular Smooth Muscle Mineralocorticoid Receptor Contributes to Coronary and Left Ventricular Dysfunction After Myocardial Infarction. *Hypertension* **2016**, *67*, 717–723. [\[CrossRef\]](http://doi.org/10.1161/HYPERTENSIONAHA.115.06709)
- 141. Abdel Hamid, M.; Bakhoum, S.W.; Sharaf, Y.; Sabry, D.; El-Gengehe, A.T.; Abdel-Latif, A. Circulating Endothelial Cells and Endothelial Function Predict Major Adverse Cardiac Events and Early Adverse Left Ventricular Remodeling in Patients With ST-Segment Elevation Myocardial Infarction. *J. Interv. Cardiol.* **2016**, *29*, 89–98. [\[CrossRef\]](http://doi.org/10.1111/joic.12269) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26864952)
- 142. Berezin, A.E.; Kremzer, A.A. Circulating endothelial progenitor cells as markers for severity of ischemic chronic heart failure. *J. Card. Fail.* **2014**, *20*, 438–447. [\[CrossRef\]](http://doi.org/10.1016/j.cardfail.2014.02.009) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24594088)
- 143. Berezin, A.E.; Kremzer, A.A.; Samura, T.A.; Martovitskaya, Y.V. Circulating endothelial-derived apoptotic microparticles in the patients with ischemic symptomatic chronic heart failure: Relevance of pro-inflammatory activation and outcomes. *Int. Cardiovasc. Res. J.* **2014**, *8*, 116–123.
- 144. Ponikowski, P.; Voors, A.A.; Anker, S.D.; Bueno, H.; Cleland, J.G.F.; Coats, A.J.S.; Falk, V.; Gonzalez-Juanatey, J.R.; Harjola, V.P.; Jankowska, E.A.; et al. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC)Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur. Heart J.* **2016**, *37*, 2129–2200. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehw128) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27206819)
- 145. Shah, A.; Gkaliagkousi, E.; Ritter, J.M.; Ferro, A. Endothelial function and arterial compliance are not impaired in subjects with heart failure of non-ischemic origin. *J. Card. Fail.* **2010**, *16*, 114–120. [\[CrossRef\]](http://doi.org/10.1016/j.cardfail.2009.10.019) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20142022)
- 146. Chen, J.W.; Ting, C.T.; Chen, Y.H.; Wu, T.C.; Hsu, N.W.; Lin, S.J.; Chang, M.S. Differential coronary microvascular function in patients with left ventricular dysfunction of unknown cause–implication for possible mechanism of myocardial ischemia in early stage of cardiomyopathy. *Int. J. Cardiol.* **1999**, *69*, 251–261. [\[CrossRef\]](http://doi.org/10.1016/S0167-5273(99)00042-X) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10402108)
- 147. Oikonomou, E.; Souvaliotis, N.; Lampsas, S.; Siasos, G.; Theofilis, P.; Korakas, E.; Lambadiari, V.; Ikonomidis, I.; Pesiridis, T.; Zakynthinos, G.; et al. The role of cardiometabolic risk factors and endothelial dysfunction in serum albumin levels of patients with COVID-19. *Cardiol. J.* **2022**, *29*, 1037–1039. [\[CrossRef\]](http://doi.org/10.5603/CJ.a2022.0088) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36117293)
- 148. Hirakawa, K.; Yamamuro, M.; Uemura, T.; Takashio, S.; Kaikita, K.; Utsunomiya, D.; Nakayama, M.; Yamamoto, E.; Yamashita, Y.; Hokimoto, S.; et al. Correlation between microvascular dysfunction and B-type natriuretic peptide levels in non-ischemic heart failure patients with cardiac fibrosis. *Int. J. Cardiol.* **2017**, *228*, 881–885. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2016.11.054)
- 149. Knapp, M.; Tu, X.; Wu, R. Vascular endothelial dysfunction, a major mediator in diabetic cardiomyopathy. *Acta Pharmacol. Sin.* **2019**, *40*, 1–8. [\[CrossRef\]](http://doi.org/10.1038/s41401-018-0042-6)
- 150. Reichman-Warmusz, E.; Brzozowa-Zasada, M.; Wojciechowska, C.; Dudek, D.; Warmusz, O.; Wojnicz, R. Decreased immunoreactivity of von Willebrand factor may reflect persistent nature of the endothelial dysfunction in non-ischemic heart failure. *Folia Histochem. Cytobiol.* **2021**, *59*, 108–113. [\[CrossRef\]](http://doi.org/10.5603/FHC.a2021.0012)
- 151. Thorin, E. Different contribution of endothelial nitric oxide in the relaxation of human coronary arteries of ischemic and dilated cardiomyopathic hearts. *J. Cardiovasc. Pharmacol.* **2001**, *37*, 227–232. [\[CrossRef\]](http://doi.org/10.1097/00005344-200102000-00010) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11210005)
- 152. Vittorio, T.J.; Zolty, R.; Garg, P.K.; Sarswat, N.; Tseng, C.H.; Jorde, U.P.; Colombo, P.C. Interdependence of cardiac and endothelial function in patients with symptomatic chronic heart failure of nonischemic etiology. *Echocardiography* **2009**, *26*, 916–921. [\[CrossRef\]](http://doi.org/10.1111/j.1540-8175.2009.00907.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19486111)
- 153. Yoshida, H.; Nakamura, M.; Akatsu, T.; Arakawa, N.; Hiramori, K. Effects of nitric oxide inhibition on basal forearm blood flow in patients with nonischemic chronic heart failure. *Heart Vessels* **1998**, *13*, 142–146. [\[CrossRef\]](http://doi.org/10.1007/BF01747831) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10328184)
- 154. Tuder, R.M.; Archer, S.L.; Dorfmuller, P.; Erzurum, S.C.; Guignabert, C.; Michelakis, E.; Rabinovitch, M.; Schermuly, R.; Stenmark, K.R.; Morrell, N.W. Relevant issues in the pathology and pathobiology of pulmonary hypertension. *J. Am. Coll. Cardiol.* **2013**, *62*, D4–D12. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2013.10.025) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24355640)
