

Multi-modal MRI reveals changes in placental function following preterm premature rupture of membranes

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Purpose: Preterm premature rupture of membranes complicates up to 40% of premature deliveries. Fetal infection may occur in the absence of maternal symptoms, delaying diagnosis and increasing morbidity and mortality. A noninvasive antenatal assessment of early signs of placental inflammation is therefore urgently required.

Methods: Sixteen women with preterm premature rupture of membranes < 34 weeks gestation and 60 women with uncomplicated pregnancies were prospectively recruited. A modified diffusion-weighted spin-echo single shot EPI sequence with a diffusion preparation acquiring 264 unique parameter combinations in < 9 min was obtained on a clinical 3 Tesla MRI scanner. The data was fitted to a 2-compartment T_2^* -intra-voxel incoherent motion model comprising fast and slowly circulating fluid pools to obtain quantitative information on perfusion, density, and tissue composition. Z values were calculated, and correlation with time from between the rupture of membranes and the scan, gestational age at delivery, and time between scan and delivery assessed.

Results: Placental T_2^* was significantly reduced in preterm premature rupture of membranes, and the 2-compartmental model demonstrated that this decline is mainly linked to the perfusion component observed in the placental parenchyma. Multi-modal MRI measurement of placental function is linked to gestational age at delivery and time from membrane rupture.

Conclusion: More complex models and data acquisition can potentially improve fitting of the underlying etiology of preterm birth compared with individual single-contrast models and contribute to additional insights in the future. This will need validation in larger cohorts. A multi-modal MRI acquisition between rupture of the membranes and delivery can be used to measure placental function and is linked to gestational age at delivery.

KEYWORDS

Diffusion MRI, pregnancy, preterm birth, Relaxometry

1 | INTRODUCTION

Preterm premature rupture of membranes (PPROM) complicates up to 40% of deliveries less than 37 weeks gestation.¹ Neonatal morbidity, including sepsis, cystic periventricular leukomalacia, intraventricular hemorrhage, and later development of cerebral palsy, are significantly higher among pregnancies with PPRM complicated by infection, confirmed by histopathological evidence of chorioamnionitis postdelivery.² Fetal infection can occur in the absence of overt clinical signs in the mother³ and is therefore often a retrospective diagnosis postdelivery with acute inflammatory and vascular lesions found in over 20% to 40% of assessed placentas.^{4,5}

The current retrospective diagnosis limits optimal timing of antenatal interventions such as corticosteroids or delivery. Current clinical practice is to delay delivery until 37 weeks in the absence of overt signs of infection.⁶ There is a pressing need for noninvasive antenatal assessment of early signs of placental inflammation that may be indicative of fetal infection during this prolonged time window. Current biomarker techniques proposed include the maternal neutrophil-lymphocyte ratio,⁷ but this is not routinely used in clinical practice and evidence for benefit is still outstanding.

Blood reaches the spiral arteries from the uterine arteries, allowing a continuous and slow supply of highly oxygenated maternal blood to the placenta. Blood enters the intervillous spaces, bathing the fetal vasculature contained in villous trees. Fetal blood resides in hairpin-like villous capillaries without mixing with the maternal blood. Inflammatory responses to maternal infection are associated with an increased concentration of neutrophils and cytokines in the placenta and increased deposition of placental fibrin in later stages.³ Changes in both the molecular properties of placental tissue as well as in the microstructure are thus expected. Multi-modal MRI techniques are ideally suited to assess the structural and functional properties of tissues and can therefore play a crucial role in helping to discriminate between acute and chronic phases of inflammation *in vivo*. MRI-based techniques have been successfully used in inflammatory processes in the liver⁸ and heart.⁹

Among the most promising placental MRI techniques are T_2^* relaxometry and diffusion MRI. T_2^* focuses on the molecular properties of tissue, especially exploiting the blood-oxygen level dependency (BOLD), which correlates the concentration of deoxygenated hemoglobin with the T_2^* values. Typically used multi-echo gradient-echo sequences sample data at multiple TEs to allow mono-exponential fitting to obtain T_2^* maps.

Diffusion MRI can reveal microstructural characteristics by fitting models to data acquired using

gradients of different strength and direction (*b* value/*b* vector).

One commonly used model is the intravoxel incoherent motion (IVIM) model, exploiting the bi-phase behavior of the MRI signal decay with growing *b* value to obtain a pseudo-diffusivity—associated with water in perfusing blood—and a diffusivity value—associated with water diffusing in tissue. The placenta's unique vascular structure—in particular, the fact that maternal blood slowly perfuses through the intra-villous space—complicates interpretation of IVIM model parameters. It is possible that a proportion of perfusing maternal blood will pseudo-diffuse slowly enough to show up in the diffusion compartment.

