

Review **Role of the Gut Microbiome in the Development of Atherosclerotic Cardiovascular Disease**

Ahmad Al Samarraie ¹ , Maxime Pichette ² and Guy Rousseau 3,*

- 1 Internal Medicine Department, Faculty of Medicine, University of Montreal, Montréal, QC H3T 1J4, Canada
- ² Cardiology Department, Faculty of Medicine, University of Montreal, Montréal, QC H3T 1J4, Canada
³ Cantre de Pierrédesine, CU ISSS NJM/Hêrital du Sarré Cenur Montréal, QC H4J 1C5 Canada
- ³ Centre de Biomédecine, CIUSSS-NÎM/Hôpital du Sacré-Cœur, Montréal, QC H4J 1C5, Canada

***** Correspondence: guy.rousseau@umontreal.ca

Abstract: Atherosclerotic cardiovascular disease (ASCVD) is the primary cause of death globally, with nine million deaths directly attributable to ischemic heart diseases in 2020. Since the last few decades, great effort has been put toward primary and secondary prevention strategies through identification and treatment of major cardiovascular risk factors, including hypertension, diabetes, dyslipidemia, smoking, and a sedentary lifestyle. Once labelled "the forgotten organ", the gut microbiota has recently been rediscovered and has been found to play key functions in the incidence of ASCVD both directly by contributing to the development of atherosclerosis and indirectly by playing a part in the occurrence of fundamental cardiovascular risk factors. Essential gut metabolites, such as trimethylamine N-oxide (TMAO), secondary bile acids, lipopolysaccharides (LPS), and short-chain fatty acids (SCFAs), have been associated with the extent of ischemic heart diseases. This paper reviews the latest data on the impact of the gut microbiome in the incidence of ASCVD.

Keywords: gut microbiota; gut microbiome; atherosclerotic cardiovascular disease; atherosclerosis; risk factors; hypertension; dyslipidemia; diabetes; trimethylamine N-oxide; secondary bile acids; lipopolysaccharides; short-chain fatty acids

Citation: Al Samarraie, A.; Pichette, M.; Rousseau, G. Role of the Gut Microbiome in the Development of Atherosclerotic Cardiovascular Disease. *Int. J. Mol. Sci.* **2023**, *24*, 5420. [https://doi.org/10.3390/](https://doi.org/10.3390/ijms24065420) [ijms24065420](https://doi.org/10.3390/ijms24065420)

Academic Editors: Dulcenombre Gómez Garre and Javier Modrego

Received: 12 February 2023 Revised: 6 March 2023 Accepted: 9 March 2023 Published: 12 March 2023

Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

1. Introduction

Despite major advances in prevention and treatment strategies, atherosclerotic cardiovascular disease (ASCVD) remains the leading cause of morbidity and mortality all around the world [\[1\]](#page-9-0). This alarming statistic brought to light the complex etiology of atherosclerosis, which has been recognized as not being solely induced by conventional risk factors, such as hypertension, diabetes, dyslipidemia, male sex, and smoking. In 2000, Haraszthy et al. proposed for the first time that the gut microbiota was associated with the occurrence of ASCVD, after finding DNA from multiple bacteria species in a plaque of cholesterol [\[2\]](#page-9-1).

Trillions of micro-organisms weighing about 1.5 kg inhabit the gut, carrying out key functions that the rest of the human body is incapable of performing [\[3\]](#page-9-2). These microorganisms have combined genomes (the microbiome) that exceed the human genome by many times [\[4–](#page-9-3)[6\]](#page-9-4). The gut microbiota is dominated by anaerobic bacteria, with *Firmicutes* (Gram-positive) and *Bacteroidetes* (Gram-negative) composing more than 90% of intestinal bacterial species [\[7\]](#page-9-5).

This paper reviews the latest available information on the role of the gut microbiome in the incidence and progression of ASCVD.

2. Metabolic Pathways

Major metabolites have been identified and linked to the development of cardiovascular diseases, including but not limited to trimethylamine N-oxide (TMAO), secondary

bile acids, lipopolysaccharides (LPS), short-chain fatty acids (SCFAs), and phenylacetyl-bile acids, lipopolysaccharides (LPS), short-chain fatty acids (SCFAs), and phenylacetylglutamine (PAGln). Figure [1](#page-1-0) illustrates major gut metabolic pathways leading to ASCVD. glutamine (PAGln). Figure 1 illustrates major gut metabolic pathways leading to ASCVD.

Major metabolites have been identified and linked to the development of cardiovas-

Figure 1. Major metabolic pathways involving the gut microbiome and leading to the development **Figure 1.** Major metabolic pathways involving the gut microbiome and leading to the development and progression of atherosclerotic cardiovascular disease. FXR: farnesoid X receptor, Gln: glutamine, mine, IL-1: interleukin-1, IL-6: interleukin-6, IL-8: interleukin-8, LPS: lipopolysaccharides, NF-κB: IL-1: interleukin-1, IL-6: interleukin-6, IL-8: interleukin-8, LPS: lipopolysaccharides, NF-κB: nuclear factor-κB, SCFAs: short-chain fatty acids, TGR5: takeda G protein-coupled receptor 5, TLR: toll-like receptor, TMA: trimethylamine, TMAO: trimethylamine N-oxide, TNF-α: tumor necrosis factor-α. factor-α.

correlation between these two entities was first reported by Wang et al. in 2011 [\[8\]](#page-9-6). TMAO goes through several enzyme modifications before being transformed into its final active form. In fact, the first step requires the intestinal conversion of one of three metabolites found in the food, namely L-carnitine, choline, and betaine, into TMA by the enzyme TMA lyase, which is derived fro[m](#page-9-7) the *Firmicutes* species found in the gut microbiota [9]. These three nutrients are naturally found in foods, such as eggs, red meat, and fish. After their conversion into TMA, this amine is absorbed into the bloodstream before being transported to the liver, where it is transformed into TMAO by the enzyme flavin-dependent mono oxygenase 3 (FMO₃) [10]. In physiologic states, the kidney excretes in the urine close to 95% of TMA oxidized into TMAO [11]. Hence, changes to any component along this metabolic pathway, from food ingestion to hepatorenal function together with the liver FMO₃ activity, could lead to increased levels of TMAO with associated complications, such as ASCVD [12]. First, TMAO directly contributes to the pathogenesis and extent of ASCVD. The

Primary bile acids are synthesized in the liver from cholesterol and are conjugated with glycine, subsequently leading to the formation of cholic acid and chenodeoxycholic acid. Then, primary bile acids are transported in the gut, in which the microbiota contributes to their deconjugation to form secondary bile acids in the distal ileum [\[13\]](#page-9-11). Secondary bile acids permit absorption of lipid nutrients and fat-soluble vitamins [\[14\]](#page-9-12). They are also involved in the activation of two key receptors, namely farnesoid X receptor (FXR) and takeda G protein-coupled receptor 5 (TGR5). These receptors modulate glucose and cholesterol metabolism. In fact, TGR5 leads to an increased secretion of glucagon-like peptide 1 (GLP-1), which contributes to an improved glucose tolerance $[15,16]$ $[15,16]$. TGR5 is also thought to possess anti-inflammatory properties by inhibiting nuclear factor-κB (NF-κB), thus decreasing the production of pro-inflammatory cytokines [\[17\]](#page-9-15). In another trial, simultaneous inhibition of both FXR and TGR5 exacerbated atherosclerotic formation,

thereby highlighting their benefits in disease control [\[18\]](#page-9-16). Their anti-inflammatory and anti-atherogenic properties arise from suppression of tumor necrosis factor- α (TNF- α) and NF-κB signaling pathways in addition to a decreased secretion of pro-inflammatory cytokines [\[19\]](#page-9-17). Therefore, secondary bile acids activate two key receptors involved in the inhibition of major atherosclerotic pathways.

Similar to TMAO, LPS are endotoxins found on the outer membrane of Gram-negative bacteria and are involved in the pathogenesis of ASCVD. LPS are recognized by the innate immune system by toll-like receptor 4 (TLR4), which is a subtype of pattern recognition receptors (PRR) [\[20\]](#page-9-18). Upon recognition of LPS, TLR4 induces a pro-inflammatory state with an increased production and secretion of cytokines and chemokines [\[21\]](#page-9-19). LPS are also identified by other receptors, such as LPS-binding protein (LBP), myeloid differentiation protein 2 (MD-2), and cluster of differentiation 14 (CD14) [\[22\]](#page-9-20). These receptors, which are mainly expressed on macrophages, activate and enhance several protein kinases, such as IL-1 receptor-associated kinase (IRAK-1) and myeloid differentiation factor 88 (MyD88). NF-κB is subsequently activated, which, along with LPS, stimulate numerous pro-atherosclerotic inflammatory pathways [\[23](#page-9-21)[–25\]](#page-9-22). In fact, LPS induce endothelial dysfunction, increase oxidative stress through production of reactive oxygen species (ROS), and produce several pro-inflammatory cytokines, such as $TNF-\alpha$, interleukin-1 (IL-1), interleukin-6 (IL-6), and interleukin-8 (IL-8) [\[26](#page-9-23)[–28\]](#page-9-24). Therefore, LPS contribute to ASCVD by promoting inflammation through various pathways.

In contrast to TMAO and LPS, SCFAs are protective against the occurrence of atherosclerosis and are the result of the ingestion and digestion of complex carbohydrates by numerous gut bacteria, including *Anaerostipes butyraticus*, *Faecalibacterium prausnitzii*, and *Roseburia intestinalis* [\[29,](#page-10-0)[30\]](#page-10-1). The most frequent SCFAs produced are acetate, butyrate, and propionate [\[31\]](#page-10-2). These nutrients serve many roles, but their primary function is to modulate the host immune system through increased production of regulatory T cells and suppression of histone deacetylases (HDACs) [\[32](#page-10-3)[,33\]](#page-10-4). By inhibiting HDACs, SCFAs inhibit inflammatory pathways owing to a decrease in NF-κB activation together with a reduced production of pro-inflammatory cytokines [\[34\]](#page-10-5). Other functions attributed to SCFAs include enhanced intestinal barrier stability and protection against pathogen invasion [\[32,](#page-10-3)[33\]](#page-10-4). Thus, SCFAs protect against atherosclerosis by modulating inflammatory pathways.

Phenylacetylglutamine (PAGln) is a metabolite that was recently discovered and was shown to be positively associated with the development of cardiovascular diseases [\[35\]](#page-10-6). PAGln is derived from a simple amino acid, phenylalanine, that undergoes a series of alterations before arriving to its active metabolite. In fact, Nemet et al. have demonstrated that the microbial *porA* gene permits the transformation of phenylalanine into phenylacetic acid, with subsequent hepatic metabolization of phenylacetic acid into PAGln [\[36\]](#page-10-7). PAGln was first reported to be positively correlated with ASCVD and overall mortality in patients suffering from chronic kidney disease (CKD) [\[37\]](#page-10-8). Several pathophysiologic mechanisms were hypothesized to explain this association. As a matter of fact, PAGln was shown to increase platelets' activation and responsiveness, resulting in increased thrombosis potential leading to ASCVD. [\[38\]](#page-10-9) PAGln also transmits cellular events via G-protein coupled receptors, specifically the α2A, α2B, and $β2$ adrenergic receptors [\[36\]](#page-10-7). Interestingly, carvedilol, a commonly used β-blocker in clinical practice, was shown to inhibit these prothrombotic effects [\[36\]](#page-10-7). Thus, PAGln is involved in the occurrence of ASCVD through an accelerated rate of thrombus generation and vessel occlusion, potentially giving rise to acute myocardial infarction.

3. Gut Microbiome and Hypertension

Arterial hypertension is a well-recognized and major risk factor for the development of ASCVD [\[39](#page-10-10)[–41\]](#page-10-11). Leading medical communities of cardiology recommend blood pressure control both pharmacologically and non-pharmacologically as part of cardiovascular diseases' primary and secondary preventions [\[42–](#page-10-12)[44\]](#page-10-13). Even though the exact cause of essential hypertension remains unclear, many risk factors are thought to contribute to its

development, including advanced age, positive family history, obesity, a high-sodium diet, and a sedentary lifestyle [\[42](#page-10-12)[–44\]](#page-10-13).

Recently, the gut microbiota has been found to play a role in the development of hypertension [\[45](#page-10-14)[–54\]](#page-11-0). Li et al. have demonstrated that high blood pressure was transferrable through fecal transplantation from hypertensive subjects to germ-free mice, thus confirming the implication of the intestinal microbiome [\[47\]](#page-10-15). Additionally, compared to healthy controls, pre-hypertensive and hypertensive individuals have decreased microbial richness and variety with an overgrowth of specific bacteria, namely *Prevotella* and *Klebsiella* [\[47\]](#page-10-15). A decrease in microbial richness constitutes an alteration in gut microbiome, thus defining dysbiosis. Dysbiosis is thought to induce low-grade inflammation, which in turn can provoke hypertension when the inflammation is persistent [\[55](#page-11-1)[,56\]](#page-11-2). Additionally, a reduction in *Lactobacillus* abundance can induce higher blood pressure values in both mice and humans when compared to healthy controls [\[57\]](#page-11-3).

