

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Contents lists available at ScienceDirect

# European Journal of Medicinal Chemistry

journal homepage: www.elsevier.com/locate/ejmech



# Review article Potential RNA-dependent RNA polymerase (RdRp) inhibitors as prospective drug candidates for SARS-CoV-2



Mohamed S. Bekheit<sup>a</sup>, Siva S. Panda<sup>b,\*</sup>, Adel S. Girgis<sup>a,\*\*</sup>

<sup>a</sup> Department of Pesticide Chemistry, National Research Centre, Dokki, Giza, 12622, Egypt
 <sup>b</sup> Department of Chemistry and Physics, Augusta University, Augusta, GA, 30912, USA

	A	R	Т	I	С	L	Е	I	Ν	F	0
--	---	---	---	---	---	---	---	---	---	---	---

Keywords: Polymerase RdRp SARS-CoV-2 COVID-19 Remdesivir Favipiravir Molnupiravir Computational studies

# ABSTRACT

The SARS-CoV-2 pandemic is considered as one of the most disastrous pandemics for human health and the world economy. RNA-dependent RNA polymerase (RdRp) is one of the key enzymes that control viral replication. RdRp is an attractive and promising therapeutic target for the treatment of SARS-CoV-2 disease. It has attracted much interest of medicinal chemists, especially after the approval of Remdesivir. This study highlights the most promising SARS-CoV-2 RdRp repurposed drugs in addition to natural and synthetic agents. Although many in silico predicted agents have been developed, the lack of in vitro and in vivo experimental data has hindered their application in drug discovery programs.

# 1. Introduction

Infectious diseases are still one of the major public health challenges. Diverse microorganisms including viruses, bacteria, fungi, and parasites are the main cause of infectious diseases [1]. Many epidemics and pandemics due to HIV/AIDS, avian flu, swine flu, Ebola, Zika, SARS-CoV-2, and monkeypox viruses have occurred in the last few decades [2,3]. The entire global population is still suffering from emergent and recurring infectious diseases caused by many microorganisms [4]. About two years ago, the WHO (World Health Organization) announced COVID-19 (coronavirus disease 2019) as a public health concern (March 11, 2020) due to the widespread global impact of an infectious SARS-CoV-2 (severe acute respiratory syndrome coronavirus-2) parasite. More than 6.6 million deaths and 639 million infected patients have been reported till the beginning of December 2022 [5,6]. The first identified case of SARS-CoV-2 was reported in the local fish and wild animal market in Wuhan City, China [7]. SARS-CoV-2 is a RNA zoonotic virus (family: Coronaviridae, order: Nidovirales, genus: Betacoronavirus). It is mainly found in bats. After this virus was transferred to humans in China, it then widely spread to all other countries, leading to a global pandemic [8–10]. Four genera ( $\alpha$ ,  $\beta$ ,  $\delta$ , and  $\gamma$ ) of coronavirus are known [11]. Several waves of SARS-CoV-2 infections have been recognized due to viral mutations. The most important variants of the virus are Beta,

Gamma, Delta, and Omicron variants. The new Omicron variant indicates that the epidemic/pandemic is far from its end due to its efficient human-to-human transmission [12]. The main symptoms due to the infection from this variant are fever, dry cough, diarrhea, and shortness of breath besides blood clotting and stroke in severe conditions [10,13]. Repurposed drugs and vaccination substantially helped to overcome the pandemic and retain the socioeconomic status and human life to normal [14].

SARS-CoV-2 belongs to a positive-sense single-stranded RNA viral group (ssRNA(+)). It can infect humans and largely spreads through close contact and by breathing droplets generated by coughing/sneezing. Its genetic material can directly act as a viral messenger RNA (mRNA) and translate into viral proteins in the host cell [15–17]. Several enzymes are involved in coronaviral replication. Thus, targeting these enzymes in drug discovery efforts might lead to promising antiviral drugs [18]. The SARS-CoV-2 nonstructural proteins including RNA-dependent RNA polymerase (RdRp) and main protease (M<sup>pro</sup>) are crucial for viral genomic transcription and replication [17,19]. The crystal structures of both RdRP and M<sup>pro</sup> are available in the Protein Data Bank (PDB) [20–22].

The enzyme RdRp is encoded in all RNA viruses. Viral RdRp is the main target for developing potent antiviral agents against SARS-CoV-2, not only due to its ability to accelerate the replication of RNA but also

\* Corresponding author.

\*\* Corresponding author.

https://doi.org/10.1016/j.ejmech.2023.115292

Received 31 December 2022; Received in revised form 14 March 2023; Accepted 15 March 2023 Available online 18 March 2023 0223-5234/© 2023 Elsevier Masson SAS. All rights reserved.

E-mail addresses: sspanda12@gmail.com, SIPANDA@augusta.edu (S.S. Panda), as.girgis@nrc.sci.eg, girgisas10@yahoo.com (A.S. Girgis).



Remdesivir

Remdesivir triphosphate (RDV-TP)

Fig. 1. Strucure of Remdesivir prodrug and its active triphosphate derivatives (RDV-TP).



Fig. 2. Structures of W116, 116-N1 and 116-NTP.



Fig. 3. Structures of GS-621763 and GS-441524.



Favipiravir

Favipiravir-ribofuranosyl-triphosphate



due to the lack of RdRp closely related host cell counterparts. Theoretically, the designed agent will selectively target the viral RdRp with no off-target side effects [23].

RdRp inhibitors are categorized into two classes based on their



Fig. 5. Molnupiravir, COVID-19 therapeutical agent.



Fig. 6. Galidesivir(BCX4430) RdRNAp replication inhibitor.

structure and location for binding to RdRp: nucleoside analog inhibitors (NIs) and non-nucleoside analog inhibitors (NNIs). Structurally, NIs are known to bind the RdRp protein at the enzyme active site, NNIs bind to the allosteric sites of RdRp. Several studies on RdRp inhibitors with various applications have been reported. Tian et al. reported a recent review for RdRp inhibitors however, most of the mentioned analogs are for pyrimidine-containing compounds (either NIs and NNIs) [24]. This current review adopts wider scope summarizing the natural and synthetic compounds as well as the repurposed drugs/active agents with RdRp inhibitory properties for treating SARS-CoV-2, with various heterocyclic scaffolds. Notably, the screening techniques for coronavirus RdRp are not as simple as those for proteases [25]. With increasing interest, a few studies on RdRp screening for coronavirus have been reported.

The anti-SARS-CoV-2 properties of Galidesivir against different cell cultures.

Cell line	SARS-CoV-2 strains	Assay	Incubation period	EC <sub>50</sub> (μM)	EC <sub>90</sub> (μM)	CC50 (µM)	SI
Caco-2	WA1/2020	VYR <sup>a</sup>	24 h	n.d. <sup>b</sup>	14.19	82.8	5.8
Vero-76	WA1/2020	VYR <sup>a</sup>	24 h	n.d. <sup>b</sup>	10.94	>295.7	>27
		CPE <sup>c</sup>			50.3		5.8
Calu-3	WA1/2020	Imaging	2 h	14.15	n.d.	>50	>3.5

<sup>a</sup> VYR = Viral Yield Reduction.

<sup>b</sup> n.d. = not determined.

<sup>c</sup> CPE = cytopathic effect.



Fig. 7. Ribavirin, a broad spectrum antiviral agent.

# 2. Drug repurposing

Development of new drug(s) usually involves many successive stages which are: design, synthesis, bio-properties investigation, formulation of prototypes, and pre-clinical and clinical trials. Due to the pandemic outbreak, drug repurposing is considered the most accessible and appealing approach for urgent identifying potential therapeutics to control the disaster and save human lives. Drug repurposing means the adoption of an existing broad-spectrum therapeutical entity of potential efficacy and minimal adverse effects for clinical application of the infected patients supported by pre-clinical establishments. Drug repurposing strategy is superior to traditional drug discovery in terms of cost and time reduction. In addition to a lower failure rate compared with the traditional approaches owing to its well-established efficacy, metabolic characteristics, dose determination, and safety or toxicity issues [26-28]. FDA (food and drug administration) approved several drugs of other pathophysiological under the emergency use authorization of which antiviral drugs (remdesivir, penciclovir and favipiravir) and antimalarial drugs (chloroquine, hydroxychloroquine) as anti-COVID-19 agents. Meanwhile, adverse effects humbled the clinical applicability of some of them [29-31].

## 2.1. Remdesivir (RDV, formerly GS-5734)

Yin et al. investigated the inhibition of the RdRp from SARS-CoV-2 by remdesivir. The complex structure of SARS-CoV-2 RdRp and remdesivir reveals that the partial double-stranded RNA template is inserted into the central channel of the RdRp, where remdesivir is covalently incorporated into the primer strand at the first replicated base pair and terminates chain elongation [32]. Remdesivir (RDV) is the first FDA-authorized drug to treat COVID-19 patients with severe conditions (on Oct. 22, 2020) [33,34]. Remdesivir is a phosphoramidite

prodrug of a 1'-cyano-substituted adenosine nucleotide analog, which acts as an RNA polymerase inhibitor (regulate genomic replication) developed initially by Gilead Sciences for Ebola viral infections treatment in 2014 [35]. It has been mentioned as a broad-spectrum antiviral property against various acute viral infections caused by different ssRNA viruses including Hendra virus, Lassa fever virus, Junin virus, Nipah virus, and coronaviruses (SARS-CoV and MERS-CoV) in addition to the efficacy in the therapeutical composition against HCV (hepatitis C virus) and HIV (human immunodeficiency virus) [36–38]. Remdesivir is the first candidate as an anti-COVID-19 drug owing to its broad spectrum as an antiviral agent, especially on coronaviruses (SARS-CoV and MERS-CoV), this encourages Gilead Sciences to repurpose it to treat patients infected with SARS-CoV-2 and approved by FDA in 2020 for treatment patient with COVID-19 infection [34,39].

The broad-spectrum antiviral properties of Remdesivir are attributed to its ability to be metabolized in the host cell as nucleoside triphosphate (RDV-TP). Consequently, it can be integrated within the nascent viral



Alovudine azidothymidine (AZT)

Fig. 9. Structural of Alovudine and azidothymidine (AZT).



Fig. 10. Structure of tenofovir.



Fig. 8. Sofosbuvir and, Daclatasvir, repurposed drugs fir COVID-19.



Fig. 11. Docking poses of Tenofovir, Ribavirin and Sofosbuvir in SARS HCoV RdRp (PDB ID: 6NUR), using AutoDock Vina software [80].



Fig. 12. Structures of AT-527 and AT-9010.

RNA chains due to the RdRp action. Thus, viral RNA synthesis is terminated (Fig. 1) [34,40]. In the VERO-E6 assay, the EC<sub>50</sub> of Remdesivir is 0.77  $\mu$ M, and the CC<sub>50</sub> = >100  $\mu$ M (SI, selectivity index = >129.87) [41].

To improve the  $t_{1/2}$  and in-vivo metabolism of the Remdesivir, Wen et al. replaced the hydrogen in the active molecular group with isotope deuterium as the carbon-deuterium bonds (C–D) are more stable than carbon-hydrogen bonds(C–H) [24,42,43].

New oral RDV derivatives were found to demonstrate SARS-CoV-2 RdRp inhibitors such as VV116 and GS-621763. VV116 (JT001) is a tri-isobutyrate ester RDV prodrug derivative. VV116 is rapidly metabolized into the parent nucleoside (116-N1) in vivo which is intracellularly converted to the active form nucleoside triphosphate that incorporates with RdRp, thus inhibiting SARS-CoV-2 replication [44] (Fig. 2). VV116 exhibited potent activity against a panel of SARS-CoV-2 variants [45,46] with satisfactory safety in phase I studies [44].

Another oral RDV derivative is GS-621763 metabolized in plasma to GS-441524 (Fig. 3). In the lung the triphosphate effectively controls the SARS-CoV2 replication. Both metabolites GS-621763 and GS-441524

possess anti-SARS-CoV-2 properties against different variants. Viral proliferation was completely terminated by oral GS-621763 [47].

#### 2.2. Favipiravir

Another nucleotide analog inhibitor of RdRp is Favipiravir (FPV, favilavi, or Avigan). Favipiravir was originally developed by Toyama Chemicals, Japan, and approved as anti-influenza in 2014 in Japan [48, 49]. Favipiravir is a RdRp inhibitor that has demonstrated antiviral activity against influenza virus H1N1 infection [48]. During the Ebola virus outbreak in West Africa, Favipiravir has been evaluated against human Ebola virus infection. It reveals a promising activity against Ebola virus infection in the mouse model however, poor activity was observed in human Ebola infections [50].

