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Delay-related activity in marmoset prefrontal cortex

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Persistent delay-period activity in prefrontal cortex (PFC) has long been regarded as a neural signature of working memory (WM). Electrophysiological investigations in macaque PFC have provided much insight into WM mechanisms; however, a barrier to understanding is the fact that a portion of PFC lies buried within the principal sulcus in this species and is inaccessible for laminar electrophysiology or optical imaging. The relatively lissencephalic cortex of the New World common marmoset (*Callithrix jacchus*) circumvents such limitations. It remains unknown, however, whether marmoset PFC neurons exhibit persistent activity. Here, we addressed this gap by conducting wireless electrophysiological recordings in PFC of marmosets performing a delayed-match-to-location task on a home cage-based touchscreen system. As in macaques, marmoset PFC neurons exhibited sample-, delay-, and response-related activity that was directionally tuned and linked to correct task performance. Models constructed from population activity consistently and accurately predicted stimulus location throughout the delay period, supporting a framework of delay activity in which mnemonic representations are relatively stable in time. Taken together, our findings support the existence of common neural mechanisms underlying WM performance in PFC of macaques and marmosets and thus validate the marmoset as a suitable model animal for investigating the microcircuitry underlying WM.

Key words: delayed-response task, electrophysiology, persistent firing, primate, working memory.

Introduction

In 1936, Jacobsen tested Old World mangabey and baboon monkeys on a spatial delayed-response task, in which the animal observed the hiding of food under 1 of 2 identical cups. After a delay of a few seconds to several minutes, the animal was allowed to choose one of the cups in order to obtain the food item. Jacobsen's now-famous observation was that bilateral prefrontal cortex (PFC) lesions completely impaired the monkeys' performance in this task. Subsequent studies narrowed the prefrontal regions responsible for the maintenance of visuospatial information to the cortex surrounding the caudal two-thirds of the principal sulcus in rhesus macaque monkeys (Goldman and Rosvold 1970; Goldman et al. 1971). By recording from single neurons in this prefrontal region in macaque monkeys during similar manual spatial delayed-response tasks, Fuster (1973) and Kubota and Niki (1971) found neurons that responded to the visual stimulus, anticipated the motor response, were active following the response, or, most surprisingly, neurons that discharged persistently during the delay period. In subsequent studies, Goldman-Rakic and colleagues took advantage of the behaviorally well-controlled oculomotor delayed-response task and demonstrated spatial tuning of delay-related activity and differences between correct and error trials (Funahashi et al. 1989, 1991). This led to the now-prevalent notion that persistent delay-period activity in the dorsolateral PFC represents the cellular basis of spatial working memory (WM; see for review Riley and Constantinidis 2016). Over the past 25 years, neural recordings in macaque monkeys have provided crucial insights into delay activity associated with various cognitive functions, and pharmacological studies in macaques have also begun to explore the influence of different neurotransmitter systems, such as dopamine (Ott and Nieder 2016) and acetylcholine (Vijayraghavan and Everling 2021), on persistent delay activity.

While macaque monkeys are an excellent animal model for studies of the neural basis of WM in the PFC, the species also has several severe shortcomings: (i) a large part of the lateral PFC in macaques is deeply buried in the principal sulcus, making it difficult to access for laminar-specific recordings and manipulations; (ii) macaques are large and difficult to handle; (iii) they are expensive to house; (iv) their low birth rate and long sexual maturation make longitudinal developmental studies challenging, and (v) pharmacological studies of new compounds are expensive due to the animal's large body size. Although rodents, which also show delayrelated activity in their frontal cortex, do not have these shortcomings, mice and rats lack a granular PFC. The so-called medial PFC of rodents likely corresponds to the primate anterior cingulate cortex and lacks the strong connectivity with parietal cortex that is characteristic of the primate lateral PFC (for reviews, see Preuss 1995;

Laubach et al. 2018). Moreover, it seems that delayrelated activity in rodents is only present for short delays and is less robust than in macaque monkeys (Preuss 1995; Delatour and Gisquet-Verrier 1996; Ragozzino et al. 1998; Delatour and Gisquest-Verrier 1999; Gisquet-Verrier and Delatour 2006).

A nonhuman primate (NHP) species that avoids many of the problems of macaques is the small New World common marmoset (Callithrix jacchus). The species is rapidly becoming popular as a powerful complement to canonical macaque and rodent models for preclinical modeling of the human brain in healthy and diseased states (Okano et al. 2016). The marmoset's fast sexual maturation, low inter-birth interval, and routinely observed chimeric twinning make it the leading candidate for transgenic primate models (Sasaki et al. 2009; Okano et al. 2012; Kishi et al. 2014; Belmonte et al. 2015; Mitchell and Leopold 2015; Sasaki 2015). The lissencephalic (smooth) marmoset cortex also offers the opportunity for laminar electrophysiological recordings and optical imaging in frontoparietal areas. Most importantly, like macaques, marmosets have a granular lateral PFC that has strong connectivity with posterior parietal cortex (Burman et al. 2006; Reser et al. 2013).

Field studies have demonstrated that marmosets exhibit spatial WM (Miles 1957; MacDonald et al. 1994; Tsujimoto and Sawaguchi 2002; Nakako et al. 2013) and touchscreen-based studies using delayed-match-tolocation (DML) tasks have demonstrated that marmosets can maintain spatial information in WM for at least several seconds (Spinelli et al. 2004, 2006; Yamazaki et al. 2016; Sadoun et al. 2019). In the DML task, a stimulus is briefly presented at one location of the screen (sample), followed by a waiting period (delay), and then the stimulus is again presented with one or several other identical stimuli (choice) and the subject has to touch the stimulus that is presented at the same location as the sample. The next step in establishing marmosets as an NHP model for PFC physiology is to test whether marmoset PFC neurons exhibit response profiles in delayed-response task similar to those in macaques. To tackle this question, we trained 3 marmosets on the DML task using a custom-developed on-cage touchscreen system and wirelessly recorded neural activity using 96-channel Utah arrays chronically implanted over the marmoset PFC.

