



OPEN

Sex-differences in prostaglandin signaling: a semi-systematic review and characterization of PTGDS expression in human sensory neurons

Breanna Q. Shen, Ishwarya Sankaranarayanan, Theodore J. Price & Diana Tavares-Ferreira

There is increasing evidence of sex differences in underlying mechanisms causing pain in preclinical models, and in clinical populations. There are also important disconnects between clinical pain populations and the way preclinical pain studies are conducted. For instance, osteoarthritis pain more frequently affects women, but most preclinical studies have been conducted using males in animal models. The most widely used painkillers, nonsteroidal anti-inflammatory drugs (NSAIDs), act on the prostaglandin pathway by inhibiting cyclooxygenase (COX) enzymes. The purpose of this study was to analyze the preclinical and clinical literature on the role of prostaglandins and COX in inflammation and pain. We aimed to specifically identify studies that used both sexes and investigate whether any sex-differences in the action of prostaglandins and COX inhibition had been reported, either in clinical or preclinical studies. We conducted a PubMed search and identified 369 preclinical studies and 100 clinical studies that matched our inclusion/exclusion criteria. Our analysis shows that only 17% of preclinical studies on prostaglandins used both sexes and, out of those, only 19% analyzed or reported data separated by sex. In contrast, 79% of the clinical studies analyzed used both sexes. However, only 6% of those reported data separated by sex. Interestingly, 14 out of 15 preclinical studies and 5 out of 6 clinical studies that analyzed data separated by sex have identified sex-differences. This builds on the increasing evidence of sex-differences in prostaglandin signaling and the importance of sex as a biological variable in data analysis. The preclinical literature identifies a sex difference in prostaglandin D₂ synthase (PTGDS) expression where it is higher in female than in male rodents in the nervous system. We experimentally validated that PTGDS expression is higher in female human dorsal root ganglia (DRG) neurons recovered from organ donors. Our semi-systematic literature review reveals a need for continued inclusivity of both male and female animals in prostaglandins studies and data analysis separated by sex in preclinical and clinical studies. Our finding of sex-differences in neuronal PTGDS expression in humans exemplifies the need for a more comprehensive understanding of how the prostaglandin system functions in the DRG in rodents and humans.

Differences in the incidence and severity of many pain disorders between men and women have been reported. The prevalence of fibromyalgia¹ migraine², and osteoarthritis pain³ is greater among women than men. Women are also reported to have more severe postoperative pain⁴. The chronic pain patient population is largely female, older, and genetically heterogeneous, while preclinical pain studies have been primarily conducted in young, male mice or rats of limited, and usually inbred strain⁵. The past decade has seen an explosion of preclinical pain research demonstrating fundamental sex differences in mechanisms causing chronic pain in mouse and rat models^{6,7}. Some female-specific pain mechanisms are now emerging in the preclinical literature, such as the more prominent effects of calcitonin gene-related peptide and prolactin in promoting pain in female rodents⁸⁻¹¹. Similar differences are now emerging at the molecular level in the human dorsal root ganglion (DRG)¹²⁻¹⁴ suggesting that sex differences in basic pain mechanisms may contribute to differential efficacy of pain therapeutics

Department of Neuroscience and Center for Advanced Pain Studies, University of Texas at Dallas, 800 W Campbell Rd, Richardson, TX 75080, USA. email: Theodore.price@utdallas.edu; Diana.TavaresFerreira@utdallas.edu

in men and women. We recently identified a sex difference in DRG neuron expression of a prostaglandin synthesizing enzyme, PTGDS, that led to differences in behavioral outcomes in response to prostaglandins and PTGDS inhibitors in male and female mice¹⁵. This finding prompted us to look more carefully at the clinical and preclinical research on prostaglandins and their receptors with the hypothesis that the very commonly used drugs that target this pathway may have differential efficacy in men versus women.

Prostaglandins (PGs) are lipid-derived signaling molecules that play an important role in pain and inflammation. PGs are produced from plasma membrane-derived arachidonic acid, and their conversion is dependent on the action of cyclooxygenase (COX) enzymes (Fig. 1). The four major prostaglandins, prostaglandin E₂ (PGE₂), prostaglandin D₂ (PGD₂), prostacyclin (PGI₂), and prostaglandin F_{2α} (PGF_{2α}) act on G-protein coupled receptors to regulate intracellular signaling pathways¹⁶. PGE₂ acts on the E prostanoid receptors EP1, EP2, EP3, and EP4. As G_s-coupled receptors, EP2 and EP4 activate adenylyl cyclase to produce cyclic adenosine monophosphate (cAMP) from adenosine triphosphate (ATP); as a second messenger, cAMP phosphorylates protein kinase A (PKA), which in turn can phosphorylate intracellular target proteins¹⁶. EP3 is a G_i- and G₁₂-coupled, lowering cAMP, increasing intracellular calcium and the activity of rho GTPases¹⁶ and this receptor has been linked to anti-nociceptive actions of PGE₂¹⁷. EP1 is a G_q-coupled receptor, whose activation causes phospholipase C to catalyze conversion of phosphatidylinositol 4,5-bisphosphate (PIP₂) into inositol triphosphate (IP₃) and diacylglycerol (DAG). All four EP receptor genes are expressed in mouse¹⁸ and human¹⁴ DRG neurons where their expression varies depending on the subtype of receptor and the type of sensory neuron. EP1, 2 and 4 receptors are known to sensitize ion channels like the TRPV1 receptor and voltage gated sodium channels that regulate the excitability of sensory neurons^{19,20}.

PGD₂ acts on two GPCRs, DP1, a G_s-coupled receptor that increases cAMP concentration, and DP2, a G_i-coupled receptor also known as chemoattractant receptor-homologous molecule expressed on T helper 2 cells (CRTH₂) that decreases cAMP concentration and increases calcium intracellularly¹⁶. While some studies have shown that PGD₂ is pro-nociceptive²¹, other studies have demonstrated that PGD₂ has anti-nociceptive effects²². PGI₂ exerts pro-inflammatory effects through the prostacyclin receptor (IP), a G_s-coupled receptor that increases cAMP concentration¹⁶. IP mediates both peripheral nociception and spinal transmission of nociceptive information¹⁶. In dorsal root ganglia (DRG) and spinal dorsal horn neurons, IP receptor activation causes increased neuronal excitability²³. In addition, IP has an important role inducing inflammation, causing vasodilation and edema¹⁶. PGF_{2α} activates the FP_A and FP_B receptors, which are G_q-coupled and increase concentrations

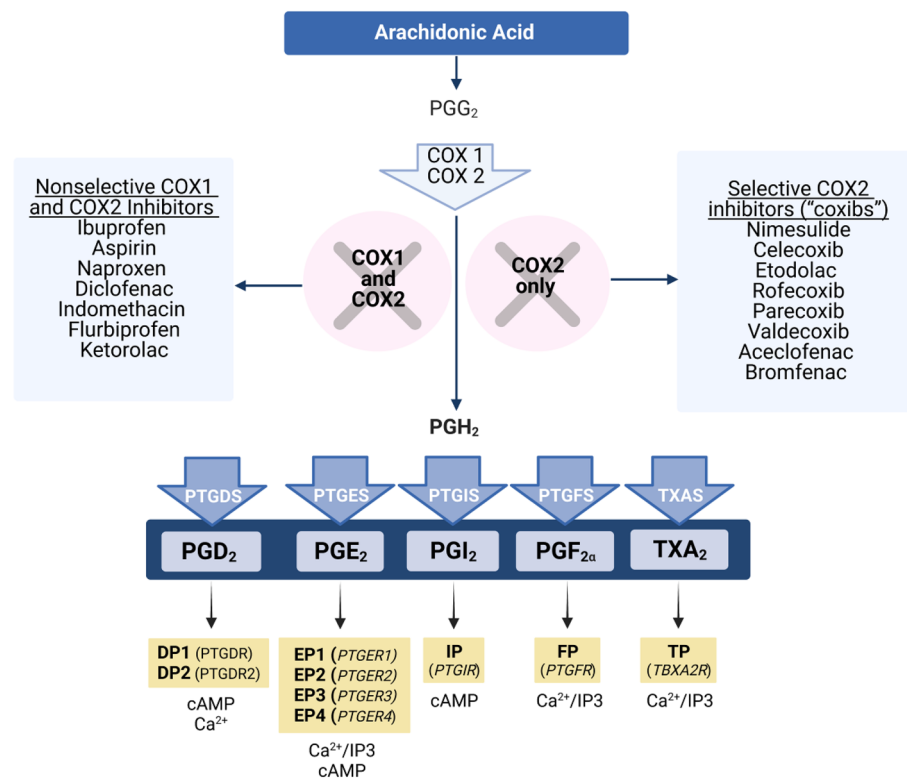


Figure 1. Prostaglandins biosynthesis pathway. Cyclooxygenases (COX) metabolize arachidonic acid first to prostaglandin G₂ (PGG₂) and then to prostaglandin H₂ (PGH₂). COX1 and 2 are targeted by non-steroidal anti-inflammatory drugs (NSAIDs), which block the synthesis of prostaglandins. PGH₂ is converted to prostaglandin D₂ (PGD₂), prostaglandin E₂ (PGE₂), prostaglandin I₂ (PGI₂), prostaglandin F_{2α} (PGF_{2α}) and thromboxane A₂ (TXA₂) by respective synthases. Each prostaglandin binds to specific receptors, which are members of the G protein-coupled receptor (GPCR) superfamily of seven transmembrane proteins, activating different downstream signaling pathways. cAMP cyclic adenosine monophosphate, IP₃ inositol triphosphate.

of IP₃, DAG, calcium, and rho activity¹⁶. PGF_{2a} has pro-inflammatory effects in arthritic conditions and can induce inflammation, but the mechanisms are unclear¹⁶. Finally, an additional prostanoid, thromboxane A₂ acts on TP_a and TP_b GPCRs to increase IP₃, DAG, calcium, and activity of RhoGEF in cells¹⁶.