Favipiravir showed effectiveness in controlling the progression of the SARS-COV-2 virus. Patients with mild COVID-19 had a potential clinical recovery rate [51]. While treating severe COVID-19 patients with Favipiravir, an improvement in the lymphocyte count was recorded [52]. Therefore, the clinical tackle was recommended for



Fig. 13. Strucure of promising nucleosides as anti-RdRNAP, anti-ExoN, and anti-SARS-CoV-2 relative to Riboprine-TP and Forodesine-TP.



Fig. 14. Baicalein (1) and Baicalin (2) polyphenolic flavonoids of potential anti-SARS-CoV-2 RdRp.

mild-to-moderate COVID-19 patients with Favipiravir [53].

Favipiravir (pro-drug) metabolized in the body to the active favipiravir-ribofuranosyl-5'-triphosphate (FPV-RTP) via ribosylation and phosphorylation which binds to the active site of the viral RdRp thus terminates the replication of viral RNA (Fig. 4) [54]. In Vero-E6 assay, Favipiravir reveals  $EC_{50} = 61.88$  and  $CC_{50} = >400 \ \mu M$  (SI = 6.46) [41].

# 2.3. Molnupiravir

Molnupiravir (EIDD-1931 or NHC) is the isopropyl ester pro-drug of the ribonucleoside analog  $\beta$ -D-N4-hydroxycytidine [55]. It can inhibit

SARS-CoV-2 replication in human airway epithelial cell cultures and various animal models [56,57]. Clinical trial studies supported the effectiveness of Molnupiravir against SARS-CoV-2 relative to Placebo (inert medication with no therapeutical value) [58]. Molnupiravir was granted emergency use authorization by FDA and MRHA (medicines and healthcare products regulatory agency) for the treatment COVID-19 patients (Fig. 5) [59].

The inhibitory mechanism of Molnupiravir is quite similar to that of Favipiravir. The oral administration of molnupiravir rapidly appears in plasma and is converted to its triphosphate form in cells. The active NHC triphosphate form is incorporated into viral RNA in place of cytosine or



Fig. 15. Baicalein and baicalin in the active site of PDB, ID: 6XQB [99].



Isoginkgetin

Bilobetin

Fig. 16. Strucures for Isoginkgetin, Bilobetin and, Afzelin.

uracil by RdRp, causing a break of viral replication [56,60,61]. The  $IC_{50}$  = 1.965, 0.7556 µM of Molnupiravir are against WT and Omicron SARS-CoV-2 variants, respectively in Calu-3 cells [62].

## 2.4. Galidesivir

Galidesivir (BCX4430) (Fig. 6) is an adenosine analog with RdRp replication inhibitory activities against many RNA viruses including Ebola, Zika, Marburg, and yellow fever (in vitro and in animal models) [63,64]. In Syrian golden hamster models, it was tested as anti-SARS-CoV-2 reducing the lung pathology upon the treatment initiated 24 h before the viral infection, compared with untreated controls [65]. The Galidesivir triphosphate is the active substrate responsible for binding to the active site of the viral RdRp and terminates the replication of viral RNA [66]. Table 1 exhibits the anti-SARS-CoV-2 activity of Galidesivir in different cell cultures [65].

#### 2.5. Ribavirin

Ribavirin (RBV,1- $\beta$ -D-ribofuranosyl-1,2,4-triazole-3-carboxamide) is a guanosine analog (Fig. 7). Ribavirin has a broad antiviral spectrum against in vitro human cell line and several animal models [67]. It was repurposed to treat COVID-19, and reflected a promising activity against SARS-CoV-2 [68,69]. RBV was reported to have a high survival rate in severe patients due to its ability for viral clearance (observational study) [70]. Another clinical trial (Phase II) demonstrated that a combination of RBV, interferon beta-1b, and lopinavir-ritonavir, is a potential treatment for mild to moderate COVID-19 patients [71]. Hemolytic anemia in addition to the reduction of calcium and magnesium in the elder besides the restriction for pregnant women are the serious adverse effects of RBV [72]. The active substrate RBV triphosphate (RBV-TP) is formed via the host cell kinases, which pairs with the uridine triphosphate or cytidine triphosphate in the RNA template resulting in lethal mutagenesis and so prevents viral RNA replication [73]. The  $EC_{50} = 7.1$ ,  $CC_{50} = 160 \mu$ M (SI = 16) are of Ribavirin in Calu-3 assay [74].

#### 2.6. Sofosbuvir and daclatasvir

The replication process of HCV is similar to that of coronavirus, especially at the start of the disease. Anti-HCV therapies such as Sofosbuvir (HCV polymerase inhibitor) [75] and Daclatasvir (RNA replication and virion assembly inhibitor) [76] were suggested to have promising potential in the treatment of COVID-19 (Fig. 8) [74]. Randomized clinical trials on moderate or severe COVID-19 patients demonstrated that sofosbuvir and daclatasvir reducing the hospital stay duration relative to the standard care alone. This is attributed to the daclatasvir/sofosbuvir antiviral efficacy on SARS-CoV-2 replication in respiratory cells [74,77]. The EC<sub>50</sub> = 7.3, 1.1 and CC<sub>50</sub> = 512, 38  $\mu$ M are for Sofosbuvir and Daclatasvir, respectively in Calu-3 assay [74].

Alovudine and AZT which are nucleotide-related structures to Sofosbuvir (2'-fluoro-2'-methyl-UTP) were investigated as anti-SARS-CoV RdRp-inhibitors (Fig. 9). They terminate RNA synthesis and



Fig. 17. 3D-conformations of (A) afzelin, (B) bilobetin, (C) isoginkgetin, (D) chloroquine, (E) remdesivir, and (F) lopinavir in PDB: 6 M71 [100].



Fig. 18. Rutin and Nicotiflorin, promising anti-SARS-CoV-2 based on in silico studies.

replication of the virus in model polymerase extension experiments [78].

# 2.7. Tenofovir

Tenofovir is a broad-spectrum antiviral drug active against the HIV and hepatitis B virus (HBV). The active triphosphate form of the tenofovir diphosphate acts as a terminator of viral RNA subsequent polymerase synthesis (Fig. 10) [79,80]. Few in vitro studies on tenofovir or its prodrug formulations have been reported. The results are not promising and are contradictory [81,82]. Although these reports, in silico studies mentioned that tenofovir binds strongly to SARS-CoV-2 RdRp, with binding energies close to other successful drugs (Remdesivir, Galidesivir, Ribavirin, and Sofosbuvir) with no supporting biochemical observations (Fig. 11) [83].



Fig. 19. Nicotiflorin and rutin in the active site of PDB: 6M71 [101].



Fig. 20. Ellagic acid, p-Coumaric acid, Kaempferol, and Quercetin, promising anti- SARS-CoV-2 supported by in silico studies.



Fig. 21. Docking confirmations of (a) ellagic acid, (b) hesperetin, and (c) kaempferol in PDB: 6M71 [103].



Theaflavin,  $R^1 = R^2 = H$ Theaflavin-3,3'-di-O-gallate,  $R^1 = R^2$  = galloyl

Fig. 22. Theaflavin and theaflavin-3,3'-di-O-gallate.



Fig. 23. 2D-interaction diagram of Theaflavin and SARS-CoV-2 RdRp by the Blind Dock server [106].

#### 2.8. AT-527

AT-527 is an orally available prodrug of a guanosine nucleotide analog that acts as a potent broad-spectrum anti-coronavirus inhibitor in a variety of cell lines by targeting the RdRp activity (Fig. 12) [84]. AT-527 is converted by cellular enzymes to the active triphosphate metabolite, AT-9010. AT-527 recently entered phase III clinical trials to treat COVID-19 [85]. AT-9010 can bind to the active site of RdRp exhibiting promising efficacy against COVID-19 [86].

Nucleoside analogs were used in searching for potent anti-SARS-CoV-2 inhibitors with both SARS-CoV-2-RdRp and SARS-CoV-2 exonuclease (ExoN) dual inhibitory enzyme effects as a strategy to combat COVID-19 [87]. Based on the pharmacodynamic/pharmacokinetic results and predicted anti-SARS-CoV-2 activities, analogs were selected for docking studies (Fig. 13). Blind docking was performed via MOE (molecular operating environment) software using SARS-CoV-2 RdRp (PDB, ID: 7BV2) and ExoN (PDB, ID: 7MC6) proteins from PDB (protein data bank). The molecular docking results revealed that some of the compounds were with promising binding energies in both enzymes relative to Riboprine-TP and Forodesine-TP (triphosphates). The assumptions were supported by in vitro testing as anti-RdRp, anti-ExoN, and anti-SARS-CoV-2 [88].

## 3. Natural RdRp inhibitors

Natural isolate compounds have a significant impact on drug development and pharmacotherapy due to their tremendous structural, and chemical variety, and relatively low toxicity. Numerous natural products were discovered to have medicinal potential, including morphine, guinine, paclitaxel, penicillin, lovastatin, and doxorubicin. Natural compounds were and still are one of the major sources for new medications, with an estimated 34% of approved new chemical entities between 2000 and 2014 coming from natural compounds isolated from plants, microorganisms, and other resources. Many analogs of potential bio-properties are mimicked based on the chemical scaffold of promising natural biologically active entities [89-91]. A famous example is Artemisinin, one of the recent naturally isolate compounds with high potential as anti-malarial (Plasmodium falciparum) properties described by Tu Youyou (Noble Prize winner in 2015). Artemisinin is extracted from the plant Artemisia annua (sweet wormwood, a herb used in traditional medicine in Chinese) [92]. Several review articles reported the antiviral properties of some natural isolate compounds [93–95].

#### 3.1. Benzopyrans

Flavonoids and their glycosides or their bioisosteres displayed antiviral properties inhibiting different stages of the virus infective cycle (inhibition of viral protease, RNA polymerase, and mRNA). Different reports mentioned the efficacy against some RNA viruses (SARS-CoV, MERS-CoV, and influenza A virus) [96–98].

Zandi et al. reported the antiviral properties of baicalein 1 and baicalin 2 (polyphenolic flavonoids accessible in traditional Chinese medicine extracted from Scutellaria baicalensis root) against SARS-CoV-2 through cell-based and biochemical studies (Fig. 14). Both compounds inhibited activity against SARS-CoV-2 RdRp with higher efficacy for baicalein than that of baicalin. Antiviral evaluation assay of baicalein and baicalin exhibited 99.8% and 98% inhibition of SARS-CoV-2, respectively (at 20 µM). Baicalein also showed more potency against SARS-CoV-2 relative to baicalin with an  $EC_{50}=4.5~\mu M$  and  $EC_{90}=7.6$  $\mu$ M. Both compounds also demonstrated anti-SARS-CoV-2 activity when tested in Vero-E6 and human Calu3 cells with safe cytotoxicity ( $EC_{50} =$ 4.5, 9.0; 1.2, 8.0;  $CC_{50}$  = 8.6, >100; 91, >100  $\mu M$  for baicalein and baicalin in Vero-E6 and Calu3 cells, respectively). In the SARS-CoV-2 RdRp testing of the compounds using Remdesivir-triphosphate (RDV-TP, reference standard), baicalein significantly revealed higher potency against RNA polymerase than that of RDV-TP and baicalin. In silico studies (PDB, ID: 6XQB) using AutoDock Vina 1.5.6 and Discovery Studio 2.5 software showed that both compounds were with higher affinity to SARS-CoV-2 RdRp compared with Remdesivir (Fig. 15) [99].

A series of 4*H*-chromen-4-one containing compounds were subjected to virtual screening studies with the coronavirus main protease ( $M^{pro}$ , also called 3CL<sup>pro</sup> "3-chymotrypsin-like cysteine protease", PDB: 6LU7) using AUTO-DOCK and VINA software. The top hits (isoginkgetin, bilobetin, and afzelin molecules, Fig. 16) were subjected to in vitro testing (SARS-CoV-2–infected Vero cell drug screening assay). Isoginkgetin was the most potent active agent discovered with IC<sub>50</sub> = 22.81  $\mu$ M, relative to IC<sub>50</sub> = 7.18, 11.63, and 11.49  $\mu$ M for the standard references Remdesivir, Chloroquine, and Iopinavir, respectively. Isoginkgetin was also docked in RdRp (PDB: 6M71) (Fig. 17). Good binding affinities were noticed in both SARS-CoV-2 M<sup>pro</sup> and RdRp docking studies supporting its efficacy [100].

