

G OPEN ACCESS

Citation: Li K, Wang H, Yang C, Li C, Xue B, Zhou J (2023) Clinical implication and potential function of ARHGEF6 in acute myeloid leukemia: An *in vitro* study. PLoS ONE 18(4): e0283934. https://doi.org/ 10.1371/journal.pone.0283934

Editor: Senthilnathan Palaniyandi, University of Missouri, UNITED STATES

Received: April 29, 2022

Accepted: March 20, 2023

Published: April 7, 2023

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: https://doi.org/10.1371/journal.pone.0283934

Copyright: © 2023 Li et al. This is an open access article distributed under the terms of the <u>Creative</u> <u>Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by Guangdong Basic and Applied Basic Research Foundation

RESEARCH ARTICLE

Clinical implication and potential function of ARHGEF6 in acute myeloid leukemia: An *in vitro* study

Kang Li¹, Haiquan Wang¹, Chaofan Yang¹, Chaojun Li¹*, Bin Xue²*, Jiankui Zhou³*

1 Medical School of Nanjing University, Nanjing, Jiangsu, China, 2 Core Laboratory, Sir Run Run Hospital, Nanjing Medical University, Nanjing, China, 3 Precise Genome Engineering Center, School of Life Sciences, Guangzhou University, Guangzhou, Guangdong, China

* zhoujiankui@126.com (JZ); xuebin@njmu.edu.cn (BX); licj@nju.edu.cn (CL)

Abstract

The roles of Rho GTPases in various types of cancer have been extensively studied, but the research of Rho guanine nucleotide exchange factors (GEFs) in cancer is not comprehensive. Rho guanine nucleotide exchange factor 6 (ARHGEF6) is an important member of the Rho GEFs family involved in cytoskeletal rearrangement, and it has not been investigated in acute myeloid leukemia (AML). Our research showed that the expression of ARHGEF6 was mainly higher in AML cell lines, meanwhile, was highest in the samples from patients with AML compared to other cancer types. High ARHGEF6 expression in AML was associated with a good prognosis. ARHGEF6^{low} cases showed significantly higher overall survival (OS) after autologous or allogeneic HSCT (auto/allo-HSCT). High expression of ARHGEF6 downregulates the negative regulation of myeloid differentiation process and upregulates G protein-coupled receptor signaling pathway-related processes, among which HOXA9, HOXB6, and TRH have significant differential expression and prognostic impact in AML. Therefore, ARHGEF6 can become a prognostic marker in AML; ARHGEF6^{low} patients can gain from auto/allo-HSCT.

Introduction

AML is characterized by accumulation of immature cells resulting from uncontrolled proliferation of myeloid progenitor cells, thus impairing myeloid differentiation and ultimately decrease the percentage of normal blood cells. In the United States, 75% of AML patients are over 65 years old, elder patients are refractory and prone to relapse, the recurrence rate can reach 10%-40% even in younger patients [1, 2]. Recently, several studies have shown that AML patients with some genes (such as *ARHGAP9* and *BCL2*) abnormal expression can benefit from auto/allo-HSCT [3, 4]. Therefore, optimal therapeutic strategy based on validated prognostic markers is important for AML patients.

Rho GTPases belong to the Ras GTPase family, and are activated at cellular membranes by Rho GEFs [5, 6]. Activated Rho GTPases participate in various biological processes, for instance, vesicle transport and cytoskeletal rearrangement [7, 8]. In cancer, Rho GTPases are (2020A1515110581 to J. Zhou), Science and Technology Program of Guangzhou (202201020209 to J. Zhou), start-up funds from Guangzhou University to J. Zhou, Chinese National Science Foundation (32071145 and 31771572 to B. Xue), the Nature Science Foundation of Jiangsu Province (BK20191356 to B. Xue). There was no additional external funding received for this study.

Competing interests: The authors have declared that no competing interests exist.

thought to be correlated with tumor development and poor prognoses [9]. And in hematopoiesis, Rho GTPases are concerned with various processes such as cell proliferation, differentiation, migration, and self-renewal [10-14]. Rho GTPases have significant impacts on both cancer and the hematopoietic system.

Rho GEFs are considered as prospective targets for cancer treatment, because of their functions to promote the GTP-bound state formation of Rho GTPases [15]. Although some Rho GEFs are overexpressed in cancer tissues and exhibit poor prognoses [16–18], the functional and clinical significance of most Rho GEFs remain undefined. ARHGEF6 (α PIX/Cool2) is identified as a GEF of Rac1/Cdc42, binds with PARVB and CAPNS1, participates in the regulation of cytoskeletal rearrangement, including cell adhesion and migration [19, 20]. In gliomas, ARHGEF6 overexpression correlates with tumor grading [21]. However, ARHGEF6 signaling has an essential role in apoptosis induction in chlorambucil-resistant ovarian carcinoma [22]. These instances demonstrate that the function of ARHGEF6 is distinctively according to the specific type of cancer.