Yang et al. recently proved that the *Firmicutes* on *Bacteroidetes* ratio was increased in spontaneously hypertensive rats, in angiotensin II-induced hypertensive rats, and in a small group of humans with hypertension [\[51\]](#page-10-16). It is noteworthy to note that by normalizing this ratio with the administration of minocycline, blood pressure of spontaneously and induced-hypertensive rats also normalizes [\[51\]](#page-10-16). Additionally, fasting for a five-day period seems to induce a modification in the gut microbiota, subsequently reducing blood pressure in hypertensive patients [\[58\]](#page-11-4).

The gut microbiota produces various metabolites with different effects on blood pressure regulation [\[59\]](#page-11-5). Beneficial metabolites include SCFAs and vitamins. Acetate, propionate, and butyrate account for 80% of the total SCFAs produced [\[60\]](#page-11-6). SCFAs are thought to be beneficial in blood pressure reduction through mainly their vasorelaxant and anti-inflammatory effects [\[61\]](#page-11-7). Indeed, Bartolomaeus et al. demonstrated that the administration of propionate in mouse models was associated with a better control of high blood pressure together with a decrease in vascular inflammation and cardiac damage [\[62\]](#page-11-8). In another mouse model, acetate was shown to be highly effective in improving cardiac function by reducing left ventricular wall thickness and body weight in addition to a decrease in systemic blood pressure [\[63\]](#page-11-9).

By contrast, TMAO, another metabolite produced by the gut microbiome, is positively associated with hypertension [\[64\]](#page-11-10). TMAO has a proatherogenic and prothrombotic effect [\[65](#page-11-11)[,66\]](#page-11-12). This toxic metabolite is thought to induce hypertension through prolongation of the hypertensive effect of angiotensin II and facilitation of angiotensin II-induced vasoconstriction [\[67](#page-11-13)[,68\]](#page-11-14). TMAO also enhances stiffening of the large arteries, namely the aorta and carotid arteries, which amplifies the risk of ASCVD both directly and indirectly through increased systolic blood pressure [\[69\]](#page-11-15).

Thus, the gut microbiome fabricates different metabolites with various effects on blood pressure. SCFAs improve its control while TMAO is deleterious.

4. Gut Microbiome and Diabetes

In 2019, diabetes was estimated to affect around 463 million people worldwide [\[70\]](#page-11-16). That alarming number is expected to rise to 700 million by 2045 [\[70\]](#page-11-16). Obesity contributes to the development of type 2 diabetes (T2D) through many pathophysiologic mechanisms, mainly insulin resistance [\[71\]](#page-11-17). Diabetes results in microvascular and macrovascular complications, with cardiovascular disease being the most common cause of morbidity and mortality among people suffering from diabetes [\[72\]](#page-11-18). Like hypertension, the exact etiology of diabetes remains unclear but many risk factors have been identified, including a positive family history, advanced age, obesity, hypertension, and a history of cardiovascular disease [\[73,](#page-11-19)[74\]](#page-11-20).

In 2004, Backhed et al. suggested for the first time that the gut microbiota could be linked to the development of T2D by inducing alterations in glucose metabolism [\[75\]](#page-11-21). Many studies have reported that obesity and alterations in glucose metabolism were associated with an altered ratio between the two most common bacteria composing the intestine, with increased *Bacteroidetes* and decreased *Firmicutes* levels [\[76](#page-11-22)[–78\]](#page-12-0). Thereafter, experiments using metagenomic sequencing in human volunteers established that people with T2D have a dysbiotic gut microbiota [\[7,](#page-9-5)[79\]](#page-12-1). Both trials reported that people with diabetes had less butyrate-producing bacteria. Butyrate is one of the three main SCFAs and is thought to possess an advantageous effect on insulin sensitivity and energy balance [\[80\]](#page-12-2). Several studies have followed and have reported that gut microbiome dysbiosis contributes to a less favorable course of T2D by inducing a rapid progression of insulin resistance [\[81–](#page-12-3)[85\]](#page-12-4).

More than 80% of patients with T2D are overweight [\[86\]](#page-12-5). The underlying pathophysiological mechanism linking these two conditions is insulin resistance induced by obesity [\[87\]](#page-12-6). Numerous studies from animal models have demonstrated that the gut microbiota is implicated in the development of obesity [\[75](#page-11-21)[–77](#page-11-23)[,88,](#page-12-7)[89\]](#page-12-8). Several trials have provided an explanation, with one interesting experiment reporting that low bacterial variety in the microbiome is associated with insulin resistance, fatty liver, low-grade inflammation, and obesity when compared to high bacterial diversity [\[90\]](#page-12-9). In another experimentation, scientists isolated the microbiota of obese animals and transplanted it into germ-free animals; obesity developed after 14 days [\[75\]](#page-11-21). Other trials have focused on the potential role of SCFAs and have found that mice suffering from diabetes exhibit lower levels of butyrate-producing bacteria, such as *Fecalibacterium prausnitzii*, *Eubacterium rectale*, and *Roseburia intestinalis*, when compared to healthy controls [\[91](#page-12-10)[,92\]](#page-12-11). Thus, SCFAs, particularly butyrate, are beneficial metabolites that seem to protect against the incidence of diabetes.

In addition, LPS have been found to be early triggers of obesity by inducing an inflammatory state through secretion of cytokines and chemokines [\[93\]](#page-12-12). In healthy individuals, the ingestion of a high-fat meal leads to a transitory increase in plasma LPS levels while in patients suffering from obesity and insulin resistance, LPS levels were found to be chronically elevated, thus contributing to the development of T2D [\[94](#page-12-13)[,95\]](#page-12-14).

Furthermore, recent animal studies have suggested that elevated levels of circulating TMAO are associated with an increased risk of developing T2D, mainly through impaired glucose tolerance, insulin resistance, and oxidative stress [\[96,](#page-12-15)[97\]](#page-12-16). Chronic high levels of TMAO are also linked with an increased risk of obesity via secretion of inflammatory cytokines, thus contributing to the occurrence of T2D [\[98\]](#page-12-17). A recently published metaanalysis confirmed previous findings and suggested a positive association between T2D and TMAO levels in a dose-dependent manner [\[99\]](#page-12-18).

Gut microbiota is strongly linked with both microvascular and macrovascular diabetic complications [\[100](#page-12-19)[–103\]](#page-13-0). Indeed, patients with end-stage diabetic nephropathy were found to have an abundance of *Haemophilus* and *Lachnospiraceae* bacteria when compared to earlier stages [\[104\]](#page-13-1). Furthermore, *Pasteurellaceae* bacteria are significantly lower in patients with diabetic retinopathy as compared to patients without this complication [\[105\]](#page-13-2). Plasma TMAO levels are also significantly increased in individuals with diabetic retinopathy, and its levels are associated with the incidence of this microvascular complication [\[106,](#page-13-3)[107\]](#page-13-4). These data highlight the importance of the intestinal microbiome and suggest that dysbiosis could play an important role in the development of diabetic complications.

5. Gut Microbiome and Dyslipidemia

Dyslipidemia is one of the major risk factors for both the occurrence and progression of cardiac diseases [\[108\]](#page-13-5). In recent decades, primary and secondary prevention strategies were implemented to decrease cholesterol levels. Despite major improvements, dyslipidemia still affects around 12% of adults in the United States [\[109\]](#page-13-6). Some of the identified risk factors for increased cholesterol levels are obesity, lack of physical activity, smoking, an unhealthy diet, and diabetes [\[110](#page-13-7)[–112\]](#page-13-8). Uncontrolled diabetes is one of the most common conditions contributing to dyslipidemia through insulin resistance, therefore leading to hyperinsulinemia. Elevated insulin concentrations contribute to increases in both lowdensity lipoprotein-cholesterol (LDL-C) and triglycerides levels in contrast to fewer highdensity lipoprotein-cholesterol (HDL-C) particles [\[113–](#page-13-9)[115\]](#page-13-10).

Gut microbiota has been shown to be involved in the occurrence of hyperlipidemia [\[116\]](#page-13-11). In fact, a recent study reported that people with dyslipidemia exhibit lower levels of fecal butyrate, acetate, and propionate when compared to healthy controls [\[117\]](#page-13-12). These metabolites represent the main SCFAs and are produced by a variety of gut bacteria, such as *Bifidobacterium*, *Lactobacillus*, *Faecalibacterium prausnitzii*, and *Roseburia* [\[118\]](#page-13-13). Indeed, they are thought to protect against obesity and diabetes by improving lipid and glucose homeostasis as well as glucose tolerance [\[80,](#page-12-2)[119–](#page-13-14)[121\]](#page-13-15).

Additionally, in contrast to a lower abundance of SCFAs, patients with high levels of cholesterol excrete feces with a higher quantity of LPS-producing bacteria, such as *Escherichia coli* and *Enterobacter cloacae* [\[118\]](#page-13-13). LPS compose the cell walls of Gram-negative bacteria and are responsible for the release of pro-inflammatory cytokines [\[122\]](#page-13-16). An overproduction of these cytokines leads to increased circulating levels of nitric oxide, subsequently triggering a global activation of inflammatory reactions resulting in cardiac, renal, hepatic, and pulmonary failures [\[123](#page-13-17)[–125\]](#page-13-18).

Furthermore, patients with dyslipidemia tend to exhibit high levels of TMAO, which reduce levels of HDL-C, therefore increasing the risk of ischemic heart disease [\[126,](#page-13-19)[127\]](#page-13-20). TMAO was also shown to reduce the expression of cytochrome P450 family 7, subfamily A member 1 (CYP7A1), which is a key enzyme in cholesterol and bile acid metabolism, in addition to inhibiting cholesterol transport, thus inducing cholesterol accumulation in cells [\[128\]](#page-14-0).

Finally, the gut microbiome is involved in the production of secondary bile acids, which were shown to be protective against the development of dyslipidemia [\[129](#page-14-1)[,130\]](#page-14-2). In fact, these metabolites modulate glucose and cholesterol metabolism through the activation of two key receptors, specifically FXR and TGR5 [\[131](#page-14-3)[,132\]](#page-14-4). Several studies have established that a deficiency in any one of these receptors, particularly FXR, leads to dyslipidemia, with increased triglycerides and non-HDL-C levels [\[133–](#page-14-5)[135\]](#page-14-6). In contrast, the activation of FXR by secondary bile acids increases the activity and expression of LDL receptors in addition to an inhibition of the activity of proprotein convertase subtilisin/kexin type 9 (PCSK9) [\[136](#page-14-7)[–138\]](#page-14-8). Thus, FXR activation by the gut microbiome could lower LDL-C levels and contribute to a better control of dyslipidemia.

Altogether, the previous data suggest that SCFAs and secondary bile acids are protective against the incidence of dyslipidemia while other metabolites, such as LPS and TMAO, are detrimental, contributing to an increase in cholesterol levels.

6. Gut Microbiome and Atherosclerotic Cardiovascular Disease

Even though there has been substantial improvement in cardiovascular disease outcomes in the past few decades, ASCVD remains the leading cause of death around the world [\[139–](#page-14-9)[141\]](#page-14-10). Insufficient prevention strategies and uncontrolled risk factors are the reasons why cardiac diseases still top the list [\[139](#page-14-9)[,140\]](#page-14-11). Uncontrolled dyslipidemia, persistent inflammation, and high levels of oxidative stress greatly contribute to atherosclerosis [\[142\]](#page-14-12).

Traditionally, ASCVD prevention strategies focused solely on lifestyle modifications, such as eating a healthy diet and doing exercise, in addition to taking beneficial medications, such as aspirin and beta-blockers [\[143–](#page-14-13)[145\]](#page-14-14). The gut microbiota was neglected until the scientific community realized its importance in playing key functions in the body, thus labeling it "the forgotten organ" [\[146\]](#page-14-15). At the beginning of the millennium, a relationship between the microbiota and atherosclerosis was demonstrated for the first time, after multiple studies reported the presence of DNA of numerous bacterial species in a plaque of cholesterol [\[2](#page-9-1)[,147\]](#page-14-16). Another trial suggested the presence of dysbiosis in individuals suffering from atherosclerosis, with an abundance of *Actinobacteria* in their intestine as compared to a large quantity of butyrate-producing bacteria in healthy controls [\[148\]](#page-14-17).

Recent studies in mice have suggested an important role of the gut microbiota in converting dietary phosphatidylcholine to TMA, which is then oxidized in the liver to TMAO [\[8](#page-9-6)[,149\]](#page-14-18). TMAO is a pro-atherosclerotic molecule, with patients suffering from ASCVD displaying significantly higher levels of TMAO as compared with healthy individuals [149[\]. Se](#page-14-18)veral experiments have demonstrated that TMAO was involved in all steps leading to atherogenesis, specifically foam cells' formations, endothelial dysfunction, thrombus generation, and plaque instability leading, ultimately, to plaque rupture and acute coronary syndrome [\[8](#page-9-6)[,65](#page-11-11)[,66](#page-11-12)[,150](#page-14-19)].