Both T_2^* and diffusion MRI have been successfully used for placental assessment *in vivo* to visualize and quantify microstructural differences in conditions such as chronic hypertension, preeclampsia,^{10,11,12} and fetal growth restriction.^{13,14} There has been recent focus on the IVIM model, showing increased perfusion fraction inside the placental parenchyma compared to control cases,¹⁵ reduced perfusion fraction in placentae with evidence of fetal vascular malperfusion,¹⁶ and reduced birth weight,^{17,18} as well as good robustness of the placental perfusion fraction measurements.¹⁹

In addition to studies focusing on T_2^* or diffusion individually, joint modeling and acquisition approaches for diffusion-relaxometry experiments have been performed. Analytic techniques include a 2-compartment model, which has been successfully used in fetal growth restriction,²⁰ as well as 2-compartment models with Laplacian approaches employed to assess preeclampsia.²¹ These more complex models reveal distinct structural and functional phenotypes. Finally, new acquisition techniques have been proposed, allowing the acquisition of data necessary for such combined experiments in 1 scan. This provides a more efficient and approach to motion.²¹⁻²³

Such combined models potentially fit the complex physiological processes expected in *in-utero* infection and inflammatory responses well but have not been deployed to date. In the context of PPRM, fetal MRI poses additional challenges. The reduced amount of amniotic fluid changes the properties of the magnetic field and thus incurs additional artifacts and changes the contrast surrounding the placenta, leading to more difficult segmentation. So far, MRI in PPRM was limited to volumetric assessment of the brain,²⁴ lung,²⁵ and thymus,²⁶ as well as to the described T_2^* and diffusion techniques in isolation, showing decreased mean T_2^* and ADC.²⁷

This study presents an efficient multimodal T_2^* -diffusion acquisition with dedicated multi-compartmental combined diffusion-relaxometry analysis to study

differences in placentas affected by PPRM compared to those from control pregnancies.

2 | METHODS

Women at high risk of preterm delivery were prospectively recruited from St Thomas' Hospital (London, UK) between April 2019 and March 2022 as part of 3 ethically approved studies. (Not all eligible women were approached during this time period, only during periods when the study team was recruiting.) Inclusion criteria were gestational age between 20 and 34 weeks and PPRM, confirmed on clinical assessment at vaginal speculum examination recruited from the antenatal ward or maternity assessment unit, or pregnancies considered low risk at the time of study entry. Exclusion criteria were known structural or chromosomal abnormalities, multiple pregnancies (twins, triplets), active labor, maternal inability to give informed consent, pregnancy complications including preeclampsia and fetal growth restriction, and contraindications to MRI such as claustrophobia or a recently sited metallic implant.

Key clinical parameters included maternal age and body mass index at booking, as well as previous medical and obstetric history such as previous term or preterm deliveries, current medication history, date of rupture of membranes, and any treatment received. Furthermore, extensive pregnancy outcome data were obtained, including birth weight, gestational age at delivery, Apgar score, and details regarding any neonatal admissions. The time between membrane rupture and scan and the time between scan and delivery were calculated as displayed in Figure A. Maternal neutrophil-lymphocyte ratio values were obtained from the majority of the PPRM participants within days of the MRI scan but not from the low-risk participants.

Participants were scanned in supine position on a 3 Tesla Philips Achieva scanner (Philips, Best, Netherlands) using a cardiac 32-channel coil after informed consent was obtained. Constant life monitoring using an in vivo device assessing heart rate, oxygen saturation, and blood pressure was performed. Frequent verbal interaction was maintained throughout the scan. The scan lasted 60 min, with a break after approximately 30 min for maternal comfort. The acquisition is illustrated schematically in Figure B.

2.1 | Acquisition protocol

After pilot and calibration scans, T_2 -weighted turbo spin echo sequences were performed covering the entire uterus in sagittal and coronal plane to maternal habitus as well as at 45-degree angles sagittal focusing on the fetal brain. The following scanning parameters were used: TR = 25 s,

TE = 80 ms, slice thickness of 2.5 mm, slice overlap of 1.25 mm, with a flip angle = 90°.

A B_0 map covering the entire uterus was acquired, and image-based shimming focusing on the placenta was performed,²⁸ restricted to the placental parenchyma and carefully avoiding maternal bowel and fetal structures. The latter are of particular importance in the absence of amniotic fluid. Data was then acquired with a multi-slice 2D multi-echo gradient echo sequence (MEGE) covering the entire uterus (resolution 3 mm isotropic, 5 TE in [18.159] ms, TR = 3 s) in under 1 min, and a modified spin-echo diffusion-weighted single-shot EPI sequence using the thus obtained shim values.²² The parameters were chosen such that 3 mm isotropic resolution could be achieved while limiting the acoustic output of the sequence, resulting in FOV = 340 mm × 400 mm × 28 mm, TR = 3.4 sec, SENSE = 2, minimal TE = 78 ms. This approach was taken based on previous work,²⁹ with measurements at iso-center performed for different echo-spacing values. Three subsequent TEs were chosen by minimizing delay time between subsequent readout trains. This resulted in 4 TEs being chosen as [78, 114, 150, and 186] ms, with the readout length of 36 ms setting the deltas. The diffusion preparation parameters were selected as previously optimized.³⁰ They include 3 diffusion gradient directions at $b = [5, 10, 25, 50, 100, 200, 400, 600, 1200, 1600]$ s mm; 8 directions at $b = 18$ s mm; 7 at $b = 36$ s mm, and 15 at $b = 800$ s mm. In total, 256 (66*4) unique preparations were acquired. Both MEGE and T_2^* -diffusion data was acquired in the coronal plane to maternal habitus, resulting in a total acquisition time for both of under 10 min.