In silico studies adopting the docking technique (AutoDock Vina software) of quercetin-3-*O*-rutinoside (rutin) and kaempferol-3-*O*-rutinoside (nicotiflorin) "flavonoid glycosides found in *Dysphania ambro-sioides*" in the SARS-CoV-2 M<sup>PRO</sup> (PDB: 6W63) and RdRp (PDB: 6M71), support the possibility as promising agents against SARS-CoV-2. However, intensive in vitro and in vivo studies are still needed for these assumptions (Figs. 18 and 19) [101].

Similarly, various flavonoids and triterpenes were considered in



Sitoindoside IX

Fig. 24. Phytochemicals from Indian medicinal plants of potential RdRp inhibitors.



Fig. 25. Cryptomisrine, cryptospirolepine, cryptoquindoline and biscryptolepine of potentional inhibitory properties against MPro and RdRp through in silico studies.

docking studies against RdRp (PDB: 7BV2) utilizing Schrödinger software revealing promising docking scores with no experimental supporting (in vitro/in vivo) observations [102]. Ellagic acid, *p*-Coumaric acid, kaempferol, and quercetin (Fig. 20) were also mentioned as anti-SARS-CoV-2 agents due to docking studies in M<sup>pro</sup> (PDB: 6LU7) and RdRp (PDB: 6M71) utilizing AutoDock Vina software (Fig. 21) [103].

Theaflavin extracted from *Camellia sinensin*, is a polyphenolic compound found in black tea with a considerable medicinal value useable in Chinese traditional medicine (Fig. 22). Theaflavin and theaflavin gallate derivatives exhibit broad-spectrum antiviral properties against several viruses, influenza A and B and hepatitis C virus [104,105]. In silico studies utilizing theaflavin revealed promising efficacy against RdRp of SARS-CoV-2. Similar observations were also noticed for SARS-CoV, and MERS-CoV, using the UCSF Chimera and SWISS-MODEL (Fig. 23) [106].

In-silico studies, including molecular docking (DOCK 6 software) and molecular dynamics simulations, identified four phytochemicals, from Indian medicinal plants, swertiapuniside, and amarogentin (from *Swertia chirayita*), cordifolide A (from *Tinospora cordifolia*) and sitoindoside IX (from *Withania Somnifera*) as RdRp inhibitors of SARS-CoV-2 (Fig. 24) [107].



Fig. 26. View of 3D- (left) and 2D-interactions (right) of (a) cryptomisrine, (b) cryptospirolepine and (c) cryptoquindoline with RdRp enzyme active pocket [109].



Fig. 27. Phycocyanobilins, riboflavin and cyanidin of potential inhibitory properties against Mpro and RdRp enzymes through in silico studies.



Fig. 28. Docked compounds "(a) Phycocyanobilin, (b) Riboflavin, (c) Cyanidin and (d) Daidzein" in RdRp [110].

# 3.2. Alkaloid

Alkaloids are found in *Cryptolepis sanguinolenta* (West African herb with antiviral, antiplasmodial and anti-inflammatory properties) [108]. In silico studies for the potential anti-SARS-CoV-2, the main protease (M<sup>pro</sup>, PDB: 6LU7) and RdRp (PDB: 6M71) were considered for the alkaloid analogs. Cryptomisrine, cryptospirolepine, cryptoquindoline, and biscryptolepine showed promising affinity towards RdRp enzyme, suggesting their potential inhibitory properties towards both proteins (Figs. 25 and 26) [109].

In another in silico molecular docking study for some of the food

bioactive compounds, three alkaloids phycocyanobilin (found in Spirulina), riboflavin (found in eggs, meat, fruits), cyanidin (found in grapes and berries) revealed high binding affinity towards SARS-CoV-2 M<sup>pro</sup> and RdRp enzymes relative to the antiviral drugs Remdesivir, Nelfinavir, and Lopinavir utilizing AutoDock Vina software (Figs. 27 and 28) [110].

# 3.3. Polycyclic aromatic

The potential inhibitory effects of phytocompounds derived from *Clerodendrum* spp (with the application against respiratory disorders problems) [111,112] against SARS-CoV-2 RdRp (PDB: 7BV2) were



Fig. 29. Phytocompounds of potential inhibitory properties aganst SARS-CoV-2Mpro and RdRp through in silico studies.

studied by molecular docking (AutoDock Vina software). Taraxerol, friedelin, and stigmasterol displayed the highest binding potential relative to Remdesivir and Favipiravir. They also exhibited promising inhibitory affinity against the M<sup>pro</sup> (SARS-CoV-2 spike protein) (Figs. 29 and 30) [113].

## 4. Synthetic non-nucleoside RdRp inhibitors

#### 4.1. Indole

A series of 2-[(indol-3-yl)thio)]-*N*-benzyl-acetamides **7** were synthesized through the reaction of the Bunte salt ethyl acetate-2-sodium thiosulfate **3**, and indole derivatives **4** to afford the indolyl thioacetates **5**. The hydrolysis of **5** yielded the corresponding acids **6**. Finally, the coupling reaction of **6** with different substituted benzylamines afforded the target products **7**. Compound **8** was also generated



Fig. 30. Interaction of Taraxerol in (A) SARS-CoV-2 spike protein, (B) SARS-CoV-2 M<sup>pro</sup> and (C) SARS-CoV-2 RdRp [113].



R<sup>1</sup> = H, 4-Br, 5-Br, 6-Br, 7-Br; R<sup>2</sup> = H, CH<sub>3</sub>; R<sup>3</sup> = NH-CH<sub>2</sub>-Ar



(a)  $Na_2S_2O_3$ ,  $MeOH/H_2O$ ; (b) iodine, DMSO, 60 °C; (c) NaOH,  $EtOH/H_2O$ ; (d) HATU, DIEA,  $CH_2Cl_2$ ; (e) Phenylboronic acid,  $Pd_2(dba)_3$ , Sphos, cesium carbonate, 1,4-dioxane

Scheme 1. Synthesis of 2-[(indol-3-yl)thio]-N-benzyl-acetamides 7 and 8.



Fig. 31. Effective candidates in the CoV-RdRp-Gluc reporter assay.



a) P<sub>2</sub>S<sub>5</sub>, THF, Reflux; b) ethyl 2- bromoacetate, K<sub>2</sub>CO<sub>3</sub>, DMF;
c) sodium S-(2-ethoxy-2-oxoethyl) sulfurothioate, iodine, DMSO, 60 °C;
d) Na, EtOH/H<sub>2</sub>O; e) substituted aniline, HATU, DIEA, CH<sub>2</sub>Cl<sub>2</sub>

Scheme 2. Synthesis of 2,3-dithioindoles 9.



Fig. 32. Compounds 7e and 7h with high potency against SARS-CoV-2 RdRp.

via Suzuki-Miyaura cross-coupling of its corresponding analog 7 with phenylboronic acid (Scheme 1) [114].

CoV-RdRp-Gluc reporter assay was considered for evaluation of SARS-CoV-2 RdRp inhibitory properties of the synthesized agents. Five compounds (**7a-7d**, **8**) showed dose-dependent inhibition of SARS-CoV-2 RdRp with IC<sub>50</sub> values of 1.11–4.55  $\mu$ M while the IC<sub>50</sub> of the positive control (Remdesivir) was 1.19  $\mu$ M (Fig. 31). The most potent five compounds (**7a-7d**, **8**) were further evaluated for their ability to inhibit the RNA synthesis by SARS-CoV-2 RdRp. All of them decreased the levels



**10a**, R = 4-bocpiperazin-1-yl **10b**, R = (S)-boc-3-amino-pyrrolidinyl **10c**, R = (R)-boc-3-amino-pyrrolidinyl

Fig. 33. Quinolines 10a-c of promising SARS-CoV-2-RdRp inhibitory properties.

of viral plus- and minus-strand Gluc RNA in a dose-dependent manner (at 5 and 10  $\mu$ M concentrations). Three compounds **7b**, **7d**, and **8** diminished the levels of both plus- and minus-strand Gluc RNA more potently than the Remdesivir. The antiviral effect of the most potent compound **7d** was evaluated against the human coronavirus strains HCoV-OC43 ( $\beta$ -coronavirus as SARS-CoV-2) and HCoVNL63 ( $\alpha$ -coronavirus as SARS-CoV-2) in a cell-based assay in vitro. It was found that compound **7d** inhibits the replication of HCoV-OC43 more effectively than Remdesivir in a dose-dependent manner [114].



Fig. 34. Suramin, a promising RdRp inhibitor.



Fig. 35. Rifapentine, Rifabutin, Fidaxomicin and Ivermectin of potential interaction with SARS-COV-2 RdRNAP protein.

The 2-[(1*H*-indol-3-yl)thio]-*N*-phenyl-acetamide, 2-[(indol-3-yl) thio)-*N*-benzyl-acetamide (Scheme 1), and 2,3-dithioindole derivatives (synthesized according to Scheme 2) were screened through CoV-RdRp-Gluc assay. Compound **7h** ( $R^1 = H$ ,  $R^2 = Boc$ ,  $R^3 = 3$ -EtOC<sub>6</sub>H<sub>4</sub>) represented the most promising agent (EC<sub>50</sub> = 1.41 µM, selectivity index = >70.92). However, the other compounds showed good to mild RdRp inhibition effect with an EC<sub>50</sub> = 1.70, 2.75, 2.53, 3.07, 1.05 µM for **7e** ( $R^1 = R^2 = H$ ,  $R^3 = 2$ -MeOC<sub>6</sub>H<sub>4</sub>), **7f** ( $R^1 = R^2 = H$ ,  $R^3 = 3$ -H<sub>3</sub>CC<sub>6</sub>H<sub>4</sub>), **9a** (R = 3,4-Cl<sub>2</sub>), and Remdesivir respectively [115].

High potency against SARS-CoV-2 RdRp was revealed by compounds **7e** and **7h** (EC<sub>50</sub> = 1.58, 1.18  $\mu$ M, respectively measured in HEK293T cells with CCK-8 kits) relative to Remdesivir (EC<sub>50</sub> = 0.73  $\mu$ M, measured in A549 cells with CCK-8 kits) [115] (Fig. 32).

## 4.2. Quinoline

Quinoline derivatives 10a-c (Fig. 33), that considered previously against the influenza A virus, were repurposed as SARS-CoV-2-RdRp inhibitors. The assumed compounds revealed promising inhibitory properties relative to Remdesivir at 10  $\mu$ M (95.03%, 92.85%, and 74.94% for 10a-10c, respectively). The CC<sub>50</sub> values for 10a-10c were

70.79, 72.44, and 79.43  $\mu$ M, respectively, with therapeutic indexes 65.55, 34.83, and 20.26, respectively reflecting their low cytotoxicity. Compound **10a** showed antiviral activity against human coronavirus strains HCoV-OC43 and HCoVNL63 determine by the cell-based assay (HCT-8 and LLC-MK2 cell lines infected with HCoV-OC43 " $\beta$ -coronavirus as SARS-CoV-2" and HCoV-NL63 ( $\alpha$ -coronavirus as SARS-CoV-2") [116].

## 4.3. Suramin

A 100 year-old-drug, Suramin (Fig. 34) is identified as a potent inhibitor of the SARS-CoV-2 RdRp and acts by blocking the binding of RNA to the enzyme. Biochemical studies suggest Suramin and its derivatives are at least 20-fold more potent than Remdesivir. The 2.6 Å cryoelectron microscopy structure of the viral RdRp bound to Suramin uncovers two binding sites of which one directly blocks the binding of the RNA template strand and the other clashes with the RNA primer strand near the RdRp catalytic site, thus inhibiting RdRp activity. The IC<sub>50</sub> values obtained from the solution-based assays of RdRp inhibition for Suramin is 0.26  $\mu$ M, and for Remdesivir in its triphosphate form (RDV-TP) is 6.21  $\mu$ M under identical assay conditions, suggesting that Suramin is at least 20-fold more potent than RDV-TP [117].

# Table 2

Predicted ADME-Tox properties.