TMAO is a proposition of the proposition of the patients suffering from \mathcal{S} and \mathcal{S} are patients suffering from \mathcal{S}

Five mechanisms linking TMAO to ASCVD have been recently proposed. Figure [2](#page-6-0) Five mechanisms linking TMAO to ASCVD have been recently proposed. Figure 2 summarizes these mechanisms. The first pro-atherosclerotic mechanism is an increased summarizes these mechanisms. The first pro-atherosclerotic mechanism is an increased migration of macrophages and an augmented formation of foam cells in cholesterol migration of macrophages and an augmented formation of foam cells in cholesterol plaques [\[151,](#page-14-20)[152\]](#page-14-21). In a trial involving mice supplemented with either choline or TMAO, plaques [151,152]. In a trial involving mice supplemented with either choline or TMAO, Park et al. found that scavenger receptor-A (SR-A) and CD36, two macrophage receptors Park et al. found that scavenger receptor-A (SR-A) and CD36, two macrophage receptors associated with atherosclerosis, were both increased when compared to control mice [\[153\]](#page-15-0). associated with atherosclerosis, were both increased when compared to control mice [153].

Figure 2. Proposed mechanisms linking trimethylamine N-oxide to atherosclerotic cardiovascular **Figure 2.** Proposed mechanisms linking trimethylamine N-oxide to atherosclerotic cardiovascular disease. HDL-C: high-density lipoprotein-cholesterol, RCT: reverse cholesterol transport, ROS: reactive oxygen species, TMAO: trimethylamine N-oxide.

The second metabolic pathway states that TMAO impacts cholesterol metabolism by The second metabolic pathway states that TMAO impacts cholesterol metabolism by inhibiting the reverse cholesterol transport (RCT) system and diminishing cholesterol excretion through the biliary system [66]. Th[e RC](#page-11-12)T system helps maintain cholesterol homeostasis by transporting cholesterol from peripheral tissues to the liver for biliary excretion [\[154\]](#page-15-1). A recently published study demonstrated a 35% decrease in the RCT system in mice fed with a diet containing TMAO when compared with healthy controls [\[66\]](#page-11-12). TMAO increases cholesterol levels by downregulating two cytochromes, namely CYP7A1 and also increases cholesterol levels by downregulating two cytochromes, namely CYP7A1 and CYP27A1. Downregulation of these enzymes leads to decreased bile acid secretion, which CYP27A1. Downregulation of these enzymes leads to decreased bile acid secretion, which reduces cholesterol excretion, thereby contributing to accelerated atherosclerosis [\[155](#page-15-2)[,156\]](#page-15-3).

The third likely pathway is TMAO-induced endothelial dysfunction [\[157\]](#page-15-4). Indeed, TMAO induces vascular inflammation by increasing recruitment of leucocytes to endothe-lial cells through a G-protein-coupled receptor (GPCR) pathway [\[158\]](#page-15-5). TMAO also causes inflammation through mitogen-activated protein kinase (MAPK) and NF-κB signaling pathways [\[158\]](#page-15-5). Finally, protein kinase C is a known mediator of endothelial dysfunction, and its activity was found to be significantly increased in response to a diet enhanced with IMAO [159,160]. TMAO [\[159](#page-15-6)[,160\]](#page-15-7).

The fourth pathway involves an increase in oxidative stress. Recent trials have demonstrated that TMAO activates the nucleotide-binding oligomerization domain-like receptor family pyrin domain-containing 3 (NLRP3) inflammasome in endothelial cells. Activation of the NLRP3 inflammasome is involved in the production of ROS through activation of the mitochondrial reactive oxygen species signaling pathway [\[161,](#page-15-8)[162\]](#page-15-9). Oxidative stress causes cell damage and is involved in the pathogenesis of multiple diseases, including ASCVD [\[163\]](#page-15-10).

The fifth and last identified pathway mediates TMAO-associated atherosclerosis through suppression of endothelial progenitor cells' (EPC) production [\[164\]](#page-15-11). Several trials have demonstrated that decreased levels of EPCs contribute to endothelial dysfunction, since EPCs are known for their role in repairing and regenerating damaged endothelium following vascular injury [\[165–](#page-15-12)[167\]](#page-15-13). Chou et al. have found that TMAO levels were proportional to plasmatic inflammatory markers, specifically high-sensitivity C-reactive protein (hsCRP), IL-6, and TNF- α [\[164\]](#page-15-11). By contrast, TMAO levels are inversely proportional to EPC levels, thus leading to impaired endothelial function [\[164\]](#page-15-11).

Other than TMAO, LPS are other pro-atherosclerotic metabolites released by Gramnegative bacteria [\[168\]](#page-15-14). In healthy individuals, butyrate is secreted by the gut microbiota in a sufficient amount to maintain the intestinal barrier [\[169\]](#page-15-15). In atherosclerosis, gut microbiome dysbiosis results in a reduced number of butyrate-producing bacteria, subsequently leading to increased intestinal permeability and increased LPS levels [\[170,](#page-15-16)[171\]](#page-15-17). LPS activate numerous inflammatory pathways that contribute to the occurrence of atherosclerosis. Indeed, LPS induce the generation of ROS by activating nicotinamide adenine dinucleotide phosphate (NADPH) oxidase [\[172\]](#page-15-18). NADPH oxidase produces ROS and induces the production of pro-inflammatory cytokines, such as $TNF-\alpha$, IL-6, and IL-8 [\[173\]](#page-15-19). Furthermore, LPS provoke expression of inflammatory mediators, resulting in increased infiltration of inflammatory cells in cholesterol plaques [\[174\]](#page-15-20). These inflammatory cells include neutrophils, monocytes, selectins, and integrins, and are involved in the progression of atherosclerosis [\[175\]](#page-15-21). Thus, LPS directly contribute to the development and progression of atherosclerosis.

Moreover, the gut microbiome produces secondary bile acids, which are involved in the activation of two key receptors, the membranous TGR5 and the nuclear FXR [\[130\]](#page-14-2). Their activation is associated with a slowed progression of atherosclerosis through an inhibition of NF-κB activity, resulting in a decreased production of pro-inflammatory cytokines, as well as inhibited LDL uptake via a lowered expression of CD36 [\[130\]](#page-14-2). In contrast, the absence of FXR in mouse models was demonstrated to be linked with a decreased survival rate owing to more severe atherosclerosis with increased atherosclerotic plaque burden [\[176\]](#page-15-22).

Likewise, SCFAs are thought to be beneficial on ASCVD by inhibiting various inflammatory mechanisms thought to induce atherosclerosis. SCFAs are produced by the gut microbiota through fermentation of dietary fibers [\[60,](#page-11-6)[177\]](#page-15-23). Ingestion of a high-fiber diet contributes to an improved glycemic control and weight loss as well as increased blood concentrations of SCFAs [\[178–](#page-15-24)[180\]](#page-16-0). SCFAs, particularly butyrate, were recently shown to suppress atherosclerotic lesions in mice supplemented with a high-fiber diet [\[181,](#page-16-1)[182\]](#page-16-2). Butyrate is thought to increase plaque stability by decreasing ROS and nitric oxide release from macrophages as well as reducing production of known inflammatory molecules, such as chemotaxis protein-1, vascular cell adhesion molecule-1, and matrix metalloproteinase-2 [\[181,](#page-16-1)[182\]](#page-16-2).

A recently published trial demonstrated that a dysbiotic microbiota is positively associated with an increase in the size of acute myocardial infarction in rats [\[183\]](#page-16-3). Probiotics were shown to attenuate the infarct size observed in the dysbiotic group suggesting that microbiota is an important component of ischemic damage. In addition to an increase in infarct size, other noteworthy findings include a higher plasma LPS concentration secondary to increased gut permeability together with an increased *Firmicutes* to *Bacteroidetes* ratio.

Thus, dysbiosis contributes to the development of atherosclerosis through increases in TMAO and LPS levels while secondary bile acids and SCFAs are protective. Figure [3](#page-8-0) illustrates the implication of the gut microbiome in the occurrence of ASCVD.

illustrates the implication of the gut microbiome in the occurrence of ASCVD.

Figure 3. Role of the gut microbiome in the incidence of atherosclerotic cardiovascular disease. A low-fiber diet is associated with a decreased production of the short-chain fatty acid butyrate, subsequently aggravating dysbiosis as well as sustaining local and systemic inflammation through leakage of bacterial toxins, notably LPS. A modern western diet rich in red meat promotes bacterial leakage of bacterial toxins, notably LPS. A modern western diet rich in red meat promotes bacterial production of TMA, which is then oxidized to the pro-atherosclerotic metabolite TMAO in the liver. production of TMA, which is then oxidized to the pro-atherosclerotic metabolite TMAO in the liver. FMO3: flavin-containing monooxygenase 3, LPS: lipopolysaccharides, TMA: trimethylamine, FMO3: flavin-containing monooxygenase 3, LPS: lipopolysaccharides, TMA: trimethylamine, TMAO: TMAO: trimethylamine N-oxide. trimethylamine N-oxide.

7. Conclusions

In the last few decades, major advances have been made in the understanding of In the last few decades, major advances have been made in the understanding of physiological and pathological functions of the gut microbiota. In the cardiovascular field, physiological and pathological functions of the gut microbiota. In the cardiovascular field, there is no doubt nowadays that the microbiome plays a crucial role in the development there is no doubt nowadays that the microbiome plays a crucial role in the development of ASCVD. The microbiome is directly involved in all steps leading to atherogenesis, cluding all major cardiovascular risk factors, specifically hypertension, obesity, diabetes, including all major cardiovascular risk factors, specifically hypertension, obesity, diabetes, and dyslipidemia. In the upcoming years, the challenge will be to transition from theoretical understanding to clinical practice as major pathophysiologic mechanisms linking the gut microbiota to ASCVD have been elucidated. In the near future, can an in-depth analysis of the gut microbiota be used as a cardiovascular risk marker for which the use of probiotics probiotics might prove beneficial when used adequately? might prove beneficial when used adequately?

Author Contributions: Writing, review, and editing: A.A.S.; M.P.; and G.R. All authors have read **Author Contributions:** Writing, review, and editing: A.A.S., M.P. and G.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding. **Funding:** This research received no external funding.

Institutional Review Board Statement: Not applicable. **Institutional Review Board Statement:** Not applicable.

Informed Consent Statement: Not applicable. **Informed Consent Statement:** Not applicable.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is **Data Availability Statement:** No new data were created or analyzed in this study. Data sharing is not applicable to this article. not applicable to this article.

Acknowledgments: The figures were made using Biorender.com (accessed on 5 March 2023).

