

PRAME Expression in Mucosal Melanoma of the Head and Neck Region

Costantino Ricci, MD,*† Maria V. Altavilla, MD,‡§ Barbara Corti, MD,‡ Ernesto Pasquini, MD,||
Livo Presutti, MD,¶ Anna M. Baietti, MD,# Luca Amorosa, MD,** Tiziana Balbi, MD,‡
Chiara Baldovini, MD, PhD,‡ Francesca Ambrosi, MD,*† Marco Grillini, BSc,‡
Antonia D'Errico, MD, PhD,†‡ Michelangelo Fiorentino, MD, PhD,*†
and Maria P. Foschini, MD††

Abstract: PRAME (PReferentially expressed Antigen in MELanoma), a cancer-testis antigen expressed in normal and neoplastic tissues with several functions, proved to be a useful diagnostic tool in the differential diagnosis between benign and malignant melanocytic lesions. The current study aims to perform PRAME stain on a retrospective case series of mucosal melanocytic tumors of the head and neck region to compare 3 different scores and evaluate the most reliable one in this diagnostic set. Immunohistochemical analysis for PRAME was performed in 54 benign and malignant mucosal melanocytic tumors of the head and neck region collected from 41 patients. The best-performing cutoff of PRAME-positive cells (nuclear stain) to differentiate benign and malignant mucosal melanocytic tumors of the head and neck region is that proposed

by Raghavan and colleagues (< 60%/ ≥ 60% of PRAME-positive cells), with 100% and 77.8% of benign lesions and malignant tumors respectively correctly identified. Applying this score, PRAME stain showed the best results (sensitivity, specificity, accuracy, and positive and negative predictive values) for the diagnosis of head and neck melanocytic tumors. However, a subset of PRAME-negative malignant tumors was identified, especially located in the palatal area (hard and soft palate). Finally, high PRAME expression (≥ 60%) was associated with specific sites (nasal cavity/nasal septum/turbinates nasopharynx, and the maxillary sinus), nodular histotype, and female sex.

Key Words: PRAME, melanocytic, melanoma, mucosa, head & neck, immunohistochemistry

(*Am J Surg Pathol* 2023;47:599–610)

Received for publication November 8, 2022; accepted January 18, 2023.

From the *Pathology Unit; **ENT Unit, Surgical Department, Maggiore Hospital-AUSL Bologna; #Maxillo-Facial Operative Unit, Bellaria and Maggiore Hospital-AUSL Bologna; †Department of Experimental, Diagnostic, and Specialty Medicine (DIMES), University of Bologna; §Department of Biomedical and Neuromotor Sciences, School of Anatomic Pathology, University of Bologna; ‡Pathology Unit; ¶Otolaryngology Unit, Department of Head and Neck Surgery, IRCCS AOUBO; ||ENT Unit, Surgical Department; and ††Pathology Unit, Department of Biomedical and Neuromotor Sciences (DIBINEM), Bellaria Hospital, University of Bologna, Bologna, Italy.

C.R., M.V.A., B.C., A.D.E., M.F., and M.P.F.: performed study concept and design and wrote the manuscript. T.B., F.A., and C.B.: contributed to the histologic revision of the cases and provided analysis and interpretation of data. E.P., L.P., A.M.B., and L.A.: provided clinical data and clinical analysis of the cases. M.G.: performed development of methodology and statistical analysis.

This work has been partially presented as a poster (smaller case series, without statistical analyses and discussion) at AIOCC 2022 Congress.

Conflicts of Interest and Source of Funding: The authors have disclosed that they have no significant relationships with, or financial interest in, any commercial companies pertaining to this article.

Correspondence: Costantino Ricci, MD, Pathology Unit, Maggiore Hospital-AUSL Bologna, Largo Nigrisoli 2, Bologna 40133 (BO), Italy (e-mail: costanricci@gmail.com).

Supplemental Digital Content is available for this article. Direct URL citations appear in the printed text and are provided in the HTML and PDF versions of this article on the journal's website, www.ajsp.com.

Copyright © 2023 The Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The diagnosis of mucosal melanoma of the head and neck region (MM-H&N) is challenging due to the peculiar anatomic, histologic, and genetic features of these melanocytic tumors.^{1–6} MM-H&N are exceedingly rare and even experienced head and neck pathologists struggle to confidently diagnose this entity.^{1–6} Besides, in this site it is difficult to benefit from clinical-dermatoscopic correlation (as in the cutaneous counterpart), the samples are often poorly cellular incisional biopsies and/or highly fragmented samples, and some atypical histologic features (epidermal effacement and pagetoid spread of melanocytes in suprabasal layers) are rarely found in the early stage of disease.^{5,6} In recent years, several molecular tools, such as fluorescence in situ hybridization (FISH), comparative genomic hybridization, and next-generation sequencing have been implemented to aid in improving the diagnosis of melanocytic tumors.^{7–10} Unfortunately, the rarity of MM-H&N and the limited literature data on specific genetic mutations (as *TERT* promoter mutations) and molecular tests (FISH) distinguishing MM-H&N from benign mucosal melanocytic lesions of the head and neck region (MBML-H&N), limits their adoption in this diagnostic field.^{1–6,11–15} The recent introduction of PRAME (PReferentially expressed Antigen in MELanoma) has again shifted interest to using immunohistochemistry for the diagnosis of melanocytic

lesions.¹⁶ PRAME is a cancer-testis antigen isolated in 1997 from autologous T-lymphocytes and direct against the tumor in a patient with metastatic cutaneous melanoma.¹⁷ In recent years, PRAME was found to be expressed in normal and neoplastic tissues (mainly germ cell tumors of the testis, ovarian cancer, sarcomas, and hematologic tumors), with functions in oncogenesis, immune response evasion, apoptosis, acquisition of metastatic phenotype, and drug resistance.^{18–24} PRAME became of great interest for the diagnosis of melanocytic tumors, as it proved to be expressed in melanomas but not in nevi, therefore an immunohistochemical marker of great help in one of the most challenging issues of surgical pathology.^{16,24–41} However, the majority of previous studies on PRAME did not enroll MM-H&N and the available data on the diagnostic value of PRAME are mainly on skin melanocytic tumors; a few studies only evaluated a limited number of MM-H&N.^{16,24–28,40–42} Toyama et al⁴⁰ tested PRAME on a series of differently-sited mucosal melanomas (head and neck region, gastrointestinal, and genitourinary) and obtained promising results. Hovander et al⁴¹ confirmed these results on a small case series of oral cavity melanomas, and recently Scheurleer et al⁴² found PRAME expression in a cohort of sinonasal melanoma. The present study aims to test and validate PRAME stain as a reliable diagnostic tool in a retrospective case series of MM-H&N, comparing MM-H&N with MBML-H&N.

MATERIALS AND METHODS

Case Series (Patients and Specimens) Selection

All MM-H&N and MBML-H&N excised between 2004 and 2022 were retrieved from the database of 3 pathology units: Bellaria Hospital of Bologna (37 cases),

Maggiore Hospital of Bologna (9 cases), and IRCCS Azienda Ospedaliero-Universitaria Policlinico di Sant’Orsola, Bologna (8 cases). The patients were selected according to the following inclusion criteria: (1) primary mucosal melanocytic lesion (each case was reviewed to verify its site and exclude metastases and cases more appropriately classifiable as cutaneous, especially for lips); (2) availability of formalin-fixed and paraffin-embedded samples with enough material to perform immunohistochemical analysis; (3) availability of clinical data. By contrast, cases judged as nonprimitive of the mucous membranes, and with no availability of formalin-fixed and paraffin-embedded samples for immunohistochemical analyses and/or clinical data were excluded. For each patient, the following clinical data were recorded: age at first diagnosis and sex. For each MM-H&N sample, the following pathologic features were recorded: type of histologic specimen (excision of the primary tumor, excision of residual tumor/relapse, incisional biopsy), site, histologic subtype, pigmentation, prevalent cytotype, ulceration, bone and/or cartilage infiltration, number of mitoses/mm², lymphovascular invasion, perineural infiltration, and pT stage. For each MBML-H&N, only the following pathologic features were recorded: type of histologic specimen, site, and histologic diagnosis. All cases were reviewed and diagnosed according to the World Health Organization Blue Book on Classification of Head and Neck Tumours (fifth edition, 2022) by a panel of 4 pathologists with specific expertise in melanocytic pathology and/or head and neck pathology (C.R., B.C., T.B., and M.P.F.); all MM-H&N were staged according to the eighth edition of the American Joint Committee on Cancer (AJCC) Cancer Staging Manual.^{43,44}