Compd.	MW	logP	HBD	HBA	RB	TPSA	BBB	HIA	P-gp substrate	hERG pIC <sub>50</sub>	BS
Remdesivir	602.6	1.77	4	14	15	203.5	_	-	yes	4.60	0.17
VV116	501.5	2.59	1	12	12	168.1	_	+	no	4.16	0.17
GS-621763	501.5	2.59	1	12	12	168.1	_	+	no	4.17	0.17
Favipiravir	157.1	-1.25	2	5	1	88.84	_	+	no	3.28	0.55
Molnupiravir	329.3	-0.34	4	10	6	143.1	-	-	no	3.85	0.55
Galidesivir	265.3	-1.20	6	8	2	140.3	-	+	yes	3.69	0.55
Ribavirin	244.2	-1.85	4	9	3	143.7	-	-	no	3.26	0.55
Sofosbuvir	529.5	1.19	3	12	11	158.2	_	+	yes	4.51	0.17
Daclatasvir	734.8	4.00	4	14	17	176.3	_	+	yes	4.57	0.17
Alovudine	244.32	-0.44	2	6	2	84.32	-	+	no	4.21	0.55
Azidothymidine	267.2	-0.02	2	9	3	134.1	-	+	no	4.09	0.55
Tenofovir	287.2	-0.005	3	9	5	136.4	-	_	yes	4.61	0.55
AT-527	581.5	2.03	4	14	12	185	-	_	yes	4.96	0.17
Vidarabine	267.2	-1.05	4	9	2	139.5	_	+	no	4.19	0.55
Tubercidin	266.3	-0.77	4	8	2	126.7	-	+	no	4.24	0.55
Aristeromycin	265.3	-0.53	4	8	2	130.3	-	-	yes	4.47	0.55
8-Chloroadenosine	301.7	-0.49	4	9	2	139.5	-	+	no	4.29	0.55
Fludarabine	285.2	-1.04	4	9	2	139.5	-	+	no	4.44	0.55
Neplanocin A	263.3	-0.77	4	8	2	130.3	-	+	yes	4.61	0.55
Psicofuranine	297.3	-1.7	5	10	3	159.8	-	+	yes	3.96	0.55
Decoyinine	279.3	-1.22	4	9	2	139.5	-	+	yes	4.15	0.55
Cladribine	285.7	0.02	3	8	2	119.3	-	-	no	4.45	0.55
Clofarabine	303.7	-0.19	3	8	2	119.3	-	+	no	4.47	0.55
Riboprine	335.4	0.23	4	9	5	125.6	-	-	yes	5.19	0.55
Tecadenoson	337.3	-0.20	4	10	4	134.8	-	-	yes	7.83	0.55
Nelarabine	297.3	-0.98	4	10	3	148.8	-	+	no	4.20	0.55
Forodesine	266.3	-1.59	6	8	2	134.3	-	+	no	3.72	0.55
Maribavir	376.2	2.09	4	7	4	99.77	-	+	yes	5.17	0.55
Forodesine-TP	506.2	-0.40	9	17	8	273.9	-	-	yes	2.35	0.11
Riboprine-TP	575.3	0.58	7	18	11	265.1	-	-	yes	3.55	0.11
Baicalein	270.2	0.31	3	5	1	90.9	-	+	no	4.54	0.55
Baicalin	446.4	2.94	6	11	4	187.1	-	+	no	3.03	0.11
Isoginkgetin	566.5	4.24	4	10	5	159.8	-	+	no	5.11	0.55
Bilobetin	552.5	3.81	5	10	4	170.8	-	+	yes	5.01	0.55
Afzelin	432.4	0.40	6	10	3	170.1	-	-	no	4.41	0.55
Rutin	610.5	-0.82	10	16	6	269.4	-	-	yes	3.89	0.17
Nicotiflorin	594.5	-0.69	9	15	6	249.2	-	-	yes	4.04	0.17
Ellagic acid	302.2	1.57	4	8	0	141.3	-	+	yes	5.16	0.55
p-Coumaric acid	164.2	1.47	2	3	2	57.53	+	+	no	3.58	0.55
Kaempferol	286.2	2.55	4	6	1	111.1	-	+	no	4.65	0.55
Quercetin	302.2	1.54	5	7	1	131.4	-	+	no	4.41	0.55
Theatlavin	564.5	1.65	9	12	2	217.6	-	-	yes	4.90	0.17
Swertiapuniside	598.5	-1.18	9	10	/	258.4	-	-	no	4.22	0.17
Amarogentin	586.5	0.81	6	13	8	201.7	-	-	yes	4.08	0.11
Cordifolide A	598.7	0.34	5	12		185.3	-	+	yes	4.00	0.17
Sitoindoside IX	632.7	0.57	5	11	0	1/5.5	-	+	yes	4.29	0.17
Cryptomisrine	462.5	4.98	2	5	2	/4.43	-	+	yes	5.65	0.17
Cryptospirolepine	504.6	5.70	1	5	1	44.27	-	+	no	6.17	0.55
Riseruntolonino	448.5 462 E	0.74	0	4	1	35.04	+	+	yes	7.01	0.55
Bisciyptolepille	402.3	7.93	5	4	11	164.7	+	Ŧ	yes	2.64	0.55
Pilycocyallobilli Diboflovin	276 /	3.79	5	10		104.7	-	_	yes	2.04	0.11
Cuonidin	370.4	-0.80	5	10	1	112.4	-	+	no	4.40	0.55
Cyalliulli Torovorol	496.7	2.09	1	1	1	20.22	-	+	no	4.7Z	0.55
Eriodolin	420.7	5.60	1	1	0	20.23	+	+	no	5.41	0.55
Friedelill Stiemesterel	420.7	5.55	0	1	0	17.07	+	+	110	5.08	0.55
	412.7 206 E	/.10	1	1	5	20.23	+	+	yes	6.28 E 40	0.55
7 d 7 h	210.4	4.50	1	3	6	26.1	+	+	no	4.01	0.55
70	310.4 46E 4	5.10	1	3	0	26.1	+	+	110	4.91	0.55
70	405.4	3.37	1	5	0	50.1	+	+	yes	5.51	0.55
70	226.4	4.00	1	3	9	54.50	-	+	yes	4.07	0.55
7e 7f	320.4	2.04	2	4	/	54.1Z	+	+	110	4.27 E 10	0.55
102	476.6	3.75	1	8	10	84	_	т +	y co	5.10	0.55
104	470.0	3.75	1	8	10	04 80.76	-	+	yes	5.21	0.55
10c	461 1	3 74	1	, 7	10	80.76	_	+	ves	5.31	0.55
Suramin	1297	1.97	12	29	22	483.8	_	_	ves	3.96	0.11
Rifabutin	847	2.90	5	15	5	205.6	_	+	ves	4.24	nd
Rifapentine	877	4.05	6	16	6	200.0	_	+	ves	6.24	nd
Fidaxomicin	1058	3.76	7	18	15	266.7	_	_	ves	4.82	nd
Ivermectin	875.1	3.33	, 3	14	8	170.1	_	_	ves	4.96	nd
	0, 0,1	0.00	•		5	1, 0,1			,		

 $Ideal \ values \ for \ oral \ drug \ candidates: \ MW \geq 500; \ log P \geq 5; \ HBA \geq 10; \ RB \geq 10; \ TPSA \geq 140; \ BBB \ (-); \ HIA \ (+); \ P-gp \ substrate \ (yes); \ hERG \ pIC_{50} \geq 5; \ BS \geq 0.55.$ 

## 5. In silico proposed compounds

In silico techniques help design bioactive agents and identify anti-SARS-CoV-2 hits [118,119]. Diverse acyclic [120] and heterocycles [121–123] of different scaffolds (triazole [124], oxadiazole [125], thiadiazole [126], indole [127], pyridine [128], pyrimidine [129] and coumarin [130]) were predicated as effective inhibitory agents for SARS-CoV-2 RdRp. However, the lack of experimental in vitro and in vivo observed results hindered the usefulness of these proposed prospective compounds. Recently, Saha et al. explored the impact of natural products (Tellimagrandin I, SaikosaponinB2, Hesperidin (–)-Epigallocatechin Gallate) on SARS-COV-2 RdRp protein [131].

In addition to the nucleoside and non-nucleoside based RdRp inhibitors, many lead candidates were identified through molecular docking and homolog model-based screening. Parvez et al. revealed that antibacterial drugs (fidaxomicin, ivermectin, rifabutin, and rifapentine) show potential interaction with SARS-COV-2 RdRp protein (Fig. 35). These drugs could be further investigated and considered as leads for further development of potential RdRp inhibitors for SARS-COV-2 [132].

# 6. Predicted ADME-Tox Properties

Drug development is always a tedious and challenging task. Several strategies have been used in recent years to expertise the process which includes molecular hybridization, molecular docking, and consideration of ADME-Tox properties. As many drug candidates fail to reach their drug target because of their poor ADME-Tox profile. It is essential to pay attention for having balanced pharmacokinetic properties while developing new RdRp inhibitors for SARS-CoV-2. The descriptor and druglikeness properties of the compounds included in this article were calculated with the Swiss ADME server [133] and STARDROP software [134]. Among the various parameters, we considered to include molecular weight (MW), logP, hydrogen bond donors (HBD), hydrogen bond acceptors (HBA), number of rotatable bonds (RB), topological polar surface area (TPSA), ability to cross the blood-brain barrier (BBB), human intestine absorption (HIA), inhibition of P-glycoprotein (P-gp), human ether-a-go-go-related gene (hERG) potassium channel inhibition and bioavailability score (BS) as these properties play critical role (see Table 2). These data could serve as additional information for the new drug development process.

## 7. Conclusion

SARS-CoV-2 is considered one of the most severe pandemics facing the global population. The development of efficient anti-SARS-CoV-2 drugs over a short time is associated with many challenges, considerable obstacles, and unknown difficulties. Among various modern approaches to adopt for developing potential drug candidates for SARS-CoV-2, rational drug design and drug repurposing strategies are key. In the past two years, RdRp is found to be a prime target as this enzyme has capable to combat viral replication. Some drugs have been identified as anti-SARS-CoV-2 RdRp accessible for mild and weak infectious patients. Numerous natural and synthetic molecules have also been investigated. However, drug candidates with higher potency are still in demand. In silico studies have been exploring the most in the search for potential drug candidates but without supporting in vitro and in vivo observations their accessibilities will be hindered for any drug discovery program. We believe the compiled information will develop an interest in this field among the research community and provide them with a deeper understanding of the requirements and importance of chemical scaffolds for designing potential RdRp inhibitors for SARS-CoV-2.

# Declaration of competing interest

interests or personal relationships that could have appeared to influence the work reported in this paper.

# Data availability

No data was used for the research described in the article.

## Acknowledgment

This work was supported financially by National Research Centre, Egypt, project ID: 13060103. We also thank the Department of Chemistry and Physics at Augusta University for their support.