Although ARHGEF6 expression has been confirmed in platelets [23], there have been no research about the expression and function of ARHGEF6 in AML. Our research showed the ARHGEF6 expression and its correlation with clinicopathological characteristics in AML. We further assessed the prognostic significance of ARHGEF6 and discussed its impact on the choice of AML treatment.

Materials and methods

ARHGEF6 expression analysis in cell lines

ARHGEF6 expression, on the mRNA level, was analyzed in the Human Protein Atlas (HPA) database (https://www.proteinatlas.org) [24]. On the entry of ARHGEF6, the "Cell Line" section based on genome-wide RNA expression was chosen, and transcriptomic data were sorted using the "Organ" parameter. ARHGEF6 expression, on the protein level, was analyzed in the Cancer Cell Line Encyclopedia (CCLE) database (https://portals.broadinstitute.org/ccle) [25]. On the entry of ARHGEF6 in the "CCLE data" section, proteomics was selected, and protein data were downloaded followed by plotting with the ggplot2 package in R 3.3.5.

Pan-cancer analysis of ARHGEF6 expression

ARHGEF6 expression pattern in pan-cancer was conducted using the UALCAN database (http://ualcan.path.uab.edu/) [26]. Briefly, "Pan-cancer view" was chosen on the entry of ARH-GEF6 and the expression of ARHGEF6 across TCGA tumors was exhibited. Pan-cancer analysis of ARHGEF6 expression pattern was also conducted by employing the Gene Expression Profiling Interactive Analysis (GEPIA) database (http://gepia.cancer-pku.cn) [27], showing the expression profile of ARHGEF6 across various tumors with paired normal tissues. The expression results in the AML are also obtained in the GEPIA database through the "Expression DIY" tool.

ARHGEF6 expression in AML with different karyotypes

The Gene Expression Omnibus (GEO) database (https://www.ncbi.nlm.nih.gov/geo) was used to get ARHGEF6 expression patterns in AML with different karyotypes. ARHGEF6 expression data in GSE14468 were retrieved from the GEO2R online software based on ARHGEF6 ID "209539_at". GSE14468 has 526 AML patients in total. The raw counts were used to generate the data in GraphPad Prism 8.0.

AML clinical data analysis

LinkedOmics database (http://www.linkedomics.orglogin.php) was used to analyze the relationship between ARHGEF6 expression and OS in AML patients [28]. Briefly, "TCGA-LAML" cancer type, "RNA-seq" data type, "ARHGEF6" gene name, and "clinical" target data type were chosen.

This study comprised a cohort of 173 AML patients having ARHGEF6 expression data from TCGA (https://cancergenome.nih.gov/ and http://www.cbioportal.org/) [29]. For consolidation treatment, 73 patients received auto/allo-HSCT, while the remaining 100 patients got just chemotherapy. Based on the mRNA level of ARHGEF6, these patients were separated into two groups (ARHGEF6^{low} and ARHGEF6^{high}) (S1 File). Table 1 summarizes the key clinical and laboratory characteristics of cases with different ARHGEF6 expressions.

Transcriptome analysis and functional annotation

RNA-seq data of AML patients were downloaded from TCGA database and normalized using the quantile normalization procedure. Differential expressed genes (DEGs) between ARH-GEF6^{high} and ARHGEF6^{low} groups were identified by t-test in the limma package. And if the adjust P value < 0.05 and log(FoldChange) (logFC) > 1, we considered the RNAs to be differentially expressed.

Enrichment analysis of Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) were performed by ClusterProfiler package in R software and we considered the terms with P value less than 0.05 to be significant.

Prognostic validation of DEGs

We validated the prognosis of DEGs between the ARHGEF6 high and low expression groups by using the survival analysis panel in GEPIA database. Briefly, we selected the "survival plot" under survival analysis, entered the gene name, selected the "LAML" dataset.

Statistical analysis

IBM SPSS 26 was used to do statistical analyses of the data. Categorical variables were compared by chi-square test and Fisher's exact tests. Because the number of samples in each group was fewer than 5000, the Shapiro–Wilk test was employed to determine if the values in each group were normally distributed for the comparison of continuous variables. A two-sample ttest or the Mann–Whitney U test was employed, depending on the values were normal/abnormal distributed, respectively. Except for the LinkedOmics database, the Log-rank in GraphPad Prism 8.0 was used to examine the prognostic impact of ARHGEF6 expression and different treatments on Disease-free survival (DFS) and OS.

Results

ARHGEF6 overexpressed in AML

To determine ARHGEF6 expression in AML cells, we analyzed RNA-seq and proteomics data in the HPA and CCLE databases, respectively. In the HPA database, ARHGEF6 mRNA expression levels in myeloid cell lines such as HEL, HL60, HMC-1, and U937 were higher than in lymphoid cell lines. Meanwhile, ARHGEF6 mRNA was almost non-existent in the lung, reproductive system, skin, and other tissues (S1A Fig). Furthermore, in the CELL database, ARH-GEF6 had the highest protein level in AML cell lines (S1B Fig).

