Increased excretion of soluble interleukin 2 receptors and free light chain immunoglobulins in the urine of patients with active lupus nephritis

Chang-Youh Tsai, Tsai-Hung Wu, Kuang-Hui Sun, Whu-Mei Lin, Chia-Li Yu

Abstract

Samples of protein from the urine of 23 patients with lupus nephropathy and 15 patients with proteinuria who did not have systemic lupus erythematosus (SLE) were studied for the presence of cytokines, soluble interleukin 2 receptors (sIL-2R), and free light chain immunoglobulins. The patients with lupus nephropathy were divided into two groups with active (nephritis) and inactive inflammation (nephrosis) based on the results of the analysis of urine samples and renal histology. The crude urine proteins (5 mg/ml) after precipitation by 80% ammonium sulphate from 14 patients with lupus nephritis contained higher concentrations of sIL-2R (4.88 (SEM 1.27 ng/ml) than those from nine patients with nephrosis (1.11 (0.52) ng/ml) or 15 patients without SLE (1.31 (0.87) ng/ml). The concentration of sIL-2R in protein from urine samples was not correlated with the concentration in plasma and was inversely correlated with the excretion of protein in urine over 24 hours in patients with SLE. It is suggested that, in addition to leakage from the circulation, the local production of sIL-2R by inflamed kidneys is possible. The crude proteins in urine were further fractionated by gel filtration on Sephacryl S-200. Arbitrarily, four fractions could be obtained from urine from patients with SLE but only three fractions were found in the urine of patients without SLE. Fraction IV derived from patients with nephritis or nephrosis augmented the pokeweed mitogen induced [³H]thymidine uptake of mononuclear cells. In addition, the positive rates of free kappa (\varkappa) (35.7%) and lambda (λ) (42.9%) chains in proteins in urine from nephritic patients were higher than those in the other two groups. These results suggest that the severity of inflammation in the kidneys of patients with lupus can be reflected by the increased excretion of sIL-2R, free light chain immunoglobulins, and cytokine-like molecules in urine.

Lupus nephritis has long been regarded as a prototype of immune complex mediated inflammatory reactions. Many investigators¹⁻³ have shown that the infiltrates of monocytes, T and B lymphocytes, and polymorphonuclear leucocytes in the glomeruli play an important role in murine and human glomerulonephritis. Cytokines and other inflammatory mediators secreted by these infiltrating mononuclear and mesangial cells^{4 5} are believed to initiate the inflammation and ultimately to lead to tissue

necrosis of the kidney. Using Northern blot analysis, Boswell *et al*^{6 7} found that expression of tumour necrosis factor (TNF) and interleukin 1 β (IL-1 β) mRNA were enhanced in glomerular macrophages in the lupus prone (MRL/1pr) mouse during the course of the disease. Noble *et al*⁸ found that the urinary excretion of cytokines, including IL-1 β and TNF, might reflect the severity of renal inflammation in experimental immune complex induced glomerulonephritis. However, there are very few reports on the amount of cytokines in the proteins from urine of patients with lupus nephritis.

Hopper and Papagiannes⁹ showed a significant amount of unbound polyclonal light chain immunoglobulins in the culture supernatant of mitogen stimulated normal human mononuclear cells. Epstein¹⁰ and Hopper *et al*¹¹ further reported that clinical relapse of patients with systemic lupus erythematosus (SLE) was correlated with an antecedent increase in free light chain immunoglobulins in urine. The interrelation between the increased free light chains in urine and the severity of lupus nephritis in humans has not yet been clearly shown.

In this study, we compared the concentrations of cytokines, cytokine inhibitor (soluble interleukin 2 receptor (sIL-2R)), free light chain immunoglobulins and immunological activity of proteins in urine from patients with and without SLE to evaluate more precisely the severity of inflammation and to understand further the immunological changes in patients with lupus nephritis.