#### References

- [1] https://www.emro.who.int/health-topics/infectious-diseases/index.html (accessed on Dec. 1, 2022).
- [2] J.M. Miroa, A. Moreno, The infectious diseases challenges facing Spain in 2040 require the specialty: A national and international survey, Enferm. Infecc. Microbiol. Clín. (2023), https://doi.org/10.1016/j.eimc.2022.08.009 (in press).
- [3] O.E. Santangelo, V. Gianfredi, S. Provenzano, Wikipedia searches and the epidemiology of infectious diseases: a systematic review, Data Knowl. Eng. 142 (2022), 102093, https://doi.org/10.1016/j.datak.2022.102093.
- [4] M. Lee, J.W. Kim, B. Jang, Dove, An infectious disease outbreak statistics visualization system, IEEE Access 6 (2018) 47206–47216. https://doi:10.1109/ ACCESS.2018.2867030.
- [5] https://covid19.who.int/ (accessed on Dec. 1, 2022).
- [6] World Health Statistics 2022: Monitoring Health for the SDGs, Sustainable Development Goals. ISBN 978-92-4-005114-0 (electronic version), ISBN 978-92-4-005115-7 (print version).
- [7] H. Lu, C.W. Stratton, Y.-W. Tang, Outbreak of pneumonia of unknown etiology in Wuhan, China: the mystery and the miracle, J. Med. Virol. 92 (2020) 401–402. https://doi:10.1002/jmv.25678.
- [8] L. Zheng, L. Zhang, J. Huang, K.S. Nandakumar, S. Liu, K. Cheng, Potential treatment methods targeting 2019-nCoV infection, Eur. J. Med. Chem. 205 (2020), 112687, https://doi.org/10.1016/j.ejmech.2020.112687.
- [9] I.A. Seliem, S.S. Panda, A.S. Girgis, Y. Moatasim, A. Kandeil, A. Mostafa, M.A. Ali, E.S. Nossier, F. Rasslan, A.M. Srour, R. Sakhuja, T.S. Ibrahim, Z.K.M. Abdel-samii, A.M.M. Al-Mahmoudy, New quinoline-triazole conjugates: synthesis, and antiviral properties against SARS-CoV-2, Bioorg. Chem. 114 (2021), 105117, https://doi.org/10.1016/j.bioorg.2021.105117.
- [10] P. Zhou, X.-L. Yang, X.-G. Wang, B. Hu, L. Zhang, W. Zhang, H.-R. Si, Y. Zhu, B. Li, C.-L. Huang, H.-D. Chen, J. Chen, Y. Luo, H. Guo, R.-D. Jiang, M.-Q. Liu, Y. Chen, X.-R. Shen, X. Wang, X.-S. Zheng, K. Zhao, Q.-J. Chen, F. Deng, L.-L. Liu, B. Yan, F.-X. Zhan, Y.-Y. Wang, G.-F. Xiao, Z.-L. Shi, A pneumonia outbreak associated with a new coronavirus of probable bat origin, Nature 579 (2020) 270–273, https://doi.org/10.1038/s41586-020-2012-7.
- [11] J.A.C. Nascimento Junior, A.M. Santos, L.J. Quintans-Junior, C.I.B. Walker, L. P. Borges, M.R. Serafini, SARS, MERS and SARS-CoV-2 (COVID-19) treatment: a patent review, Expert Opin. Ther. Pat. 30 (2020) 567tic, https://doi.org/ 10.1080/13543776.2020.1772231.
- [12] Y. Araf, F. Akter, Y.-d. Tang, R. Fatemi, S.A. Parvez, C. Zheng, G. Hossain, Omicron variant of SARS-CoV-2: genomics, transmissibility, and responses to current COVID-19 vaccines, J. Med. Virol. 94 (2022) 1825–1832. https://do i:10.1002/jmv.27588.
- [13] N.G. Fawazy, S.S. Panda, A. Mostafa, B.M. Kariuki, M.S. Bekheit, Y. Moatasim, O. Kutkat, W. Fayad, M.A. El-Manawaty, A.A.F. Soliman, R.A. El-Shiekh, A. M. Srour, R.F. Barghash, A.S. Girgis, Development of spiro-3-indolin-2-one containing compounds of antiproliferative and anti-SARS-CoV-2 properties, Sci. Rep. 12 (2022), 13880, https://doi.org/10.1038/s41598-022-17883-9.
- [14] R. Krause, J. Smolle, Covid-19 mortality and local burden of infectious diseases: a worldwide country-by-country analysis, J. Infect. Public Health 15 (2022) 1370–1375, https://doi.org/10.1016/j.jiph.2022.10.018.
- [15] L. Buonaguro, M. Tagliamonte, M.L. Tornesello, F.M. Buonaguro, SARS-CoV-2 RNA polymerase as target for antiviral therapy, J. Transl. Med. 18 (2020) 185, https://doi.org/10.1186/s12967-020-02355-3.
- [16] S. Durmuş, K.Ö. Ülgen, Comparative interactomics for virus-human proteinprotein interactions: DNA viruses versus RNA viruses, FEBS Open Bio 7 (2017) 96–107. https://doi:10.1002/2211-5463.12167.
- [17] W. Zhu, C.Z. Chen, K. Gorshkov, M. Xu, D.C. Lo, W. Zheng, RNA-dependent RNA polymerase as a target for COVID-19 drug discovery, SLAS Discovery 25 (2020) 1141–1151, https://doi.org/10.1177/24725552209421.
- [18] R. Banerjee, L. Perera, L.M.V. Tillekeratne, Potential SARS-CoV-2 main protease inhibitors, Drug Discov. Today 26 (2021) 804–816, https://doi.org/10.1016/j. drudis.2020.12.005.
- [19] S. Ullrich, C. Nitsche, The SARS-CoV-2 main protease as drug target, Bioorg. Med. Chem. Lett 30 (2020), 127377, https://doi.org/10.1016/j.bmcl.2020.127377.
- [20] A.D. Elmezayen, A. Al-Obaidi, A.T. Şahin, K. Yelekçi, Drug repurposing for coronavirus (COVID-19): *in silico* screening of known drugs against coronavirus 3CL hydrolase and protease enzymes, J. Biomol. Struct. Dynam. 39 (2021) 2980–2992, https://doi.org/10.1080/07391102.2020.1758791.

The authors declare that they have no known competing financial

- [21] Y. Gao, L. Yan, Y. Huang, F. Liu, Y. Zhao, L. Cao, T. Wang, Q. Sun, Z. Ming, L. Zhang, J. Ge, L. Zheng, Y. Zhang, H. Wang, Y. Zhu, C. Zhu, T. Hu, T. Hua, B. Zhang, X. Yang, J. Li, H. Yang, Z. Liu, W. Xu, L.W. Guddat, Q. Wang, Z. Lou, Z. Rao, Structure of the RNA-dependent RNA polymerase from COVID-19 virus, Science 368 (2020) 779–782. https://doi10.1126/science.abb7498.
- [22] I.A. Seliem, A.S. Girgis, Y. Moatasim, A. Kandeil, A. Mostafa, M.A. Ali, M. S. Bekheit, S.S. Panda, New pyrazine conjugates: synthesis, computational studies, and antiviral properties against SARS-CoV-2, ChemMedChem 16 (2021) 3418–3427, https://doi.org/10.1002/cmdc.202100476.
- [23] X. Xu, Y. Chen, X. Lu, W. Zhang, W. Fang, L. Yuan, X. Wang, An update on inhibitors targeting RNA-dependent RNA polymerase for COVID-19 treatment: promises and challenges, Biochem. Pharmacol. 205 (2022), 115279, https://doi. org/10.1016/j.bcp.2022.115279.
- [24] L. Tian, T. Qiang, C. Liang, X. Ren, M. Jia, J. Zhang, J. Li, M. Wan, X.Y. Wen, H. Li, W. Cao, H. Liu, RNA-dependent RNA polymerase (RdRp) inhibitors: the current landscape and repurposing for the COVID-19 pandemic, Eur. J. Med. Chem. 213 (2021), 113201, https://doi.org/10.1016/j.ejmech.2021.113201.
- [25] K. Zandi, K. Musall, A. Oo, D. Cao, B. Liang, P. Hassandarvish, S. Lan, R.L. Slack, K.A. Kirby, L. Bassit, F. Amblard, B. Kim, S. AbuBakar, S.G. Sarafianos, R. F. Schinazi, Baicalein and baicalin inhibit SARS-CoV-2 RNA-dependent-RNA polymerase, Microorganisms 9 (2021) 893, https://doi.org/10.3390/ microorganisms9050893.
- [26] S.M.V. de Almeida, J.C.S. Soares, K.L. dos Santos, J.E.F. Alves, A.G. Ribeiro, I.T. T. Jacob, C.J.d.S. Ferreira, J.C. dos Santos, J.F. de Oliveira, L.B. de Carvalho Junior, M.d.C.A. de Lima, COVID-19 therapy: what weapons do we bring into battle? Bioorg. Med. Chem. 28 (2020), 115757 https://doi.org/10.1016/j. bmc.2020.115757.
- [27] C. Wu, Y. Liu, Y. Yang, P. Zhang, W. Zhong, Y. Wang, Q. Wang, Y. Xu, M. Li, X. Li, M. Zheng, L. Chen, H. Li, Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods, Acta Pharm. Sinica B10 (2020) 766–788, https://doi.org/10.1016/j.apsb.2020.02.008.
- [28] C. Gil, A. Martinez, Is drug repurposing really the future of drug discovery or is new innovation truly the way forward? Expet Opin. Drug Discov. 16 (2021) 829–831, https://doi.org/10.1080/17460441.2021.1912733.
- [29] M.A. Martinez, Efficacy of repurposed antiviral drugs: lessons from COVID-19, Drug Discov. Today 27 (2022) 1954–1960, https://doi.org/10.1016/j. drudis.2022.02.012.
- [30] X. Gao, X. Jing, J. Wang, Y. Zheng, Y. Qiu, H. Ji, L. Peng, S. Jiang, W. Wu, D. Guo, Safety considerations of chloroquine in the treatment of patients with diabetes and COVID-19, Chem. Biol. Interact. 361 (2022), 109954, https://doi.org/ 10.1016/j.cbi.2022.109954.
- [31] A. Cortegiani, G. Ingoglia, M. Ippolito, A. Giarratano, S. Einav, A systematic review on the efficacy and safety of chloroquine for the treatment of COVID-19, J. Crit. Care 57 (2020) 279–283, https://doi.org/10.1016/j.jcrc.2020.03.005.
- [32] W. Yin, C. Mao, X. Luan, D.-D. Shen, Q. Shen, H. Su, X. Wang, F. Zhou, W. Zhao, M. Gao, S. Chang, Y.-C. Xie, G. Tian, H.-W. Jiang, S.-C. Tao, J. Shen, Y. Jiang, H. Jiang, Y. Xu, S. Zhang, Y. Zhang, H.E. Xu, Structural basis for inhibition of the RNA-dependent RNA polymerase from SARS-CoV-2 by remdesivir, Science 368 (2020) 1499–1504.
- [33] https://www.drugs.com/history/veklury.html (accessed on Dec. 7, 2022).
- [34] Y.N. Lamb, Remdesivir: first approval, Drugs 80 (2020) 1355–1363, https://doi. org/10.1007/s40265-020-01378-w.
- [35] T.K. Warren, R. Jordan, M.K. Lo, A.S. Ray, R.L. Mackman, V. Soloveva, D. Siegel, M. Perron, R. Bannister, H.C. Hui, N. Larson, R. Strickley, J. Wells, K.S. Stuthman, S.A. Van Tongeren, N.L. Garza, G. Donnelly, A.C. Shurtleff, C.J. Retterer, D. Gharaibeh, R. Zamani, T. Kenny, B.P. Eaton, E. Grimes, L.S. Welch, L. Gomba, C.L. Wilhelmsen, D.K. Nichols, J.E. Nuss, E.R. Nagle, J.R. Kugelman, G. Palacios, E. Doerffler, S. Neville, E. Carra, M.O. Clarke, L. Zhang, W. Lew, B. Ross, Q. Wang, K. Chun, L. Wolfe, D. Babusis, Y. Park, K.M. Stray, I. Trancheva, J.Y. Feng, O. Barauskas, Y. Xu, P. Wong, M.R. Braun, M. Flint, L.K. McMullan, S.-S. Chen, R. Fearns, S. Swaminathan, D.L. Mayers, C.F. Spiropoulou, W.A. Lee, S.T. Nichol, T. Cihlar, S. Bavari, Therapeutic efficacy of the small molecule GS-5734 against Ebola virus in rhesus monkeys, Nature 531 (2016) 381–385. https://doi: 10.1038/nature17180.
- [36] M.K. Lo, R. Jordan, A. Arvey, J. Sudhamsu, P. Shrivastava-Ranjan, A.L. Hotard, M. Flint, L.K. McMullan, D. Siegel, M.O. Clarke, R.L. Mackman, H.C. Hui, M. Perron, A.S. Ray, T. Cihlar, S.T. Nichol, C.F. Spiropoulou, GS-5734 and its parent nucleoside analog inhibit Filo-, Pneumo-, and Paramyxoviruses, Sci. Rep. 7 (2017), 43395. https://doi:10.1038/srep43395.
- [37] T.P. Sheahan, A.C. Sims, R.L. Graham, V.D. Menachery, L.E. Gralinski, J.B. Case, S.R. Leist, K. Pyrc, J.Y. Feng, I. Trantcheva, R. Bannister, Y. Park, D. Babusis, M. O. Clarke, R.L. Mackman, J.E. Spahn, C.A. Palmiotti, D. Siegel, A.S. Ray, T. Cihlar, R. Jordan, M.R. Denison, R.S. Baric, Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses, Sci. Transl. Med. 9 (396) (2017) eaal3653. https://doi.10.1126/scitranslmed.aal3653.
- [38] A.J. Brown, J.J. Won, R.L. Graham, K.H. Dinnon III, A.C. Sims, J.Y. Feng, T. Cihlar, M.R. Denison, R.S. Baric, T.P. Sheahan, Broad spectrum antiviral remdesivir inhibits human endemic and zoonotic deltacoronaviruses with a highly divergent RNA dependent RNA polymerase, Antivir. Res. 169 (2019), 104541, https://doi.org/10.1016/j.antiviral.2019.104541.
- [39] R.E. Ferner, J.K. Aronson, Remdesivir in covid-19, A drug with potential-don't waste time on uncontrolled observations, BMJ 369 (2020) m1610. https://do i:10.1136/bmj.m1610.
- [40] A.P. Cardile, T.K. Warren, K.A. Martins, R.B. Reisler, S. Bavari, Will there Be a cure for Ebola? Annu. Rev. Pharmacol. Toxicol. 57 (2017) 329–348. https://doi :10.1146/annurev-pharmtox-010716-105055.