We next examined the mRNA expression of ARHGEF6 in various human tumor samples using the UALCAN and GEPIA databases. ARHGEF6 mRNA expression level was the highest

Patient characteristics	ARHGEF6 expression		
	Low (n = 87)	High (n = 86)	р
Sex, male/female	47/40	45/41	0.823
Median age, years (range)	61 (18-88)	55.5 (21–77)	0.103
Median BM blasts, % (range)	73 (30–99)	72 (32–100)	0.709
Median WBC,(range) $\times 10^{9}$ /L	14.9 (0.7–297.4)	19.15 (0.4–223.8)	0.773
Median PB blasts, % (range)	29 (0-98)	41 (0-97)	0.162
WHO classifications AML with certain genetic abnormalities	12	26	0.009
RUNX1-RUNX1T1	1	6	0.064
CBFB-MYH11	3	7	0.211
PML-RARA	5	11	0.110
MLLT3-KMT2A	1	0	1
RBM15-MKL1	1	0	1
BCR-ABL1	1	2	0.621
AML-MRC	30	26	0.55
t-AML	NA	NA	NA
NOS	45	33	0.078
M0	0	3	0.121
M1	17	8	0.056
M2	9	12	0.467
M4	9	6	0.431
M5	9	3	0.132
M6	0	0	0
M7	1	0	1
No data	0	1	0.497
Risk level Good	9	23	0.005
Intermediate	55	46	0.194
Poor	22	15	0.208
NA	1	2	0.621

Table 1. Correlations between ARHGEF6 expression and clinicopathological characteristics in AML from the TCGA cohort.

n number of patients, *WHO* World Health Organization

AML-MRC AML with myelodysplasia-related changes

t-AML Therapy-related AML, NOS not otherwise specified

BM-blast bone marrow blast

WBC white blood cell

PB-blast peripheral blood blast

https://doi.org/10.1371/journal.pone.0283934.t001

across all kinds of cancers (Fig 1A and 1B). Moreover, in the GEPIA database, a significantly higher ARHGEF6 expression level was found in AML compared to normal tissues (Fig 1C). Then microarray data (GSE14468) was utilized to find out whether ARHGEF6 expression was associated with major recurrent chromosomal translocations in the GEO database. The result showed that AML patients with t(8;21) had the highest ARHGEF6 expression compared with other karyotypes (Fig 1D).

The association of ARHGEF6 with clinicopathological characteristics of AML patients

Table 1 summed up the clinical characteristics of patients according to the clinical data from the TCGA database. The WHO classification and risk stratification were significantly different

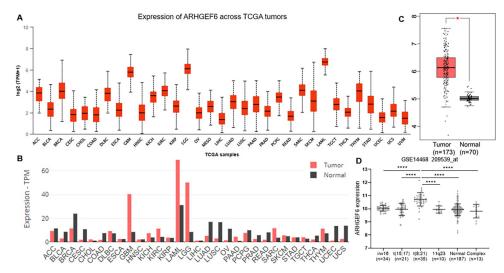


Fig 1. ARHGEF6 expression in AML. (A) ARHGEF6 expression in pan-cancer, result from UALCAN. (B) The expression of ARHGEF6 in different cancerous and normal tissues using the GEPIA. Bar height refers to the value of the median expression. (C) ARHGEF6 expression between AML and normal samples, result was obtained by GEPIA analysis. *, p < 0.05. (D) ARHGEF6 expression in AML among various karyotypes, according to GSE14468 in the GEO database. ****, p < 0.0001.

https://doi.org/10.1371/journal.pone.0283934.g001

in patients with different ARHGEF6 expressions. In the WHO classification distribution, high ARHGEF6 expression was significantly associated with genetic abnormalities. In addition, patients with high ARHGEF6 expression tended to have good prognoses. Meanwhile, there was no significant difference in age, sex, BM blasts, WBC, or PB cells between the ARHGE-F6^{low} and ARHGEF6^{high} cases (Table 1).

Prognostic values of ARHGEF6 in AML

To investigate the relations between ARHGEF6 expression and prognosis in AML patients, we assessed the effect of ARHGEF6 expression on OS by employing the LinkedOmics database. The result indicated that low ARHGEF6 expression in AML was associated with poor OS (Fig 2A). Meanwhile, the survival data from the TCGA database was analyzed, as a result, ARH-GEF6 overexpression was significantly associated with higher OS (Fig 2B). The DFS was higher in the ARHGEF6^{high} cases compared with the ARHGEF6^{low} cases, although it did not reach statistical significance (Fig 2C).

