

Functional and Metabolic Imaging in Heart Failure with Preserved Ejection Fraction: Promises, Challenges, and Clinical Utility

Matthew K Burrage¹ · Andrew J Lewis^{1,2} · Jack J J. Miller^{1,2,3,[4](http://orcid.org/0000-0002-6258-1299)}

Accepted: 8 June 2022 / Published online: 26 July 2022 © The Author(s) 2022

Abstract

Heart failure with preserved ejection fraction (HFpEF) is recognised as an increasingly prevalent, morbid and burdensome condition with a poor outlook. Recent advances in both the understanding of HFpEF and the technological ability to image cardiac function and metabolism in humans have simultaneously shone a light on the molecular basis of this complex condition of diastolic dysfunction, and the infammatory and metabolic changes that are associated with it, typically in the context of a complex patient. This review both makes the case for an integrated assessment of the condition, and highlights that metabolic alteration may be a measurable outcome for novel targeted forms of medical therapy. It furthermore highlights how recent technological advancements and advanced medical imaging techniques have enabled the characterisation of the metabolism and function of HFpEF within patients, at rest and during exercise.

Keywords Metabolic imaging · HFpEF · Cardiac metabolism · C13 · PET · Heart failure · Diastolic dysfunction · Systematic review

Introduction

The Demographics of Heart Failure

Half of patients clinically presenting with heart failure (HF) have a preserved left ventricular ejection fraction (HFpEF). Despite this, the symptoms of HFpEF are typically sufficient to cause a substantial reduction in their quality of life. Their

 \boxtimes Jack J J. Miller jack.miller@physics.org

> Matthew K Burrage matthew.burrage@cardiov.ox.ac.uk

Andrew J Lewis andrew.lewis@cardiov.ox.ac.uk

- ¹ Oxford Centre for Clinical Cardiovascular Magnetic Resonance Research (OCMR); Radclife Department of Medicine, University of Oxford, John Radclife Hospital, Oxford, UK
- Department of Physiology, Anatomy and Genetics, University of Oxford, Parks Road, Oxford, UK
- ³ The PET Research Centre and The MR Research Centre, Aarhus University, Aarhus, Denmark
- ⁴ Department of Physics, Clarendon Laboratory, University of Oxford, Parks Road, Oxford, UK

disease follows a substantially similar course to patients suffering heart failure with a reduced ejection fraction (HFrEF), with a similar morbidity and mortality burden. HFpEF presents a large and currently unmet need for effective medical therapy and is increasingly recognised as a major and growing pathology with a complex aetiology, increasing at a rate of approximately 1% per year [\[1](#page-15-0)]. As its major risk factors, including hypertension, obesity, and type 2 diabetes mellitus (T2DM), are increasing in most populations, HFpEF is expected to emerge as the most prevalent HF phenotype worldwide [[2\]](#page-15-1). It is worth highlighting that patients with HFpEF are as functionally limited as their counterparts with HFrEF: requiring frequent hospitalisation and living with a poor quality of life $[3-5]$ $[3-5]$, and their overall mortality is similar to that of most HFrEF populations, with observational studies reporting a dismal 5-year survival of only 35–40% post-hospitalisation for HF—comparable to many cancers.

The demographic of the HFpEF patient is varied and typically comorbid. Population-based studies (e.g. the US Veterans Afairs cohort of 1.8 million individuals [\[6](#page-15-4)]) have reported that HFpEF patients are predominantly older women [[1\]](#page-15-0), with established cardiovascular risk factors such as obesity, hypertension, chronic kidney disease, coronary artery disease (CAD), anaemia, hyperlipidaemia, T2DM, and atrial fbrillation [[7–](#page-15-5)[9\]](#page-15-6) shown via multivariable logistic regression to be associated with increased odds ratios (OR) of developing HFpEF. Whilst the existence of these risk factors is arguably unsurprising, their relative magnitudes difer at times substantially between the conditions: for example, the comparative risk for developing HFpEF following previous MI is approximately half that compared with HFrEF (OR / 95% confdence interval: 2.85 (2.78–2.93) for HFpEF compared to 4.18 (4.10–4.26) for HFrEF); with BMI and diabetes further independently hinting at a distinct mechanism of pathology (c.f. Fig. [1\)](#page-1-0) on a population level alone [\[10\]](#page-15-7).

The Link Between Pressure, Function, and Metabolism

This information, replicated through many diferent patient studies, provides a strong but subtle cue to examine the molecular pathogenesis of HFpEF: its risk factors such as type 2 diabetes, obesity, and hypertension, are all indepedently associated with impaired cardiac energy metabolism [\[11](#page-15-8)[–17\]](#page-16-0).

From a fundamental physical point of view, this is selfconsistent: the heart as a pump does hydraulic work whilst iterating through a pressure-volume $(P - V)$ loop and,

from basic thermodynamics, the integral area of that loop, $\oint P \cdot dV$, necessarily has to be equal to the stroke work undertaken in a cardiac cycle. Broadly speaking, HFpEF is a disease characterised by high flling pressures and mechanically stif ventricles, with patients falling on a spectrum of diastolic dysfunction [[18\]](#page-16-1). Approximately a third of the patients presenting with HFpEF have a normal collagen volume fraction as ascertained through endomyocardial leftventricular biopsies [[19\]](#page-16-2) (the remainder raised), but all possess LVEDP, LV end-systolic wall stress, and LV stifness modulus consistent with patients presenting with a raised collagen volume fraction. This suggests that in addition to collagen deposition, intrinsic cardiomyocyte stifness also contributes directly to diastolic LV dysfunction in HFpEF, a fnding experimentally borne out in patient groups where further biopsies are available [[19,](#page-16-2) [20](#page-16-3)]. The biomechanical efect of these alterations in myocardial stifness is a marked increase in pulmonary arterial pressure: an increase in myocardial stifness results in both impaired LV relaxation and flling. This increases the left atrial pressure, exposing the whole of the pulmonary vascular system and ultimately the right heart to an increased pressure workload accordingly, independent of other (comorbid) associated changes in the peripheral vascular resistance that may be present in the

Fig. 1 Multivariable logistic regression derived odds ratios (OR) for risk factors in developing HFpEF and HFrEF in the 1.8 million US Veterans Affairs population (comprising $n = 66 831$ HFpEF and $n = 92233$ HFrEF patients). Data are reproduced (with permission) from Gaziano et al. [[6\]](#page-15-4), and have been corrected for age, sex, race, and ethnicity. A thin blue line highlights the diferences between HFpEF and HFrEF. Abbreviations: ACE, angiotensin converting enzyme; ARB, angiotensin receptor blocker; BMI, body mass index

(coefficient per 5 kg/m^2); CAD, coronary artery disease; COPD, chronic obstructive pulmonary disease; DBP, diastolic blood pressure (per 10 mmHg); eGFR, estimated glomerular fltration rate (per 15 ml∕min∕1.73m²; HDL-c, high density lipoprotein cholesterol (per 15 mg/dl); HFpEF, heart failure with preserved ejection fraction; HFrEF, heart failure with reduced ejection fraction; LDL-c, low density lipoprotein cholesterol (per 35 mg/dl); MI, myocardial infarction; SBP, systolic blood pressure (per 20 mmHg)

patient. This leads to increased pulmonary transcapillary hydrostatic pressure, which drives fuid transudation, potentially increasing capillary diameter from the Young-Laplace relation, and hence leading to pulmonary congestion and breathlessness [[21\]](#page-16-4). What is perhaps often under-appreciated is the role of the right heart in this process. Reduced right ventricular contractile reserve and reduced coupling of the right ventricle to the pulmonary circulation or RV-PA coupling results in increased right-sided pressures, with right atrial dilatation and increased right atrial and systemic venous pressures. This exacerbates pulmonary congestion by reducing clearance of lung water via pulmonary lymphatics [\[22](#page-16-5)[–25](#page-16-6)].

Together with an increase in myocyte and ventricular stifness (i.e. mechanically increasing stroke work), the metabolome in HFpEF patients is significantly altered, and again diferentially altered in comparison to HFrEF patients. Plasma metabolomics reveals that, accounting for diferences in blood biochemistry known to be due to comorbidities such as T2DM, HFpEF is associated with indices of increased infammation and oxidative stress (such as higher levels of hydroxyproline and symmetric dimethyl arginine, alanine, cystine), impaired lipid metabolism (lower lysophophatidylcholine, potentially altered coupling between glucose oxidation and glycolysis [[26\]](#page-16-7)), increased collagen synthesis, and downregulated nitric oxide signalling. Together, these fndings suggest a more predominant systemic microvascular endothelial dysfunction and infammation linked to increased fbrosis in HFpEF compared with HFrEF [[27](#page-16-8)].

As summarised in Fig. [2A](#page-2-0), HFpEF therefore remains a disease that is (a) prevalent and expected to become increasingly so; (b) complex and necessarily associated with numerous comorbid pathologies that may help defne its molecular milieu; (c) deeply connected with the biophysical mechanics of the heart; and (d) a disease with a high mortality and morbidity (Fig. [2B\)](#page-2-0).

Cardiac Metabolism in Heart Failure

The Heart as a Metabolic Omnivore

The healthy heart has an enormous energy requirement and consumes more energy and oxygen than any other organ [[30\]](#page-16-9). It continuously produces large amounts of adenosine triphosphate (ATP), which is necessary to sustain both active myocardial contraction and active diastolic relaxation. This is achieved by metabolising a variety of fuels (primarily fatty acids and glucose with additional contributions from lactate, ketones, and amino acids) and oxidative phosphorylation within the cardiac mitochondrial respiratory electron transport chain [[31,](#page-16-10) [32\]](#page-16-11). In health, the bulk of mitochondrial ATP production, which contributes approximately \sim 95% of myocardial ATP requirements, is derived from fatty acid oxidation (∼ 40–70%), with the remainder originating from glucose and lactate via the oxidation of pyruvate [[32,](#page-16-11) [33](#page-16-12)]. Only about 5% of myocardial ATP demand is met by glycolysis, which does not require oxygen [[32,](#page-16-11) [34,](#page-16-13) [35\]](#page-16-14). The heart demonstrates a degree of metabolic fexibility and is able to utilise diferent energy substrates under diferent conditions to maintain function, although this may become lost in the failing heart $[36]$ $[36]$. Given the reliance of actin-myosin interactions on ATP and oxygen availability, it is clear that disruption in cardiac energy metabolism pathways and altered

Fig. 2 A Taken together, these factors are sufficient to delineate HFpEF as a very distinct disease from HFrEF. As detailed by Guazzi et al. [[28](#page-16-16)], in HFpEF, concentric myocyte remodelling with increased left ventricular (LV) diastolic stifness, atrial functional MR, and a stiff left atrium (LA) are the major driving forces to alveolar-capillary stress failure and vascular remodelling, with corresponding diferential changes in the end-systolic/end-diastolic pressure-volume relation

(ESPVR/EDPVR) compared to healthy controls (shown in blue). **B** It is worth further stressing that, although not typically caused by an ischaemic origin, HFpEF has a similar mortality to HFrEF following MI, as illustrated here in an "all-comers" Ethiopian cohort HF study reporting outcomes on all HF patients treated at the University of Gondar Referral Hospital between 2010 and 2015 [\[29\]](#page-16-17)

myocardial energetics have signifcant implications for cardiac function and the pathogenesis of heart failure [\[30](#page-16-9), [32](#page-16-11)].