Nonsteroidal anti-inflammatory drugs (NSAIDs) reduce inflammation and its symptoms through either nonselective inhibition of both COX isoforms or selective inhibition of COX2. COX inhibition then reduces the production of PGs. COX1 is ubiquitously expressed and plays a key role in maintenance of mucosal tissues, and COX2 is upregulated by injury and contributes to inflammation¹⁶. NSAIDs were first used in the form of myrtle and willow tree bark by the Egyptian civilization around 1500 BC for their analgesic and antipyretic functions, and salicin was first isolated from the willow bark in 1828²⁴. Salicylic acid (aspirin) was subsequently synthesized and marketed, followed by ibuprofen and around 50 other NSAIDs. Globally, NSAIDs are some of the most widely used and prescribed drugs²⁴. While aspirin is a noncompetitive COX inhibitor, non-aspirin NSAIDs competitively inhibit active sites on COX enzymes²⁴. In an attempt to eliminate the gastrointestinal complications of COX1 inhibition, COX2 selective inhibitors, also known as coxibs, were developed in the 1990s²⁵.

The primary goal of our semi-systematic review was to first identify if males, females or both sexes have been used in studies investigating PGs in pain and inflammation. Second, we aimed to identify any reports of sex-differences in PG signaling in the preclinical literature (studies using non-human subjects) and at the clinical level. Our semi-systematic review found that there is a large bias towards using male animals in prostaglandin preclinical studies. While most clinical studies include both male and female subjects, only a small proportion of studies reported data separated by sex. To provide further evidence of the need to look at the impact of sex differences in studies on prostaglandins and pain, we characterized the expression of PTGDS, an enzyme that converts PGH₂ to PGD₂, in human DRG neurons. We found that PTGDS protein expression is higher in DRG neurons recovered from female organ donors compared to male organ donors. We conclude that potential sex differences in the action of prostaglandins should be studied more carefully in preclinical and clinical studies moving forward.

Results

We conducted a semi-systematic review of the literature on prostaglandins (PGs), inflammation and pain. Papers were categorized as preclinical studies, which included animal models and/or human tissues, and as clinical studies, which included human subjects in clinical or experimental settings.

Preclinical studies. Details on the preclinical studies included in our semi-systematic review can be found in Supplementary Files 1 and 2. Here we summarize the articles that report results separated by sex.

In several studies, male and female animals demonstrated differences in response to noxious substance exposure related to prostaglandins. In a model of IL-1b induced temporomandibular joint (TMJ) inflammation, female rats showed greater sensitivity in the TMJ region than male rats before and after implantation of a cannula. Female rats also had a lower head withdrawal threshold after intratrigeminal ganglionic IL-1b injection but this difference was not statistically significant. IL-1b upregulates COX2 mRNA and protein, resulting in upregulation of PGE₂, increased EP2 activation, and increased Nav1.7 expression. No significant differences were observed in expression fold change for Nav1.7 and COX2 mRNA and protein between males and females²⁶. While there is inconclusive evidence on sex differences in the endogenous production of PGE₂ in response to introduction of IL-1b, application of exogenous PGE₂ as a component of a combination of inflammatory mediators in another study caused sex-dependent sensitization of neurons. In this study, the proportion of dural afferents sensitized by an inflammatory mediator solution of bradykinin, histamine, and PGE₂ was significantly greater in female rats than male rats. The investigators suggest this may be caused by different changes in TTX-R sodium current of males and females²⁷. Since three different inflammatory mediators were used in the study, it is not possible to know whether sex-differences are solely caused by PGE₂.

Sex differences in the transcriptomes (RNAs bound to ribosomes) and transcriptomes of male and female mice DRG have been reported. In the prostaglandin pathway, prostaglandin D₂ synthase (*Ptgds*) was found to be upregulated in female DRG neurons. Female mice also exhibited higher levels of PTGDS protein and PGD₂¹⁵. PTGDS blockade produced more intense grimacing in male compared to female mice, suggesting that endogenous PGD₂ might reduce nociception in the absence of injury¹⁵. In the same study, it was also observed that female mice displayed more mechanical allodynia and grimacing after PGE₂ injection than male mice¹⁵. This study highlights the need to investigate sex-differences in the context of the unique function and expression of individual prostaglandins, as well as their interactions.

In studies investigating COX enzymes, female COX1 knockout mice and female COX2 knockout mice showed reduced edema and joint destruction in a model of Freund's adjuvant-induced arthritis compared to male mice. Female COX1 knockout mice also showed reduced contralateral allodynia compared with males²⁸. Similarly, another study demonstrated a reduced writhing response to acetic acid in COX2 deficient female mice. However, the anti-nociceptive effect was not seen in COX2 deficient male mice²⁹. On the other hand, NSAIDs that antagonize COX action appear to be less effective in females. The COX2 inhibitor celecoxib generated shorter and less prominent effects in female than male mice, when administered after Complete Freund's Adjuvant injection³⁰. COX inhibitors may also have different anti-inflammatory effects in males and females. In rats, indomethacin, a COX inhibitor, administered after experimental surgery decreased the elevated interleukin-6 levels in males only³¹. These seemingly conflicting results present an interesting paradox: while reduction/ablation of COX2 expression through genetic manipulation may be more effective in preventing nociceptive effects in females, reduction of COX2 through pharmacological inhibitors, such as NSAIDs, may be more effective in males. Several factors may contribute to this effect, including sex-differences in expression of genes related to drug metabolism and drug transporter expression³². In fact, the activity level of cytochrome p450 3A4, an enzyme involved in

drug metabolism, is higher in females³³. This could be consistent with lower COX-inhibitor efficacy in females, but the effects may also be mediated by differences in signaling at the level of the DRG nociceptor, as suggested by other studies described above.

Clinical studies. Clinical trials were subdivided in two ways. First, studies were categorized by how the pain conditions occurred: experimentally in the laboratory (n = 26) or clinically due to a disease process and/or procedural intervention (n = 74). Second, they were categorized by clinical model of disease and compared within these groups. Clinical trials were grouped by clinical model as follows: oral health (n = 10), surgical extraction of third molars (n = 14), hip and knee surgeries (n = 14), general surgeries (n = 13), tendon and joint disorders (n = 7), muscle pain (n = 7), experimentally induced skin hyperalgesia (n = 20), vascular system diseases (n = 5), experimental models of sleep deprivation (n = 2), headache (n = 2), and general studies of NSAIDs (n = 6). This grouping scheme allowed us to compare and contrast conclusions derived from relatively uniform settings. More details on the included clinical studies can be found in Supplementary Files 3 and 4.

Most clinical studies using the oral health model investigated whether levels of PGE₂ can be correlated with severity of gum disease and patient-reported pain. Other studies focused on the efficacy of COX inhibitors. However, no studies reported data separated by sex. One study found that there is variability in response to COX inhibitors. In third molar extraction surgery, some participants are partial responders to ibuprofen (4 men, 5 women), and others are complete responders (7 men, 3 women)³⁴. While the ratio of men to women in the complete responder group is greater than in the partial responder group to ibuprofen, the study did not analyze the difference in complete and partial responders in terms of potential sex differences, discussing only differences in levels of PG metabolites, cytokines, peripheral blood mononuclear cells gene expression between the two groups (Suppl. File 3)³⁴.

The analgesic and anti-inflammatory effects of various coxibs and nonselective COX inhibitors were compared in patients undergoing hip and knee treatment, e.g. total knee arthroplasty and hip replacement due to osteoarthritis. Only one study on knee arthroscopy observed that the women were at greater risk of developing moderate or severe pain³⁵. Other studies involving patients undergoing hip and knee procedures included both men and women but did not analyze data separated by sex or report sex-differences.

Experimental skin hyperalgesia was induced in several studies by injections of PGE₂ or low pH solutions into the muscle and skin, potentiating nociceptor activation³⁶. In a model of experimentally induced sunburn (UVB-induced erythema), ibuprofen (COX inhibitor) increased heat pain threshold and heat pain tolerance overall³⁷. However, the authors noted that men were more responsive to ibuprofen compared to women. Similarly, in a model of electrically induced pain, only men showed response to ibuprofen treatment and had higher pain tolerance^{38,39}.