Patients and methods

PATIENTS AND CONTROLS

Twenty three urine samples from patients fulfilling the 1982 revised criteria of the American Rheumatism Association for the classification of SLE were studied. These patients (19 women and four men with a mean age of 31.7 (SEM 11.4) years) were further divided into two groups according to the results of urine analysis and renal histology. Fourteen patients were classified as having active lupus nephritis (nephritis group) owing to significantly increased white and red blood cell counts, casts in the urinary sediments and hypercellularity with mononuclear cell infiltration in the glomeruli. Nine patients were classified as having inactive lupus nephritis (nephrosis group) with the presence of normal urinary sediments and normocellularity with scanty mononuclear cell infiltration in the renal glomeruli. Fifteen

Immunology, and Rheumatology, Department of Medicine, Veterans General Hospital Taipei, 201 Section 2, Shih-Pai Road, Taipei, Taiwan 11217, **Republic of China** C-Y Tsai W-M Lin C-L Yu Section of Nephrology, Department of Medicine, Veterans General Hospital Taipei, 201 Section 2, Shih-Pai Road, Taipei, Taiwan 11217, Republic of China T-H Wu School of Medical Technology, National Yang-Ming Medical College, 201 Section 2, Shih-Pai Road, Taipei, Taiwan 11217 Republic of China K-H Sun

Section of Allergy,

Correspondence to: Dr Chia-Li Yu. Accepted for publication 18 March 1991 patients without SLE with proteinuria (five with diabetic nephropathy, four with hypertensive nephrosclerosis, three with idiopathic membranous glomerulonephritis, and three with minimal nephrotic syndrome) served as controls. Urine was collected throughout a 24 hour period in the presence of 0.01% sodium azide as a preservative.

COLLECTION OF URINE PROTEIN

The urine sample was initially precipitated with 80% ammonium sulphate followed by centrifugation at 10 000 g for 30 minutes.¹² ¹³ The pellet was constituted with 50 mM TRIS buffer in 100 mM NaCl, pH 7.5. The solution was then intensively dialysed against at least 100 volumes of the same buffer for 48 hours with several changes. After dialysis, the protein solution was centrifuged at 300 g for 10 minutes and the clear supernatant (crude urine protein) was collected. The concentration of the crude urine protein was determined by the Bio-Rad protein assay kit (Bio-Rad Laboratories, Richmond, CA, USA). For further purification, the crude urine protein was fractionated by eluting with 0.15 M TRIS buffer (pH 7.5) through an 80×2.5 cm Sephacryl S-200 gel filtration column at a flow rate of 20 ml/hour. Four millilitres of eluent were collected in each tube. The different fractions were pooled, dialysed and concentrated.

DETERMINATION OF sIL-2R, TNF- α AND IL-6 IN CRUDE PROTEINS FROM URINE

Commercially available enzyme linked immunosorbent assay (ELISA) kits were used for the detection of sIL-2R (AMAC, Westbrook, Maine, USA), TNF- α (T cell · Sciences, Cambridge, MA, USA) and IL-6 (Genzyme, Boston, MA, USA). The detailed procedures are described by the manufacturers. Concentrations of TNF- α and IL-6 higher than 4.8 pg/ml or 0.16 ng/ml, respectively, in 5 mg/ml protein from urine were considered positive in this study.

QUANTITATION OF FREE KAPPA AND LAMBDA LIGHT CHAIN IMMUNOGLOBULINS IN CRUDE PROTEIN FROM URINE

As free light chain specific antisera were not

available, urinary free light chain immunoglobulin was determined indirectly by calculation. The concentrations of IgG, IgA, IgM, and total light chains in crude protein in urine (5 mg/ml) were measured by laser nephelometry (Hoechst-Behringwerke, Marburg, Germany). The free kappa (\varkappa) and lambda (λ) light chain immunoglobulins were then calculated according to the following equations: (A) total Ig(mg/l)=IgG+ IgA+IgM; (B) calculated bound \varkappa chain=total Ig $\times 2/3$, calculated bound λ chain=total Ig $\times 1/3$; (C) free light chains, free \varkappa chain= (measured \varkappa -calculated bound \varkappa) \times l/6 (or 17%), free λ chain=(measured λ -calculated bound λ) \times l/6 (or 17%).

