

First-in-Human Studies of Pharmacokinetics and Safety of Utreloxastat (PTC857), a Novel 15-Lipoxygenase Inhibitor for the Treatment of Amyotrophic Lateral Sclerosis

Clinical Pharmacology
in Drug Development
2023, 12(2) 141–151
© 2022 PTC Therapeutics. *Clinical Pharmacology in Drug Development*
published by Wiley Periodicals LLC
on behalf of American College of
Clinical Pharmacology.
DOI: 10.1002/cpdd.1203

Lan Gao, Peter Giannousis, Martin Thoolen, Diksha Kaushik, Joey Latham, Aaron Tansy, Jiyuan Ma, Mayzie Johnston, Mandar Dali, Lee Golden, Matthew Klein, Ronald Kong, and Jeffrey Trimmer

Abstract

Utreloxastat (PTC857) is a 15-lipoxygenase inhibitor being developed to treat amyotrophic lateral sclerosis. This first-in-human study investigated the safety and pharmacokinetics of utreloxastat in healthy volunteers (N = 82) in a double-blind, placebo-controlled trial. The effects of a single ascending dose (100–1000 mg), multiple ascending doses (150–500 mg), and food (500 mg) on the pharmacokinetics and safety of utreloxastat were evaluated. Following single doses, the time to maximum plasma concentration (C_{max}) was observed \approx 4 hours after dosing and the terminal half-life ranged from 20 to 25.3 hours. The C_{max} and area under the concentration-time curve (AUC) increased slightly over dose proportionally. Following multiple doses (once daily/twice daily), the apparent clearance reduced and terminal half-life was \geq 33 hours. There was no apparent difference of exposure following morning or evening doses. Varying diets increased the C_{max} and AUCs of utreloxastat but did not alter time to C_{max} . There were no gender-based differences in exposure. Utreloxastat showed no marked safety signal following single doses up to 1000 mg and multiple doses over 14 days of 500 mg once daily or 250 mg twice daily. The results support further development of utreloxastat for the treatment of patients with amyotrophic lateral sclerosis at a 250-mg twice-daily dose administered with food.

Keywords

15-lipoxygenase inhibitor, ALS, amyotrophic lateral sclerosis, pharmacokinetic, utreloxastat (PTC857)

Amyotrophic lateral sclerosis (ALS) is a fatal neurodegenerative disease that affects the motor neuron system. ALS results in progressive paralysis and eventual death due to respiratory failure,¹ typically within 3–5 years of symptom onset.² The disease is estimated to affect \approx 1–2.6 in 100,000 people per year worldwide.³ Currently, only two approved treatments (riluzole and edaravone) for ALS are available in the United States, and in the European Union only riluzole is available. These therapies are aimed at slowing disease progression but do so with limited efficacy.⁴

Oxidative stress is a central component of ALS and is hypothesized to be responsible for ferroptosis, an iron-dependent oxidative mode of cell death, which contributes to the disease pathology.² Oxidative

stress results in production of reactive oxygen species (ROS). ROS attacks polyunsaturated fatty acids contained in cellular membrane phospholipids, triggering

PTC Therapeutics, Inc., South Plainfield, New Jersey, USA

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

Submitted for publication 8 June 2022; accepted 27 October 2022.

Corresponding Author:

Jeffrey Trimmer, PhD, PTC Therapeutics, Inc., 100 Corporate Court, South Plainfield, NJ 07080
(e-mail: jtrimmer@ptcbio.com)

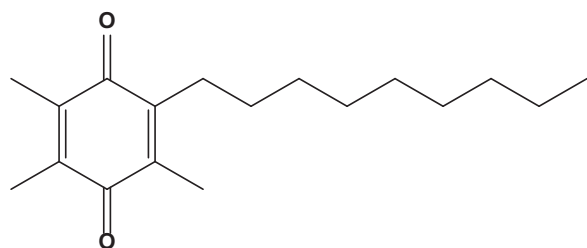


Figure 1. Structure of utreloxastat.