The Failing Heart and Its Fuels

As outlined above, heart failure presents a major societal impact, is increasingly prevalent, and is associated with substantial morbidity and mortality [[37\]](#page-16-18). However, the role of cardiac energy metabolism is perhaps more intuitive in the pathophysiology of HFrEF, in that it is easier to conceptualise that the failing heart with contractile dysfunction is starved of fuel [[30](#page-16-9)]. Impaired cardiac energy metabolism as demonstrated with non-invasive phosphorus magnetic resonance spectroscopy (31P-MRS) has been shown to correlate with the degree of heart failure and predict mortality in patients with HFrEF due to ischaemic and dilated cardiomyopathy [\[38](#page-16-19)[–41](#page-16-20)]. In contrast, HFpEF remains challenging to diagnose, vastly heterogeneous in terms of pathophysiology and clinical risk factors and, until recently [\[42](#page-16-21)], had few therapeutic options [[43](#page-16-22)]. Because of this, there is increasing interest to more carefully phenotype individual patients with HFpEF [[44\]](#page-16-23). Recent basic scientifc and translational studies have provided further evidence that impaired cardiac energy metabolism also plays a central role in the pathogenesis of HFpEF, with alterations in lipid metabolism, and an increase in mitochondrial stress and ROS together with other infammatory-related changes in mitochondrial function [[45](#page-16-24)[–49](#page-16-25)]. This is plausible, given that active diastolic relaxation has the greatest energy demand of any phase of the cardiac cycle [\[50](#page-16-26)]: rather than a metabolic disorder of contraction, HFpEF is, if anything, a disease of relaxation (but one that is distinct from diastolic dysfunction alone [[21\]](#page-16-4)) and moreover one with few universally recognised animal models of the

Table 1 A brief summary of some of the main metabolic changes associated with HFpEF (in comparison with HFrEF). The detailed metabolic changes that occur in heart failure are complex, and are

condition, in contrast to HFrEF in which ischaemia-reperfusion models of MI progress routinely to HFrEF. In both conditions, cardiac dysfunction further manifests as a series of profound shifts in the utilisation of fuels through specifc pathways, as summarised in Table [1,](#page-3-0) which potentially appear to be distinct between HFpEF and HFrEF.

Multiple cardiometabolic and other risk factors for HFpEF, such as T2DM, obesity, and hypertension, are all independently associated with impaired cardiac energy metabolism, and can be shown through magnetic resonance spectroscopy studies in human patients and volunteers under a variety of cardiometabolic states to quantitatively afect cardiac energetics $[11-17]$ $[11-17]$ $[11-17]$. Diastolic dysfunction is also associated with impaired myocardial energetics in translational and clinical models, but represents a more widely spread disease in patients [\[71](#page-17-0), [72](#page-17-1)].

It is therefore clear that metabolic modulation may thus be a promising therapeutic strategy for the treatment of heart failure in appropriately phenotyped patients [[32,](#page-16-11) [73](#page-17-2)]. As detailed subsequently, novel metabolic and functional imaging techniques may help identify those patients who could derive most beneft from such strategies, as well as act as appropriate tools to use in future clinical trials.

Infammation and HFpEF

Alongside impaired myocardial energetics, there is a strong body of pre-clinical and clinical evidence that HFpEF is a multi-system disease characterised by systemic infammation that affects the myocardium, skeletal muscles, vasculature, and kidneys [[74\]](#page-17-3), ultimately resulting in increased left ventricular myocardial stifness and fbrosis. This is one of

dependent not only on the severity and type of heart failure present, but also on the coexistence of common comorbidities such as obesity and type 2 diabetes, and reviewed in detail elsewhere [\[70\]](#page-17-4)

the hallmarks of HFpEF physiology, where increased myocardial stifness and impaired diastolic relaxation results in high LV flling pressures and exertional breathlessness. It has been proposed that systemic infammation is linked to comorbidities associated with HFpEF, particularly diabetes mellitus and obesity, which are associated with a systemic proinfammatory state, haematopoietic activation and systemic leukocytosis [[75](#page-17-17)], and the subsequent development of myocardial stifness and fbrosis [\[76\]](#page-17-18).

Titin as a Link Between Infammation and Stifness

Paulus et al. have presented a clear series of pathophysiological mechanisms that link systemic infammation and myocardial stifness [\[74\]](#page-17-3). Metabolic load associated with comorbidities, such as obesity, diabetes, and chronic kidney disease, induces proinfammatory signalling and systemic infammation as evidenced by raised plasma levels of infammatory cytokines [\[77](#page-17-19)[–80](#page-17-20)]. Haemodynamic load, as seen in arterial hypertension, also activates proinfammatory and profbrotic signalling pathways [\[74\]](#page-17-3). Increased haemodynamic load is sensed by cardiomyocytes, fbroblasts, and resident macrophages resulting in a cascade of structural remodelling and leukocyte recruitment that promotes fbroblast activation and collagen deposition, thereby increasing myocardial stifness [[74](#page-17-3)]. Modifcations in the titin protein are a major contributor to cardiomyocyte stifness. Titin is the largest protein in the body, spans multiple regions within the human sarcomere, and is responsible for diastolic distensibility. It consists of two main isoforms, the short stif N2B isoform and the longer, more compliant N2BA isoform. Post-translational titin oxidation or phosphorylation modulates distensibility and contribute to the high myocardial stifness seen in HFpEF. The N2BA/N2B titin isoform ratio is lower and the N2B isoform is hypophosphorylated in HFpEF [[81,](#page-17-21) [82](#page-17-22)].

Additionally, reactive oxygen species (ROS) cause formation of disulfde bonds within titin, also contributing to increased cardiomyocyte stifness [[83](#page-17-23), [84\]](#page-17-24). Changes in myocardial collagen homeostasis also lead to abnormalities of the extracellular matrix, resulting in myocardial fbrosis and stifness [[76\]](#page-17-18). This has been evidenced by an increase in collagen volume fraction on myocardial biopsy in patients with HFpEF [[84,](#page-17-24) [85\]](#page-17-25). Finally, the myocardial accumulation of degraded proteins, which occurs due to infammationtriggered expression of inducible nitric oxide synthase in cardiomyocytes, is also thought to drive myocardial stifness [\[45](#page-16-24), [76](#page-17-18)]. This increase in stifness directly causes haemodynamic efects that lead ultimately to the development of the symptoms of the disease, as summarised in Fig. [3.](#page-6-0)

It is worth refecting that many studies which have provided mechanistic and molecular detail on the mechanobiology of HFpEF have done so through the acquisition of myocardial biopsy samples taken from patients listed for operations related either to their condition or their comorbidities, typically at the more severe end of the phenotype. These samples provide histological evidence supporting the twin hypotheses of both diferential extracellular matrix content (with HFpEF patients having higher titin expression in their myocytes and a higher collagen content in the surrounding extracellular matrix), leading to mechanically stifer myocytes with a stronger passive force per unit area (*F*passive); and inflammation associated with HFpEF. In addition, endomyocardial biopsy provides a gold-standard method to diagnose or rule out cardiac amyloidosis, a prevalent aetiology of "HFpEF" that is mechanistically distinct, caused by the extracellular deposition of autologous protein as amyloid fbrils from a variety of more than 30 molecular sources [[90\]](#page-17-26) that initially presents as a restrictive cardiomyopathy characterized by progressive diastolic dysfunction that subsequently becomes systolic biventricular dysfunction with oft-fatal arrhythmia. A summary of histological fndings in HFpEF is shown in Table [2,](#page-5-0) which are consistent with the pattern described above. Whilst this tissue is highly valuable, and can be subject to a large number of related diferential analyses, it is prudent to be mindful of the fact that human biopsy samples are typically only obtained from patients at the more developed end of the disease spectrum and for whom the surgical intervention is otherwise justifed. This selection bias combined with the risks and cost of biopsy motivates the investigation of both non-invasive, or less-invasive diagnostic techniques such as imaging methods, and furthermore the scientifc study of less severe disease.

Imaging Infammation

The ability to non-invasively characterise myocardial infammation and fbrosis has important implications for management. Emerging imaging tracers using hyperpolarized magnetic resonance or positron emission tomography enable non-invasive assessment of major cardiac leukocyte populations, including macrophages [[91](#page-17-27), [92](#page-17-28)]. Novel PET tracers targeting the cardiac stroma including collagen could be combined with cardiovascular MR for a complete assessment of the extracellular volume fraction. A recent study which identifed patients with HFpEF and myocardial fbrosis on cardiovascular magnetic resonance imaging showed that the oral antifibrotic agent pirfenidone may reduce myocardial fibrosis [\[93](#page-17-29)]. The effects of this on clinical outcomes and patient symptoms still needs to be confrmed. Other immune-mediated conditions may more directly cause cardiac infammation and heart failure symptoms. Multiple systemic autoimmune or immune-mediated disorders may cause myocarditis, where infammation is the primary driver of cardiac dysfunction, while this is most commonly

Table 2 Human biopsy studies in HFpEF reveal a complex, multifacted disease with increased collagen deposition, alterations in the ECM, and a signifcant infammatory component. Reported n is for HFpEF patients in each study; studies listed have substantially different designs and control arms. Abbreviations: ECM, extracellular matrix; EM, electron microscopy; FA, force analysis; HM, histomorphometry; IF, immunofluorescence; IHC, immunohistochemistry; RT-PCR, real-time reverse transcription-polymerase chain reaction

attributable to viral infection [\[94\]](#page-18-0). Identifcation of active primary cardiac infammation is important as patients are far more likely to respond to immunosuppressive therapies. Advanced myocardial tissue characterisation techniques have shown considerable promise in the detection of myocardial infammation [[95\]](#page-18-1), having identifed cardiac involvement in a wide variety of systemic conditions [[96–](#page-18-2)[104](#page-18-3)].

Functional Imaging and HFpEF

It is therefore highly desirable to attempt to simultaneously ascertain both the function of the heart and measures of its metabolome and microstructure in the context of patients who may well be suffering from relevant comorbid diseases (e.g. T2DM). It is therefore recognised that, in a clinical context, the diagnosis of HFpEF remains challenging, with several diagnostic criteria proposed by learned bodies and in clinical trials [[18\]](#page-16-1).

Imaging approaches span a spectrum of complexity, invasiveness, availability, and cost, and these competing concerns have to be balanced against either the scientifc rationale for their engagement, or the clarity which they could add to the diagnosis of a condition that exhibits a high misdiagnosis rate [\[105](#page-18-4), [106\]](#page-18-5). Furthermore, HFpEF is a condition where comparatively subtle physiological changes can be manifest even at the early stages of the disease [\[18](#page-16-1)]. The haemodynamic response of the heart to increased myocardial stifness includes left atrial dilatation (with a volume index *>* 32 ml/m2 shown to increase the risk of cardiovascular mortality), and a reduction of diastolic function as typically quantifed by the mitral E velocity (*>* 90 cm/s) and septal e' velocity (*<* 9 cm/s) on echocardiography, or equivalently an increase in their ratio (*>* 9). Whilst some degree of left ventricular hypertrophy is supportive of HFpEF, its absence does not exclude its diagnosis [[18\]](#page-16-1).

What is perhaps under-appreciated is the variability present within the condition and the degree to which it can be masked by existing co-morbidities or be confused with other forms of heart failure; one study by Kanagala et al. [[107\]](#page-18-6) on

Fig. 3 A very brief schematic overview of the currently understood pathogenesis of HFpEF. It is important to remember that the HFpEF syndrome is a biventricular process and that the contributions of the right heart are equally important. (1) Increased left ventricular stifness, and a decreased functional reserve, results in (2) increased left-sided flling pressures with left atrial dilatation and increased left atrial pressure. This leads to increased pulmonary transcapillary hydrostatic pressure, which drives fuid transudation, potentially increasing capillary diameter from the Young-Laplace relation, and resulting in pulmonary congestion (3). Similar maladaptive right heart processes may also occur in parallel. (4) Reduced right ventric-

154 patients with a provisional HFpEF diagnosis underwent a comprehensive CMR examination (stress/rest perfusion, cine function, and late gadolinium enhancement). Forty-two patients (27%) were found to have an alternative diagnosis (coronary artery disease, microvascular dysfunction, hypertrophic cardiomyopathy, or constrictive pericarditis), with worse outcomes.

