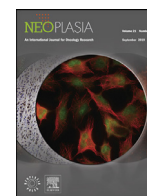


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Why haven't we solved intracranial pediatric ependymoma? Current questions and barriers to treatment advances.

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ABSTRACT

Pediatric intracranial ependymoma has seen a recent exponential expansion of biological findings, rapidly dividing the diagnosis into several subgroups, each with specific molecular and clinical characteristics. While such subdivision may complicate clinical conclusions from historical trials, this knowledge also provides an opportunity for interrogating the major clinical and biological questions preventing near-term translation into effective therapy for children with ependymoma. In this article, we briefly review some of the most critical clinical questions facing both patient management and the construct of future trials in childhood ependymoma, as well as explore some of the current barriers to efficient translation of preclinical discovery to the clinic.

Introduction

The management of childhood intracranial ependymomas remains an ongoing challenge for pediatric neuro-oncologists. Although these tumors are relatively common, their heterogeneity, in both clinical presentation and biology, has clouded best practice guidelines and left some clinical questions unanswered. The ongoing molecular classification of ependymomas, which has allowed for the correlation of molecular findings to clinical outcomes, suggests that subsets of these tumors may require radically different therapeutic approaches.

While the current ambiguity in treatment guidelines may understandably create apprehension for both families and providers, the re-

cent discoveries in ependymoma also present an opportunity to dramatically shift the landscape of these challenging tumors. Through better understanding of ependymoma biology, careful analysis of clinical data, and thoughtful trial design, physicians and researchers can shape the future treatment of ependymomas and provide evidence-based care that is uniquely tailored to each individual case. Comprehensive biological and clinical reviews have been recently published and are beyond the scope of this article; [1–11] instead, this review, from the Pacific Pediatric Neuro-oncology Consortium (PNOC) ependymoma working group, will aim to briefly highlight some of the clinical and biological questions currently being actively investigated and describe some barriers to translational therapeutic efforts.

Abbreviations: Alpha thalassemia/mental retardation syndrome X-linked, ATRX; Anti-programmed death ligand 1, PDL1; Associazione Italiana Ematologia Oncologia Pediatrica, AIEOP; Bromodomain and extra terminal motif, BET; Chimeric antigen receptor, CAR; Central nervous system, CNS; Children's Cancer Group, CCG; Children's Oncology Group, COG; Clustered Regularly Interspaced Short Palindromic Repeats, CRISPR; Collaborative Ependymoma Research Network, CERN; Confidence Interval, CI; Cyclin dependent inhibitor 2A, CDKN2A; Diffuse midline glioma, DMG; Deoxyribonucleic acid, DNA; Enhancer of zeste homolog 2, EZH2; Ependymoma, EPN; Event free survival, EFS; EZH2 interacting protein, EZHIP; Fibroblast growth factor receptor, FGFR; Global Ependymoma Network of Excellence, GENE; Gross total resection, GTR; Human epidermal growth factor receptor 2, HER2; Insulin-like growth factor receptor, IGFR; International Society of Pediatric Oncology, SIOP; In utero electroporation, IUE; Pacific Pediatric Neuro-oncology Consortium, PNOC; Patient derived xenograft, PDX; Pediatric Oncology Group, POG; Platelet derived growth factor receptor A, PDGFRA; Posterior fossa, PF; Posterior fossa group A, PFA; Posterior fossa group B, PFB; Progression free survival, PFS; Receptor tyrosine kinase, RTK; Replication-Competent Avian Sarcoma-leukosis virus long-terminal repeat with splice acceptor, RCAS-TVa; Societe Francaise d'Oncologie Pediatrique, SFOP; TEA domain, TEAD; TF, transcription factor; World Health Organization, WHO; Yes associated protein 1, YAP1; Zinc finger translocation associated, ZFTA.

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General background

Ependymoma is the third most common pediatric central nervous system (CNS) tumor, accounting for 8-10% of all diagnoses [12], and occurring most frequently in the posterior fossa, followed by the supratentorial area and, less commonly, the spinal cord [13]. Unfortunately, outcomes for ependymomas are suboptimal, with five-year progression-free and overall survival of 23-45% and 50-64%, respectively [14-17]. The current standard of care for ependymoma is maximal surgical resection followed by radiotherapy to the primary site. Management, outcomes, and tumor biology are distinct from adult tumors, typically with more rapid and prevalent recurrence, which may be delayed although portend poor outcomes.