lipid peroxidation, which leads to cell death through ferroptosis.⁵

ROS upregulates 15-lipoxygenase (15-LO), an enzyme that typically uses polyunsaturated fatty acids to facilitate the removal of apoptotic cells and activate inflammatory processes.^{6,7} This upregulation results in the production of lipid peroxide products that increase oxidative stress, activate proinflammatory glial cells, deplete reduced glutathione, accumulate iron, and ultimately cause ferroptosis.^{7,8}

These biochemical events, in particular the production of oxidized lipids, inflammation, oxidative stress, and depletion of reduced glutathione, are known contributors to the neuronal cell death that characterizes ALS.⁹ ROS and the resultant oxidative damage are increased in samples of ALS patients and ALS animal models.^{10–13} The role of 15-LO in ferroptosis-related cell death makes it a promising potential treatment target in ALS.

Utreloxastat (alkyl-substituted cyclohexadiene-1,4-dione) (Figure 1) is an orally bioavailable small molecule being developed for the treatment of neurodegenerative diseases characterized by high levels of oxidative stress and mitochondrial pathology, such as ALS. Pharmacologically, utreloxastat can be classified as a redox-active inhibitor of ferroptosis; it functions as an inhibitor of 15-LO to reduce oxidative stress and block depletion of reduced glutathione. By inhibiting 15-LO, utreloxastat is expected to slow or prevent neurodegeneration in ALS by preventing ferroptosis.

The pharmacokinetics (PK) of utreloxastat has been evaluated in nonclinical studies of mice, rats, and monkeys. Utreloxastat was found to be orally bioavailable in all nonclinical studies, and following oral doses to mice, utreloxastat was well distributed into brain tissues with a favorable brain-to-plasma ratio of ≈ 10 -fold after time to maximum plasma concentration (t_{\max}).

In vitro studies were conducted to understand utreloxastat metabolism and drug-drug interaction potential. Multiple cytochrome P450 (CYP) enzymes, potentially but not limited to CYP1A2, 2B6, 2C8, 2C19, 3A4, and 4F2, involved in the metabolism of utreloxastat were investigated using pooled human liver microsomes or recombinant human CYP enzymes. In

vitro utreloxastat was a weak inhibitor for CYP1A2 and 2B6 with half maximal inhibitory concentration $> 5.3 \mu\text{M}$ and inhibition toward other CYP450 enzymes was minor (half maximal inhibitory concentration $> 46 \mu\text{M}$). In human hepatocytes, utreloxastat at $20 \mu\text{M}$ induced messenger RNA expression of CYP2B6 and 3A4 but not CYP1A2. Utreloxastat metabolism in rats was extensive. Following a single oral dose of ^{14}C -radiolabeled utreloxastat, unchanged utreloxastat accounted for $< 5\%$ total radioactivity in plasma but was not detected in urine and bile. The safety of utreloxastat was evaluated in vitro and in rats and monkeys following repeat doses over 28 days. Overall, nonclinical data were favorable and support further development and evaluation of utreloxastat in humans.

The current study assessed the safety, tolerability, and PK of utreloxastat in a phase 1, 3-part, single-center, double-blind, randomized, placebo-controlled, single-ascending-dose (SAD), multiple-ascending-dose (MAD), and food-effect (FE) study in healthy human subjects.

Methods

Study Design

The study was conducted in compliance with the principles of the Declaration of Helsinki and the current Dutch Medical Research Involving Human Subjects Act (Wet medisch-wetenschappelijk onderzoek met mensen) and was approved by the appropriate institutional review boards. One clinical site (QPS Netherlands B.V, Groningen, Netherlands) participated in the study. The clinical protocol and informed consent statements were reviewed and approved by the Independent Ethics Committee of the Foundation Evaluation of Ethics in Biomedical Research (Stichting Beoordeling Ethiek Biomedisch Onderzoek; Assen, the Netherlands). Written informed consent was obtained from each subject before enrollment in the study.