Two studies on headache investigated whether headache is induced by inflammatory mediators^{40,41}. The authors did not report data separated by sex.

Subject selection and data analysis separated by sex. *Approximately 17% of preclinical studies examined included both sexes in subject selection and, out of those, 19% analyzed data separated by sex.* Preclinical studies analyzed in this semi-systematic review were published in years ranging from 1973 to November 2020. Among the 369 total studies analyzed, 202 studies involved the use of male animals only, 38 studies involved the use of female animals only, and 64 studies included the use of both male and female animals (Fig. 2A,B; Suppl. File 3). Several of these studies included both males and females, but in different experiments within the study, resulting in no direct comparisons. Additionally, 43 studies contained unclear information on the sex of the experimental animals, and 22 studies involved experimental setups where the sex information was not applicable, such as cell cultures (Suppl. File 3).

Out of the 369 total studies analyzed, only 64 studies were conducted using both male and female experimental animals. Out of the 64 studies that included male and female subjects, only 15 studies analyzed data separated by sex, regardless of whether they found significant sex differences or not. While 14 of these 15 studies included comments comparing prostaglandin-related action in males and females, one study presented data from each human tissue donor separately without further analyses⁴². We note that although studies in our analysis ranged from 1973–2020, all 15 studies that analyzed data separated by sex were published after 2000 (one in 2001, one in 2002, two in 2003, one in 2005, one in 2006, one in 2008, one in 2011, one in 2017, three in 2018, and three before our cutoff date in 2020). This suggests that the relevance of sex as a biological variable in data analysis is becoming more widely recognized. In addition, 12 out of the 15 studies that had sex as a biological variable in data analysis included animals only, and the remaining 3 out of these 15 studies included human tissues only. However, a much larger proportion of the studies that included both male and female subjects made at least some use of human tissues, at 25 out of 64 studies. The inclusion of human tissues from both male and female donors did not necessarily translate into greater data analysis with sex as a biological variable. An alternative interpretation is that there were no sex differences, but the authors did not report their negative findings on the lack of sex difference. Fourteen out of the 15 preclinical studies that analyzed data separated by sex found many differences between male and female animals in the prostaglandin pathways, behavior in response to noxious stimuli, etc. which we described in the first part of our results.

79% of clinical studies examined included both sexes and, out of those, approximately 7% analyzed data separated by sex. Contrary to preclinical studies, most clinical trials include both male and female participants (Fig. 2C,D; Suppl. File 4). However, few studies reported results separately for men and women. It is unclear whether the data from men and women were merged because no sex differences were found or if potential sex differences were not assessed at all. Some studies included subjects of only one biological sex. Similar to preclinical studies,

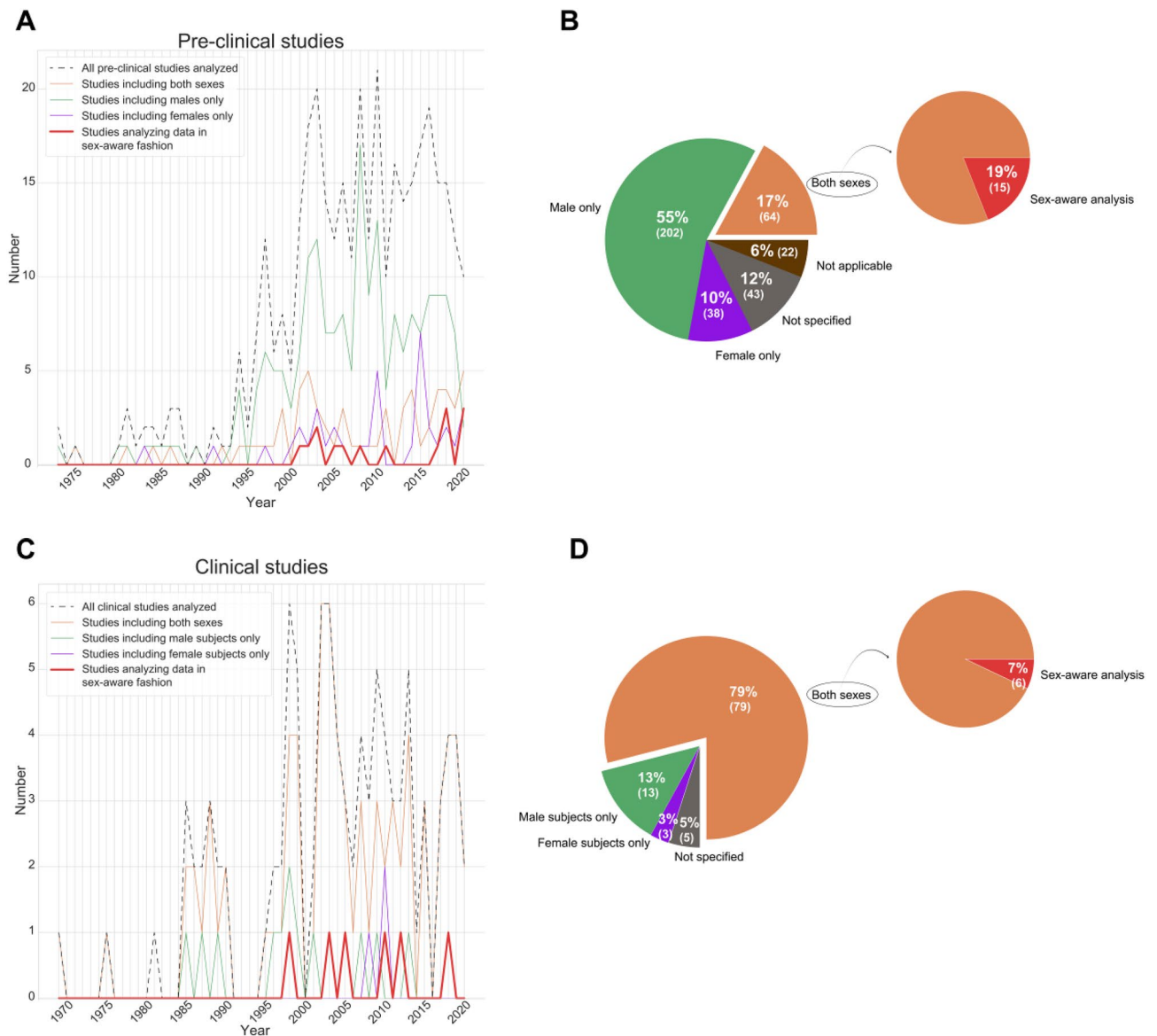


Figure 2. Most prostaglandin studies have not been analyzed with sex as a biological variable. **(A)** Line plot showing the inclusion of male, female or both sexes in pre-clinical studies from 1973 to 2020 and the number of studies that have been analyzed separated by sex (red line). **(B)** Pie charts showing the overall percentage of pre-clinical studies that include male, female or both sexes from 1973 to 2020. **(C)** Line plot showing the inclusion of male, female or both sexes in clinical studies from 1969 to 2020, including the studies that have analyzed the data separated by sex (red line). **(D)** Pie charts showing the overall percentage of clinical studies that include male, female or both sexes from 1969 to 2020. Note: studies where the sex of animals/subjects is not clear or specified are not shown in the line plots, please refer to supplementary files 1–4.

these single-sex studies in the experimental pain category showed a bias towards using male subjects (six studies involved men only, and none involved women only). In clinical pain conditions, the 71 studies that included both male and female subjects showed highly variable proportions of male to female participants. Numbers range from 187 men: 9 women (95.4% male)⁴³ to an equal split (10 men: 10 women)⁴⁴ to 3 men: 25 women⁴⁵. Additionally, only one study using a clinical pain model analyzed data separated by sex. This single study noted that female biological sex was a risk factor for moderate/severe postoperative pain after minor arthroscopic knee surgery³⁵. Another study focusing on the investigation of NSAID use found that women report using more over the counter pain medications⁴⁶. The remaining studies analyzed data from male and female participants together. In models of experimentally induced pain, the number of single-sex studies including male subjects only ($n = 13$) and female subjects only ($n = 3$) were also biased toward males only. Among the 4 experimental pain studies that analyzed data separated by sex, there were some findings that appeared to be contradictory. In an eccentric exercise model, no sex differences were noted in changes in muscle function, muscle soreness, or histological data⁴⁷, even though serum creatine kinase pre-values and change after the first bout of exercise were lower in females⁴⁷. On the other hand, a sunburn model showed that ibuprofen had a significantly greater effect

on lowering skin temperature in men compared with women³⁷. This finding in an inflammatory model agrees with similar conclusions made from two electrically-induced pain models. In these two electrical pain studies, ibuprofen had an analgesic effect only in males^{38,39}.

PTGDS is expressed in human DRG neurons and is enriched in neurons from female organ donors. Prostaglandin-D2 synthase (PTGDS) catalyzes the synthesis of prostaglandin D₂ (PGD₂), which is the most abundant prostaglandin in the brain^{48,49}. PGD₂ controls nociception, sleep and temperature^{21,22,50–58}. Previous research in our lab showed that PTGDS is enriched in female mouse DRG neurons¹⁵. We sought to investigate PTGDS expression in human DRG of age- and sex-matched donors using immunohistochemistry (IHC). We found that PTGDS is colocalized with the neuronal marker peripherin and is expressed in human DRG neurons (Fig. 3A). Next, we quantified PTGDS expression in human DRG from 12 organ donors (Table 1).

