ORIGINAL ARTICLE



Comparison of platelet count results on the Sysmex XN between citrate or MgSO₄ and K2 EDTA anticoagulants

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[Correction added on 22 December 2022, after first online publication: The copyright line was changed.]

Abstract

Introduction: The aim of this study performed on Sysmex XN is to compare platelet values on citrate and MgSO₄ (TBX) in patients with K2EDTA-induced platelet clusters and to identify platelet biases of these matrices compared to K2EDTA.

Methods: Sixty patients with K2EDTA-induced platelet clusters were re-sampled with K2EDTA, citrate and TBX. Platelet results were then compared, and smears were analysed for clumping. Platelet results from 120 patients without K2EDTAinduced platelet clusters were compared between K2 EDTA, citrate, and MgSO₄ with impedance and fluorescence modes. Biases from regressions were analysed.

Results: Out of the 60 patients with K2EDTA-induced platelet clusters, none showed platelet clusters with MgSO₄ whereas 50% still showed clusters with citrate. Among those without platelet clusters on citrate, the mean relative difference between (citrate- $MgSO_4$)/ $MgSO_4$ was -12.7% in impedance and -9.8% in fluorescence. Among the 120 patients without K2EDTA-induced platelet clusters, in fluorescence the mean relative bias with respect to K2EDTA was -2.06% for MgSO₄ and -10.3% for Citrate. For the MgSO₄ versus K2 EDTA regressions, the maximum absolute values of the 95% CI of the relative biases at $150 \times 10^9/L$ (5.45%) and $450 \times 10^9/L$ (3.56%) were below the desirable analytical objectives of the EFLM.

Conclusion: In patients with K2EDTA-induced platelet clusters, MgSO₄ is preferable to citrate. MgSO₄ provides a bias with XN in fluorescence when compared to EDTA which is within analytical tolerance.

KEYWORDS

citrate, ethylene diamine tetracetic acid, magnesium sulphate, platelet, Sysmex

INTRODUCTION 1

EDTA dependent pseudothrombocytopenia (PTP) is a problem encountered relatively frequently in daily routine laboratory activity, as the prevalence of K2 EDTA-induced PTP ranges from 0.03% to 0.27% in the general population.¹⁻³ PTP represents a challenge because of the risk of delivering an erroneous result of the platelet count, as overlooking a severe thrombocytopenia can have serious consequences for the patient. It is therefore essential to obtain reliable platelet counts in order to make clinically important decisions with confidence.

PTP is characterized by the in vitro agglutination of platelets in the presence of EDTA leading to a falsely lowered platelet count. PTP is confirmed by observation of the May Grunwald Giemsa (MGG) stained blood smear for the presence of platelet clusters and by the disappearance of these platelet clusters using an anticoagulant other than EDTA.

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PTP is a temperature and time dependent phenomenon.⁴⁻⁶ It is related to the existence of predominantly IgG autoantibodies (IgG1) to cryptic epitopes of GpllbIlla unmasked by EDTA calcium chelation.^{4,5}

To prevent platelet agglutination in vitro, different anticoagulants can be used. Sodium citrate is the most widely used. It is recommended to perform the platelet count within a maximum of 3 h after collection.⁵

Several publications recommend the use of MgSO₄ for platelet counts in EDTA-induced PTP.^{7.8} The anticoagulant effect of MgSO₄ results on the one hand from the inhibition of fibrinogen binding to the GpIlbIIIa complex as well as the inhibition of thromboxane A2 formation, and on the other hand from the formation of cAMP, which blocks platelet activation by inhibiting the mobilization of intra-tubular calcium.⁹

The Sysmex XN series automated system allows platelets to be counted in two different ways. The impedance after hydrodynamic focalization produces a histogram limited by two flexible thresholds located between 2 and 6×10^{-15} L for the lower threshold and between 12 and 30×10^{-15} L for the upper threshold. In addition, the use of a method based on the combination of optics and fluorescence allows better discrimination of platelets from leukocyte cytoplasm fragments and improves the accuracy of counting in thrombocytopenic patients with results directly comparable to the reference method CD41/CD61 in flow cytometry. $^{10-13}$

Previous studies have shown that on the XE 5000 analyser, MgSO₄ underestimated platelet counts in impedance compared to K3 EDTA but produced similar results in fluorescence using polymethine.^{7,8}

The objectives of this study performed on the Sysmex XN analyser were:

- In patients with K2EDTA-induced platelet clusters, to compare the platelet count results obtained when using sodium citrate and MgSO₄.
- In patients without K2EDTA-induced platelet clusters, to evaluate the regressions and biases at decision thresholds between platelet count results obtained with either sodium citrate or magnesium sulphate and those obtained with K2EDTA using impedance or fluorescence with oxazine.