Methods

Subjects

Data were collected from 3 adult female common marmosets (*C. jacchus*; Marmoset L, 41 months; Marmoset A, 26 months; Marmoset B, 24 months). All experimental procedures conducted were in accordance with the Canadian Council of Animal Care policy on the care and use of laboratory animals and a protocol approved by the Animal Care Committee of the University of Western Ontario Council on Animal Care. The animals were under the close supervision of university veterinarians.

Delayed-match-to-location task

Marmosets performed a DML task (Fig. 1) with varying delay lengths (2-8 s) on an in-house developed touchscreen testing box attached to the home cage (see Section 2.3). The touchscreen testing box measured 30 \times 20×20 cm and had an opaque ceiling and floor. One end of the box was fitted with a metal docking plate designed to interface with the home cage and sliding door to allow access to the touchscreen testing box. Directly opposite to this was a slot for the touchscreen (254.80 \times 177.50 mm, Elo 1002L). The remaining walls were constructed of transparent plexiglass, which allowed light in and the marmosets to freely view the surrounding room space. For Marmosets A and B, we covered the box with a blanket as the room view appeared to cause stress and reduce their motivation to perform the task. A reward spout made from brass tubing was mounted on the floor with the outlet directly in front of the screen, 5–7 cm from the floor. As some marmosets preferred to use their snout to interact with the screen, to maintain consistency between marmosets we encouraged the use of the forelimbs by affixing an array of vertical plexiglass bars (0.8 cm width, 2 cm spacing) in front of the screen. The task and reward delivery were controlled by a Raspberry PI 3 Model B running a custom-written Python script (marmtouch v0.0.1, Selvanayagam et al. 2021). Each trial began with the presentation of a sample stimulus (filled blue or pink circle, 3 cm diameter) on a gray background at one of 4 corner locations for 2.5 s (Fig. 1). This was followed by a delay period in which the screen remained blank for 2-8 s. After the delay period, choice stimuli (filled blue or pink circles, 3 cm diameter) were presented at each of the 4 corner locations and the animal was required to touch the location matching the previously presented stimulus to obtain 0.075–0.1 mL of liquid reward per trial (50/50 mix of 1:1 acacia gum powder and water with liquid marshmallow). The reward was delivered via an infusion pump (model NE-510; New Era Pump Systems) through a liquid spout placed in front of the touchscreen monitor. Trials were separated by 5s intertrial periods. Animals performed 50–160 correct trials (lower and upper bound, respectively) per session for approximately 5–15 mL of liquid reward.

Training protocol

Marmosets participated in a sequence of training phases derived via a process of successive approximations in order to establish their performance on the DML task. In a first training stage, we aimed to establish the association between touching a stimulus on the screen and obtaining a reward. To accomplish this, a small piece of marshmallow was placed at the center of the screen, and a visual stimulus consisting of a large blue or pink circle (6 cm diameter) was presented at the same location. Touching the screen, and consequently the stimulus, allowed the



Fig. 1. DML task. Successive panels indicate initial appearance of the sample stimulus, delay period, and response (choice) phase. The sample was presented randomly in 1 of 4 possible locations.

animals to harvest the marshmallow and additionally triggered delivery of a liquid reward through the reward spout, which was paired with an auditory cue. Once they reliably performed this behavior, we progressed to a stage in which the screen was no longer baited with a marshmallow, and touches to an identical large blue or pink circle were rewarded with liquid reward as in the previous stage. Because the DML task required the animals to touch stimuli at varying locations on the touchscreen, we next familiarized them with touching stimuli at varying locations. In this training phase, stimuli were presented randomly at one of the 4 corners of the touchscreen, and the animals received a liquid reward delivered from the reward spout for stimulus touches. Once this behavior was performed reliably, we then introduced the concept of a short delay into the training protocol. In this phase, a visual sample stimulus was flashed at one location on the touchscreen. This was followed by a delay of 500 ms (incrementally increased to longer durations), after which a second comparison stimulus was flashed at the same location as the sample. Marmosets received a reward for touching this second comparison stimulus. To encourage exploratory responding and promote acquisition of the task concept, the animals were not restricted from touching the screen during the cue or delay periods. In addition, to simplify the task and training, we added the constraint that, in this stage and those following, cue and comparison stimuli were always presented at one of the 4 corners of the screen, rather than at locations randomized to any screen location on a trial-by-trial basis. We reasoned that this would make it easier for marmosets to acquire the task rule while still requiring the use of mnemonic resources to correctly perform the task. Once the above steps were completed, animals advanced to the DML task proper, in which the sample and delay periods were identical to the previous training stage, but comparison stimuli were presented simultaneously at each of the 4 corners of the screen, forcing the animals to rely upon their mnemonic representations of the earlier sample location and knowledge of the location matching rule to perform the task correctly. However, an additional step was required for Marmoset A, as this animal remained at chance levels in the DML task proper. In this additional step, the animal was given a luminance cue to the correct location during the response period. A visual

sample stimulus was presented at one location on the screen, followed by a delay during which the screen was blank. Following this, comparison stimuli were presented simultaneously at each of the 4 corners of the screen. Of these, the stimulus matching the earlier sample stimulus was presented at a slightly lower luminance to act as an additional aid in acquiring the task rule of matching stimulus location. Once above-chance levels, Marmoset A advanced to the DML task proper. Animals were able to complete this training protocol in 6–8 weeks.