Cardiovascular MR in HFpEF

Cardiovascular magnetic resonance (CMR) is the goldstandard imaging modality for assessing atrial and ventricular volumes and accurately quantifying ejection fraction (in contrast to echocardiography, which sufers both from an increased measurement variability and systematic bias in underestimating cardiac volumes, owing both to increased inter- and intra-observer variability and the conventional assumption that the LV forms a prolate ellipsoid when its mass is estimated through linear measurements made of the wall on the parasternal long-axis view $[108-110]$ $[108-110]$ $[108-110]$). As a consequence, although the full role of CMR for diastolic function assessment is evolving, it currently allows accurate assessment of the structural changes associated with HFpEF, such as left atrial (LA) enlargement, and pathological LV hypertrophy, along with mitral infow and pulmonary venous

ular contractile reserve, and reduced coupling of the right ventricle to the pulmonary circulation or RV-PA coupling, results in increased right-sided pressures, with right atrial dilatation and increased right atrial and systemic venous pressures (5). This exacerbates pulmonary congestion by reducing clearance of lung water via pulmonary lymphatics. The underlying pathogenesis of HFpEF is still the subject of ongoing research, but pro-infammatory changes result in a mechanical increase in the stifness of the myocardium mediated through alterations in collagen deposition and the biophysics of titin, which occurs concomitantly with metabolic changes leading to an increased oxidative stress on the myocyte

flow velocity. These MR flow measurements are obtained via phase-contrast (PC) methods, wherein a bipolar gradient of magnetic feld is applied along a given axis, and a change in the phase of the MR signal produced in proportion to the velocity of blood fowing along it. For the most part, the interpretation of these measures largely follows that of Doppler echocardiography, with a comparatively more coarse temporal resolution and a requirement to acquire data over several heartbeats as opposed to in real-time, meaning that CMR measurements of flow are hence more susceptible to arrhythmias. However, in addition to the measurement of mitral and pulmonary fow in one direction, it is possible to quantify the three-dimensional velocity vector feld of blood through with CMR, using "4D-fow" sequences in which the direction of the spatial magnetic feld gradient that encodes fow information in the phase of the MR signal is varied in 3D space. Over the course of a longer, more complex acquisition, whole-heart fow patterns can be obtained throughout the cardiac cycle which can both reveal complex valvular insufficiency, retained residual blood volume, and the degree of vorticity and turbulent fow in diastole [[111](#page-18-9), [112](#page-18-10)]. HFpEF patients are understood to have increased diastolic infow vortex strength and greater turbulent flow, again consistent with a biomechanical dysfunction of pumping function [[113\]](#page-18-11), leading to complex and subtly diferent responses to

fuid-dynamical alterations brought about by, for example, the efect of a presence of an interventricular septal shunt or vasodilation, compared to either healthy individuals or those with HFrEF [\[114](#page-18-12), [115\]](#page-18-13).

Additionally, diastolic dysfunction can also be inferred through alterations of the flling dynamics of each chamber of the heart. The increased availability of dedicated semiautomatic cardiac image analysis and segmentation software (oft using "AI" algorithms in the process) has recently enabled these to be computed relatively more rapidly by those reading the images, removing the requirement to manually segment, e.g. endo- and epicardial contours for the quantifcation of the LV time-volume flling curve. The resulting peak flling rates and time to peak flling can therefore easily be obtained. Both time to peak flling, peak flling rate, and other novel indices aimed at quantifying these curves have been demonstrated to be of utility in quantifying left ventricular diastolic dysfunction [\[116\]](#page-18-14).

Furthermore, myocardial strain (and inferred stifness) can be directly be determined through feature tracking or tagging techniques—which can either be determined post hoc after the acquisition of a conventional cine image or by the use of the spin-physics behind MR to impose structured tags within the myocardial tissue that then deform over time [\[117\]](#page-18-15). These thus therefore infer the motion of individual voxels of myocardial tissue, either by imprinting upon the physical myocardium or by estimating its motion in the image domain. As strain is efectively defned as extension over original length, longitudinal, radial, and circumferential strain can be estimated for the heart by the use of either a 2D or 3D mathematical model of deformation [\[118](#page-18-16)], once a computational model of the deformation over time of each individual voxel has been determined. This process remains an active research area for both techniques, owing to the fact that multiple complex cardiac motions may result in the same observed deformation [[119](#page-18-17), [120\]](#page-18-18). The utility of the technique in HFpEF comes from that fact that, if considered as a rigid body, a stifer myocardium will be unable to deform as much under the same force, and increased myocardial stifness can be quantifed directly (c.f. Table [3](#page-8-0)). Perhaps refecting this, multi-layer feature-tracking derived strain metrics have thus been shown to correlate with NTproBNP and efectively be able to diagnose HFpEF in a signal scan with an 89% sensitivity and a specifcity of 100% [\[121\]](#page-18-19), and global longitudinal strain has itself been associated with mortality and other hard cardiovascular outcomes in HFpEF [[122\]](#page-18-20).

Finally, it is possible to use CMR to assess fbrosis directly through either late gadolinium-enhancement MR, or the increasingly available technique of T_1 mapping with and without gadolinium and the concurrent estimation of the extracellular volume (ECV) fraction $[116]$ $[116]$ $[116]$. These related methods utilise the known fact that gadolinium contrast agents extravasate and are retained in interstitial spaces. Whilst conventionally used for the comparatively routine detection of fbrotic scar (typically following myocardial infarction) through late gadolinium-enhancement MR, by measuring the nuclear T_1 time of the myocardium pre and post the addition of gadolinium subtle, not visually obvious differences in T_1 and hence gadolinium uptake can be determined. If the patient's haematocrit is known, their extracellular volume fraction can be correspondingly estimated, and expressed in intuitive units of per cent [[123](#page-18-21)]. An increase in ECV is almost always due either to excessive collagen deposition or cardiac amyloidosis, and ECV has been shown to act as an independent predictor of intrinsic LV stifness in HFpEF patients as measured via invasive pressure-volume loops [[124](#page-18-22)]. Fortunately for the diferential diagnosis of cardiac amyloidosis, the appearance of global, subendocardial late gadolinium is characteristic [[125](#page-18-23)], as is an inability to null the enhanced signal in an inversion-recovery late gadolinium MR scan, and an increase in the native T_1 of the amyloid myocardium [\[90,](#page-17-26) [126\]](#page-18-24). These CMR changes appear to be present for both AL and ATTR forms of cardiac amyloidosis, highlighting that CMR is a complementary technique to SPECT imaging with the bone-seeking diphosphate radionuclide 99*m*Tc-DPD, a sensitive scintigraphy tracer that is highly effective at detecting the ATTR form of cardiac amyloidosis at an early stage, but only efective in approximately a third of patients with cardiac AL amyloid [[90](#page-17-26)].

Advanced forms of proton cardiac imaging, therefore, have an important and growing role in the diagnosis of HFpEF from a clinical perspective, able to probe and quantify both the morphological adaptations in the HFpEF heart arising from increased flling pressures and impaired diastolic relaxation, and also probe changes due to increased mechanical stifness and altered collagen content. Furthermore, compared to the use of echocardiography alone, the wider availability of CMR may help address the underdiagnosis of cardiac amyloidosis as a prevalent aetiology of "HFpEF" given the practical limits of the accessibility of complementary imaging (i.e. 99*m*Tc-DPD SPECT-CT) for amyloid specifcally.

Exercise (In‑)Tolerance

Together with functional changes at rest, HFpEF, like other forms of heart failure, is associated with a signifcant decrease in the exercise reserve of the patient. This arises as a biophysical consequence of increased LV stifness: during hyperaemic stress there is an inability to augment the left-ventricular end-diastolic volume despite increasing the left-ventricular end-diastolic pressure for a given cardiac workload. The outcome of this is an increased stress on both the pulmonary vasculature and the heart as

a whole: as is well known in HFrEF, exercise acts therefore as a cardiopulmonary stress test under which pathology may reveal itself in HFpEF as well, with patients with early-stage HFpEF displaying symptoms and invasively measured flling pressure increases only present on exertion [\[129,](#page-18-25) [130](#page-19-0)]. Exercise testing with concurrent echocardiography (ExE) has been used to provide further useful pieces of information that are of relevance in the diagnosis of HFpEF. Firstly, the awareness and perception of dyspnoea may be highly variable between patients and value therefore added by an objective measure both of cardiac parameters and exercise capacity [[131](#page-19-1)[–133](#page-19-2)]. Secondly, it is possible to acquire myocardial E/e' during exercise, which has been shown to increase under stress through a study of $n = 436$ patients presenting with fatigue or dsypnoea and considered for the diagnosis of HFpEF [\[129\]](#page-18-25) (as illustrated in Fig. [4A-C](#page-9-0)). Thirdly, the formation of hyperechoic "B" line artefacts is directly detectable in the lungs of patients with acute pulmonary congestion [[134](#page-19-3)], and shown to correlate with B-type natriuretic peptide (BNP), respiratory rate, clinical congestion, and systolic pulmonary arterial pressure (Fig. [4D\)](#page-9-0) as a measure of pulmonary congestion. In HFpEF, the development of pulmonary congestion upon exercise is mostly concomitant with exercise-induced worsening of diastolic function [[128](#page-18-26)], and CMR measures of cardiac function during exercise indicate that the HFpEF patient sufers atrial dilatation, indicative of higher flling pressures (Fig. [5\)](#page-9-1). Tricuspid regurgitation has additionally been shown to be present on ExE, which, together with E/E', can predict future mortality [\[135,](#page-19-4) [136](#page-19-5)].

Fig. 5 In an "in-magnet" CMR exercise study using a stepping ergometer (**A**) with whole-heart coverage (**B**) during 20 W submaximal exercise on a population of patients spanning the spectrum of diastolic dysfunction, it was found that the absolute change in right (**C**) and left (**D**) atrial volumes during exercise was highly maladaptive in HFpEF: greater dilation during exercise is suggestive of increased flling pressures [\[137](#page-19-6)]

Metabolic Imaging Methods

These profound changes in the morphology, function,

Fig. 4 Echocardiography prior to and post exercise can reveal comparatively subtle increases in diastolic dysfunction. Reproduced here is the case of a 75-year-old male with current exertional dyspnoea and a history of percutaneous coronary interventional therapy. Wall motion analysis revealed no exercise-induced wall motion abnormality in either view (**A**, **B**). The measured mitral flow pattern and tissue Doppler was consistent with that of delayed relaxation (**C**) and E/E' increased from 14 at rest to 16 with exercise [[127](#page-18-27)]. Similarly, hyperechoic b-lines that arise in the lung from transudated fuid (**D**) that can be detected in HFpEF patients with dyspnoea, with good prognostic power [[128\]](#page-18-26)

and structure of the heart in HFpEF occur concomitantly with alterations in metabolism and a shift in substrates used by the heart, as inferred by biopsy and metabolomic approaches outlined previously in Table [1](#page-3-0). These invasive techniques necessarily are difficult to apply to a larger number of patients and are typically reserved either to studying preclinical models or patients who have efectively reached the end stage of the disease and undergo surgery either for LVAD implantation or transplant. In contrast, non-invasive or less-invasive molecular imaging techniques have recently been applied to both confrm or discover that these changes exist *in vivo* and at an earlier stage of the disease. All of these techniques provide a consistent picture of the disease becoming unmasked under exercise stress, summarised in Table [4.](#page-11-0)

Broadly speaking, CMR with nuclei other than protons, X-nuclei MR, and radionuclide imaging techniques are those that are able to best interrogate these changes in humans. Phosphorus-31 cardiac magnetic resonance spectroscopy is an established X-nuclear MR technique that is able to quantify the presence and concentration of ³¹P-containing moeities within the human heart, most notably phosphocreatine (PCr) and ATP. This refects the energetic status of the heart as phosphocreatine acts as a labile energy buffer and the endpoint of multiple metabolic pathways. Accordingly, the PCr/ATP ratio is well known to be lowered in advanced heart failure [[30\]](#page-16-9), diabetes [\[146,](#page-19-7) [147\]](#page-19-8), and HFpEF [\[148\]](#page-19-9).