2 | MATERIAL AND METHODS

2.1 | Patients

The patients come from the laboratory's patient base.

We collected samples from 60 patients (33 males, 27 females, median age 67 years, min 19 years, max 92 years) with initial platelet clusters on K2EDTA without fibrin flakes. These samples were collected using 3 tubes and in the following order: first K2EDTA, 7.2 mg, 4 ml, BD Vacutainer; then Sodium Citrate 0.109 M, 2.7 ml, BD Vacutainer; then MgSO₄, 33.8 μ mol, 2.7 ml, Thromboexact (TBX) Sarstedt.

One hundred and twenty patients (65 males, 55 females, median 66 years, min 18 years, max 95 years) without platelet clusters on the blood smear, were sampled for platelet count. For each patient, we collected 1 K2EDTA tube, 1 Sodium Citrate tube and 1 TBX tube. These patients were chosen with a K2EDTA mean corpuscular volume between 75×10^{-15} L and 106×10^{-15} L, median 91.8×10^{-15} L, and leucocytes between 1.42×10^9 /L and 25.53×10^9 /L, median 7.21×10^9 /L, without cytoplasmic fragments of nucleated cells on

the May Grunwald Giemsa (MGG) stained smear. In addition, no images suggestive of cryoglobulin were observed on the smears. One patient had platelet aggregates on the K2EDTA blood smear that were grouped by 2 or 3 at most on 4 out of 10 fields. This patient, whose largest clusters were less than 5 platelets, was not excluded from the study. On K2EDTA, this patient had $213 \times 10^9/L$ platelets in impedance and $249 \times 10^9/L$ platelets in fluorescence; and on MgSO4 he had $259 \times 10^9/L$ platelets in impedance and $258 \times 10^9/L$ platelets in fluorescence.

This study was approved by the Ethics Committee (Ile de France IV) (2021-A02741-40) and was conducted according to the Declaration of Helsinki. All patients gave written informed consent for participation in the study.

2.2 | Sample processing

Samples were transported at 15–25°C and analysed within a maximum of 3 h after collection. Platelets from the 60 patients with platelet clusters were analysed using an XN 9000 (XN10 version 22.16). Platelets from the 120 patients without platelet clusters were analysed in replicate using the same XN3000 (XN10 version 22.16) in impedance and fluorescence with oxazine. None of the 120 samples showed alarms on the XN suggesting the presence of platelet clusters.

During the study period, three levels of internal quality control materials were routinely evaluated for the XN 3000 and XN 9000 and all the required maintenance procedures were performed in accordance with the Sysmex's manual. The laboratory also obtained acceptable results on external quality control samples.

Platelet counts on citrate were corrected by multiplying the raw result by the ratio of haemoglobin levels on K2EDTA to citrate. Platelet clusters were detected by staining smears with MGG on K2EDTA, citrate, and TBX tubes from patients. When observing 10 fields under the microscope (50X, 10X), the presence of platelet clusters was characterized by the observation on each of at least four fields of platelets attached by five. Blood smears were read under a Zeiss light microscope (Primo Star) by a laboratory technician and verified by a senior haematopathologist.

2.3 | Analysis of statistical results

The results of the platelet count on citrate and MgSO4 were compared for each mode with the result obtained on K2EDTA. Three regressions were analysed: least squares (O.L.S), Deming, and Passing-Bablok (PB). For Deming, the ratio of analytical variances was calculated according to Combleet and Gochman. 14 For each regression, the slope, intercept, R2 coefficient, and biases at the $150\times10^9/L$ and $450\times10^9/L$ thresholds were evaluated. The 95% CI of the intercept, slope, and biases were calculated for O.L.S according to Clinical and Laboratory Standards Institute 15 with Excel 2010, for Deming according to Jacknife $^{16.17}$ with Excel 2010, and for PB according to Bootstrap with MedCalc $^{(8)}$ Statistical Software version 20.011 (MedCalc Software Ltd, Ostend, Belgium). Relative biases were compared to the desirable EFLM 18 (European Federation of Clinical Chemistry and Laboratory Medicine Biological variation) biases and the maximum bounds of the 95%CI of the biases were compared to the reference data. 19

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30 64 93 138 60 66 150 172	29	113	196	272	59	68	143	148
50 04 75 150 00 00 139 175	30	64	93	138	60	66	159	173

TABLE 1 Comparison of platelet count results $(10^9/L)$ obtained on the XN impedance analyser using the anticoagulants citrate and MgSO₄ in 60 patients with K2EDTA-induced platelet clusters

Note: Platelet count results with platelet clusters are shown in bold.