2.2 | Reconstruction, motion correction, and selection of the region of interest

The data was reconstructed using in-house tools³¹ and motion corrected using Ants.³² For the motion correction, a template was constructed from the average of volumes acquired with the lowest TE. Then, registration was performed, resulting in a set of transformations correcting the data from the first echo. These were then applied to the other echoes, exploiting the fact that the data for all echoes for each slice is acquired in < 200 ms, hence not presenting significant motion artifacts. The placental parenchyma was manually segmented on all slices of the second TE of the MEGE and on the motion-corrected T_2^* -diffusion dataset, avoiding amniotic fluid on the chorionic plate and maternal vasculature on the basal plate.

Mono-exponential fitting was performed on the MEGE dataset, resulting in T_2^* maps (MEGE- T_2^*) and proton density (MEGE- S_0) maps. Analysis of the T_2^* -diffusion datasets

was performed using in-house python scripts (version 1.0) and our extensions to the dmipy python library for diffusion models.^{33,34} These included both a simpler biexponential model of T_2^* and ADC decay (T_2^* ADC model; see Equation 1, where T_2^* and ADC are the unknowns), and a T_2^* -IVIM model including slow and fast diffusion in 2 compartments and representing diffusing and perfusing blood within the placenta (IVIM model; see Equation 2).

$$T_2^* \text{ ADC model: } S(T_E, b) = S_0 e^{-(T_E - T_{Emin})/T_2^*} e^{-bADC} \quad (1)$$

$$T_2^* \text{ IVIM Model: } S(T_E, b) = S_0 \left[f e^{-(T_E - T_{Emin})/T_{2fast}^*} e^{-bD^*} + (1-f) e^{-(T_E - T_{Emin})/T_{2slow}^*} e^{-bADC} \right], \quad (2)$$

whereby T_E is the TE; T_{Emin} is the lowest TE acquired; b is the b value; ADC is the apparent diffusion coefficient; T_2^* is the effective transverse relaxation time; and S_0 is the signal at the lowest TE with zero diffusion weighting. The resulting parameters are the T_2^* of the perfusion compartment T_{2fast}^* and of the diffusion compartment T_{2slow}^* , the pseudo-diffusivity D^* (or ADC1) describing the perfusion, and the ADC in the diffusion compartment ADC (ADC2), as well as the fraction f expressing the percentage of each voxel belonging to the perfusion compartment.

2.3 | Statistical analysis

Linear regression was performed on all the control cases in python using scipy, resulting in the slope, intercept, P value, and R2 score. A P value < 0.005 was considered significant. Next, z-scores were calculated based on the control cohort using the method by DeVore et al. for cross-sectional cohorts.³⁵ The z-scores for the PPRM cohort were obtained, and the correlation between these and 3 key measures regarding PPRM (gestation at delivery, time between scan and delivery, and time between ROM and scan) were calculated using linear regression.

Slope controls over gestational age	R ² /P value value controls over gestational age	P value from the z-scores cohort difference
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3 | RESULTS

Sixteen PPRM participants and 60 healthy control participants were included in this study, and clinical

characteristics are given in Supporting Information Figure 1 and Supporting Information Table 4, highlighting the lower gestational age at delivery (29 vs. 40 weeks gestational age) as expected, the maintained birth weight centile (59 vs. 60), similar age, and slightly increased body mass index (24 vs. 21) in the PPRM group, as well as decreased time between scan and delivery in the PPRM group.

Figure A–2B display the signal evolution curves for 1 exemplary case over all dynamics, varying TE while also varying the diffusion preparation for each echo as outlined above for 3 depicted regions: lobule center, septa, and chorionic plate.

Parametric maps for both considered models for the T_2^* -ADC and the T_2^* -IVIM models for 1 case are shown in Figure C, displaying reformatted views in all 3 orientations for each measure. The perfusion compartment thus dominates in the centers of the lobules where the spiral arteries enter the placental parenchyma as well as closer to the chorionic plate. The slow diffusion compartment dominates in the intervillous spaces but distant from the spiral arteries inflow.

The quantitative results for all 3 models in Figure 3 display the changes over gestation and between the cohorts; these formed the basis for the calculated z-scores. Thereby, significant correlation for the control cohort over gestational age could be found for T_2^* , T_2^* from T_2^* -ADC, T_{2fast}^* from T_2^* -IVIM, and ADC_{slow} from T_2^* -IVIM (all < 0.005), as well as for the perfusion fraction. No correlation with gestational age was found for the PPRM cohort (see Table 1).

The z-scores for the PPRM cases were thereby significantly reduced for T_2^* ($P < 0.05$) (Figure A), T_2^* from T_2^* -ADC ($P < 0.005$) (Figure C), and for the T_{2fast}^* (perfusion compartment) ($P < 0.005$) (Figure E). The T_{2slow}^* (diffusion compartment) showed a trend toward significant reduction ($P = 0.069$). The z-scores for all ADC values did not show a significant difference ($P = 0.42$ for ADC from T_2^* -ADC, $P = 0.59$ and 0.15 for ADC for perfusing and diffusing compartment, respectively), neither did the perfusion fraction ($P = 0.39$).

Next, the correlation between the z-scores for all the quantitative measures and 3 key measures regarding PPRM, gestation at delivery, time between scan and delivery, and time between ROM and scan were investigated. The results show significant correlations for the ADC in the compartment mainly dominated by the perfusion compartment, that is, with the time of delivery, the time since ROM and between the T_{2slow}^* (diffusing compartment), and time between scan and delivery ($P < 0.05$).