In cytogenetically normal AML (CN-AML), patients with different ARHGEF6 expressions had no significant difference in OS or DFS (Fig 2D and 2E). Furthermore, no significant differences in OS or DFS was detected between the ARHGEF6^{low} and ARHGEF6^{high} groups with either chemotherapy or chemotherapy plus auto/allo-HSCT treatment (Fig 2F–2I). Overall, low ARHGEF6 expression in AML exhibited a poor prognosis. However, neither chemotherapy alone nor chemotherapy combined with auto/allo-HSCT improved the prognosis of ARH-GEF6^{low} patients. Finally, in ARHGEF6^{low} groups, patient survival with or without (WOW) auto/allo-HSCT were analyzed. The result showed that ARHGEF6^{low} patients who received auto/allo-HSCT had significantly improved OS, but not DFS, than patients undergoing only chemotherapy (Fig 2J and 2K).

Functional annotation of DEGs between ARHEGF6^{low} and ARHGEF6^{high} group

To find the function of ARHGEF6 in AML, we analyzed the RNA-seq data of the ARHGEF6 low and high expression groups. We obtained a total of 504 DEGs, of which 163 genes were

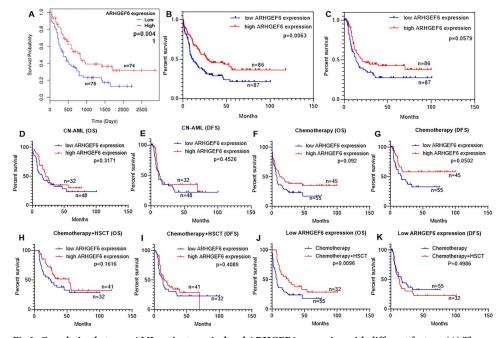


Fig 2. Correlation between AML patient survival and ARHGEF6 expression with different factors. (A) The prognostic value between ARHGEF6^{low} and ARHGEF6^{high} group using the LinkedOmics database. (B) OS and (C) DFS in AML patients with different ARHGEF6 expressions. (D) OS and (E) DFS in CN-AML patients with different ARHGEF6 expressions. (F) OS and (G) DFS of ARHGEF6^{low} and ARHGEF6^{high} patients treated with chemotherapy. (H) OS and (I) DFS of ARHGEF6^{low} and ARHGEF6^{high} patients undergoing chemotherapy + auto/allo-HSCT. (J) OS and (K) DFS of patients treated WOW auto/allo-HSCT in the ARHGEF6^{low} group.

https://doi.org/10.1371/journal.pone.0283934.g002

significantly up-regulated and 341 genes were significantly down-regulated (S2 File), and we marked the top 9 significantly highly expressed and top 10 significantly lowly expressed genes, respectively (Fig 3A).

GO analysis annotated Molecular Function (MF), Cellular Component (CC), and Biological Process (BP) that were significantly up- and down-regulated with increasing ARHGEF6 expression levels, respectively (Fig <u>3B</u> and <u>3C</u>, <u>S3</u> File). The results showed that the significantly up-regulated genes were mainly involved in the classical regulatory pathways of GEFs, such as G protein-coupled receptor signaling pathway, synapse assembly, and vesicle transport. The significantly down-regulated genes were mainly involved in the differentiation and development of the embryonic skeletal system and the negative regulation of myeloid differentiation.

KEGG pathway annotation showed that the significantly up-regulated pathways mainly involved circadian entrainment, renin secretion, vascular smooth muscle contraction, transcriptional misregulation in cancer, etc.; the significantly down-regulated pathways included osteoclast differentiation, B cell receptor signaling pathway, cytokine receptor interaction, signaling pathways regulating pluripotency of stem cells, hippo and wnt signaling pathway, etc (Fig 3D, S4 File).

Prognosis validation of DEGs

We validated the prognostic profile of AML for the top 10 genes that were significantly upand down-regulated, and the results showed that high expression of the top 10 genes that were significantly up-regulated in patients were all associated with higher OS, among which, *TRH* significantly high expressed in AML samples compared to normal samples (Fig 3E), and high

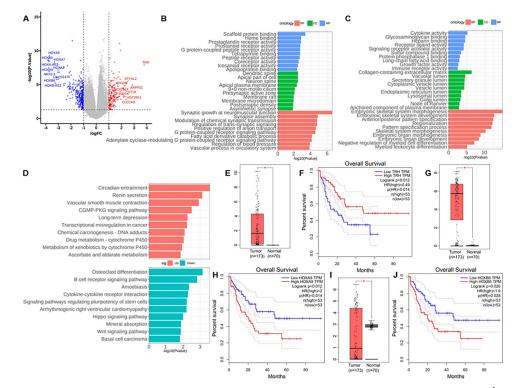


Fig 3. Functional annotation and prognosis validation of differentially expressed genes between ARHEGF6^{low} and **ARHGEF6**^{high} group. (A) Volcano plot of DEGs between the ARHGEF6^{low} and ARHGEF6^{high} groups. (B) GO analysis of significantly upregulated genes. (C) GO analysis of significantly downregulated genes. (D) KEGG analysis of DEGs. (E) and (F) TRH expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05. (G) and (H) HOXA9 expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05. (I) and (J) HOXB6 expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05. (I) and (J) HOXB6 expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05. (I) and (J) HOXB6 expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05. (I) and (J) HOXB6 expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05. (I) and (J) HOXB6 expression levels in AML and normal tissues, and the effect on OS in AML patients, result from GEPIA. *, p < 0.05.