The Shapiro-Wilk test was used to analyse the normality of the distribution of results. Non-normally distributed data was expressed as medians and interquartile ranges (IQR).

Means were compared using Student's t-test when the difference in values was normally distributed, otherwise the Wilcoxon test was performed. Medians were compared with the Friedman test. The significance level is p < 0.05.

3 | RESULTS

3.1 | Comparison of platelet results with citrate or $MgSO_4$ in patients with K2EDTA induced platelet clusters

Platelet counts were performed in impedance mode in all the 60 patients sampled on K2EDTA, Citrate and $MgSO_4$.

On K2EDTA, the median was 67 \times 10 $^9/L$, on citrate the mean was 148.8 \times 10 $^9/L$, on MgSO4 the mean was 209.6 \times 10 $^9/L$.

All patients had platelet clusters on K2EDTA. Thirty of them still showed clusters on citrate (50%) and none showed clusters on MgSO4. All platelet results obtained with MgSO4 were superior to those of K2EDTA and citrate. It should be noted that eight patients had lower platelet results on citrate than on K2EDTA (Table 1).

Of the 60 patients whose platelet results were associated with the presence of platelet clusters, 56 had EDTA-induced pseudothrombopenia. With citrate, 33 out of the 56 patients had platelet counts below $150\times10^9/L$, 10 of which were confirmed by the absence of platelet clusters on the smear. In contrast, with MgSO₄, 13 patients had platelet counts below $150\times10^9/L$ with no clumps on the smear. For patient 47, the thrombocytopenia diagnosed with citrate disappeared with MgSO₄.

Among the 30 patients without platelet clusters on citrate, the mean of the relative differences in impedance analysed platelets

between Citrate and MgSO₄ versus MgSO₄ for these patients was -12.7%, 95% CI (-10.4% to -14.9%).

Out of the previous 60 patients, 30 were analysed by fluorescence (eight with clusters on citrate and 22 without clusters).

The median was 98.5 \times 10 $^9/L$ on K2EDTA, while the medians were 207.6 \times 10 $^9/L$ on citrate and 255.3 \times 10 $^9/L$ on MgSO4.

Among the 22 patients without platelet clusters on citrate, the mean relative difference in platelets between Citrate and MgSO₄ versus MgSO₄ was -9.8%, 95%CI (-5.7% to -13.9%) in fluorescence -11.3%, 95%CI (-7.9% to -14.7%) in impedance.

For these 22 patients the mean relative difference in platelets between fluorescence and impedance versus impedance was 12.17%, 95% CI (7.03%–17.31%) on citrate and 10.97%, 95% CI (3.65 to 18.28) on MgSO₄.

These results suggested that in patients with EDTA-induced platelet clusters, $MgSO_4$ was more effective than citrate to perform platelet counts. Furthermore, on XN with $MgSO_4$ the platelet results obtained with fluorescence were about 10% higher than those obtained with impedance.

3.2 | Comparison of platelet results between MgSO₄ and K2EDTA in patients without K2EDTA induced platelet clusters

One hundred and twenty patients were sampled with K2 EDTA and MgSO₄. Platelet counts were performed on a single XN using impedance and fluorescence modes.

For K2EDTA the range of values was $18-767 \times 10^9/L$ in impedance and $15-801 \times 10^9/L$ in fluorescence.

For all these patients the mean relative bias of the platelet results between fluorescence and impedance compared to impedance was -0.45% 95%CI (-1.85 to 0.95) with K2EDTA and 9.45% 95%CI (7.81-11.09) with MgSO₄.

Therefore, the results of platelet count in the fluorescence correlated better with K2EDTA than with $MgSO_4$ in impedance.

Comparison of platelet count results in fluorescence mode between MgSO $_4$ and K2EDTA versus K2EDTA showed a mean relative bias of -2.06% 95%CI (-3.41 to -0.71).

In fluorescence the MgSO₄ versus K2EDTA regressions showed correlation coefficients greater than 0.995. The slope of the Deming regression was not statistically different from 1. At the decision threshold of $150\times10^9/L$, the average bias was between $4.6\times10^9/L$ and $5.4\times10^9/L$ and the maximums from 95% CI were between $6.7\times10^9/L$ and $8.2\times10^9/L$. At the $150\times10^9/L$ and $450\times10^9/L$ bounds, the mean relative bias and the maximum relative bias derived from the 95%CI were lower than the specifications of the EFLM¹⁸ and Buoro et al., 19 respectively (Table 2 Figure 1A).

By way of comparison, at a threshold of $150\times10^9/L$, in impedance, the MgSO₄ versus K2EDTA regressions generated mean biases between $15.7\times10^9/L$ and $16.8\times10^9/L$ and maximum biases between $19\times10^9/L$ and $20.6\times10^9/L$ (Table 2, Figure 1B).