FIGURE 1 Overview of study showing (A) the time course of investigations, and (B) the MRI protocol and processing schemata describing the data flow

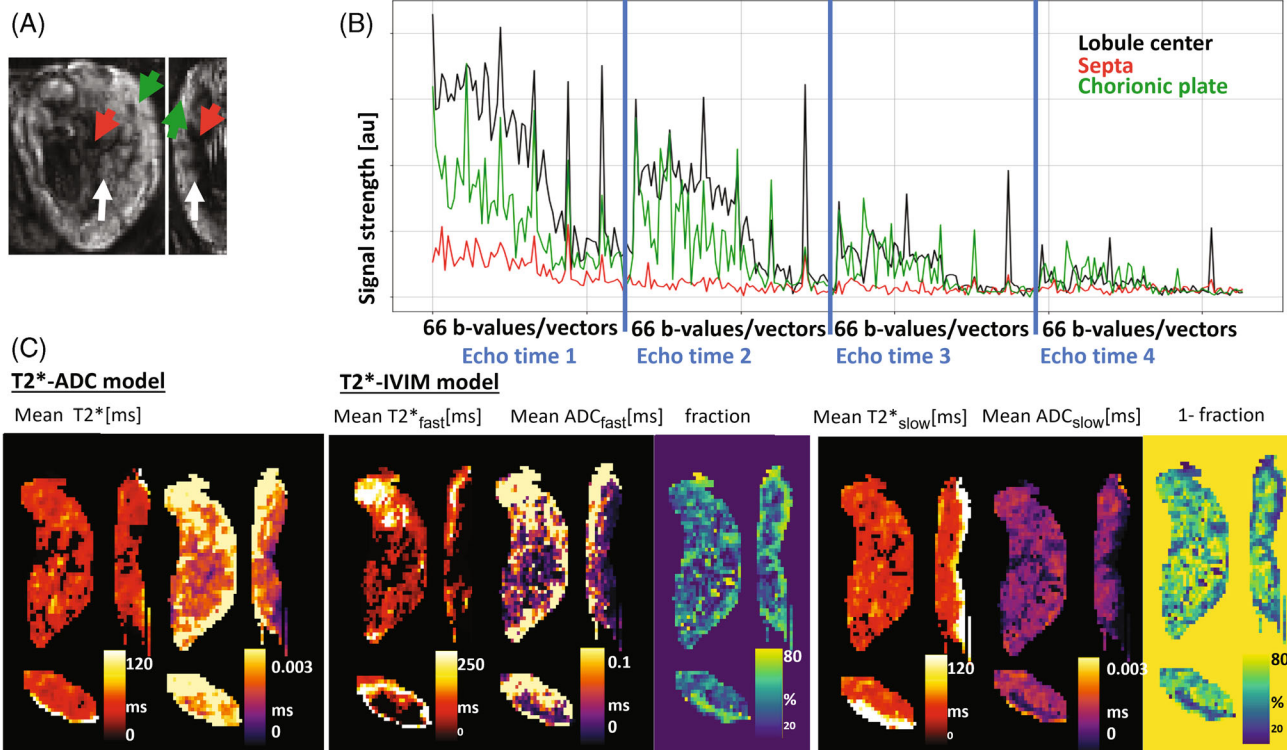
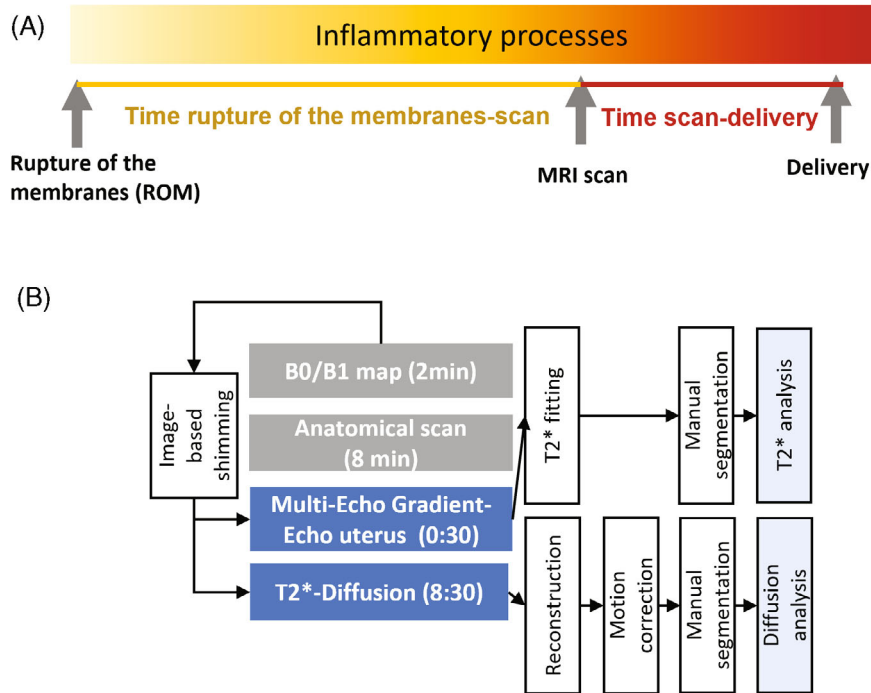


FIGURE 2 Boxplots for all considered quantities. Top row: T_2^* , ADC from T_2^* -ADC fit, T_2^* from T_2^* -ADC fit, ADC1 from IVIM fit. Bottom row: ADC2 from IVIM fit, T_2^* 1 from IVIM fit, T_2^* 2 from IVIM fit, fraction from IVIM