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Khan, M.A.; Hashim, M.J.; Mustafa, H.; Baniyas, M.Y.; Al Suwaidi, S.K.B.M.; AlKatheeri, R.; Alblooshi, F.M.K.; Almatrooshi, M.E.A.H.; Alzaabi, M.E.H.; Al Darmaki, R.S.; et al. Global Epidemiology of Ischemic Heart Disease: Results from the Global Burden of Disease Study. *Cureus* **2020**, *12*, e9349. [\[CrossRef\]](http://doi.org/10.7759/cureus.9349)
- 2. Haraszthy, V.I.; Zambon, J.J.; Trevisan, M.; Zeid, M.; Genco, R.J. Identification of periodontal pathogens in atheromatous plaques. *J. Periodontol.* **2000**, *71*, 1554–1560. [\[CrossRef\]](http://doi.org/10.1902/jop.2000.71.10.1554) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11063387)
- 3. Allin, K.H.; Nielsen, T.; Pedersen, O. Mechanisms in endocrinology: Gut microbiota in patients with type 2 diabetes mellitus. *Eur. J. Endocrinol.* **2015**, *172*, R167–R177. [\[CrossRef\]](http://doi.org/10.1530/EJE-14-0874)
- 4. Lepage, P.; Leclerc, M.C.; Joossens, M.; Mondot, S.; Blottière, H.M.; Raes, J.; Ehrlich, D.; Doré, J. A metagenomic insight into our gut's microbiome. *Gut* **2013**, *62*, 146–158. [\[CrossRef\]](http://doi.org/10.1136/gutjnl-2011-301805)
- 5. Flint, H.J.; Scott, K.P.; Louis, P.; Duncan, S.H. The role of the gut microbiota in nutrition and health. *Nat. Rev. Gastroenterol. Hepatol.* **2012**, *9*, 577–589. [\[CrossRef\]](http://doi.org/10.1038/nrgastro.2012.156) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22945443)
- 6. Shanahan, F. The gut microbiota—A clinical perspective on lessons learned. *Nat. Rev. Gastroenterol. Hepatol.* **2012**, *9*, 609–614. [\[CrossRef\]](http://doi.org/10.1038/nrgastro.2012.145)
- 7. Qin, J.; Li, R.; Raes, J.; Arumugam, M.; Burgdorf, K.S.; Manichanh, C.; Nielsen, T.; Pons, N.; Levenez, F.; Yamada, T.; et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* **2010**, *464*, 59–65. [\[CrossRef\]](http://doi.org/10.1038/nature08821) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20203603)
- 8. Wang, Z.; Klipfell, E.; Bennett, B.J.; Koeth, R.; Levison, B.S.; Dugar, B.; Feldstein, A.E.; Britt, E.B.; Fu, X.; Chung, Y.M.; et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* **2011**, *472*, 57–63. [\[CrossRef\]](http://doi.org/10.1038/nature09922)
- 9. Spencer, M.D.; Hamp, T.J.; Reid, R.W.; Fischer, L.M.; Zeisel, S.H.; Fodor, A.A. Association between composition of the human gastrointestinal microbiome and development of fatty liver with choline deficiency. *Gastroenterology* **2011**, *140*, 976–986. [\[CrossRef\]](http://doi.org/10.1053/j.gastro.2010.11.049) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21129376)
- 10. Krueger, S.K.; Williams, D.E. Mammalian flavin-containing monooxygenases: Structure/function, genetic polymorphisms and role in drug metabolism. *Pharmacol. Ther.* **2005**, *106*, 357–387. [\[CrossRef\]](http://doi.org/10.1016/j.pharmthera.2005.01.001)
- 11. Fennema, D.; Phillips, I.R.; Shephard, E.A. Trimethylamine and Trimethylamine N-Oxide, a Flavin-Containing Monooxygenase 3 (FMO3)-Mediated Host-Microbiome Metabolic Axis Implicated in Health and Disease. *Drug. Metab. Dispos.* **2016**, *44*, 1839–1850. [\[CrossRef\]](http://doi.org/10.1124/dmd.116.070615)
- 12. Gatarek, P.; Kaluzna-Czaplinska, J. Trimethylamine N-oxide (TMAO) in human health. *EXCLI J.* **2021**, *20*, 301–319. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33746664)
- 13. Chiang, J.Y. Bile acid metabolism and signaling. *Compr. Physiol.* **2013**, *3*, 1191–1212. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23897684)
- 14. Porez, G.; Prawitt, J.; Gross, B.; Staels, B. Bile acid receptors as targets for the treatment of dyslipidemia and cardiovascular disease. *J. Lipid. Res.* **2012**, *53*, 1723–1737. [\[CrossRef\]](http://doi.org/10.1194/jlr.R024794)
- 15. Watanabe, M.; Houten, S.M.; Mataki, C.; Christoffolete, M.A.; Kim, B.W.; Sato, H.; Messaddeq, N.; Harney, J.W.; Ezaki, O.; Kodama, T.; et al. Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. *Nature* **2006**, *439*, 484–489. [\[CrossRef\]](http://doi.org/10.1038/nature04330)
- 16. Thomas, C.; Gioiello, A.; Noriega, L.; Strehle, A.; Oury, J.; Rizzo, G.; Macchiarulo, A.; Yamamoto, H.; Mataki, C.; Pruzanski, M.; et al. TGR5-mediated bile acid sensing controls glucose homeostasis. *Cell. Metab.* **2009**, *10*, 167–177. [\[CrossRef\]](http://doi.org/10.1016/j.cmet.2009.08.001)
- 17. Yoo, J.Y.; Sniffen, S.; McGill Percy, K.C.; Pallaval, V.B.; Chidipi, B. Gut Dysbiosis and Immune System in Atherosclerotic Cardiovascular Disease (ACVD). *Microorganisms* **2022**, *10*, 108. [\[CrossRef\]](http://doi.org/10.3390/microorganisms10010108)
- 18. Miyazaki-Anzai, S.; Masuda, M.; Kohno, S.; Levi, M.; Shiozaki, Y.; Keenan, A.L.; Miyazaki, M. Simultaneous inhibition of FXR and TGR5 exacerbates atherosclerotic formation. *J. Lipid. Res.* **2018**, *59*, 1709–1713. [\[CrossRef\]](http://doi.org/10.1194/jlr.M087239)
- 19. Hu, Y.B.; Liu, X.Y.; Zhan, W. Farnesoid. X receptor. *agonist. INT-767 attenuates liver steatosis and inflammation in rat model of nonalcoholic steatohepatitis. Drug. Des. Devel. Ther.* **2018**, *12*, 2213–2221. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30038487)
- 20. Mullen, L.M.; Chamberlain, G.; Sacre, S. Pattern recognition receptors as potential therapeutic targets in inflammatory rheumatic disease. *Arthritis. Res. Ther.* **2015**, *17*, 122. [\[CrossRef\]](http://doi.org/10.1186/s13075-015-0645-y) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25975607)
- 21. Guha, M.; Mackman, N. LPS induction of gene expression in human monocytes. *Cell. Signal.* **2001**, *13*, 85–94. [\[CrossRef\]](http://doi.org/10.1016/S0898-6568(00)00149-2)
- 22. Gorabi, A.M.; Kiaie, N.; Khosrojerdi, A.; Jamialahmadi, T.; Al-Rasadi, K.; Johnston, T.P.; Sahebkar, A. Implications for the role of lipopolysaccharide in the development of atherosclerosis. *Trends. Cardiovasc. Med.* **2022**, *32*, 525–533. [\[CrossRef\]](http://doi.org/10.1016/j.tcm.2021.08.015)
- 23. Watts, C.; West, M.A.; Zaru, R. TLR signalling regulated antigen presentation in dendritic cells. *Curr. Opin. Immunol.* **2010**, *22*, 124–130. [\[CrossRef\]](http://doi.org/10.1016/j.coi.2009.12.005)
- 24. Karnati, H.K.; Pasupuleti, S.R.; Kandi, R.; Undi, R.B.; Sahu, I.; Kannaki, T.R.; Subbiah, M.; Gutti, R.K. TLR-4 signalling pathway: MyD88 independent pathway up-regulation in chicken breeds upon LPS treatment. *Vet. Res. Commun.* **2015**, *39*, 73–78. [\[CrossRef\]](http://doi.org/10.1007/s11259-014-9621-2)
- 25. Chen, T.; Huang, W.; Qian, J.; Luo, W.; Shan, P.; Cai, Y.; Lin, K.; Wu, G.; Liang, G. Macrophage-derived myeloid differentiation protein 2 plays an essential role in ox-LDL-induced inflammation and atherosclerosis. *EBioMedicine* **2020**, *53*, 102706. [\[CrossRef\]](http://doi.org/10.1016/j.ebiom.2020.102706)
- 26. Griendling, K.K.; Sorescu, D.; Ushio-Fukai, M. NAD(P)H oxidase: Role in cardiovascular biology and disease. *Circ. Res.* **2000**, *86*, 494–501. [\[CrossRef\]](http://doi.org/10.1161/01.RES.86.5.494) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10720409)
- 27. Sweet, M.J.; Hume, D.A. Endotoxin signal transduction in macrophages. *J. Leukoc. Biol.* **1996**, *60*, 8–26. [\[CrossRef\]](http://doi.org/10.1002/jlb.60.1.8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/8699127)
- 28. Diks, S.H.; van Deventer, S.J.; Peppelenbosch, M.P. Lipopolysaccharide recognition, internalisation, signalling and other cellular effects. *J. Endotoxin. Res.* **2001**, *7*, 335–348. [\[CrossRef\]](http://doi.org/10.1179/096805101101532909) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11753202)
- 29. Parada Venegas, D.; De la Fuente, M.K.; Landskron, G.; González, M.J.; Quera, R.; Dijkstra, G.; Harmsen, H.J.M.; Faber, K.N.; Hermoso, M.A. Short Chain Fatty Acids (SCFAs)-Mediated Gut Epithelial and Immune Regulation and Its Relevance for Inflammatory. *Bowel. Dis. Front. Immunol.* **2019**, *10*, 277. [\[CrossRef\]](http://doi.org/10.3389/fimmu.2019.00277)
- 30. Flint, H.J.; Scott, K.P.; Duncan, S.H.; Louis, P.; Forano, E. Microbial degradation of complex carbohydrates in the gut. *Gut. Microbes.* **2012**, *3*, 289–306. [\[CrossRef\]](http://doi.org/10.4161/gmic.19897) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22572875)
- 31. Chambers, E.S.; Preston, T.; Frost, G.; Morrison, D.J. Role of Gut Microbiota-Generated Short-Chain Fatty Acids in Metabolic and Cardiovascular Health. *Curr. Nutr. Rep.* **2018**, *7*, 198–206. [\[CrossRef\]](http://doi.org/10.1007/s13668-018-0248-8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30264354)
- 32. Arpaia, N.; Campbell, C.; Fan, X.; Dikiy, S.; van der Veeken, J.; deRoos, P.; Liu, H.; Cross, J.R.; Pfeffer, K.; Coffer, P.J.; et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* **2013**, *504*, 451–455. [\[CrossRef\]](http://doi.org/10.1038/nature12726)
- 33. Zheng, X.X.; Zhou, T.; Wang, X.A.; Tong, X.H.; Ding, J.W. Histone deacetylases and atherosclerosis. *Atherosclerosis.* **2015**, *240*, 355–366. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2014.12.048) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25875381)
- 34. Usami, M.; Kishimoto, K.; Ohata, A.; Miyoshi, M.; Aoyama, M.; Fueda, Y.; Kotani, J. Butyrate and trichostatin A attenuate nuclear factor kappaB activation and tumor necrosis factor alpha secretion and increase prostaglandin E2 secretion in human peripheral blood mononuclear cells. *Nutr. Res.* **2008**, *28*, 321–328. [\[CrossRef\]](http://doi.org/10.1016/j.nutres.2008.02.012) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19083427)
- 35. Ottosson, F.; Brunkwall, L.; Smith, E.; Orho-Melander, M.; Nilsson, P.M.; Fernandez, C.; Melander, O. The gut microbiota-related metabolite phenylacetylglutamine associates with increased risk of incident coronary artery disease. *J. Hypertens.* **2020**, *38*, 2427–2434. [\[CrossRef\]](http://doi.org/10.1097/HJH.0000000000002569)
- 36. Nemet, I.; Saha, P.P.; Gupta, N.; Zhu, W.; Romano, K.A.; Skye, S.M.; Cajka, T.; Mohan, M.L.; Li, L.; Wu, Y.; et al. A Cardiovascular Disease-Linked Gut Microbial Metabolite Acts via Adrenergic Receptors. *Cell* **2020**, *180*, 862–877e22. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2020.02.016)
- 37. Poesen, R.; Claes, K.; Evenepoel, P.; de Loor, H.; Augustijns, P.; Kuypers, D.; Meijers, B. Microbiota-Derived Phenylacetylglutamine Associates with Overall Mortality and Cardiovascular Disease in Patients with CKD. *J. Am. Soc. Nephrol.* **2016**, *27*, 3479–3487. [\[CrossRef\]](http://doi.org/10.1681/ASN.2015121302) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27230658)
- 38. Liu, Y.; Liu, S.; Zhao, Z.; Song, X.; Qu, H.; Liu, H. Phenylacetylglutamine is associated with the degree of coronary atherosclerotic severity assessed by coronary computed tomographic angiography in patients with suspected coronary artery disease. *Atherosclerosis* **2021**, *333*, 75–82. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2021.08.029) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34438323)
- 39. Camen, S.; Csengeri, D.; Geelhoed, B.; Niiranen, T.; Gianfagna, F.; Vishram-Nielsen, J.K.; Costanzo, S.; Söderberg, S.; Vartiainen, E.; Börschel, C.S.; et al. Risk Factors, Subsequent Disease Onset, and Prognostic Impact of Myocardial Infarction and Atrial Fibrillation. *J. Am. Heart. Assoc.* **2022**, *11*, e024299. [\[CrossRef\]](http://doi.org/10.1161/JAHA.121.024299)