- [41] M. Wang, R. Cao, L. Zhang, X. Yang, J. Liu, M. Xu, Z. Shi, Z. Hu, W. Zhong, G. Xiao, Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro, Cell Res. 30 (2020) 269–271, https:// doi.org/10.1038/s41422-020-0282-0.
- [42] S. Hughes, O. Troise, H. Donaldson, N. Mughal, L.S.P. Moore, Bacterialand fungal coinfection among hospitalized patients with COVID-19: aretro- spective cohortstudy in a UKsecondary-care setting, Clin. Microbiol. Infect. 26 (2020) 1395–1399.
- [43] W.D. Wen, Q. Chen, W.Q. Shi, C. Deng, Y. Shi, W. Tang, K. Chen, X. Wu, M.X. Dai, Y.J. Tang, Deuterium A Nucleoside Analogue and Preparation Method and Use Thereof, 5 June 2020. CN111233929A.
- [44] H.-j. Qian, Y. Wang, M.-q. Zhang, Y.-c. Xie, Q.-q. Wu, L.-y. Liang, Y. Cao, H.q. Duan, G.-h. Tian, J. Ma, Z.-b. Zhang, N. Li, J.-y. Jia, J. Zhang, H.A. Aisa, J.s. Shen, C. Yu, H.-I. Jiang, W.-h. Zhang, Z. Wang, G.-y. Liu, Safety, tolerability, and pharmacokinetics of VV116, an oral nucleoside analog against SARS-CoV-2, in Chinese healthy subjects, Acta Pharmacol. Sin. (2022) 1–9, https://doi.org/ 10.1038/s41401-022-00895-6.
- [45] Y. Xie, W. Yin, Y. Zhang, W. Shang, Z. Wang, X. Luan, G. Tian, H.A. Aisa, Y. Xu, G. Xiao, J. Li, H. Jiang, S. Zhang, L. Zhang, H.E. Xu, J. Shen, Design and development of an oral remdesivir derivative VV116 against SARS-CoV-2, Cell Res. 31 (2021) 1212–1214, https://doi.org/10.1038/s41422-021-00570-1.
- [46] Y. Shen, J. Ai, N. Lin, H. Zhang, Y. Li, H. Wang, S. Wang, Z. Wang, T. Li, F. Sun, Z. Fan, L. Li, Y. Lu, X. Meng, H. Xiao, H. Hu, Y. Ling, F. Li, H. Li, C. Xi, L. Gu, W. Zhang, X. Fan, An open, prospective cohort study of VV116 in Chinese participants infected with SARS-CoV-2 omicron variants, Emerg. Microb. Infect. 11 (2022) 1518–1523, https://doi.org/10.1080/22221751.2022.2078230.
- [47] R.M. Cox, J.D. Wolf, C.M. Lieber, J. Sourimant, M.J. Lin, D. Babusis, V. DuPont, J. Chan, K.T. Barrett, D. Lye, R. Kalla, K. Chun, R.L. Mackman, C. Ye, T. Cihlar, L. Martinez-Sobrido, A.L. Greninger, J.P. Bilello, R.K. Plemper, Oral prodrug of remdesivir parent GS-441524 is efficacious against SARS-CoV-2 in ferrets, Nat. Commun. 12 (2021) 6415, https://doi.org/10.1038/s41467-021-26760-4.
- [48] Y. Furuta, T. Komeno, T. Nakamura, Favipiravir (T-705), a broad spectrum inhibitor of viral RNA polymerase, Proc. Jpn. Acad. Ser. B 93 (2017) 449–463. htt ps://doi:10.2183/pjab.93.027.
- [49] K. Shiraki, T. Daikoku, Favipiravir, an anti-influenza drug against life-threatening RNA virus infections, Pharmacol. Therapeut. 209 (2020), 107512, https://doi. org/10.1016/j.pharmthera.2020.107512.
- [50] D. Gatherer, The 2014 Ebola virus disease outbreak in West Africa, J. General Virol. 95 (2014) 1619–1624. https://doi:10.1099/vir.0.067199-0.
- [51] O.S. Reddy, W.-F. Lai, Tackling COVID-19 using Remdesivir and Favipiravir as therapeutic options, Chembiochem 22 (2021) 939–948, https://doi.org/10.1002/ cbic.202000595.
- [52] N. Dauby, S. Van Praet, C. Vanhomwegen, I. Veliziotis, D. Konopnicki, A. Roman, Tolerability of favipiravir therapy in critically ill patients with COVID-19: a report of four cases, J. Med. Virol. 93 (2021) 689–691. https://doi:10.1002/jmv.26488.
- [53] Z.F. Udwadia, P. Singh, Ha Barkate, S. Patil, S. Rangwala, A. Pendse, J. Kadam, W. Wu, C.F. Caracta, M. Tandon, Efficacy and safety of favipiravir, an oral RNAdependent RNA polymerase inhibitor, in mild-to-moderate COVID-19: a randomized, comparative, open-label, multicenter, phase 3 clinical trial, Int. J. Infect. Dis. 103 (2021) 62–71, https://doi.org/10.1016/j.ijid.2020.11.142.
- [54] K. Naydenova, K.W. Muir, L.-F. Wu, Z. Zhang, F. Coscia, M.J. Peet, P. Castro-Hartmann, P. Qian, K. Sader, K. Dent, D. Kimanius, J.D. Sutherland, J. Löwe, D. Barford, C.J. Russo, Structure of the SARS-CoV-2 RNA-dependent RNA polymerase in the presence of favipiravir-RTP, Proc. Natl. Acad. Sci. USA 118 (2021), e2021946118, https://doi.org/10.1073/pnas.2021946118.
- [55] I. Vicenti, M. Zazzi, F. Saladini, SARS-CoV-2 RNA-dependent RNA polymerase as a therapeutic target for COVID-19, Expert Opin. Ther. Pat. 31 (2021) 325–337, https://doi.org/10.1080/13543776.2021.1880568.
- [56] F. Pourkarim, S. Pourtaghi-Anvarian, H. Rezaee, Molnupiravir: a new candidate for COVID-19 treatment, Pharmacol. Res. Perspect. 10 (2022), e00909. https://d oi:10.1002/prp2.909.
- [57] A. Wahl, L.E. Gralinski, C.E. Johnson, W. Yao, M. Kovarova, K.H. Dinnon II, H. Liu, V.J. Madden, H.M. Krzystek, C. De, K.K. White, K. Gully, A. Schäfer, T. Zaman, S.R. Leist, P.O. Grant, Gr R. Bluemling, A.A. Kolykhalov, M.G. Natchus, F.B. Askin, G. Painter, E.P. Browne, C.D. Jones, R.J. Pickles, R.S. Baric, J. V. Garcia, SARS-CoV-2 infection is effectively treated and prevented by EIDD-2801, Nature 591 (2021) 451–457. https://doi:10.1038/s41586-021-03312-w.
- [58] A.J. Bernal, M.M. Gomes da Silva, D.B. Musungaie, E. Kovalchuk, A. Gonzalez, V. D. Reyes, A. Martin-Quiros, Y. Caraco, A. Williams-Diaz, M.L. Brown, J. Du, A. Pedley, C. Assaid, J. Strizki, J.A. Grobler, H.H. Shamsuddin, R. Tipping, H. Wan, A. Paschke, J.R. Butterton, M.G. Johnson, C. De Anda, Molnupiravir for oral treatment of Covid-19 in nonhospitalized patients, N. Engl. J. Med. 386 (2022) 509–520. https://doi.10.1056/NEJMoa2116044.
- [59] D.V. Parums, Editorial: current status of oral antiviral drug treatments for SARS-CoV-2 infection in non-hospitalized patients, Med. Sci. Mon. Int. Med. J. Exp. Clin. Res. 28 (2022), e935952. https://doi:10.12659/MSM.935952.
- [60] N. Urakova, V. Kuznetsova, D.K. Crossman, A. Sokratian, Da B. Guthrie, A. A. Kolykhalov, M.A. Lockwood, M.G. Natchus, M.R. Crowley, G.R. Painter, E. I. Frolova, I. Frolov, β-D-N<sup>4</sup>Hydroxycytidine is a potent anti-alphavirus compound that induces a high level of mutations in the viral genome, J. Virol. 92 (2018), https://doi.org/10.1128/JVI.01965-17 e01965-17.
- [61] W.P. Painter, W. Holman, J.A. Bush, F. Almazedi, H. Malik, N.C.J.E. Eraut, M. J. Morin, L.J. Szewczyk, G.R. Painter, Human safety, tolerability, and pharmacokinetics of Molnupiravir, a novel broad-spectrum oral antiviral agent with activity against SARS-CoV-2, Antimicrob. Agents Chemother. 65 (2021), https://doi.org/10.1128/AAC.02428-20 e02428-20.