Interestingly, it is possible to perform exercise experiments within the confnes of the bore of the MR scanner, permitting the determination of PCr/ATP at rest and during submaximal exercise, together with the observation of functional changes in cardiac haemodynamics [[148](#page-19-9)]. This therefore highlights multinuclear CMR as a powerful technique with the ability to simultaneously interrogate the energetic status of the heart through ³¹P-spectroscopy, and probe functional and microstructural methods as outlined above in the same scan session. Some authors have developed magnet-safe exercise devices, together with advanced cardiac and pulmonary imaging CMR sequences (retrospectively gated compressed-sensing cine; and a novel radial ultrashort echo time lung water proton density mapping sequence) that permit the quantifcation of cardiac function and pulmonary free fuid during exercise $([137];$ $([137];$ $([137];$ summarised in Fig. [6\)](#page-12-0). It was found that under a constant low workload of 20 W, patients across a spectrum of diastolic dysfunction (11 age-matched controls, 9 with diabetes mellitus, 14 with clinical HFpEF, and 9 with amyloid cardiomyopathy) had a gradient of deficit in the PCr/ATP, an increase in the detected lung water again increasing across the groups, and (as expected) changes in atrial volumes that are likely associated with increased flling pressures.

These factors paint a strong picture of a maladaptive cardiac response to exercise that is associated with a potentially causative metabolic shift: as well as reporting reduced PCr/ ATP ratios and increased exercise lung water, previous work has aimed to characterise cardiac metabolism in HFpEF with radiolabelled 11 C-acetate as a PET tracer [\[150](#page-19-10), [151\]](#page-19-11). Injected free acetate is rapidly taken up by the heart and then mitochondrially converted to acetyl coenzyme A and metabolised to carbon dioxide through the TCA cycle through oxidative phosphorylation; and thus, 11 C-acetate PET imaging provides a reproducible proxy measurement of cardiac oxidative metabolism (i.e. $MVO₂$) at centres that have the cyclotron required to synthesise this tracer with a 20-min half-life. In a study of $n = 11$ HFpEF patients and $n = 10$ controls, decreased cardiac update of 11 C-acetate was inversely correlated with E/e' in patients only $[150]$ $[150]$. In a more recent study $[149]$ $[149]$ that additionally included dobutamine-induced exercise mimetic stress with prospectively enrolled HF_pEF patients ($n = 19$) and matched controls $(n = 19)$ that evaluated myocardial blood flow (MBF) and mechanical cardiac work in addition to $MVO₂$, it was found that, at rest, compared with controls, patients with HFpEF had higher cardiac work, $MVO₂$, and MBF. During dobutamine stress, cardiac work, $MVO₂$, and MBF increased in both HFpEF and controls, but the magnitude of increases was signifcantly smaller in HFpEF. This is indicative, as before, of the impaired 'exercise reserve' of the HFpEF heart. In both groups, MBF increased with dobutamine stress in relation to cardiac work, but the magnitude of this increase was signifcantly reduced in HFpEF patients, while HFpEF patients with LV hypertrophy had a signifcant reduction in computed left ventricular mechanical efficiency compared to controls, summarised in Fig. [7](#page-13-0). This implies a mismatch of both cardiac energetic demand and blood fow—pointing to some degree of coronary microvascular disease—and moreover an alteration in the metabolic efficiency of the heart. It is worth stressing that pulmonary hypertension alone has been indicated in driving the heart towards glycolysis and the role of the right-heart in HFpEF cannot be understated [\[152](#page-19-13)]—but, as ever, isolating cause, effect, and symptom remains challenging owing to the exquisitely integrated nature of cardiovascular physiology.

A Future Role for Advanced Imaging in HFpEF?

An abnormal metabolism is clearly implicated in HFpEF and HFrEF alike, and the question of whether (or not) metabolic changes drive HFrEF is often described as a decades-old chicken-and-egg problem in the context of HFrEF [[30\]](#page-16-9). The roles of the immune system, of titin, and of maladaptation to the resultant alterations in the mechanical stifness of the HFpEF heart are also clear: once a patient has started down

 $\underline{\textcircled{\tiny 2}}$ Springer

elimination; VO_2 , oxygen uptake [145] capillary wedge pressure; *VE/VCO*₂, ratio of minute ventilation to carbon dioxide elimination; *VO*₂, oxygen uptake [[145](#page-19-14)] 100 $\times 100$ carbon ventilation to

Fig. 6 Patients with increasing diastolic dysfunction progressively dilated atrial volumes, most likely as a result of increased flling pressures as alluded to in Fig. [3.](#page-6-0) ³¹ P-magnetic resonance spectroscopy (obtaining from the mid interventricular septum**A** spectra showing the presence of high-energy metabolites such as phosphocreatine (PCr), three resonances corresponding to the three ³¹P nuclei in ATP; and 2,3-diphosphoglycerate, **B**). This subsequently revealed in that these patients a discrete energetic deficit (i.e. a reduction in the detectable PCr/ATP) that was progressively worse across the groups (**C**). This was accompanied by an increase in detected pulmonary fuid as detected by a novel protondensity mapping MR sequence (**D**, [\[137\]](#page-19-6))

the path of diastolic dysfunction, increased flling pressures, and other comorbidities that defne the HFA-PEFF diagnostic scoring system, the manifest condition is relatively easy to study. What is less clear, however, is exactly why the epidemiologically relevant risk factors for HFpEF are obesity and T2DM. HFpEF is not always the most straightforward of conditions to diagnose: relevant masquerading conditions to rule out via a number of distinct investigations include cardiac amyloidosis, sarcoidosis, hypertrophic cardiomyopathy, valvular heart disease, high-output heart failure, myocarditis, constrictive pericarditis, and the various toxin-mediated cardiomyopathies [\[153\]](#page-19-22).

Advanced metabolic imaging, therefore, might fulfl two separate purposes. Firstly, in a clinical research or pre-clinical setting, the ability to determine uniquely the balance of substrates used by the myocardium would directly inform the research of the underlying shifts in cardiac metabolism and whether they precede, occur contemporaneously with, or follow alterations in myocardial stifness (in HFrEF, myocardial metabolic changes precede the descent into heart failure [[30,](#page-16-9) [154](#page-19-23), [155\]](#page-19-24)). Moreover, trials investigating pharmacologic regimens aimed at improving the functional outcomes of HFpEF patients, or prevent those at high risk from developing the disease, would beneft from the ability to have a direct readout of their metabolic efect. Many such therapies have been proposed and investigated for HF in general, with somewhat mixed efects: etomoxir and perhexiline (reported as targeting carnitine palmitoyl transferase, decreasing myocardial fatty acid metabolism, and favouring a reciprocal increase in glycolysis) have shown beneft in patients but may be limited by side efects; trimetazidine and L-propionylcarnitine, decreasing fatty-acid oxidation, increasing glucose oxidation; and increasing fatty acid transport across the mitochondrial membrane directly may have a survival and functional benefit [[156\]](#page-19-25). Despite their success in pre-clinical trials, most of these molecules have not found widespread adaptation in patients. One potential reason for this is that "all-comers" trials based on those with a clinical deficit in functional scores such as the New York Heart Association functional class may not select patients with an underlying pathogenic mechanism immediately compatible with the metabolic modulation each drug provides. Being able to directly image the specifc alterations involved in the disease is therefore highly desirable.

Secondly, beyond scientifc inquiry into the molecular origins of HFpEF, there are several reasons to believe that multimodal metabolic imaging may have a patient-specifc role in the diagnosis and management of the condition.

Fig. 7 Myocardial oxidative metabolism inferred from the kinetics of the PET tracer ¹¹C-acetate, which is visible in the RV and LV before being taken up into the myocardium and later excreted as it decays (**A**). By ftting these data to an appropriate mathematical model (**B**), it is possible to estimate myocardial mechanical work (denoted external work, EW) and infer $MVO₂$ and myocardial blood fow (MBF). In a trial of *n* = 19 HFpEF patients undergoing dobutamine exercisemimetic stress, it was found that the ability of the heart to augment these values was reduced in HFpEF. This is consistent with an earlier indication in $n = 11$ HFpEF patients at rest that myocardial metabolic capacity is decreased as E/E' increases (**D**) and as the left ventricular end-diastolic dimension decreases (LV EDV mass index is a diagnostic criterion of HFpEF), which highlights a metabolic impairment in the disease (panels **A**–**C** reproduced from [\[149](#page-19-12)]; **D** from [\[150\]](#page-19-10) with permission)

Metabolic imaging—either in the form of advanced MRI or PET with appropriate tracers—has long been recognised of being valuable in cardiology where the metabolic demands of the organ are inherently linked to its function with disease-varied efficiencies $[157-159]$ $[157-159]$ $[157-159]$. Novel technological progress, in the form of either advanced PET reconstruction algorithms and improved multi-modal imaging, or hyperpolarised MR (which uses endogenous metabolites such as $[1 - {^{13}C}]$ pyruvate together with low-temperature spinphysics to make their metabolism transiently visible to the scanner), heralds considerable promise for improving the ability of sites to routinely and rapidly quantify more aspects of cardiac function. For that reason, more than 42 clinical trials registered on clinicaltrials.gov (with 21 currently recruiting) aim to use advanced MR, multimodal cardiac MRI, and measures of metabolism directly to prognosticate HFpEF (for example, by quantifying its microvascular milieu through the use of frst-pass perfusion gadolinium scanning at rest and under dobutamine stress, with radioactive H_2 ¹⁵O PET perfusion scans as a gold-standard perfusion readout [\[160](#page-19-28)]). Hyperpolarised MR, meanwhile, has already

been shown to be of utility in investigating ischaemic heart disease [[161\]](#page-19-29), resolving both regional alterations in glucose utilisation caused by coronary artery disease, and in the diabetic myocardium, wherein insensitivity to glucose uptake and altered postprandial response was directly quantifed [[162](#page-19-30)]. The changes in energetic status within the HFpEF myocardium found via 31P spectroscopy are intriguing and point towards a disease on a spectrum of diastolic dysfunction potentially being both unifed by an energetic defcit and ultimately prognosticated by it. Hyperpolarised pyruvate imaging can, in principle, achieve higher resolution imaging than similar PET scans within a minute $[163]$ $[163]$ and may therefore be of utility in further characterising the HFpEF heart. This technique is currently in international clinical trials in a variety of indications, and ofers the promise of a rapid and comprehensive assessment of myocardial metabolism which may be of beneft both in HFpEF and additionally for identifying the ischaemic region at risk in ischaemic cardiomyopathies. At present is a comparatively expensive and rare technique, but one with a wide number of potential applications both in cardiology and in other conditions, such as monitoring tumour response to therapy, and a similar cost base to PET. Should its value and utility be demonstrated conclusively in forthcoming trials, it is at least possible and at best probable that hyperpolarised MR could have clinical availability in the medium-term future.