Tumor location may play an important role in patient outcomes due to its impact on neurosurgical management as well as location-associated molecular drivers that determine the biological aggressiveness of these tumors [12]. Due to difficulties with obtaining complete surgical resection, ependymomas arising from the floor and the lateral aspect of the fourth ventricle have a worse prognosis than those arising from the roof [8]. Supratentorial ependymomas are typically seen in younger patients (age 0-12 years) and grow more uncommon with age [18], but with an increased ability to achieve gross-total resection, as well as distinct biological drivers, they are associated with an improved prognosis [8]. Because of the strong impact of extent of resection on prognosis, second look surgeries or cytoreductive chemotherapy should strongly be considered to achieve a complete response, especially as, upon review, they do not appear to increase mortality or permanent morbidity and provide an avenue to significantly reduce tumor volume [19]. Following surgical resection of their ependymoma, patients are commonly treated with radiotherapy with possible adjuvant chemotherapy, although some patients with completely resected, more indolent entities such as supratentorial ependymomas or posterior fossa B-type tumors may be able to be managed with observation alone [2].

Over the past decade, molecular features have revealed substantial differences in driver biology and epigenetic characteristics, leading to significant revisions in tumor stratification and identification of novel potential targets. While a comprehensive review of the current landscape is best approached in already-published articles [1], currently there exist at least six different intracranial subtypes in pediatric, including posterior fossa ependymoma group A (PFA or PF-EPN-A), or B (PFB or PF-EPN-B), supratentorial ependymoma *YAPI* (ST-EP-*YAPI*), supratentorial ependymoma *RELA* (ST-EP-*RELA*, now termed *ZFTA*), and both posterior fossa or supratentorial subependymoma [20]. Some subtypes are particularly distinctive in their outcomes or biology. For instance, within the posterior fossa, distinct cytogenetic patterns have been associated with prognostic outcomes. Tumors with a chromosome 1q gain and other partial structural chromosomal alterations have a worse prognosis, while those displaying a balanced profile without alterations have a slightly better prognosis [8,21-23]. These findings have, for example, opened the possibility for distinct treatment strategies that may achieve improved efficacy and reduce the significant morbidity associated with ependymoma treatment.

There is currently no standard of care for recurrent ependymoma in children. Children who receive incomplete resections of their tumor are at much higher risk for recurrence than those who receive a gross-total resection. Treatment strategies for children with recurrent ependymoma include re-resection and re-irradiation when feasible. [22] Chemotherapy has typically not demonstrated significant benefit in patients with recurrent tumor. [23] There have been documented long-term responses to metronomic anti-angiogenic regimens, but these treatments are generally not felt to be curative [24,25]. As understanding of the molecular and biologic features of pediatric ependymoma improves, targeted therapies for relapsed ependymoma may be identified and investigated through early-phase clinical trials.

Current questions in pediatric ependymoma management

As understanding of the heterogeneity of ependymoma matures, questions about interpretation of clinical results lacking molecular annotation present an obstacle to subtype-specific treatment strategies. Interpretation of small and large retrospective studies containing molecular annotation but lacking a cohesive common treatment strategy also present difficulties regarding sweeping conclusions in appropriate management. In the following section, a few of the more commonly debated questions will be addressed.

Chemotherapy: is it helpful in ependymoma?

Chemotherapy has long held a prominent role in the treatment of childhood cancer; however, its utility in ependymoma has been questioned. Historically, although not in the context of known molecular subtypes, several general chemotherapy-induced responses generated hope for potential benefit. The Children's Cancer Group (CCG) CCG9942 study showed that many of the 34 patients with residual disease who received pre-radiation chemotherapy (cisplatin, vincristine, cyclophosphamide, etoposide) had significant responses (42% complete response, 18% partial response, 26% stable disease), although in this study 15% progressed prior to irradiation; further, the 3-year event-free survival of these patients (58%) was not significantly different from patients who received a gross total resection (GTR) followed by irradiation (62%). [24] These results were replicated in the International Society for Pediatric Oncology (SIOP) Ependymoma I trial, with a 65% response rate in newly diagnosed, subtotally-resected ependymoma. [25] Other studies likewise demonstrated some promise, primarily to platinum-based chemotherapy regimens, although these were typically in small series of young patients seeking to avoid irradiation [26,27]. One of the more promising was the Pediatric Oncology Group (POG) 9233/34 study in infants with ependymoma (n=82), demonstrating a 2-year improved event free survival (EFS) with chemotherapy, 42.1% vs 19.6%; [28] together, these studies highlighted the possibility of chemosensitivity, especially as a tool to obtain complete response (and subsequent improved EFS), or to delay irradiation in very young children.