Regression analysis of Passing Bablok (PB), MgSO₄ in fluorescence versus K2EDTA in impedance, showed a slope that was not statistically different from 1 and a mean bias at 150×10^9 /L of 5.1×10^9 /L.

As a result, platelet counts on MgSO₄ analysed in fluorescence correlate with platelet counts on K2EDTA analysed in impedance and were similar to those obtained with K2EDTA in fluorescence.

3.3 | Comparison of platelet results between Citrate and K2EDTA in patients without K2EDTA induced platelet clusters

For the 120 patients, the mean relative bias of platelets on citrate between fluorescence and impedance compared to impedance was 9.49% 95% CI (7.60%–11.39%).

The results of platelets collected on citrate were always lower than those of platelets collected on K2 EDTA regardless of the mode of analysis.

For the 120 patients, the median platelet (citrate-K2EDTA)/K2EDTA result ratio was in impedance -18.7% IQR (-24.6%; -14.5%) and in fluorescence -10.5% IQR (-15.5%; -7%).

Analysis of the results of the platelet regressions between citrate and K2EDTA showed correlation coefficients ranging from 0.942 to 0.96 in impedance and from 0.953 to 0.977 in fluorescence. The mean biases of the citrate versus K2EDTA regressions at the decision threshold of 150 \times 10 $^9/L$ ranged from 15.4 \times 10 $^9/L$ to 21.5 \times 10 $^9/L$ in fluorescence and from 27.9 \times 10 $^9/L$ to 30.4 \times 10 $^9/L$ in impedance (Table 3, Figure 1C, D).

Although the regression results for citrate versus K2EDTA platelets were slightly better in the fluorescence mode than in the impedance mode, at thresholds of 150×10^9 /L, the mean relative biases and their 95%CI exceeded the analytical specifications of the EFLM¹⁸ and Buoro et al.¹⁹

3.4 | Mean platelet volume

Among the 113 patients for whom platelet volume was available, a comparison of results between K2EDTA, citrate, and MgSO₄ matrices was performed.

Medians of platelet volume with interquartile ranges (IQR) were 10.3×10^{-15} /L (9.8; 10.94), 9.75×10^{-15} /L (9.3; 10.40), and 9.50×10^{-15} /L (9.1; 10.08) for K2EDTA, citrate, and MgSO4 respectively. They differed significantly (Friedman test p < 0.0001).

The mean relative bias between citrate and EDTA K2 was -5.48%, 95%CI (-6.207 to -4.748) while the mean relative bias between TBX and EDTA K2 was -7.4%, 95%CI (-8.117 to -6.682).

The bias of citrate over K2EDTA from the Passing Bablok (PB) regression was $-0.6\times10^{-15}/L$ 95%Cl (-0.71 to -0.49) for MPV K2EDTA = $9\times10^{-15}/L$ and was $-0.6\times10^{-15}/L$ 95%Cl (-0.73 to -0.39) for MPV K2EDTA = $12\times10^{-15}/L$.

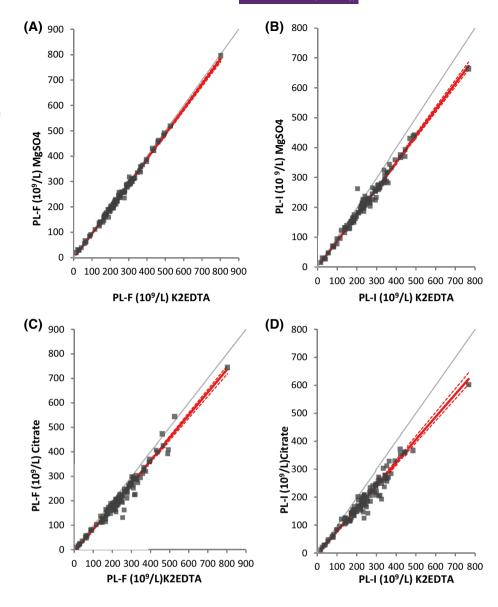
Comparison of platelet results obtained between magnesium sulfate (MgSO₄) and K2EDTA (EDTA) in 120 patients without platelet clumps on XN analyser with impedance or fluorescence mode using least squares, Deming and Passing-Bablok regressions TABLE 2