- 155. Ranchoux, B.; Harvey, L.D.; Ayon, R.J.; Babicheva, A.; Bonnet, S.; Chan, S.Y.; Yuan, J.X.; Perez, V.J. Endothelial dysfunction in pulmonary arterial hypertension: An evolving landscape (2017 Grover Conference Series). *Pulm. Circ.* **2018**, *8*, 2045893217752912. [\[CrossRef\]](http://doi.org/10.1177/2045893217752912) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29283043)
- 156. Southgate, L.; Machado, R.D.; Graf, S.; Morrell, N.W. Molecular genetic framework underlying pulmonary arterial hypertension. *Nat. Rev. Cardiol.* **2020**, *17*, 85–95. [\[CrossRef\]](http://doi.org/10.1038/s41569-019-0242-x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31406341)
- 157. Santos-Gomes, J.; Le Ribeuz, H.; Bras-Silva, C.; Antigny, F.; Adao, R. Role of Ion Channel Remodeling in Endothelial Dysfunction Induced by Pulmonary Arterial Hypertension. *Biomolecules* **2022**, *12*, 484. [\[CrossRef\]](http://doi.org/10.3390/biom12040484)
- 158. Friedman, D.; Szmuszkovicz, J.; Rabai, M.; Detterich, J.A.; Menteer, J.; Wood, J.C. Systemic endothelial dysfunction in children with idiopathic pulmonary arterial hypertension correlates with disease severity. *J. Heart Lung Transplant.* **2012**, *31*, 642–647. [\[CrossRef\]](http://doi.org/10.1016/j.healun.2012.02.020)
- 159. Loscalzo, J. Endothelial dysfunction in pulmonary hypertension. *N. Engl. J. Med.* **1992**, *327*, 117–119. [\[CrossRef\]](http://doi.org/10.1056/NEJM199207093270209)
- 160. Huertas, A.; Perros, F.; Tu, L.; Cohen-Kaminsky, S.; Montani, D.; Dorfmuller, P.; Guignabert, C.; Humbert, M. Immune dysregulation and endothelial dysfunction in pulmonary arterial hypertension: A complex interplay. *Circulation* **2014**, *129*, 1332–1340. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.113.004555)
- 161. Levy, M.; Maurey, C.; Celermajer, D.S.; Vouhe, P.R.; Danel, C.; Bonnet, D.; Israel-Biet, D. Impaired apoptosis of pulmonary endothelial cells is associated with intimal proliferation and irreversibility of pulmonary hypertension in congenital heart disease. *J. Am. Coll. Cardiol.* **2007**, *49*, 803–810. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2006.09.049) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17306711)
- 162. Zhou, C.; Townsley, M.I.; Alexeyev, M.; Voelkel, N.F.; Stevens, T. Endothelial hyperpermeability in severe pulmonary arterial hypertension: Role of store-operated calcium entry. *Am. J. Physiol. Lung Cell Mol. Physiol.* **2016**, *311*, L560–L569. [\[CrossRef\]](http://doi.org/10.1152/ajplung.00057.2016) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27422996)
- 163. Dai, Z.; Zhu, M.M.; Peng, Y.; Machireddy, N.; Evans, C.E.; Machado, R.; Zhang, X.; Zhao, Y.Y. Therapeutic Targeting of Vascular Remodeling and Right Heart Failure in Pulmonary Arterial Hypertension with a HIF-2alpha Inhibitor. *Am. J. Respir. Crit. Care Med.* **2018**, *198*, 1423–1434. [\[CrossRef\]](http://doi.org/10.1164/rccm.201710-2079OC)
- 164. Morrell, N.W.; Adnot, S.; Archer, S.L.; Dupuis, J.; Lloyd Jones, P.; MacLean, M.R.; McMurtry, I.F.; Stenmark, K.R.; Thistlethwaite, P.A.; Weissmann, N.; et al. Cellular and molecular basis of pulmonary arterial hypertension. *J. Am. Coll. Cardiol.* **2009**, *54*, S20–S31. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2009.04.018) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19555855)
- 165. Rabinovitch, M.; Guignabert, C.; Humbert, M.; Nicolls, M.R. Inflammation and immunity in the pathogenesis of pulmonary arterial hypertension. *Circ. Res.* **2014**, *115*, 165–175. [\[CrossRef\]](http://doi.org/10.1161/CIRCRESAHA.113.301141) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24951765)
- 166. Stenmark, K.R.; Yeager, M.E.; El Kasmi, K.C.; Nozik-Grayck, E.; Gerasimovskaya, E.V.; Li, M.; Riddle, S.R.; Frid, M.G. The adventitia: Essential regulator of vascular wall structure and function. *Annu. Rev. Physiol.* **2013**, *75*, 23–47. [\[CrossRef\]](http://doi.org/10.1146/annurev-physiol-030212-183802)
- 167. Frid, M.G.; Kale, V.A.; Stenmark, K.R. Mature vascular endothelium can give rise to smooth muscle cells via endothelialmesenchymal transdifferentiation: In vitro analysis. *Circ. Res.* **2002**, *90*, 1189–1196. [\[CrossRef\]](http://doi.org/10.1161/01.RES.0000021432.70309.28)
- 168. Cober, N.D.; VandenBroek, M.M.; Ormiston, M.L.; Stewart, D.J. Evolving Concepts in Endothelial Pathobiology of Pulmonary Arterial Hypertension. *Hypertension* **2022**, *79*, 1580–1590. [\[CrossRef\]](http://doi.org/10.1161/HYPERTENSIONAHA.122.18261)
- 169. Wermuth, P.J.; Li, Z.; Mendoza, F.A.; Jimenez, S.A. Stimulation of Transforming Growth Factor-beta1-Induced Endothelial-To-Mesenchymal Transition and Tissue Fibrosis by Endothelin-1 (ET-1): A Novel Profibrotic Effect of ET-1. *PLoS ONE* **2016**, *11*, e0161988. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0161988)
- 170. Guo, Y.; Li, P.; Bledsoe, G.; Yang, Z.R.; Chao, L.; Chao, J. Kallistatin inhibits TGF-beta-induced endothelial-mesenchymal transition by differential regulation of microRNA-21 and eNOS expression. *Exp. Cell Res.* **2015**, *337*, 103–110. [\[CrossRef\]](http://doi.org/10.1016/j.yexcr.2015.06.021)
- 171. Boucherat, O.; Potus, F.; Bonnet, S. microRNA and Pulmonary Hypertension. *Adv. Exp. Med. Biol.* **2015**, *888*, 237–252. [\[CrossRef\]](http://doi.org/10.1007/978-3-319-22671-2_12) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26663186)