Conversely, other studies have failed to demonstrate a significant benefit of chemotherapy. In cases of first recurrence, 138 patients in the German HIT-REZ studies (HIT-REZ 97 and HIT-REZ 2005) failed to demonstrate response to over 40 different chemo- and targeted agents of systemic therapy, including high-dose chemotherapy with autologous stem cell rescue, although some responses were recorded. [29] Similarly, several small case series demonstrated only small numbers of responses in children with relapsed ependymoma, with roughly 10% of patients exhibiting response. [30]

Thus, while evidence of response in some patients exists, data regarding actual progression-free or overall survival benefit remains elusive. This analysis is further complicated by the clusters of late relapses which may fall outside the survival windows for historical study outcomes. While initial reports from the Children's Oncology Group (COG) study ACNS0831, which enrolled 451 eligible patients and randomized to receipt of maintenance chemotherapy, showed that there might be a potential role for chemotherapy in some patients, subsequent analysis is required for definitive conclusions across the entire cohort and within subgroups. [6] These study results, which employed molecular phenotyping in a prospective trial, are eagerly awaited.

In conclusion, use of chemotherapy to obtain complete response prior to irradiation (with the known caveat of potential interval progression) or in the context of a radiation-delaying or sparing strategy in infants seem to have some consensus utility; however, chemotherapy's use as a consolidative strategy is less clear. ACNS0831 results combined with an evolving understanding of molecularly high-risk ependymoma, however, may push some providers to offer either intensive or metronomic chemotherapy, although the data to support their use currently remains limited.

Anaplasia: controversy in ependymoma

Anaplastic histology has variably been supported in past clinical trials as a marker of high-risk disease. This was in large part due to considerable literature describing anaplastic ependymoma as having worse clinical outcomes, leading to the incorporation of anaplasia as a high-risk stratification marker [31–34] in clinical care as well as in prospective trial design. However, with the advent of molecular phenotyping and the separation of ependymoma into compartment- and biologically-relevant groups, the utility of anaplasia as an independent risk factor has been debated.

One motive for continued review was concern about reproducibility. In a retrospective study, five separate experienced neuropathologists reviewed the pathology of 229 children with ependymoma enrolled on four separate European trials (SIOP CNS9204, CNS9904; Societe Francaise d'Oncologie Pediatrique (SFOP), Associazione Italiana Ematologia Oncologia Pediatrica (AIEOP)). [35] This study showed significant variability in calling grade 2 versus grade 3 amongst the expert neuropathologists, reinforcing concerns about the reliability of a histopathological characteristic which may not be reliably reproduced. Subsequent recommendations by several neuropathology teams leaned heavily towards location- and molecularly-based assessment.

The 2021 World Health Organization has abandoned the usage of 'anaplasia' to define ependymoma, instead relying on location-based molecular phenotyping with allowance for grading, but specifically noting that the impact of anaplasia on prognosis was controversial when considered in the context of molecular information (n.b., the newest version of the WHO criteria also removed histologically-described ependymoma from its lexicon: papillary, tanycytic, and clear cell) [3,11].

Currently, while anaplastic ependymoma may represent a molecularly aggressive phenotype, in the context of complete molecular phenotyping, there is no clear data that it represents an additional independent risk factor. Thus, clinical practice has begun to shift away from responding to anaplasia to drive clinical decisions, and future therapeutic trials are likely to hinge more directly on molecular biology.

Ultra-high-risk Ependymoma: time to consider early phase trials at diagnosis?

Of all the genetic alterations frequently seen in pediatric posterior fossa ependymoma, the gain of chromosome 1q and loss of chromosome 6q are by far the most clinically impactful. Multiple series have reported 1q gain to be the most common structural aberration in pediatric ependymoma (17–20% of all cases), followed by 6q loss (8–9%) [11,26,27]. Unfortunately, these rather common alterations also confer a poorer prognosis, which is further compounded when both occur within the same tumor. In a cohort of 663 posterior fossa ependymoma type A (PF-EPN-A) tumors, the 5-year progression-free survival (PFS) was strikingly worse tumors harboring 1q gain or 6q loss, with a 5-year PFS of 50% (95% CI 45%–55%) for balanced tumors, compared with 32% (95% CI 24%–44%) for 1q gain only, 7.3% (95% CI 2.0%–27%) for 6q loss only and 0% for both 1q gain and 6q loss [27]. Tumors demonstrating 1q gain/6q loss were found also to progress rapidly with a median PFS of 0.75 years compared to 2.42 years for those with 1q gain who were 6q balanced. Recurrence of tumors with 1q gain/6q loss is often metastatic (38.5% local, 53.8% distant only, 7.7% combined, $P = .03$).