https://doi.org/10.1371/journal.pone.0283934.g003

expression of *TRH* in AML patients was a good prognostic factor that was significantly associated with higher OS (Fig 3F); High expression of the significantly down-regulated genes were shown to be associated with lower OS in AML patients, among which, *HOXA9* and *HOXB6* having significantly different expression profiles in AML samples compared to normal samples (Fig 3G and 3I, respectively), and high expression in AML were poor prognostic factors (Fig 3H and 3J).

Discussion

Rho GEFs activate Rho GTPases by exchanging bound GDP with GTP [30]. Numerous studies have pointed out that abnormal expression of Rho GEFs has been found in human cancers [31, 32]. Rho GEFs expression is distinct in different cancer types. For example, DOCK2 is upregulated in follicular lymphoma and downregulated in NSCLC (non-small cell lung cancer) [33, 34]. Tiam1 is downregulated in colorectal cancer and highly expressed in various cancers such as gastric, laryngeal squamous cell carcinoma, and ovarian cancers [35–37]. In the current study, ARHGEF6 was highly expressed among AML tissues and cell lines, especially in t(8;21) AML patients with relatively good prognoses.

Some studies on Rho GEFs have shown that a large part of Rho GEFs are correlated with poor prognosis in various tumors [38–40]. For example, ABR, PREX1, DOCK2, and DOCK4 showed poor prognosis in NSCLC [31]. But, in AML, the high expression group of ARHGEF6

had a good prognosis. Thus, Rho GEFs play different roles in different cancers. On the other hand, ARHGAP9, which belongs to the Rho GAP family, inactivates GTPases and oppositely function to that of Rho GEFs. ARHGAP9 is an adverse prognostic factor for AML OS [3]. Thus, Rho GEFs and Rho GAPs may play other roles in cancers.

Despite the significantly lower expression of ARHGEF6 in CN-AML, no relationship between ARHGEF6 expression and CN-AML prognosis has been found. Furthermore, several researchers found that some genes, for example, ARHGAP9 and BCL2 were correlated with the prognosis of auto/allo-HSCT and/or chemotherapy in AML patients [3, 4]. Our findings showed that auto/allo-HSCT significantly improves prognosis in patients with low ARHGEF6 expression. However, in the high ARHGEF6 expression group, patients treated with chemotherapy + auto/allo-HSCT had a significant decrease in DFS, although there was no significant change in OS. These results suggest that patients with low ARHGEF6 expression could benefit from autologous/allogeneic HSCT, but autologous/allogeneic HSCT is not recommended for patients with high ARHGEF6 expression because of the significantly higher tendency to relapse and progression.

Interestingly, after analysis of patient RNA-seq data, we found that high expression of ARHGEF6 was associated with the downregulation of several HOX gene family members. the HOX gene family plays a crucial regulatory role in animal development [41], and in hematopoiesis, the HOX gene family is involved in regulating the differentiation and developmental process of hematopoietic stem cells to different cell types [42], and the disturbances in their expression levels are associated with the development of AML [43]. Previous studies have identified that HOXA9 overexpressed in AML and is a poor prognostic factor [44], consistent with our results. In vivo, HOXB6 promotes the development of AML by promoting the proliferation of hematopoietic stem cells and myeloid precursor cells while inhibiting the production of erythroid and lymphocytes [45]. On the other hand, the significantly upregulated gene TRH was found to be associated with a good prognosis in a study of t(8;21) acute myeloid leukemia [46]. However, there are no studies about the role of TRH in AML.

Overall, we first reported that ARHGEF6 overexpressed in AML cell lines, tissues, and the t (8; 21) patients. Elevated ARHGEF6 expression was significantly correlated with a favorable prognosis in AML. A combination of auto/allo-HSCT and chemotherapy, instead of only chemotherapy, can improve poor prognosis related to low ARHGEF6 expression. High expression of ARHGEF6 downregulated the expression level of the poor-prognosis HOXA9 and HOXB6, while increasing the expression of the good-prognosis TRH, thus improving the OS of patients.

Supporting information

S1 Fig. ARHGEF6 expression in cell lines. (A) The mRNA levels of ARHGEF6 in human cell lines using the HPA. (B) The relative protein levels of ARHGEF6 in human cell lines using the CCLE. Labels of x-axis were sorted from large to small according to the median expression of protein.