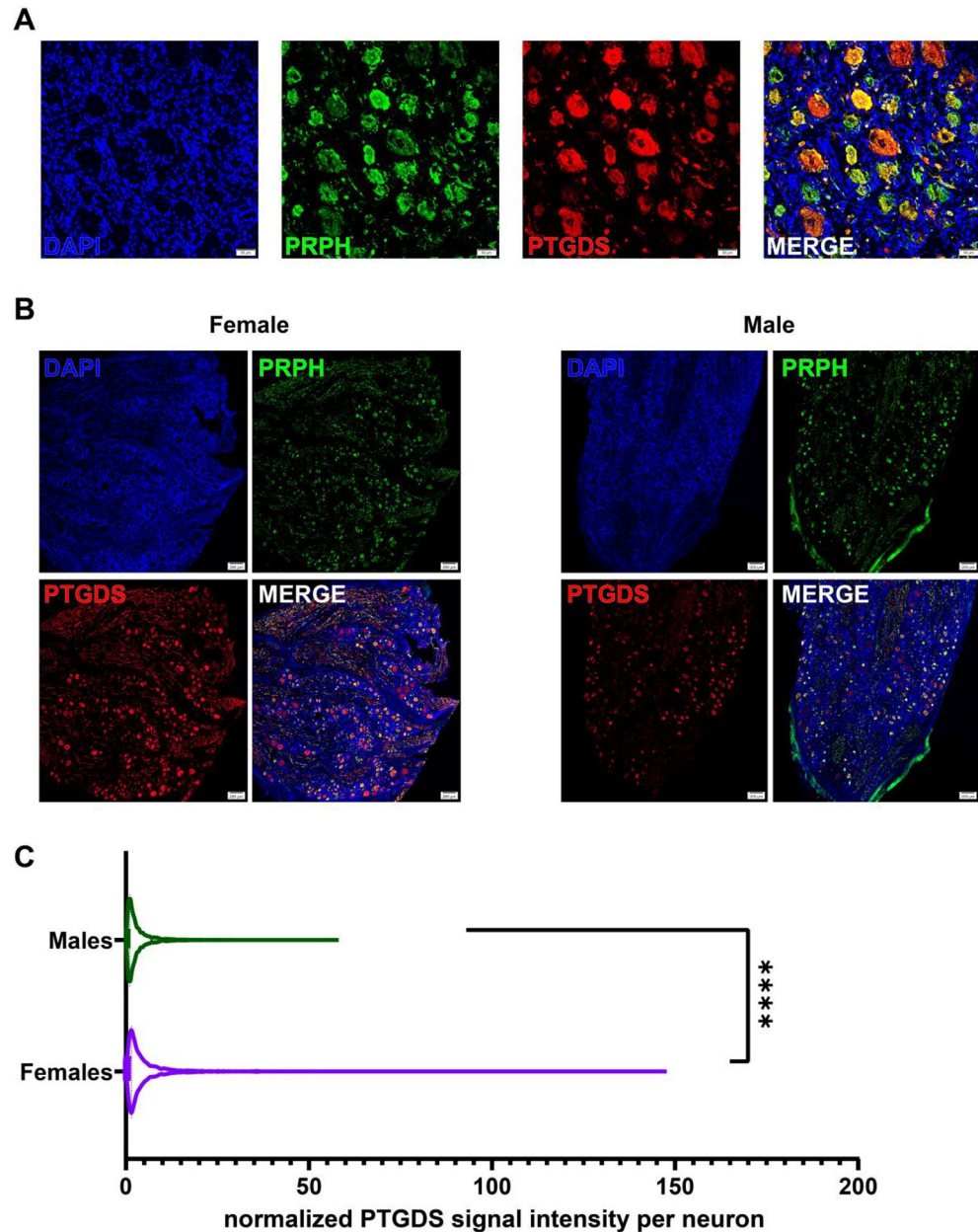


Figure 3. PTGDS is expressed in human DRG neurons. (A) Confocal images show co-localization of PTGDS (red) and neuronal marker peripherin (PRPH, green). Cell nuclei are labelled with DAPI (blue). Scale bar = 50 μ m. (B) Expression of PTGDS in female (left) and male (right) DRG neurons. Scale bar = 200 μ m. (C) PTGDS has higher expression in female DRG neurons compared to male DRGs (Unpaired *t* test with Welch's correction, $t = 11.08$, $df = 4299$, p -value < 0.0001). **** p -value < 0.0001 . Signal intensity was normalized by the area of the neurons. Data from donor #12 was excluded (post-menopause, 61-year-old female). In total 6813 neurons were analyzed (2920 from 5 female DRG samples and 3893 from 6 male DRG samples).

Donor number	Sex	Age	Cause of death
1	M	29	Head trauma/blunt injury
2	M	29	Head trauma/Gun shot wound
3	M	34	cerebral vascular accident/stroke
4	M	51	CVA/stroke
5	M	53	Anoxia/cardiac arrest
6	M	56	Anoxia/cardiac arrest
7	F	29	Anoxia/drug overdose
8	F	34	Anoxia/overdose
9	F	36	Head trauma/blunt injury/motor vehicle accident
10	F	44	CVA/stroke
11	F	53	CVA/stroke
12	F	61	Anoxia/cardiac arrest

Table 1. Donor information.

We observed that PTGDS expression is higher in sensory neurons from female organ donors when compared to male organ donors (Fig. 3B,C).

DP1 receptor expression in human DRG. PGD₂ acts through two receptors, DP1 and DP2. Previous transcriptomic studies demonstrate that the gene that encodes DP2, *PTGDR2*, is scarcely detected in human DRG^{14,59–61}. Therefore, we decided to focus on characterizing the expression of DP1 in human DRG. Transcriptomic studies on human DRG do not suggest sex differences in *PTGDR1* expression, but the cell type expressing the gene is not clear from these studies^{13,14,62}. Using IHC on both male and female DRG samples, our results suggest that DP1 is expressed mostly in glial cells surrounding neurons in human DRG, as it is highly colocalized

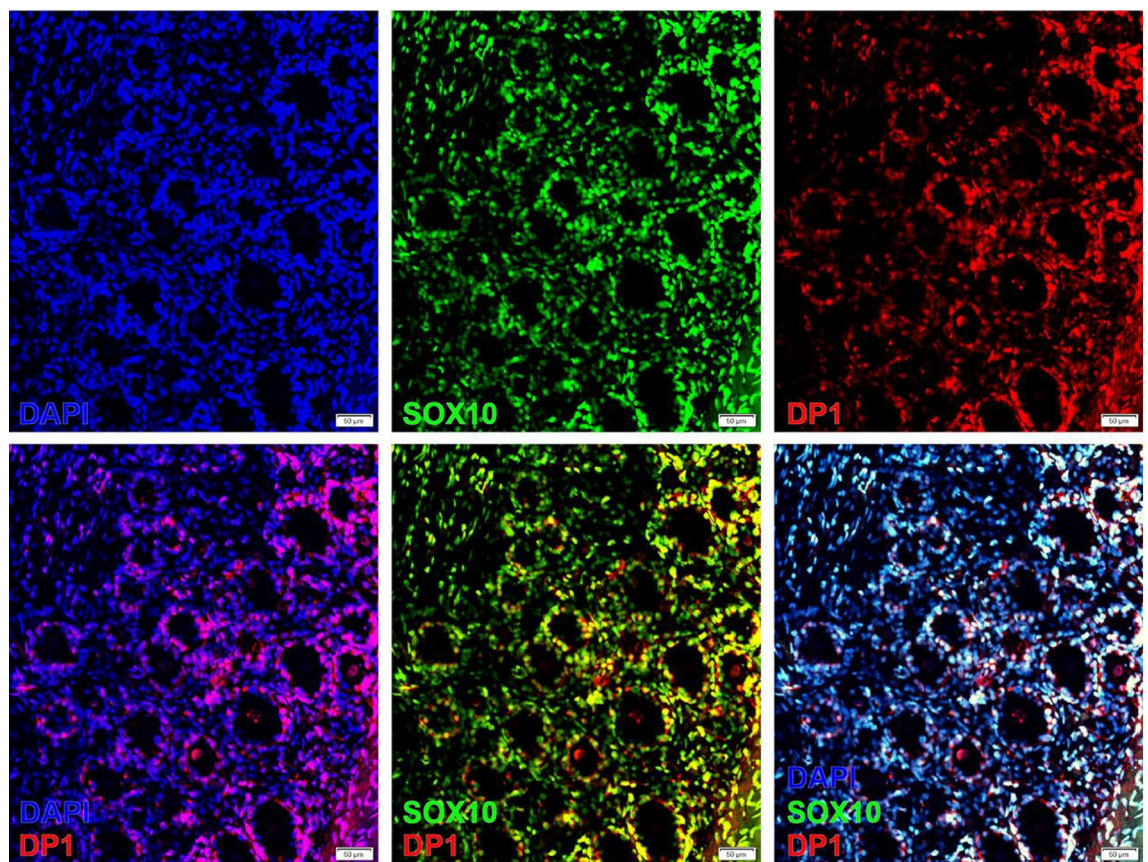


Figure 4. PGD₂ receptor, DP1, is expressed in cells surrounding human DRG neurons. DP1 (red) is co-localized with SOX10+ (green) cells in human DRG. Cell nuclei are labelled with DAPI (blue). Scale bar = 50 μm.

with SOX10, a glial cell marker (Fig. 4). These results suggest that PGD_2 may be released by neurons and acts via DP1 that is expressed in glial cells surrounding neurons. Glial cells and their inter-communication with neurons have been previously reported to have a role in pain processing^{63–65}.