The aggressive behavior of this molecularly-distinct subgroup has earned it the unofficial label of "ultra-high-risk" ependymoma. These tumors account for approximately 10% of all PF-EPN-A tumors, making them a substantial contributor to the mortality seen in this tumor group [27]. The rapid progression and extremely high rate of relapse in tumors with 1q gain/6q loss has prompted discussions about initiating a radically new upfront treatment approach for this group, as current treatment standards are clearly ineffective.

The ethics of enrolling on early phase clinical trials has revolved around patients who otherwise do not have a reasonable expectation of cure or prolonged disease stabilization, engaging with the ethical concept of 'beneficence.' These situations are typically clearest in the context of relapsed childhood brain cancer, or in rarer situations where survival is poor, even at diagnosis. The subgroup of tumors described here replicate the significant mortality of some of the worse pediatric brain cancers, such as diffuse midline glioma, TP53-mutant sonic hedgehog medulloblastoma, and MYC-amplified Group 3 medulloblastoma. Thus, akin to those scenarios, prioritization of children with ultra-high-risk ependymoma may be considered for participation in Phase I studies at diagnosis. Furthermore, the dramatic prognostic implications of these (and other) molecular findings highlight the importance of genetic profiling of ependymoma at diagnosis.

Radiation: required for everyone?

Radiation has clearly been demonstrated to be an effective modality of treatment in ependymoma, with most studies demonstrating prolonged EFS with adjuvant radiotherapy. However, irradiation also may cause significant long-term morbidity, including secondary malignancy, vasculopathy, and cognitive impairment [36]. With the advent of molecular subgroups and refined risk stratification based on patient outcomes, some patients have excellent outcomes and may be considered for radiation-sparing treatment strategies.

In supratentorial ependymoma, small case series also provided rationale for potential radiation sparing in gross totally resected, non-anaplastic disease. Initially reported in 1998, ten patients with supratentorial ependymoma were observed after GTR, with only three patients experiencing relapse, and two of those being salvaged with repeat resection and focal irradiation [37]. The possibility of supratentorial ependymoma surviving without radiotherapy was seemingly confirmed in a SEER database study that included children and adults with supratentorial EP after GTR, half of whom received radiotherapy, with no statistically significant outcome differences [38]. Later, ACNS0121 utilized observation for supratentorial ependymoma after a GTR, accruing eleven patients. In this study, 5-year EFS was 61.4%, similar to the general population, with over half of the patients with durable response and only one patient experiencing distant failure; the group exhibited a -year OS of 100% [2]. Conversely, a study of Canadian centers found that of 14 ST ependymoma that received GTR alone, 57% experienced relapse, most occurring within 1 year of diagnosis, although the pattern of relapse mimicked that of other studies, with only one patient exhibiting distant relapse not salvageable by re-resection and focal irradiation [39]. Further, follow-up for these studies was relatively brief given the possibility of delayed progression for ependymoma, lending some equipoise to the concept of surgery-only treatment in GTR ST ependymoma. However, it is worthwhile to note that molecularly analyzed cases hypothesize that ST-EPN-YAP1 in particular, may be amenable to this strategy.

Similarly, in patients with posterior fossa group B tumors who receive a total resection, survival without irradiation may be feasible. This data is reported in both relatively small case series, such as a report with 9 patients (no failures), [36] as well as a large combined series collating the Global Ependymoma Network of Excellence (GENE), St Jude RT1, and Collaborative Ependymoma Research Network (CERN) cases and demonstrating that, in the context of a GTR, patients with non-irradiated PFB tumors harbored a 10 year OS of 82.3%, although the PFS was only 45.1% [40]. Thus, while irradiation did increase 10-year PFS to 74%, this data suggested that at least some children with completely-resected PFB ependymoma were curable with surgery alone. Subsequent analysis further refined PFB ependymoma, postulating that 13-q balanced PFB had an even better survival, supporting the intriguing hypothesis of a surgery-only initial approach [1,41].

Coupled with clear and significant declines in full scale IQ from even focal upfront conformal radiotherapy, consideration for radiation-sparing trials in selected cases of ependymoma seem reasonable.