Array surgery

Once an animal reached 60% correct task performance, it underwent an aseptic surgical procedure under general anesthesia in which a 96-channel Utah array (4 mm \times 4 mm; 1 mm or 1.5 mm electrode length; 400 μ m pitch; iridium oxide tips) was implanted in the left PFC (see Selvanayagam et al. 2019 for details). During this surgery, a microdrill was used to open a 4 mm burr hole in the skull and was enlarged as necessary using a rongeur. The dura was removed, and the array was manually inserted into the lateral PFC; wires and connectors were fixed to the skull using dental adhesive resin cement (All-Bond Universal and Duo-Link, Bisco Dental Products). Once implanted, the array site was covered with a thin layer of silicone adhesive (Kwik Sil; World Precision Instruments). A screw hole was drilled into the skull on the right side to place a stainless steel ground screw. The ground wire of the array was then tightly wound around the base of the screw to ensure good electrical connection. A combination recording chamber/head holder (Johnston et al. 2018) was placed around the array and connectors and fixed in place using further layers of dental adhesive resin cement. Finally, a removable protective cap was placed on the chamber to protect the 3×32 channel omnetics connector.

Neural recordings

After recovery from array implantation, we verified that electrode contacts were within the cortex by monitoring extracellular neural activity using the SpikeGadgets' data acquisition system, Trodes (v2.2.2). Upon observing single- or multiunit activity at multiple sites of the array after about 3 weeks, we commenced unrestrained datalogger recordings of extracellular activity from the 96 implanted electrodes while the animal performed the DML task on a touchscreen attached to the home cage. Prior to a recording session, the animal was removed from the home cage. The protective chamber cap was removed, exposing the 3×32 omnetics connector. A custom-built routing board to 3 omnetics connector, horizontal headstage, and an untethered datalogger (SpikeGadgets, San Francisco, USA) was attached on top of the animal's head. Once secured, the animal was returned to a smaller restricted area of the home cage where there was access to the touchscreen testing box for 2 h. Subsequently, the animal was removed from the home cage for the removal of the routing board,

headstage, and datalogger. The protective chamber cap was placed on, and the animal was returned to the home cage. During the recording, the Logger Dock (SpikeGadgets, San Francisco, USA) was used for untethered data acquisition and received synchronization pulses from the Raspberry Pi, aligned to sample onset. Camera(s) (Raspberry Pi camera (G) with fisheye lens or Arducam IMX477 Synchronized Stereo Camera with fisheye lens) were mounted on the in-house developed touchscreen testing box to record videos (aligned to sample onset) of the animal performing the DML task. In most recording sessions, animals were separated from their cage mate. However, in a few sessions where the animals were not separated (to reduce stress from separation), recorded videos were used to score each trial for animal identity.

Neural data were first filtered with a common median filter to remove large movement-related artifacts. This was then filtered with a 4-pole Butterworth 500 Hz highpass filter. Spike detection and sorting were performed offline (Plexon Offline Sorter v3). Only clearly isolated single units with baseline discharge rates greater than 0.5 Hz were included in the analysis (149/1,853 units were excluded). Videos of all trials in all sessions were manually scored to determine whether the animal looked at the sample during the sample-presentation period. Only experimental sessions with at least 10 correct trials where the animal looked at the sample during the sample-presentation period and did not touch the screen during the delay period were included (5 of 33 sessions were excluded).

Data analysis

Analysis was performed using custom code written in Matlab (MathWorks).

Activity within distinct task epochs was assessed using analyses of variance (ANOVA) to compare the mean discharge rates of each neuron for each condition (four spatial locations) and each task epoch (baseline, sample, delay, pre-response, and post-response). The baseline epoch was defined as the 1.5 s prior to the onset of the sample stimulus to the time of sample stimulus presentation. The sample epoch was defined as the 100 ms after the onset of the sample stimulus to the time of stimulus offset (2.4 s). We excluded the first 100 ms to avoid any potential sluggish sample-related activity contaminating estimates of delay-related activity. The delay epoch was 4 s for Marmoset L and 2 s for Marmoset A and B. The pre-response epoch was defined as the 300 ms period prior to the touch response and the post-response epoch was defined as the 1,000 ms immediately after the touch response. Statistical significance was evaluated at an alpha level of P < 0.02.

To investigate differences in activity between correct and error trials, a modulation index was computed by subtracting the discharge rate of the non-preferred condition from the discharge rate of the preferred condition, divided by the sum of the discharge rate of the preferred and non-preferred condition. For each unit, we classified the preferred and non-preferred conditions as those in which the unit had the highest and lowest mean discharge rates during the delay epoch in the correct trials, respectively.

Waveform preprocessing and cell type classification

For each single unit, the mean waveform was interpolated (cubic spline) from an original sampling rate of 30 kHz–1 MHz. For cell class classification, we computed 2 measures of the resultant waveform: trough-to-peak duration and time for repolarization. The time for repolarization was defined as the time at which the waveform amplitude decayed 25% from its peak value (Ardid et al. 2015). Here we clustered cell classes into broad and narrow spiking cells using an unsupervised algorithm by Trainito et al. (2019), which correspond to putative pyramidal cells and interneurons. These cell classes have previously been demonstrated to contribute differentially to spatial tuning in the oculomotor delayed-response task (Rao et al. 1999). In this method, the expectationmaximization (EM) algorithm is used to estimate the parameters of the Gaussian mixture model (GMM), a statistical model which describes the mean and variance of underlying subpopulations.