Healthy subjects between 18 and 55 years of age were eligible to participate in the study. Subjects were excluded if they had a history of coagulopathy or fat malabsorption or had any prior medical or psychological history that could jeopardize their safety in the opinion of the investigator. Subjects were also excluded if they had dietary restrictions preventing them from adhering to the dietary requirements of the study, had a history of drug abuse, used nicotine-containing products, or had a positive drug urine screen. Diet, exercise, caffeine, and smoking were restricted during the study. Known CYP3A inhibitors/inducers (eg, grapefruit) were not permitted.

This was a randomized, double-blind, placebo-controlled study in healthy volunteers. The study

consisted of 3 parts: part 1 was a SAD study, part 2 was a MAD study; and part 3 was a FE study.

The starting dose of 100 mg was based on the most sensitive species' no observed adverse effect level from 1-month toxicology studies in rats and monkeys. The no observed adverse effect level found in rats (100 mg/kg) has a human equivalent dose of 16 mg/kg. The 100-mg starting dose in humans represents one-tenth of the human equivalent dose for an adult with a median body weight of 60 kg.

Utreloxastat was prepared in sesame oil (US Pharmacopeia/National Formulary) and packed in hard gelatin 50 mg capsules. Placebo to utreloxastat was sesame oil (US Pharmacopeia/National Formulary) packed in the same type of hard gelatin capsules.

Part 1: SAD

In part 1 (SAD), 40 subjects were randomized into 1 of 4 cohorts (10 subjects each) to receive either placebo treatment ($n = 2$) or single doses of utreloxastat at 100, 250, 500, or 1000 mg ($n = 8$). For each cohort, initial sentinel dosing was performed in 2 subjects (1 subject with utreloxastat and 1 subject with placebo) before the remaining subjects were dosed (7 subjects with utreloxastat and 1 subject with placebo).

Subjects received oral utreloxastat or placebo on day 1 after an overnight fast and after the consumption of a medium-fat breakfast. The medium-fat diet consisted of 500–800 total calories with 25%–50% of calories from fat. An example of a medium-fat meal consisted of 3 slices of bread with 30 g of diet margarine; 25 g of jam, honey, or apple syrup; 1 slice (20 g) of meat (chicken filet, ham, or smoked beef); 1 slice (20 g) of cheese; 200 mL of demi milk; and 150 mL of decaffeinated or thein-free tea. Subjects were released from the clinic after all study procedures had been completed on day 2 and returned to the clinic for sampling on day 3 (48 hours) and day 4 (72 hours). Escalation to the next dose level was based on a review of safety and tolerability data from the previous cohort by the safety review committee.

Part 2: MAD

In part 2 (MAD), 30 subjects were randomized into 1 of 3 cohorts (10 subjects each) to receive 14 days of treatment with either placebo ($n = 2$) or utreloxastat ($n = 8$) at doses of either 250 mg twice daily, 500 mg once daily, or 150 mg twice daily/once daily. Utreloxastat was administered with a medium-fat diet as in part 1. For subjects receiving a twice-daily dose, the evening dose was administered 12 hours after the morning dose with a medium-fat diet.

In both parts 1 and 2, progression to the next dose level was based on a review of the safety and

tolerability data from at least 7 evaluable subjects in the applicable cohort and all available data from the previous cohort(s).

Part 3: FE

Part 3 (FE) consisted of a 3×3 crossover design with randomized, sequential treatment periods of 12 healthy subjects. Subjects were randomized into 1 of 3 sequences and would receive 1 of the 3 treatments in a period with a single dose of utreloxastat 500 mg on day 1 of each period in either a fasted, low-fat fed, or high-fat fed condition. Subjects were released from the clinic on day 2 of each treatment period and returned to the clinic for sampling on day 3 (48 hours) and day 4 (72 hours). Each treatment period was separated by a 7-day washout period, which could be shortened or prolonged depending on the PK results of part 1.