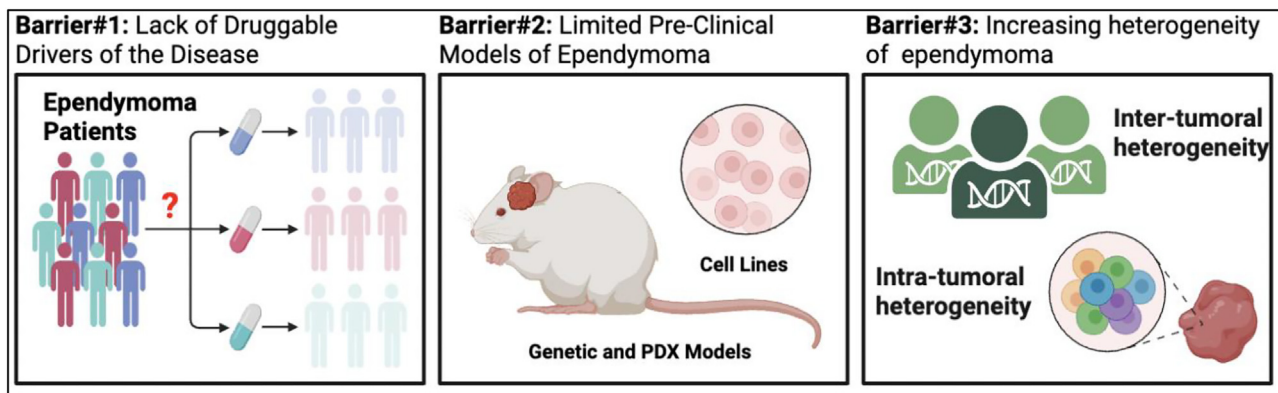


Fig 1. Major obstacles to rapid translation of novel biologic findings to effective clinical interventions.

However, due to the relatively small overall numbers and some conflicting data, investigators at an ependymoma consensus meeting concluded that radiation-sparing strategies should be tested in clinical trials in these relatively low-risk groups in order to reduce the rate of treatment-related morbidity without sacrificing survival [42].

The advent of advanced radiation delivery, such as with proton beam irradiation, must also be considered when weighing the benefits and consequences of primary or repeat irradiation. Thus far, proton-based irradiation does not seem to compromise efficacy, [43] and may improve negative cognitive and other late effects. Other questions regarding optimal radiation treatment remain, such as whether craniospinal radiation is necessary given some reports of distant relapse [43,44], whether radiation doses may be mitigated in some children to $<59\text{Gy}^{2,43}$ and other compelling questions outside the scope of this article.

Regardless, the significant survival differences discussed in these brief vignettes, namely, extremely poor prognoses in patients with $1q+/6q-$, contrasted against the extremely good prognosis of ST-EPN-YAP1 and PFB ependymoma, highlight one agreed upon conclusion: upfront molecular phenotyping is essential for both clinical care and design in future clinical trials.

Challenges in therapeutic translation

The explosion of knowledge regarding ependymoma subtypes and biology has led to significant revisions in clinical perspective. However, translation of this data into improved outcomes for children with ependymoma has not yet materialized.

Several major challenges have hampered development of clinical trials and treatments for ependymoma (Fig. 1). Few genetic and patient-derived xenograft models of disease have limited pre-clinical studies and efforts to delineate the molecular basis of ependymoma. A lack of druggable-driver alterations has prevented application of precision medicine-based approaches [45]. Finally, despite few genetic alterations, the transcriptional heterogeneity of ependymoma is recognized as increasingly diverse both between patients and within a given tumor sample [20,41,42,46-48]. Recent advancements in animal modeling epigenomics, and single cell genomics have attempted to address some of these hurdles, providing new therapeutic targets and an improved understanding of the disease [4,5,7,49-53]. We will review recent advances in ependymoma biology according to the following key barriers and discuss opportunities for future advancement and discovery.

Barrier 1: lack of druggable drivers of the disease

Like most pediatric cancers, ependymomas are characterized by relative 'quiet' genomes [45,52]. In the case of supratentorial ependymoma, *ZFTA* or *YAP1* associated gene fusions are sufficient to initiate tumors, with co-occurring *CDKN2A* deletions occurring in about 20-30% of cases

[40]. There are currently no small molecules capable of inhibiting or degrading *ZFTA* gene fusion proteins, likely due to their highly disordered protein structure (commonly observed in transcription factors). However, the presence of at least 1-3 C2H2 zinc finger binding domains in *ZFTA* may provide a potential opportunity for molecular glues to be utilized in a protein degradation type strategy [52]. Before drug discovery approaches are initiated, however, a key unanswered question is whether *ZFTA*- and *YAP1*- fusions are still required for tumor progression or rather these gene fusions serve as initiating events. Alternate (although indirect) targeting strategies may be effective such as probing the dependency on transcriptional co-regulators critical for maintaining the neoplastic transcriptional state [7,50]. These include inhibitors of chromatin regulators such as BET bromodomain proteins, EP300/CBP lysine acetyl-transferases, and transcriptional elongation factors [54-56]. A targetable strategy may exist for *YAP1* fusion driven ependymoma, with the advancement of TEAD inhibitors and degraders. Such approaches could be readily tested in novel genetic models of *YAP1* fusion ependymoma (see next section) [57].