Pattern classifier

To determine whether delay activity was sustained throughout the entire delay period, for each session we trained linear supply vector machines (SVM) to predict the sample stimulus location using the discharge activity of each neuron in a sliding window over the task interval. We constructed models for 1 s time bins starting 1.5 s before sample onset (baseline) to 1 s after the end of the delay epoch in steps of 500 ms. Models were trained with 80% of the data at a specific time bin and tested at the same time bin with the remaining 20% of data for a classification accuracy. The same model was used to test the same 20% of data at all other time bins. This process of separating the data into a training and testing set was repeated 100 times, and the classification accuracies were averaged to provide a more robust estimate. These values were then averaged across sessions for each animal separately.

To investigate differences in the 2 populations of neurons (BS vs. NS), we trained a single model over the entire delay period using the entire dataset for each session. Note that since we used a one versus one linear SVM, each feature (neuron) has a coefficient for each pair of conditions (4 choose 2, 6 coefficients; i.e. condition 1 vs. 2, condition 1 vs. 3, ... condition 3 vs. 4). As a measure of each unit's contribution to the model, we computed the magnitude of the 6-dimensional vector constructed from these coefficients. A larger magnitude suggests that a neuron's activity is better able to separate between all pairs of conditions. We then compared these weights between BS and NS cells in an independent samples t-test.

Array localization

Marmosets were euthanized at the end of the data acquisition process to prepare the brains for ex vivo magnetic resonance imaging (MRI) scan. The animals were deeply anesthetized with 20 mg/kg of ketamine plus 0.025 mg/kg medetomidine and 5% isoflurane in 1.4%-2% oxygen to reach beyond the surgical plane (i.e. no response to toe pinching or cornea touching). They were then transcardially perfused with 0.9% sodium chloride irrigation solution, followed by 4% paraformaldehyde in 0.1 M phosphate buffer solution or 10% buffered formalin. The brain was then extracted and stored in 10% buffered formalin for more than a week before ex vivo MRI. On the day of the scan, the brain was transferred to another container for imaging and immersed in a fluorine-based lubricant (Christo-lube; Lubrication Technology) to improve homogeneity and avoid susceptibility artifacts at the boundaries. Ex vivo MRI was performed on a 9.4T 31 cm horizontal bore magnet (Varian/Agilent, Yarnton, UK) and Bruker BioSpec Avance III console with the software package Paravision-7 (Bruker BioSpin Corp, Billerica, MA), a custom-built high-performance 15cm-diameter gradient coil with 400 mT/m maximum gradient strength (xMR, London, CAN; Peterson et al. 2018), and an mp30 (Varian Inc., Palo Alto, USA) transmit/receive coil. High-resolution $(100 \times 100 \times 100 \mu m \text{ for})$ Marmoset A and Marmoset B, $100 \times 100 \times 200 \ \mu m$ for Marmoset L) T2-weighted images were acquired for each animal.

The raw MRI images were converted to NifTI format using dcm2niix (Li et al. 2016) and the MRIs were nonlinearly registered to the ultra-high-resolution ex vivo NIH template brain (Liu et al. 2018), which contains the location of cytoarchitectonic boundaries of the Paxinos atlas (Paxinos et al. 2012), using Advanced Normalization Tools (Avants et al. 2011) software. The resultant transformation matrices were then applied to the cytoarchitectonic boundary image included with the NIH template brain atlas. These cytoarchitectonic boundaries overlaid on the registered ex vivo anatomical T2 images were used to reconstruct the location of the implanted array in each marmoset (Supplementary Fig. 1).

Results

Task performance

Marmosets performed a total of 28 sessions (Marmoset L, 18 sessions; Marmoset B, 8 sessions; Marmoset A, 2 sessions). Overall, we observed that marmosets were able to perform the DML task at above chance accuracy at a range of delay durations. Data were collected from 3 animals at a 2 s delay, (Marmoset L = 70.6%, Marmoset B = 64.1%, Marmoset A = 64.2%). Marmoset L additionally performed the task at 2 longer delay durations (4 s and 8 s). Consistent with many reports, performance declined with increasing durations, falling to 63.4% and 54.2%, respectively. This observation was confirmed statistically via one-way ANOVA (F(2,23) = 12.49, P < 0.001). A post

hoc Bonferroni corrected t-test revealed that mean task accuracy was significantly lower following 8 s delays as compared to 2 or 4 s delays (P < 0.05).

To further investigate task performance, we compared marmosets' reaction times (RTs) on correct and error trials at the 2 s delay duration. The logic of the DML task, as in other delayed-response tasks, is that animals acquire a mnemonic representation of the spatial location of the cue stimulus during cue presentation, maintain this representation during the delay period of the task, and subsequently compare this representation with the stimuli presented during the response period to select the correct response. Generally speaking, if animals are relying on mnemonic processes to guide response selection, it is expected that RTs on error trials will be similar to or longer than those on correct trials, since they should reflect "diligent guesses" regarding the correct location (Link 1982). For the 2 s delay condition, we found that in all cases, RTs on error trials were equal to or greater than those on correct trials, with RTs significantly greater for error than correct trials for Marmosets L and A (ttest, P < 0.05) (Marmoset L: \overline{x} correct RT = 1.23 s, \overline{x} error RT = 1.99 s; Marmoset B: \overline{x} correct RT = 1.43 s, \overline{x} error RT = 1.57 s; Marmoset A: \overline{x} correct RT = 1.20s, \overline{x} error RT = 1.84 s). These results are consistent with a reliance of marmosets on mnemonic processes during DML task performance. Taken together with the findings of above chance task performance for all animals, and the decline in task performance as a function of delay duration observed in Marmoset L, these data indicate that performance on the DML task was an accurate reflection of marmosets' spatial WM abilities.