(TIF)

S1 File. ARHGEF6^{low} **and ARHGEF6**^{high} **group information.** Patients were divided into ARHGEF6^{low} group (n = 87) and ARHGEF6^{high} group (n = 86) according to the mRNA expression level of ARHGEF6. (TXT)

S2 File. DEGs list. DEGs between ARHGEF6^{low} and ARHGEF6^{high} groups. (XLSX)

S3 File. DEGs GO. GO analysis of the DEGs in S2 File. (XLSX)

S4 File. DEGs KEGG. KEGG analysis of the DEGs in S2 File. (XLSX)

Acknowledgments

We thank Guangzhou University for supporting the study.

Author Contributions

Conceptualization: Kang Li.

Data curation: Haiquan Wang, Chaofan Yang.

Formal analysis: Chaofan Yang.

Funding acquisition: Bin Xue, Jiankui Zhou.

Investigation: Kang Li.

Project administration: Bin Xue, Jiankui Zhou.

Resources: Chaojun Li, Jiankui Zhou.

Supervision: Chaojun Li.

Visualization: Haiquan Wang.

Writing – original draft: Kang Li.

Writing - review & editing: Jiankui Zhou.

References

- Turbeville S, Francis KM, Behm I, Chiu GR, Sanchez H, Morrison BA, et al. Prevalence and incidence of acute myeloid leukemia may be higher than currently accepted estimates among the ≥65 year-old population in the United States. Blood 2014; 124(21):958–958.
- Dohner H, Weisdorf DJ, Bloomfield CD. Acute Myeloid Leukemia. N Engl J Med 2015; 373(12):1136– 52. https://doi.org/10.1056/NEJMra1406184 PMID: 26376137
- Han C, He S, Wang R, Gao X, Wang H, Qiao J, et al. The role of ARHGAP9: clinical implication and potential function in acute myeloid leukemia. Journal of Translational Medicine 2021; 19(1). https://doi. org/10.1186/s12967-021-02733-5 PMID: 33579308
- Zhou J, Zhang T, Xu Z, Gu Y, Ma J, Li X, et al. BCL2 overexpression: clinical implication and biological insights in acute myeloid leukemia. Diagnostic Pathology 2019; 14(1). <u>https://doi.org/10.1186/s13000-019-0841-1</u> PMID: <u>31253168</u>
- Wennerberg K, Rossman KL, Der CJ. The Ras superfamily at a glance. J Cell Sci 2005; 118(Pt 5):843– 6. https://doi.org/10.1242/jcs.01660 PMID: 15731001
- Heasman SJ, Ridley AJ. Mammalian Rho GTPases: new insights into their functions from in vivo studies. Nat Rev Mol Cell Biol 2008; 9(9):690–701. https://doi.org/10.1038/nrm2476 PMID: 18719708
- Buchsbaum RJ. Rho activation at a glance. J Cell Sci 2007; 120(Pt 7):1149–52. https://doi.org/10.1242/ jcs.03428 PMID: 17376960
- Schwartz M. Rho signalling at a glance. J Cell Sci 2004; 117(Pt 23):5457–8. https://doi.org/10.1242/jcs. 01582 PMID: 15509861
- Crosas-Molist E, Samain R, Kohlhammer L, Orgaz JL, George SL, Maiques O, et al. Rho GTPase signaling in cancer progression and dissemination. Physiol Rev 2022; 102(1):455–510. https://doi.org/10. 1152/physrev.00045.2020 PMID: 34541899
- David MD, Petit D, Bertoglio J. The RhoGAP ARHGAP19 controls cytokinesis and chromosome segregation in T lymphocytes. J Cell Sci 2014; 127(Pt 2):400–10. https://doi.org/10.1242/jcs.135079 PMID: 24259668

