

The function of S100A4 in pulmonary disease

A review

Ting Wang, MD^{a,*} 

Abstract

S100 protein family, which represents 25 relatively small calcium binding proteins, is involved in many intracellular and/or extracellular processes, including differentiation, apoptosis, migration/invasion, Ca²⁺ homeostasis, inflammation, and tissue repair. As an important member, S100A4 was reported to have an abnormal expression in several lung diseases, such as lung cancer, pulmonary hypertension, idiopathic pulmonary fibrosis (IPF), etc. For example, in lung cancer, S100A4 was demonstrated to be associated to metastatic tumor progression and epithelial to mesenchymal transition (EMT). In IPF, S100A4 was considered as a promising serum biomarker predicting disease progression. Various studies in recent years focused on the S100A4 function in lung diseases, showing researchers' interests on this protein. It is necessary to focus on relative studies, and make a comprehensive understanding of S100A4 in common pulmonary diseases. By doing this, this paper provides a review of the evidence for S100A4 in lung cancer, chronic obstructive pulmonary disease (COPD), asthma, IPF and pulmonary hypertension.

Abbreviation: COPD = chronic obstructive pulmonary disease, EMT = epithelial to mesenchymal transition, HA = hyaluronan, IPF = idiopathic pulmonary fibrosis, MPC = mesenchymal progenitor cells, PAH = pulmonary arterial hypertension, PA-SMC = protein in pulmonary arterial smooth muscle cells.

Keywords: asthma, chronic obstructive pulmonary disease, idiopathic pulmonary fibrosis, lung cancer, S100A4

1. Introduction

The S100 protein family, which was discovered firstly by Moore et al in 1965, have been proved to have 25 small acidic Ca²⁺ combined with cytotoxic proteins.^[1] Two founding members, S100A1 and S100B, were found to be soluble in 100% saturated ammonium sulfate at that time, thus this family was named S100.^[2,3] As a largest one in the EF-hand superfamily, S100 proteins implicated in multiple intracellular and/or extracellular regulatory activities. According to the roles in Calcium regulation, S100 members could be divided into 2 groups.^[4] The first group gets involved in translating the signals via detecting the Ca²⁺ ions levels by calcium sensors. The second one, which consists of Ca²⁺ buffer, could bind free cell cytoplasm Ca²⁺ ions and modulate the calcium signals.^[5] Thus, these second messenger mediators participate in the control of an array of cellular processes ranging from muscle contraction to cell behaviors. In a variety of diseases, S100s have been reported to have an abnormal expression. S100A2, for example, was firstly found to act as a tumor suppressor gene, having a downregulated expression in skin, lung, kidney and prostate tumors.^[6–8] On the other hand, S100A2 was found to have an upregulation in some cancer types as well, including pancreatic cancer, gastric cancer and epithelial ovarian cancer. S100A6 has an elevated expression in the serum of gastric cancer patients^[9]; S100A7 levels have been found to be augmented in cerebrospinal fluid of patients with Alzheimer disease^[10]; blood levels of S100A8/9 were reported

to be increased in obesity and coronary artery diseases^[11]; S100A12 has been reported to have an enhanced expression in inflammatory diseases and diabetes.^[12] In pulmonary diseases, such as asthma, chronic obstructive pulmonary disease (COPD), idiopathic pulmonary fibrosis (IPF), cystic fibrosis, pulmonary hypertension, and lung cancer, S100 family members have also been observed to have dysregulated responses. A quantity of studies in recent years were undertaken to identify the exact role of this family in pulmonary disease pathogenesis and therapy. Of all the S100 members, S100A4 is the most extensively studied, and has been given many names, such as PEL-98, 18A2, 42A, CAPL, P9KA, metastasin (MTS-1), etc.^[13] The human S100A4 gene is located in the epidermal differentiation complex on chromosome 1q21, which is prone to chromosomal rearrangements, thus the encoded protein is involved in many physiological functions, including cell proliferation, invasion and metastasis.^[14] Meanwhile, its function in multiple pulmonary diseases has been confirmed.^[15] In this review, we summarize the evidence concerning S100A4 and pulmonary diseases and discuss the mechanisms through which S100A4 plays its diverse functions in common lung diseases.