4 | DISCUSSION

This study demonstrated the first results of a simultaneous diffusion-relaxometry MRI acquisition and

multi-compartmental analysis in pregnant women with PPROM. Reduced mean placental T_2^* was significantly correlated with PPROM both for the values obtained with a T_2^* model alone ($P < 0.05$), a T_2^* -Diffusion model

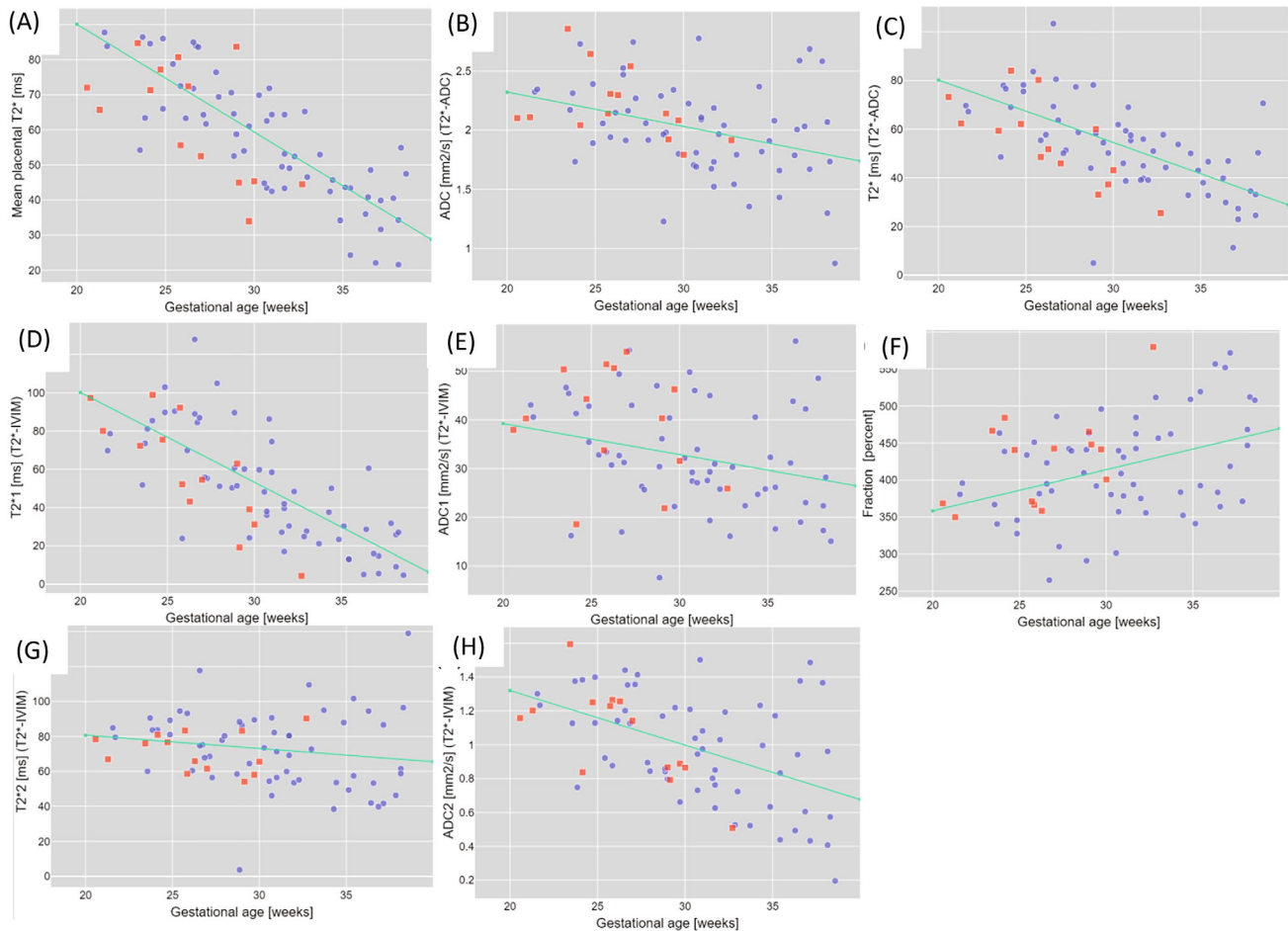


FIGURE 3 (A) Selected regions of interest in the central lobule areas (indicated by white arrow), the septa (indicated by red arrow), and the basal plate (indicated by green arrow) together with (B) signal evolution over all acquired data points. (C) Quantitative maps obtained using T_2^* -ADC and the T_2^* -IVIM model, displaying T_2^* and ADC map; the T_2^* , ADC map and tissue fraction map of the perfusion compartment; and the T_2^* , ADC, and fraction map of the diffusion compartment from the IVIM (intravoxel incoherent motion) model

TABLE 1 Quantitative results for the control values over gestation and the P values for the z -score cohort difference

$N = 16$ PPROM $N = 60$ controls	Slope controls over Gestational Age	R^2/P value controls over Gestational Age	p-value from the z-scores cohort difference
T_2^*	-3.004 ms/week	0.81/<0.005	0.014
ADC from T_2^*-ADC	-0.035	0.37/<0.005	0.429
T_2^* from T_2^* -ADC	-2.615 ms/week	0.65/<0.005	<0.005
ADC1 from T_2^*-IVIM	-0.764	0.31/0.016	0.589
ADC2 from T_2^*-IVIM	-0.033	0.49/<0.005	0.147
T_2^*1 from T_2^* -IVIM	-4.791 ms/week	0.57/<0.005	<0.005
T_2^*2 from T_2^* -IVIM	-0.671 ms/week	0.16/0.214	0.069
Fraction perfusion	5.726%/week	0.39/<0.005	0.394

Note: The bold indicates cut-off for significance at $p < 0.005$.