$Max\;Bias\;(\%)\\Xc=450\;10^9/L$	5	4	9:	13.58		е;		61	52
Σ×	3.05	2.84	3.56	13	13	13.3	2.7	1.89	3.55
Bias (%) Xc = 450 10°/L	2.23	2.08	2.47	12.13	11.75	12.02	0.88	0	1.35
Max Bias (%) Xc = 150 10 ⁹ /L	5.45	5.00	4.45	13.73	12.73	12.7	8.35	8.53	6.03
Bias (%) Xc = 150 10 ⁹ /L	3.58	3.81	3.08	10.48	10.96	11.19	4.29	5.41	3.41
C.C.	0.997	0.997	0.995	0.990	0.990	0.979	0.988	0.988	0.981
Intercept 95Cl	-3.037 (-6.415 to 0.341)	-3.907 (-7.318 to -0.497)	-1.374 (-4.937 to 1.555)	3.7 05 (-2.261 to 9.671)	1.792 (-2.420 to 6.004)	1.874 (-1.740 to 6.152)	-7.664 (-15.122 to -0.206)	-12.165 (-19.538 to -4.791)	-4.624 (-12.488 to 3.089)
Slope 95CI	0.984 (0.972 to 0.997)	0.988 (0.974 to 1.002)	0.978 (0.962 to 0.992)	0.870 (0.848 to 0.893)	0.878 (0.860 to 0.897)	0.876 (0.856 to 0.896)	1.008 (0.980 to 1.036)	1027 (0.995 to 1059)	0.997 (0.960 to 1026)
Regression	OLS	Deming	PB	OLS	Deming	PB	OLS	Deming	PB
X-max/ X-min (10 ⁹ /L)	801/15	801/15	801/15	767/18	767/18	767/18	767/18	767/18	767/18
Comparison	$\begin{array}{l} Y = MgSO_4 \text{ versus} \\ X = EDTA \end{array}$	$\begin{array}{l} Y = MgSO_4 \text{ versus} \\ X = EDTA \end{array}$	$\begin{array}{l} Y = MgSO_4 \text{ versus} \\ X = EDTA \end{array}$	$\begin{array}{l} Y = MgSO_4 \text{ versus} \\ X = EDTA \end{array}$	$\begin{array}{l} Y = MgSO_4 \text{ versus} \\ X = EDTA \end{array}$	$\begin{array}{l} Y = MgSO_4 \text{ versus} \\ X = EDTA \end{array}$	$Y = MgSO_4 F versus$ X = EDTA I	$Y = MgSO_4 F versus$ X = EDTAI	$Y = MgSO_4 F versus$ X = EDTA I
Méthod	Fluo	Fluo	Fluo	dml	dml	dml	Fluo Imp	Fluo Imp	Fluo Imp

Note: Shown in bold are the mean relative biases that are less than the desirable specifications of the EFLM platelet bias and the maximum relative biases that are less than the 95% confidence interval limits of the platelet bias according to Buoro et al. 19

Abbreviations: Bias (%) Xc, absolute value of the relative bias measured at medical decision level Xc; C.C., correlation coefficient; Deming, Deming regression; F, Fluo, Fluorescence; I, Imp, Impedance, OLS, ordinary least squares regression; Max Bias (%), absolute value of the maximum of the 95% confidence interval of the relative bias at medical decision level Xc; PB, Passing-Bablok regression.

FIGURE 1 Comparison using Passing-Bablok regression of platelet results from 120 patients without platelet clumps between magnesium sulfate and K2EDTA (A. B) and between sodium citrate and EDTA K2 (C, D) in fluorescence (A, C), and impedance (B, D) modes. Regression lines are presented with solid red lines, 95%CI lines are presented with dashed red lines, and identity lines are presented with solid grey lines. PL-I, platelets measured on the XN analyser in impedance mode; PL-F, platelets measured on the XN analyser in fluorescence mode; Citrate, sodium citrate; magnesium sulfate, MgSO₄.



The bias of MgSO4 vs EDTA from PB regression was $-0.67\times10^{-15}/L$ 95Cl (-0.8 to -0.56) for MPV K2EDTA $=~9\times10^{-15}/L$ and was $-0.95\times10^{-15}/L,$ 95Cl (-1.12 to -0.79) for MPV EDTA $=12\times10^{-15}/L$

At the 9×10^{-15} /L and 12×10^{-15} /L thresholds, the absolute value of the relative biases obtained between MgSO4 and K2EDTA and those obtained between citrate and K2EDTA exceeded the relative bias of 1.9% (95%CI: 1.4%–2.7%) established by Buoro et al.¹⁹

4 | DISCUSSION

PTP is a relatively common in vitro phenomenon which leads to falsely low platelet counts determined automatically. Usually, the thrombocytopenia is noticed accidentally without a corresponding tendency to bleeding. But if this interference is not recognized, misinterpretation may have dramatic consequences by way of diagnostical and therapeutical mistakes. Besides, in many patients, the use of another matrix such as sodium citrate does not overcome this interference and

does not solve the problem. This is the reason why it is important to have a matrix that allows the laboratory to deliver a correct platelet count on which the clinician can base his diagnostic and/or therapeutic attitude.