TABLE 1. Clinicopathologic Features of the Case Series

	MM-H&N and MBML-H&N, n (%)	MM-H&N, n (%)	MBML-H&N, n (%)
	Patients (N = 41)	Patients (N = 23)	Patients (N = 18)
Clinical features			
Age, median value (range)	60 (10-96)	69.9 (29-96)	47.4 (10-79)
Sex			
Male	16 (39)	9 (39.1)	7 (38.9)
Female	25 (61)	14 (60.9)	11 (61.1)
	Histologic samples (N = 54)	Histologic samples (N = 36)	Histologic samples (N = 18)
Pathologic features			
Site			
Nasal cavity/nasal septum/turbinates	22 (40.7)	22 (61.1)	0 (0)
Nasopharynx	2 (3.7)	2 (5.6)	0 (0)
Palate (hard and soft)	12 (22.2)	6 (16.7)	6 (33.3)
Maxillary sinus	4 (7.4)	4 (11)	0 (0)
Tonsil	1 (1.9)	1 (2.8)	0 (0)
Gum (upper and lower)	8 (14.8)	0 (0)	8 (44.4)
Lip (upper and lower)	3 (5.6)	0 (0)	3 (16.7)
Tongue	2 (3.7)	1 (2.8)	1 (5.6)
Type of histologic specimen			
Excision of the primary tumor	37 (68.5)	24 (66.7)	13 (72.2)
Excision of residual tumor/relapse	10 (18.5)	10 (27.8)	0 (0)
Incisional biopsy	7 (13)	2 (5.5)	5 (27.8)

MM-H&N: mucosal melanoma of the head and neck region; MBML-H&N: mucosal benign melanocytic lesions of the head and neck region.

TABLE 2. Pathologic Features of MM-H&N and Histologic Diagnosis of MBML-H&N

	MM-H&N, n (%)	
	Patients (N = 23)	Samples (N = 36)
Histologic subtype		
Nodular	14 (60.9)	16 (44.4)
Mucosal lentiginous	9 (39.1)	20 (55.6)
Pigmentation		
Yes	15 (65.2)	9 (25)
No	8 (34.8)	27 (75)
Prevalent cytotype		
Epithelioid	14 (60.9)	20 (55.6)
Fused	5 (21.7)	11 (30.5)
Mixed	4 (17.4)	5 (13.9)
Ulceration		
Yes	18 (78.3)	7 (19.4)
No	5 (21.7)	29 (80.6)
Bone and/or cartilage infiltration		
Yes	6 (26.1)	7 (19.4)
No	17 (73.9)	29 (80.6)
Mitoses/mm ² , median value (range)	—	5 (1-13)
LVI		
Yes	10 (43.5)	23 (63.9)
No	13 (56.5)	13 (36.1)
PNI		
Yes	2 (8.7)	34 (94.4)
No	21 (91.3)	2 (5.6)
pT stage		
pT3	18 (78.3)	29 (80.6)
pT4a	4 (17.4)	6 (16.7)
pT4b	1 (4.3)	1 (2.8)
	MBML-H&N, n (%)	
	Patients and samples (N = 18)	
Histologic diagnosis		
Melanotic macula	10 (55.6)	
Blue nevus	6 (33.3)	
Common nevus	2 (11.1)	

Herein, we reported values for patients and samples because, for the cases of MM-H&N with multiple samples, a global (column: patients) and a single-sample (column: samples) assessment of the histologic features was performed.

LVI indicates lymphovascular invasion; PNI, perineural infiltration.

Immunohistochemistry and PRAME Scoring

For each case, a representative slide was selected, and from the corresponding paraffin-embedded tissue block one 3- μ m-thick section was cut and stained with Melan A/PRAME (BenchMark ULTRA automated immunostainer; Ventana Medical Systems-Roche Diagnostics; Melan A: clone A103, PRAME: clone EPR20330) according to the previously described and published protocol.^{35,36} Cytoplasmatic Melan A stain was identified with the Red detection kit (red cytoplasmatic stain), whereas nuclear PRAME stain with the 3,3'-diaminobenzidine detection kit (brown nuclear stain).^{35,36} The double stain Melan A/PRAME was here adopted as it allows to better score PRAME in melanocytic cells and differentiate them from keratinocytes and the background of inflammatory cells.^{35,36} In MM-H&N with <76% of PRAME-positive cells (negative according to Lezcan

et al¹⁶) an additional section was stained with single stain for PRAME to evaluate potential discrepancies with Melan A/PRAME.^{35,36} To identify the cutoff of PRAME-positive cells more suitable to discriminate MM-H&N and MBML-H&N, 3 different scores utilized in routine practice and/or proposed in the recent literature were tested: (1) score according to Lezcano et al¹⁶: 0 = no expression, 1+ = 1% to 25%, 2+ = 26% to 50%, 3+ = 51% to 75%, 4+ = \geq 76%, with cases dichotomized in negative (0, 1+, 2+, 3+) and positive (4+); (2) score according to Raghavan et al³⁰: 0 = no expression—59%, 1+ = \geq 60%, with cases dichotomized in negative (0) and positive (1+); (3) score according to Santandrea et al³⁴: adding the quartile of positive tumor cells (0, 1+, 2+, 3+, 4+ sec. Lezcano et al.) to PRAME expression intensity in tumor cells (0 [no expression], 1+ [weak], 2+ [moderate], 3+ [strong]), with cases dichotomized in negative (< 5) and positive (\geq 5).

Statistical Analysis

For each score sensitivity (SE), specificity (SP), positive predictive value (PPV), negative predictive value (NPV), and accuracy (AC) were evaluated. The score with the highest values of these parameters has been chosen to dichotomize the cases (low and high expression of PRAME) and analyze the association with the other dichotomous/categorical clinicopathologic features using the χ^2 test. The statistical tests were performed using the IBM SPSS software, with a *P*-value <0.05 (2-sided) indicating statistical significance.

Ethical Approval

The study has been approved by the Review Board of the Area Vasta Emilia Centro-AVEC (protocol n. 03-2022-OSS-AUSLBO).

RESULTS

Case Series (Patients and Specimens)

A total of 54 histologic samples from 41 patients were collected. Samples consisted of 37 (68.5%) excisions of the primary tumor, 10 (18.5%) excisions of residual tumor/relapse, and 7 (8.5%) incisional biopsies. Twenty-five (61%) patients were females and 16 (39%) were males; the age at diagnosis ranged from 10 to 96 years (median value: 60 y). Clinicopathologic features of the case series are summarized in Tables 1 and 2. A graphical representation of MM-H&N and MBML-H&N sites is provided in Figure 1.