- [62] P. Li, Y. Wang, M. Lavrijsen, M.M. Lamers, A.C. de Vries, R.J. Rottier, M.J. Bruno, M.P. Peppelenbosch, B.L. Haagmans, Q. Pan, SARS-CoV-2 Omicron variant is highly sensitive to molnupiravir, nirmatrelvir, and the combination, Cell Res. 32 (2022) 322–324, https://doi.org/10.1038/s41422-022-00618-w.
- [63] J.G. Julander, V. Siddharthan, J. Evans, R. Taylor, K. Tolbert, C. Apuli, J. Stewart, P. Collins, M. Gebre, S. Neilson, A. Van Wettere, Y.-M. Lee, W.P. Sheridan, J. D. Morrey, Y.S. Babu, Efficacy of the broad-spectrum antiviral compound BCX4430 against Zika virus in cell culture and in a mouse model, Antivir. Res. 137 (2017) 14–22. https://doi:10.1016/j.antiviral.2016.11.003.
- [64] R. Taylor, P. Kotian, T. Warren, R. Panchal, S. Bavari, J. Julander, S. Dobo, A. Rose, Y. El-Kattan, B. Taubenheim, Y. Babu, W.P. Sheridan, BCX4430 – A broad-spectrum antiviral adenosine nucleoside analog under development for the treatment of Ebola virus disease, J. Inf. Public Health 9 (2016) 220–226, https:// doi.org/10.1016/j.jiph.2016.04.002.
- [65] R. Taylor, R. Bowen, J.F. Demarest, M. DeSpirito, A. Hartwig, H. Bielefeldt-Ohmann, D.M. Walling, A. Mathis, Y.S. Babu, Activity of Galidesivir in a hamster model of SARS-CoV-2, Viruses 14 (2022) 8, https://doi.org/10.3390/v14010008.
- [66] T.K. Warren, J. Wells, R.G. Panchal, K.S. Stuthman, N.L. Garza, S.A. Van Tongeren, L. Dong, C.J. Retterer, B.P. Eaton, G. Pegoraro, S. Honnold, S. Bantia, P. Kotian, X. Chen, B.R. Taubenheim, L.S. Welch, D.M. Minning, Y.S. Babu, W. P. Sheridan, S. Bavari, Protection against filovirus diseases by a novel broadspectrum nucleoside analogue BCX4430, Nature 508 (2014) 402–405. https: //doi:10.1038/nature13027.
- [67] R.W. Sidwell, J.H. Huffman, G.P. Khare, L.B. Allen, J.T. Witkowski, R.K. Robins, Broad-spectrum antiviral activity of Virazole: 1-beta-D-ribofuranosyl-1,2,4triazole-3-carboxamide, Science 177 (1972) 705–706, https://doi.org/10.1126/ science.177.4050.705.
- [68] A.A. Elfiky, SARS-CoV-2 RNA dependent RNA polymerase (RdRp) targeting: an in silico perspective, J. Biomol. Struct. Dyn. 39 (2021) 3204–3212, https://doi.org/ 10.1080/07391102.2020.1761882.
- [69] A.A. Elfiky, Anti-HCV, nucleotide inhibitors, repurposing against COVID-19, Life Sci. 248 (2020), 117477, https://doi.org/10.1016/j.lfs.2020.117477.
- [70] Y. Xu, M. Li, L. Zhou, D. Liu, W. He, W. Liang, Q. Sun, H. Sun, Y. Li, X. Liu, Ribavirin treatment for critically Ill COVID-19 patients: an observational study, Infect. Drug Resist. 14 (2021) 5287–5291, https://doi.org/10.2147/IDR. S330743.
- [71] I.F.-N. Hung, K.-C. Lung, E.Y.-K. Tso, R. Liu, T.W.-H. Chung, M.-Y. Chu, Y.-Y. Ng, J. Lo, J. Chan, An R. Tam, H.-P. Shum, V. Chan, A.K.-L. Wu, K.-M. Sin, W.-S. Leung, W.-L. Law, D.C. Lung, S. Sin, P. Yeung, C.C.-Y. Yip, R.R. Zhang, Ag Y.-F. Fung, E.Y.-W. Yan, K.-H. Leung, J.D. Ip, A.W.-H. Chu, W.-M. Chan, An C.-K. Ng, R. Lee, K. Fung, A. Yeung, T.-C. Wu, J.W.-M. Chan, W.-W. Yan, W.-M. Chan, J.F.-W. Chan, A.K.-W. Lie, O.T.-Y. Tsang, V.C.-C. Cheng, T.-L. Que, C.-S. Lau, K.-H. Chan, K.K.-W. To, K.-Y. Yuen, Triple combination of interferon beta-1b, lopinavir–ritonavir, and ribavirin in the treatment of patients admitted to hospital with COVID-19: an open-label, randomised, phase 2 trial, Lancet 395 (2020) 1695–1704, https://doi.org/10.1016/S0140-6736(20)31042-4.
- [72] E. Brochot, S. Castelain, G. Duverlie, D. Capron, E. Nguyen-Khac, C. François, Ribavirin monitoring in chronic hepatitis C therapy: anaemia versus efficacy, Antivir. Ther. 15 (2010) 687–695. https://doi:10.3851/IMP1609.
- [73] S. Crotty, D. Maag, J.J. Arnold, W. Zhong, J.Y.N. Lau, Z. Hong, R. Andino, C. E. Cameron, The broad-spectrum antiviral ribonucleoside ribavirin is an RNA virus mutagen, Nat. Med. 6 (2000) 1375–1379. https://doi:10.1038/82191.
- [74] C.Q. Sacramento, N. Fintelman-Rodrigues, J.R. Temerozo, A.d.P.D. Da Silva, S.d. S. Gomes Dias, C.d.S. da Silva, A.C. Ferreira, M. Mattos, C.R.R. Pão, C.S. de Freitas, V.C. Soares, L.V.B. Hoelz, T.V.A. Fernandes, F.S.C. Branco, M.M. Bastos, N. Boechat, F.B. Saraiva, M.A. Ferreira, S. Jockusch, X. Wang, C. Tao, M. Chien, W. Xie, D. Patel, A. Garzia, T. Tuschl, J.J. Russo, R.K.R. Rajoli, C.S.G. Pedrosa, G. Vitória, L.R.O. Souza, L. Goto-Silva, M.Z. Guimarães, S.K. Rehen, A. Owen, F. A. Bozza, D.C. Bou-Habib, J. Ju, P.T. Bozza, T.M.L. Souza, In vitro antiviral activity of the anti-HCV drugs daclatasvir and sofoshuvir against SARS-CoV-2, the aetiological agent of COVID-19, J. Antimicrob. Chemother. 76 (2021) 1874–1885. https://doi.log3/iac/dkab072.
- [75] Y.-A. Heo, E.D. Deeks, Sofosbuvir/Velpatasvir/Voxilaprevir: A Review in chronic hepatitis C, Drugs 78 (2018) 577–587, https://doi.org/10.1007/s40265-018-0895-5.
- [76] M.A. Smith, R.E. Regal, R.A. Mohammad, Daclatasvir: a NS5A replication complex inhibitor for hepatitis C infection, Ann. Pharm. (Poznan) 50 (2016) 39–46. https://doi:10.1177/1060028015610342.
- [77] A. Sadeghi, A.A. Asgari, A. Norouzi, Z. Kheiri, A. Anushirvani, M. Montazeri, H. Hosamirudsai, S. Afhami, E. Akbarpour, R. Aliannejad, A.R. Radmard, A. H. Davarpanah, J. Levi, H. Wentzel, A. Qavi, A. Garratt, B. Simmons, A. Hill, S. Merat, Sofosbuvir and daclatasvir compared with standard of care in the treatment of patients admitted to hospital with moderate or severe coronavirus infection (COVID-19): a randomized controlled trial, J. Antimicrob. Chemother. 75 (2020) 3379–3385. https://doi:10.1093/jac/dkaa334.
- [78] J. Ju, X. Li, S. Kumar, S. Jockusch, M. Chien, C. Tao, I. Morozova, S. Kalachikov, R.N. Kirchdoerfer, J.J. Russo, Nucleotide analogues as inhibitors of SARS-CoV polymerase, Pharmacol. Res. Perspect. (2020), e00674, https://doi.org/10.1002/ prp2.674.
- [79] E. De Clercq, Tenofovir alafenamide (TAF) as the successor of tenofovir disoproxil fumarate (TDF), Biochem. Pharmacol. 119 (2016) 1–7. https://doi:10.1016/j. bcp.2016.04.015.
- [80] L. Lou, Advances in nucleotide antiviral development from scientific discovery to clinical applications: tenofovir disoproxil fumarate for hepatitis B, J. Clin. Transl. Hepatol. 1 (2013) 33–38, https://doi.org/10.14218/JCTH.2013.004XX.

- [81] J.Y. Feng, V. Du Pont, D. Babusis, C.J. Gordon, E.P. Tchesnokov, J.K. Perry, V. Duong, A. Vijiapurapu, X. Zhao, J. Chan, C. Cohen, K. Juneja, T. Cihlar, M. Götte, J.P. Bilello, The nucleoside/nucleotide analogs Tenofovir and Emtricitabine are inactive against SARS-CoV-2, Molecules 27 (2022) 4212, https://doi.org/10.3390/molecules27134212.
- [82] I. Zanella, D. Zizioli, F. Castelli, E. Quiros-Roldan, Tenofovir, another inexpensive, well-known and widely available old drug repurposed for SARS-COV-2 infection, Pharmaceuticals 14 (2021) 454, https://doi.org/10.3390/ ph14050454.
- [83] A.A. Elfiky, Ribavirin, remdesivir, sofosbuvir, Galidesivir, and tenofovir against SARSCoV-2 RNA dependent RNA polymerase (RdRp): a molecular docking study, Life Sci. 253 (2020), 117592, https://doi.org/10.1016/j.lfs.2020.117592.
- [84] S.S. Good, J. Westover, K.H. Jung, X.-J. Zhou, A. Moussa, P. La Colla, G. Collu, B. Canard, J.-P. Sommadossi, AT-527, a Double prodrug of a guanosine nucleotide analog, is a potent inhibitor of SARS-CoV-2 in vitro and a promising oral antiviral for treatment of COVID-19, Antimicrob. Agents Chemother. 65 (2021), https://doi.org/10.1128/AAC.02479-20 e02479-20.
- [85] Y.J. Han, K.H. Lee, S. Yoon, S.W. Nam, S. Ryu, D. Seong, J.S. Kim, J.Y. Lee, J. W. Yang, J. Lee, A. Koyanagi, S.H. Hong, E. Dragioti, J. Radua, L. Smith, H. Oh, R. Abou Ghayda, A. Kronbichler, M. Effenberger, D. Kresse, S. Denicolò, W. Kang, L. Jacob, H. Shin, J.I. Shin, Treatment of severe acute respiratory syndrome (SARS), Middle East respiratory syndrome (MERS), and coronavirus disease 2019 (COVID-19): a systematic review of *in vitro*, *in vivo*, and clinical trials, Theranostics 11 (2021) 1207–1231. https://doi:10.7150/thno.48342.
- [86] A. Shannon, V. Fattorini, B. Sama, B. Selisko, M. Feracci, C. Falcou, P. Gauffre, P. El Kazzi, A. Delpal, E. Decroly, K. Alvarez, C. Eydoux, J.-C. Guillemot, A. Moussa, S.S. Good, P. La Colla, K. Lin, J.-P. Sommadossi, Y. Zhu, X. Yan, H. Shi, F. Ferron, B. Canard, A dual mechanism of action of AT-527 against SARS-CoV-2 polymerase, Nat. Commun. 13 (2022) 621, https://doi.org/10.1038/s41467-022-28113-1.
- [87] S. Khater, P. Kumar, N. Dasgupta, G. Das, S. Ray, A. Prakash, Combining SARS-CoV-2 proofreading exonuclease and RNA-dependent RNA polymerase inhibitors as a strategy to combat COVID-19: a high-throughput in silico screening, Front. Microbiol. 12 (2021), 647693. https://doi:10.3389/fmicb.2021.647693.
- [88] A.M. Rabie, M. Abdalla, Forodesine and Riboprine exhibit strong anti-SARS-CoV-2 repurposing potential: *In silico* and *in vitro* studies, ACS Bio. Med. Chem (2022), https://doi.org/10.1021/acsbiomedchemau.2c00039. Au (in press).
- [89] D.J. Newman, G.M. Cragg, Natural products as sources of new drugs from 1981 to 2014, J. Nat. Prod. 79 (2016) 629–661. https://doi:10.1021/acs.jnatpro d.5b01055.
- [90] E. De Clercq, G. Li, Approved antiviral drugs over the past 50 years, Clin. Microbiol. Rev. 29 (2016) 695i747, https://doi.org/10.1128/CMR.00102-15.
- [91] A.S. Girgis, P. D'Arcy, D.R. Aboshouk, M.S. Bekheit, Synthesis and bio-properties of 4-piperidone containing compounds as curcumin mimics, RSC Adv. 12 (2022) 31102–31123. http://doi.10.1039/d2ra05518j.
- [92] K.M. Nelson, J.L. Dahlin, J. Bisson, Ja Graham, G.F. Pauli, M.A. Walters, The essential medicinal chemistry of curcumin, J. Med. Chem. 60 (2017) 1620–1637. http://doi:10.1021/acs.jmedchem.6b00975.
- [93] R. Chakravarti, R. Singh, A. Ghosh, D. Dey, P. Sharma, R. Velayutham, S. Roy, D. Ghosh, A review on potential of natural products in the management of COVID-19, RSC Adv. 11 (2021) 16711-16735. http://doi:10.1039/d1ra00644d.
- [94] E. Thomas, L.E. Stewart, B.A. Darley, A.M. Pham, I. Esteban, S.S. Panda, Plantbased natural products and extracts: potential source to develop new antiviral drug candidates, Molecules 26 (2021) 6197, https://doi.org/10.3390/ molecules26206197.
- [95] L. Owen, K. Laird, M. Shivkumar, Antiviral plant-derived natural products to combat RNA viruses: targets throughout the viral life cycle, Lett. Appl. Microbiol. 75 (2021) 476–499. http://doi:10.1111/lam.13637.
- [96] J.S. Mani, J.B. Johnson, J.C. Steel, D.A. Broszczak, P.M. Neilsen, K.B. Walsh, M. Naiker, Natural product-derived phytochemicals as potential agents against coronaviruses: a review, Virus Res. 284 (2020), 197989, https://doi.org/ 10.1016/j.virusres.2020.197989.
- [97] J. Solnier, J.-P. Fladerer, Flavonoids: a complementary approach to conventional therapy of COVID-19? Phytochemistry Rev. 20 (2021) 773–795, https://doi.org/ 10.1007/s11101-020-09720-6.
- [98] F. Limanaqi, C.L. Busceti, F. Biagioni, G. Lazzeri, M. Forte, S. Schiavon, S. Sciarretta, G. Frati, F. Fornai, Cell clearing systems as targets of polyphenols in viral infections: potential implications for COVID-19 pathogenesis, Antioxidants 9 (2020) 1105. https://doi:10.3390/antiox9111105.
- [99] K. Zandi, K. Musall, A. Oo, D. Cao, B. Liang, P. Hassandarvish, S. Lan, R.L. Slack, K.A. Kirby, L. Bassit, F. Amblard, B. Kim, S. AbuBakar, S.G. Sarafianos, R. F. Schinazi, Baicalein and baicalin inhibit SARS-CoV-2 RNA-dependent-RNA polymerase, Microorganisms 9 (2021) 893, https://doi.org/10.3390/ microorganisms9050893.
- [100] V. Raj, J.-H. Lee, J.-J. Shim, J. Lee, Antiviral activities of 4H-chromen-4-one scaffold-containing flavonoids against SARS-CoV-2 using computational and in vitro approaches, J. Mol. Liquids 353 (2022), 118775, https://doi.org/10.1016/j. molliq.2022.118775.
- [101] F.M.A. da Silva, K.P.A. da Silva, L.P.M. de Oliveira, E.V. Costa, H.H.F. Koolen, M. L.B. Pinheiro, A.Q.L. de Souza, A.D.L. de Souza, Flavonoid glycosides and their putative human metabolites as potential inhibitors of the SARS-CoV-2 main protease (Mpro) and RNA-dependent RNA polymerase (RdRp), Mem. Inst. Oswaldo Cruz, Rio de Janeiro 115 (2020), e200207. https://doi:10.1590/ 0074-02760200207.
- [102] R.K. Goyal, J. Majeed, R. Tonk, M. Dhobi, B. Patel, K. Sharma, S. Apparsundaram, Current targets and drug candidates for prevention and treatment of SARS-CoV-2

(COVID-19) infection, Rev. Cardiovasc. Med. 21 (2020) 365–384. https://doi:10 .31083/j.rcm.2020.03.118.