Abbreviations: IVIM, intravoxel incoherent motion; PPROM, preterm premature rupture of membranes.

($P < 0.005$), and the more complex IVIM model for the perfusion compartment ($P < 0.005$). The latter bicompartimental model result and stronger level of significance demonstrated in this study give evidence that the decay

is driven by the fast-flowing perfusion compartment, mostly attributed to maternal inflowing blood. Interestingly, the perfusion fraction, describing how many voxels are predominantly perfusing fast-flowing blood, trends

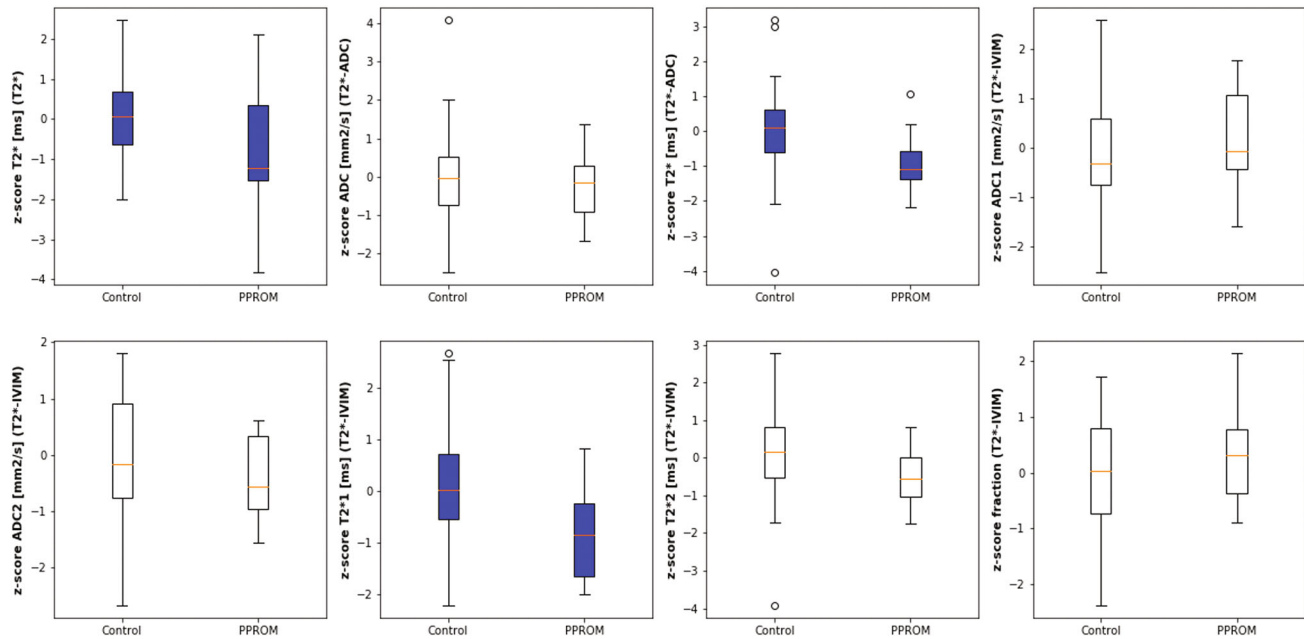


FIGURE 4 Quantitative results for the T_2^* data (A) for the T_2^* (B), and ADC (C) for the T_2^* -ADC fit and for the T_2^* -IVIM fit (D, E, F, G, H, I) The control subjects are marked with blue squares and the PPRM cases with red dots; the green line represents the regression line obtained for the controls PPRM, preterm premature rupture of membranes.

toward higher values for PPRM. This could potentially correspond to an increase in neutrophils and cytokines in the maternal arteries toward the intervillous spaces as described in histopathological studies of PPRM placentas.³ However, the perfusion compartment dominated also on the fetal chorionic plate. Potentially this could hint toward an increase in fetal blood supply, in itself a sign of fetal inflammatory response. Further investigations would however be required to assert this. A larger cohort would be required to investigate these spatial differences in more detail. The use of such more complex models might thus allow us in the future to work toward a better understanding of the pathophysiology underlying PPRM and open new focused avenues to understand the observed differences.

We observed in our data also an increase in perfusion fraction for healthy controls over gestation, in accordance with some previous studies.^{15,36} An increase in perfusion in the intervillous space would be in line with studies using ultrasound bubbles showing an increase over gestation,³⁷ as well as an increase in blood flow to the fetoplacental compartment as the pregnancy progresses.