MBML-H&N (Clinicopathologic Features and PRAME Expression)

Eighteen histologic samples from 18 patients were collected: 13 (72.2%) excisions of the primary lesion and 5 (27.8%) incisional biopsies. Eleven (61.1%) patients were females and 7 (38.9%) males; the age at diagnosis ranged from 10 to 79 years (median value: 47.4 y). The most represented sites were gum (8, 44.4%) and palate (6, 33.3%). The cases were diagnosed as follows: 10 (55.6%) melanotic macules, 6 (33.3%) blue nevi, and 2 (11.1%)

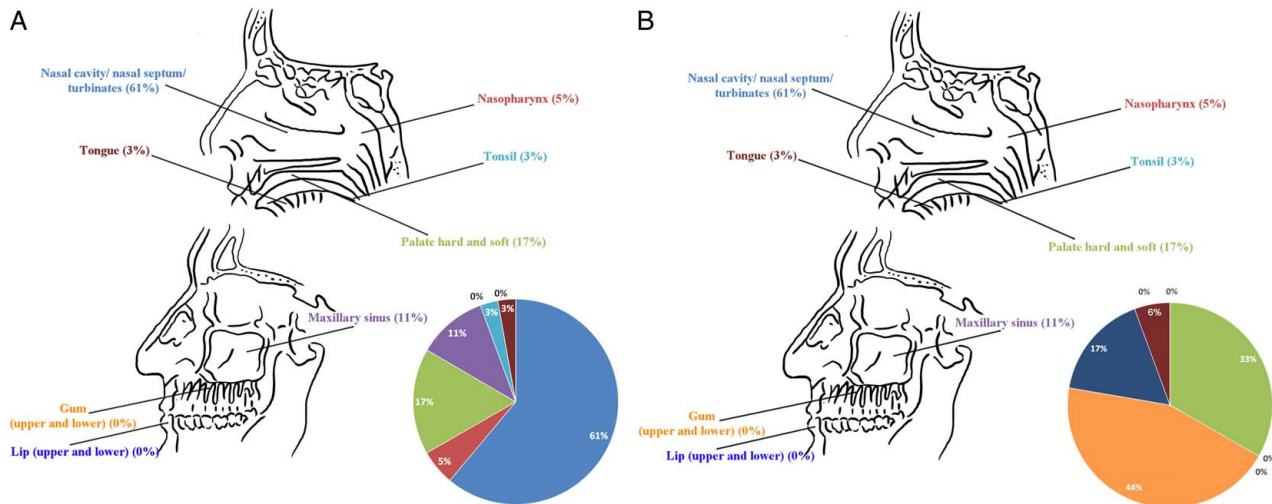


FIGURE 1. Graphical representation of MM-H&N (A) and MBML-H&N (B) in our case series.

common nevi. All MBML-H&N resulted negative for nuclear PRAME expression according to the 3 tested scores. Thirteen (72.2%) cases were completely negative, 5 (27.8%) showed focal and weak immunoreactivity in a low percentage of cells (5% to 15%) with a random distribution and no intralésional intensity variation (defined as at least 2 adjacent high-power fields with >75% stain in a tumor with an overall uniform stain <75%) (Fig. 2).²⁷

MM-H&N (Clinicopathologic Features and PRAME Expression)

Thirty-six histologic samples from 23 patients were collected: 24 (66.72%) excisions of the primary tumor, 10 (27.8%) excisions of residual tumor/relapse, and 2 (5.5%) incisional biopsies, both followed by the surgical excision of the primary tumor. Fourteen (60.9%) patients were females and 9 (39.1%) males; the age at diagnosis ranged from 29 to 96 years (median value: 69.9 y). The most represented sites were the nasal cavity/nasal septum/turbinates (22, 40.7%) and palate (6, 16.7%). The most frequent histologic subtype was the nodular one (14, 60.9%), with a high percentage of cases showing ulceration (18, 78.3%) and prevalent epithelioid cytology (14, 60.9%). According to the eighth edition of the AJCC Cancer Staging Manual, the pT stages included 18 (78.3%) pT3, 4 (17.4%) pT4a, and 1 (4.3%) pT4b. MM-H&N were characterized by a diffuse ($\geq 60\%$ of neoplastic cells) nuclear expression of PRAME in 28 (77.8%) cases, and with a moderate/intense expression (intensity 2+ and 3+) in 20 (55.6%) (Figs. 3, 4). A single stain for PRAME was additionally performed in all cases of MM-H&N with <76% of PRAME-positive cells (negative according to Lezcano et al¹⁶) and the results were superimposable to those observed with double stain for Melan A/PRAME.³⁵ The two incisional biopsies of MM-H&N were both positive only adopting the score of Raghavan et al³⁰ (PRAME-positive cells: 70% and 65%, respectively). Noteworthy, both patients showed higher expression of PRAME (percentage of positive cells and/or intensity) in

the subsequent surgical excisions resulting positive with all 3 tested scores.^{16,30,34} Two cases of MM-H&N were completely negative (0% of positive cells) for PRAME (Fig. 5), and both were located in the palate. In one of them, the negativity was probably related to the quality of the histologic sample (extensive necrosis and poor fixation).

Comparison of PRAME Scores for the Diagnosis of MBML-H&N and MM-H&N

All the tested scores^{16,30,34} showed SP and PPV of 100%, since no MBML-H&N turned out positive with any score. The score of Raghavan et al³⁰ exhibited the highest values of SE, NPV, and AC: 77.8%, 69.2%, and 81.5%, respectively. The values of PRAME (percentage of positive cells and intensity) are summarized in Supplemental Digital Content 1 (<http://links.lww.com/PAS/B522>). The overview of PRAME results in MM-H&N and MBML-H&N, and the comparison between the 3 different scores are provided in Table 3; a graphical representation of MM-H&N and MBML-H&N cases distribution according to PRAME stain is shown in Figure 6.

Association Between PRAME Expression and Clinicopathologic Features

Based on the obtained results, the cutoff of 60% of PRAME-positive cells (nuclear stain) was chosen to dichotomize MM-H&N cases in high ($\geq 60\%$) and low (<60%) expression, and analyze the association of PRAME expression with the other dichotomous/categorical clinicopathologic features. Accordingly, high expression of PRAME was associated with female sex (21/23 [91.3%] vs. 7/13 [53.8%] in male sex, $P=0.016$) and nodular histologic subtype (19/20 [95%] vs. 9/16 [56.8%] in mucosal lentiginous one, $P=0.005$). Notably, although the mucosal lentiginous histotype showed lower rates of positivity, PRAME highlighted the intra-epithelial component/growth, so being potentially useful for the appropriate evaluation of the mucosal resection