A utreloxastat dose of 500 mg QD was selected on the basis of the mean exposure in part 1 (SAD) that would be equivalent to or greater than the nonclinical model-projected efficacious exposure, as measured by reduction of lipoxygenase-mediated ferroptotic activity. The low-fat and high-fat diets were comparable to those suggested by guidance from the Food and Drug Administration.¹⁴

Safety Assessment

The overall safety profile of utreloxastat in the SAD, MAD, and FE parts of the study was characterized by type, frequency, severity, timing, and relationship to study treatment of any adverse events (AEs), vital signs, laboratory abnormalities, physical examination abnormalities, Columbia-Suicide Severity Rating Scale scores, or electrocardiogram (ECG) abnormalities. AEs were collected from screening of each part of the study until the end of that part of the study.

Pharmacokinetic Sample Collection and Analysis

Approximately 2.0 mL of blood for PK samples were collected before dosing and at 0.5, 1, 2, 3, 4, 6, 8, 10, 12, 24, 48, and 72 hours after dosing following utreloxastat oral administration on day 1 (SAD and FE) and before dosing and at 0.5, 1, 2, 3, 4, 6, 8, 10, 12, 14, 16, 18, and 24 hours on days 1 and 14 following the morning dose, with an additional predose sample on day 7 before the morning dose and additional postdose samples on day 14 at 48 and 72 hours (MAD).

The concentrations of utreloxastat in plasma were determined using validated liquid chromatography with tandem mass spectrometry (LC-MS/MS) assays. Utreloxastat concentrations were calculated by interpolation from a calibration curve. The calibration range for quantitation of utreloxastat in human plasma was 0.5–500 ng/mL. Utreloxastat in dipotassium

ethylenediaminetetraacetic acid human plasma (100 μ L) and the internal standard (IS), utreloxastat-d₆, were extracted by a liquid-liquid extraction procedure using tertbutyl methyl ether. The mixture was mixed and centrifuged, and the organic layer was transferred to a clean glass tube. After evaporation of the organic layer under nitrogen at 40 °C, the residue was reconstituted into a small amount of reconstitution solution (0.5% formic acid in 80:20, v/v acetonitrile/water). A 5- μ L sample of the reconstituted extract was injected on a high-performance liquid chromatography system and quantified using tandem mass spectrometers. The chromatographic separation was performed using Acquity UPLC CSH Phenyl-hexyl column (1.7 μ m, 2.1 \times 50 mm, Waters Corp., Milford, Massachusetts), maintained at 0.8 mL/min flow rate and using mobile phase as water and acetonitrile (60:40, v/v). The multiple reaction monitoring transition ions monitored were m/z 277.2 \rightarrow 165.2 and m/z 283.3 \rightarrow 168.2 for utreloxastat and the IS (utreloxastat d₆), respectively. Analytes to IS peak area ratios for the standards were used to obtain a linear calibration curve using 1/x² weighted least-squares regression analysis. Owing to the photolabile nature of utreloxastat, the method was validated in yellow light conditions (in the absence of white light). Due to high concentrations expected in escalating-dose cohorts in the first-in-human study, an additional LC-MS/MS method with a concentration range of 400–40000 ng/mL was fully validated with methodology like the low-range method.

The method for quantitation of utreloxastat in human urine that had been pretreated with 2% human serum albumin was validated in the range of 0.1–100 ng/mL using LC-MS/MS. In addition to maintaining a yellow light environment, extraction was performed in glass tubes to avoid nonspecific adsorption of utreloxastat to urine collection containers. Similar chromatographic and mass spectrometric conditions were used for quantification of urine samples as for plasma specimen.