- 172. Aldred, M.A.; Vijayakrishnan, J.; James, V.; Soubrier, F.; Gomez-Sanchez, M.A.; Martensson, G.; Galie, N.; Manes, A.; Corris, P.; Simonneau, G.; et al. BMPR2 gene rearrangements account for a significant proportion of mutations in familial and idiopathic pulmonary arterial hypertension. *Hum. Mutat.* **2006**, *27*, 212–213. [\[CrossRef\]](http://doi.org/10.1002/humu.9398) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16429403)
- 173. Gomez-Puerto, M.C.; van Zuijen, I.; Huang, C.J.; Szulcek, R.; Pan, X.; van Dinther, M.A.; Kurakula, K.; Wiesmeijer, C.C.; Goumans, M.J.; Bogaard, H.J.; et al. Autophagy contributes to BMP type 2 receptor degradation and development of pulmonary arterial hypertension. *J. Pathol.* **2019**, *249*, 356–367. [\[CrossRef\]](http://doi.org/10.1002/path.5322) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31257577)
- 174. Han, Y.; Ali, M.K.; Dua, K.; Spiekerkoetter, E.; Mao, Y. Role of Long Non-Coding RNAs in Pulmonary Arterial Hypertension. *Cells* **2021**, *10*, 1892. [\[CrossRef\]](http://doi.org/10.3390/cells10081892) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34440661)
- 175. Xu, D.; Hu, Y.H.; Gou, X.; Li, F.Y.; Yang, X.Y.; Li, Y.M.; Chen, F. Oxidative Stress and Antioxidative Therapy in Pulmonary Arterial Hypertension. *Molecules* **2022**, *27*, 3724. [\[CrossRef\]](http://doi.org/10.3390/molecules27123724) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35744848)
- 176. Ryan, J.J.; Archer, S.L. Emerging concepts in the molecular basis of pulmonary arterial hypertension: Part I: Metabolic plasticity and mitochondrial dynamics in the pulmonary circulation and right ventricle in pulmonary arterial hypertension. *Circulation* **2015**, *131*, 1691–1702. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.114.006979) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25964279)
- 177. Cuthbertson, I.; Morrell, N.W.; Caruso, P. BMPR2 Mutation and Metabolic Reprogramming in Pulmonary Arterial Hypertension. *Circ. Res.* **2023**, *132*, 109–126. [\[CrossRef\]](http://doi.org/10.1161/CIRCRESAHA.122.321554) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36603064)
- 178. Mumby, S.; Perros, F.; Hui, C.; Xu, B.L.; Xu, W.; Elyasigomari, V.; Hautefort, A.; Manaud, G.; Humbert, M.; Chung, K.F.; et al. Extracellular matrix degradation pathways and fatty acid metabolism regulate distinct pulmonary vascular cell types in pulmonary arterial hypertension. *Pulm. Circ.* **2021**, *11*, 2045894021996190. [\[CrossRef\]](http://doi.org/10.1177/2045894021996190)
- 179. Kobayashi, T.; Kim, J.D.; Naito, A.; Yanagisawa, A.; Jujo-Sanada, T.; Kasuya, Y.; Nakagawa, Y.; Sakao, S.; Tatsumi, K.; Suzuki, T. Multi-omics analysis of right ventricles in rat models of pulmonary arterial hypertension: Consideration of mitochondrial biogenesis by chrysin. *Int. J. Mol. Med.* **2022**, *49*, 1–12. [\[CrossRef\]](http://doi.org/10.3892/ijmm.2022.5124)
- 180. Legchenko, E.; Chouvarine, P.; Borchert, P.; Fernandez-Gonzalez, A.; Snay, E.; Meier, M.; Maegel, L.; Mitsialis, S.A.; Rog-Zielinska, E.A.; Kourembanas, S.; et al. PPARgamma agonist pioglitazone reverses pulmonary hypertension and prevents right heart failure via fatty acid oxidation. *Sci. Transl. Med.* **2018**, *10*, eaao0303. [\[CrossRef\]](http://doi.org/10.1126/scitranslmed.aao0303)
- 181. Omura, J.; Satoh, K.; Kikuchi, N.; Satoh, T.; Kurosawa, R.; Nogi, M.; Ohtsuki, T.; Al-Mamun, M.E.; Siddique, M.A.H.; Yaoita, N.; et al. ADAMTS8 Promotes the Development of Pulmonary Arterial Hypertension and Right Ventricular Failure: A Possible Novel Therapeutic Target. *Circ. Res.* **2019**, *125*, 884–906. [\[CrossRef\]](http://doi.org/10.1161/CIRCRESAHA.119.315398)
- 182. Zuchi, C.; Tritto, I.; Carluccio, E.; Mattei, C.; Cattadori, G.; Ambrosio, G. Role of endothelial dysfunction in heart failure. *Heart Fail. Rev.* **2020**, *25*, 21–30. [\[CrossRef\]](http://doi.org/10.1007/s10741-019-09881-3)
- 183. Bank, A.J.; Lee, P.C.; Kubo, S.H. Endothelial dysfunction in patients with heart failure: Relationship to disease severity. *J. Card. Fail.* **2000**, *6*, 29–36. [\[CrossRef\]](http://doi.org/10.1016/S1071-9164(00)00009-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10746816)
- 184. Popovic, B.; Zannad, F.; Louis, H.; Clerc-Urmes, I.; Lakomy, C.; Gibot, S.; Denis, C.V.; Lacolley, P.; Regnault, V. Endothelial-driven increase in plasma thrombin generation characterising a new hypercoagulable phenotype in acute heart failure. *Int. J. Cardiol.* **2019**, *274*, 195–201. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2018.07.130) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30089551)
- 185. Heitzer, T.; Baldus, S.; von Kodolitsch, Y.; Rudolph, V.; Meinertz, T. Systemic endothelial dysfunction as an early predictor of adverse outcome in heart failure. *Arterioscler. Thromb. Vasc. Biol.* **2005**, *25*, 1174–1179. [\[CrossRef\]](http://doi.org/10.1161/01.ATV.0000166516.52477.81) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15831810)
- 186. Katz, S.D.; Hryniewicz, K.; Hriljac, I.; Balidemaj, K.; Dimayuga, C.; Hudaihed, A.; Yasskiy, A. Vascular endothelial dysfunction and mortality risk in patients with chronic heart failure. *Circulation* **2005**, *111*, 310–314. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.0000153349.77489.CF)
- 187. Akar, J.G.; Al-Chekakie, M.O.; Fugate, T.; Moran, L.; Froloshki, B.; Varma, N.; Santucci, P.; Wilber, D.J.; Matsumura, M.E. Endothelial dysfunction in heart failure identifies responders to cardiac resynchronization therapy. *Heart Rhythm.* **2008**, *5*, 1229–1235. [\[CrossRef\]](http://doi.org/10.1016/j.hrthm.2008.05.027)