- 40. Hajar, R. Risk Factors for Coronary Artery Disease: Historical Perspectives. *Heart Views* **2017**, *18*, 109–114. [\[CrossRef\]](http://doi.org/10.4103/HEARTVIEWS.HEARTVIEWS_106_17)
- 41. Wang, Y.; Li, J.; Zheng, X.; Jiang, Z.; Hu, S.; Wadhera, R.K.; Bai, X.; Lu, J.; Wang, Q.; Li, Y.; et al. Risk Factors Associated With Major Cardiovascular Events 1 Year After Acute Myocardial Infarction. JAMA Netw. *Open.* **2018**, *1*, e181079.
- 42. Rabi, D.M.; McBrien, K.A.; Sapir-Pichhadze, R.; Nakhla, M.; Ahmed, S.B.; Dumanski, S.M.; Butalia, S.; Leung, A.A.; Harris, K.C.; Cloutier, L.; et al. Hypertension Canada's 2020 Comprehensive Guidelines for the Prevention, Diagnosis, Risk Assessment, and Treatment of Hypertension in Adults and Children. *Can. J. Cardiol.* **2020**, *36*, 596–624. [\[CrossRef\]](http://doi.org/10.1016/j.cjca.2020.02.086)
- 43. Whelton, P.K.; Carey, R.M.; Aronow, W.S.; Casey, D.E., Jr.; Collins, K.J.; Dennison Himmelfarb, C.; DePalma, S.M.; Gidding, S.; Jamerson, K.A.; Jones, D.W.; et al. 2017 ACC/AHA/AAPA/ABC/ACPM/AGS/APhA/ASH/ASPC/NMA/PCNA Guideline for the Prevention, Detection, Evaluation, and Management of High Blood Pressure in Adults: Executive Summary: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. *Hypertension* **2018**, *71*, 1269–1324.
- 44. Williams, B.; Mancia, G.; Spiering, W.; Agabiti Rosei, E.; Azizi, M.; Burnier, M.; Clement, D.L.; Coca, A.; de Simone, G.; Dominiczak, A.; et al. 2018 ESC/ESH Guidelines for the management of arterial hypertension. *Eur. Heart J.* **2018**, *39*, 3021–3104. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehy339) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30165516)
- 45. Dan, X.; Mushi, Z.; Baili, W.; Han, L.; Enqi, W.; Huanhu, Z.; Shuchun, L. Differential analysis of hypertension-associated intestinal microbiota. *Int. J. Med. Sci.* **2019**, *16*, 872–881. [\[CrossRef\]](http://doi.org/10.7150/ijms.29322) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31337961)
- 46. de la Cuesta-Zuluaga, J.; Mueller, N.T.; Alvarez-Quintero, R.; Velásquez-Mejía, E.P.; Sierra, J.A.; Corrales-Agudelo, V.; Carmona, J.A.; Abad, J.M.; Escobar, J.S. Higher fecal short-chain fatty acid levels are associated with gut microbiome dysbiosis, obesity, hypertension and cardiometabolic disease risk fac- tors. *Nutrients* **2018**, *11*, 51. [\[CrossRef\]](http://doi.org/10.3390/nu11010051) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30591685)
- 47. Li, J.; Zhao, F.; Wang, Y.; Chen, J.; Tao, J.; Tian, G.; Wu, S.; Liu, W.; Cui, Q.; Geng, B.; et al. Gut microbiota dysbiosis contributes to the development of hypertension. *Microbiome* **2017**, *5*, 14. [\[CrossRef\]](http://doi.org/10.1186/s40168-016-0222-x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28143587)
- 48. Sun, S.; Lulla, A.; Sioda, M.; Winglee, K.; Wu, M.C.; Jacobs, D.R., Jr.; Shikany, J.M.; Lloyd-Jones, D.M.; Launer, L.J.; Fodor, A.A.; et al. Gut microbiota com- position and blood pressure. *Hypertension* **2019**, *73*, 998–1006. [\[CrossRef\]](http://doi.org/10.1161/HYPERTENSIONAHA.118.12109)
- 49. Verhaar, B.J.H.; Collard, D.; Prodan, A.; Levels, J.H.M.; Zwinderman, A.H.; Bäckhed, F.; Vogt, L.; Peters, M.J.L.; Muller, M.; Nieuwdorp, M.; et al. Associations between gut microbiota, faecal short-chain fatty acids, and blood pressure across ethnic groups: The HELIUS study. *Eur. Heart J.* **2020**, *41*, 4259–4267. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehaa704)
- 50. Yan, Q.; Gu, Y.; Li, X.; Yang, W.; Jia, L.; Chen, C.; Han, X.; Huang, Y.; Zhao, L.; Li, P.; et al. Alterations of the gut microbiome in hypertension. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 381. [\[CrossRef\]](http://doi.org/10.3389/fcimb.2017.00381)
- 51. Yang, T.; Santisteban, M.M.; Rodriguez, V.; Li, E.; Ahmari, N.; Carvajal, J.M.; Zadeh, M.; Gong, M.; Qi, Y.; Zubcevic, J.; et al. Gut dysbiosis is linked to hypertension. *Hypertension* **2015**, *65*, 1331–1340. [\[CrossRef\]](http://doi.org/10.1161/HYPERTENSIONAHA.115.05315) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25870193)
- 52. Kim, S.; Goel, R.; Kumar, A.; Qi, Y.; Lobaton, G.; Hosaka, K.; Mohammed, M.; Handberg, E.M.; Richards, E.M.; Pepine, C.J.; et al. Imbalance of gut microbiome and intestinal epithelial barrier dysfunction in patients with high blood pressure. *Clin. Sci.* **2018**, *132*, 701–718. [\[CrossRef\]](http://doi.org/10.1042/CS20180087)
- 53. Huart, J.; Leenders, J.; Taminiau, B.; Descy, J.; Saint-Remy, A.; Daube, G.; Krzesinski, J.M.; Melin, P.; de Tullio, P.; Jouret, F. Gut microbiota and fecal levels of short-chain fatty acids differ upon 24-hour blood pressure levels in men. *Hypertension* **2019**, *74*, 1005–1013. [\[CrossRef\]](http://doi.org/10.1161/HYPERTENSIONAHA.118.12588)
- 54. Jackson, M.A.; Verdi, S.; Maxan, M.E.; Shin, C.M.; Zierer, J.; Bowyer, R.C.E.; Martin, T.; Williams, F.M.K.; Menni, C.; Bell, J.T.; et al. Gut microbiota associations with common diseases and prescription medications in a population-based cohort. *Nat. Commun.* **2018**, *9*, 2655. [\[CrossRef\]](http://doi.org/10.1038/s41467-018-05184-7) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29985401)
- 55. Schiffrin, E.L. Immune mechanisms in hypertension and vascular injury. *Clin. Sci.* **2014**, *126*, 267–274. [\[CrossRef\]](http://doi.org/10.1042/CS20130407)
- 56. Cotillard, A.; Kennedy, S.P.; Kong, L.C.; Prifti, E.; Pons, N.; Le Chatelier, E.; Almeida, M.; Quinquis, B.; Levenez, F.; Galleron, N.; et al. Dietary intervention impact on gut microbial gene richness. *Nature* **2013**, *500*, 585–858. [\[CrossRef\]](http://doi.org/10.1038/nature12480) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23985875)
- 57. Wilck, N.; Matus, M.G.; Kearney, S.M.; Olesen, S.W.; Forslund, K.; Bartolomaeus, H.; Haase, S.; Mähler, A.; Balogh, A.; Markó, L.; et al. Salt-responsive gut commensal modulates TH17 axis and disease. *Nature* **2017**, *551*, 585–589. [\[CrossRef\]](http://doi.org/10.1038/nature24628) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29143823)
- 58. Maifeld, A.; Bartolomaeus, H.; Löber, U.; Avery, E.G.; Steckhan, N.; Markó, L.; Wilck, N.; Hamad, I.; Šušnjar, U.; Mähler, A.; et al. Fasting alters the gut microbiome reducing blood pressure and body weight in metabolic syndrome patients. *Nat. Commun.* **2021**, *12*, 1970. [\[CrossRef\]](http://doi.org/10.1038/s41467-021-22097-0)
- 59. Wikoff, W.R.; Anfora, A.T.; Liu, J.; Schultz, P.G.; Lesley, S.A.; Peters, E.C.; Siuzdak, G. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 3698–3703. [\[CrossRef\]](http://doi.org/10.1073/pnas.0812874106) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19234110)
- 60. Cummings, J.H.; Pomare, E.W.; Branch, W.J.; Naylor, C.P.; Macfarlane, G.T. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut.* **1987**, *28*, 1221–1227. [\[CrossRef\]](http://doi.org/10.1136/gut.28.10.1221)
- 61. Marques, F.Z.; Mackay, C.R.; Kaye, D.M. Beyond gut feelings: How the gut microbiota regulates blood pressure. *Nat. Rev. Cardiol.* **2018**, *15*, 20–32. [\[CrossRef\]](http://doi.org/10.1038/nrcardio.2017.120)
- 62. Bartolomaeus, H.; Balogh, A.; Yakoub, M.; Homann, S.; Markó, L.; Höges, S.; Tsvetkov, D.; Krannich, A.; Wundersitz, S.; Avery, E.G.; et al. Short-Chain Fatty Acid Propionate Protects From Hypertensive Cardiovascular Damage. *Circulation* **2019**, *139*, 1407–1421. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.118.036652) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30586752)
- 63. Marques, F.Z.; Nelson, E.; Chu, P.Y.; Horlock, D.; Fiedler, A.; Ziemann, M.; Tan, J.K.; Kuruppu, S.; Rajapakse, N.W.; El-Osta, A.; et al. High-Fiber Diet and Acetate Supplementation Change the Gut Microbiota and Prevent the Development of Hypertension and Heart Failure in Hypertensive Mice. *Circulation* **2017**, *135*, 964–977. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.116.024545)
- 64. Ge, X.; Zheng, L.; Zhuang, R.; Yu, P.; Xu, Z.; Liu, G.; Xi, X.; Zhou, X.; Fan, H. The gut microbial metabolite trimethylamine N-oxide and hypertension risk: A systematic review and dose-response meta-analysis. *Adv. Nutr.* **2020**, *11*, 66–76. [\[CrossRef\]](http://doi.org/10.1093/advances/nmz064) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31269204)
- 65. Zhu, W.; Gregory, J.C.; Org, E.; Buffa, J.A.; Gupta, N.; Wang, Z.; Li, L.; Fu, X.; Wu, Y.; Mehrabian, M.; et al. Gut microbial metabolite TMAO enhances platelet hyperreactivity and thrombosis risk. *Cell* **2016**, *165*, 111–124. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2016.02.011) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26972052)
- 66. Koeth, R.A.; Wang, Z.; Levison, B.S.; Buffa, J.A.; Org, E.; Sheehy, B.T.; Britt, E.B.; Fu, X.; Wu, Y.; Li, L.; et al. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat. Med.* **2013**, *19*, 576–585. [\[CrossRef\]](http://doi.org/10.1038/nm.3145) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23563705)
- 67. Jiang, S.; Shui, Y.; Cui, Y.; Tang, C.; Wang, X.; Qiu, X.; Hu, W.; Fei, L.; Li, Y.; Zhang, S.; et al. Gut microbiota dependent trimethylamine N-oxide aggravates angiotensin II-induced hypertension. *Redox. Biol.* **2021**, *46*, 102115. [\[CrossRef\]](http://doi.org/10.1016/j.redox.2021.102115)
- 68. Ufnal, M.; Jazwiec, R.; Dadlez, M.; Drapala, A.; Sikora, M.; Skrzypecki, J. Trimethylamine-N-oxide: A carnitine-derived metabolite that prolongs the hypertensive effect of angiotensin II in rats. *Can. J. Cardiol.* **2014**, *30*, 1700–1705. [\[CrossRef\]](http://doi.org/10.1016/j.cjca.2014.09.010) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25475471)
- 69. Brunt, V.E.; Gioscia-Ryan, R.A.; Richey, J.J.; Zigler, M.C.; Cuevas, L.M.; Gonzalez, A.; Vázquez-Baeza, Y.; Battson, M.L.; Smithson, A.T.; Gilley, A.D.; et al. Suppression of the gut microbiome ameliorates age-related arterial dysfunction and oxidative stress in mice. *J. Physiol.* **2019**, *597*, 2361–2378. [\[CrossRef\]](http://doi.org/10.1113/JP277336)
- 70. Saeedi, P.; Petersohn, I.; Salpea, P.; Malanda, B.; Karuranga, S.; Unwin, N.; Colagiuri, S.; Guariguata, L.; Motala, A.A.; Ogurtsova, K.; et al. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045 Results from the International Diabetes Federation Diabetes Atlas, 9th edition. *Diabetes. Res. Clin. Pract.* **2019**, *157*, 107843. [\[CrossRef\]](http://doi.org/10.1016/j.diabres.2019.107843)
- 71. Shuldiner, A.R.; Yang, R.; Gong, D.W. Resistin, obesity, and insulin resistance – the emerging role of the adipocyte as an endocrine organ. *N. Engl. J. Med.* **2001**, *345*, 1345–1346. [\[CrossRef\]](http://doi.org/10.1056/NEJM200111013451814) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11794158)
- 72. Matheus, A.S.; Tannus, L.R.; Cobas, R.A.; Palma, C.C.; Negrato, C.A.; Gomes, M.B. Impact of diabetes on cardiovascular disease: An update. *Int. J. Hypertens.* **2013**, *2013*, 653789. [\[CrossRef\]](http://doi.org/10.1155/2013/653789) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23533715)
- 73. Fletcher, B.; Gulanick, M.; Lamendola, C. Risk factors for type 2 diabetes mellitus. *J. Cardiovasc. Nurs.* **2002**, *16*, 17–23. [\[CrossRef\]](http://doi.org/10.1097/00005082-200201000-00003)