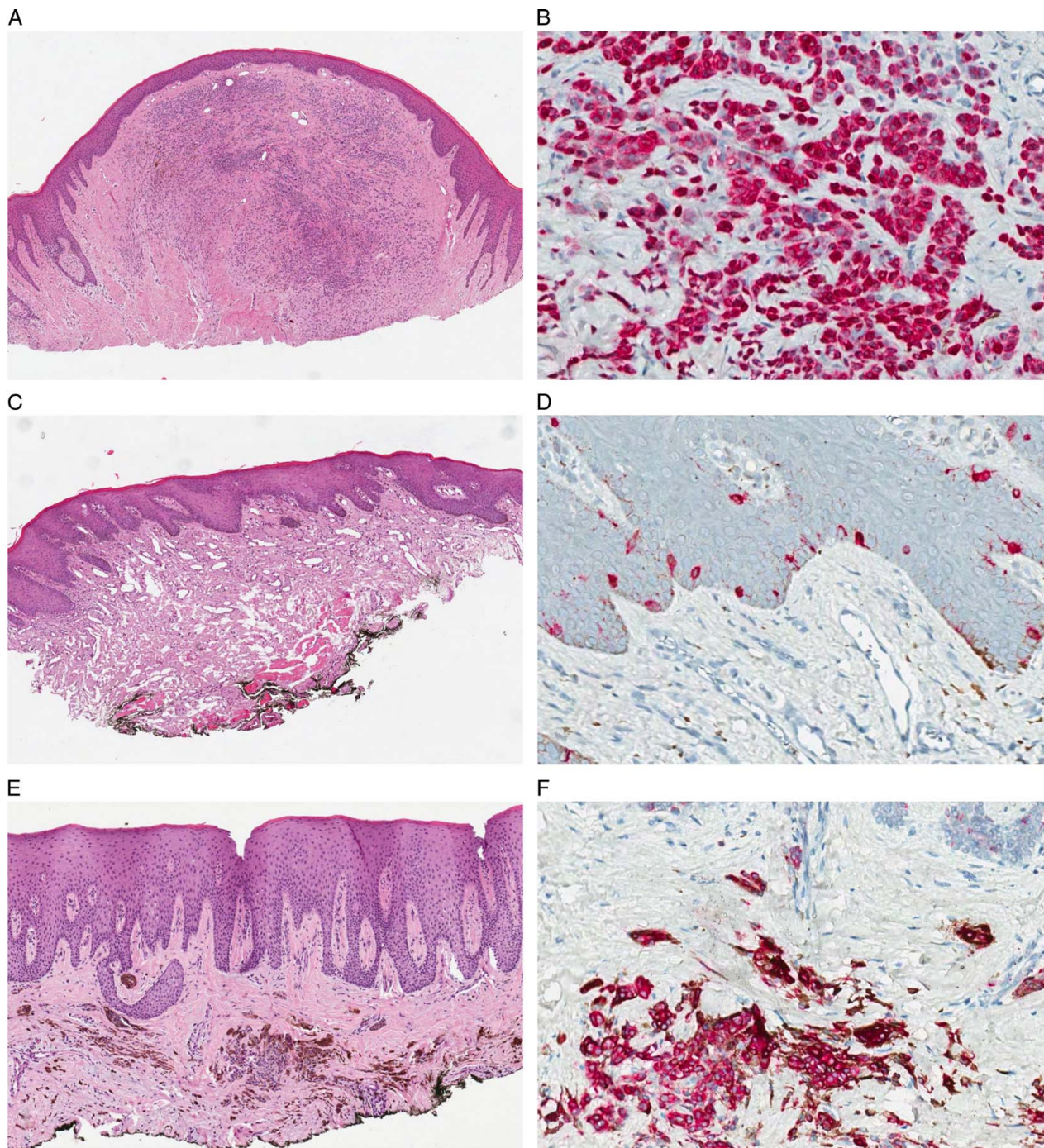


FIGURE 2. MBML-H&N: mucosal benign melanocytic lesions of the head and neck region; PRAME expression in MBML-H&N. Three cases of histologically straightforward MBML-H&N: common nevus (A: hematoxylin and eosin), melanotic macula (C: hematoxylin and eosin), and blue nevus (E: hematoxylin and eosin), respectively. Immunohistochemistry for Melan A/PRAME shows no or focal PRAME expression in the nuclei of melanocytes (B, D, F: PRAME).

margins (Fig. 3). High expression of PRAME was also significantly associated with specific anatomic sites ($P < 0.001$), as follows: MM-H&N of the nasal cavity/nasal septum/turbinates showed high expression of PRAME in 20/22 (90.9%) cases, whereas MM-H&N of the palate in 0/6 (0%). Albeit affected by much fewer

cases, all the cases of the nasopharynx (2/2, 100%) and the maxillary sinus (4/4, 100%) exhibited high expression of PRAME. No statistically significant association was found between high expression of PRAME and the other clinicopathologic features (type of histologic specimen, pigmentation, prevalent cytotype, ulceration,

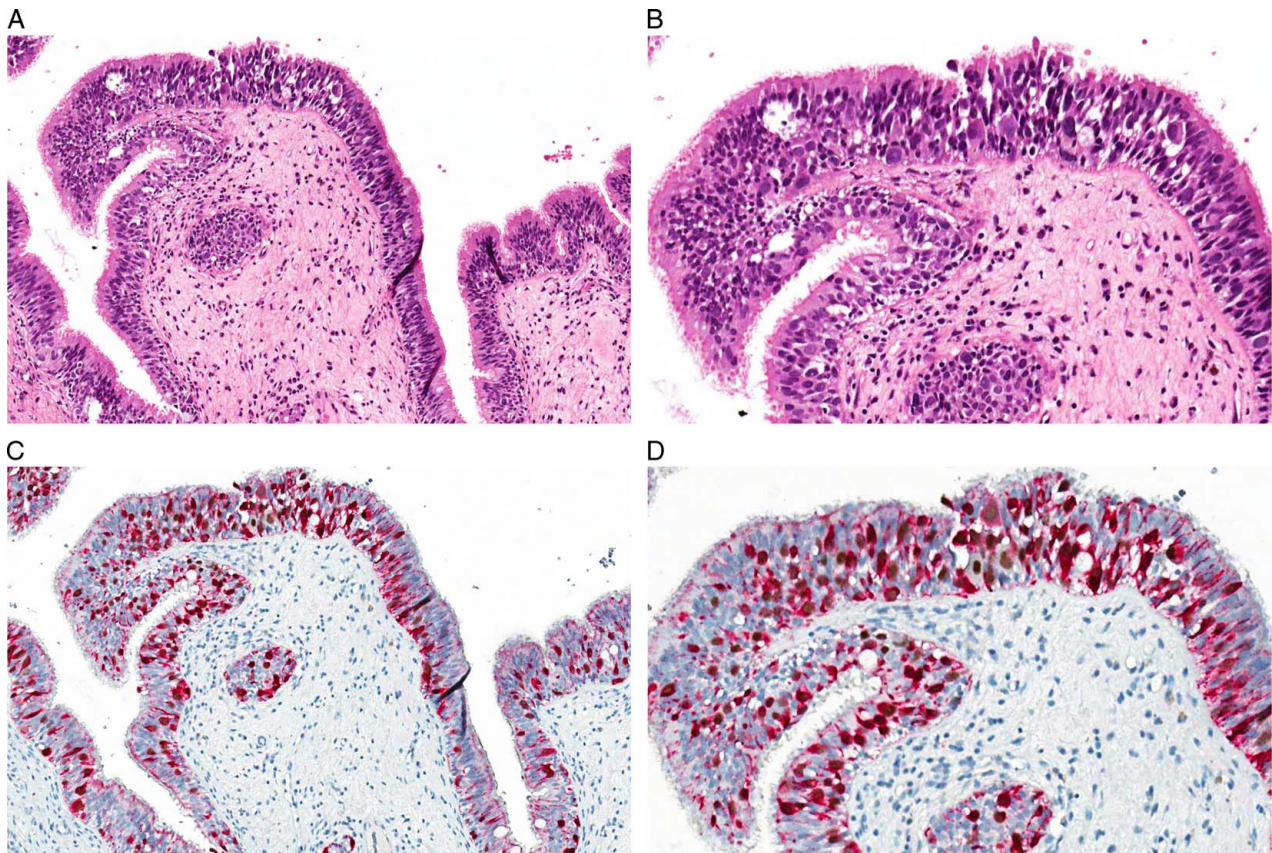


FIGURE 3. PRAME expression in a case of mucosal lentiginous MM-H&N. The histologic examination shows the intraepithelial component (A, B: hematoxylin and eosin) nicely highlighted by nuclear PRAME stain (brown) (C: hematoxylin and eosin; D: PRAME), and so being potentially useful for the assessment of mucosal resection margins.

bone and/or cartilage infiltration, lymphovascular invasion, perineural infiltration, and pT stage) (Table 4).

DISCUSSION

After the introduction of immunohistochemistry for PRAME in the diagnosis of melanocytic lesions, many researchers focused on its application in several diagnostic scenarios (Spitz lesions, lentigo maligna, acral lesions, etc.) with encouraging but also partially discordant results.^{16,24-41} First, it is questionable whether PRAME stain could be interchangeable and therefore completely replace biomolecular tests for the assessment of “difficult-to-classify tumors,” even if it showed high correlation (about 90%) with cytogenetic tests (FISH and single-nucleotide polymorphism array) in case series enrolling a large spectrum of challenging melanocytic tumors.²⁶⁻³⁰ Furthermore, despite the most diffuse and applied immunohistochemical score of PRAME is that proposed by Lezcano and colleagues, several authors found that alternative scores (different cutoffs and/or a combination of a qualitative and quantitative assessment) are more performing for the assessment of specific histotypes of melanocytic lesions.^{16,24-42} As a result, at the state of the art, it is not known whether the PRAME score should be one in

all cases, or if different scores depending on the analyzed tumor are more appropriate.^{16,24-42} To the best of our knowledge, this is the first study testing PRAME and comparing different scores for the appropriate diagnosis of MM-H&N and MBML-H&N. A heterogeneous case series of MBML-H&N, including nodular and mucosal lentiginous mucosal melanoma, different histologic subtypes of MBML-H&N (melanotic macula, blue nevus, and common nevus), and covering a large spectrum of sites (nasal cavity/nasal septum/turbinates, palate, nasopharynx, etc.) constituted the basis of the present study. The most relevant data emerging is that all MBML-H&N are negative for PRAME with all 3 tested scores.^{16,30,34} Namely, the majority of MBML-H&N showed no PRAME immunostaining, whereas a minority of cases showed rare positive cells only, scattered through the lesion without intralesional intensity variation and hotspots.²⁷ As a result, SP and PPV were 100% (0 false-positive cases) with all 3 tested scores.^{16,30,34} This result can be relevant in a diagnostic scenario where each false-positive case risks being subjected to aggressive therapies impacting on the quality of life.¹⁻⁶ According to the data here shown, the most performing score for differentiating MM-H&N from MBML-H&N (SE: 77.8%; NPV: 69.2%; AC: 81.5%) was that proposed by Raghavan et al.³⁰ They analyzed a