- 188. Suhrs, H.E.; Schroder, J.; Bove, K.B.; Mygind, N.D.; Frestad, D.; Michelsen, M.M.; Lange, T.; Gustafsson, I.; Kastrup, J.; Prescott, E. Inflammation, non-endothelial dependent coronary microvascular function and diastolic function-Are they linked? *PLoS ONE* **2020**, *15*, e0236035. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0236035)
- 189. Hage, C.; Michaelsson, E.; Kull, B.; Miliotis, T.; Svedlund, S.; Linde, C.; Donal, E.; Daubert, J.C.; Gan, L.M.; Lund, L.H. Myeloperoxidase and related biomarkers are suggestive footprints of endothelial microvascular inflammation in HFpEF patients. *ESC Heart Fail.* **2020**, *7*, 1534–1546. [\[CrossRef\]](http://doi.org/10.1002/ehf2.12700)
- 190. Chong, A.Y.; Freestone, B.; Patel, J.; Lim, H.S.; Hughes, E.; Blann, A.D.; Lip, G.Y. Endothelial activation, dysfunction, and damage in congestive heart failure and the relation to brain natriuretic peptide and outcomes. *Am. J. Cardiol.* **2006**, *97*, 671–675. [\[CrossRef\]](http://doi.org/10.1016/j.amjcard.2005.09.113)
- 191. Maio, R.; Perticone, M.; Suraci, E.; Sciacqua, A.; Sesti, G.; Perticone, F. Endothelial dysfunction and C-reactive protein predict the incidence of heart failure in hypertensive patients. *ESC Heart Fail.* **2021**, *8*, 399–407. [\[CrossRef\]](http://doi.org/10.1002/ehf2.13088) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33236853)
- 192. Anastasiou, M.; Oikonomou, E.; Zagouri, F.; Siasos, G.; Antonopoulos, A.S.; Psaltopoulou, T.; Bamias, A.; Dimopoulos, M.A.; Tousoulis, D. Flow-Mediated Dilation of Brachial Artery as a Screening Tool for Anthracycline-Induced Cardiotoxicity. *J. Am. Coll. Cardiol.* **2017**, *70*, 3072. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2017.09.1140) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29241496)
- 193. Keteyian, S.J.; Michaels, A. Heart Failure in Cardiac Rehabilitation: A REVIEW AND PRACTICAL CONSIDERATIONS. *J. Cardiopulm. Rehabil. Prev.* **2022**, *42*, 296–303. [\[CrossRef\]](http://doi.org/10.1097/HCR.0000000000000713) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35836338)
- 194. Dickstein, K.; Cohen-Solal, A.; Filippatos, G.; McMurray, J.J.; Ponikowski, P.; Poole-Wilson, P.A.; Stromberg, A.; van Veldhuisen, D.J.; Atar, D.; Hoes, A.W.; et al. ESC guidelines for the diagnosis and treatment of acute and chronic heart failure 2008: The Task Force for the diagnosis and treatment of acute and chronic heart failure 2008 of the European Society of Cardiology. Developed in collaboration with the Heart Failure Association of the ESC (HFA) and endorsed by the European Society of Intensive Care Medicine (ESICM). *Eur. J. Heart Fail.* **2008**, *10*, 933–989. [\[CrossRef\]](http://doi.org/10.1016/j.ejheart.2008.08.005)
- 195. Leggio, M.; Fusco, A.; Loreti, C.; Limongelli, G.; Bendini, M.G.; Mazza, A.; Coraci, D.; Padua, L. Effects of exercise training in heart failure with preserved ejection fraction: An updated systematic literature review. *Heart Fail. Rev.* **2020**, *25*, 703–711. [\[CrossRef\]](http://doi.org/10.1007/s10741-019-09841-x)
- 196. Piepoli, M.F.; Conraads, V.; Corra, U.; Dickstein, K.; Francis, D.P.; Jaarsma, T.; McMurray, J.; Pieske, B.; Piotrowicz, E.; Schmid, J.P.; et al. Exercise training in heart failure: From theory to practice. A consensus document of the Heart Failure Association and the European Association for Cardiovascular Prevention and Rehabilitation. *Eur. J. Heart Fail.* **2011**, *13*, 347–357. [\[CrossRef\]](http://doi.org/10.1093/eurjhf/hfr017)
- 197. Pearson, M.J.; Smart, N.A. Effect of exercise training on endothelial function in heart failure patients: A systematic review meta-analysis. *Int. J. Cardiol.* **2017**, *231*, 234–243. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2016.12.145)
- 198. Sandri, M.; Viehmann, M.; Adams, V.; Rabald, K.; Mangner, N.; Hollriegel, R.; Lurz, P.; Erbs, S.; Linke, A.; Kirsch, K.; et al. Chronic heart failure and aging—Effects of exercise training on endothelial function and mechanisms of endothelial regeneration: Results from the Leipzig Exercise Intervention in Chronic heart failure and Aging (LEICA) study. *Eur. J. Prev. Cardiol.* **2016**, *23*, 349–358. [\[CrossRef\]](http://doi.org/10.1177/2047487315588391)
- 199. Chen, J.; Gu, S.; Song, Y.; Ji, X.; Zeng, W.; Wang, X.; Wang, Y.; Feng, Q. The impact of cardiomotor rehabilitation on endothelial function in elderly patients with chronic heart failure. *BMC Cardiovasc. Disord.* **2021**, *21*, 524. [\[CrossRef\]](http://doi.org/10.1186/s12872-021-02327-5)
- 200. Sales, A.R.K.; Azevedo, L.F.; Silva, T.O.C.; Rodrigues, A.G.; Oliveira, P.A.; Jordao, C.P.; Andrade, A.C.M.; Urias, U.; Guimaraes, G.V.; Bocchi, E.A.; et al. High-Intensity Interval Training Decreases Muscle Sympathetic Nerve Activity and Improves Peripheral Vascular Function in Patients With Heart Failure With Reduced Ejection Fraction. *Circ. Heart Fail.* **2020**, *13*, e007121. [\[CrossRef\]](http://doi.org/10.1161/CIRCHEARTFAILURE.120.007121)
- 201. Jaconiano, E.; Moreira-Goncalves, D. Unveiling the role of exercise training in targeting the inflammatory paradigm of heart failure with preserved ejection fraction: A narrative review. *Heart Fail. Rev.* **2022**, *27*, 163–190. [\[CrossRef\]](http://doi.org/10.1007/s10741-021-10138-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34244870)