2. Lung cancer

S100A4 was firstly reported to have an association with metastatic tumor in 1989. Ebralidze MS et al detected the

This study was supported by Key Research and Development Program of Shaanxi Province (Grant No. 2022SF-539; 2022JM-531).

The authors have no conflicts of interest to disclose.

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

^a Department of Respiratory Medicine, Xi'an People's Hospital (Xi'an No. 4 Hospital), Xi'an, China.

*Correspondence: Ting Wang, Department of Respiratory Medicine, Xi'an People's Hospital (Xi'an No. 4 Hospital), Xi'an 710004, China (e-mail: hdtw.ok@163.com).

Copyright © 2023 the Author(s). Published by Wolters Kluwer Health, Inc.

This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Wang T. The function of S100A4 in pulmonary disease: A review. *Medicine* 2023;102:14(e33466).

Received: 21 February 2023 / Accepted: 16 March 2023

<http://dx.doi.org/10.1097/MD.00000000000033466>

expression of *mts1* in the metastatic phenotype of different transformed and normal cells using Northern blot analysis, and results showed that the overexpression of *mts1* was associated with a high degree of metastasis.^[16] Relative researches later on found that the high level of S100A4 implicates in metastasis and progression of various malignancies; it could also be used as a marker in epithelial to mesenchymal transition (EMT).^[17-19] In lung cancer, Takenaga K et al firstly demonstrated that pEL98 (*mts1*) expression was related to invasive and motile abilities in clones derived from lung carcinoma, and they drew a conclusion that pEL98 (*mts1*) had a function in regulating cell invasiveness and tumor cell motility.^[20] Their later research found cell motility and in vitro invasiveness were suppressed in the antisense S100A4 RNA-expressing Lewis lung carcinoma, supporting the above conclusion from the reverse side.^[21] Compared with leading-edge peripheral parts, S100A4, along with other mesenchymal markers, had a lower expression in the central region of non-small cell lung cancer. Moreover, its high expression was proved to be related to poor differentiation and advanced stage of adeno- and squamous cell carcinoma. Concerning of the exact mechanism of S100A4 in lung cancer, Stewart RL et al demonstrated the knockdown of S100A4 could inhibit NF- κ B activity and decreased TNF α -induced MMP9 expression (Fig. 1), indicating these downstream molecules in its regulation of lung cancer cellular biological activities.^[22] A recent study found that the overexpression of S100A4 could also enhance cell proliferation and inhibit starvation-induced autophagy via the Wnt/ β -catenin signaling pathway in A549 cells.^[23] In another study of NSCLC cell lines, S100A4 was also proved to up-regulate mitochondrial complex I subunit NADH dehydrogenase (ubiquinone) Fe-S protein 2, thus promoting cells' invasion and altering metabolism (Fig. 1).^[24] Researches focusing on the molecules that regulate S100A4, on the other hand, found that plakoglobin, a tumor/metastasis suppressor, restored its tumor suppressor activity in NSCLC cells by upregulating S100A4 with P53 mutants (H1299).^[25] In another research, a long noncoding RNA (linc01833) was proved to adsorb miR-519e-3p through a sponge and regulate S100A4 in lung adenocarcinoma progression.^[26]