- Florian MC, Dorr K, Niebel A, Daria D, Schrezenmeier H, Rojewski M, et al. Cdc42 activity regulates hematopoietic stem cell aging and rejuvenation. Cell Stem Cell 2012; 10(5):520–30. https://doi.org/10. 1016/j.stem.2012.04.007 PMID: 22560076
- Mulloy JC, Cancelas JA, Filippi MD, Kalfa TA, Guo F, Zheng Y. Rho GTPases in hematopoiesis and hemopathies. Blood 2010; 115(5):936–47. <u>https://doi.org/10.1182/blood-2009-09-198127</u> PMID: 19965643
- Wang L, Yang L, Filippi MD, Williams DA, Zheng Y. Genetic deletion of Cdc42GAP reveals a role of Cdc42 in erythropoiesis and hematopoietic stem/progenitor cell survival, adhesion, and engraftment. Blood 2006; 107(1):98–105. https://doi.org/10.1182/blood-2005-05-2171 PMID: 16174757
- Yang L, Wang L, Geiger H, Cancelas JA, Mo J, Zheng Y. Rho GTPase Cdc42 coordinates hematopoietic stem cell quiescence and niche interaction in the bone marrow. Proc Natl Acad Sci U S A 2007; 104 (12):5091–6. https://doi.org/10.1073/pnas.0610819104 PMID: 17360364
- Maldonado M, Medina JI, Velazquez L, Dharmawardhane S. Targeting Rac and Cdc42 GEFs in Metastatic Cancer. Front Cell Dev Biol 2020; 8:201. <u>https://doi.org/10.3389/fcell.2020.00201</u> PMID: 32322580
- Wertheimer E, Gutierrez-Uzquiza A, Rosemblit C, Lopez-Haber C, Sosa MS, Kazanietz MG. Rac signaling in breast cancer: a tale of GEFs and GAPs. Cell Signal 2012; 24(2):353–362. https://doi.org/10. 1016/j.cellsig.2011.08.011 PMID: 21893191
- Porter AP, Papaioannou A, Malliri A. Deregulation of Rho GTPases in cancer. Small GTPases 2016; 7 (3):123–38. https://doi.org/10.1080/21541248.2016.1173767 PMID: 27104658
- Kazanietz MG, Caloca MJ. The Rac GTPase in Cancer: From Old Concepts to New Paradigms. Cancer Res 2017; 77(20):5445–5451. https://doi.org/10.1158/0008-5472.CAN-17-1456 PMID: 28807941
- Rosenberger G, Jantke I, Gal A, Kutsche K. Interaction of alphaPIX (ARHGEF6) with beta-parvin (PARVB) suggests an involvement of alphaPIX in integrin-mediated signaling. Hum Mol Genet 2003; 12(2):155–67. https://doi.org/10.1093/hmg/ddg019 PMID: 12499396
- Rosenberger G, Gal A, Kutsche K. αPIX Associates with Calpain 4, the Small Subunit of Calpain, and Has a Dual Role in Integrin-mediated Cell Spreading. Journal of Biological Chemistry 2005; 280 (8):6879–6889.
- Yokota T, Kouno J, Adachi K, Takahashi H, Teramoto A, Matsumoto K, et al. Identification of histological markers for malignant glioma by genome-wide expression analysis: dynein, α-PIX and sorcin. Acta Neuropathologica 2006; 111(1):29–38.
- Maiti AK. Gene network analysis of oxidative stress-mediated drug sensitivity in resistant ovarian carcinoma cells. The pharmacogenomics journal 2010; 10(2):94–104. <u>https://doi.org/10.1038/tpj.2009.49</u>
 PMID: 19918261
- Nagy Z, Wynne K, von Kriegsheim A, Gambaryan S, Smolenski A. Cyclic Nucleotide-dependent Protein Kinases Target ARHGAP17 and ARHGEF6 Complexes in Platelets. J Biol Chem 2015; 290 (50):29974–83. https://doi.org/10.1074/jbc.M115.678003 PMID: 26507661
- 24. Thul PJ, Lindskog C. The human protein atlas: A spatial map of the human proteome. Protein Sci 2018; 27(1):233–244. https://doi.org/10.1002/pro.3307 PMID: 28940711
- Barretina J, Caponigro G, Stransky N, Venkatesan K, Margolin AA, Kim S, et al. The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. Nature 2012; 483(7391):603– 7. https://doi.org/10.1038/nature11003 PMID: 22460905
- Chandrashekar DS, Bashel B, Balasubramanya S, Creighton CJ, Ponce-Rodriguez I, Chakravarthi B, et al. UALCAN: A Portal for Facilitating Tumor Subgroup Gene Expression and Survival Analyses. Neoplasia 2017; 19(8):649–658. https://doi.org/10.1016/j.neo.2017.05.002 PMID: 28732212
- Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. Nucleic Acids Res 2017; 45(W1):W98–W102. https://doi.org/10. 1093/nar/gkx247 PMID: 28407145
- Vasaikar SV, Straub P, Wang J, Zhang B. LinkedOmics: analyzing multi-omics data within and across 32 cancer types. Nucleic Acids Res 2018; 46(D1):D956–D963. <u>https://doi.org/10.1093/nar/gkx1090</u> PMID: 29136207
- Ley TJ, Miller C, Ding L, Raphael BJ, Mungall AJ, Robertson A, et al. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. N Engl J Med 2013; 368(22):2059–74. <u>https://doi.org/ 10.1056/NEJMoa1301689 PMID: 23634996</u>
- Zheng Y. Dbl family guanine nucleotide exchange factors. Trends Biochem Sci 2001; 26(12):724–32. https://doi.org/10.1016/s0968-0004(01)01973-9 PMID: 11738596
- Zeng RJ, Xie WJ, Zheng CW, Chen WX, Wang SM, Li Z, et al. Role of Rho guanine nucleotide exchange factors in non-small cell lung cancer. Bioengineered 2021; 12(2):11169–11187. https://doi. org/10.1080/21655979.2021.2006519 PMID: 34783629