Discussion

Our semi-systematic review reveals that although there are a small number of studies that either used both sexes or reported data analysis separated by sex, there is evidence for sex differences in prostaglandin-mediated effects in both animals and humans. A previous study has shown that PGE_2 signaling promotes pain more efficaciously in female mice¹⁵. There are also reports that COX inhibitors are more effective in men than in women in experimental pain models^{37–39}. Our experimental data confirms a finding from the rodent literature in humans where we observed that the expression of PTGDS is higher in female human DRG neurons from organ donors. Our findings highlight that there are likely important sex differences in signaling for one of the most widely studied classes of inflammatory mediators, the PGs.

In this review, we analyzed 100 clinical trial studies on the role of PGs and COX inhibitors in pain and inflammation. Most of these studies demonstrated the analgesic efficacy of selective and/or nonselective COX inhibitors. Patient PG levels were often compared with healthy controls, and changes in PG levels were tracked after administering COX inhibitors. Overall, both selective and nonselective COX inhibitors appear to be effective in treating inflammatory pain. However, it is not clear if no sex-differences were found or if the data was simply not analyzed with sex as a biological variable. In a study of NSAID prescriptions in the Danish population, women received more prescriptions for every NSAID than men⁶⁶. Additionally, a study included in our semi-systematic review found that women report more the use NSAIDs than men⁴⁶. In another study of patients receiving oxycodone, females received higher morphine equivalent daily dose of opioid than men⁶⁷. However, a study on postoperative opioid prescriptions after surgery showed that patient and provider demographic characteristics influenced doses prescribed, with males prescribed higher morphine milligram equivalents⁶⁸. Thus, from the data on prescriptions of analgesics, from NSAIDs to opioids, it is unclear whether women require greater amounts of painkillers to achieve equal relief as men, reflecting sex-differences in drug efficacy, or whether prescribing differences are influenced by social factors.

In addition to clinical pain studies, we analyzed 26 experimental pain studies. While two electrically-induced pain models showed a significant difference in ibuprofen efficacy between men and women^{38,39}, no difference was seen in a sunburn model of inflammatory pain³⁷. Some studies in both the experimental and clinical pain categories involved participants of only one sex. Interestingly, all the experimental pain studies using only one sex in this analysis happened to include male participants only. While less than a fifth of experimental pain studies analyzed data with sex as a biological variable, almost no clinical pain studies did so. As mentioned above, the experimental pain studies found sex differences in COX action with a greater effect in men. The single clinical pain study that reported data separated by sex found a difference in pain levels between men and women but did not report any difference in NSAID efficacy. This study on arthroscopic knee surgery observed that the women were at greater risk of developing moderate or severe pain³⁵. It is unclear whether the remaining papers did not report sex as a biological variable for data analysis because investigators determined there were no differences in the data from male and female participants, or simply because no analyses separated by sex were attempted.

In light of the evidence we present here for sex differences in PG action and COX inhibitor efficacy, we propose that clinical pain studies in this area should analyze and report data with sex as a biological variable. While most published clinical pain studies do not present data separated by sex, these studies often involve a large number of participants allowing for exploratory analyses that can form new hypotheses for further testing. If re-analyzed with sex as a biological variable, this trove of data could yield significant findings on whether COX inhibitors have a different efficacy between men and women. The findings of our review would lead us to hypothesize greater efficacy in men.

Functional differences in pain and COX-inhibitor efficacy may be at least partially explained by molecular differences in PG pathway receptor and enzyme gene expression. Previous work demonstrates that PTGDS is expressed more highly in female than male mouse DRG, so we evaluated whether this difference is conserved in humans¹⁵. Our findings show that in human DRG, PTGDS is localized within neurons, and expression is higher in female DRG neurons. Interestingly, among human donors of the same sex, there was some variation in DRG neuron PTGDS expression. Age of the donors, especially in females, as well as disease conditions, could potentially contribute to the variation. In our study, neuronal PTGDS expression in donor #12, a 61-year-old female, was nearly two times lower than that in the five other female donors, whose ages were in the pre-menopausal range. While it is difficult to reach firm conclusions from one donor, the finding could suggest that female sex hormones regulate PTGDS protein expression, a hypothesis that can be tested in future experiments. We also show that DP1 protein is highly colocalized in glial cells surrounding neurons in human DRG. This localization suggests that glial cells, likely satellite glial cells, may have an important role in mediating the functions of PGD_2 and regulating signaling effects by this prostaglandin within the DRG, and further studies are required to elucidate the exact mechanisms.

Reviews in other areas of biomedical research reported that there is a disconnect between the sex of study subjects and the respective disease prevalence rates^{69–71}. For instance, reviews of the pre-clinical⁷² and clinical⁷³ literature on depression have not only identified potential sex-differences but also noted the lack of inclusion of female subjects and analysis of data separated by sex. These examples show that a lack of sex-specific data analysis despite inclusion of both men and women subjects, is not limited to the prostaglandin signaling field.

Our findings highlight sex differences in prostaglandin effects on pain in both the preclinical and clinical literature that may have implications for development of future pain therapeutics targeting the prostaglandin system. Our work also points to the possibility that commonly used NSAIDs may have less efficacy in women

than in men, an effect that may also apply to other pain medications like opioids. Finally, our work validates a sex difference observed in PTGDS expression in rodent DRG in the human DRG. Collectively, this mix of semi-systematic review and experimental data points to shortcomings in our understanding of basic pain mechanisms in women. More emphasis is needed on understanding pain mechanisms in women if we are to adequately treat pain in the population that most frequently suffers from chronic pain disorders.

Materials and methods

This work was approved by the University of Texas at Dallas (UTD) Institutional Review Board (IRB). This is classified as IRB exempt research and the UTD IRB approval for this exemption is protocol #15-237 (The Human Dorsal Root Ganglion Transcriptome). Informed written consent for recovery of sensory tissue for research purposes was obtained from next to kin at Southwest Transplant Alliance.

Preclinical studies. *PubMed search.* A total of 1825 articles were retrieved from PubMed on November 20, 2020 using the following keywords: “prostaglandins, inflammation, and pain”. 369 papers were included for analysis (Fig. 5A).

Inclusion criteria. Articles of preclinical studies were found using the keywords mentioned above. Only papers that involved animal models and/or the use of human tissues were included. Only articles with PMID were included. A total of 369 papers were included.

Exclusion criteria. Papers were excluded if they were reviews ($n=488$), not written in English ($n=94$), had no full text available ($n=38$), studied plant extracts, alternative medicines, and other non-NSAID compounds ($n=337$), focused on pathways unrelated to PGs or COXs ($n=168$), studied drug dosing, delivery, or pharmacokinetics only ($n=99$), presented models or procedures only ($n=59$), or were clinical trials ($n=173$).

Data collection. A semi-systematic analysis was conducted for papers that analyzed data separated by sex. Information such as the specific PGs, COX enzymes, NSAIDs, and receptors investigated in each study were recorded.

Data analysis. The number of studies conducted each year with males, females, or both was recorded. For papers that included both male and female subjects, we also noted whether data was analyzed separated by sex. Finally, we examined any sex differences in PG or COX inhibitor actions (Suppl. Files 1 and 2).

Clinical Trials. *PubMed search.* A total of 104 papers on prostaglandins, inflammation, and pain were retrieved from PubMed on Feb 2, 2021. Papers were clinical trials, clinical trial protocols, clinical trial phases I–IV, and randomized controlled trials (Fig. 5B). An additional 34 papers on the same topic were retrieved from a PubMed search on November 20, 2020, without the above filters.