- 74. Grarup, N.; Sandholt, C.H.; Hansen, T.; Pedersen, O. Genetic susceptibility to type 2 diabetes and obesity: From genome-wide association studies to rare variants and beyond. *Diabetologia* **2014**, *57*, 1528–1541. [\[CrossRef\]](http://doi.org/10.1007/s00125-014-3270-4) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24859358)
- 75. Backhed, F.; Ding, H.; Wang, T.; Hooper, L.V.; Koh, G.Y.; Nagy, A.; Semenkovich, C.F.; Gordon, J.I. The gut microbiota as an environmental factor that regulates fat storage. *PNAS* **2004**, *101*, 15718–15723. [\[CrossRef\]](http://doi.org/10.1073/pnas.0407076101) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15505215)
- 76. Ley, R.E.; Turnbaugh, P.J.; Klein, S.; Gordon, J.I. Microbial ecology: Human gut microbes associated with obesity. *Nature* **2006**, *444*, 1022–1023. [\[CrossRef\]](http://doi.org/10.1038/4441022a)
- 77. Turnbaugh, P.J.; Ley, R.E.; Mahowald, M.A.; Magrini, V.; Mardis, E.R.; Gordon, J.I. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* **2006**, *444*, 1027–1031. [\[CrossRef\]](http://doi.org/10.1038/nature05414)
- 78. Chávez-Carbajal, A.; Pizano-Zárate, M.L.; Hernández-Quiroz, F.; Ortiz-Luna, G.F.; Morales-Hernández, R.M.; De Sales-Millán, A.; Hernández-Trejo, M.; García-Vite, A.; Beltrán-Lagunes, L.; Hoyo-Vadillo, C.; et al. Characterization of the Gut Microbiota of Individuals at Different T2D Stages Reveals a Complex Relationship with the Host. *Microorganisms* **2020**, *8*, 94. [\[CrossRef\]](http://doi.org/10.3390/microorganisms8010094) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31936722)
- 79. Karlsson, F.H.; Tremaroli, V.; Nookaew, I.; Bergström, G.; Behre, C.J.; Fagerberg, B.; Nielsen, J.; Bäckhed, F. Gut metagenome in European women with normal, impaired and diabetic glucose control. *Nature* **2013**, *498*, 99–103. [\[CrossRef\]](http://doi.org/10.1038/nature12198) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23719380)
- 80. Gao, Z.; Yin, J.; Zhang, J.; Ward, R.E.; Martin, R.J.; Lefevre, M.; Cefalu, W.T.; Ye, J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* **2009**, *58*, 1509–1517. [\[CrossRef\]](http://doi.org/10.2337/db08-1637)
- 81. Kuitunen, M.; Kukkonen, K.; Juntunen-Backman, K.; Korpela, R.; Poussa, T.; Tuure, T.; Haahtela, T.; Savilahti, E. Probiotics prevent IgE-associated allergy until age 5 years in cesarean-delivered children but not in the total cohort. *J. Allergy. Clin. Immunol.* **2009**, *123*, 335–341. [\[CrossRef\]](http://doi.org/10.1016/j.jaci.2008.11.019)
- 82. McLoughlin, R.M.; Mills, K.H. Influence of gastrointestinal commensal bacteria on the immune responses that mediate allergy and asthma. *J. Allergy. Clin. Immunol.* **2011**, *127*, 1097–1107. [\[CrossRef\]](http://doi.org/10.1016/j.jaci.2011.02.012) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21420159)
- 83. Henao-Mejia, J.; Elinav, E.; Jin, C.; Hao, L.; Mehal, W.Z.; Strowig, T.; Thaiss, C.A.; Kau, A.L.; Eisenbarth, S.C.; Jurczak, M.J.; et al. Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* **2012**, *482*, 179–185. [\[CrossRef\]](http://doi.org/10.1038/nature10809) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22297845)
- 84. Musso, G.; Gambino, R.; Cassader, M. Interactions between gut microbiota and host metabolism predisposing to obesity and diabetes. *Annu. Rev. Med.* **2011**, *62*, 361–380. [\[CrossRef\]](http://doi.org/10.1146/annurev-med-012510-175505)
- 85. Sharma, S.; Tripathi, P. Gut microbiome and type 2 diabetes: Where we are and where to go? J. *Nutr. Biochem.* **2019**, *63*, 101–108. [\[CrossRef\]](http://doi.org/10.1016/j.jnutbio.2018.10.003)
- 86. Tilg, H.; Moschen, A.R. Microbiota and diabetes: An evolving relationship. *Gut* **2014**, *63*, 1513–1521. [\[CrossRef\]](http://doi.org/10.1136/gutjnl-2014-306928) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24833634)
- 87. Tilg, H.; Moschen, A.R. Inflammatory mechanisms in the regulation of insulin resistance. *Mol. Med.* **2008**, *14*, 222–231. [\[CrossRef\]](http://doi.org/10.2119/2007-00119.Tilg) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18235842)
- 88. Ley, R.E.; Backhed, F.; Turnbaugh, P.; Lozupone, C.A.; Knight, R.D.; Gordon, J.I. Obesity alters gut microbial ecology. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 11070–11075. [\[CrossRef\]](http://doi.org/10.1073/pnas.0504978102)
- 89. Turnbaugh, P.J.; Hamady, M.; Yatsunenko, T.; Cantarel, B.L.; Duncan, A.; Ley, R.E.; Sogin, M.L.; Jones, W.J.; Roe, B.A.; Affourtit, J.P.; et al. A core gut microbiome in obese and lean twins. *Nature* **2009**, *457*, 480–484. [\[CrossRef\]](http://doi.org/10.1038/nature07540) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19043404)
- 90. Le Chatelier, E.; Nielsen, T.; Qin, J.; Prifti, E.; Hildebrand, F.; Falony, G.; Almeida, M.; Arumugam, M.; Batto, J.M.; Kennedy, S.; et al. Richness of human gut microbiome correlates with metabolic markers. *Nature* **2013**, *500*, 541–546. [\[CrossRef\]](http://doi.org/10.1038/nature12506)
- 91. Qin, J.; Li, Y.; Cai, Z.; Li, S.; Zhu, J.; Zhang, F.; Liang, S.; Zhang, W.; Guan, Y.; Shen, D.; et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* **2012**, *490*, 55–60. [\[CrossRef\]](http://doi.org/10.1038/nature11450)
- 92. Wang, L.; Li, C.; Huang, Q.; Fu, X. Polysaccharide from Rosa roxburghii Tratt Fruit Attenuates Hyperglycemia and Hyperlipidemia and Regulates Colon Microbiota in Diabetic db/db Mice. *J. Agric. Food Chem.* **2020**, *68*, 147–159. [\[CrossRef\]](http://doi.org/10.1021/acs.jafc.9b06247)
- 93. Cani, P.D.; Amar, J.; Iglesias, M.A.; Poggi, M.; Knauf, C.; Bastelica, D.; Neyrinck, A.M.; Fava, F.; Tuohy, K.M.; Chabo, C.; et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* **2007**, *56*, 1761–1772. [\[CrossRef\]](http://doi.org/10.2337/db06-1491)
- 94. Erridge, C.; Attina, T.; Spickett, C.M.; Webb, D.J. A high-fat meal induces low-grade endotoxemia: Evidence of a novel mechanism of postprandial inflammation. *Am. J. Clin. Nutr.* **2007**, *86*, 1286–1292. [\[CrossRef\]](http://doi.org/10.1093/ajcn/86.5.1286) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17991637)
- 95. Pussinen, P.J.; Havulinna, A.S.; Lehto, M.; Sundvall, J.; Salomaa, V. Endotoxemia is associated with an increased risk of incident diabetes. *Diabetes Care* **2011**, *34*, 392–397. [\[CrossRef\]](http://doi.org/10.2337/dc10-1676) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21270197)
- 96. Chen, S.; Henderson, A.; Petriello, M.C.; Romano, K.A.; Gearing, M.; Miao, J.; Schell, M.; Sandoval-Espinola, W.J.; Tao, J.; Sha, B.; et al. Trimethylamine N-Oxide Binds and Activates PERK to Promote Metabolic Dysfunction. *Cell. Metab.* **2019**, *30*, 1141–1151.e5. [\[CrossRef\]](http://doi.org/10.1016/j.cmet.2019.08.021)
- 97. Naghipour, S.; Cox, A.J.; Peart, J.N.; Du Toit, E.F.; Headrick, J.P. Trimethylamine N-oxide: Heart of the microbiota-CVD nexus? Nutr. *Res. Rev.* **2021**, *34*, 125–146. [\[CrossRef\]](http://doi.org/10.1017/S0954422420000177) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32718365)
- 98. Dehghan, P.; Farhangi, M.A.; Nikniaz, L.; Nikniaz, Z.; Asghari-Jafarabadi, M. Gut microbiota-derived metabolite trimethylamine N-oxide (TMAO) potentially increases the risk of obesity in adults: An exploratory systematic review and dose-response metaanalysis. *Obes. Rev.* **2020**, *21*, e12993. [\[CrossRef\]](http://doi.org/10.1111/obr.12993)
- 99. Zhuang, R.; Ge, X.; Han, L.; Yu, P.; Gong, X.; Meng, Q.; Zhang, Y.; Fan, H.; Zheng, L.; Liu, Z.; et al. Gut microbe-generated metabolite trimethylamine N-oxide and the risk of diabetes: A systematic review and dose-response meta-analysis. *Obes. Rev.* **2019**, *20*, 883–894. [\[CrossRef\]](http://doi.org/10.1111/obr.12843) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30868721)
- 100. Salguero, M.V.; Al-Obaide, M.A.I.; Singh, R.; Siepmann, T.; Vasylyeva, T.L. Dysbiosis of Gram-negative gut microbiota and the associated serum lipopolysaccharide exacerbates inflammation in type 2 diabetic patients with chronic kidney disease. *Exp. Ther. Med.* **2019**, *18*, 3461–3469. [\[CrossRef\]](http://doi.org/10.3892/etm.2019.7943)
- 101. Jayasudha, R.; Das, T.; Kalyana Chakravarthy, S.; Sai Prashanthi, G.; Bhargava, A.; Tyagi, M.; Rani, P.K.; Pappuru, R.R.; Shivaji, S. Gut mycobiomes are altered in people with type 2 Diabetes Mellitus and Diabetic Retinopathy. *PLoS ONE.* **2020**, *15*, e0243077. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0243077)
- 102. Xie, J.; Song, W.; Liang, X.; Zhang, Q.; Shi, Y.; Liu, W.; Shi, X. Protective effect of quercetin on streptozotocin-induced diabetic peripheral neuropathy rats through modulating gut microbiota and reactive oxygen species level. *Biomed. Pharmacother.* **2020**, *127*, 110147. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2020.110147) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32559841)
- 103. Zhang, Y.; Lu, S.; Yang, Y.; Wang, Z.; Wang, B.; Zhang, B.; Yu, J.; Lu, W.; Pan, M.; Zhao, J.; et al. The diversity of gut microbiota in type 2 diabetes with or without cognitive impairment. Aging Clin. *Exp. Res.* **2021**, *33*, 589–601.
- 104. Du, X.; Liu, J.; Xue, Y.; Kong, X.; Lv, C.; Li, Z.; Huang, Y.; Wang, B. Alteration of gut microbial profile in patients with diabetic nephropathy. *Endocrine* **2021**, *73*, 71–84. [\[CrossRef\]](http://doi.org/10.1007/s12020-021-02721-1)
- 105. Huang, Y.; Wang, Z.; Ma, H.; Ji, S.; Chen, Z.; Cui, Z.; Chen, J.; Tang, S. Dysbiosis and Implication of the Gut Microbiota in Diabetic Retinopathy. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 646348. [\[CrossRef\]](http://doi.org/10.3389/fcimb.2021.646348) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33816351)
- 106. Liu, W.; Wang, C.; Xia, Y.; Xia, W.; Liu, G.; Ren, C.; Gu, Y.; Li, X.; Lu, P. Elevated plasma trimethylamine-N-oxide levels are associated with diabetic retinopathy. *Acta Diabetol.* **2021**, *58*, 221–229. [\[CrossRef\]](http://doi.org/10.1007/s00592-020-01610-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33064205)
- 107. Yakar, B.; Onalan, E.; Kaymaz, T.; Donder, E.; Gursu, M.F. The role of trimethylamine-N-oxide level in the diagnosis of diabetic retinopathy and the differential diagnosis of diabetic and nondiabetic retinopathy. *Arq. Bras. Oftalmol.* **2022**. [\[CrossRef\]](http://doi.org/10.5935/0004-2749.2021-0527)
- 108. Pol, T.; Held, C.; Westerbergh, J.; Lindbäck, J.; Alexander, J.H.; Alings, M.; Erol, C.; Goto, S.; Halvorsen, S.; Huber, K.; et al. Dyslipidemia and risk of cardiovascular events in patients with atrial fibrillation treated with oral anticoagulation therapy: Insights from the ARISTOTLE (Apixaban for Reduction in Stroke and Other Thromboembolic Events in Atrial Fibrillation) Trial. *J. Am. Heart Assoc.* **2018**, *7*, e007444. [\[CrossRef\]](http://doi.org/10.1161/JAHA.117.007444)
- 109. Carroll, M.D.; Fryar, C.D.; Nguyen, D.T. Total and high-density lipoprotein cholesterol in adults: United States, 2015-2016. *NCHS Data Brief.* **2017**, *290*, 1–8.
- 110. Facchini, F.S.; Hollenbeck, C.B.; Jeppesen, J.; Chen, Y.D.; Reaven, G.M. Insulin resistance and cigarette smoking. *Lancet* **1992**, *339*, 1128–1130. [\[CrossRef\]](http://doi.org/10.1016/0140-6736(92)90730-Q)