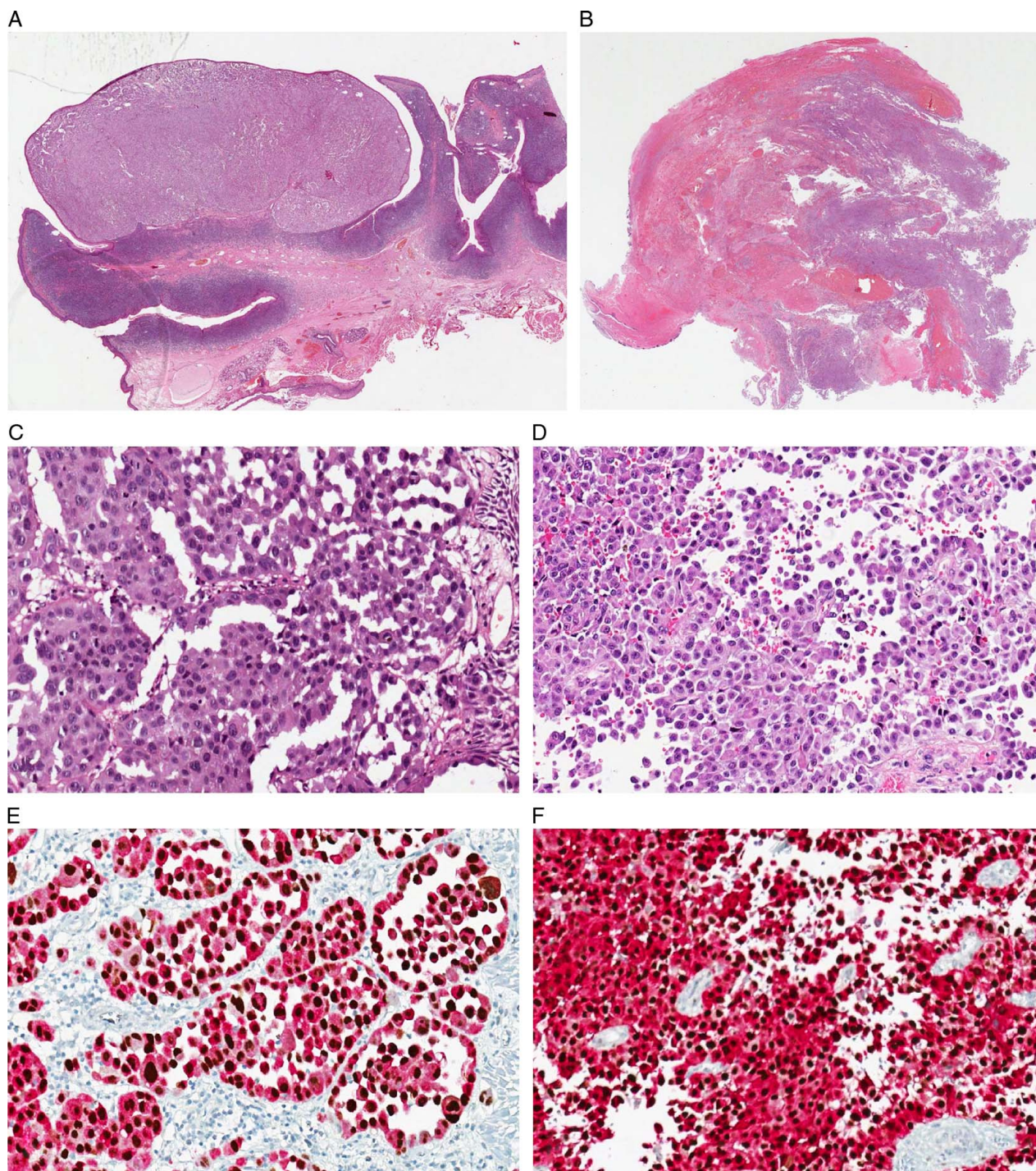


FIGURE 4. Two cases of nodular MM-H&N. Large and polypoid lesion (A: hematoxylin and eosin), with a nodular growth of atypical melanocytes (C: hematoxylin and eosin) diffusely positive for PRAME (brown nuclei) and Melan A (red cytoplasm) (E: PRAME). Highly fragmented sample of an ulcerated MM-H&N (B: hematoxylin and eosin) with a diffuse growth of atypical melanocytes (D: hematoxylin and eosin) diffusely positive for PRAME (brown nuclei) and Melan A (red cytoplasm) (F: PRAME).

heterogenous case series of melanocytic tumors with intermediate histopathologic or spitzoid features, and found that the best threshold to differentiate benign from malignant tumors was 60% of PRAME-positive cells.³⁰

According to these authors, the loss of SE obtained with the other scores would impair the utility of PRAME.^{16,30,34} This finding highlights as scores alternative to that of Lezcano and colleagues may be more

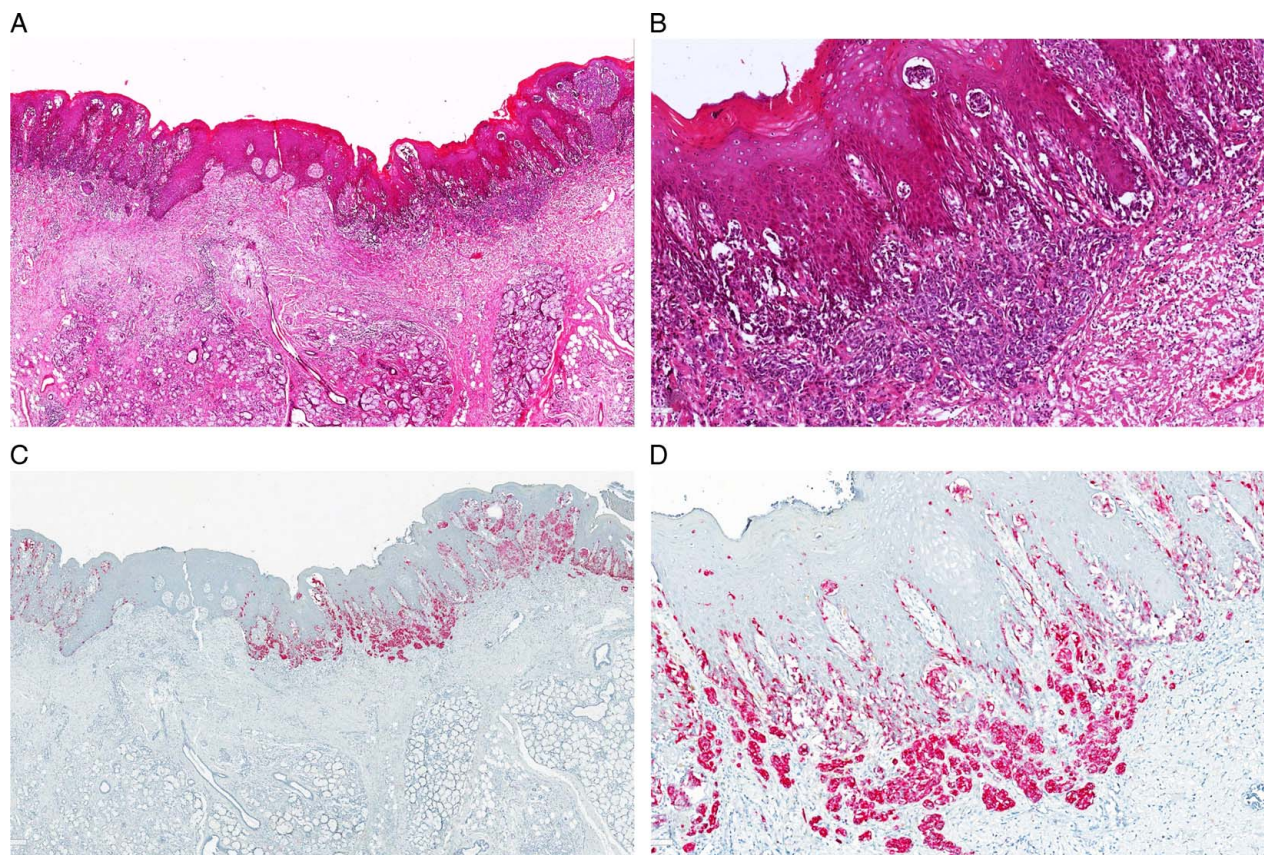


FIGURE 5. MBML-H&N: mucosal benign melanocytic lesions of the head and neck region; Negative PRAME expression in a case of mucosal lentiginous MM-H&N of the palate. The histologic examination shows a contiguous proliferation of atypical non-nested junctional melanocytes and irregular/dyscohesive nests at the dermoepidermal junction and in the superficial portion of the lamina propria, with marked epidermal effacement and diffuse pagetoid spread (A, C: hematoxylin and eosin). Double stain for Melan A/PRAME shows as the entire melanocytic population is PRAME-negative, with scattered positive fibroblasts as a positive control (B, D: hematoxylin and eosin).