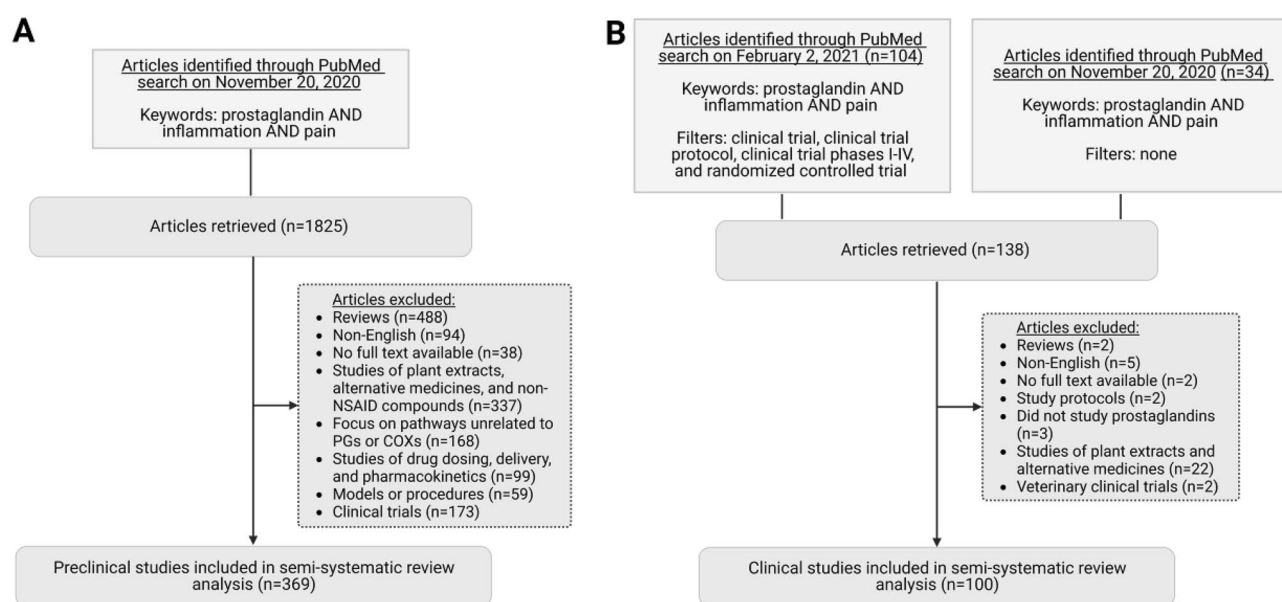


Figure 5. Semi-systematic review of articles retrieved on PubMed. **(A)** Workflow of pre-clinical studies identified through PubMed. **(B)** Workflow of clinical studies identified through PubMed.

Inclusion criteria. Clinical trial articles were included upon mention of all three search criteria, prostaglandins, inflammation, and pain. Included studies must also involve human subjects. Only articles with PMID were included. A total of 100 papers were included.

Exclusion criteria. Papers were excluded if they were reviews (n = 2), not written in English (n = 5), had no full text available (n = 2), described study protocols only (n = 2), did not study prostaglandins (n = 3), studied plant extracts or alternative medicine treatments (n = 22), or were veterinary clinical trials (n = 2).

Data collection. A semi-systematic analysis was conducted for all articles regardless of whether analysis included sex as a biological variable. We recorded the specific PGs, COX enzymes, NSAIDs, and receptors investigated. We also identified the purpose, experimental design, model, sex, number, group design, doses, injection type, measurements and tissues collected.

The papers analyzed include either clinical pain models (studies involving patients diagnosed with a pain condition, n = 74) or experimental pain conditions (n = 26). Experimental pain conditions include eccentric exercise-induced muscle pain (n = 4), experimentally induced skin hyperalgesia (n = 20), and experimental models of sleep deprivation (n = 2). All other studies involve clinical pain conditions, such as surgeries, joint pain, and painful dental conditions.

Data analysis. We categorized papers by clinical model and compared the main findings. Any discrepancies and/or agreements between studies were investigated in terms of divergent experimental methodology. We also recorded the number of clinical trials conducted each year with males, females, or both. Finally, we examined any sex differences in PG or COX inhibitor action (Suppl. Files 3 and 4).

Immunohistochemistry. Human dorsal root ganglia (DRG) from the L4 and L5 levels were recovered from organ donors, frozen immediately on crushed dry ice, and stored in a -80 °C freezer as previously described⁷⁴. The DRGs were embedded in OCT by gradually adding layers of OCT on the tissue, which was kept frozen over dry ice. Tissues were sectioned in the cryostat at 20 µm and adhered onto SuperFrost Plus charged slides (Thermo Fisher Scientific). Slides were kept in the -20 °C cryostat chamber for 15 min following completion of sectioning. The slides were then immediately fixed in ice-cold formalin (10%) for 1 min followed by dehydration in 50% ethanol (1 min), 70% ethanol (1 min), and 100% ethanol (2 min) at room temperature. The slides were briefly air dried. A hydrophobic pen (ImmEdge PAP Pen; Vector Labs) was used to draw boundaries around each tissue section, and boundaries were allowed to air dry.

Slides were incubated with blocking buffer (10% Normal Goat Serum, Atlanta Biologicals, Cat #S13150h, 0.3% Triton X-100 in 0.1 M PB) for 1 h at room temperature. Sections were then incubated overnight with a primary antibody cocktail. Following primary antibody incubation, sections were washed with 0.1 M phosphate buffer and incubated with Alexa Fluor secondary antibodies (Fisher Scientific/Invitrogen, dilutions 1:1000) for 1 h at room temperature. Sections were washed in 0.1 M phosphate buffer. To remove lipofuscin signal, Trublack (1:20 in 70% ethanol; Biotium #23007) was pipetted to cover each section for 1 min before being rinsed off. Finally, slides were air dried and cover slipped with Prolong Gold Antifade reagent (Fisher Scientific; P36930). The PTGDS antibody (ab18214, dilution 1:100) was obtained from Abcam and the peripherin antibody (P5117, dilution 1:500) was obtained from Sigma-Aldrich. The DP1 antibody (101640, dilution 1:200) was obtained from Cayman Chemicals and the SOX10 antibody (ab216020, dilution 1:40) was obtained from Abcam.

Image acquisition and PTGDS quantification. DRG sections with PTGDS staining (n = 12) were imaged on an Olympus VS120 Virtual Slide Microscope and Olympus FluoView 1200 confocal microscope, using the same settings for all images. Images were analyzed using Olympus CellSens software. The mean gray intensity of all neurons in a full DRG section from each donor was quantified (on average, we quantified 619 neurons per DRG section; the lowest number of neurons per DRG was 268). DRG sections with DP1 staining were imaged on an Olympus FV3000RS Confocal Laser Scanning Microscope.

Statistics. Statistical analysis for PTGDS quantification was done in GraphPad Prism 9.3.1. The mean gray intensity values were normalized by the area of the neurons. Single comparisons were performed on all neurons using Student's *t* test with Welch's correction (which accounts for the unequal variances per group). For statistical analysis, data from the 61-year-old female (donor #12) was excluded (post-menopause). In total 6813 neurons were analyzed (2920 from 5 female DRG samples and 3893 from 6 male DRG samples). Statistical results can be found in Fig. 3 legend.

Data visualization for Fig. 2 was done in Python (version 3.8.5 with Anaconda distribution). Figures 1 and 5 were generated using Biorender (BioRender.com).

Ethics approval. This work was approved by the University of Texas at Dallas (UTD) Institutional Review Board (IRB). This is classified as IRB exempt research and the UTD IRB approval for this exemption is protocol #15-237 (The Human Dorsal Root Ganglion Transcriptome). Informed written consent for recovery of sensory tissue for research purposes was obtained from next to kin at Southwest Transplant Alliance.

Data availability

Data is available from corresponding authors upon request.