- 111. Criqui, M.H.; Cowan, L.D.; Tyroler, H.A.; Bangdiwala, S.; Heiss, G.; Wallace, R.B.; Cohn, R. Lipoproteins as mediators for the effects of alcohol consumption and cigarette smoking on cardiovascular mortality: Results from the Lipid Research Clinics Follow-up Study. *Am. J. Epidemiol.* **1987**, *126*, 629–637. [\[CrossRef\]](http://doi.org/10.1093/oxfordjournals.aje.a114702) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/3631053)
- 112. Hubert, H.B.; Feinleib, M.; McNamara, P.M.; Castelli, W.P. Obesity as an independent risk factor for cardiovascular disease: A 26-year follow-up of participants in the Framingham Heart Study. *Circulation* **1983**, *67*, 968–977. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.67.5.968) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/6219830)
- 113. Zavaroni, I.; Dall'Aglio, E.; Alpi, O.; Bruschi, F.; Bonora, E.; Pezzarossa, A.; Butturini, U. Evidence for an independent relationship between plasma insulin and concentration of high-density lipoprotein cholesterol and triglyceride. *Atherosclerosis* **1985**, *55*, 259–266. [\[CrossRef\]](http://doi.org/10.1016/0021-9150(85)90105-4) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/3893447)
- 114. Garg, A.; Grundy, S.M. Nicotinic acid as therapy for dyslipidemia in non-insulin-dependent diabetes mellitus. *JAMA* **1990**, *264*, 723–726. [\[CrossRef\]](http://doi.org/10.1001/jama.1990.03450060069031)
- 115. Howard, B.V. Insulin resistance and lipid metabolism. *Am. J. Cardiol.* **1999**, *84*, 28J–32J. [\[CrossRef\]](http://doi.org/10.1016/S0002-9149(99)00355-0)
- 116. He, K.; Hu, Y.; Ma, H.; Zou, Z.; Xiao, Y.; Yang, Y.; Feng, M.; Li, X.; Ye, X. Rhizoma Coptidis alkaloids alleviate hyperlipidemia in B6 mice by modulating gut microbiota and bile acid pathways. *Biochim. Biophys. Acta* **2016**, *1862*, 1696–1709. [\[CrossRef\]](http://doi.org/10.1016/j.bbadis.2016.06.006) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27287254)
- 117. Gargari, G.; Deon, V.; Taverniti, V.; Gardana, C.; Denina, M.; Riso, P.; Guardamagna, O.; Guglielmetti, S. Evidence of dysbiosis in the intestinal microbial ecosystem of children and adolescents with primary hyperlipidemia and the potential role of regular hazelnut intake. *FEMS. Microbiol. Ecol.* **2018**, *94*, fiy045. [\[CrossRef\]](http://doi.org/10.1093/femsec/fiy045)
- 118. Moreno-Indias, I.; Sánchez-Alcoholado, L.; Pérez-Martínez, P.; Andrés-Lacueva, C.; Cardona, F.; Tinahones, F.; Queipo-Ortuño, M.I. Red wine polyphenols modulate fecal microbiota and reduce markers of the metabolic syndrome in obese patients. *Food Funct.* **2016**, *7*, 1775–1787. [\[CrossRef\]](http://doi.org/10.1039/C5FO00886G) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26599039)
- 119. Yamashita, H.; Fujisawa, K.; Ito, E.; Idei, S.; Kawaguchi, N.; Kimoto, M.; Hiemori, M.; Tsuji, H. Improvement of obesity and glucose tolerance by acetate in Type 2 diabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. *Biosci. Biotechnol. Biochem.* **2007**, *71*, 1236–1243. [\[CrossRef\]](http://doi.org/10.1271/bbb.60668)
- 120. De Vadder, F.; Kovatcheva-Datchary, P.; Goncalves, D.; Vinera, J.; Zitoun, C.; Duchampt, A.; Bäckhed, F.; Mithieux, G. Microbiotagenerated metabolites promote metabolic benefits via gut-brain neural circuits. *Cell* **2014**, *156*, 84–96. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2013.12.016)
- 121. Koh, A.; De Vadder, F.; Kovatcheva-Datchary, P.; Bäckhed, F. From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. *Cell* **2016**, *165*, 1332–1345. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2016.05.041) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27259147)
- 122. Tucureanu, M.M.; Rebleanu, D.; Constantinescu, C.A.; Deleanu, M.; Voicu, G.; Butoi, E.; Calin, M.; Manduteanu, I. Lipopolysaccharide-induced inflammation in monocytes/macrophages is blocked by liposomal delivery of Gi-protein inhibitor. *Int. J. Nanomed.* **2017**, *13*, 63–76. [\[CrossRef\]](http://doi.org/10.2147/IJN.S150918) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29317816)
- 123. Feihl, F.; Waeber, B.; Liaudet, L. Is nitric oxide overproduction the target of choice for the management of septic shock? Pharmacol. *Ther.* **2001**, *91*, 179–213.
- 124. Weigand, M.A.; Hörner, C.; Bardenheuer, H.J.; Bouchon, A. The systemic inflammatory response syndrome. *Best. Pract. Res. Clin. Anaesthesiol.* **2004**, *18*, 455–475. [\[CrossRef\]](http://doi.org/10.1016/j.bpa.2003.12.005)
- 125. Wang, H.; Xu, T.; Lewin, M.R. Future possibilities for the treatment of septic shock with herbal components. *Am. J. Emerg. Med.* **2009**, *27*, 107–112. [\[CrossRef\]](http://doi.org/10.1016/j.ajem.2008.08.003) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19041541)
- 126. Agerholm-Larsen, B.; Nordestgaard, B.G.; Steffensen, R.; Jensen, G.; Tybjaerg-Hansen, A. Elevated HDL cholesterol is a risk factor for ischemic heart disease in white women when caused by a common mutation in the cholesteryl ester transfer protein gene. *Circulation* **2000**, *101*, 1907–1912. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.101.16.1907) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10779455)
- 127. Qi, J.; You, T.; Li, J.; Pan, T.; Xiang, L.; Han, Y.; Zhu, L. Circulating trimethylamine N-oxide and the risk of cardiovascular diseases: A systematic review and meta-analysis of 11 prospective cohort studies. *J. Cell. Mol. Med.* **2018**, *22*, 185–194. [\[CrossRef\]](http://doi.org/10.1111/jcmm.13307)
- 128. Pathak, P.; Helsley, R.N.; Brown, A.L.; Buffa, J.A.; Choucair, I.; Nemet, I.; Gogonea, C.B.; Gogonea, V.; Wang, Z.; Garcia-Garcia, J.C.; et al. Small molecule inhibition of gut microbial choline trimethylamine lyase activity alters host cholesterol and bile acid metabolism. *Am. J. Physiol. Heart. Circ. Physiol.* **2020**, *318*, H1474–H1486. [\[CrossRef\]](http://doi.org/10.1152/ajpheart.00584.2019)
- 129. Dabke, K.; Hendrick, G.; Devkota, S. The gut microbiome and metabolic syndrome. *J. Clin. Investig.* **2019**, *129*, 4050–4057. [\[CrossRef\]](http://doi.org/10.1172/JCI129194)
- 130. Matey-Hernandez, M.L.; Williams, F.M.K.; Potter, T.; Valdes, A.M.; Spector, T.D.; Menni, C. Genetic and microbiome influence on lipid metabolism and dyslipidemia. *Physiol. Genom.* **2018**, *50*, 117–126. [\[CrossRef\]](http://doi.org/10.1152/physiolgenomics.00053.2017)
- 131. Kaska, L.; Sledzinski, T.; Chomiczewska, A.; Dettlaff-Pokora, A.; Swierczynski, J. Improved glucose metabolism following bariatric surgery is associated with increased circulating bile acid concentrations and remodeling of the gut microbiome. *World J. Gastroenterol.* **2016**, *22*, 8698–8719. [\[CrossRef\]](http://doi.org/10.3748/wjg.v22.i39.8698) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27818587)
- 132. Ryan, K.K.; Tremaroli, V.; Clemmensen, C.; Kovatcheva-Datchary, P.; Myronovych, A.; Karns, R.; Wilson-Pérez, H.E.; Sandoval, D.A.; Kohli, R.; Bäckhed, F.; et al. FXR is a molecular target for the effects of vertical sleeve gastrectomy. *Nature* **2014**, *509*, 183–188. [\[CrossRef\]](http://doi.org/10.1038/nature13135)
- 133. Lambert, G.; Amar, M.J.; Guo, G.; Brewer, H.B.; Gonzalez, F.J., Jr.; Sinal, C.J. The farnesoid X-receptor is an essential regulator of cholesterol homeostasis. *J. Biol. Chem.* **2003**, *278*, 2563–2570. [\[CrossRef\]](http://doi.org/10.1074/jbc.M209525200)
- 134. Sinal, C.J.; Tohkin, M.; Miyata, M.; Ward, J.M.; Lambert, G.; Gonzalez, F.J. Targeted disruption of the nuclear receptor FXR/BAR impairs bile acid and lipid homeostasis. *Cell* **2000**, *102*, 731–744. [\[CrossRef\]](http://doi.org/10.1016/S0092-8674(00)00062-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11030617)
- 135. Cariou, B.; van Harmelen, K.; Duran-Sandoval, D.; van Dijk, T.H.; Grefhorst, A.; Abdelkarim, M.; Caron, S.; Torpier, G.; Fruchart, J.C.; Gonzalez, F.J.; et al. The farnesoid X receptor modulates adiposity and peripheral insulin sensitivity in mice. *J. Biol. Chem.* **2006**, *281*, 11039–11049. [\[CrossRef\]](http://doi.org/10.1074/jbc.M510258200) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16446356)
- 136. Taniguchi, T.; Chen, J.; Cooper, A.D. Regulation of cholesterol 7 alpha-hydroxylase gene expression in Hep-G2 cells. Effect of serum, bile salts, and coordinate and noncoordinate regulation with other sterol-responsive genes. *J. Biol. Chem.* **1994**, *269*, 10071–10078. [\[CrossRef\]](http://doi.org/10.1016/S0021-9258(17)36991-0) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/8144506)
- 137. Nakahara, M.; Fujii, H.; Maloney, P.R.; Shimizu, M.; Sato, R. Bile acids enhance low density lipoprotein receptor gene expression via a MAPK cascade-mediated stabilization of mRNA. *J. Biol. Chem.* **2002**, *277*, 37229–37234. [\[CrossRef\]](http://doi.org/10.1074/jbc.M206749200) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12149270)
- 138. Langhi, C.; Le May, C.; Kourimate, S.; Caron, S.; Staels, B.; Krempf, M.; Costet, P.; Cariou, B. Activation of the farnesoid X receptor represses PCSK9 expression in human hepatocytes. *FEBS. Lett.* **2008**, *582*, 949–955. [\[CrossRef\]](http://doi.org/10.1016/j.febslet.2008.02.038)
- 139. Weir, H.K.; Anderson, R.N.; Coleman King, S.M.; Soman, A.; Thompson, T.D.; Hong, Y.; Moller, B.; Leadbetter, S. Heart disease and cancer deaths-trends and projections in the United States, 1969-2020. Prev. *Chronic. Dis.* **2016**, *13*, E157.
- 140. Johnson, N.B.; Hayes, L.D.; Brown, K.; Hoo, E.C.; Ethier, K.A.; Centers for Disease Control and Prevention (CDC). CDC National Health Report: Leading causes of morbidity and mortality and associated behavioral risk and protective factors-United States, 2005–2013. *MMWR Suppl.* **2014**, *63*, 3–27.
- 141. Xu, J.; Murphy, S.L.; Kochanek, K.D.; Arias, E. Mortality in the United States, 2015. *NCHS Data Brief.* **2016**, *267*, 1–8.
- 142. Rader, D.J.; Daugherty, A. Translating molecular discoveries into new therapies for atherosclerosis. *Nature* **2008**, *451*, 904–913. [\[CrossRef\]](http://doi.org/10.1038/nature06796)
- 143. Mancini, G.B.; Gosselin, G.; Chow, B.; Stone, J.; Yvorchuk, K.J.; Abramson, B.L.; Cartier, R.; Huckell, V.; Tardif, J.C.; Connelly, K.; et al. Canadian Cardiovascular Society guidelines for the diagnosis and management of stable ischemic heart disease. *Can. J. Cardiol.* **2014**, *30*, 837–849. [\[CrossRef\]](http://doi.org/10.1016/j.cjca.2014.05.013) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25064578)
- 144. Collet, J.P.; Thiele, H.; Barbato, E.; Barthélémy, O.; Bauersachs, J.; Bhatt, D.L.; Dendale, P.; Dorobantu, M.; Edvardsen, T.; Folliguet, T.; et al. 2020 ESC Guidelines for the management of acute coronary syndromes in patients presenting without persistent ST-segment elevation. *Eur. Heart J.* **2021**, *42*, 1289–1367. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehaa575)
- 145. Gulati, M.; Levy, P.D.; Mukherjee, D.; Amsterdam, E.; Bhatt, D.L.; Birtcher, K.K.; Blankstein, R.; Boyd, J.; Bullock-Palmer, R.P.; Conejo, T.; et al. 2021 AHA/ACC/ASE/CHEST/SAEM/SCCT/SCMR Guideline for the Evaluation and Diagnosis of Chest Pain: A Report of the American College of Cardiology/American Heart Association Joint Committee on Clinical Practice Guidelines Circulation. *J. Am. Coll. Cardiol.* **2021**, *144*, e368–e454.