- 202. Kitzman, D.W.; Brubaker, P.H.; Herrington, D.M.; Morgan, T.M.; Stewart, K.P.; Hundley, W.G.; Abdelhamed, A.; Haykowsky, M.J. Effect of endurance exercise training on endothelial function and arterial stiffness in older patients with heart failure and preserved ejection fraction: A randomized, controlled, single-blind trial. *J. Am. Coll. Cardiol.* **2013**, *62*, 584–592. [\[CrossRef\]](http://doi.org/10.1016/j.jacc.2013.04.033) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23665370)
- 203. Hibbert, B.; Simard, T.; Ramirez, F.D.; Pourdjabbar, A.; Raizman, J.E.; Maze, R.; Wilson, K.R.; Hawken, S.; O'Brien, E.R. The effect of statins on circulating endothelial progenitor cells in humans: A systematic review. *J. Cardiovasc. Pharmacol.* **2013**, *62*, 491–496. [\[CrossRef\]](http://doi.org/10.1097/FJC.0b013e3182a4027f)
- 204. Zou, G.; Song, L.; Chang, L.; Wang, H. Comparison and analysis of statins drug use in the treatment of diastolic dysfunction in patients. *Pak. J. Pharm. Sci.* **2018**, *31*, 1725–1730. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30203770)
- 205. Oikonomou, E.; Siasos, G.; Zaromitidou, M.; Hatzis, G.; Mourouzis, K.; Chrysohoou, C.; Zisimos, K.; Mazaris, S.; Tourikis, P.; Athanasiou, D.; et al. Atorvastatin treatment improves endothelial function through endothelial progenitor cells mobilization in ischemic heart failure patients. *Atherosclerosis* **2015**, *238*, 159–164. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2014.12.014) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25525743)
- 206. Tousoulis, D.; Andreou, I.; Tsiatas, M.; Miliou, A.; Tentolouris, C.; Siasos, G.; Papageorgiou, N.; Papadimitriou, C.A.; Dimopoulos, M.A.; Stefanadis, C. Effects of rosuvastatin and allopurinol on circulating endothelial progenitor cells in patients with congestive heart failure: The impact of inflammatory process and oxidative stress. *Atherosclerosis* **2011**, *214*, 151–157. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2010.11.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21122851)
- 207. Erbs, S.; Beck, E.B.; Linke, A.; Adams, V.; Gielen, S.; Krankel, N.; Mobius-Winkler, S.; Hollriegel, R.; Thiele, H.; Hambrecht, R.; et al. High-dose rosuvastatin in chronic heart failure promotes vasculogenesis, corrects endothelial function, and improves cardiac remodeling–results from a randomized, double-blind, and placebo-controlled study. *Int. J. Cardiol.* **2011**, *146*, 56–63. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2010.02.019)
- 208. Tousoulis, D.; Antoniades, C.; Bosinakou, E.; Kotsopoulou, M.; Pitsavos, C.; Vlachopoulos, C.; Panagiotakos, D.; Stefanadis, C. Effects of atorvastatin on reactive hyperemia and inflammatory process in patients with congestive heart failure. *Atherosclerosis* **2005**, *178*, 359–363. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2004.08.037)
- 209. Tousoulis, D.; Oikonomou, E.; Siasos, G.; Chrysohoou, C.; Zaromitidou, M.; Kioufis, S.; Maniatis, K.; Dilaveris, P.; Miliou, A.; Michalea, S.; et al. Dose-dependent effects of short term atorvastatin treatment on arterial wall properties and on indices of left ventricular remodeling in ischemic heart failure. *Atherosclerosis* **2013**, *227*, 367–372. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2013.01.015)
- 210. Gounari, P.; Tousoulis, D.; Antoniades, C.; Kampoli, A.M.; Stougiannos, P.; Papageorgiou, N.; Roulia, G.; Stefanadi, E.; Siasos, G.; Tsioufis, C.; et al. Rosuvastatin but not ezetimibe improves endothelial function in patients with heart failure, by mechanisms independent of lipid lowering. *Int. J. Cardiol.* **2010**, *142*, 87–91. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2008.12.067)
- 211. do Vale, G.T.; Ceron, C.S.; Gonzaga, N.A.; Simplicio, J.A.; Padovan, J.C. Three Generations of beta-blockers: History, Class Differences and Clinical Applicability. *Curr. Hypertens. Rev.* **2019**, *15*, 22–31. [\[CrossRef\]](http://doi.org/10.2174/1573402114666180918102735) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30227820)
- 212. Peller, M.; Ozieranski, K.; Balsam, P.; Grabowski, M.; Filipiak, K.J.; Opolski, G. Influence of beta-blockers on endothelial function: A meta-analysis of randomized controlled trials. *Cardiol. J.* **2015**, *22*, 708–716. [\[CrossRef\]](http://doi.org/10.5603/CJ.a2015.0042) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26202651)
- 213. Ambrosioni, E.; Bacchelli, S.; Esposti, D.D.; Borghi, C. Beta-blockade in hypertension and congestive heart failure. *J. Cardiovasc. Pharmacol.* **2001**, *38* (Suppl. 3), S25–S31. [\[CrossRef\]](http://doi.org/10.1097/00005344-200112003-00005) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11811389)
- 214. Chin, B.S.; Langford, N.J.; Nuttall, S.L.; Gibbs, C.R.; Blann, A.D.; Lip, G.Y. Anti-oxidative properties of beta-blockers and angiotensin-converting enzyme inhibitors in congestive heart failure. *Eur. J. Heart Fail.* **2003**, *5*, 171–174. [\[CrossRef\]](http://doi.org/10.1016/S1388-9842(02)00251-9)
- 215. Bernjak, A.; Clarkson, P.B.; McClintock, P.V.; Stefanovska, A. Low-frequency blood flow oscillations in congestive heart failure and after beta1-blockade treatment. *Microvasc. Res.* **2008**, *76*, 224–232. [\[CrossRef\]](http://doi.org/10.1016/j.mvr.2008.07.006)
- 216. Turchetti, V.; Bellini, M.A.; Boschi, L.; Postorino, G.; Pallassini, A.; Richichi, M.G.; Trabalzini, L.; Guerrini, M.; Forconi, S. Haemorheological and endothelial-dependent alterations in heart failure after ACE inhibitor, calcium antagonist and beta blocker. *Clin. Hemorheol. Microcirc.* **2002**, *27*, 209–218.