The need for an advanced understanding of cardiac metabolism *in vivo* in human HFpEF patients is well illustrated by recent successful clinical trials. The surprising (yet positive) results of studies such as EMPA-REG which showed a signifcant beneft to cardiovascular mortality in T2DM patients with improved glycaemic control provided by empaglifozin, a sodium-glucose co-transporter 2 inhibitor [[164](#page-19-32)], potentially by procuring a shift in cardiac metabolism [[165\]](#page-19-33), has led to a recent clinical trial aiming to replicate these fndings in HFpEF patients independent of diabetes status. In the EMPEROR-Preserved trial of nearly 6000 patients with HFpEF and NYHA class II to IV functional impairment, empaglifozin reduced by around 21% the combined risk of cardiovascular death or hospitalisation for HFpEF patients, regardless of the presence or absence of diabetes [[166\]](#page-19-34). This follows from a string of broadly speaking failures in wide HFpEF demographic groups: PEP-CHF (antihypertensive perindopril, [\[167](#page-20-0)]); CHARM-Preserved (antihypertensive candesartan, [[168\]](#page-20-1)), I-PRESERVE (anti-hypertensive irbesartan, [[169\]](#page-20-2)), TOPCAT (potassium-sparing diuretic spironolactone, [[170](#page-20-3)]), and PARAGON-HF (angiotensin receptor-neprilysin inhibitor sacubitril/ valsartan, [[171\]](#page-20-4)) all failed to show substantial beneft with hard patient end-points. The fact that a metabolic modulator, albeit one with a variety of as yet-to-be-elucidated cardiac efects, can signifcantly improve patient mortality in both HFpEF and HFrEF demographics is puzzling, and requires further scientifc study. Even small benefcial changes in cardiac energetic utilisation within a beat-to-beat timeframe may translate into large differences in efficiency, known to be associated with outcomes [[165](#page-19-33)]. The sister studies to EMPEROR-Preserved evaluating SGLT2 inhibition in HFrEF, the DAPA-HF and EMPEROR-Reduced trials, have similarly shown signifcant reductions in morbidity and mortality with SGLT2 inhibition (dapaglifozin and empaglifozin respectively), again independent of diabetic state [[172](#page-20-5)[–174](#page-20-6)]. This is strongly suggestive of an underlying metabolic mechanism, not least because the heart does not express SGLT2 and radiolabelled 14C autoradiographic studies indicate that it is not detectable directly in the heart [[175](#page-20-7)]. Off-target effects involving either sodium homeostasis, or action on the sodium-proton exchanger, are not reported in isolated ventricular cardiomyocytes over a wide range of doses that may otherwise explain a direct cardiac mechanism of action [\[176\]](#page-20-8). The need to fully understand the metabolic changes that SGLT2 inhibitors can induce in patients with either HFrEF or HFpEF is therefore highlighted neatly: by altering systemic metabolism we can improve mortality, although we as yet do not know how.

The mechanisms underpinning the beneft of SGLT2 inhibitors (SGLT2i) in HFpEF remain elusive, though several compelling candidate pathways have been postulated. Restoration of myocardial energetics and normalisation of metabolism are a key candidate mechanism, linked to the observation that SGLT2is promotes mild ketosis, via an increase in production of the ketone body beta-hydroxybutyrate as reported in both invasive animal studies [\[177,](#page-20-9) [178](#page-20-10)] and patient populations [[179](#page-20-11), [180](#page-20-12)]. Ketones may offer a more efficient myocardial metabolic substrate, requiring fewer moles of oxygen per mole of ATP produced [[181](#page-20-13)], thus counteracting metabolic infexibility arising from an over-reliance on non-esterifed fatty acids in HFpEF [\[182,](#page-20-14) [183\]](#page-20-15). Alternatively, ketones might abrogate pro-hypertrophic signalling pathways and prevent MAP kinase activation resulting in changes in left ventricular mass [[184](#page-20-16), [185](#page-20-17)]. Importantly, multiple facets of these mechanisms are amenable to non-invasive assessment, using the arsenal of imaging tools described in earlier sections. For example, this could include hyperpolarised 13 C MRI and/or PET for cardiac substrate metabolism $[186]$ $[186]$ $[186]$, ³¹P MRS for energetics, and ¹H spectroscopy for myocardial lipid content. Ongoing clinical trials and investigations will aim to directly reproduce and test these fndings in patient populations.

A second key candidate mechanism relates to modulation of the cardiac stroma, including resident innate immune cell and fbroblast function, which play a key role in maintaining normal cardiac function through the regulation of cardiac metabolism and the cardiac renin-angiotensin-aldosterone system, as well as the release of soluble paracrine factors with anti-fibrotic, pro-angiogenic, and anti-apoptotic effects [[187,](#page-20-19) [188](#page-20-20)]. SGLT2is has been linked in other settings with amelioration of pro-infammatory and pro-fbrotic signalling, and it is conceivable that these may have relevance to clinical beneft in HFpEF [\[189\]](#page-20-21). Again, using novel molecular imaging probes as described above in Sections "[Imaging](#page-4-0) [Infammation"](#page-4-0) and "[Cardiovascular MR in HFpEF"](#page-6-1), it is now possible not only to assess key leukocyte populations within the myocardium, but also to measure myocardial fbrosis via cardiac extracellular volume. Additionally, the modulation of adipokine function (including a reduction in serum leptin) has been linked with altered epicardial fat deposition profles [[190](#page-20-22), [191](#page-20-23)], the changes of which can be straightforwardly assessed using imaging.

Given the undeniable disease-modifying benefit that SGLT2i has been shown to have, elucidating their molecular mechanisms non-invasively with metabolic imaging methods in patients would provide a signifcant improvement in the understanding of HFpEF in general.

Key Conclusions

HFpEF is a complex yet prevalent condition that is driven by an increase in the bio-mechanical stifness of the heart, microscopically originating from changes in titin expression and alterations in the extracellular matrix and an increased collagen volume fraction of the myocardium. This leads to impaired relaxation, and hence maladaptive changes in the pressure/volume relationship of the heart. It is therefore often referred to as being defned by diastolic dysfunction.

Whilst sharing many common cardiovascular risk factors and co-morbidities with HFrEF, the HFpEF heart has itself been shown to be metabolically distinct. Advanced non-invasive imaging techniques are able to accurately quantify both the biomechanical phenotype of the HFpEF heart and its blunted haemodynamic response to exercise, and point towards energetic impairment. The recent emergence of SGLT2 inhibition as a disease-modifying therapy arguably provides further evidence that both myocardial and systemic metabolic efects should be investigated in order to further elucidate its mechanistic role in ameliorating this morbid and prevalent condition.

Author Contributions All the authors wrote, reviewed, and edited this manuscript. We additionally would like to acknowledge Damian Tyler for his advice and interesting discussions whilst internally reviewing this manuscript.

Funding All the authors would like to acknowledge the support of the British Society for Cardiovascular Research; of the British Heart Foundation and the British Heart Foundation Oxford Centre of Research Excellence. MKB would like to acknowledge a BHF CRTF, ref. FS/19/65/34692. AJL would like to acknowledge support from the Academy of Medical Sciences (SGL021/1038). JJM would like to acknowledge support from a Novo Nordisk Postdoctoral Fellowship run in conjunction with the University of Oxford; and a Starter Grant from the Novo Foundation, ref. NNF21OC0068683.

Data availability Please see referenced primary manuscripts (if appropriate).

Code Availability Please see referenced primary manuscripts (if appropriate).

Declarations

Ethics Approval Please see appropriate cited primary publications where relevant.

Consent to Participate Please see appropriate cited primary publications where relevant.

Consent for Publication Please see appropriate cited publications where relevant.

Conflict of Interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by/4.0/>.

References

- 1. Lam CSP, Donal E, Kraigher-Krainer E, Vasan RS. Epidemiology and clinical course of heart failure with preserved ejection fraction. European Journal of Heart Failure. 2011;13(1):18–28.
- 2. Owan TE, et al. Trends in prevalence and outcome of heart failure with preserved ejection fraction. The New England Journal of Medicine. 2006;355(3):251–9.
- 3. Hoekstra T, Lesman-Leegte I, van Veldhuisen DJ, Sanderman R, Jaarsma T. Quality of life is impaired similarly in heart failure patients with preserved and reduced ejection fraction. European Journal of Heart Failure. 2011;13(9):1013–8.
- 4. Shah SJ, et al. Baseline characteristics of patients in the treatment of preserved cardiac function heart failure with an aldosterone antagonist trial. Circulation Heart Failure. 2013;6(2):184–92.
- 5. Steinberg BA, et al. Trends in patients hospitalized with heart failure and preserved left ventricular ejection fraction: Prevalence, therapies, and outcomes. Circulation. 2012;126(1):65–75.
- 6. Gaziano L, et al. Risk factors and prediction models for incident heart failure with reduced and preserved ejection fraction. ESC Heart Failure. 2021;8(6):4893–903.
- 7. Bhatia RS, et al. Outcome of heart failure with preserved ejection fraction in a population-based study. The New England Journal of Medicine. 2006;355(3):260–9.
- 8. Lam CSP, et al. Sex diferences in clinical characteristics and outcomes in elderly patients with heart failure and preserved ejection fraction: The Irbesartan in Heart Failure with Preserved Ejection Fraction (I-PRESERVE) trial. Circulation Heart Failure. 2012;5(5):571–8.
- 9. Yancy CW, et al. Clinical presentation, management, and inhospital outcomes of patients admitted with acute decompensated heart failure with preserved systolic function: A report from the Acute Decompensated Heart Failure National Registry (ADHERE) Database. Journal of the American College of Cardiology. 2006;47(1):76–84.
- 10. Lindman BR, et al. Cardiovascular phenotype in HFpEF patients with or without diabetes: A RELAX trial ancillary study. Journal of the American College of Cardiology. 2014;64(6):541–9.
- 11. Lamb H, et al. Diastolic dysfunction in hypertensive heart disease is associated with altered myocardial metabolism. Circulation. 1999;99(17):2261–7.
- 12. Rider OJ, et al. Efects of catecholamine stress on diastolic function and myocardial energetics in obesity. Circulation. 2012;125(12):1511–9.
- 13. Levelt E, et al. Cardiac energetics, oxygenation, and perfusion during increased workload in patients with type 2 diabetes mellitus. European Heart Journal. 2015;37(46):3461–9.
- 14. Rayner J, et al. The relative contribution of metabolic and structural abnormalities to diastolic dysfunction in obesity. International Journal of Obesity. 2018;42(3):441–7.
- 15. Rayner J, et al. Myocardial energetics in obesity: Enhanced ATP delivery through creatine kinase with blunted stress response. Circulation. 2020;141(14):1152–63.
- 16. Rayner J, et al. Obesity modifes the energetic phenotype of dilated cardiomyopathy. Eur heart J. 2021.
- 17. Scheuermann-Freestone M, et al. Abnormal cardiac and skeletal muscle energy metabolism in patients with type 2 diabetes. Circulation. 2003;107(24):3040–6.
- 18. McDonagh T, et al. 2021 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: Developed by the Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) With the special contribution of the Heart Failure Association (HFA) of the ESC. European Heart Journal. 2021;42(36):3599–726.
- 19. Borbély A, et al. Cardiomyocyte stifness in diastolic heart failure. Circulation. 2005;111(6):774–81.
- 20. Borlaug BA, Paulus WJ. Heart failure with preserved ejection fraction: Pathophysiology, diagnosis, and treatment. Eur Heart J. 2010;32(6):670–9.
- 21. A AO, Shah SJ. Diagnosis and management of Heart Failure with Preserved Ejection Frac-tion : 10 Key Lessons. Current Cardiology Reviews. 2015;11(1):42–52.
- 22. Gorter TM, Obokata M, Reddy YNV, Melenovsky V, Borlaug BA. Exercise unmasks distinct pathophysiologic features in heart failure with preserved ejection fraction and pulmonary vascular disease. European Heart Journal. 2018;39(30):2825–35.
- 23. Obokata M, et al. Haemodynamics, dyspnoea, and pulmonary reserve in heart failure with preserved ejection fraction. European Heart Journal. 2018;39(30):2810–21.
- 24. Lamb HJ, et al. Diastolic dysfunction in Hypertensive heart disease is associated with altered myocardial metabolism. Circulation. 1999;99(17):2261–7.
- 25. Reddy YNV, et al. The haemodynamic basis of lung congestion during exercise in heart failure with preserved ejection fraction. European Heart Journal. 2019;40(45):3721–30.
- 26. Fillmore N, et al. Uncoupling of glycolysis from glucose oxidation accompanies the development of heart failure with preserved ejection fraction. Molecular Medicine (Cambridge, Mass). 2018;24(1):3.
- 27. Hage C, et al. Metabolomic profile in HFpEF vs HFrEF patients. Journal of Cardiac Failure. 2020;26(12):1050–9.
- 28. Guazzi M, Ghio S, Adir Y. Pulmonary hypertension in HFpEF and HFrEF : JACC review topic of the week. Journal of the American College of Cardiology. 2020;76(9):1102–11.
- 29. Abebe TB, Gebreyohannes EA, Tefera YG, Abegaz TM. Patients with HFpEF and HFrEF have diferent clinical characteristics but similar prognosis: A retrospective cohort study. BMC Cardiovascular Disorders. 2016;16(1):232.
- 30. Neubauer S. The failing heart – an engine out of fuel. New England Journal of Medicine. 2007;356(11):1140–51.
- 31. Stanley W, Recchia F, Lopaschuk G. Myocardial substrate metabolism in the normal and failing heart. Physiological Reviews. 2005;85(3):1093–129.
- 32. Lopaschuk G, Karwi Q, Tian R, Wende A, Abel E. Cardiac energy metabolism in heart failure. Circulation Research. 2021;128(10):1487–513.
- 33. Watson WD, et al. Use of cardiac magnetic resonance to detect changes in metabolism in heart failure. Cardiovascular Diagnosis and Therapy. 2020;10(3):583–97.
- 34. Wisneski J, Stanley W, Neese R, Gertz E. Efects of acute hyperglycemia on myocardial glycolytic activity in humans. The Journal of Clinical Investigation. 1990;85(5):1648–56.
- 35. Saddik M, Lopaschuk G. Myocardial triglyceride turnover and contribution to energy substrate utilization in isolated