DETERMINATION OF POKEWEED MITOGEN ENHANCING ACTIVITY OF PROTEIN IN URINE ON NORMAL MONONUCLEAR CELLS

Mononuclear cells were isolated from heparinised venous blood after centrifugation at 300 g for 30 minutes in a Ficoll-Hypaque density gradient (specific gravity 1.077). One hundred microlitres of mononuclear cells $(1 \times 10^{6}/\text{ml})$ were placed in flat bottomed microwells in triplicate. Twenty microlitres of pokeweed mitogen (100 µg/ml), 40 µl of various fractions of urine protein (0.2 mg/ml), human IgG (0.2 mg/ml), or RPMI 1640 medium, and 40 µl of 10% fetal bovine serum in RPMI medium were added to the wells to give a total volume of 200 µl in each well. The mixtures were incubated at 37°C in 5% carbon dioxide/95% air for 68 hours. The cells were then pulsed with 18.5 kBq methyl[³H]thymidine (specific activity 248 G Bq/mmol; NEN, Boston) per well for four hours. The cells were harvested and the radioactivity was measured by a beta counter. The percentage of pokeweed mitogen (PWM) enhancement was calculated by the following formula using the counts per minute (cpm):

%PWM enhancing =
$$\left(\frac{\text{cpm with samples-cpm with medium}}{\text{cpm with medium}}\right) \times 100$$

ELECTROPHORETIC ANALYSIS OF CRUDE PROTEINS FROM URINE AND THEIR ELUTION FRACTIONS

Ten microlitres of various samples of protein from urine (protein concentration 2 mg/ml)

Table I	Clinical, laboratory	, and renal histo	logical findin	gs in patient	s with active	lupus nephritis
---------	----------------------	-------------------	----------------	---------------	---------------	-----------------

Patient No	Sex	Age (years)	24 Hour urine protein excretion (g)	Urinary sediments (/HPF)			Renal	C3c	C4	Antibody to
				Red blood cells (n)	White blood cells (n)	Casts	pathology*	(mg / l)	(mg / l)	dsDNA (IU/ml)
1	м	39	0.74	6-8	10-15	Hvaline	IV	40	211	89.4
2	F	30	3.1	5-10	15-25	Granular	IV	279	76	15.8
3	F	42	12.0	0-4	5-10	Hyaline	III	846	364	40.5
4	F	39	0.19	0-4	10-30		II	837	328	28.6
5	F	24	0.22	10-15	20-35	Granular	IV	266	41	2100
6	F	34	2.5	0-4	10-15	_	Ш	830	309	36.7
7	F	16	8·64	20-40	10-20	Hyaline	IV	401	151	214
8	F	28	8 ∙05	2040	25–50	Hyaline,	ĪV	135	48	962
9	F	17	0.7	0-4	5 10	granular				
10	L L	45	1.15	5-10	5-10		II	603	198	36.0
11	Г Б	24	1.13	10-20	5-10		III	597	218	1259
12	r E	38	0.33	10-20	20-40	Hyaline	IV	258	91	1054
13	M	26	14.6		15-30	Hyaline	III	304	98	126
13	F	20	0.86	0-5	6-15	2	II.	209	116	1575
17	Τ.	24	0.90	58	1035	Granular	IV	409	105	326

*According to the World Health Organisation classification of lupus nephritis.

STATISTICAL ANALYSIS

The results were represented as the mean (SEM) in the whole study. The statistical significance was assessed by the non-parametric Wilcoxon signed rank test or Fisher's exact test.

Results

DEMOGRAPHIC AND LABORATORY ASSESSMENT OF THE PATIENTS WITH SLE

Tables 1 and 2 show the clinical and laboratory data for the patients with lupus nephropathy. The inflammatory activity of the kidney was mainly judged by the urinary sediments and renal histology. The changes in the urine sediments and renal biopsy were more active in the nephritis group than those in the nephrosis group. However, the serum abnormalities were the same in the two groups. All of the patients in the nephrotic group showed hypercholesterolaemia (>9.1 mmol/l) and decreased serum albumin (<30 g/l). In addition, the 24 hour excretion of urine protein in the nephritis group (3.87 (4.86) g) was less than that of the nephrosis group (11.40 (7.67) g), but was not different from that of the group without SLE (6.51 (4.43) g). Two patients with positive antinuclear antibodies (1/160 diffuse and 1/320 diffuse) in serum and five patients with increased white blood cells (>five cells/high powered field) in the urine were also found in the group without SLE.