- [103] M.A. Shaldam, G. Yahya, N.H. Mohamed, M.M. Abdel-Daim, Y. Al Naggar, In silico screening of potent bioactive compounds from honeybee products against COVID-19 target enzymes, Env. Sci. Poll. Research 28 (2021) 40507–40514, https://doi.org/10.1007/s11356-021-14195-9.
- [104] Z.-F. Yang, L.-P. Bai, W.-b. Huang, X.-Z. Li, S.-S. Zhao, N.-S. Zhong, Z.-H. Jiang, Comparison of in vitro antiviral activity of tea polyphenols against influenza A and B viruses and structure-activity relationship analysis, Fitoterapia 93 (2014) 47–53, https://doi.org/10.1016/j.fitote.2013.12.011.
- [105] P. Chowdhury, M.-E. Sahuc, Y. Rouillé, C. Rivière, N. Bonneau, A. Vandeputte, P. Brodin, M. Goswami, T. Bandyopadhyay, J. Dubuisson, K. Séron, Theaflavins, polyphenols of black tea, inhibit entry of hepatitis C virus in cell culture, PLoS One 13 (2018), e0198226, https://doi.org/10.1371/journal.pone.0198226.
- [106] J. Lung, Y.-S. Lin, Y.-H. Yang, Y.-L. Chou, L.-H. Shu, Y.-C. Cheng, H.T. Liu, C.-Y. Wu, The potential chemical structure of anti-SARS-CoV-2 RNA-dependent RNA polymerase, J. Med. Virol. 92 (2020) 693–697. https://doi:10.1002/jmv.25761.
- [107] S. Koulgi, V. Jani, M. Uppuladinne V, N.U. Sonavane, R. Joshi, Natural plant products as potential inhibitors of RNA dependent RNA polymerase of Severe Acute Respiratory Syndrome Coronavirus-2, PLoS One 16 (2021), e0251801, https://doi.org/10.1371/journal.pone.0251801.
- [108] N. Osafo, K.B. Mensah, O. Kofi Yeboah, Phytochemical and pharmacological review of *Cryptolepis sanguinolenta* (lindl.) schlechter, hindawi, Adv. Pharmaceut. Sci. 3026370 (2017), https://doi.org/10.1155/2017/3026370.
- [109] L.S. Borquaye, E.N. Gasu, G.B. Ampomah, L.K. Kyei, M.A. Amarh, C.N. Mensah, D. Nartey, M. Commodore, A.K. Adomako, P. Acheampong, J.O. Mensah, D. B. Mormor, C.I. Aboagye, Alkaloids from *Cryptolepis sanguinolenta* as potential inhibitors of SARS-CoV-2 viral proteins: an *in silico* study, Hindawi BioMed. Res. Intern 5324560 (2020), https://doi.org/10.1155/2020/5324560.
- [110] B. Pendyalaa, A. Patras, In Silico Screening of Food Bioactive Compounds to Predict Potential Inhibitors of COVID-19 Main Protease (M<sup>pro</sup>) and RNAdependent RNA Polymerase (RdRp), 2020. ChemRxiv, https://doi:10.26434/c hemrxiv.12051927.v2.
- [111] M.G. Jang, J.M. Oh, H.C. Ko, J.-W. Kim, S. Baek, Y.J. Jin, S.-P. Hur, S.-J. Kim, *Clerodendrum trichotomum* extract improves metabolic derangements in high fructose diet-fed rats, Anim. Cell Syst. 25 (2021) 396–404, https://doi.org/ 10.1080/19768354.2021.2004221.
- [112] J. Jagruti, a Patel, R. Sanjeev, b Acharya, NiyatiS. Acharya, *Clerodendrum serratum* (L.) Moon. – a review on traditional uses, phytochemistry and pharmacological activities, J. Ethnopharmacol. 154 (2014) 268–285, https://doi.org/10.1016/j. jep.2014.03.071.
- [113] P. Kar, N.R. Sharma, B. Singh, A. Sen, A. Roy, Natural compounds from *Clerodendrum* spp. as possible therapeutic candidates against SARS-CoV-2: an *in silico* investigation, J. Biomol. Struct. Dyn. 39 (2021) 4774–4785, https://doi.org/ 10.1080/07391102.2020.1780947.
- [114] G.-N. Zhang, J. Zhao, Q. Li, M. Wang, M. Zhu, J. Wang, S. Cen, Y. Wang, Discovery and optimization of 2-((1H-indol-3-yl)thio)-N-benzylacetamides as novel SARS-CoV-2 RdRp inhibitors, Eur. J. Med. Chem. 223 (2021), 113622, https://doi.org/10.1016/j.ejmech.2021.113622.
- [115] J. Zhao, G. Zhang, Y. Zhang, D. Yi, Q. Li, L. Ma, S. Guo, X. Li, F. Guo, R. Lin, G. Luu, Z. Liu, Y. Wang, S. Cen, 2-((1H-indol-3-y))thio)-N-phenyl-acetamides: SARS-CoV-2 RNA-dependent RNA polymerase inhibitors, Antivir. Res. 196 (2021), 105209, https://doi.org/10.1016/j.antiviral.2021.105209.
- (2021), 105209, https://doi.org/10.1016/j.antiviral.2021.105209.
  [116] J. Zhao, Y. Zhang, M. Wang, Q. Liu, X. Lei, M. Wu, S. Guo, D. Yi, Q. Li, L. Ma, Z. Liu, F. Guo, J. Wang, X. Li, Y. Wang, S. Cen, Quinoline and derivatives inhibit viral RNA synthesis by SARS-CoV-2 RdRp, ACS Infect. Dis. 7 (2021) 1535–1544, https://doi.org/10.1021/acsinfecdis.1c00083.
- [117] W. Yin, X. Luan, Z. Li, Z. Zhou, Q. Wang, M. Gao, X. Wang, F. Zhou, J. Shi, E. You, M. Liu, Q. Wang, Y. Jiang, H. Jiang, G. Xiao, L. Zhang, X. Yu, S. Zhang, H.E. Xu, Structural basis for inhibition of the SARS-CoV-2 RNA polymerase by suramin, Nat. Struct. Mol. Biol. 28 (2021) 319–325.
- [118] A.S. Girgis, S.S. Panda, M.N. Aziz, P.J. Steel, C.D. Hall, A.R. Katritzky, Rational design, synthesis, and 2D-QSAR study of anti-oncological alkaloids against hepatoma and cervical carcinoma, RSC Adv. 5 (2015) 28554–28569. https://doi :10.1039/c4ra16663a.

- [119] P. Subhaswaraj, B. Siddhardha, Molecular docking and molecular dynamic simulation approaches for drug development and repurposing of drugs for severe acute respiratory syndrome-Coronavirus-2, chapter 11, Computational approaches for novel therapeutic and diagnostic designing to mitigate SARS-CoV-2 infection (2022) 207–246, https://doi.org/10.1016/B978-0-323-91172-6.00007-8. Elsevier Inc.
- [120] H. Alici, H. Tahtaci, K. Demir, Design and various in silico studies of the novel curcumin derivatives as potential candidates against COVID-19 -associated main enzymes, Comput. Biol. Chem. 98 (2022), 107657, https://doi.org/10.1016/j. compbiolchem.2022.107657.
- [121] P.R. Nagar, N.D. Gajjar, T.M. Dhameliya, In search of SARS CoV-2 replication inhibitors: virtual screening, molecular dynamics simulations and ADMET analysis, J. Mol. Struct. 1246 (2021), 131190, https://doi.org/10.1016/j. molstruc.2021.131190.
- [122] S. Mahmoudi, M.M. Dehkordi, M.H. Asgarshamsi, The effect of various compounds on the COVID mechanisms, from chemical to molecular aspects, Biophys. Chem. 288 (2022), 106824, https://doi.org/10.1016/j. bpc.2022.106824.
- [123] F. Nabati, A. kamyabiamineh, R. Kosari, F. Ghasemi, S. Seyedebrahimi, S. Mohammadi, M. Moradi, Virtual screening based on the structure of more than 10<sup>5</sup> compounds against four key proteins of SARS-CoV-2: M<sub>Pro</sub>, S<sub>RBD</sub>, RdRp, and PL<sub>pro</sub>, Inform. Med. Unlocked 35 (2022), 101134, https://doi.org/10.1016/j. imu.2022.101134.
- [124] A.Y. Alzahrani, M.M. Shaaban, B.H. Elwakil, M.T. Hamed, N. Rezki, M.R. Aouad, M.A. Zakaria, M. Hagar, Anti-COVID-19 activity of some benzofused 1,2,3-triazolesulfonamide hybrids using in silico and in vitro analyses, Chemometr. Intell. Lab. Syst. 217 (2021), 104421, https://doi.org/10.1016/j. chemolab.2021.104421.
- [125] A.M. Rabie, Two antioxidant 2,5-disubstituted-1,3,4-oxadiazoles (CoViTris2020 and ChloViD2020): successful repurposing against COVID-19 as the first potent multitarget anti-SARS-CoV-2 drugs, New J. Chem. 45 (2021) 761–771, https:// doi.org/10.1039/d0nj03708g.
- [126] M.A. Said, S.M. Riyadh, N.S. Al-Kaff, A.A. Nayl, K.D. Khalil, S. Bräse, S.M. Gomha, Synthesis and greener pastures biological study of bis-thiadiazoles as potential Covid-19 drug candidates, Arab. J. Chem. 15 (2022), 104101, https://doi.org/ 10.1016/j.arabjc.2022.104101.
- [127] T.A. Farghaly, I. Althagafi, M.H. Ibrahim, N.T. Al-Qurashi, U. Farooq, Synthesis under microwaves irradiation, structure elucidation, docking study for inhibiting COVID-19 and DFT calculations of novel azoles incorporated indole moiety, J. Mol. Struct. 1244 (2021), 131263, https://doi.org/10.1016/j. molstruc.2021.131263.
- [128] A. Ramalingam, M. Kuppusamy, S. Sambandam, M. Medimagh, O.E. Oyeneyin, A. Shanmugasundaram, N. Issaoui, N.D. Ojo, Synthesis, spectroscopic, topological, hirshfeld surface analysis, and anti-covid-19 molecular docking investigation of isopropyl 1-benzoyl-4-(benzoyloxy)-2,6-diphenyl-1,2,5,6tetrahydropyridine-3-carboxylate, Heliyon 8 (2022), e10831, https://doi.org/ 10.1016/j.heliyon.2022.e10831.
- [129] Y. El Bakri, M.M. Kurbanova, S.A. Siddique, S. Ahmad, S. Goumri-Said, One-pot synthesis, X-ray crystal structure, and identification of potential molecules against COVID-19 main protease through structure-guided modeling and simulation approach, Arab. J. Chem. 15 (2022), 104230, https://doi.org/10.1016/j. arabjc.2022.104230.
- [130] C.S. Mun, L.Y. Hui, L.C. Sing, R. Karunakaran, V. Ravichandran, Multi-targeted molecular docking, pharmacokinetics, and drug-likeness evaluation of coumarin based compounds targeting proteins involved in development of COVID-19, Saudi J. Biol. Sci. 29 (2022), 103458, https://doi.org/10.1016/j.sjbs.2022.103458.
- [131] S. Saha, R. Nandi, P. Vishwakarma, A. Prakash, D. Kumar, Discovering potential RNA dependent RNA polymerase inhibitors as prospective drugs against COVID-19: an in silico approach, Front. Pharmacol. 12 (2021), 634047.
- [132] M.S.A. Parvez, M.A. Karim, M. Hasan, J. Jaman, Z. Karim, T. Tahsin, M.N. Hasan, M.J. Hosen, Prediction of potential inhibitors for RNA-dependent RNA polymerase of SARS-CoV-2 using comprehensive drug repurposing and molecular docking approach, Int. J. Biol. Macromol. 163 (2020) 1787e1797.

[133] http://www.swissadme.ch (accessed March 08, 2023).

[134] StarDrop, version 7.3; Optibrium Ltd.: Cambridge, U.K.; https://www.optibrium. com/stardrop (accessed March 08, 2023).