- 217. Poelzl, G.; Frick, M.; Lackner, B.; Huegel, H.; Alber, H.F.; Mair, J.; Herold, M.; Schwarzacher, S.P.; Pachinger, O.; Weidinger, F. Short-term improvement in submaximal exercise capacity by optimized therapy with ACE inhibitors and beta blockers in heart failure patients is associated with restoration of peripheral endothelial function. *Int. J. Cardiol.* **2006**, *108*, 48–54. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2005.04.003)
- 218. Castro, P.; Vukasovic, J.L.; Chiong, M.; Diaz-Araya, G.; Alcaino, H.; Copaja, M.; Valenzuela, R.; Greig, D.; Perez, O.; Corbalan, R.; et al. Effects of carvedilol on oxidative stress and chronotropic response to exercise in patients with chronic heart failure. *Eur. J. Heart Fail.* **2005**, *7*, 1033–1039. [\[CrossRef\]](http://doi.org/10.1016/j.ejheart.2004.11.009)
- 219. Alfieri, A.B.; Briceno, L.; Fragasso, G.; Spoladore, R.; Palloshi, A.; Bassanelli, G.; Montano, C.; Arioli, F.; Cuko, A.; Ruotolo, G.; et al. Differential long-term effects of carvedilol on proinflammatory and antiinflammatory cytokines, asymmetric dimethylarginine, and left ventricular function in patients with heart failure. *J. Cardiovasc. Pharmacol.* **2008**, *52*, 49–54. [\[CrossRef\]](http://doi.org/10.1097/FJC.0b013e31817e0edd)
- 220. Falskov, B.; Hermann, T.S.; Raunso, J.; Christiansen, B.; Rask-Madsen, C.; Major-Pedersen, A.; Kober, L.; Torp-Pedersen, C.; Dominguez, H. Endothelial function is unaffected by changing between carvedilol and metoprolol in patients with heart failure–a randomized study. *Cardiovasc. Diabetol.* **2011**, *10*, 91. [\[CrossRef\]](http://doi.org/10.1186/1475-2840-10-91)
- 221. Ding, Y.; Wei, Z.; Li, J.; Zhu, L. Effects of Metoprolol Succinate Combined with Entresto on Cardiac Function Indexes and Coagulation Function in Patients with Congestive Heart Failure. *Comput. Math Methods Med.* **2022**, *2022*, 9765884. [\[CrossRef\]](http://doi.org/10.1155/2022/9765884)
- 222. Gibbs, C.R.; Blann, A.D.; Watson, R.D.; Lip, G.Y. Abnormalities of hemorheological, endothelial, and platelet function in patients with chronic heart failure in sinus rhythm: Effects of angiotensin-converting enzyme inhibitor and beta-blocker therapy. *Circulation* **2001**, *103*, 1746–1751. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.103.13.1746)
- 223. Fortini, F.; Vieceli Dalla Sega, F.; Marracino, L.; Severi, P.; Rapezzi, C.; Rizzo, P.; Ferrari, R. Well-Known and Novel Players in Endothelial Dysfunction: Updates on a Notch(ed) Landscape. *Biomedicines* **2021**, *9*, 997. [\[CrossRef\]](http://doi.org/10.3390/biomedicines9080997) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34440201)
- 224. Joannides, R.; Bizet-Nafeh, C.; Costentin, A.; Iacob, M.; Derumeaux, G.; Cribier, A.; Thuillez, C. Chronic ACE inhibition enhances the endothelial control of arterial mechanics and flow-dependent vasodilatation in heart failure. *Hypertension* **2001**, *38*, 1446–1450. [\[CrossRef\]](http://doi.org/10.1161/hy1201.096529)
- 225. Hryniewicz, K.; Dimayuga, C.; Hudaihed, A.; Androne, A.S.; Zheng, H.; Jankowski, K.; Katz, S.D. Inhibition of angiotensinconverting enzyme and phosphodiesterase type 5 improves endothelial function in heart failure. *Clin. Sci.* **2005**, *108*, 331–338. [\[CrossRef\]](http://doi.org/10.1042/CS20040266) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15574127)
- 226. Safonova, J.I.; Kozhevnikova, M.V.; Danilogorskaya, Y.A.; Zheleznykh, E.A.; Ilgisonis, I.S.; Privalova, E.V.; Khabarova, N.V.; Belenkov, Y.N. Possible pathway for heart failure with preserved ejection fraction prevention and treatment: The angiotensinconverting enzyme inhibitor effect on endothelial function in comorbid patients. *Kardiologiia* **2022**, *62*, 65–71. [\[CrossRef\]](http://doi.org/10.18087/cardio.2022.1.n1952) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35168535)
- 227. Besler, C.; Doerries, C.; Giannotti, G.; Luscher, T.F.; Landmesser, U. Pharmacological approaches to improve endothelial repair mechanisms. *Expert. Rev. Cardiovasc. Ther.* **2008**, *6*, 1071–1082. [\[CrossRef\]](http://doi.org/10.1586/14779072.6.8.1071)
- 228. Imanishi, T.; Hano, T.; Nishio, I. Angiotensin II accelerates endothelial progenitor cell senescence through induction of oxidative stress. *J. Hypertens.* **2005**, *23*, 97–104. [\[CrossRef\]](http://doi.org/10.1097/00004872-200501000-00018)
- 229. Chuang, S. Mathematic models for cancer chemotherapy: Pharmacokinetic and cell kinetic considerations. *Cancer Chemother. Rep.* **1975**, *59*, 827–842.
- 230. Levi, A.; Leshem-Lev, D.; Weissler-Snir, A.; Hasin, T.; Mats, I.; Murninkas, D.; Kornowski, R.; Lev, E.I.; Ben-Gal, T. The Effect of Mineralocorticoid Receptor Antagonists on Recruitment and Function of Endothelial Progenitor Cells in Patients with Congestive Heart Failure. *Isr. Med. Assoc. J.* **2018**, *20*, 233–238.
- 231. Marumo, T.; Uchimura, H.; Hayashi, M.; Hishikawa, K.; Fujita, T. Aldosterone impairs bone marrow-derived progenitor cell formation. *Hypertension* **2006**, *48*, 490–496. [\[CrossRef\]](http://doi.org/10.1161/01.HYP.0000235681.25685.cf) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16847146)
- 232. Farquharson, C.A.; Struthers, A.D. Spironolactone increases nitric oxide bioactivity, improves endothelial vasodilator dysfunction, and suppresses vascular angiotensin I/angiotensin II conversion in patients with chronic heart failure. *Circulation* **2000**, *101*, 594–597. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.101.6.594)
- 233. Abiose, A.K.; Mansoor, G.A.; Barry, M.; Soucier, R.; Nair, C.K.; Hager, D. Effect of spironolactone on endothelial function in patients with congestive heart failure on conventional medical therapy. *Am. J. Cardiol.* **2004**, *93*, 1564–1566. [\[CrossRef\]](http://doi.org/10.1016/j.amjcard.2004.03.015) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15194040)