SOLUBLE IL-2 RECEPTOR IN CRUDE PROTEINS FROM URINE

The concentration of sIL-2R in crude proteins

from urine (5 mg/ml) in the nephritis group (4.88 (1.27) ng/ml) was significantly higher than in the nephrosis group (1.11 (0.52) ng/ml) or the group without SLE (1.31 (0.87) ng/ml) as determined by a commercial ELISA kit (fig 1). To determine if the high level of urinary sIL-2R in the nephritis group was due to local production by the inflamed kidney, we compared the concentrations of sIL-2R in plasma and urine in these patients, but no correlation could be found (data not shown). In addition, the sIL-2R concentration in protein from urine was inversely correlated with the 24 hour protein excretion in patients with SLE (fig 2). It appears that local production of sIL-2R in the active inflamed kidney is possible.

PRESENCE OF FREE LIGHT CHAIN

IMMUNOGLOBULINS, TNF- α and IL-6 in CRUDE PROTEINS FROM URINE

Table 3 shows that the positive rate of free kappa (35.7%) and free lambda (42.9%) chains in proteins from nephritic urine (5 mg/ml) was significantly higher than that in the other two groups. As only a few samples of protein from

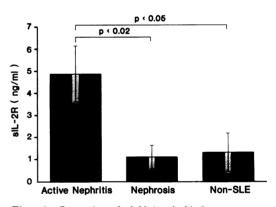


Figure 1 Comparison of soluble interleukin 2 receptor (sIL-2R) concentration in proteins from urine (5 mg/ml) of patients with active lupus nephritis, inactive lupus nephritis (nephrosis), and nephropathy without SLE (non-SLE).

Table 2	Clinical, la	aboratory and r	enal histological	findings in patients v	with inactive lupus nephritis	
---------	--------------	-----------------	-------------------	------------------------	-------------------------------	--

Patient No	Sex	Age (years)	24 Hour urine protein excretion (g)	Urinary sediments (/HPF)			Renal	C3c	C4	Antibody to dsDNA
				<i>RBC</i> (<i>n</i>)	WBC (n)	Cast	pathology*	(mg / l)	(mg / l)	(IU/ml)
1	F	34	8.68	0-2	0-5	_	III	398	210	17.8
2	Ē	24	7.55	0-5	0-5	_	IV	218	172	105.4
3	Ê	17	7.94	0-5	0-5		IV	198	127	215.5
4	M	64	25.6	0-2	0-2	Hyaline	I	852	654	<8
Ś	F	30	10.0	1-5	0-2	_	v	158	43	48 ·7
6	F	42	8.64	0-5	0-5	_	V	415	157	36.5
7	Ē	18	7.49	0-3	0-2		III	319	80	63.5
8	F	43	23.42	0-2	0-3		II	459	140	44·7
9	М	30	3.3	0-2	0-5	_	IV	923	442	<8

*According to the WHO classification of lupus nephritis.

Table 3 Presence of free light chains, TNF-a, and IL-6 in crude proteins in urine from patients with active and inactive lupus nephritis and nephropathy without SLE. Results are shown as number (percentage)

Disease group	Free light chains		TNF-α	IL-6	
	Kappa	Lambda			
Active lupus nephritis Inactive lupus nephritis Without SLE	5/14 (36) 0/9* (0) 0/15† (0)	6/14 (43) 0/9* (0) 0/15† (0)	2/14 (14) 1/9 (11) 0/15 (0)	3/14 (21) 1/9 (11) 0/15 (0)	

*Compared with patients with active lupus nephritis, p=0.0036 by Fisher's exact test. †Compared with patients with active lupus nephritis, p=0.0297 by Fisher's exact test. urine contained TNF- α (two of 14 in the nephritis group, one of nine in the nephrosis group) and IL-6 (three of 14 in the nephritis group, one of nine in the nephrosis group), the differences among these three groups were not significant.

SEPHACRYL S-200 FRACTIONATION OF PROTEIN FROM URINE

Figure 3 shows a typical chromatogram for protein in urine from a patient with lupus nephritis (patient 14) after Sephacryl S-200 gel filtration. Arbitrarily, four fractions could be recognised in proteins from urine of nephritic or nephrotic patients, whereas only the first three fractions were seen in samples from patients without SLE. Fractions I, II, and III represent macromolecules (IgM and/or immune complexes), IgG, and albumin, respectively. Fraction IV is presumed to contain cytokines or

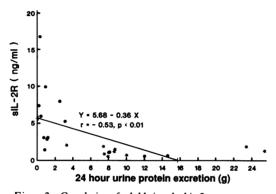


Figure 2 Correlation of soluble interleukin 2 receptor (sIL-2R) concentration and 24 hour urine protein excretion in patients with active (\bullet) and inactive (\star) lupus nephritis.