Marmoset PFC neurons exhibit sample-, delay-, and response-period activity

To determine whether marmoset PFC neurons exhibit task-related activity in the DML task, we simultaneously recorded the activity of a total of 1,704 PFC neurons (502 in Marmoset L, 908 in Marmoset B, 294 in Marmoset A) in 3 monkeys over 28 experimental sessions in which they performed this task. We included in our analyses only well-isolated single units with baseline discharge rates greater than 0.5 Hz. Many neurons were modulated during the DML task (460 in Marmoset L, 782 in Marmoset B, 209 in Marmoset A), which we defined as a significant difference in mean discharge rate between a task epoch and a baseline period. Representative single neurons exhibiting modulations during the sample epoch, delay epoch, both sample and delay epochs, as well as during the response epochs are presented in Figs 2 and 3. Many neurons exhibited task-related activity modulations, and these modulations were observed in all epochs of the DML task, with a bias toward units responding in the 2 response epochs. Of the 460 neurons modulated during the task in Marmoset L, 135 neurons (26.9% of overall neurons) displayed sample-related activity, 238 neurons (47.4%) displayed delay-period activity, 359 neurons (71.5%) displayed pre-response-related activity, and 402



Fig. 2. PFC neurons exhibit sample- and delay-period activity during the DML task. Representative single neurons exhibiting excited (A) or suppressed (B) activity during the sample epoch, excited (C) or suppressed (D) activity during the delay epoch, and excited (E) or suppressed (F) activity during both sample and delay epochs. Rasters are aligned to sample onset. Red lines depict RT.

neurons (80.1%) displayed post-response related activity. In Marmoset B, 247 (27.2%), 228 (25.1%), 557 (61.3%), and 630 (69.4%) neurons, as well as in Marmoset A, 83 (28.2%), 67 (22.8%), 130 (44.2%), and 159 (54.1%) neurons displayed sample-related activity, delay-period activity, pre-response-related activity, and post-response-related activity, respectively. Overall, we found that, across the 3 animals, well-isolated single units were recorded across a broad set of prefrontal subregions including areas 46D, 46 V, 8aD, 8aV, 9, and 10 (see Fig. 4). We observed task-related activity in all epochs in all subregions but noted that the proportion of units with delay activity was relatively lower in areas 9 and 10.

Prior work in macaque monkeys performing delayedresponse tasks has revealed that modulations of sample-, delay-, and response-related activity may take the form of either increases or decreases from the baseline discharge rate (Funahashi et al. 1989, 1991). To



Fig. 3. PFC neurons exhibit response-related activity during the DML task. Representative single neurons exhibiting excited (A) or suppressed (B) activity during the pre-response epoch, and excited (E) or suppressed (F) activity during both pre- and post-response epochs. Rasters are aligned to response.

investigate such modulations in marmoset PFC, we further classified whether each neuron was significantly excited or suppressed in each task epoch with respect to baseline (Table 1). The proportions of neurons excited and suppressed significantly varied across task epochs (chi-square test, Marmoset L: X^2 (3, N = 1134) = 37.49, P < 0.001; Marmoset B: X^2 (3, N = 1662) = 19.41, P < 0.001; Marmoset A: X^2 (3, N = 439) = 5.57, P < 0.001). Overall, these data show that marmoset PFC neurons are

modulated in a similar manner to those in macaque during the DML task.

Marmoset PFC neurons exhibit spatial tuning during DML task performance

A now-classic observation in macaque PFC is that single units exhibit spatial tuning of discharge rates in the sample, delay, and response epochs of spatial WM tasks. We similarly observed such tuning in marmoset



Fig. 4. Distribution of task-modulated units across recording arrays. Array locations were reconstructed using high-resolution MRIs and superimposed on a standardized marmoset brain, area boundaries from Paxinos et al. (2012; left panel). For each marmoset, task-modulated units were distributed across the array in locations where well-isolated single units were observed, followed by locations on the array where sample-, delay-, and response-related units were observed (from left to right). Grey color depicts where on the array well-isolated single units were not observed.

Table 1. Number of neurons excited or suppressed in each task epoch for each marmoset.

Marmoset	Modulation	Epoch				
		Sample	Delay	Pre-response	Post-response	
L	Excited	63 (47%)	95 (40%)	91 (25%)	99 (25%)	
	Suppressed	72 (53%)	143 (60%)	268 (75%)	303 (75%)	
В	Excited	97 (39%)	49 (221%)	171 (31%)	176 (28%)	
	Suppressed	150 (61%)	179 (79%)	386 (69%)	454 (72%)	
А	Excited	36 (43%)	19 (28%)	39 (30%)	49 (31%)	
	Suppressed	47 (57%)	48 (72%)	91 (70%)	110 (69%)	

PFC neurons in all animals from which we recorded. To investigate this statistically, we carried out separate oneway ANOVAs with sample location at 4 levels for each task epoch separately. An example neuron exhibiting such tuning is depicted in Fig. 5. Overall, in Marmoset L, 32 neurons (23.7%, proportion in respect to the number of neurons modulated in the epoch) displayed tuning during the sample period, 58 neurons (24.4%) displayed tuning during the delay period, 80 neurons (22.3%) displ ayed tuning during the pre-response period, and 135 neurons (33.6%) displayed tuning during the post-response period. In Marmoset B, 45 (18.2%), 20 (8.8%), 143 (25.7%), and 204 (32.4%) neurons, as well as in Marmoset A, 2 (2.4%), 10 (14.9%), 16 (12.3%), and 18 (11.3%) neurons displayed tuning activity during the sample period, delay period, pre-response period, and post-response period, respectively.

Neural activity reflects mnemonic processes rather than simple orienting

One possibility, noted also in earlier work (see Goldman-Rakic 1995), was that animals were able to correctly perform the task not by relying on WM per se, but rather by simply orienting the head or whole body toward the location of the stimulus during the sample presentation and maintaining that orientation throughout the delay period until responding. This strategy would not only circumvent the cognitive process of WM but additionally potentially contaminate any observations of delayperiod activity with a sustained signal indicating body or head orientation.