3. Chronic obstructive pulmonary disease

COPD is characterized with chronic airway inflammation and progressive fixed airflow limitation, usually having an association

with tobacco smoke inhalation.^[27] An explanation for airway remodeling in chronic inflammatory respiratory diseases, such as COPD, is involving with the differentiation of airway epithelial cells to a mesenchymal phenotype, which is also named EMT.^[28] In 2010, researchers firstly examined EMT markers, including S100A4, in airway biopsy tissue from subjects with COPD.^[29] Compared with control groups, obviously increased reticular basement membrane cell S100A4 staining could be seen in smokers with COPD. Their later studies in 2011 and 2014 found that S100A4 had a significantly high expression in basal epithelium compared to infiltrating macrophages, fibroblasts, as well as immune cells, and S100A4 positive cell numbers sharply decreased after the treatment of inhaled fluticasone propionate, providing additional support for active EMT and anti-EMT effects of inhaled corticosteroids in COPD.^[30,31] Further study conducted by Mahmood MQ et al showed S100A4 tended to have an increased expression in small airway, compared to large airways, of subjects with chronic airflow limitation.^[32] An elevated expression of S100A4 could also be found in serum, tissues and vasculature of patients with COPD. Reimann S et al performed real-time RT-PCR analysis and immunohistochemistry to investigate the expression of S100A4 in laser-microdissected intrapulmonary arteries of COPD patients, and data revealed S100A4 had an enhanced expression at both levels.^[33] These findings were mirrored by Enzyme-Linked Immunosorbent Assay analysis of S100A4 in the serum of patients with COPD.^[34] Moreover, serum S100A4 was inversely related to pulmonary function among COPD patients. Concerning of the mechanism, the study of Jiang B et al demonstrated that exposure of the epithelium to cigarette smoke extract and exposure of the mice to cigarette smoke can induce EMT by activating the Akt signaling pathway, suggesting this pathway could be a regulator for S100A4 in COPD (Fig. 1).^[35]

4. Asthma

Asthma, which is one of the most common forms of respiratory disease, is characterized by immune hyper-responsiveness, airway inflammation, eosinophilic infiltration and mucus hypersecretion.^[36,37] It has been demonstrated that damage-associated molecular pattern can activate the innate immune response through interacting with pattern recognition receptors. S100A4 could act as damage-associated molecular pattern once secreted by the action of specific chemokines, stress or necrosis.^[38] Related studies found that S100A4 had an elevated

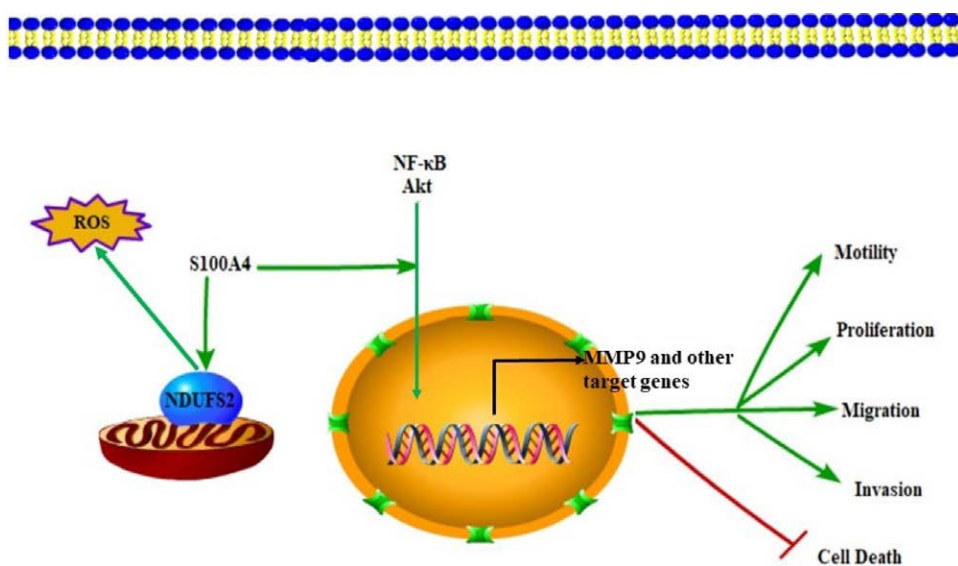


Figure 1. S100A4 proteins get involved in NF- κ B and AKT signaling, affecting the cell activities.