This study showed that in 60 patients with K2EDTA-induced clumping, the use of MgSO4 was more effective than citrate in removing platelet clumps. In addition, on XN, fluorescence counting of platelets collected on MgSO4 generated an average relative bias of -2.06% compared to K2EDTA and relative biases at the threshold of $150\times10^9/L$ which, regardless of the regression procedure, remained below the desirable specifications of the EFLM. 18 On the other hand, the use of citrate in fluorescence or MgSO4 in impedance did not achieve these objectives.

Comparison of the mean values of the platelet ratios (citrate-MgSO4)/MgSO4 measured by impedance: -12.7% (30 patients) and -11.3% (22 patients), and fluorescence: -9.8% (22 patients), suggested that irrespective of the mode of analysis, impedance or fluorescence, the results of platelets count on citrate were on average about 10% lower than those on MgSO₄ measured in the same mode.

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Comparison of platelet results obtained between sodium citrate (Cit.) and EDTAK2 (EDTA) in 120 patients without platelet clumps on XN analyser with impedance or fluorescence mode using least squares, Deming and Passing-Bablok regressions TABLE 3

		X max -X min		i d	Č i	Ç		Max Bias (%)	Bias (%)	Max Bias (%)
Method	Method Comparaison ((10.7L)	Kegression Slope	Slope 93Cl	Intercept 95CI	ز ز	Blas (%) $XC = 150 \ 10^{\circ} / L$ $XC = 150 \ 10^{\circ} / L$ $XC = 450 \ 10^{\circ} / L$	$XC = 150 \ 10^{\circ}/L$	XC = 450 IU'/L	XC = 450 IU / L
dwl	Y = Cit. versus X = EDTA 767-18	767-18	OLS	0.793 (0.757-0.829)	0.635 (-8.817 to 10.087 0.942 20.30	0.942	20.30	25.44	20.58	22.88
lmp	Y = Cit. versus X = EDTA 767-18	767-18	Deming	0.870 (0.823-0.916)	4.853 (-6.164 to 15.870) 0.956 19.16	0.956	19.16	21.9	19.8	22.2
dwl	Y = Cit. versus X = EDTA 767-18	767-18	PB	0.812 (0.782-0.847)	0.318 (-7.171 to 4.518 0.96	96.0	18.61	21.39	18.76	21.03
Fluo	Y = Cit. versus X = EDTA 801-15	801-15	OLS	0.920 (0.833-0.958)	-9.624 (-19.532 to 0.283) 0.953 14.31	0.953	14.31	19.82	10.06	14.28
Fluo	Y = Cit. versus X = EDTA 801-15	801-15	Deming	0.935 (0.889-0.980)	0.935 (0.889-0.980) -10.595 (-21.485 to 0.294) 0.977 13.6	0.977	13.6	17.12	8.89	11.4
Fluo	Y = Cit. versus X = EDTA 801-15	801-15	PB	0.924 (0.894-0.948)	0.924 (0.894-0.948) -3.94 (-11.458 to -0.734) 0.97	0.97	10.24	13.11	8.49	10.41

Abbreviations: Bias (%) Xc, absolute value of the relative bias measured at medical decision level Xc; C.C., correlation coefficient; Deming, Deming regression; Fluo, Fluorescence; Imp, Impedance, OLS, ordinary least squares regression; Max Bias (%), absolute value of the maximum of the 95% confidence interval of the relative bias at medical decision level Xc; PB, Passing-Bablok regression

Comparison of the mean values of the platelet ratios (fluorescence-impedance)/impedance obtained on citrate: 12.7% (22 patients) and 9.49% (120 patients), and on MgSO₄: 10.97% (22 patients) and 9.45% (120 patients), suggested that regardless of the matrix, MgSO₄ or citrate, the platelet results on one of these matrices were on average about 10% higher on fluorescence than on impedance. In contrast, for EDTA, the platelet fluorescence results were on average about 0.45% lower than the platelet impedance results.

This latter ratio, which reflects the overestimation of platelets in impedance, must be interpreted according to the selection criteria of the 120 patients. First, samples that showed platelet clusters on the smear consisting of at least 5 agglutinated platelets were eliminated. In addition, for red blood cells, only two patients had an K2EDTA MCV of less than 80×10^{-15} L: one with an K2EDTA MCV of 75×10^{-15} L with platelets at $305 \times 10^9/L$ in impedance and $296 \times 10^9 / L$ in fluorescence, and the other with an K2EDTA MCV of 77×10^{-15} L and platelets at $336 \times 10^9/L$ in impedance and $301 \times 10^9 / L$ in fluorescence. Finally, for red blood cells, the RDW-CV ranged from 11.5% to 19% and the fragment alarm was never generated.