This study, however, has a number of important limitations. These include the number of subjects; recruiting pregnant women with PPRM is a significant challenge both due to the high level of clinical surveillance required and the unpredictable interval between rupture of membranes and delivery. This complicates the scheduling of

an MRI scans. This limitation furthermore makes longitudinal within-subject follow-up very complicated and thus hampers our ability to study the progression of inflammatory processes. However, as outlined in the introduction, the clinical utility of MRI may be in cases with diagnostic uncertainty where no maternal signs of infection are apparent, and deferral of delivery until 37 weeks is often advocated.

Next, the choice of the region of interest for the analysis is critical. This study has focused on the placental parenchyma, carefully avoiding maternal tissues, for example, in the basal plate. This was chosen with the aim to understand both the placental involvement and to avoid inaccuracy in the segmentation of the thin maternal tissues. Future studies could focus on regions of interest in the basal plate and inclusion of perfusion techniques, measuring noninvasively the inflow of blood into the tissue directly.³⁸⁻⁴⁰

5 | CONCLUSION

Data was successfully acquired in a challenging group of patients and analysis revealed the first correlations between quantitative modal results from the placenta and the time between rupture of the membranes and delivery. This combination of efficient data acquisition and multi-compartmental modeling allows a comprehensive

study of microstructure and function and can thus potentially play a role in assessing the impact of inflammation and infection on placental function. Future work will link to other longitudinal ultrasound and serum markers to further enhance information about these pregnancies and thus further optimize delivery and intervention.

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DATA AVAILABILITY STATEMENT

Access to the data is available to all academic parties by email to the corresponding author in line with the ethical approval of this study. The code used for the analysis is available under <https://github.com/Jamahu/pypla.git>.

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REFERENCES

- Morris JM, Roberts CL, Bowen JR, et al. Immediate delivery compared with expectant management after preterm pre-labour rupture of the membranes close to term (PPROMT trial): a randomised controlled trial. *Lancet*. 2016;387:444-452.
- Ramsey PS, Lieman JM, Brumfield CG, Carlo W. Chorioamnionitis increases neonatal morbidity in pregnancies complicated by preterm premature rupture of membranes. *Am J Obstet Gynecol*. 2005;192:1162-1166.
- Romero R, Espinoza J, Gonçalves LF, Kusanovic JP, Friel LA, Nien JK. Inflammation in preterm and term labour and delivery. *Semin Fetal Neonatal Med*. 2006;11:317-326.
- Arias F, Victoria A, Cho K, Kraus F. Placental histology and clinical characteristics of patients with preterm premature rupture of membranes. *Obstet Gynecol*. 1997;89:265-271.
- Armstrong-Wells J, Post MD, Donnelly M, Manco-Johnson MJ, Fisher BM, Winn VD. Patterns of placental pathology in preterm premature rupture of membranes. *J Dev Orig Health Dis*. 2013;4:249-255.
- Thomson AJ, Royal College of Obstetricians and Gynaecologists. Care of Women Presenting with suspected preterm Prelabour rupture of membranes from 24+0 weeks of gestation: green-top guideline no. 73. *BJOG*. 2019;126:e152-e166.
- Balciuniene G, Kvederaite-Budre G, Gulbiniene V, et al. Neutrophil-lymphocyte ratio for the prediction of histological chorioamnionitis in cases of preterm premature rupture of membranes: a case-control study. *BMC Pregnancy Childbirth*. 2021;21:656.
- Guimaraes AR, Siqueira L, Uppal R, et al. T₂ relaxation time is related to liver fibrosis severity. *Quant Imaging Med Surg*. 2016;6:103-114.
- Antonini-Canterin F, Faganello G, Mantero A, et al. Cardiovascular multimodality imaging: it is time to get on board! A 'Società Italiana di Ecocardiografia e CardioVascular imaging' statement. *J Cardiovasc Echogr*. 2018;28:1-8.
- Ho AEP, Hutter J, Jackson LH, et al. T₂* placental magnetic resonance imaging in preterm preeclampsia. *Hypertension*. 2020;75:1523-1531.
- Ho A, Hutter J, Slator P, et al. Placental magnetic resonance imaging in chronic hypertension: a case-control study. *Placenta*. 2020;104:138-145.
- Slator PJ, Ho A, Bakalis S, et al. Anisotropy in the human placenta in pregnancies complicated by fetal growth restriction. In: Özarslan E, Schultz T, Zhang E, Fuster A, eds. *Anisotropy Across Fields and Scales*: Springer; 2021; 263-276.
- Sinding M, Sørensen A, Hansen DN, Peters DA, Frøkjær JB, Petersen AC. T₂* weighted placental MRI in relation to placental histology and birth weight. *Placenta*. 2021;114:52-55.
- Sinding M, Peters DA, Frøkjær JB, et al. Placental magnetic resonance imaging T₂* measurements in normal pregnancies and in those complicated by fetal growth restriction. *Ultrasound Obstet Gynecol*. 2016;47:748-754.
- Moore RJ, Strachan BK, Tyler DJ, et al. In utero perfusing fraction maps in normal and growth restricted pregnancy measured using IVIM echo-planar MRI. *Placenta*. 2000;21:726-732.