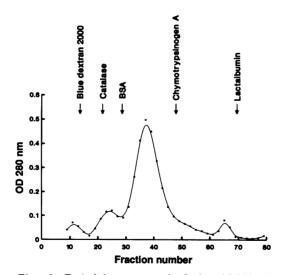


Figure 3 Typical chromatogram after Sephacryl S-200 gel filtration of protein in urine from the patient with active lupus nephritis (patient 14 in the nephritis group).

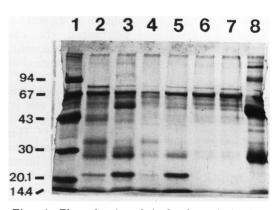


Figure 4 Electrophoretic analysis of crude proteins in urine and the fraction IV molecule from patients with active and inactive lupus nephritis using 10% sodium dodecyl sulphate-polyacrylamide gel elecytrophoresis. Lane 1, calibration molecules; lane 2 fraction IV from patients with inactive nephritis (sample 1); lane 3, fraction IV from patients with active nephritis (sample I); lane 4, fraction IV from patients with inactive nephritis (sample II); lane 5, fraction IV from patients with active nephritis (sample II); lane 6, crude proteins in urine from patients with active nephritis; lane 7, crude proteins in urine from patients with inactive nephritis; lane 8 normal human IgG.

cytokine related molecules. These fractions were pooled, dialysed and sterilised for determining the immunological activity. At a concentration of 40 μ g/ml, fraction IV from urine from the nephritic and nephrotic groups contained potent pokeweed mitogen enhancing activity on normal human mononuclear cells (table 4) but fractions I, II, and III were inactive. The estimated molecular weight of fraction IV determined under reducing condition by 10% sodium dodecyl sulphate-polyacrylamide gel electrophoresis was about 23 kilodaltons (fig 4).

Discussion

We analysed and compared the composition of proteins in urine from patients with and without SLE and found that increased sIL-2R and free light chain immunoglobulin concentrations in urine might reflect the severity of inflammation in the lupus kidney. Although certain cytokines such as IL-1,⁶ ¹⁴ TNF,⁷ IL-6,¹⁵ and transform-ing growth factor- β^{16} may play a role in the pathogenesis of glomerular damage, we could only find a few samples containing TNF- α (7.9%) or IL-6 (10.5%) in our 38 samples of proteins from urine. Noble et al⁸ reported that the number of mononuclear cells in glomeruli was well correlated with the amount of urinary protein excretion and the concentrations of IL-1 β and TNF- α in urine in experimental proliferative immune complex glomerulonephritis. The inconsistency between our results and other reports is a result of (a) different disease (lupus nephropathy v immune complex glomerulonephritis), **(b)** different species

Table 4 Pokeweed mitogen (PWM) enhancing activity of fractions III and IV (40 μ g/ml) derived from proteins in urine of patients with active and inactive lupus nephritis

	Human IgG (40 μg/ml)	Active nephritis		Inactive nephritis	5
		Fraction III	Fraction IV	Fraction III	Fraction IV
Mean (SEM) PWM enhancing activity (%) p Value*	1.2 (1.3)	10·1 (7·7) 0·051	85·8 (10·5) 0·0004	1·7 (0·5) 0·31	58·4 (8·9) 0·001

*Calculated by Wilcoxon signed rank test.

(human v rat), and (c) a different method, as we determined the cytokine concentration in proteins concentrated to 5 mg/ml in urine rather than in the original urine sample.