levels in several inflammatory diseases such as arthritis, kidney fibrosis and neuronal injury.^[39,40] In chondrocytes, stimulation of T cells with S100A4 was proved to have an increased production of cytokines, especially eotaxin-2 and granulocyte colony-stimulating factor, both of which are important factors in the pathogenesis of asthma. In 2018, Huang X et al tested the expression in induced sputum and plasma from asthmatics and healthy controls. Data showed that S100A4 was overexpressed in the sputum rather than in plasma in asthmatics, and its high expression was associated negatively with some lung function parameters and were correlated positively with sputum lymphocyte and eosinophilia. In the asthma mouse model, the expression of S100A4 was also significantly higher in the lung as well as in BALF. Further study showed that LY294002, a PI3K inhibitor, could decrease S100A4 in both lung and BLAF markedly in asthmatic mice.^[41] In mast cell, S100A4 gene deficiency was identified to dampen its activation both in vitro and in vivo, suggesting S100A4 may participate in the regulation of allergic responses through regulating the activation of mast cells. Furthermore, the airway remodeling in the chronic asthma model was confirmed to be attenuated by inhibition of soluble epoxide hydrolase, dapagliflozin and ZDHXB-101 (3',5-Diallyl-2, 4'-dihydroxy-[1,1'-biphen-yl]-3,5'-dicarbaldehyde), and the expression of remodeling-related molecular markers reduced, including S100A4.^[42-44] An in-depth study showed that the synthesis and secretion of S100A4 in airway smooth muscle tissues could be stimulated by inflammatory mediators and that extracellular S100A4 acted via RAGE to mediate airway smooth muscle inflammation.^[45]

5. Idiopathic pulmonary fibrosis

IPF is a progressive and lethal fibrotic lung disease characterized by alveolar epithelial cell injury and activation, formation of myofibroblast foci, and exaggerated extracellular matrix accumulation in the lung parenchyma.^[46,47] Patients are often behaved progressive dyspnea and restrictive physiology on pulmonary function testing.^[48] In 2010, Degryse AL et al developed lung fibrosis mice model by intratracheal bleomycin. Data showed that 50% of S100A4 + lung fibroblasts were derived from epithelial mesenchymal transition in those mice models established by repetitive bleomycin, which had greater fibrosis by scoring, morphometry and collagen content, compared with 33% in the single-dose model. In bronchoalveolar lavage fluid, numbers of S100A4 + macrophages were proved to be correlated well with S100A4 protein levels and the occurrence of IPF.^[48] The study of Zhang W et al, data suggested that S100A4 was produced and secreted by M2 polarized alveolar macrophages.^[49] Moreover, in vitro, extracellular S100A4 were revealed to activate both mouse and human lung fibroblasts via upregulating α -SMA and type I collagen, during which sphingosine-1-phosphate increased.^[50] As for its clinical application, researchers detected S100A4 levels in the sera and tissues of patients with IPF and health controls. Results showed serum S100A4 levels were undetectable in all health controls but were detectable in 26/95 IPF cases. In the lung tissues from IPF patients, aggregation of numerous S100A4-expressing cells was found around the fibroblastic foci and mature fibrotic regions. Those patients with higher serum S100A4 levels tended to have a significantly worse prognosis than their counterparts.^[51] Similar conclusions were also confirmed by real-time PCR and immunoblotting in fibroblasts from IPF patients and controls.^[52] In another study, hyaluronan (HA) was found to present in the fibroblastic focus together with CD44-expressing fibrogenic mesenchymal progenitor cells (MPC) and that ligation of CD44 by HA triggered S100A4 nuclear translocation to support IPF MPC self-renewal, suggesting that S100A4-mediated MPC fibrogenicity in IPF was regulated by HA/CD44 axis.^[53]