To address this, in several sessions (3 in Marmoset L, 2 in Marmoset B, 2 in Marmoset A), videos of all correct trials were manually scored with respect to whether the animal was orienting (head-oriented) toward the sam-



Fig. 5. Example neuron exhibiting spatial tuning during the DML task. Rasters are aligned to sample onset.

ple during the delay period (Supplementary Video 1) or not (Supplementary Video 2). In the majority of trials (95.8% for Marmoset L, 92.3% for Marmoset B, 94.9% for Marmoset A), the animals did not fixate on the location of the sample during the delay period. We removed the trials on which the animals fixated on the location of the sample and performed an ANOVA on condition and task epoch again to compare the differences. For these sessions, the number of neurons that displayed delayperiod activity was similar (Marmoset L: 55/59 neurons [93.2%]; Marmoset B: 81/85 neurons [95.3%]; Marmoset A: 62/67 neurons [92.5%]). Additionally, in 23/28 sessions where we recorded the touch location during the response period, we performed analysis on same versus opposite side error. 37.5% of errors were made on the same side and 62.5% of errors were made on the opposite side. Together, this indicates that our observations were not contaminated by orienting or strategic processes and reflect instead the retention of mnemonic information.



Fig. 6. Delay-period activity is reduced on error trials. Example neuron for which the discharge rate during the delay period is significantly different between correct and error trials (A). Delay-period modulation index computed from the preferred and non-preferred conditions separately for correct and error trials (B–D). Average modulation indices of neurons recorded at each electrode contact of the Utah array across sessions for Marmoset L and A (E). Grey color depicts where on the array well-isolated single units were not observed.

Persistent delay-period activity in marmoset PFC reflects task performance

A seminal observation linking persistent activity and task performance is that persistent delay-period activity is attenuated on trials on which animals make performance errors relative to those on which they perform correctly (Funahashi et al. 1989). To investigate this in marmoset PFC, we compared discharge rates between correct and error trials for neurons which were responsive during the delay period (e.g. exhibited discharge rates significantly elevated during the delay epoch relative to baseline). In Marmoset L, 119/204 neurons (58.3%) exhibited a significant difference in discharge rate between correct and incorrect trials during the delay period. In Marmoset B, 48/212 (22.6%) and in Marmoset A, 28/63 (44.4%) neurons had a significant difference (independent samples t-test, P < 0.05). Figure 6A depicts an example neuron for which the discharge rate during the delay period was significantly different between correct and incorrect trials.

To further investigate the magnitude of differences in neural activity between correct and incorrect trials during the delay period, we computed a modulation index from the preferred and non-preferred conditions separately for correct and incorrect trials (Fig. 6B). For Marmoset L, the modulation index was higher for correct trials than incorrect trials (0.2414 vs. -0.0065, P < 0.001). This trend was similar for Marmoset B (0.222 vs. 0.034, P < 0.001) and Marmoset A (0.230 vs. -0.018, P < 0.001). Overall, these data are consistent with previous reports indicating that delay-period activity is reduced on error trials, and they provide evidence for a link between the magnitude of persistent delay-period activity and WM task performance.

To determine the PFC locations at which we observed the strongest, spatially selective delay activity, we averaged the modulation indices of neurons recorded at each electrode contact of the Utah array across sessions. We pooled across the neurons recorded from Marmosets L and B to improve our sampling of common PFC subregions as the arrays in these animals shared a high degree of overlap with respect to the PFC areas sampled (See Fig. 4). We observed the greatest mean modulation indices at the center of the array along the anteriorposterior axis, roughly corresponding to areas 46, 8aV, and 8aD (see Fig. 6E).

Broad and narrow spiking neurons are modulated in all task epochs

To investigate the contributions of putative pyramidal cells and interneurons to mnemonic processes in marmoset PFC, we used the EM algorithm for GMM clustering method on the trough-to-peak duration and

Marmoset	Modulation	Cell type	Epoch				
			Sample	Delay	Pre-response	Post-response	
L	Excited	BS	34	58	58	62	
		NS	29	37	33	37	
	Suppressed	BS	60	95	165	192	
		NS	12	48	103	111	
В	Excited	BS	79	45	118	132	
		NS	18	4	53	44	
	Suppressed	BS	101	117	289	334	
		NS	49	62	97	120	
A	Excited	BS	29	17	31	40	
		NS	7	2	8	9	
	Suppressed	BS	38	42	77	92	
		NS	9	6	14	18	

Table 2. Number of neurons excited or suppressed in each task epoch for each marmoset separated by cell type.