Received: 14 December 2022; Accepted: 14 March 2023

Published online: 22 March 2023

References

- Arout, C. A., Sofuoglu, M., Bastian, L. A. & Rosenheck, R. A. Gender differences in the prevalence of fibromyalgia and in comorbid medical and psychiatric disorders: A National Veterans Health Administration study. *J. Womens Health (Larchmt)* **27**, 1035–1044. <https://doi.org/10.1089/jwh.2017.6622> (2018).
- Vetvik, K. G. & MacGregor, E. A. Sex differences in the epidemiology, clinical features, and pathophysiology of migraine. *Lancet Neurol.* **16**, 76–87. [https://doi.org/10.1016/S1474-4422\(16\)30293-9](https://doi.org/10.1016/S1474-4422(16)30293-9) (2017).
- Vina, E. R. & Kwok, C. K. Epidemiology of osteoarthritis: Literature update. *Curr. Opin. Rheumatol.* **30**, 160–167. <https://doi.org/10.1097/BOR.0000000000000479> (2018).
- Fillingim, R. B., King, C. D., Ribeiro-Dasilva, M. C., Rahim-Williams, B. & Riley, J. L. 3rd. Sex, gender, and pain: A review of recent clinical and experimental findings. *J. Pain* **10**, 447–485. <https://doi.org/10.1016/j.jpain.2008.12.001> (2009).
- Mogil, J. S. Laboratory environmental factors and pain behavior: The relevance of unknown unknowns to reproducibility and translation. *Lab. Anim. (NY)* **46**, 136–141. <https://doi.org/10.1038/labana.1223> (2017).
- Mogil, J. S. Qualitative sex differences in pain processing: Emerging evidence of a biased literature. *Nat. Rev. Neurosci.* **21**, 353–365. <https://doi.org/10.1038/s41583-020-0310-6> (2020).
- Rosen, S., Ham, B. & Mogil, J. S. Sex differences in neuroimmunity and pain. *J. Neurosci. Res.* **95**, 500–508. <https://doi.org/10.1002/jnr.23831> (2017).
- Avona, A. *et al.* Meningeal CGRP-prolactin interaction evokes female-specific migraine behavior. *Ann. Neurol.* **89**, 1129–1144. <https://doi.org/10.1002/ana.26070> (2021).
- Patil, M. *et al.* Prolactin receptor expression in mouse dorsal root ganglia neuronal subtypes is sex-dependent. *J. Neuroendocrinol.* **31**, e12759. <https://doi.org/10.1111/jne.12759> (2019).
- Paige, C. *et al.* A female-specific role for calcitonin gene-related peptide (CGRP) in rodent pain models. *J. Neurosci.* **42**, 1930–1944. <https://doi.org/10.1523/JNEUROSCI.1137-21.2022> (2022).
- Avona, A. *et al.* Dural calcitonin gene-related peptide produces female-specific responses in rodent migraine models. *J. Neurosci.* **39**, 4323–4331. <https://doi.org/10.1523/JNEUROSCI.0364-19.2019> (2019).
- Ray, P. R. *et al.* Transcriptome analysis of the human tibial nerve identifies sexually dimorphic expression of genes involved in pain, inflammation, and neuro-immunity. *Front. Mol. Neurosci.* **12**, 37. <https://doi.org/10.3389/fnmol.2019.00037> (2019).
- Ray, P. R. *et al.* RNA profiling of human dorsal root ganglia reveals sex-differences in mechanisms promoting neuropathic pain. *Brain* <https://doi.org/10.1093/brain/awac266> (2022).
- Tavares-Ferreira, D. *et al.* Spatial transcriptomics of dorsal root ganglia identifies molecular signatures of human nociceptors. *Sci. Transl. Med.* **14**, eabj8186. <https://doi.org/10.1126/scitranslmed.abj8186> (2022).
- Tavares-Ferreira, D. *et al.* Sex differences in nociceptor transcriptomes contribute to divergent prostaglandin signaling in male and female mice. *Biol. Psychiatry* **91**, 129–140. <https://doi.org/10.1016/j.biopsych.2020.09.022> (2022).
- Ricciotti, E. & FitzGerald, G. A. Prostaglandins and inflammation. *Arterioscler. Thromb. Vasc. Biol.* **31**, 986–1000. <https://doi.org/10.1161/ATVBAHA.110.207449> (2011).
- Natura, G. *et al.* Neuronal prostaglandin E2 receptor subtype EP3 mediates antinociception during inflammation. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 13648–13653. <https://doi.org/10.1073/pnas.1300820110> (2013).
- Zeisel, A. *et al.* Molecular architecture of the mouse nervous system. *Cell* **174**, 999–1014 e1022. <https://doi.org/10.1016/j.cell.2018.06.021> (2018).
- Chen, L., Yang, G. & Grosser, T. Prostanoids and inflammatory pain. *Prostaglandins Other Lipid Mediat.* **104–105**, 58–66. <https://doi.org/10.1016/j.prostaglandins.2012.08.006> (2013).
- Davidson, S. *et al.* Group II mGluRs suppress hyperexcitability in mouse and human nociceptors. *Pain* **157**, 2081–2088. <https://doi.org/10.1097/j.pain.0000000000000621> (2016).
- Eguchi, N. *et al.* Lack of tactile pain (allodynia) in lipocalin-type prostaglandin D synthase-deficient mice. *Proc. Natl. Acad. Sci. U.S.A.* **96**, 726–730. <https://doi.org/10.1073/pnas.96.2.726> (1999).
- Minami, T., Okuda-Ashitaka, E., Nishizawa, M., Mori, H. & Ito, S. Inhibition of nociceptin-induced allodynia in conscious mice by prostaglandin D2. *Br. J. Pharmacol.* **122**, 605 (1997).
- Buisseret, B., Alhouayek, M., Guillemot-Legrès, O. & Muccioli, G. G. Endocannabinoid and prostanoid crosstalk in pain. *Trends Mol. Med.* **25**, 882–896. <https://doi.org/10.1016/j.molmed.2019.04.009> (2019).
- Nonsteroidal anti-inflammatory drugs: Adverse effects and their prevention. *Semin. Arthritis Rheum.* **39**, 294–312. <https://doi.org/10.1016/j.semarthrit.2008.08.001> (2010).
- Langford, R. M. & Mehta, V. Selective cyclooxygenase inhibition: Its role in pain and anaesthesia. *Biomed. Pharmacother.* **60**, 323–328. <https://doi.org/10.1016/j.biopha.2006.06.017> (2006).
- Zhang, P., Bi, R. Y. & Gan, Y. H. Glial interleukin-1 β upregulates neuronal sodium channel 1.7 in trigeminal ganglion contributing to temporomandibular joint inflammatory hypernociception in rats. *J. Neuroinflammation* **15**, 117. <https://doi.org/10.1186/s12974-018-1154-0> (2018).
- Scheff, N. N. & Gold, M. S. Sex differences in the inflammatory mediator-induced sensitization of dural afferents. *J. Neurophysiol.* **106**, 1662–1668. <https://doi.org/10.1152/jn.00196.2011> (2011).
- Chillingworth, N. L., Morham, S. G. & Donaldson, L. F. Sex differences in inflammation and inflammatory pain in cyclooxygenase-deficient mice. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **291**, R327–R334. <https://doi.org/10.1152/ajpregu.00901.2005> (2006).
- Racz, I., Schutz, B., Abo-Salem, O. M. & Zimmer, A. Visceral, inflammatory and neuropathic pain in glycine receptor alpha 3-deficient mice. *NeuroReport* **16**, 2025–2028. <https://doi.org/10.1097/00001756-200512190-00011> (2005).
- Liu, L. *et al.* Sex differences revealed in a mouse CFA inflammation model with macrophage targeted nanotherapeutics. *Theranostics* **10**, 1694–1707. <https://doi.org/10.7150/thno.41309> (2020).
- Page, G. G. & Ben-Eliyahu, S. Indomethacin attenuates the immunosuppressive and tumor-promoting effects of surgery. *J. Pain* **3**, 301–308. <https://doi.org/10.1054/jpai.2002.125184> (2002).
- Fisher, J. L. *et al.* Considerations and challenges for sex-aware drug repurposing. *Biol. Sex Differ.* **13**, 13. <https://doi.org/10.1186/s13293-022-00420-8> (2022).
- Sukoff Rizzo, S. J., McTighe, S. & McKinzie, D. L. Genetic background and sex: Impact on generalizability of research findings in pharmacology studies. *Handb. Exp. Pharmacol.* **257**, 147–162. https://doi.org/10.1007/164_2019_282 (2020).
- Theken, K. N. *et al.* Variability in the analgesic response to ibuprofen is associated with cyclooxygenase activation in inflammatory pain. *Clin. Pharmacol. Ther.* **106**, 632–641. <https://doi.org/10.1002/cpt.1446> (2019).
- Solheim, N. *et al.* Randomized controlled trial of intra-articular ketorolac on pain and inflammation after minor arthroscopic knee surgery. *Acta Anaesthesiol. Scand.* **62**, 829–838. <https://doi.org/10.1111/aas.13104> (2018).
- Namer, B. *et al.* Differential sensitization of silent nociceptors to low pH stimulation by prostaglandin E2 in human volunteers. *Eur. J. Pain* **19**, 159–166. <https://doi.org/10.1002/ejp.532> (2015).
- Syha, T. *et al.* A simple pain model for the evaluation of analgesic effects of NSAIDs in healthy subjects. *Br. J. Clin. Pharmacol.* **56**, 165–172. <https://doi.org/10.1046/j.0306-5251.2003.01869.x> (2003).