6. Pulmonary arterial hypertension

Pulmonary arterial hypertension (PAH), which is characterized by pulmonary vessels' remodeling and a persistent increase in the pulmonary vascular resistance, is a complex pulmonary vasculature disease with poor prognosis.^[54,55] EMT was considered as a critical process in PAH etiology.^[56] Thus, some factors implicated in EMT, including S100A4, have been verified to be associated with PAH. A recent study conducted by Laggner M et al quantified S100A4, EGF, and EGFR in patients suffering from chronic thromboembolic pulmonary hypertension and idiopathic PAH. Data analysis revealed S100A4 tissue expression positively correlated with higher grades of Heath-Edwards histopathological lesions of idiopathic PAH-derived lung tissue, while was devoid in pulmonary thrombo-endarterectomized samples.^[57] In the mice models of chronic hypoxia, which is considered as a significant factor in the occurrence of pulmonary hypertension, S100A4, along with CD36 were confirmed to be positively expressed in the vascular tunica media.^[58] Concerning of the mechanism, Lawrie A et al investigated the codependence of 5-HT receptors and serotonin transporter in regulating S100A4/Mts1 in human pulmonary artery smooth muscle cells (hPA-SMC). They found that 5-HT elevated S100A4/Mts1 mRNA levels and increased S100A4/Mts1 PA-SMC lysates and culture media, indicating a mechanistic link between the 5-HT pathway and S100A4/Mts1 in pulmonary hypertension.^[59] Besides that, advanced glycation endproducts (RAGE) binding S100A4 released by activated leukocytes results in the generation of reactive oxygen species and further activation of NF- κ B.^[60] This leads to reduced bioavailability of the labile vasodilator nitric oxide, reducing its anti-inflammatory effects and possibly compromising control of vascular tone directly. Furthermore, RAGE antagonism could also prevent migration and proliferation of PA-SMC in response to 5-HT.^[60] Accordingly, Farmer DG et al pointed out that S100-RAGE signaling may be of key importance in pulmonary vascular homeostasis and/or disease in a review.^[60]

7. Conclusion

S100A4 was demonstrated to be involved in several lung diseases, including lung cancer, COPD, asthma, pulmonary hypertension, and IPF. With more and more studies, its mechanism of action has been constantly confirmed. In the future, this protein may be used in the diagnosis, targeted treatment, and prognosis evaluation of several pulmonary diseases.

Author contributions

Data curation: Ting Wang.
Writing – original draft: Ting Wang.
Writing – review & editing: Ting Wang.

References

- Donato R, Cannon BR, Sorci G, et al. Functions of S100 proteins. *Curr Mol Med.* 2013;13:24–57.
- Bresnick AR, Weber DJ, Zimmer DB. S100 proteins in cancer. *Nat Rev Cancer.* 2015;15:96–109.
- Gonzalez LL, Garrie K, Turner MD. Role of S100 proteins in health and disease. *Biochim Biophys Acta Mol Cell Res.* 2020;1867:118677.
- Sadigh AR, Mihanfar A, Fattahi A, et al. S100 protein family and embryo implantation. *J Cell Biochem.* 2019;120:19229–44.
- Heizmann CW, Fritz G, Schäfer BW. S100 proteins: structure, functions and pathology. *Front Biosci.* 2002;7:d1356–68.
- Zhu L, Okano S, Takahara M, et al. Expression of S100 protein family members in normal skin and sweat gland tumors. *J Dermatol Sci.* 2013;70:211–9.
- Sugiyama T, Ozono S, Miyake H. Expression profile of S100A2 and its clinicopathological significance in renal cell carcinoma. *Anticancer Res.* 2020;40:6337–43.