Two patients excluded from the study had a deep microcytosis: one with a K2EDTA MCV of 62×10^{-15} /L had platelets in fluorescence at $161 \times 10^9 / L$ on K2EDTA and $159 \times 10^9 / L$ on MgSO₄, the other one with a K2EDTA MCV of $55 \times 10^{-15}/L$ had platelets at $171 \times 10^9 / L$ on K2EDTA and $168 \times 10^9 / L$ on MgSO₄. These results suggest that even in the case of microcytosis, platelet fluorescence results on MgSO₄ correlate well with those obtained on K2EDTA although this needs to be confirmed in a larger patient population.

Since the average relative bias between MgSO₄ Fluorescence and K2EDTA Fluorescence compared to K2EDTA Fluorescence was low (-2.06%), all platelets detected in fluorescence by K2EDTA were likely to be detected with MgSO₄. The underestimation of MgSO₄ platelets analysed by impedance compared to K2EDTA might be related to the reduced volume of platelets collected on MgSO₄ compared to K2EDTA which would result in the loss of platelets below the low detection limit. The underestimation of citrate platelets analysed by impedance compared to K2EDTA was not explained by the decrease in platelet volume of citrate compared to K2EDTA, as the MPV of citrate was higher than the MPV of MgSO₄ and therefore the platelet counts in citrate impedance should be higher than those obtained in MgSO₄.

Conversely, it can be seen that at a threshold of $150 \times 10^9/L$ in impedance, the mean bias between MgSO₄ and K2EDTA was 17×10^9 /L, whereas it was 26×10^9 /L between citrate and K2EDTA.

The mean relative bias in fluorescence between Citrate and K2EDTA compared to K2EDTA (-10.3%) revealed that citrate, unlike MgSO₄, detected a lower number of platelets in fluorescence compared to K2EDTA. This difference could be explained either by a lower reactivity of the fluorochrome oxazine with platelets in the presence of citrate compared to K2EDTA, or by the fact that some platelets aggregated in the presence of citrate and were excluded from the count due to the large size of the platelet clusters.²⁰

In a study of 23 patients, François et al.²¹ showed that with ADVIA, the median platelet count on K3EDTA (255 \times 10 9 /L) was higher than on MgSO₄ (215 \times 10⁹/L) and that the median platelet volume on K3EDTA (8.6 \times 10⁻¹⁵ L) was lower than on MgSO₄ (10.45 \times $10^{-15}\,$ L). These results suggest, according to the authors, that the decrease in platelets on MgSO₄ compared to K3EDTA is related to platelet activation and fragmentation. Our results obtained on XN in oxazine fluorescence and those of Mannuß obtained on XE in optics show much lower average platelet deviations between the 2 matrices and the MPV measured in impedance are higher for K2EDTA or K3EDTA than for MgSO₄. These discrepancies are related to the ADVIA assay method, which relies on the interaction between the laser beam and the platelets to obtain the volume and refractive index which are then converted to platelet volume and density. The greater increase in MPV over time with K3EDTA compared to MgSO₄ on ADVIA could result in altered optical properties and a defect in platelet counting.8 In contrast, the hypothesis of platelet activation on MgSO₄ was refuted by the observation that the activation marker CD62 is less expressed on platelets in the presence of MgSO₄ or citrate compared to EDTA.⁷

In summary, in K2EDTA induced platelet clusters, MgSO $_4$ is more effective than citrate in removing platelet clusters. Consequently, in a PTP context, we advocate the use of the MgSO $_4$ matrix rather than citrate to overcome this interference and deliver a correct platelet count. In addition, unlike citrate, the use of MgSO $_4$ results in a bias on fluorescence XN compared to K2EDTA that is compatible with the desirable analytical specifications of the EFLM. Further studies will be required to compare the accuracy of platelet results obtained with MgSO $_4$ versus K2EDTA and to clarify the stability of platelets collected on MgSO $_4$ when analysed in fluorescence with XN.

AUTHOR CONTRIBUTIONS

Michel Soulard designed the study. Pascale Croix performed the experiments. Michel Soulard, Patrick Cohen analysed the data and wrote the manuscript. All the authors read and approved the content of the manuscript.

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CONFLICT OF INTEREST

The author declares that there is no conflict of interest.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ETHICS STATEMENT

This study received the approval of the Ethics Committee (Ile de France IV) (2021-A02741-40).

PATIENT CONSENT STATEMENT

All patients gave written informed consent for participation in the study.