time for repolarization of single unit waveforms to identify BS and NS cell clusters in an unsupervised manner (Supplementary Fig. 2). This resulted in 414 cells (28.5%) being classified as narrow spiking and 1,037 cells (71.5%) being classified as broad spiking cells (Marmoset L: BS = 286 cells, NS = 174 cells; Marmoset B: BS = 575 cells, NS = 207 cells; Marmoset A: BS = 176 cells, NS = 33cells). For neurons modulated in each epoch of the DML task, we identified whether the neuron was classified as BS or NS. In Marmoset L, 94 BS (69.6%) and 41 NS (30.4%) cells displayed sample-related activity; 153 BS (64.3%) and 85 NS (35.7%) cells displayed delay-period activity; 223 BS (62.1%) and 136 NS (37.9%) cells displayed preresponse-related activity; and 254 BS (63.2%) and 148 NS (36.8%) cells displayed post-response-related activity. In Marmoset B, 180 (72.9%), 162 (71.1%), 407 (73.1%), and 466 (74.0%) BS cells, as well as in Marmoset A, 67 (80.7%), 59 (88.1%), 108 (83.1%), and 132 (83.0%) BS cells displayed sample-related activity, delay-period activity, pre-response-related activity, and post-response-related activity, respectively. In Marmoset B, 67 (27.1%), 66 (28.9%), 150 (26.9%), and 164 (26.0%) NS cells, as well as in Marmoset A, 16 (19.3%), 8 (11.9%), 22 (16.9%), and 27 (17.0%) NS cells displayed sample-related activity, delay-period activity, pre-response-related activity, and post-response-related activity, respectively. For the taskmodulated cells, chi-square goodness of fit tests were conducted to determine if the proportion of each cell type varied across the task epochs. No significant differences were observed (Marmoset L: X^2 (3, N = 1,134) = 2.52, P > 0.05; Marmoset B: X^2 (3, N = 1,662) = 0.73, P > 0.05; Marmoset A: X^2 (3, N = 439) = 1.50, P > 0.05). We further identified whether BS and NS cells were excited or suppressed in each task epoch compared to baseline (Table 2). Overall, consistent with previous reports in macaque PFC (Wilson et al. 1994; Rao et al. 1999; Constantinidis et al. 2002), we observed that both BS and NS neurons were modulated in all task epochs and that this modulation could take the form of excitation or suppression.

Stable versus dynamic mnemonic representations in persistent activity

To determine whether delay activity was stable in time or temporally dynamic, for each session we trained a pattern classifier to predict the sample stimulus location from the population activity in a sliding window over the task interval. The classifier can be trained on data from a specific time point in the trial and can be tested on the same time point, or different time points, for a measure of the stability of the representation over the course of WM maintenance (Sreenivasan and D'Esposito 2019). Above chance classification accuracy (25%) indicates that a representation of the information being classified exists in the population. In Marmosets L and B, classifiers trained at a given delay epoch time bin predicted stimulus location in other delay epoch time bins more accurately than chance; this was not the case for Marmoset A (see Fig. 7). In Marmoset L, where the delay period was 4 s, we observed a robust pattern of stable population coding. A similar pattern was observed in Marmoset B albeit weaker, likely due to the shorter delay period (2 s). Finally, in Marmoset A, where the array was further anterior and medial (see Fig. 4), stable population coding was not observed.

We separated the population of neurons within each session into 2 categories (neurons that display delayperiod activity and neurons that did not as determined above) to determine their influence on the trained model. As we had used a one versus one, linear SVM with 4 classes (stimulus location) where each feature (neuron) had 6 (4 choose 2) coefficients, to determine a measure of each unit's contribution to the model, we computed the magnitude of the 6-dimensional vector constructed from these coefficients. These weights were greater for neurons that displayed delay-period activity (Marmoset L: Mean = 1.67, SE = 0.06; Marmoset B: Mean = 0.98, SE = 0.03) as compared to neurons that did not display delay-period activity (Marmoset L: Mean = 1.43, SE = 0.04; Marmoset B: Mean = 0.90, SE = 0.02; t-test: Marmoset L: P < 0.001; Marmoset B:



Fig. 7. Stable versus dynamic mnemonic representations at the population level. For each session, we trained a pattern classifier to predict the sample stimulus location from the population activity in a sliding window over the task interval (trained on data from a specific time point and tested on the same and different time points). Above chance classification accuracy (25%) indicates that a representation of the information being classified exists in the population. Stable population coding was present during the delay period in Marmoset L and B, but not in Marmoset A.

P < 0.05). The results indicated that neurons that display delay-period activity are mostly responsible for the stability of the representation over the course of WM maintenance.

We additionally separated the population of neurons within each session into cell types and performed the same analysis during the delay period. These weights were greater for broad spiking cells (Marmoset L: Mean=1.62, SE=0.04; Marmoset B: Mean=0.97, SE=0.02) as compared to narrow spiking cells (Marmoset L: Mean=1.40, SE=0.05; Marmoset B: Mean=0.79, SE=0.02; t-test: Marmoset L: P < 0.01; Marmoset B: P < 0.001). The results suggested that broad spiking cells to the population's representation of stimulus location throughout the delay period.

Discussion

Activity during delay periods is a hallmark of macaque PFC (Fuster and Alexander 1971; Fuster 1973; Niki and Watanabe 1976; Quintana et al. 1988; Funahashi et al. 1989, 1990). Although delay-related activity has been reported in rodents, it is only present for short delay periods and it is substantially less robust than in macaques. Whether small New World common marmosets show delay-related activity comparable to Old World macaques remains unanswered. Here we recorded single-neuron activity in the marmoset PFC while unrestrained monkeys performed a touchscreenbased DML task to address this question. We found that common marmosets possess robust delay-related activity throughout their PFC and that this activity seems to be sustained throughout delay periods.