- [8] Kwon YW, Chang IH, Kim KD, et al. Significance of S100A2 and S100A4 expression in the progression of prostate adenocarcinoma. *Korean J Urol.* 2010;51:456–62.
- [9] Zhang J, Zhang K, Jiang X, et al. S100A6 as a potential serum prognostic biomarker and therapeutic target in gastric cancer. *Dig Dis Sci.* 2014;59:2136–44.
- [10] Qin W, Ho L, Wang J, et al. S100A7, a novel Alzheimer's disease biomarker with non-amyloidogenic alpha-secretase activity acts via selective promotion of ADAM-10. *PLoS One.* 2009;4:e4183.
- [11] Zandstra J, van de Geer A, Tanck MWT, et al. Biomarkers for the discrimination of acute Kawasaki disease from infections in childhood. *Front Pediatr.* 2020;8:355.
- [12] van de Logt F, Day AS. S100A12: a noninvasive marker of inflammation in inflammatory bowel disease. *J Dig Dis.* 2013;14:62–7.
- [13] Ambartsumian N, Klingelhöfer J, Grigorian M. The multifaceted S100A4 protein in cancer and inflammation. *Methods Mol Biol.* 2019;1929:339–65.
- [14] Li Z, Li Y, Liu S, et al. Extracellular S100A4 as a key player in fibrotic diseases. *J Cell Mol Med.* 2020;24:5973–83.
- [15] Chen N, Sato D, Saiki Y, et al. S100A4 is frequently overexpressed in lung cancer cells and promotes cell growth and cell motility. *Biochem Biophys Res Commun.* 2014;447:459–64.
- [16] Grigorian MS, Tulchinsky EM, Zain S, et al. The mts1 gene and control of tumor metastasis. *Gene.* 1993;135:229–38.
- [17] Hua T, Liu S, Xin X, et al. S100A4 promotes endometrial cancer progress through epithelial-mesenchymal transition regulation. *Oncol Rep.* 2016;35:3419–26.
- [18] Ning Q, Li F, Wang L, et al. S100A4 amplifies TGF- β -induced epithelial-mesenchymal transition in a pleural mesothelial cell line. *J Investig Med.* 2018;66:334–9.
- [19] Tochimoto M, Oguri Y, Hashimura M, et al. S100A4/non-muscle myosin II signaling regulates epithelial-mesenchymal transition and stemness in uterine carcinosarcoma. *Lab Invest.* 2020;100:682–95.
- [20] Takenaga K, Nakamura Y, Endo H, et al. Involvement of S100-related calcium-binding protein pEL98 (or mts1) in cell motility and tumor cell invasion. *Jpn J Cancer Res.* 1994;85:831–9.
- [21] Takenaga K, Nakamura Y, Sakiyama S. Expression of antisense RNA to S100A4 gene encoding an S100-related calcium-binding protein suppresses metastatic potential of high-metastatic Lewis lung carcinoma cells. *Oncogene.* 1997;14:331–7.
- [22] Stewart RL, Carpenter BL, West DS, et al. S100A4 drives non-small cell lung cancer invasion, associates with poor prognosis, and is effectively targeted by the FDA-approved anti-helminthic agent niclosamide. *Oncotarget.* 2016;7:34630–42.
- [23] Hou S, Tian T, Qi D, et al. S100A4 promotes lung tumor development through β -catenin pathway-mediated autophagy inhibition. *Cell Death Dis.* 2018;9:277.
- [24] Liu L, Qi L, Knifley T, et al. S100A4 alters metabolism and promotes invasion of lung cancer cells by up-regulating mitochondrial complex I protein NDUFS2. *J Biol Chem.* 2019;294:7516–27.
- [25] Alaei M, Nool K, Pasdar M. Plakoglobin restores tumor suppressor activity of p53R175H mutant by sequestering the oncogenic potential of β -catenin. *Cancer Sci.* 2018;109:1876–88.
- [26] Zhang Y, Li W, Lin Z, et al. The long noncoding RNA Linc01833 enhances lung adenocarcinoma progression via MiR-519e-3p/S100A4 Axis. *Cancer Manag Res.* 2020;12:11157–67.