- 146. O'Hara, A.M.; Shanahan, F. The gut flora as a forgotten organ. *EMBO. Rep.* **2006**, *7*, 688–693. [\[CrossRef\]](http://doi.org/10.1038/sj.embor.7400731)
- 147. Ott, S.J.; El Mokhtari, N.E.; Musfeldt, M.; Hellmig, S.; Freitag, S.; Rehman, A.; Kühbacher, T.; Nikolaus, S.; Namsolleck, P.; Blaut, M.; et al. Detection of diverse bacterial signatures in atherosclerotic lesions of patients with coronary heart disease. *Circulation* **2006**, *113*, 929–937. [\[CrossRef\]](http://doi.org/10.1161/CIRCULATIONAHA.105.579979)
- 148. Karlsson, F.H.; Fak, F.; Nookaew, I.; Tremaroli, V.; Fagerberg, B.; Petranovic, D.; Bäckhed, F.; Nielsen, J. Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat. Commun.* **2012**, *3*, 1245. [\[CrossRef\]](http://doi.org/10.1038/ncomms2266) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23212374)
- 149. Tang, W.H.; Wang, Z.; Levison, B.S.; Koeth, R.A.; Britt, E.B.; Fu, X.; Wu, Y.; Hazen, S.L. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N. Engl. J. Med.* **2013**, *368*, 1575–1584. [\[CrossRef\]](http://doi.org/10.1056/NEJMoa1109400) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23614584)
- 150. Fu, Q.; Zhao, M.; Wang, D.; Hu, H.; Guo, C.; Chen, W.; Li, Q.; Zheng, L.; Chen, B. Coronary Plaque Characterization Assessed by Optical Coherence Tomography and Plasma Trimethylamine-N-oxide Levels in Patients With Coronary Artery Disease. *Am. J. Cardiol.* **2016**, *118*, 1311–1315. [\[CrossRef\]](http://doi.org/10.1016/j.amjcard.2016.07.071)
- 151. Zeisel, S.H.; Warrier, M. Trimethylamine N-Oxide, the Microbiome, and Heart and Kidney Disease. *Annu. Rev. Nutr.* **2017**, *37*, 157–181. [\[CrossRef\]](http://doi.org/10.1146/annurev-nutr-071816-064732)
- 152. Wolf, D.; Ley, K. Immunity and Inflammation in Atherosclerosis. *Circ. Res.* **2019**, *124*, 315–327. [\[CrossRef\]](http://doi.org/10.1161/CIRCRESAHA.118.313591)
- 153. Park, Y.M. CD36, a scavenger receptor implicated in atherosclerosis. *Exp. Mol. Med.* **2014**, *46*, e99. [\[CrossRef\]](http://doi.org/10.1038/emm.2014.38)
- 154. Ohashi, R.; Mu, H.; Wang, X.; Yao, Q.; Chen, C. Reverse cholesterol transport and cholesterol efflux in atherosclerosis. *QJM.* **2005**, *98*, 845–856. [\[CrossRef\]](http://doi.org/10.1093/qjmed/hci136)
- 155. Charach, G.; Rabinovich, A.; Argov, O.; Weintraub, M.; Rabinovich, P. The role of bile acid excretion in atherosclerotic coronary artery disease. *Int. J. Vasc. Med.* **2012**, *2012*, 949672. [\[CrossRef\]](http://doi.org/10.1155/2012/949672) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21918722)
- 156. Lu, Y.; Feskens, E.J.; Boer, J.M.; Müller, M. The potential influence of genetic variants in genes along bile acid and bile metabolic pathway on blood cholesterol levels in the population. *Atherosclerosis* **2010**, *210*, 14–27. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2009.10.035)
- 157. Zheng, Y.; He, J.Q. Pathogenic Mechanisms of Trimethylamine N-Oxide-induced Atherosclerosis and Cardiomyopathy. *Curr. Vasc. Pharmacol.* **2022**, *20*, 29–36. [\[CrossRef\]](http://doi.org/10.2174/1570161119666210812152802) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34387163)
- 158. Seldin, M.M.; Meng, Y.; Qi, H.; Zhu, W.; Wang, Z.; Hazen, S.L.; Lusis, A.J.; Shih, D.M. Trimethylamine N-Oxide Promotes Vascular Inflammation Through Signaling of Mitogen-Activated Protein Kinase and Nuclear Factor-κB. J. *Am. Heart Assoc.* **2016**, *5*, e002767. [\[CrossRef\]](http://doi.org/10.1161/JAHA.115.002767) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26903003)
- 159. Ma, G.; Pan, B.; Chen, Y.; Guo, C.; Zhao, M.; Zheng, L.; Chen, B. Trimethylamine N-oxide in atherogenesis: Impairing endothelial self-repair capacity and enhancing monocyte adhesion. *Biosci. Rep.* **2017**, *37*, BSR20160244. [\[CrossRef\]](http://doi.org/10.1042/BSR20160244) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28153917)
- 160. Durpès, M.C.; Morin, C.; Paquin-Veillet, J.; Beland, R.; Paré, M.; Guimond, M.O.; Rekhter, M.; King, G.L.; Geraldes, P. PKC-β activation inhibits IL-18-binding protein causing endothelial dysfunction and diabetic atherosclerosis. *Cardiovasc. Res.* **2015**, *106*, 303–313. [\[CrossRef\]](http://doi.org/10.1093/cvr/cvv107)
- 161. Zhou, X.; Chen, M.; Zeng, X.; Yang, J.; Deng, H.; Yi, L.; Mi, M.T. Resveratrol regulates mitochondrial reactive oxygen species homeostasis through Sirt3 signaling pathway in human vascular endothelial cells. *Cell. Death. Dis.* **2014**, *5*, e1576. [\[CrossRef\]](http://doi.org/10.1038/cddis.2014.530)
- 162. Chen, M.L.; Zhu, X.H.; Ran, L.; Lang, H.D.; Yi, L.; Mi, M.T. Trimethylamine-N-Oxide Induces Vascular Inflammation by Activating the NLRP3 Inflammasome Through the SIRT3-SOD2-mtROS Signaling Pathway. *J. Am. Heart Assoc.* **2017**, *6*, e006347. [\[CrossRef\]](http://doi.org/10.1161/JAHA.117.006347)
- 163. Ray, P.D.; Huang, B.W.; Tsuji, Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell. Signal.* **2012**, *24*, 981–990. [\[CrossRef\]](http://doi.org/10.1016/j.cellsig.2012.01.008)
- 164. Chou, R.H.; Chen, C.Y.; Chen, I.C.; Huang, H.L.; Lu, Y.W.; Kuo, C.S.; Chang, C.C.; Huang, P.H.; Chen, J.W.; Lin, S.J. Trimethylamine N-Oxide, Circulating Endothelial Progenitor Cells, and Endothelial Function in Patients with Stable Angina. *Sci. Rep.* **2019**, *9*, 4249. [\[CrossRef\]](http://doi.org/10.1038/s41598-019-40638-y)
- 165. Toya, T.; Ozcan, I.; Corban, M.T.; Sara, J.D.; Marietta, E.V.; Ahmad, A.; Horwath, I.E.; Loeffler, D.L.; Murray, J.A.; Lerman, L.O.; et al. Compositional change of gut microbiome and osteocalcin expressing endothelial progenitor cells in patients with coronary artery disease. *PLoS ONE* **2021**, *16*, e0249187. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0249187)
- 166. Michowitz, Y.; Goldstein, E.; Wexler, D.; Sheps, D.; Keren, G.; George, J. Circulating endothelial progenitor cells and clinical outcome in patients with congestive heart failure. *Heart* **2007**, *93*, 1046–1050. [\[CrossRef\]](http://doi.org/10.1136/hrt.2006.102657)
- 167. Rauscher, F.M.; Goldschmidt-Clermont, P.J.; Davis, B.H.; Wang, T.; Gregg, D.; Ramaswami, P.; Pippen, A.M.; Annex, B.H.; Dong, C.; Taylor, D.A. Aging, progenitor cell exhaustion, and atherosclerosis. *Circulation.* **2003**, *108*, 457–463. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.0000082924.75945.48)
- 168. Shen, X.; Li, L.; Sun, Z.; Zang, G.; Zhang, L.; Shao, C.; Wang, Z. Gut Microbiota and Atherosclerosis-Focusing on the Plaque Stability. *Front. Cardiovasc. Med.* **2021**, *8*, 668532. [\[CrossRef\]](http://doi.org/10.3389/fcvm.2021.668532)
- 169. Peng, L.; Li, Z.R.; Green, R.S.; Holzman, I.R.; Lin, J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. *J. Nutr.* **2009**, *139*, 1619–1625. [\[CrossRef\]](http://doi.org/10.3945/jn.109.104638)
- 170. Moreira, A.P.B.; Texeira, T.F.S.; Ferreira, A.B.; Peluzio, M.; Peluzio, M.D.C.G.; Alfenas, R.D.C.G. Influence of a high-fat diet on gut microbiota, intestinal permeability and metabolic endotoxaemia. *Br. J. Nutr.* **2012**, *108*, 801–809. [\[CrossRef\]](http://doi.org/10.1017/S0007114512001213)
- 171. Sturm, A.; Dignass, A.U. Epithelial restitution and wound healing in inflammatory bowel disease. World, J. *Gastroenterol.* **2008**, *14*, 348–353.
- 172. Ulevitch, R.J.; Tobias, P.S. Receptor-dependent mechanisms of cell stimulation by bacterial endotoxin. *Annu. Rev. Immunol.* **1995**, *13*, 437–457. [\[CrossRef\]](http://doi.org/10.1146/annurev.iy.13.040195.002253)
- 173. Marumo, T.; Schini-Kerth, V.B.; Fisslthaler, B.; Busse, R. Platelet-derived growth factor-stimulated superoxide anion production modulates activation of transcription factor NF-kappaB and expression of monocyte chemoattractant protein 1 in human aortic smooth muscle cells. *Circulation.* **1997**, *96*, 2361–2367. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.96.7.2361)
- 174. Sawa, Y.; Ueki, T.; Hata, M.; Iwasawa, K.; Tsuruga, E.; Kojima, H.; Ishikawa, H.; Yoshida, S. LPS-induced IL-6, IL-8, VCAM-1, and ICAM-1 expression in human lymphatic endothelium. *J. Histochem. Cytochem.* **2008**, *56*, 97–109. [\[CrossRef\]](http://doi.org/10.1369/jhc.7A7299.2007)
- 175. Kim, S.J.; Park, J.H.; Kim, K.H.; Lee, W.R.; Pak, S.C.; Han, S.M.; Park, K.K. The Protective Effect of Apamin on LPS/Fat-Induced Atherosclerotic Mice. *Evid. Based Complement. Alternat. Med.* **2012**, *2012*, 305454. [\[CrossRef\]](http://doi.org/10.1155/2012/305454) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22645626)
- 176. Hanniman, E.A.; Lambert, G.; McCarthy, T.C.; Sinal, C.J. Loss of functional farnesoid X receptor increases atherosclerotic lesions in apolipoprotein E-deficient mice. *J. Lipid. Res.* **2005**, *46*, 2595–2604. [\[CrossRef\]](http://doi.org/10.1194/jlr.M500390-JLR200)
- 177. Macfarlane, G.T.; Macfarlane, S. Bacteria, colonic fermentation, and gastrointestinal health. *J. AOAC. Int.* **2012**, *95*, 50–60. [\[CrossRef\]](http://doi.org/10.5740/jaoacint.SGE_Macfarlane)
- 178. Trompette, A.; Gollwitzer, E.S.; Yadava, K.; Sichelstiel, A.K.; Sprenger, N.; Ngom-Bru, C.; Blanchard, C.; Junt, T.; Nicod, L.P.; Harris, N.L.; et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat. Med.* **2014**, *20*, 159–166. [\[CrossRef\]](http://doi.org/10.1038/nm.3444) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24390308)
- 179. Howarth, N.C.; Saltzman, E.; Roberts, S.B. Dietary fiber and weight regulation. *Nutr. Rev.* **2001**, *59*, 129–139. [\[CrossRef\]](http://doi.org/10.1111/j.1753-4887.2001.tb07001.x)
- 180. Silva, F.M.; Kramer, C.K.; de Almeida, J.C.; Steemburgo, T.; Gross, J.L.; Azevedo, M.J. Fiber intake and glycemic control in patients with type 2 diabetes mellitus: A systematic review with meta-analysis of randomized controlled trials. *Nutr. Rev.* **2013**, *71*, 790–801. [\[CrossRef\]](http://doi.org/10.1111/nure.12076) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24180564)
- 181. Aguilar, E.C.; Leonel, A.J.; Teixeira, L.G.; Silva, A.R.; Silva, J.F.; Pelaez, J.M.; Capettini, L.S.; Lemos, V.S.; Santos, R.A.; Alvarez-Leite, J.I. Butyrate impairs atherogenesis by reducing plaque inflammation and vulnerability and decreasing NFκB activation. *Nutr. Metab. Cardiovasc. Dis.* **2014**, *24*, 606–613. [\[CrossRef\]](http://doi.org/10.1016/j.numecd.2014.01.002)
- 182. Aguilar, E.C.; Santos, L.C.; Leonel, A.J.; de Oliveira, J.S.; Santos, E.A.; Navia-Pelaez, J.M.; da Silva, J.F.; Mendes, B.P.; Capettini, L.S.; Teixeira, L.G.; et al. Oral butyrate reduces oxidative stress in atherosclerotic lesion sites by a mechanism involving NADPH oxidase down-regulation in endothelial cells. *J. Nutr. Biochem.* **2016**, *34*, 99–105. [\[CrossRef\]](http://doi.org/10.1016/j.jnutbio.2016.05.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27261536)
- 183. Gagné, M.A.; Barbeau, C.; Frégeau, G.; Gilbert, K.; Mathieu, O.; Auger, J.; Tompkins, T.A.; Charbonney, E.; Godbout, R.; Rousseau, G. Dysbiotic microbiota contributes to the extent of acute myocardial infarction in rats. *Sci. Rep.* **2022**, *12*, 16517. [\[CrossRef\]](http://doi.org/10.1038/s41598-022-20826-z) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36192578)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.