In the case of PF-A ependymoma, over 90% of cases lack a detectable recurrently mutated gene [45]. Despite *EZH2* interacting protein (*EZH1P*) over-expression in nearly all PF-A tumors, *EZH1P* is mutated in less than 10% of cases, and the functional contribution of those mutations still unclear [48,58]. Similar to *ZFTA/YAP1* fusions the continued dependence on *EZH1P* is an outstanding question. Expression of *EZH1P* phenocopies the effects of *H3K27M* mutation seen in diffuse midline glioma (DMG), specifically the global loss of the histone repressive mark, *H3K27me3* [58-61]. *EZH1P* over-expression and the subsequent hypomethylation on *H3K27* interestingly is only found in PF-A ependymomas, and in stark contrast to the more differentiated and clinically less aggressive PF-B ependymomas. Tied to wide-spread epigenomic alterations in PFA ependymoma are significant alterations to metabolic pathways and a dependency on several nodes such as methionine pathway, glucose, and glutamine regulation [9,55]. These nutrient pathways may represent important vulnerabilities and sources for drug discovery against the most clinically aggressive forms of PFA ependymoma. Furthermore, PFA ependymomas have been shown to grow preferentially in "hypoxic" environments, highlighting a sensitivity to changes in oxygen tension. These distinct epigenomic and metabolic programs observed in PFA ependymomas have revealed new opportunities for treatment including agents that modulate glucose regulation (Metformin [9]), *H3K27me3* modifications (*EZH2* inhibitors [45]), *H3* acetylation (HDAC inhibitors such as valproic acid in the SIOP Ependymoma Program II) and global methylation levels (5-Azacytidine, [62] and *MAT2A* inhibitors [55]). Drugs potentially closest to translation include Metformin, which has been shown to extend PFA ependymoma survival in a mouse model, through restoration of *H3K27me3* levels. Furthermore, *EZH2* inhibitors are in clinical trial evaluation for other brain tumors including atypical teratoid rhabdoid tumors. However, expansion

Table 1
Current molecularly-targeted strategies in development for ependymoma.

Ependymoma Driver Gene	Cellular Dependency	Druggable?	Candidate Therapeutic Approaches
ZFTA Fusion Variants	Not Determined	Not Yet, Disordered Protein Structure	<ul style="list-style-type: none"> Inhibition of ZFTA associated transcriptional co-activator proteins, such as BRD4 and EP300/CBP. Targets elevated in ZFTA fusion driven tumors. (i.e. FGFR), or observed across all EPN tumors (i.e. ERBB2 and B7H3)
YAP1 Fusion Variants	Not Determined	Possibly	<ul style="list-style-type: none"> YAP pathway inhibitors including Inhibition of TEAD family of transcription factors
EZH1P	Not Determined	Not Yet, Disordered Protein Structure	<ul style="list-style-type: none"> EZH1P associated epigenetic and metabolic programs (i.e. PRC2 inhibition, hypoxia, glucose, glutamine, methionine pathway modulation).
MYCN	Not Determined	Not Yet, Disordered Protein Structure	<ul style="list-style-type: none"> Inhibition of MYCN associated transcriptional co-activator proteins, such as BRD4 and EP300/CBP.
ACVR1	Not Determined	Yes	<ul style="list-style-type: none"> ACVR1 inhibitors

of pre-clinical studies is needed for these promising drugs beyond single cell lines and patient-derived xenograft (PDX) models to establish broad applicability of these findings. Finally, investigating key downstream transcriptional targets of ependymoma by mining the chromatin landscape and cellular architecture of these tumors has yielded new targets and insights about drivers of the disease [48]. Active chromatin mapping has revealed essential transcriptional circuits critical for ependymoma tumor initiation, as well as novel dependency genes that are amenable to small molecule inhibition such as FGFR inhibitors [63]. While traditional application of precision medicine approaches that pairs gene mutation with a specific drug is likely to be ineffective against ependymoma, recognition of the distinct epi-metabolic networks of these unique tumors is an active area of therapeutic discovery. A small number of PF ependymomas cluster separately from PFA and PFB tumors on methylation, and typically have ACVR1 mutations, while lacking EZHIP expression [64]. While targeted therapies have yet to achieve significant clinical effect, these varied approaches remain in development and are summarized in Table 1.