The soluble IL-2 receptor is a cytokine binder released mainly from activated T lymphocytes¹⁷ and is found at increased concentrations in serum samples from patients with active SLE.¹⁸ ¹⁹ This soluble peptide is also present in the urine of normal subjects²⁰ and its concentration is increased in patients with adult T cell leukaemia.²¹ In this study, we found that sIL-2R was also increased in the proteins from urine of patients with active lupus nephritis compared with patients with inactive nephropathy or without SLE. However, this increment was not parallel with the concentration in serum and inversely correlated with the 24 hour protein excretion. It appears that sIL-2R in urine may not all come from the circulation through damaged glomeruli. Local production by infiltrating mononuclear or mesangial cells in the inflamed kidney is possible as these cells can actively secrete immune mediators such as IL-1, TNF- α , IL-6, TGF- β^{67} ¹⁴⁻¹⁶ or eicosanoid metabolites²² to exert nephrotoxic effects on renal parenchymal tissue. Werber $et al^{14}$ and Boswell and coworkers⁶⁷ showed that the renal cortical cells or macrophages from autoimmune MRL/1pr mice with nephritis contained several times more IL-1 β and TNF mRNA compared with normal mice. Accordingly, the direct determination of cytokines or related molecules in protein from urine may provide a reliable method of evaluating the severity of lupus nephritis. Although we did not find a significant difference in the IL-6 and TNF- α concentrations among the three groups, we did note a 23 kilodalton molecule with IL-4 like activity in urine from nephritic and nephrotic patients, but not in urine from patients without SLE. The properties of this pokeweed mitogen enhancing factor were not defined in this study.

Another interesting finding in this study was the presence of free light chain immunoglobulins in six samples of protein from urine from the nephritis group. The concentrations of urinary sIL-2R and serum titres of antibody to dsDNA were also fairly high in these patients. Epstein¹⁰ and Hopper and coworkers⁹ ¹¹ clearly showed that the presence of urinary free light chain immunoglobulins preceded clinical exacerbation of SLE. Turnover studies in humans by Waldmann et al^{23} showed that only 60% of newly synthesised light chains were eventually incorporated into complete antibody molecules; the remaining 40% of the light chain immunoglobulins were released into serum and subsequently catabolised in the renal proximal tubules. In the absence of interstitial renal disease, urinary free light chain concentrations might reflect the quantitative degree of in vivo B cell Ig production.¹⁰ However, for active lupus nephritis, both polyclonal B cell activation and renal tubule impairment may be responsible for the presence of urinary free light chains. The significance of this observation needs further evaluation.

In conclusion, this is the first study to analyse the composition of proteins from urine in

human lupus nephropathy. We have found that increased sIL-2R, free light chains, and the presence of cytokine like molecules in protein from urine can directly reflect the severity of inflammatory and immunological reactions in patients with lupus nephritis and may serve as supplemental tests to histological examination of the renal tissue.

This work is supported by grants from the National Science Council and from the Institute of Biomedical Sciences, Academia Sinica, Republic of China. We are indebted to Miss Lina Lin for her technical assistance and Miss Chun-Shia Chong for typing this manuscript.