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REFERENCES

- Bizzaro N. EDTA-dependent pseudothrombocytopenia: a clinical and epidemiological study of 112 cases, with 10-year follow-up. Am J Hematol. 1995;50:103-109. doi:10.1002/ajh.2830500206
- Bartels PCM, Schoorl M, Lombarts AJPF. Screening for EDTAdependent deviations in platelet counts and abnormalities in platelet distribution histograms in pseudothrombocytopenia. Scand J Clin Lab Invest. 1997;57:629-636. doi:10.3109/00365519709055287
- Xiao Y, Yu S, Xu Y. The prevalence and biochemical profiles of EDTAdependent pseudothrombocytopenia in a generally healthy population. Acta Haematol. 2015;134:177-180. doi:10.1159/000373915
- Baccini V, Geneviève F, Jacqmin H, et al. Platelet counting: ugly traps and good advice. Proposals from the French-speaking cellular hematology group (GFHC). J Clin Med. 2020;9:808. doi:10.3390/jcm9030808
- Lardinois B, Favresse J, Chatelain B, Lippi G, Mullier F. Pseudothrombocytopenia—a review on causes, occurrence and clinical implications. J Clin Med. 2021;10:594. doi:10.3390/jcm10040594
- Mannuß S. Influence of different methods and anticoagulants on platelet parameter measurement. J Lab Med. 2020;44:255-272. doi: 10.1515/labmed-2020-0037
- Mannuß S, Schuff-Werner P, Dreißiger K, Kohlschein P. Magnesium sulfate as an alternative in vitro anticoagulant for the measurement of platelet parameters? Am J Clin Pathol. 2016;145:806-814. doi:10. 1093/ajcp/aqw066
- Mannuß S, Kohlschein P, Dreißiger K, Schuff-Werner P. Measurement of platelet counts and volume using magnesium sulfate as an anticoagulant: comparison of impedance and light-scatter technology. Am J Clin Pathol. 2016;146:538-545. doi:10.1093/ajcp/aqw146
- Sheu JR, Hsiao G, Shen MY, et al. Mechanisms involved in the antiplatelet activity of magnesium in human platelets. *Br J Haematol.* 2002; 119:1033-1041. doi:10.1046/j.1365-2141.2002.03967.x
- Seo JY, Lee ST, Kim SH. Performance evaluation of the new hematology analyzer Sysmex XN-series. Int J Lab Hematol. 2015;37:155-164. doi:10.1111/ijlh.12254
- Park SH, Park CJ, Kim MJ, et al. The Sysmex XN-2000 hematology AutoAnalyzer provides a highly accurate platelet count than the former Sysmex XE-2100 system based on comparison with the CD41/CD61 Immunoplatelet reference method of flow cytometry. Ann Lab Med. 2014;34:471-474, doi:10.3343/alm.2014.34.6.471
- Schoorl M, Schoorl M, Oomes J, Van Pelt J. New fluorescent method (PLT-F) on Sysmex XN2000 hematology analyzer achieved higher accuracy in low platelet counting. Am J Clin Pathol. 2013;140:495-499. doi:10.1309/AJCPUAGGB4URL5XO
- Cornbleet PJ, Gochman N. Incorrect least-squares regression coefficients in method-comparison analysis. Clin Chem. 1979;25(3): 432-438.
- Tanaka Y, Tanaka Y, Gondo K, et al. Performance evaluation of platelet counting by novel fluorescent dye staining in the XN-series automated hematology analyzers. J Clin Lab Anal. 2014;28:341-348. doi: 10.1002/jcla.21691
- 15. Clinical and Laboratory Standards Institute, ed. *EPO9c: measurement procedure comparison and bias estimation using patient samples.* 3rd ed. Clinical and Laboratory Standards Institute; 2018.
- Wu CFJ. Jackknife, bootstrap and other resampling methods in regression analysis. Ann Statist. 1986;14:1261-1295. doi:10.1214/ aos/1176350142

- 17. Armitage P, Berry G, Matthews JNS. Statistical Methods in Medical Research. 4th ed. Blackwell Science; 2002.
- 18. EFLM Biological Variation Database. https://biologicalvariation.eu
- Buoro S, Seghezzi M, Manenti B, et al. Biological variation of platelet parameters determined by the Sysmex XN hematology analyzer. *Clin Chim Acta*. 2017;470:125-132. doi:10.1016/j.cca. 2017.05.004
- Weber D, Nakashima MO. Platelet count in sodium citrateanticoagulated whole blood: comparison to EDTA-anticoagulated results and stability over time. *Int J Lab Hematol*. 2021;43:e35-e37. doi:10.1111/ijlh.13350
- François D, Masure A, Atallah N, Touil L, Vasse M. Underestimation of platelet count on magnesium salt-anticoagulated samples. Clin Chem Lab Med. 2014;52:e95-e97. doi:10.1515/cclm-2013-1016

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