Despite the challenges of druggable driver identification, other areas of therapeutic innovation that rely less on cellular signaling may also be helpful, such as novel immunotherapeutic approaches. Ependymomas that display prolonged stability have been shown to have up-regulation of immune-related genes, increased T cell infiltration [65], and increased PDL1 expression. [66] For instance, expression of HER2 has been described in a large number of pediatric ependymomas with only limited expression on normal brain cells. Identification of these cellular targets may provide a vulnerability towards adoptive cellular therapy, such as chimeric antigen receptor (CAR) T cells, and has been shown preclinically to be potentially useful [67,68], and has generated several currently active clinical trials. Similarly, oncolytic virotherapy [69], vaccines [70], and immune checkpoint blockade [71] are currently being developed for ependymoma.

Barrier 2: limited pre-clinical models of ependymoma

Compared to other pediatric brain tumors such as medulloblastoma and diffuse midline glioma (DMG), genetic models of ependymoma have only recently been developed (Fig. 2). ZFTA (formerly C11orf95) gene fusions constitute the majority of supratentorial ependymomas and are sufficient to drive tumorigenesis when expressed in mouse neural stem cells through viral transduction, in utero electroporation (IUE), or the RCAS-TV system [7,51–53]. In similar vein, YAP1 gene fusions have also been modeled using similar approaches and are also sufficient to initiate the disease [57]. Advancements of ZFTA driven genetic models have revealed new insight into the role of ZFTA fusion proteins and their

Genetic Models of Fusion Ependymoma	Human PDX/Cell Models of ZFTA-Fusion Ependymoma	Human PDX/Cell Models of PF-A Ependymoma
<ol style="list-style-type: none"> IUE RCAS-TV Viral Transduction 	<ol style="list-style-type: none"> EP1-NS 1425 SJEPST-16-06903 EPD613FH MDT-ST2 MDT-ST4 VBT73 VBT145 VBT211 VBT242 VBT371 BT165 	<ol style="list-style-type: none"> SJEPPF-16-02472 SJEPPF-16-09238 SJEPPF-15-8710 SJEPPF-16-08404 EPN811 EPN928 EPD210FH EPD710FH MDT-PFA-2 MDT-PFA-4 MDT-PFA-5 MDT-PFA-9 VBT78 VBT131 VBT96 VBT160 BT214

Fig. 2. A summary of available preclinical murine models of childhood ependymoma.

importance in regulating oncogenic gene expression [7,50–53]. These new animal models have also provided a framework for investigating the ependymoma microenvironment and contribution of distinct tumor cell populations, immune cells, and normal neural cells that may contribute to disease pathogenesis. *In vivo* modeling of ZFTA fusion driven ependymomas has also incorporated gene editing based approaches to study genetic dependencies, such as the role of master (i.e., core regulatory circuit) transcription factors and their key roles in ependymoma initiation [48]. A key example includes SOX9, which was found to be one of the most active transcription factors (TFs) in ependymoma and essential for ependymoma cell proliferation and tumor initiation [72].

The successes in genetic modeling of supratentorial ependymoma have not been seen in the case of posterior fossa ependymoma. Efforts to this date have failed to generate EZHIP-driven mouse models, possibly due to several factors: i) Lack of knowledge of the cellular compartment of origin (and its existence in mice), and ii) Lack of additional genetic alterations that may participate with EZHIP unlike the TP53, ATRX, and PDGFRA alterations seen in DMG. These challenges may be overcome with advances in murine and human hindbrain-cerebellar organoids that may enable EZHIP driven transformation of cells within the correct cellular compartment. Furthermore, CRISPR-CAS9 based functional genomics screens may facilitate identification of co-operating events with

Table 2

A brief overview of pediatric ependymoma classes and major drivers.