working rat hearts. The Journal of Biological Chemistry. 1991;266(13):8162–70.

- 36. Karwi Q, Uddin G, Ho K, Lopaschuk G. Loss of metabolic fexibility in the failing heart. Frontiers in Cardiovascular Medicine. 2018;5:68.
- 37. Savarese G, Lund L. Global public health burden of heart failure. Cardiac Failure Review. 2017;3(1):7–11.
- 38. Hardy C, Weiss R, Bottomley P, Gerstenblith G. Altered myocardial high-energy phosphate metabolites in patients with dilated cardiomyopathy. American Heart Journal. 1991;1:795–801.
- 39. Neubauer S, et al. 31P magnetic resonance spectroscopy in dilated cardiomyopathy and coronary artery disease. Altered cardiac high-energy phosphate metabolism in heart failure. Circulation. 1992;86(6):1810–8.
- 40. Neubauer S, et al. Contributions of 31P-magnetic resonance spectroscopy to the understanding of dilated heart muscle disease. European Heart Journal. 1995;16:115–8.
- 41. Neubauer S, et al. Myocardial phosphocreatine-to-atp ratio is a predictor of mortality in patients with dilated cardiomyopathy. Circulation. 1997;96(7):2190–6.
- 42. Anker SD, et al. Empagliflozin in heart failure with a preserved ejection fraction. New England Journal of Medicine. 2021;385(16):1451–61.
- 43. Borlaug B. Evaluation and management of heart failure with preserved ejection fraction. Nature reviews Cardiology. 2020;17(9):559–73.
- 44. Shah S, et al. Phenotype-specifc treatment of heart failure with preserved ejection fraction: A multiorgan roadmap. Circulation. 2016;134(1):73–90.
- 45. Schiattarella G, et al. Nitrosative stress drives heart failure with preserved ejection fraction. Nature. 2019;568(7752):351–6.
- 46. Kumar A, Kelly D, Chirinos J. Mitochondrial dysfunction in heart failure with preserved ejection fraction. Circulation. 2019;139(11):1435–50.
- 47. Chaanine A, et al. Mitochondrial morphology, dynamics, and function in human pressure overload or ischemic heart disease with preserved or reduced ejection fraction. Circulation Heart Failure. 2019;1(12):005131.
- 48. Aslam M, et al. Reduced right ventricular sarcomere contractility in HFpEF with severe obesity. Circulation. 2020;143(9):965–7.
- 49. Hahn V, et al. Myocardial gene expression signatures in human heart failure with preserved ejection fraction. Circulation. 2021;143(2):120–34.
- 50. Ingwall J. ATP and the Heart. 1st ed. Boston, MA: Springer; 2002.
- 51. Peterson LR, et al. Efect of obesity and insulin resistance on myocardial substrate metabolism and efficiency in young women. Circulation. 2004;109(18):2191–6.
- 52. Rijzewijk LJ, et al. Altered myocardial substrate metabolism and decreased diastolic function in nonischemic human diabetic cardiomyopathy : studies with cardiac positron emission tomography and magnetic resonance imaging. Journal of the American College of Cardiology. 2009;54(16):1524–32.
- 53. Dávila-Román VG, et al. Altered myocardial fatty acid and glucose metabolism in idiopathic dilated cardiomyopathy. Journal of the American College of Cardiology. 2002;40(2):271–7.
- 54. Tuunanen H, et al. Free fatty acid depletion acutely decreases cardiac work and efficiency in cardiomyopathic heart failure. Circulation. 2006;114(20):2130–7.
- 55. Aubert G, et al. The failing heart relies on Ketone bodies as a fuel. Circulation. 2016;133(8):698–705.
- 56. Mori J, et al. Agonist-induced hypertrophy and diastolic dysfunction are associated with selective reduction in glucose oxidation. Circulation: Heart Failure. 2012;5(4):493–503.
- 57. Mori J, et al. ANG II causes insulin resistance and induces cardiac metabolic switch and inefficiency: A critical role of PDK4. Am J Phys Heart Circ Phys. 2013.
- 58. Weitzel LB, et al. Left ventricular assist device effects on metabolic substrates in the failing heart. PLoS ONE. 2013;8(4):e60292.
- 59. Moravec, J., El Alaoui-Talibi, Z., Moravec, M. & Guendouz, A. Oxygen transport to tissue XVII advances in experimental medicine and biology, 205–212. In: Control of oxidative metabolism in volume-overloaded rat hearts. In: Ince C, Kesecioglu J, Telci L, Akpir K, editors Boston, MA: Springer US; 1996. [https://link.](https://link.springer.com/chapter/10.1007/978-1-4613-0333-6_25) [springer.com/chapter/10.1007/978-1-4613-0333-6_25](https://link.springer.com/chapter/10.1007/978-1-4613-0333-6_25)
- 60. Zhabyeyev P, et al. Pressure-overload-induced heart failure induces a selective reduction in glucose oxidation at physiological afterload. Cardiovas Res. 2013
- 61. Zhang L, et al. Cardiac insulin-resistance and decreased mitochondrial energy production precede the development of systolic heart failure after pressure-overload hypertrophy. Circulation: Heart Failure. 2013;6(5):1039–48.
- 62. Jaswal JS, Keung W, Wang W, Ussher JR, Lopaschuk GD. Molecular defects in cardiovascular disease 153–175. In: Dhalla NS, Nagano M, Ostadal B, editors. Molecular changes in fatty acid oxidation in the failing heart. New York, NY: Springer; 2011. [https://link.springer.com/chapter/10.1007/978-1-4419-](https://link.springer.com/chapter/10.1007/978-1-4419-7130-2_12) [7130-2_12](https://link.springer.com/chapter/10.1007/978-1-4419-7130-2_12)
- 63. Leong HS, et al. Accelerated rates of glycolysis in the hypertrophied heart: Are they a methodological artifact. American Journal of Physiology Endocrinology and Metabolism. 2002;282(5):E1039-45.
- 64. Diakos N, et al. vidence of glycolysis upregulation and pyruvate mitochondrial oxidation mismatch during mechanical unloading of the failing human heart: Implications for cardiac reloading and conditioning. Journal of the American College of Cardiology. 2016;67(13-Supplement):1274.
- 65. Masoud WG, et al. Failing mouse hearts utilize energy inefficiently and beneft from improved coupling of glycolysis and glucose oxidation. Cardiovas Res. 2014;101(1)
- 66. Bedi KC, et al. Evidence for intramyocardial disruption of lipid metabolism and increased myocardial ketone utilization in advanced human heart failure. Circulation. 2016;133(8):706–16.
- 67. Funada J, et al. Substrate utilization by the failing human heart by direct quantifcation using arterio-venous blood sampling. PLoS ONE. 2009;4(10):e7533.
- 68. Sun H, et al. Catabolic defect of branched-chain amino acids promotes heart failure. Circulation. 2016;133(21):2038–49.
- 69. Wang W, et al. Defective branched chain amino acid catabolism contributes to cardiac dysfunction and remodeling following myocardial infarction. American Journal of Physiology-Heart and Circulatory Physiology. 2016;311(5):H1160–9.
- 70. Jong KAD, Lopaschuk GD. Complex energy metabolic changes in heart failure with preserved ejection fraction and heart failure with reduced ejection fraction. Canadian Journal of Cardiology. 2017;33(7):860–71.
- 71. Spindler M, et al. Diastolic dysfunction and altered energetics in the alphaMHC403 /+ mouse model of familial hypertrophic cardiomyopathy. The Journal of clinical investigation. 1998;101(8):1775–83.
- 72. Phan T, et al. Heart failure with preserved ejection fraction is characterized by dynamic impairment of active relaxation and contraction of the left ventricle on exercise and associated with myocardial energy defciency. Journal of the American College of Cardiology. 2009;54(5):402–9.
- 73. Heggermont W, Papageorgiou A, Heymans S, van Bilsen M. Metabolic support for the heart: Complementary therapy for heart failure? European Journal of Heart Failure. 2016;18(12):1420–9.
- 74. Paulus W, Zile M. From systemic infammation to myocardial fbrosis. Circulation Research. 2021;128(10):1451–67.
- 75. Hulsmans M, et al. Cardiac macrophages promote diastolic dysfunction. Journal of Experimental Medicine. 2018;215(2):423–40.
- 76. Paulus W, Tschöpe C. A novel paradigm for heart failure with preserved ejection fraction: Comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial infammation. Journal of the American College of Cardiology. 2013;62(4):263–71.
- 77. DuBrock H, AbouEzzeddine O, Redfeld M. High-sensitivity C-reactive protein in heart failure with preserved ejection fraction. PLoS ONE. 2018;13(8):0201836.
- 78. S S.-v, W, et al. Proteomic evaluation of the comorbidity-infammation paradigm in heart failure with preserved ejection fraction. Circulation. 2020;142(21):2029–44.
- 79. Tromp J, et al. Identifying pathophysiological mechanisms in heart failure with reduced versus preserved ejection fraction. Journal of the American College of Cardiology. 2018;72(10):1081–90.
- 80. Van Wijk S, et al. Circulating biomarkers of distinct pathophysiological pathways in heart failure with preserved vs. reduced left ventricular ejection fraction. European Journal of Heart Failure. 2015;17(10):1006–14.
- 81. Borbély A, et al. Hypophosphorylation of the stif N2B titin isoform raises cardiomyocyte resting tension in failing human myocardium. Circulation Research. 2009;104(6):780–6.
- 82. L vH, et al. Myocardial structure and function difer in systolic and diastolic heart failure. Circulation. 2006;25(113):1966–73.
- 83. Zeng H, Chen J-X. Sirtuin 3, endothelial metabolic reprogramming and heart failure with preserved ejection fraction. Journal of Cardiovascular Pharmacology. 2019;74(4):315–23.
- 84. Zile M, et al. Myocardial stiffness in patients with heart failure and a preserved ejection fraction. Circulation. 2015;131(14):1247–59.
- 85. Hahn V, et al. Endomyocardial biopsy characterization of heart failure with preserved ejection fraction and prevalence of cardiac amyloidosis. JACC Heart failure. 2020;8(9):712–24.
- 86. Hahn VS, et al. Endomyocardial biopsy characterization of heart failure with preserved ejection fraction and prevalence of cardiac Amyloidosis. JACC: Heart Failure. 2020;8(9):712–24.
- 87. Das S, et al. Transcriptomics of cardiac biopsies reveals differences in patients with or without diagnostic parameters for heart failure with preserved ejection fraction. Scientifc Reports. 2019;9(1):3179.
- 88. Franssen C, et al. Myocardial microvascular inflammatory endothelial activation in heart failure with preserved ejection fraction. JACC: Heart Failure. 2016;4(4):312–24.
- 89. Westermann D, et al. Cardiac inflammation contributes to changes in the extracellular matrix in patients with heart failure and normal ejection fraction. Circulation Heart Failure. 2011;4(1):44–52.
- 90. Banypersad SM, Moon JC, Whelan C, Hawkins PN, Wechalekar AD. Updates in Cardiac Amyloidosis : A review. Journal of the American Heart Association. 2012;1(2):e000364.
- 91. Lewis AJM, et al. Noninvasive immunometabolic cardiac infammation imaging using hyperpolarized magnetic resonance. Circulation Research. 2018;122(8):1084–93.
- 92. Nahrendorf M, et al. Imaging cardiovascular and lung macrophages with the positron emission tomography sensor 64Cu-Macrin in Mice, Rabbits, and Pigs. Circulation Cardiovascular Imaging. 2020;13(10):e010586.
- 93. Lewis G, et al. Pirfenidone in heart failure with preserved ejection fraction: A randomized phase 2 trial. Nature Medicine. 2021;27(8):1477–82.
- 94. Lewis AJM, Burrage MK, Ferreira VM. Cardiovascular magnetic resonance imaging for infammatory heart diseases. Cardiovascular Diagnosis and Therapy. 2020;10(3):598–609.
- 95. Ferreira V, et al. Cardiovascular magnetic resonance in nonischemic myocardial inflammation: Expert recommendations. Journal of the American College of Cardiology. 2018;72(24):3158–76.
- 96. Ferreira V, et al. Native T1-mapping detects the location, extent and patterns of acute myocarditis without the need for gadolinium contrast agents. Journal of cardiovascular magnetic resonance: Official Journal of the Society for Cardiovascular Magnetic Resonance. 2014;16(1):36–36.
- 97. Ferreira V, et al. Pheochromocytoma is characterized by catecholamine-mediated myocarditis, focal and difuse myocardial fbrosis, and myocardial dysfunction. Journal of the American College of Cardiology. 2016;67(20):2364–74.
- 98. Greulich S, et al. T1 and T2 mapping for evaluation of myocardial involvement in patients with ANCA-associated vasculitides. J Cardiovasc Magn Reson Off J Soc Cardiovasc Magn Reson. 2017;19(1):6.
- 99. Guo Q, et al. Early detection of silent myocardial impairment in drug-naive patients with new-onset systemic lupus erythematosus: A three-center prospective study. Arthritis Rheumatol Hoboken NJ. 2018;70(12):2014–24.
- 100. Mavrogeni S, et al. Cardiovascular magnetic resonance detects silent heart disease missed by echocardiography in systemic lupus erythematosus. Lupus. 2018;27(4):564–71.
- 101. Ntusi N, et al. Subclinical myocardial infammation and difuse fbrosis are common in systemic sclerosis – a clinical study using myocardial T1-mapping and extracellular volume quantification. Journal of cardiovascular magnetic resonance: Official Journal of the Society for Cardiovascular Magnetic Resonance. 2014;16(1):21.
- 102. Ntusi NAB, et al. Difuse myocardial fbrosis and infammation in rheumatoid arthritis : Insights From CMR T1 Mapping. JACC Cardiovascular Imaging. 2015;8(5):526–36.
- 103. Ntusi N, et al. Anti- TNF modulation reduces myocardial infammation and improves cardiovascular function in systemic rheumatic diseases. International Journal of Cardiology. 2018;270:253–9.
- 104. Puntmann V, et al. T1 and T2 mapping in recognition of early cardiac involvement in systemic sarcoidosis. Radiology. 2017;285(1):63–72.
- 105. Zakeri R, Cowie MR. Heart failure with preserved ejection fraction: Controversies, challenges and future directions. Heart (British Cardiac Society). 2018;104(5):377–84.
- 106. Quarta G, et al. Cardiac magnetic resonance in heart failure with preserved ejection fraction: Myocyte, interstitium, microvascular, and metabolic abnormalities. European Journal of Heart Failure. 2020;22(7):1065–75.
- 107. Kanagala P, et al. Diagnostic and prognostic utility of cardiovascular magnetic resonance imaging in heart failure with preserved ejection fraction – implications for clinical trials. Journal of Cardiovascular Magnetic Resonance. 2018;20(1):4.
- 108. Hundertmark M. Should CMR be the default imaging modality in clinical trials for heart failure? Cardiovascular Diagnosis and Therapy. 2020;10(3):554.
- 109. Muraru D, Badano LP, Ermacora D, Piccoli G, Iliceto S. Sources of variation and bias in assessing left ventricular volumes and dyssynchrony using three-dimensional echocardiography. The International Journal of Cardiovascular Imaging. 2012;28(6):1357–68.
- 110. Shimada YJ, Shiota T. A Meta-analysis and investigation for the source of bias of left ventricular volumes and function by three-dimensional echocardiography in comparison with