effective for specific categories of melanocytic lesions, as previously found in other scenarios (metastatic melanoma, nodal nevi vs. nodal metastatic melanoma, nevus-associated melanoma, conjunctival, acral, and nail unit melanocytic lesions).^{31–39} Additional aspects emerging from the present data, that should be analyzed before the adoption of PRAME for the routine diagnosis of lesions are: (1) different PRAME results according to the site; (2) PRAME results in the incisional biopsies. The data here shown highlight that PRAME must always be analyzed according to the topography of the melanocytic tumor and that a negative stain should be carefully interpreted in a palatal lesion morphologically suggestive for MM-H&N. In the present case series, the MM-H&N incisional biopsies were two only, but both positive only adopting the cutoff of Raghavan et al³⁰ (Supplemental Digital Content 1, <http://links.lww.com/PAS/B522>). By contrast, the subsequent surgical excisions of both patients turned out positive with all 3 scores. This finding suggests that, due to the well-known heterogeneity of PRAME expression and the little amount of tissue often obtained with incisional biopsies, PRAME could be more appropriately scored in

the incisional biopsies by adopting a lower threshold of PRAME-positive cells. Notably, all 5 incisional biopsies of MBML-H&N resulted negative with all 3 tested scores. Based on the present findings, it is proposed that positive PRAME stain strongly encourages a diagnosis of MM-H&N, since we did not find MBML-H&N positive for PRAME. By contrast, if the histopathologic features are strongly suggestive for MM-H&N but PRAME stain is negative, as it happens in a not negligible number of MM-H&N and especially in specific sites (palate), the possibility of malignancy should not be discarded. Finally, if the histopathologic features are supportive for MBML-H&N, PRAME stain is expected to be negative and a positive result should lead to reconsidering the possibility of malignancy. An additional benefit of PRAME could be the assessment of the mucosal resection margins, especially in cases of lentiginous MM-H&N. This latter, although with a lower positivity (9/16, 56.8%) compared with the nodular one (19/20, 95%), exhibited an intraepithelial component nicely depicted by Melan A/PRAME (Fig. 3). This scenario mirrors what found by Gradecki et al³² in skin lentigo maligna, where the authors

TABLE 3. The Overview of PRAME Results and the Comparison Between the 3 Different Scores for the Diagnosis of MBML-H&N and MM-H&N

	MBML-H&N, n/N (%)			All	MM-H&N, n/N (%)		All	
	Common nevus	Blue nevus	Melanotic macula		Mucosal lentiginous	Nodular		
Lezcano et al ¹⁶ 0, 1+, 2+, 3+	2/2 (100)	6/6 (100)	10/10 (100)	18/18 (100)	8/16 (50)	3/20 (15)	11/36 (30.6)	SE: 69.4% SP: 100% PPV: 100% NPV: 62% AC: 79.6%
4+	0 (0)	0 (0)	0 (0)	0 (0)	8/16 (50)	17/20 (85)	25/36 (69.4)	
Raghavan et al ³⁰ 0	2/2 (100)	6/6 (100)	10/10 (100)	18/18 (100)	7/16 (43.7)	1/20 (5)	8/36 (22.2)	SE: 77.8% SP: 100% PPV: 100% NPV: 69.2% AC: 85.2%
1+	0 (0)	0 (0)	0 (0)	0 (0)	9/16 (56.3)	19/20 (95)	28/36 (77.8)	
Santandrea et al ³⁴ <5	2/2 (100)	6/6 (100)	10/10 (100)	18/18 (100)	8/16 (50)	2/20 (10)	10/36 (27.8)	SE: 72.2% SP: 100% PPV: 100% NPV: 64.3% AC: 81.5%
≥5	0 (0)	0 (0)	0 (0)	0 (0)	8/16 (50)	18/20 (90)	26/36 (72.2)	

MM-H&N: mucosal melanoma of the head and neck region; MBML-H&N: mucosal benign melanocytic lesions of the head and neck region.

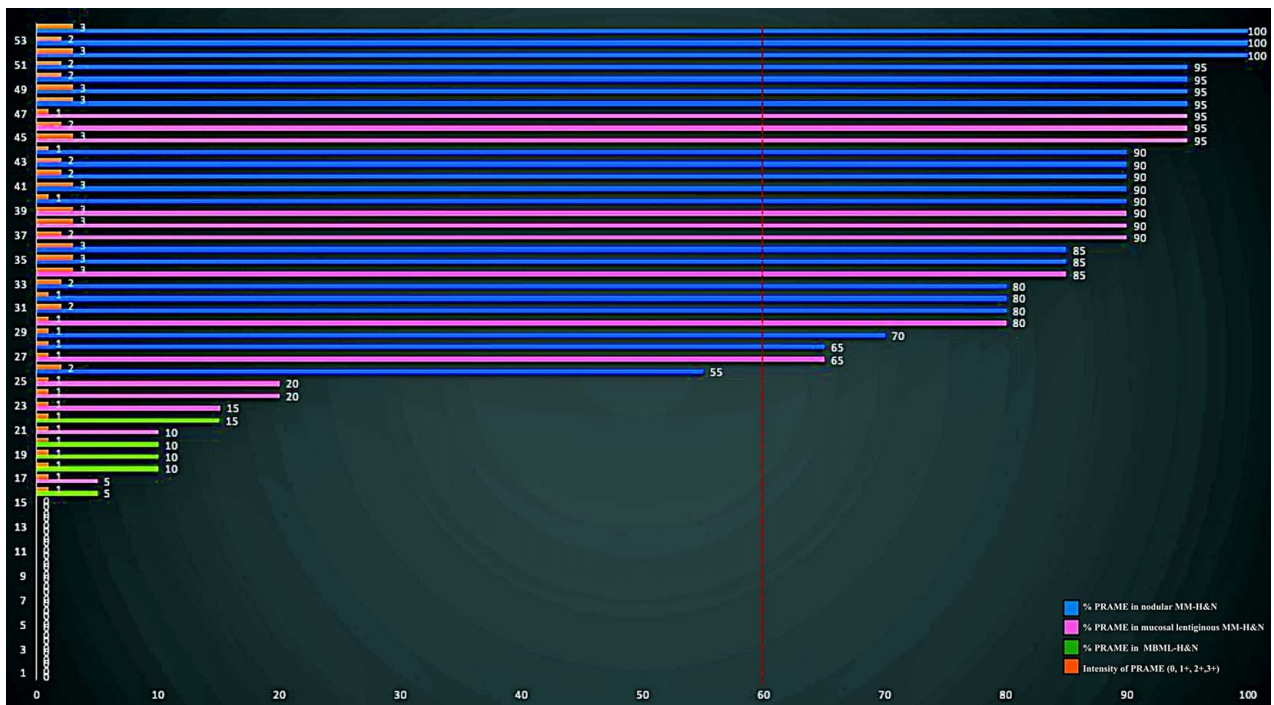


FIGURE 6. Graphical representation of PRAME expression (% of positive cells and intensity) in MBML-H&N and MM-H&N. Each case corresponds to 2 bars, one for the % of PRAME-positive cells (green: MBML-H&N, pink: mucosal lentiginous MM-H&N, blue: nodular MM-H&N) and one for the intensity (orange bars). Fifteen cases showed 0% of PRAME-positive cells (13 MBML-H&N and 2 mucosal lentiginous MM-H&N; see the Results section and Fig. 5 for more explanations). Notably, the majority of MM-H&N below the cutoff of 60% of PRAME-positive cells (red dotted line) are mucosal lentiginous (only 1 case was nodular); all MBML-H&N show % of PRAME-positive cells below this threshold.