Posterior Fossa Ependymoma			Supratentorial Ependymoma			Spinal Ependymoma			
PF-EPN-A (EZHIP+)	PF-EPN-B	SE	ZFTA Fusion	YAP1 Fusion	SE	MYCN	SP-EPN	MPE	SE
Poor	Favorable		Varied	Favorable	Favorable	Poor	Favorable		
Balanced Genome Chr 1q gain and Chr6q loss poor prognostic factors	Chromosomal Instability	Balanced Genome	Chromosome 11 Chromothripsis CDKN2A loss a poor prognostic factor	Balanced Genome	Balanced Genome	MYCN Amplification	NF2 Mutations	Chromosomal Instability	6q deletion

Ependymoma (EPN), Sub-Ependymoma (SE), Myxopapillary Ependymoma (MPE)

EZHIP over-expression. Encompassing these proposed future studies is the finding that PFA ependymomas grow preferentially in hypoxic environments and exhibit dramatically altered cellular metabolism [55]. Mouse modeling of PFA ependymoma may depend on multiple factors coming together including oncogene expression in appropriate cell types, consideration of additional co-operating driver genes, and growth within specific nutrient availability and oxygen tension. Immunocompetent model development, essential for comprehensive development of immune-based treatment strategies, has likewise been challenging without identification of clear transformative genetic aberrations that can be translated to murine models with normal immune systems.

In the past, pre-clinical efforts to identify effective therapies for ependymoma have been limited by the lack of patient-derived xenograft models [42]. In recent years, multiple groups have established several *ZFTA* fusion driven models, which have enabled drug- and CRISPR-based genetic screening [73–76]. While fewer models for PF-A ependymoma have been established, continued efforts have led to a growing number of representative models, particularly of the most aggressive forms of the disease, such as tumors harboring 1q gain and 6q loss [55,76]. One would anticipate with continued scientific community efforts to establish ependymoma PDX models, that additional PF-A models will be developed, but also including development of rare variants such as PF-B or YAP1 altered ependymoma. A major challenge is the length of time ependymoma mouse models take to grow *in vivo*, with some cell lines requiring 8 months to >1 year to develop brain tumors. This hurdle has made pre-clinical drug efficacy studies particularly challenging to perform in ependymoma, combined with the high variability in tumor formation within a given model.

Barrier 3: increasing heterogeneity of the disease that constitutes ependymoma

While our review focuses on the major subgroups of ependymoma, additional subtypes of ependymoma have been discovered such as *ZFTA* (non-*RELA*) gene fusions, and heterogeneity within PF-A, PF-B, and spinal tumors as defined by DNA methylation-based classification (Table 2) [40,77]. These subgroups/subtypes are directly tied to distinct clinical outcomes and will shape future clinical trial design for ependymoma [2,46,78,79]. Complicating the observed inter-tumoral (between patients) heterogeneity is a recognition of the diverse cellular landscape of ependymoma, composed of distinct tumor populations, microglia, infiltrating immune cells, and neurons [4,5,80]. The ependymoma microenvironment and functional interactions between cell types is poorly defined but may hold opportunities for identifying effective therapies and understanding the mechanisms of treatment resistance and relapse. Early lessons from single-cell genomic characterization of ependymoma has revealed distinct cell populations responsive to disparate receptor tyrosine kinase (RTK) inhibitors such as those that block the FGFR and IGF1R pathways [5]. These data underscore the importance of tailored combinatorial strategies against different cellular populations within a given patient tumor. Furthermore, transcriptional trajectories that reflect ependymoma cell state and differentiation capacity may hold clues

to understanding the mechanistic basis of ependymoma progression [4,81]. Understanding the functional consequences of these transitions, such as epithelial-to-mesenchymal and hypoxic programs, will be important to uncovering therapeutic vulnerabilities. Other stem-cell properties, like telomerase re-activation, might also be at play in ependymoma stem cells, and might represent not only a biomarker of the disease, but also a future therapeutic target [82,83]. These studies have also provided critical insight on the developmental basis of ependymoma, which are likely to improve our understanding to the processes of tumor initiation and lead to improved mouse modeling of the disease.

Conclusion

The relatively recent advent of molecular studies has enabled paradigm-shifting assessment of childhood ependymoma, dividing a historically single-disease into many separate subtypes. This enhancement has engendered optimism for the development of more effective treatment for what is now widely understood to be a tumor with a poor overall prognosis and has highlighted the need for comprehensive molecular phenotyping in every child at diagnosis and, potentially, recurrence. While both scientific and clinical barriers to rapid improvement in outcomes exist, identification of these obstacles allows for appropriate resource investment and consensus building to propel the field towards rapid and rational improvements in survival and morbidity.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

CRediT authorship contribution statement

Eugene I. Hwang: Conceptualization, Data curation, Writing – original draft, Writing – review & editing. **Derek Hanson:** Conceptualization, Data curation, Writing – original draft, Writing – review & editing. **Mariella G. Filbin:** Conceptualization, Data curation, Writing – original draft, Writing – review & editing. **Stephen C. Mack:** Conceptualization, Data curation, Writing – original draft, Writing – review & editing.

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