38. Walker, J. S. & Carmody, J. J. Experimental pain in healthy human subjects: Gender differences in nociception and in response to ibuprofen. *Anesth. Analg.* **86**, 1257–1262. <https://doi.org/10.1097/0000539-199806000-00023> (1998).
39. Butcher, B. E. & Carmody, J. J. Sex differences in analgesic response to ibuprofen are influenced by expectancy: A randomized, crossover, balanced placebo-designed study. *Eur. J. Pain* **16**, 1005–1013. <https://doi.org/10.1002/j.1532-2149.2011.00104.x> (2012).
40. Ashina, M. *et al.* Tender points are not sites of ongoing inflammation—In vivo evidence in patients with chronic tension-type headache. *Cephalalgia* **23**, 109–116. <https://doi.org/10.1046/j.1468-2982.2003.00520.x> (2003).
41. Antonova, M., Wienecke, T., Olesen, J. & Ashina, M. Pro-inflammatory and vasoconstricting prostanoid PGF₂α causes no headache in man. *Cephalalgia* **31**, 1532–1541. <https://doi.org/10.1177/0333102411423314> (2011).
42. Burke, J. G. *et al.* Human nucleus pulposus can respond to a pro-inflammatory stimulus. *Spine (Phila Pa 1976)* **28**, 2685–2693. <https://doi.org/10.1097/01.BRS.0000103341.45133.F3> (2003).
43. Gottesdiener, K. *et al.* Efficacy and tolerability of the specific cyclooxygenase-2 inhibitor DFP compared with naproxen sodium in patients with postoperative dental pain. *Clin. Ther.* **21**, 1301–1312. [https://doi.org/10.1016/s0149-2918\(99\)80031-9](https://doi.org/10.1016/s0149-2918(99)80031-9) (1999).
44. Cullen, L., Kelly, L., Connor, S. O. & Fitzgerald, D. J. Selective cyclooxygenase-2 inhibition by nimesulide in man. *J. Pharmacol. Exp. Ther.* **287**, 578–582 (1998).
45. Nishimura, M., Segami, N., Kaneyama, K., Suzuki, T. & Miyamaru, M. Relationships between pain-related mediators and both synovitis and joint pain in patients with internal derangements and osteoarthritis of the temporomandibular joint. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* **94**, 328–332. <https://doi.org/10.1067/moe.2002.124106> (2002).
46. Wilcox, C. M., Cryer, B. & Triadafilopoulos, G. Patterns of use and public perception of over-the-counter pain relievers: Focus on nonsteroidal antiinflammatory drugs. *J. Rheumatol.* **32**, 2218–2224 (2005).
47. Paulsen, G. *et al.* A COX-2 inhibitor reduces muscle soreness, but does not influence recovery and adaptation after eccentric exercise. *Scand. J. Med. Sci. Sports* **20**, e195–e207. <https://doi.org/10.1111/j.1600-0838.2009.00947.x> (2010).
48. Shaik, J. S., Miller, T. M., Graham, S. H., Manole, M. D. & Poloyac, S. M. Rapid and simultaneous quantitation of prostanoids by UPLC-MS/MS in rat brain. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **945–946**, 207–216. <https://doi.org/10.1016/j.jchromb.2013.11.041> (2014).
49. Ogorochi, T. *et al.* Regional distribution of prostaglandins D₂, E₂, and F₂α and related enzymes in postmortem human brain. *J. Neurochem.* **43**, 71–82 (1984).
50. Ito, S., Okuda-Ashitaka, E. & Minami, T. Central and peripheral roles of prostaglandins in pain and their interactions with novel neuropeptides nociceptin and nocistatin. *Neurosci. Res.* **41**, 299–332 (2001).
51. Liang, X., Wu, L., Hand, T. & Andreasson, K. Prostaglandin D₂ mediates neuronal protection via the DP₁ receptor. *J. Neurochem.* **92**, 477–486. <https://doi.org/10.1111/j.1471-4159.2004.02870.x> (2005).
52. Minami, T., Okuda-Ashitaka, E., Mori, H., Ito, S. & Hayaishi, O. Prostaglandin D₂ inhibits prostaglandin E₂-induced allodynia in conscious mice. *J. Pharmacol. Exp. Ther.* **278**, 1146–1152 (1996).
53. Ohkubo, T., Shibata, M., Takahashi, H. & Inoki, R. Effect of prostaglandin D₂ on pain and inflammation. *Jpn. J. Pharmacol.* **33**, 264–266. <https://doi.org/10.1254/jjp.33.264> (1983).
54. Sekeroglu, A. *et al.* Effect of PGD₂ on middle meningeal artery and mRNA expression profile of L-PGD₂ synthase and DP receptors in trigeminovascular system and other pain processing structures in rat brain. *Pharmacol. Rep.* **69**, 50–56. <https://doi.org/10.1016/j.pharep.2016.09.015> (2017).
55. Shimizu, T., Mizuno, N., Amano, T. & Hayaishi, O. Prostaglandin D₂, a neuromodulator. *Proc. Natl. Acad. Sci.* **76**, 6231–6234 (1979).
56. Uda, R., Horiguchi, S., Ito, S., Hyodo, M. & Hayaishi, O. Nociceptive effects induced by intrathecal administration of prostaglandin D₂, E₂, or F₂ α to conscious mice. *Brain Res.* **510**, 26–32. [https://doi.org/10.1016/0006-8993\(90\)90723-o](https://doi.org/10.1016/0006-8993(90)90723-o) (1990).
57. Urade, Y. & Hayaishi, O. Prostaglandin D₂ and sleep regulation. *Biochim. Biophys. Acta (BBA) Mol. Cell Biol. Lipids* **1436**, 606–615 (1999).
58. Wang, T. A. *et al.* Thermoregulation via temperature-dependent PGD₂ production in mouse preoptic area. *Neuron* **103**, 309–322. e307 (2019).
59. Ray, P. *et al.* Comparative transcriptome profiling of the human and mouse dorsal root ganglia: An RNA-seq-based resource for pain and sensory neuroscience research. *Pain* **159**, 1325–1345. <https://doi.org/10.1097/j.pain.0000000000001217> (2018).
60. Wangzhou, A. *et al.* Pharmacological target-focused transcriptomic analysis of native vs cultured human and mouse dorsal root ganglia. *Pain* **161**, 1497–1517. <https://doi.org/10.1097/j.pain.0000000000001866> (2020).
61. Wangzhou, A. *et al.* A ligand-receptor interactome platform for discovery of pain mechanisms and therapeutic targets. *Sci. Signal.* **14**, eabe1648. <https://doi.org/10.1126/scisignal.abe1648> (2021).
62. North, R. Y. *et al.* Electrophysiological and transcriptomic correlates of neuropathic pain in human dorsal root ganglion neurons. *Brain* **142**, 1215–1226. <https://doi.org/10.1093/brain/awz063> (2019).
63. Yuan, Q. *et al.* Satellite glia activation in dorsal root ganglion contributes to mechanical allodynia after selective motor fiber injury in adult rats. *Biomed. Pharmacother.* **127**, 110187 (2020).
64. Chen, Z. *et al.* Purinergic signaling between neurons and satellite glial cells of mouse dorsal root ganglia modulates neuronal excitability in vivo. *Pain* **163**, 1636–1647 (2022).
65. Ji, R.-R., Berta, T. & Nedergaard, M. Glia and pain: Is chronic pain a gliopathy? *Pain* **154**, S10–S28 (2013).
66. Fosbol, E. L. *et al.* The pattern of use of non-steroidal anti-inflammatory drugs (NSAIDs) from 1997 to 2005: A nationwide study on 4.6 million people. *Pharmacoepidemiol. Drug Saf.* **17**, 822–833. <https://doi.org/10.1002/pds.1592> (2008).
67. Barrachina, J. *et al.* Sex differences in oxycodone/naloxone vs. tapentadol in chronic non-cancer pain: An observational real-world study. *Biomedicines* **10**, 2468. <https://doi.org/10.3390/biomedicines10102468> (2022).
68. Zaveri, S., Nobel, T. B., Khetan, P., Srinivasan, M. & Divino, C. M. Surgeon bias in postoperative opioid prescribing. *World J. Surg.* **46**, 1660–1666. <https://doi.org/10.1007/s00268-022-06532-x> (2022).
69. Weinberger, A. H., McKee, S. A. & Mazure, C. M. Inclusion of women and gender-specific analyses in randomized clinical trials of treatments for depression. *J. Womens Health* **19**, 1727–1732 (2010).
70. Steinberg, J. R. *et al.* Analysis of female enrollment and participant sex by burden of disease in US clinical trials between 2000 and 2020. *JAMA Netw. Open* **4**, e2113749 (2021).
71. Dekker, M. J. *et al.* Sex proportionality in pre-clinical and clinical trials: An evaluation of 22 marketing authorization application dossiers submitted to the European medicines Agency. *Front. Med.* **8**, 643028 (2021).
72. Alshammari, T. K. Sexual dimorphism in pre-clinical studies of depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **105**, 110120 (2021).
73. Labaka, A., Goñi-Balentiaga, O., Lebeña, A. & Pérez-Tejada, J. Biological sex differences in depression: A systematic review. *Biol. Res. Nurs.* **20**, 383–392 (2018).
74. Shiers, S. I. *et al.* Convergence of peptidergic and non-peptidergic protein markers in the human dorsal root ganglion and spinal dorsal horn. *J. Comp. Neurol.* **529**, 2771–2788. <https://doi.org/10.1002/cne.25122> (2021).

Acknowledgements

The authors thank the organ donors and their families for their enduring gift. We thank employees of the Southwest Transplant Alliance and members of the Price Lab for coordinating and assisting in DRGs recoveries, respectively.

Author contributions

B.Q.S. and D.T.-F. conducted PubMed searches and defined inclusion and exclusion criteria. B.Q.S. performed the review and analysis of articles. B.Q.S. conducted immunohistochemistry, imaging and quantification. B.Q.S. and T.J.P. conducted the statistical analysis. B.Q.S. and D.T.-F. created figures. I.S. assisted in immunohistochemistry experiments and PTGDS quantification. B.Q.S., D.T.-F. and T.J.P. wrote the manuscript. D.T.-F. and T.J.P. designed and supervised the study. All authors read and edited the paper.

Funding

Funding to TJP NIH grant NS065926 and TJP and DT-F NIH grant U19NS130608.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-023-31603-x>.

Correspondence and requests for materials should be addressed to T.J.P. or D.T.-F.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2023