TABLE 4. The Association Between Clinicopathologic Features and PRAME Expression (Low: <60%, High \geq 60) in MM-H&N

	MM-H&N, samples (N = 36), n (%)		P
	Low expression of PRAME (<60)	High expression of PRAME (\geq 60)	
Type of histologic specimen			
Excision of the primary tumor	7 (19.4)	17 (47.2)	0.349
Excision of residual tumor/relapse	1 (2.8)	9 (25)	
Incisional biopsy	0 (0)	2 (5.6)	
Sex			
Male	6 (16.7)	7 (19.4)	0.009
Female	2 (5.6)	21 (58.3)	
Site			
Nasal cavity/nasal septum/turbinates	2 (5.6)	20 (55.6)	<0.001
Nasopharynx	0 (0)	2 (5.6)	
Palate (hard and soft)	6 (16.7)	0 (0)	
Maxillary sinus	0 (0)	4 (14.3)	
Tonsil	0 (0)	1 (2.8)	
Tongue	0 (0)	1 (2.8)	
Histologic subtype			
Mucosal lentiginous	7 (19.4)	9 (25)	0.005
Nodular	1 (2.8)	19 (52.8)	
Pigmentation			
No	2 (5.6)	7 (19.4)	1.000
Yes	6 (16.7)	21 (58.3)	
Prevalent cytotype			
Epithelioid	4 (14.3)	16 (44.4)	0.890
Fused	3 (8.3)	8 (22.2)	
Mixed	1 (2.8)	4 (14.3)	
Ulceration			
No	2 (5.6)	5 (13.9)	0.497
Yes	6 (16.7)	23 (63.9)	
Bone and/or cartilage infiltration			
No	0 (0)	7 (19.4)	0.115
Yes	8 (22.2)	21 (58.3)	
LVI			
No	5 (13.9)	18 (50)	0.926
Yes	3 (8.3)	10 (27.8)	
PNI			
No	7 (19.4)	27 (75)	0.331
Yes	1 (2.8)	1 (2.8)	
pT stage			
pT3	5 (13.9)	24 (66.7)	0.184
pT4a	3 (8.3)	3 (8.3)	
pT4b	0 (0)	1 (2.8)	

LVI indicates lymphovascular invasion; PNI, perineural infiltration.

suggested that the double stain Melan A/PRAME is helpful. Finally, the present findings indicate that high PRAME expression (\geq 60%) is significantly associated with specific sites, nodular histotype, and female sex (Table 4). Although the involved pathogenetic mechanisms for justifying these associations have been poorly investigated and are beyond the scope of this article, they should be kept in mind to avoid misdiagnoses. High nuclear PRAME expression (\geq 60%) was preferentially detected in MM-H&N of the nasal cavity/nasal septum/turbinates (20/22, 90.9%), nasopharynx (2/2,

100%) and maxillary sinus (4/4, 100%) rather than palate (0/6, 0%). Scheurleer et al⁴² found PRAME expression in all tested cases (23/23, 100%) of MM-H&N of the sinonasal region, but they did not test MM-H&N of other sites (palate, nasopharynx, etc.). In the study of Hovander et al,⁴¹ 6/7 (86%) MM-H&N of the oral cavity/palate resulted positive for PRAME; nevertheless, the authors did not report the criteria used for PRAME assessment (score and cutoff) and so it is not possible to compare their results with the present ones. Toyama et al⁴⁰ did not find statistically significant associations between PRAME expression and tumor site in mucosal melanomas. However, Toyama et al's⁴⁰ study differs from the present one for the tested case series (sinonasal vs. nonsinonasal sites), PRAME assessment (*H*-score), and adopted statistical test (comparison of the mean values with a *t* test calculator). A large amount of evidences emerging from recent literature data suggests that, although MM-H&N are grouped as such unique subgroup for classification purposes, they should be more appropriately considered as a heterogenous family of tumors, with marked differences from clinical, histologic, immunohistochemical, molecular, and pathogenetic sides.^{1-6,11,14,40} According with this theory and with a classification of melanomas progressively based on their molecular profile, MM-H&N of different sites exhibit divergent mutational landscapes and should be probably designed and classified as different lesions.^{14,15} Öztürk Sari et al¹⁵ found that 91% of sinonasal but only 9% of oral MM-H&N harbored mutations in the tested genes (*BRAF*, *NRAS*, *KIT*, *TERT*, *GNAQ/GNA11*); besides, *NRAS* and *TERT* promoter mutation were significantly higher in sinonasal than in oral location. More recently, Chłopek et al¹⁴ analyzed a large case series of sinonasal melanomas and found molecular divergences between paranasal sinuses (10/14 [71%] *BRAF/RAS* mutants) and nasal (26/64 [41%] *BRAF/RAS* mutants) cases. The present results support the existence of differences among MM-H&N arising in different mucosal sites and suggest that PRAME, as a pivot molecule in the biology of melanoma, could be differently involved and expressed in distinct sites.¹⁴⁻¹⁷ The high expression of PRAME primarily found in nodular (19/20, 95%) rather than mucosal lentiginous histotype (9/16, 56.8%) reflects what previously found in the skin for other subtypes of melanocytic lesions (acral melanocytic lesions, spitzoid lesions, lentigo maligna, nevus-associated melanomas, etc.).^{16,26-42} It could be explained by assuming that histologic differences reflects clinical, immunohistochemical, but primarily molecular differences, thus potentially justifying a selective involvement of PRAME in their pathogenesis.^{1-6,11,14,16,26,43} The higher PRAME expression in the female sex (21/23, 91.3%) rather than in male one (7/13, 53.8%) is more difficult to justify. It is possible that the peculiar role of sex hormones (androgen and estrogen) in the development of melanomas, as well as the complex intracellular mechanisms regulated by these 2 molecules, could explain the different PRAME expression observed in females and males.^{45,46} To conclude, the data here obtained indicate that PRAME is a useful tool for the

appropriate diagnosis of MBML-H&N and MM-H&N. The most reliable score to differentiate MM-H&N and MBML-H&N is that proposed by Raghavan and colleagues ($< 60\%$ vs. $\geq 60\%$ of PRAME-positive cells), but a subgroup of specifically sited MM-H&N (palate) can be PRAME-negative (although this assumption is based on a low number of cases and future studies are needed to verify this finding). High PRAME expression ($\geq 60\%$) was found in association with specific mucosal sites, nodular histotype, and female sex, suggesting a distinct involvement of this molecule in the pathogenesis of these subgroups of tumors.