- 32. Robles-Valero J, Lorenzo-Martín LF, Fernández-Pisonero I, Bustelo XR. Rho guanosine nucleotide exchange factors are not such bad guys after all in cancer. Small GTPases 2020; 11(4):233–239.
- Wang L, Nishihara H, Kimura T, Kato Y, Tanino M, Nishio M, et al. DOCK2 regulates cell proliferation through Rac and ERK activation in B cell lymphoma. Biochem Biophys Res Commun 2010; 395 (1):111–5. https://doi.org/10.1016/j.bbrc.2010.03.148 PMID: 20350533
- Hu N, Pang Y, Zhao H, Si C, Ding H, Chen L, et al. High expression of DOCK2 indicates good prognosis in acute myeloid leukemia. J Cancer 2019; 10(24):6088–6094. <u>https://doi.org/10.7150/jca.33244</u> PMID: <u>31762818</u>
- Diamantopoulou Z, White G, Fadlullah M, Dreger M, Pickering K, Maltas J, et al. TIAM1 Antagonizes TAZ/YAP Both in the Destruction Complex in the Cytoplasm and in the Nucleus to Inhibit Invasion of Intestinal Epithelial Cells. Cancer Cell 2017; 31(5):621–634.e6. https://doi.org/10.1016/j.ccell.2017.03. 007 PMID: 28416184
- 36. Wang S, Li S, Tang Q, Yang S, Wang S, Liu J, et al. Overexpression of Tiam1 promotes the progression of laryngeal squamous cell carcinoma. Oncol Rep 2015; 33(4):1807–14. <u>https://doi.org/10.3892/or.</u> 2015.3785 PMID: 25672412
- Li H, Cui X, Chen D, Yang Y, Piao J, Lin Z, et al. Clinical implication of Tiam1 overexpression in the prognosis of patients with serous ovarian carcinoma. Oncol Lett 2016; 12(5):3492–3498. <u>https://doi.org/10.3892/ol.2016.5091</u> PMID: 27900026
- Komiya Y, Onodera Y, Kuroiwa M, Nomimura S, Kubo Y, Nam JM, et al. The Rho guanine nucleotide exchange factor ARHGEF5 promotes tumor malignancy via epithelial-mesenchymal transition. Oncogenesis 2016; 5(9):e258. https://doi.org/10.1038/oncsis.2016.59 PMID: 27617642
- Li X, Jiang M, Chen D, Xu B, Wang R, Chu Y, et al. miR-148b-3p inhibits gastric cancer metastasis by inhibiting the Dock6/Rac1/Cdc42 axis. J Exp Clin Cancer Res 2018; 37(1):71. <u>https://doi.org/10.1186/</u> s13046-018-0729-z PMID: 29587866
- Lane J, Martin TA, Mansel RE, Jiang WG. The expression and prognostic value of the guanine nucleotide exchange factors (GEFs) Trio, Vav1 and TIAM-1 in human breast cancer. Int Semin Surg Oncol 2008; 5:23. https://doi.org/10.1186/1477-7800-5-23 PMID: 18925966
- 41. Mallo M, Alonso CR. The regulation of Hox gene expression during animal development. Development (Cambridge) 2013; 140(19):3951–3963. https://doi.org/10.1242/dev.068346 PMID: 24046316
- Alsayegh K, Cortes-Medina LV, Ramos-Mandujano G, Badraiq H, Li M. Hematopoietic Differentiation of Human Pluripotent Stem Cells: HOX and GATA Transcription Factors as Master Regulators. Curr Genomics 2019; 20(6):438–452. https://doi.org/10.2174/1389202920666191017163837 PMID: 32194342
- De Braekeleer E, Douet-Guilbert N, Basinko A, Le Bris M, Morel F, De Braekeleer M. Hox gene dysregulation in acute myeloid leukemia. Future oncology (London, England) 2014; 10(3):475–495. <u>https://</u> doi.org/10.2217/fon.13.195 PMID: 24559452
- ANDREEFF M, RUVOLO V, GADGIL S, ZENG C, COOMBES K, CHEN W, et al. HOX expression patterns identify a common signature for favorable AML. Leukemia 2008; 22(11):2041–2047. https://doi. org/10.1038/leu.2008.198 PMID: 18668134
- 45. Fischbach NA, Rozenfeld S, Shen W, Fong S, Chrobak D, Ginzinger D, et al. HOXB6 overexpression in murine bone marrow immortalizes a myelomonocytic precursor in vitro and causes hematopoietic stem cell expansion and acute myeloid leukemia in vivo. Blood 2005; 105(4):1456–66. <u>https://doi.org/10. 1182/blood-2004-04-1583 PMID: 15522959</u>
- 46. Li X, Dai Y, Chen B, Huang J, Chen S, Jiang L. Clinical significance of CD34(+)CD117(dim)/CD34(+) CD117(bri) myeloblast-associated gene expression in t(8;21) acute myeloid leukemia. Front Med 2021; 15(4):608–620. https://doi.org/10.1007/s11684-021-0836-7 PMID: 33754282