16. Malmberg M, Kragsterman E, Sinding M, et al. Perfusion fraction derived from IVIM analysis of diffusion-weighted MRI in the assessment of placental vascular malperfusion antenatally. *Placenta*. 2022;119:1-7.
17. Siauve N, Hayot PH, Deloison B, et al. Assessment of human placental perfusion by intravoxel incoherent motion MR imaging. *J Matern Fetal Neonatal Med*. 2019;32:293-300.
18. Derwig I, Lythgoe DJ, Barker GJ, et al. Association of placental perfusion, as assessed by magnetic resonance imaging and uterine artery Doppler ultrasound, and its relationship to pregnancy outcome. *Placenta*. 2013;34:885-891.
19. Jakab A, Tuura R, Kottke R, Kellenberger CJ, Scheer I. Intra-voxel incoherent motion MRI of the living human foetus: technique and test-retest repeatability. *Eur Radiol Exp*. 2017;1:26.
20. Aughwane R, Mufti N, Flouri D, et al. Magnetic resonance imaging measurement of placental perfusion and oxygen saturation in early-onset fetal growth restriction. *BJOG*. 2021;128:337-345.
21. Slator PJ, Hutter J, Marinescu RV, et al. Data-driven multi-contrast spectral microstructure imaging with InSpect: INtegrated SPECTral component estimation and mapping. *Med Image Anal*. 2021;71:102045.
22. Hutter J, Slator PJ, Christiaens D, et al. Integrated and efficient diffusion-relaxometry using ZEBRA. *Sci Rep*. 2018;8:15138.
23. Slator P, Hutter J, Marinescu R, Palombo M, Alexander DC. InSpect: INtegrated SPECTral component estimation and mapping for multi-contrast microstructural MRI. In *Proceedings of 26th International Conference on Information Processing in Medical Imaging (IPMI)*. IPMI conference; 2019. doi:10.1007/978-3-030-20351-1_59
24. Story L, Davidson A, Patkee P, et al. Brain volumetry in fetuses that deliver very preterm: an MRI pilot study. *Neuroimage Clin*. 2021;30:102650.
25. Story L, Zhang T, Steinweg JK, et al. Foetal lung volumes in pregnant women who deliver very preterm: a pilot study. *Pediatr Res*. 2020;87:1066-1071.
26. Story L, Zhang T, Uus A, et al. Antenatal thymus volumes in fetuses that delivered <32 weeks' gestation: an MRI pilot study. *Acta Obstet Gynecol Scand*. 2021;100:1040-1050.
27. Hutter J, Jackson L, Ho A, et al. The use of functional placental magnetic resonance imaging for assessment of the placenta after prolonged preterm rupture of the membranes in vivo: a pilot study. *Acta Obstet Gynecol Scand*. 2021;100:2244-2252.
28. Gaspar, A. S., Nunes R. G., Ferrazzi G., et al. Optimizing maternal fat suppression with constrained image-based shimming in fetal MR. *Magn Reson Med*. 2019;81:477-485.
29. Hutter J, Price AN, Cordero-Grande L, et al. Quiet echo planar imaging for functional and diffusion MRI. *Magn Reson Med*. 2018;79:1447-1459.
30. Slator PJ, Hutter J, McCabe L, et al. Placenta microstructure and microcirculation imaging with diffusion MRI. *Magn Reson Med*. 2018;80:756-766.
31. Cordero-Grande L, Christiaens D, Hutter J, Price AN, Hajnal JV. Complex diffusion-weighted image estimation via matrix recovery under general noise models. *Neuroimage*. 2019;200:391-404.
32. Avants B, Epstein C, Grossman M, Gee J. Symmetric diffeomorphic image registration with cross-correlation: evaluating automated labeling of elderly and neurodegenerative brain. *Med Image Anal*. 2008;12:26-41.
33. Garyfallidis E, Brett M, Amirbekian B, et al. Dipy, a library for the analysis of diffusion MRI data. *Front Neuroinform*. 2014;8:8.
34. Fick RHJ, Wassermann D, Deriche R. The Dmipy toolbox: diffusion MRI multi-compartment modeling and microstructure recovery made easy. *Front Neuroinform*. 2019;13:64.
35. DeVore GR. Computing the Z score and centiles for cross-sectional analysis: a practical approach. *J Ultrasound Med*. 2017;36:459-473.
36. Sohlberg S, Mulic-Lutvica A, Olovsson M, et al. Magnetic resonance imaging-estimated placental perfusion in fetal growth assessment. *Ultrasound Obstet Gynecol*. 2015;46:700-705.
37. Roberts VHJ, Lo JO, Salati JA, et al. Quantitative assessment of placental perfusion by contrast-enhanced ultrasound in macaques and human subjects. *Am J Obstet Gynecol*. 2016;214:e1-e8.
38. Hartevelde AA, Hutter J, Franklin SL, et al. Systematic evaluation of velocity-selective arterial spin labeling settings for placental perfusion measurement. *Magn Reson Med*. 2020;84:1828-1843.
39. Hutter J, Hartevelde AA, Jackson LH, et al. Perfusion and apparent oxygenation in the human placenta (PERFOX). *Magn Reson Med*. 2020;83:549-560.
40. Deloison B, Salomon LJ, Quibel T, et al. Non-invasive assessment of placental perfusion in vivo using arterial spin labeling (ASL) MRI: a preclinical study in rats. *Placenta*. 2019;77:39-45.

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of the article at the publisher's website.

Figure S1: Cohort characteristics for the PPRM subjects (blue) and the control subjects (red) depicting the distribution of birth weight centile, gestational age @ delivery, time between rupture of the membranes and scan and time between scan and delivery.

Table S1: Patient cohort characteristics for the control and the PPRM cohort. Significant results ($P < 0.005$) are colored in blue.

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