REFERENCES

- Rojas-Lechuga MJ, Gras-Cabrerizo JR, Aviles-Jurado FX, et al. Sinonasal mucosal melanomas: defining profiles for better survival outcomes. *Rhinology*. 2022;1:347–356.
- López F, Rodrigo JP, Cardesa A, et al. Update on primary head and neck mucosal melanoma. *Head Neck*. 2016;38:147–155.
- Ascierto PA, Accorona R, Botti G, et al. Mucosal melanoma of the head and neck. *Crit Rev Oncol Hematol*. 2017;112:136–152.
- Tacastacas JD, Bray J, Cohen YK, et al. Update on primary mucosal melanoma. *J Am Acad Dermatol*. 2014;71:366–375.
- Hernandez-Prera JC. Update from the 5th Edition of the World Health Organization Classification of Head and Neck Tumors: The Neck and Lymph Nodes, Metastasis, and Melanocytic Tumors. *Head Neck Pathol*. 2022;16:110–122.
- Dika E, Lambertini M, Pellegrini C, et al. Cutaneous and mucosal melanomas of uncommon sites: where do we stand now? *J Clin Med*. 2021;10:478.
- Gerami P, Li G, Pouryazdanparast P, et al. A highly specific and discriminatory FISH assay for distinguishing between benign and malignant melanocytic neoplasms. *Am J Surg Pathol*. 2012;36:808–817.
- Bastian BC, Olshen AB, LeBoit PE, et al. Classifying melanocytic tumors based on DNA copy number changes. *Am J Pathol*. 2003;163:1765–1770.
- Benton S, Zhao J, Zhang B, et al. Impact of next-generation sequencing on interobserver agreement and diagnosis of spitzoid neoplasms. *Am J Surg Pathol*. 2021;45:1597–1605.
- Lee CY, Gerami P. Molecular techniques for predicting behaviour in melanocytic neoplasms. *Pathology*. 2016;48:142–146.
- Tacastacas JD, Bray J, Cohen YK, et al. Update on primary mucosal melanoma. *J Am Acad Dermatol*. 2014;71:366–375.
- Ma Y, Xia R, Ma X, et al. Mucosal melanoma: pathological evolution, pathway dependency and targeted therapy. *Front Oncol*. 2021;11:702287.
- Zebary A, Jangard M, Omholt K, et al. KIT, NRAS and BRAF mutations in sinonasal mucosal melanoma: a study of 56 cases. *Br J Cancer*. 2013;109:559–564.
- Chlopek M, Lasota J, Thompson LDR, et al. Alterations in key signaling pathways in sinonasal tract melanoma. A molecular genetics and immunohistochemical study of 90 cases and comprehensive review of the literature. *Mod Pathol*. 2022;35:1609–1617.
- Öztürk Sari Ş, Yılmaz İ, Taşkın OÇ, et al. BRAF, NRAS, KIT, TERT, GNAQ/GNA11 mutation profile analysis of head and neck mucosal melanomas: a study of 42 cases. *Pathology*. 2017;49:55–61.
- Lezcano C, Jungbluth AA, Nehal KS, et al. PRAME expression in melanocytic tumors. *Am J Surg Pathol*. 2018;42:1456–1465.
- Ikeda H, Lethé B, Lehmann F, et al. Characterization of an antigen that is recognized on a melanoma showing partial HLA loss by CTL expressing an NK inhibitory receptor. *Immunity*. 1997;6:199–208.
- Ricci C, Franceschini T, Giunchi F, et al. Immunohistochemical expression of preferentially expressed antigen in melanoma (PRAME) in the uninvolved background testis, germ cell neoplasia in situ, and germ cell tumors of the testis. *Am J Clin Pathol*. 2022;157:644–648.
- Orsatti A, Sirolli M, Ambrosi F, et al. SOX2 and PRAME in the “reprogramming” of seminoma cells. *Pathol Res Pract*. 2022;237:154044.
- Zhang W, Barger CJ, Eng KH, et al. PRAME expression and promoter hypomethylation in epithelial ovarian cancer. *Oncotarget*. 2016;7:45352–45369.
- Roszik J, Wang WL, Livingston JA, et al. Overexpressed PRAME is a potential immunotherapy target in sarcoma subtypes. *Clin Sarcoma Res*. 2017;7:11.
- Matsushita M, Yamazaki R, Ikeda H, et al. Preferentially expressed antigen of melanoma (PRAME) in the development of diagnostic and therapeutic methods for hematological malignancies. *Leuk Lymphoma*. 2003;44:439–444.
- Goodison S, Urquidi V. The cancer testis antigen PRAME as a biomarker for solid tumor cancer management. *Biomark Med*. 2012;6:629–632.
- Kaczorowski M, Chlopek M, Kruczak A, et al. PRAME expression in cancer. a systematic immunohistochemical study of > 5800 epithelial and nonepithelial tumors. *Am J Surg Pathol*. 2022;46:1467–1476.
- LeBlanc RE, Miller DM, Zegans ME, et al. PRAME immunohistochemistry is useful in the evaluation of conjunctival melanomas, nevi, and primary acquired melanosis. *J Cutan Pathol*. 2021;48:1442–1448.
- Lezcano C, Jungbluth AA, Busam KJ. PRAME immunohistochemistry as an ancillary test for the assessment of melanocytic lesions. *Surg Pathol Clin*. 2021;14:165–175.
- Alomari AK, Tharp AW, Umphress B, et al. The utility of PRAME immunohistochemistry in the evaluation of challenging melanocytic tumors. *J Cutan Pathol*. 2021;48:1115–1123.
- Lezcano C, Jungbluth AA, Busam KJ. Comparison of immunohistochemistry for PRAME with cytogenetic test results in the evaluation of challenging melanocytic tumors. *Am J Surg Pathol*. 2020;44:893–900.
- Lezcano C, Pulitzer M, Moy AP, et al. Immunohistochemistry for PRAME in the distinction of nodal nevi from metastatic melanoma. *Am J Surg Pathol*. 2020;44:503–508.
- Raghavan SS, Wang JY, Kwok S, et al. PRAME expression in melanocytic proliferations with intermediate histopathologic or spitzoid features. *J Cutan Pathol*. 2020;47:1123–1131.
- Gradecki SE, Slingluff CL Jr, Gru AA. PRAME expression in 155 cases of metastatic melanoma. *J Cutan Pathol*. 2021;48:479–485.
- Gradecki SE, Valdes-Rodriguez R, Wick MR, et al. PRAME immunohistochemistry as an adjunct for diagnosis and histological margin assessment in lentigo maligna. *Histopathology*. 2021;78:1000–1008.
- McBride JD, McAfee JL, Piliang M, et al. Preferentially expressed antigen in melanoma and p16 expression in acral melanocytic neoplasms. *J Cutan Pathol*. 2022;49:220–230.
- Santandrea G, Valli R, Zanetti E, et al. Comparative analysis of PRAME expression in 127 acral and nail melanocytic lesions. *Am J Surg Pathol*. 2022;46:579–590.
- Grillini M, Ricci C, Pino V, et al. HMB45/PRAME, a novel double staining for the diagnosis of melanocytic neoplasms: technical aspects, results, and comparison with other commercially available staining (PRAME and Melan A/PRAME). *Appl Immunohistochem Mol Morphol*. 2022;30:14–18.
- Ricci C, Dika E, Ambrosi F, et al. Cutaneous melanomas: a single center experience on the usage of immunohistochemistry applied for the diagnosis. *Int J Mol Sci*. 2022;23:5911.
- Kim YJ, Jung CJ, Na H, et al. Cyclin D1 and PRAME expression in distinguishing melanoma in situ from benign melanocytic proliferation of the nail unit. *Diagn Pathol*. 2022;17:41.
- Lohman ME, Steen AJ, Grekin RC, et al. The utility of PRAME staining in identifying malignant transformation of melanocytic nevi. *J Cutan Pathol*. 2021;48:856–862.

39. See SHC, Finkelman BS, Yeldandi AV. The diagnostic utility of PRAME and p16 in distinguishing nodal nevi from nodal metastatic melanoma. *Pathol Res Pract*. 2020;216:153105.
40. Toyama A, Siegel L, Nelson AC, et al. Analyses of molecular and histopathologic features and expression of PRAME by immunohistochemistry in mucosal melanomas. *Mod Pathol*. 2019;32:1727–1733.
41. Hovander D, Allen J, Oda D, et al. PRAME immunohistochemistry is useful in the diagnosis of oral malignant melanoma. *Oral Oncol*. 2022;124:105500.
42. Scheurleer WFJ, Braunius WW, Tijink BM, et al. PRAME staining in sinonasal mucosal melanoma: a single-center experience. *Head Neck Pathol*. 2022. doi:10.1007/s12105-022-01515-9.[Epub ahead of print]
43. WHO Classifications of Tumors Editorial Board. *WHO Classification of Tumours Series: Head and Neck Tumours [Internet; beta version ahead of print]*, 5th ed. International Agency for Research on Cancer; 2022.
44. Amin MB, Edge S, Greene F, et al. *AJCC Cancer Staging Manual*, 8th ed. Springer; 2017.
45. Dika E, Lambertini M, Lauriola M, et al. Female melanoma and estrogen receptors expression: an immunohistochemical pilot study. *Melanoma Res*. 2022;32:231–240.
46. Dika E, Patrizi A, Lambertini M, et al. Estrogen receptors and melanoma: a review. *Cells*. 2019;8:1463.