magnetic resonance imaging. The American Journal of Cardiology. 2011;107(1):126–38.

- 111. Browning JR, Hertzberg JR, Schroeder JD, Fenster BE. 4D fow assessment of vorticity in right ventricular diastolic dysfunction. Bioengineering. 2017;4(2):30.
- 112. Garg P, et al. Left ventricular blood fow kinetic energy after myocardial infarction - insights from 4D flow cardiovascular magnetic resonance. Journal of Cardiovascular Magnetic Resonance. 2018;20(1):61.
- 113. Abe H, et al. Contrast echocardiography for assessing left ventricular vortex strength in heart failure: A prospective cohort study. European Heart Journal Cardiovascular Imaging. 2013;14(11):1049–60.
- 114. Cherry Kemmerling, E. In vitro pressure measurements across an interatrial shunt for HFpEF treatment. Cardiovasc Eng Technol (2022)
- 115. Chirinos JA, et al. Efects of organic and inorganic nitrate on aortic and carotid haemodynamics in heart failure with preserved ejection fraction. European Journal of Heart Failure. 2017;19(11):1507–15.
- 116. Chamsi-Pasha MA, Zhan Y, Debs D, Shah DJ. CMR in the evaluation of diastolic dysfunction and phenotyping of HFpEF : Current role and future perspectives. JACC: Cardiovascular Imaging. 2020;13(1, Part 2):283–96.
- 117. Khan JN, et al. Comparison of cardiovascular magnetic resonance feature tracking and tagging for the assessment of left ventricular systolic strain in acute myocardial infarction. European Journal of Radiology. 2015;84(5):840–8.
- 118. Satriano A, et al. Clinical feasibility and validation of 3D principal strain analysis from cine MRI : Comparison to 2D strain by MRI and 3D speckle tracking echocardiography. The International Journal of Cardiovascular Imaging. 2017;33(12):1979–92.
- 119. Bhalodiya JM, et al. Hierarchical template matching for 3D myocardial tracking and cardiac strain estimation. Scientifc Reports. 2019;9(1):12450.
- 120. Nitzken, M, et al. Improving full-cardiac cycle strain estimation from tagged CMR by accurate modeling of 3D image appearance characteristics. 2012 9th IEEE International Symposium on Biomedical Imaging (ISBI); 2012. p. 462–5
- 121. Tanacli R, et al. Multilayer myocardial strain improves the diagnosis of heart failure with preserved ejection fraction. ESC Heart Failure. 2020;7(5):3240–5.
- 122. Kammerlander AA, et al. Global longitudinal strain by CMR feature tracking is associated with outcome in HFPEF. JACC: Cardiovascular Imaging. 2019;12(8-Part-1):1585–7.
- 123. Haaf P, et al. Cardiac T1 mapping and extracellular volume (ECV) in clinical practice: A comprehensive review. Journal of Cardiovascular Magnetic Resonance. 2016;18(1):89.
- 124. Rommel K-P, et al. Extracellular volume fraction for characterization of patients with heart failure and preserved ejection fraction. Journal of the American College of Cardiology. 2016;67(15):1815–25.
- 125. Vogelsberg H, et al. Cardiovascular magnetic resonance in clinically suspected cardiac Amyloidosis. Journal of the American College of Cardiology. 2008;51(10):1022–30.
- 126. Karamitsos TD, et al. Noncontrast T1 mapping for the diagnosis of cardiac Amyloidosis. JACC: Cardiovascular Imaging. 2013;6(4):488–97.
- 127. Takagi T. Diastolic stress echocardiography. Journal of Echocardiography. 2017;15(3):99–109.
- 128. Simonovic D, et al. Exercise-induced B-lines in heart failure with preserved ejection fraction occur along with diastolic function worsening. ESC Heart Failure. 2021;8(6):5068–80.
- 129. Holland DJ, Prasad SB, Marwick TH. Contribution of exercise echocardiography to the diagnosis of heart failure with preserved ejection fraction (HFpEF). Heart. 2010;96(13):1024–8.
- 130. Skaluba SJ, Litwin SE. Mechanisms of exercise intolerance. Circulation. 2004;109(8):972–7.
- 131. Gulati M, et al. The prognostic value of a Nomogram for exercise capacity in women. New England Journal of Medicine. 2005;353(5):468–75.
- 132. Morris CK, et al. Nomogram based on metabolic equivalents and age for assessing aerobic exercise capacity in men. Journal of the American College of Cardiology. 1993;22(1):175–82.
- 133. Wilson JR, Rayos G, Yeoh TK, Gothard P, Bak K. Dissociation between exertional symptoms and circulatory function in patients with heart failure. Circulation. 1995;92(1):47–53.
- 134. Palazzuoli A, Ruocco G, Beltrami M, Nuti R, Cleland JG. Combined use of lung ultrasound, B-type natriuretic peptide, and echocardiography for outcome prediction in patients with acute HFrEF and HFpEF. Clinical Research in Cardiology. 2018;107(7):586–96.
- 135. Donal E, et al. Value of exercise echocardiography in heart failure with preserved ejection fraction: A substudy from the KaRen study. European Heart Journal Cardiovascular Imaging. 2016;17(1):106–13.
- 136. Platz E, et al. Dynamic changes and prognostic value of pulmonary congestion by lung ultrasound in acute and chronic heart failure: A systematic review. European Journal of Heart Failure. 2017;19(9):1154–63.
- 137. Burrage MK, et al. Energetic basis for exercise-induced pulmonary congestion in heart failure with preserved ejection fraction. Circulation. 2021;144(21):1664–78.
- 138. Pieske B, et al. How to diagnose heart failure with preserved ejection fraction: The HFA-PEFF diagnostic algorithm: A consensus recommendation from the Heart Failure Association (HFA) of the European Society of Cardiology (ESC). European Heart Journal. 2019;40(40):3297–317.
- 139. Sugimoto T, et al. Left atrial function dynamics during exercise in heart failure : Pathophysiological implications on the right heart and exercise ventilation inefficiency. JACC Cardiovascular Imaging. 2017;10(10 Pt B):1253–64.
- 140. Borlaug BA, Nishimura RA, Sorajja P, Lam CS, Redfeld MM. Exercise hemodynamics enhance diagnosis of early heart failure with preserved ejection fraction. Circulation: Heart Failure. 2010;3(5):588–95.
- 141. Ho JE, et al. Diferential clinical profles, exercise responses, and outcomes associated with existing HFpEF defnitions. Circulation. 2019;140(5):353–65.
- 142. Borlaug BA, et al. Impaired chronotropic and vasodilator reserves limit exercise capacity in patients with heart failure and a preserved ejection fraction. Circulation. 2006;114(20):2138–47.
- 143. Nadruz W, et al. Prognostic value of cardiopulmonary exercise testing in heart failure with reduced, midrange, and preserved ejection fraction. Journal of the American Heart Association. 2017;6(11):e006000.
- 144. Nayor M, et al. Clinical and hemodynamic associations and prognostic implications of ventilatory efficiency in patients with preserved left ventricular systolic function. Circulation: Heart Failure. 2020;13(5):e006729.
- 145. Ho JE, Nguyen C, Lewis GD. Unmasking nonpreserved heart structure, function, and energetics in heart failure with preserved ejection fraction with magnetic resonance imaging coupled with exercise. Circulation. 2021;144(21):1679–82.
- 146. Levelt E, et al. Cardiac energetics, oxygenation, and perfusion during increased workload in patients with type 2 diabetes mellitus. European Heart Journal. 2015;23(1522–9645 (Electronic)):ehv442.
- 147. Levelt E, et al. Relationship between left ventricular structural and metabolic remodeling in type 2 diabetes. Diabetes. 2016;65(1):44–52.
- 148. Phan TT, et al. Heart failure with preserved ejection fraction is characterized by dynamic impairment of active relaxation and contraction of the left ventricle on exercise and associated with myocardial energy deficiency. Journal of the American College of Cardiology. 2009;54(5):402–9.
- 149. AbouEzzeddine OF, et al. Myocardial energetics in heart failure with preserved ejection fraction. Circulation: Heart Failure. 2019;12(10):e006240.
- 150. Hasegawa S, et al. Effects of cardiac energy efficiency in diastolic heart failure : Assessment with positron emission tomography with 11C-Acetate. Hypertension Research. 2008;31(6):1157–62.