- [27] Smith MC, Wrobel JP. Epidemiology and clinical impact of major comorbidities in patients with COPD. *Int J Chron Obstruct Pulmon Dis.* 2014;9:871–88.
- [28] Hou W, Hu S, Li C, et al. Cigarette smoke induced lung barrier dysfunction, EMT, and tissue remodeling: a possible link between COPD and lung cancer. *Biomed Res Int.* 2019;2019:2025636.
- [29] Sohal SS, Reid D, Soltani A, et al. Reticular basement membrane fragmentation and potential epithelial mesenchymal transition is exaggerated in the airways of smokers with chronic obstructive pulmonary disease. *Respirology.* 2010;15:930–8.
- [30] Sohal SS, Reid D, Soltani A, et al. Evaluation of epithelial mesenchymal transition in patients with chronic obstructive pulmonary disease. *Respir Res.* 2011;12:130.
- [31] Sohal SS, Soltani A, Reid D, et al. A randomized controlled trial of inhaled corticosteroids (ICS) on markers of epithelial-mesenchymal transition (EMT) in large airway samples in COPD: an exploratory proof of concept study. *Int J Chron Obstruct Pulmon Dis.* 2014;9:533–42.
- [32] Mahmood MQ, Ward C, Muller HK, et al. Epithelial mesenchymal transition (EMT) and non-small cell lung cancer (NSCLC): a mutual association with airway disease. *Med Oncol.* 2017;34:45.
- [33] Reimann S, Fink L, Wilhelm J, et al. Increased S100A4 expression in the vasculature of human COPD lungs and murine model of smoke-induced emphysema. *Respir Res.* 2015;16:127.
- [34] Qin HY, Li MD, Xie GF, et al. Associations among S100A4, sphingosine-1-phosphate, and pulmonary function in patients with chronic obstructive pulmonary disease. *Oxid Med Cell Longev.* 2022;2022:6041471.
- [35] Jiang B, Guan Y, Shen HJ, et al. Akt/PKB signaling regulates cigarette smoke-induced pulmonary epithelial-mesenchymal transition. *Lung Cancer.* 2018;122:44–53.
- [36] Kwah JH, Peters AT. Asthma in adults: principles of treatment. *Allergy Asthma Proc.* 2019;40:396–402.
- [37] Gans MD, Gavrilova T. Understanding the immunology of asthma: pathophysiology, biomarkers, and treatments for asthma endotypes. *Paediatr Respir Rev.* 2020;36:118–27.
- [38] Maremanda KP, Sundar IK, Rahman I. Protective role of mesenchymal stem cells and mesenchymal stem cell-derived exosomes in cigarette smoke-induced mitochondrial dysfunction in mice. *Toxicol Appl Pharmacol.* 2019;385:114788.
- [39] Schneider M, Hansen JL, Sheikh SP. S100A4: a common mediator of epithelial-mesenchymal transition, fibrosis and regeneration in diseases? *J Mol Med (Berl).* 2008;86:507–22.
- [40] D'Ambrosi N, Milani M, Apolloni S. S100A4 in the physiology and pathology of the central and peripheral nervous system. *Cells.* 2021;10:798.
- [41] Huang X, Qu D, Liang Y, et al. Elevated S100A4 in asthmatics and an allergen-induced mouse asthma model. *J Cell Biochem.* 2019;120:9667–76.
- [42] Jiang JX, Guan Y, Shen HJ, et al. Inhibition of soluble epoxide hydrolase attenuates airway remodeling in a chronic asthma model. *Eur J Pharmacol.* 2020;868:172874.
- [43] Tabaa MME, Fattah AMK, Shaalan M, et al. Dapagliflozin mitigates ovalbumin-prompted airway inflammatory-oxidative successions and associated bronchospasm in a rat model of allergic asthma. *Expert Opin Ther Targets.* 2022;26:487–506.