- 1 Becker G B, Hancock W W, Stow J L, Glasgow E F, Atkins R C, Thomson N M. Involvement of the macrophage in experimental chronic immune complex glomerulonephritis. Nephron 1982; 32: 227–33.
 Hooke D H, Hancock W W, Gee D C, Kraft N, Atkins R C.
- Monoclonal antibody analysis of glomerular hypercellularity in human glomerulonephritis. *Clin Nephrol* 1984; 22: 163-8.
 Nolasco F E B, Cameron J S, Hartley B, Coelho A, Hildreth G, Reuben R. Intraglomerular T cells and monocytes in
- nephritis: study with monoclonal antibodies. *Kidney Int* 1987; 31: 1160–6.
 Lovett D H, Ryan J L, Sterzel R B. A thymocyte-activating factor derived from glomerular mesangial cells. *J Immunol* 1983; 130: 1796–801.
- 5 Lovett D H, Sterzel R B, Ryan J L, Atkins E. Production of an endogenous pyrogen by glomerular mesangial cells. *J Immunol* 1985; **134**: 670–2. Boswell J M, Yui M A, Endres S, Burt D W, Kelley V E.
- b Joswell J M, Yul M A, Endres S, Burt D W, Kelley V E. Novel and enhanced IL-1 gene expression in autoimmune mice with lupus. *J Immunol* 1988; 141: 118-24.
 7 Boswell J M, Yui M A, Burt D W, Kelley V E. Increased tumor necrosis factor and IL-1β gene expression in the kidneys of mice with lupus nephritis. *J Immunol* 1988; 141: 2060 4 3050-4
- 3050-4.
 8 Noble B, Ren K, Taverne J, et al. Mononuclear cells in glomeruli and cytokines in urine reflect the severity of experimental proliferative immune complex glomerulonephritis. Clin Exp Immunol 1990; 80: 281-7.
 9 Hopper J E, Papagiannes E. Evidence by radioimmunoassay that mitogen-activated human blood mononuclear cells accessed a complex glometra of light chein J and a several complex glometra.
- that mitogen-activated human blood mononuclear cells secrete significant amounts of light chain Ig unassociated with heavy chain. Cell Immunol 1986; 101: 122–31.
 10 Epstein W V. Immunologic events preceding clinical exacerbation of systemic lupus erythematosus. Am J Med 1972; 54, 631 6
- 1973; 54: 631-6.
 11 Hopper J E, Sequeira W, Martellotto J, Papagiannes E, Perna L, Skosey J L. Clinical relapse in systemic lupus erythematosus:correlation with antecedent elevation of urinary free light-chain immunoglobulin. J Clin Immunol 1989; 9: 338-50.
- Seckinger P, Williamson K, Balavoine J-F, et al. A urine inhibitor of interleukin 1 activity affects both interleukin 1α and 1β but not tumor necrosis factor α . *J Immunol* 1987; **139:** 1541–5.
- 139: 1541–5.
 13 Seckinger P, Isaaz S, Dayer J-M. A human inhibitor of tumor necrosis factor α. *J Exp Med* 1988; 167: 1511–6.
 14 Werber H I, Emancipator S N, Tykocinski M L, Sedor J R. The inter-leukin 1 gene is expressed by rat glomerular mesangial cells and is augmented in immune complex glomerulonephritis. *J Immunol* 1987; 138: 3207–12.
 15 Van Oers M H J, Van Der Heyden A A P A M, Aarden L A. Interleukin 6 (IL-6) in serum and urine of renal transplant recipients. *Clin Exp Immunol* 1988; 71: 314–9.
 16 Okuda S, Languino L R, Ruoslahti E, Border W A. Elevated expression of transforming growth factor-fa and proteo-
- Okuda S, Languino L R, Ruoslahti E, Border W A. Elevated expression of transforming growth factor-β and proteo-glycan production in experimental glomerulonephritis: possible role in expansion of mesangial extracellular matrix. *J Clin Invest* 1990; 86: 453-62.
 Rubin L A, Kurman C C, Fritz M E, et al. Soluble inter-leukin 2 receptors are released from activated human lymphoid cells in vitro. *J Immunol* 1985; 135: 3172-7.
 Wolf R E, Brelsford W G. Soluble interleukin-2 receptors in systemic lunus eruthematogue. *Archivite Determ* 1988: 31:

- Wolf R E, Breistord W G. Soluble interleukin-2 receptors in systemic lupus erythematosus. Arthritis Rheum 1988; 31: 729-35.
 Tokano Y, Murashima A, Takasaki Y, Hashimoto H, Okumura K, Hirose S. Relation between soluble inter-leukin 2 receptor and clinical findings in patients with systemic lupus erythematosus. Ann Rheum Dis 1989; 48: 803-9.
- 20 Novick D, Engelmann H, Wallach D, Rubinstein M. Soluble cytokine receptors are present in normal human urine. J Exp Med 1989; 170: 1409–14.
- *J Exp Med* 1989; 170: 1409–14.
 Marcon L, Fritz M E, Kurman C C, Jensen J C, Nelson D L. Soluble Tac peptide is present in the urine of normal individuals and at elevated levels in patients with adult T cell leukaemia (ATL). Clin Exp Immunol 1988; 73: 20.32 20 22
- 29-33.
 22 Stahl R A K, Thaiss F, Kahf S, Schoeppe W, Helmchen U M. Immune-mediated mesangial cell injury—biosynthesis and function of prostanoids. *Kidney Int* 1990; 38: 273-81.
 23 Waldmann T A, Strober W, Mogielnicki R P. The renal handling of low molecular weight proteins. II. Disorders of serum protein catabolism in patients with tubular proteinuria, the nephrotic syndrome, or uremia. J Clin Invest 1972; 51: 2162-74.