- 151. Brown M, Marshall DR, Sobel BE, Bergmann SR. Delineation of myocardial oxygen utilization with carbon-11-labeled acetate. Circulation. 1987;76(3):687–96.
- 152. Chatterjee, N A, Steiner J, Lewis GD. It is time to look at heart failure with preserved ejection fraction from the right side. Circulation. 2014;130(25):2272–7.
- 153. Omote K, Verbrugge FH, Borlaug BA. Heart failure with preserved ejection fraction: Mechanisms and treatment strategies. Annual Review of Medicine. 2021;73(1):null.
- 154. Rider OJ, Tyler DJ, O R, D T. Clinical implications of cardiac hyperpolarized magnetic resonance imaging. Journal of cardiovascular magnetic resonance: official journal of the Society for Cardiovascular Magnetic Resonance. 2013;15(1):93.
- 155. Zelt JG, Chaudhary KR, Cadete VJ, Mielniczuk LM, Stewart DJ. Medical therapy for heart failure associated with pulmonary hypertension. Circulation Research. 2019;124(11):1551–67.
- 156. Heggermont WA, Papageorgiou A-P, Heymans S, van Bilsen M. Metabolic support for the heart: Complementary therapy for heart failure? European Journal of Heart Failure. 2016;18(12):1420–9.
- 157. Apps A, et al. Hyperpolarised magnetic resonance for in vivo real-time metabolic imaging. Heart. 2018;104(18):1484–91.
- 158. Bizino MB, Hammer S, Lamb HJ. Metabolic imaging of the human heart: Clinical application of magnetic resonance spectroscopy. Heart. 2014;100(11):881–90.
- 159. Di Carli MF, et al. Value of metabolic imaging with positron emission tomography for evaluating prognosis in patients with coronary artery disease and left ventricular dysfunction. The American Journal of Cardiology. 1994;73(8):527–33.
- 160. DiBella E. New MRI methods applied to heart failure with preserved ejection fraction (HFpEF). Clinical Trial Registration, study/NCT04600115, clinicaltrials.gov. 2020
- 161. Apps A, et al. Proof-of- principle demonstration of direct metabolic imaging following myocardial infarction using hyperpolarized 13C CMR. JACC: Cardiovasc Imaging. 2021;0(0)
- 162. Rider OJ, et al. Noninvasive in vivo assessment of cardiac metabolism in the healthy and diabetic human heart using hyperpolarized 13C MRI. Circ Res. 2020;725–736
- 163. Miller JJ, et al. Robust and high resolution hyperpolarized metabolic imaging of the rat heart at 7 T with 3D spectral-spatial EPI. Magnetic Resonance in Medicine. 2016;75(4):1515–24.
- 164. Zinman B, et al. Empagliflozin, cardiovascular outcomes, and mortality in type 2 diabetes. N Engl J Med. 2015;373(22):2117–28.
- 165. Mudaliar S, Alloju S, Henry RR. Can a shift in fuel energetics explain the beneficial cardiorenal outcomes in the EMPA-REG OUTCOME study? A unifying hypothesis. Diabetes Care. 2016;39(7):1115–22.
- 166. Wagdy K, Nagy S. EMPEROR-Preserved : SGLT2 inhibitors breakthrough in the management of heart failure with preserved ejection fraction. Global Cardiology Science & Practice. 2021;2021(3):e202117.
- 167. Cleland JGF, et al. The perindopril in elderly people with chronic heart failure (PEP-CHF) study. European Heart Journal. 2006;27(19):2338–45.
- 168. Granger CB, et al. Effects of candesartan in patients with chronic heart failure and reduced left-ventricular systolic function intolerant to angiotensin-converting-enzyme inhibitors: The CHARM-Alternative trial. Lancet (London, England). 2003;362(9386):772–6.
- 169. Massie BM, et al. Irbesartan in patients with heart failure and preserved ejection fraction. The New England Journal of Medicine. 2008;359(23):2456–67.
- 170. Pitt B, et al. Spironolactone for heart failure with preserved ejection fraction. The New England Journal of Medicine. 2014;370(15):1383–92.
- 171. Solomon SD, et al. Angiotensin- neprilysin inhibition in heart failure with preserved ejection fraction. The New England Journal of Medicine. 2019;381(17):1609–20.
- 172. McMurray JJV, et al. Dapaglifozin in patients with heart failure and reduced ejection fraction. The New England Journal of Medicine. 2019;381(21):1995–2008.
- 173. Packer M, et al. Cardiovascular and renal outcomes with empaglifozin in heart failure. The New England Journal of Medicine. 2020;383(15):1413–24.
- 174. Zelniker TA, et al. SGLT2 inhibitors for primary and secondary prevention of cardiovascular and renal outcomes in type 2 diabetes: A systematic review and meta-analysis of cardiovascular outcome trials. Lancet (London, England). 2019;393(10166):31–9.
- 175. Chen C, et al. Pharmacokinetics, biotransformation, distribution and excretion of empaglifozin, a sodium-glucose co-transporter (SGLT 2) Inhibitor, in Mice, Rats, and Dogs. Journal of Pharmaceutics & Drug Development. 2015;3(3):1.
- 176. Chung YJ, et al. Off-target effects of sodium-glucose cotransporter 2 blockers: Empaglifozin does not inhibit Na +/ H + exchanger-1 or lower [Na +]i in the heart. Cardiovascular Research. 2020;117(14):2794–806.
- 177. Santos GCG, et al. Empaglifozin induces a myocardial metabolic shift from glucose consumption to ketone metabolism that mitigates adverse cardiac remodeling and improves myocardial contractility. Journal of the American College of Cardiology. 2018;71(11–Supplement):A674.
- 178. Santos GCG, et al. Empaglifozin ameliorates adverse left ventricular remodeling in nondiabetic heart failure by enhancing myocardial energetics. Journal of the American College of Cardiology. 2019;73(15):1931–44.
- 179. Kappel BA, et al. Efect of empaglifozin on the metabolic signature of patients with type 2 diabetes mellitus and cardiovascular disease. Circulation. 2017;136(10):969–72.
- 180. Nishimura R, et al. Efect of empaglifozin on free fatty acids and ketone bodies in Japanese patients with type 2 diabetes mellitus : A randomized controlled trial. Advances in Therapy. 2019;36(10):2769–82.
- 181. Sato K, et al. Insulin, ketone bodies, and mitochondrial energy transduction. FASEB J. 1995;9(8):651–8.
- 182. Leichman JG, et al. Association of plasma free fatty acids and left ventricular diastolic function in patients with clinically severe obesity. The American Journal of Clinical Nutrition. 2006;84(2):336–41.
- 183. Verma S, McMurray JJV. SGLT2 inhibitors and mechanisms of cardiovascular beneft: A state-of-the-art review. Diabetologia. 2018;61(10):2108–17.
- 184. Dyck JRB, et al. Cardiac mechanisms of the beneficial effects of SGLT2 inhibitors in heart failure: Evidence for potential oftarget efects. Journal of Molecular and Cellular Cardiology. 2022;167:17–31.
- 185. Berezin AE, Berezin AA, Lichtenauer M. Emerging role of adipocyte dysfunction in inducing heart failure among obese patients with prediabetes and known diabetes Mellitus. Front Cardiovasc Med. 2020;7
- 186. Abdurrachim D, et al. Empaglifozin reduces myocardial ketone utilization while preserving glucose utilization in diabetic hypertensive heart disease: A hyperpolarized 13C magnetic resonance spectroscopy study. Diabetes, Obesity and Metabolism. 2019;21(2):357–65.
- 187. Madonna R. Exploring the mechanisms of action of glifozines in heart failure and possible implications in pulmonary hypertension. Vascular Pharmacology. 2021;138:106839.
- 188. Tadic M, et al. New antidiabetic therapy and HFpEF : Light at the end of tunnel? Heart Failure Rev. 2021
- 189. Kondo H, et al. Efects of canaglifozin on human myocardial redox signalling: Clinical implications. European Heart Journal. 2021;42(48):4947–60.
- 190. Antonopoulos AS, Antoniades C. The role of epicardial adipose tissue in cardiac biology: Classic concepts and emerging roles. The Journal of Physiology. 2017;595(12):3907–17.
- 191. Santos D, Carvalho E. Adipose-related microRNAs as modulators of the cardiovascular system: The role of epicardial adipose tissue. The Journal of Physiology. 2022;600(5):1171–87.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.