- [44] Jiang JX, Shen HJ, Guan Y, et al. ZDHXB-101 (3',5'-Diallyl-2, 4'-dihydroxy-[1,1'-biphen-yl]-3,5'-dicarbaldehyde) protects against airway remodeling and hyperresponsiveness via inhibiting both the activation of the mitogen-activated protein kinase and the signal transducer and activator of transcription-3 signaling pathways. *Respir Res.* 2020;21:22.
- [45] Wu Y, Zhang W, Gunst SJ. S100A4 is secreted by airway smooth muscle tissues and activates inflammatory signaling pathways via receptors for advanced glycation end products. *Am J Physiol Lung Cell Mol Physiol.* 2020;319:L185–95.
- [46] Sharif R. Overview of idiopathic pulmonary fibrosis (IPF) and evidence-based guidelines. *Am J Manag Care.* 2017;23(11 Suppl):S176–82.
- [47] Biondini D, Balestro E, Sverzellati N, et al. Acute exacerbations of idiopathic pulmonary fibrosis (AE-IPF): an overview of current and future therapeutic strategies. *Expert Rev Respir Med.* 2020;14:405–14.
- [48] Saito S, Alkhatib A, Kolls JK, et al. Pharmacotherapy and adjunctive treatment for idiopathic pulmonary fibrosis (IPF). *J Thorac Dis.* 2019;11(Suppl 14):S1740–54.
- [49] Zhang W, Ohno S, Steer B, et al. S100a4 is secreted by alternatively activated alveolar macrophages and promotes activation of lung fibroblasts in pulmonary fibrosis. *Front Immunol.* 2018;9:1216.
- [50] Li Y, Bao J, Bian Y, et al. S100A4+ macrophages are necessary for pulmonary fibrosis by activating lung fibroblasts. *Front Immunol.* 2018;9:1776.
- [51] Akiyama N, Hozumi H, Isayama T, et al. Clinical significance of serum S100 calcium-binding protein A4 in idiopathic pulmonary fibrosis. *Respirology.* 2020;25:743–9.
- [52] Lee JU, Chang HS, Shim EY, et al. The S100 calcium-binding protein A4 level is elevated in the lungs of patients with idiopathic pulmonary fibrosis. *Respir Med.* 2020;171:105945.
- [53] Xia H, Herrera J, Smith K, et al. Hyaluronan/CD44 axis regulates S100A4-mediated mesenchymal progenitor cell fibrogenicity in idiopathic pulmonary fibrosis. *Am J Physiol Lung Cell Mol Physiol.* 2021;320:L926–41.
- [54] Coons JC, Pogue K, Kolodziej AR, et al. Pulmonary arterial hypertension: a pharmacotherapeutic update. *Curr Cardiol Rep.* 2019;21:141.
- [55] Vazquez ZGS, Klinger JR. Guidelines for the treatment of pulmonary arterial hypertension. *Lung.* 2020;198:581–96.
- [56] Zeng M, Chen S, Li H, et al. The role of β -catenin in pulmonary artery endothelial-mesenchymal transformation in rats with chronic thromboembolic pulmonary hypertension. *J Thromb Thrombolysis.* 2021;52:454–65.

- [57] Laggner M, Hacker P, Oberndorfer F, et al. The roles of S100A4 and the EGF/EGFR signaling axis in pulmonary hypertension with right ventricular hypertrophy. *Biology (Basel)*. 2022;11:118.
- [58] Kwapiszewska G, Wilhelm J, Wolff S, et al. Expression profiling of laser-microdissected intrapulmonary arteries in hypoxia-induced pulmonary hypertension. *Respir Res*. 2005;6:109.
- [59] Lawrie A, Spiekerkoetter E, Martinez EC, et al. Interdependent serotonin transporter and receptor pathways regulate S100A4/Mts1, a gene associated with pulmonary vascular disease. *Circ Res*. 2005;97:227–35.
- [60] Farmer DG, Kennedy S. RAGE, vascular tone and vascular disease. *Pharmacol Ther*. 2009;124:185–94.