The first delay-related activity in macaques was recorded in head-restrained monkeys performing a manual spatial delayed-response task (Fuster and Alexander 1971; Kubota and Niki 1971). Subsequently, an eye movement version of the spatial delayed-response task, the oculomotor delayed response (ODR) task was popularized by Goldman-Rakic's lab (Funahashi et al. 1989, 1990, 1991). The ODR task has a distinct advantage over previous manual versions in which eye positions can be controlled and samples can be presented and maintained in known retinotopic coordinates (Goldman-Rakic 1995). Although marmosets can be trained to perform saccadic eye movement tasks, it has thus far proven difficult to train these monkeys on the ODR task. To date, there are only a few conference reports regarding training marmosets on the ODR task, and the consensus is that the performance of marmosets is quite low and that delay length is limited to short periods under 400 ms

(Amly et al. 2021). Another group (Carney et al. 2019) delivered reward throughout the sample and delay periods as a means to encourage marmosets to maintain fixation, which often presents a challenge for this primate species and detrimentally affects task performance. In that case, they reported performance comparable to macaques, although the delivery of rewards concurrent with the delay period of task potentially confounds the interpretation of persistent delay activity since rewardrelated and mnemonic signals would be intermixed. We have also not succeeded in training marmosets on the ODR task. In contrast to the difficulties inherent in employing the ODR task in marmosets, several studies have shown that marmosets can be trained on the touchscreen version of a DML task with delay periods of up to 68 s (Spinelli et al. 2004, 2006; Yamazaki et al. 2016; Sadoun et al. 2019). We did note that the performance of marmosets on this task was modest in some cases, which may have been due to individual differences in anxiety or distractibility, though we did not investigate these factors systematically.

The delay-related activity that we observed in marmoset PFC neurons is remarkably similar to the initial reports by Fuster and Kubota (Fuster and Alexander 1971; Kubota and Niki 1971) in the macaque. Like macaque PFC neurons, marmoset PFC neurons exhibit sample-, delay-, and response-related activity. The profiles of individual neurons closely resembled those described in macaques. Some were active in the sample and response periods, some were active during the sample and delay periods, and others were active just during the delay period (Fig. 2) While we found many neurons that increased their activity during these periods, we also found an even larger proportion of neurons that decreased their activity from baseline during the different task epochs. This is also very similar to reports in macaques.

Due to the small size of the PFC in marmosets, each of our implanted 4 × 4 mm arrays covered multiple prefrontal areas. This allowed us to compare the density of task-related activity during different epochs across different areas. Although we found delay-related activity in all 3 animals in all sampled areas, including area 8Ad, 8Av, 47, 46 V, 46D, and 10, it was considerably weaker in marmoset A in which the array was implanted further anterior and medial than in the other 2 animals. This suggests that delay activity is less robust in marmoset areas 9 and 10. The strongest sample- and delay-related activity was found in areas 46, 47, and 8Av. Parts of area 8Av correspond to the frontal eye fields (FEF) in the marmoset (Selvanayagam et al. 2019). In fact, Marmoset L was also one of the subjects in our previous electrical microstimulation study and fixed vector saccades could be evoked at posterior lateral electrodes of this array. This is consistent with findings in macaques, which also show robust delay-related activity at sites corresponding to the FEF in area 8 (Funahashi et al. 1989).

A distinguishing feature of delay cells in primate lateral PFC is spatial tuning for the cued location (Funahashi et al. 1989, 1990; Funahashi 2006). Here, we observed spatial tuning in many of our delay cells and this effect was significantly stronger on correct than on error trials supporting the hypothesis that delay-related activity is linked to task performance in marmosets. Additionally, the large number of electrodes on our Utah arrays allowed us to sample a large number of cells simultaneously facilitating population-level analyses. We were able to predict stimulus location from the population activity above chance levels, demonstrating for the first time that delay-related activity in marmoset PFC represents mnemonic information at the population and single-neuron levels. Since animals were unrestrained in our task and often moved considerably during the delay period, it seems most likely that this tuning reflected an allocentric frame of reference. In human functional MRI studies of WM, greater activation is often observed for egocentric than allocentric representations (Moraresku and Vlcek 2020). Further studies comparing WM performance and neural activity in head-restrained versus freely moving conditions may prove illuminating with respect to whether such differences are apparent in electrophysiology.

Since the first recording studies by Fuster and Kubota (Fuster and Alexander 1971; Kubota and Niki 1971), WM has been thought to be mediated by persistent sustained neuronal spiking during the delay period (Goldman-Rakic 1995), and many circuit models of recurrent prefrontal circuits have been developed to understand the mechanisms underlying such activity (see Wang 2021). An alternative proposal is that WM is instantiated by discrete spiking bursts rather than sustained activity (Lundqvist et al. 2016). Here our results suggest that persistent activity was sustained throughout the delay period supporting a framework of delay activity in which mnemonic representations remain relatively stable in time.

We additionally observed that broad spiking neurons contributed more to the population's representation of the sample stimulus' spatial location than narrow spiking neurons. This finding dovetails with previous reports in macaques performing the ODR task demonstrating broader tuning of narrow than broad spiking neurons and substantially higher noise correlations between narrow spiking putative interneurons than broad spiking putative pyramidal neurons (Constantinidis and Goldman-Rakic 2002). This is also consistent with the notion that the presence of correlated noise reduced the representation of spatial location in narrow relative to broad spiking neurons (Averbeck and Lee 2006).

In summary, we have shown that neural activity in the marmoset PFC can be chronically recorded in touchscreen tasks using completely unrestrained datalogger-based recording technology. In this first study, our goal was to characterize the activity profiles of PFC neurons in marmosets and to determine the distribution of sample-, delay-, and response-related activity across prefrontal regions. Subsequent studies can exploit the lissencephalic marmoset PFC to employ laminar electrophysiological (Jun et al. 2017; Johnston et al. 2019) or miniscope recordings using implanted prism lenses (Kondo et al. 2018) in touchscreen tasks to characterize the functional microcircuitry during WM tasks in different PFC regions in unrestrained marmosets. Combined experiments with optogenetic manipulations promise to provide further insights into the microcircuit mechanisms of delay-related activity in the primate PFC.

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Supplementary material

Supplementary material is available at Cerebral Cortex online.

Author contributions

RKW performed experiments, analyzed data, prepared figures, and wrote the manuscript. JS assisted in performing experiments and data analysis. KDJ assisted in surgeries and data analysis. SE designed experiments and performed surgeries. All authors edited the manuscript and SE approved the final version.

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