- 234. Gager, G.M.; von Lewinski, D.; Sourij, H.; Jilma, B.; Eyileten, C.; Filipiak, K.; Hulsmann, M.; Kubica, J.; Postula, M.; Siller-Matula, J.M. Effects of SGLT2 Inhibitors on Ion Homeostasis and Oxidative Stress associated Mechanisms in Heart Failure. *Biomed. Pharmacother.* **2021**, *143*, 112169. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2021.112169) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34560555)
- 235. Cappetta, D.; De Angelis, A.; Bellocchio, G.; Telesca, M.; Cianflone, E.; Torella, D.; Rossi, F.; Urbanek, K.; Berrino, L. Sodium-Glucose Cotransporter 2 Inhibitors and Heart Failure: A Bedside-to-Bench Journey. *Front. Cardiovasc. Med.* **2021**, *8*, 810791. [\[CrossRef\]](http://doi.org/10.3389/fcvm.2021.810791) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35004918)
- 236. Packer, M. Autophagy stimulation and intracellular sodium reduction as mediators of the cardioprotective effect of sodiumglucose cotransporter 2 inhibitors. *Eur. J. Heart Fail.* **2020**, *22*, 618–628. [\[CrossRef\]](http://doi.org/10.1002/ejhf.1732)
- 237. Packer, M. SGLT2 Inhibitors Produce Cardiorenal Benefits by Promoting Adaptive Cellular Reprogramming to Induce a State of Fasting Mimicry: A Paradigm Shift in Understanding Their Mechanism of Action. *Diabetes Care* **2020**, *43*, 508–511. [\[CrossRef\]](http://doi.org/10.2337/dci19-0074)
- 238. Dyck, J.R.B.; Sossalla, S.; Hamdani, N.; Coronel, R.; Weber, N.C.; Light, P.E.; Zuurbier, C.J. Cardiac mechanisms of the beneficial effects of SGLT2 inhibitors in heart failure: Evidence for potential off-target effects. *J. Mol. Cell Cardiol.* **2022**, *167*, 17–31. [\[CrossRef\]](http://doi.org/10.1016/j.yjmcc.2022.03.005)
- 239. Theofilis, P.; Sagris, M.; Oikonomou, E.; Antonopoulos, A.S.; Siasos, G.; Tsioufis, K.; Tousoulis, D. Pleiotropic effects of SGLT2 inhibitors and heart failure outcomes. *Diabetes Res. Clin. Pract.* **2022**, *188*, 109927. [\[CrossRef\]](http://doi.org/10.1016/j.diabres.2022.109927)
- 240. Kolijn, D.; Pabel, S.; Tian, Y.; Lodi, M.; Herwig, M.; Carrizzo, A.; Zhazykbayeva, S.; Kovacs, A.; Fulop, G.A.; Falcao-Pires, I.; et al. Empagliflozin improves endothelial and cardiomyocyte function in human heart failure with preserved ejection fraction via reduced pro-inflammatory-oxidative pathways and protein kinase Galpha oxidation. *Cardiovasc. Res.* **2021**, *117*, 495–507. [\[CrossRef\]](http://doi.org/10.1093/cvr/cvaa123)
- 241. Koyani, C.N.; Plastira, I.; Sourij, H.; Hallstrom, S.; Schmidt, A.; Rainer, P.P.; Bugger, H.; Frank, S.; Malle, E.; von Lewinski, D. Empagliflozin protects heart from inflammation and energy depletion via AMPK activation. *Pharmacol. Res.* **2020**, *158*, 104870. [\[CrossRef\]](http://doi.org/10.1016/j.phrs.2020.104870)
- 242. Heidenreich, P.A.; Bozkurt, B.; Aguilar, D.; Allen, L.A.; Byun, J.J.; Colvin, M.M.; Deswal, A.; Drazner, M.H.; Dunlay, S.M.; Evers, L.R.; et al. 2022 AHA/ACC/HFSA Guideline for the Management of Heart Failure: A Report of the American College of Cardiology/American Heart Association Joint Committee on Clinical Practice Guidelines. *Circulation* **2022**, *145*, e895–e1032. [\[CrossRef\]](http://doi.org/10.1161/CIR.0000000000001063)
- 243. Menendez, J.T. The Mechanism of Action of LCZ696. *Card Fail. Rev.* **2016**, *2*, 40–46. [\[CrossRef\]](http://doi.org/10.15420/cfr.2016:1:1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28785451)
- 244. Amore, L.; Alghisi, F.; Pancaldi, E.; Pascariello, G.; Cersosimo, A.; Cimino, G.; Bernardi, N.; Calvi, E.; Lombardi, C.M.; Sciatti, E.; et al. Study of endothelial function and vascular stiffness in patients affected by dilated cardiomyopathy on treatment with sacubitril/valsartan. *Am. J. Cardiovasc. Dis.* **2022**, *12*, 125–135. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/suab139.058) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35873182)
- 245. Li, B.H.; Fang, K.F.; Lin, P.H.; Zhang, Y.H.; Huang, Y.X.; Jie, H. Effect of sacubitril valsartan on cardiac function and endothelial function in patients with chronic heart failure with reduced ejection fraction. *Clin. Hemorheol. Microcirc.* **2021**, *77*, 425–433. [\[CrossRef\]](http://doi.org/10.3233/CH-201032) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33386797)
- 246. Nathaniel, S.; McGinty, S.; Witman, M.A.H.; Edwards, D.G.; Farquhar, W.B.; Hosmane, V.; Wenner, M.M. Impact of angiotensin receptor-neprilysin inhibition on vascular function in heart failure with reduced ejection fraction: A pilot study. *Physiol. Rep.* **2022**, *10*, e15209. [\[CrossRef\]](http://doi.org/10.14814/phy2.15209)
- 247. Angadi, S.S.; Mookadam, F.; Lee, C.D.; Tucker, W.J.; Haykowsky, M.J.; Gaesser, G.A. High-intensity interval training vs. moderateintensity continuous exercise training in heart failure with preserved ejection fraction: A pilot study. *J. Appl. Physiol. (1985)* **2015**, *119*, 753–758. [\[CrossRef\]](http://doi.org/10.1152/japplphysiol.00518.2014)
- 248. Winzer, E.B.; Gaida, P.; Hollriegel, R.; Fischer, T.; Linke, A.; Schuler, G.; Adams, V.; Erbs, S. Impact of Rosuvastatin Treatment on HDL-Induced PKC-betaII and eNOS Phosphorylation in Endothelial Cells and Its Relation to Flow-Mediated Dilatation in Patients with Chronic Heart Failure. *Cardiol. Res. Pract.* **2016**, *2016*, 4826102. [\[CrossRef\]](http://doi.org/10.1155/2016/4826102)
- 249. Ellis, G.R.; Nightingale, A.K.; Blackman, D.J.; Anderson, R.A.; Mumford, C.; Timmins, G.; Lang, D.; Jackson, S.K.; Penney, M.D.; Lewis, M.J.; et al. Addition of candesartan to angiotensin converting enzyme inhibitor therapy in patients with chronic heart failure does not reduce levels of oxidative stress. *Eur. J. Heart Fail.* **2002**, *4*, 193–199. [\[CrossRef\]](http://doi.org/10.1016/S1388-9842(02)00002-8)
- 250. Nakamura, M.; Saito, S.; Yoshida, H.; Arakawa, N.; Sugawara, S.; Hiramori, K. Effects of chronic subdepressor dose of angiotensin II type 1 receptor antagonist on endothelium-dependent vasodilation in patients with congestive heart failure. *J. Cardiovasc. Pharmacol.* **2002**, *40*, 411–419. [\[CrossRef\]](http://doi.org/10.1097/00005344-200209000-00010)
- 251. Macdonald, J.E.; Kennedy, N.; Struthers, A.D. Effects of spironolactone on endothelial function, vascular angiotensin converting enzyme activity, and other prognostic markers in patients with mild heart failure already taking optimal treatment. *Heart* **2004**, *90*, 765–770. [\[CrossRef\]](http://doi.org/10.1136/hrt.2003.017368) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15201246)
- 252. Correale, M.; Mazzeo, P.; Mallardi, A.; Leopizzi, A.; Tricarico, L.; Fortunato, M.; Magnesa, M.; Tucci, S.; Maiellaro, P.; Pastore, G.; et al. Switch to SGLT2 Inhibitors and Improved Endothelial Function in Diabetic Patients with Chronic Heart Failure. *Cardiovasc. Drugs Ther.* **2022**, *36*, 1157–1164. [\[CrossRef\]](http://doi.org/10.1007/s10557-021-07254-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34519913)
- 253. Sezai, A.; Sekino, H.; Unosawa, S.; Taoka, M.; Osaka, S.; Tanaka, M. Canagliflozin for Japanese patients with chronic heart failure and type II diabetes. *Cardiovasc. Diabetol.* **2019**, *18*, 76. [\[CrossRef\]](http://doi.org/10.1186/s12933-019-0877-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31167663)
- 254. Bunsawat, K.; Ratchford, S.M.; Alpenglow, J.K.; Park, S.H.; Jarrett, C.L.; Stehlik, J.; Smith, A.S.; Richardson, R.S.; Wray, D.W. Sacubitril-valsartan improves conduit vessel function and functional capacity and reduces inflammation in heart failure with reduced ejection fraction. *J. Appl. Physiol. (1985)* **2021**, *130*, 256–268. [\[CrossRef\]](http://doi.org/10.1152/japplphysiol.00454.2020)
- 255. Du, H.; Li, X.; Zhao, W.; Jiang, N. The Difference between Sacubitril Valsartan and Valsartan on Vascular Endothelial Function, APN, MMP-9, and BNP Levels in Patients with Hypertension and Chronic Heart Failure. *J. Healthc. Eng.* **2022**, *2022*, 9494981. [\[CrossRef\]](http://doi.org/10.1155/2022/9494981)
- 256. Karavolias, G.K.; Georgiadou, P.; Gkouziouta, A.; Kariofillis, P.; Karabela, G.; Tsiapras, D.; Sbarouni, E.; Chaidaroglou, A.; Degiannis, D.; Adamopoulos, S.; et al. Short and long term anti-inflammatory effects of bosentan therapy in patients with pulmonary arterial hypertension: Relation to clinical and hemodynamic responses. *Expert Opin. Ther. Targets* **2010**, *14*, 1283–1289. [\[CrossRef\]](http://doi.org/10.1517/14728222.2010.523421)
- 257. Iannone, F.; Riccardi, M.T.; Guiducci, S.; Bizzoca, R.; Cinelli, M.; Matucci-Cerinic, M.; Lapadula, G. Bosentan regulates the expression of adhesion molecules on circulating T cells and serum soluble adhesion molecules in systemic sclerosis-associated pulmonary arterial hypertension. *Ann. Rheum. Dis.* **2008**, *67*, 1121–1126. [\[CrossRef\]](http://doi.org/10.1136/ard.2007.080424)
- 258. Sfikakis, P.P.; Papamichael, C.; Stamatelopoulos, K.S.; Tousoulis, D.; Fragiadaki, K.G.; Katsichti, P.; Stefanadis, C.; Mavrikakis, M. Improvement of vascular endothelial function using the oral endothelin receptor antagonist bosentan in patients with systemic sclerosis. *Arthritis Rheum.* **2007**, *56*, 1985–1993. [\[CrossRef\]](http://doi.org/10.1002/art.22634)
- 259. Eroglu, E.; Kocyigit, I.; Lindholm, B. The endothelin system as target for therapeutic interventions in cardiovascular and renal disease. *Clin. Chim. Acta* **2020**, *506*, 92–106. [\[CrossRef\]](http://doi.org/10.1016/j.cca.2020.03.008)
- 260. Schneider, M.P.; Boesen, E.I.; Pollock, D.M. Contrasting actions of endothelin ET(A) and ET(B) receptors in cardiovascular disease. *Annu. Rev. Pharmacol. Toxicol.* **2007**, *47*, 731–759. [\[CrossRef\]](http://doi.org/10.1146/annurev.pharmtox.47.120505.105134)
- 261. Price, L.C.; Howard, L.S. Endothelin receptor antagonists for pulmonary arterial hypertension: Rationale and place in therapy. *Am. J. Cardiovasc. Drugs* **2008**, *8*, 171–185. [\[CrossRef\]](http://doi.org/10.2165/00129784-200808030-00004) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18533738)
- 262. Bisserier, M.; Pradhan, N.; Hadri, L. Current and emerging therapeutic approaches to pulmonary hypertension. *Rev. Cardiovasc. Med.* **2020**, *21*, 163–179. [\[CrossRef\]](http://doi.org/10.31083/j.rcm.2020.02.597) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32706206)
- 263. Ilsar, R.; Levitt, J.; Ng, M.K.; Kritharides, L.; Adams, M.R.; Celermajer, D.S. Bosentan and improved pulmonary endothelial function in pulmonary arterial hypertension. *Eur. Respir. J.* **2010**, *36*, 1483–1485. [\[CrossRef\]](http://doi.org/10.1183/09031936.00083910) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21119207)
- 264. Pearl, J.M.; Wellmann, S.A.; McNamara, J.L.; Lombardi, J.P.; Wagner, C.J.; Raake, J.L.; Nelson, D.P. Bosentan prevents hypoxiareoxygenation-induced pulmonary hypertension and improves pulmonary function. *Ann. Thorac. Surg.* **1999**, *68*, 1714–1721, discussion 1712–1721. [\[CrossRef\]](http://doi.org/10.1016/S0003-4975(99)00988-1)
- 265. Yagi, S.; Akaike, M.; Iwase, T.; Kusunose, K.; Niki, T.; Yamaguchi, K.; Koshiba, K.; Yoshida, S.; Sumitomo-Ueda, Y.; Aihara, K.; et al. Bosentan ameliorated exercise-induced pulmonary arterial hypertension complicated with systemic sclerosis. *Intern. Med.* **2010**, *49*, 2309–2312. [\[CrossRef\]](http://doi.org/10.2169/internalmedicine.49.3812)
- 266. Cella, G.; Vianello, F.; Cozzi, F.; Marotta, H.; Tona, F.; Saggiorato, G.; Iqbal, O.; Fareed, J. Effect of bosentan on plasma markers of endothelial cell activity in patients with secondary pulmonary hypertension related to connective tissue diseases. *J. Rheumatol.* **2009**, *36*, 760–767. [\[CrossRef\]](http://doi.org/10.3899/jrheum.080542)

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