Plasma specimens were stored deep frozen at –80 °C in polypropylene containers, whereas urine specimen glass containers were stored at –20 °C until analysis instead of –80 °C to avoid the potential breaking of glass containers at ultra-low temperature. Quality control samples were analyzed throughout the study, and their measured concentrations were used to determine between-run and overall precision and accuracy of the assays. The lower limit of quantitation of plasma and urine assays was 0.5 and 0.1 ng/mL, respectively. The assay performance during the study was demonstrated by specificity, precision, and accuracy of plasma quality control samples, and reproducibility was demonstrated by successful performance of incurred sample

reanalysis that met acceptance criteria. PK parameters were calculated using a noncompartmental model with Phoenix WinNonlin (Certara, Princeton, New Jersey). Terminal half-life ($t_{1/2}$) was calculated with at least 3 data points after t_{max} in the regression and reported only when the adjusted R^2 was ≥ 0.80 and the ratio of the area under the concentration-time curve (AUC) from time 0 to the last measured (or measurable) concentration (AUC_{0-t}) to the AUC from time 0 to infinity (AUC_{0-inf}) was ≥ 0.80 .

Dose proportionality was examined across dose groups in part 1 (SAD) using a power model¹⁵ with ln-transformed maximum plasma concentration (C_{max}), AUC_{0-t} , and AUC_{0-inf} with the following equation:

$$\ln(\text{parameter}) = \alpha + \beta \times \ln(\text{dose})$$

where α was the intercept and β was the slope. Dose proportionality was accepted if the 90%CI constructed on the estimate of slope parameter β included 1.

Results

Subject Demographics

A total of 40 subjects (19 men, 21 women) participated in part 1 (SAD), 30 subjects (19 men, 11 women) participated in part 2 (MAD), and 12 subjects (6 men, 6 women) participated in part 3 (FE) of the study (Table 1). All subjects completed the study aside from 1 subject in part 2 (MAD) who withdrew for family reasons. The median age of participants was between 29.5 and 35.2 years for subjects in each part of the study. The majority of subjects were White (79%) and non-Hispanic or Latino (98%).

Pharmacokinetic Results

Single Dose. Following the first dose of utreloxastat between 100 and 1000 mg, the first plasma sample with measurable utreloxastat concentration ranged from the first sampling time 0.5 hour to 4 hours after dosing (Table 2). Utreloxastat plasma t_{max} was ≈ 4 hours, with a large range from 1 to 6 hours following a single dose. The elimination of utreloxastat followed a biphasic model, with a rapid distribution $t_{1/2}$ of ≈ 1.8 hours and a relative slower $t_{1/2} \approx 23$ hours after dosing. A second concentration peak (shoulder peak) of utreloxastat was observed around 10 hours for some subjects. The likelihood of the appearance of the second peak was higher with higher utreloxastat dose (Figure 2). The elimination rate was consistent in the dose range from 100 to 1000 mg.

The utreloxastat exposures (C_{max} , AUC_{0-t} , and AUC_{0-inf}) increased slightly over dose proportionally in the dose range from 100 to 1000 mg following a single dose of utreloxastat (Figure 3). The slope β was ≈ 1.225 , 1.257, and 1.223 for C_{max} , AUC_{0-t} , and AUC_{0-inf} ,

Table 1. Summary of Subject Demographics

Variable/Status	Part 1 (SAD) All Subjects	Part 2 (MAD) All Subjects	Part 3 (FE) All Subjects
N	40	30	12
Age, y			
Mean (SD)	29.5 (9.54)	35.0 (8.82)	35.2 (8.04)
Median (min-max)	27.0 (18–55)	32.0 (21–53)	36.0 (23–46)
Sex, n (%)			
Female	21 (52.50)	11 (36.67)	6 (50)
Male	19 (47.50)	19 (63.33)	6 (50)
Ethnicity, n (%)			
Hispanic or Latino	1 (2.50)	1 (3.33)	0
Not Hispanic or Latino	39 (97.50)	29 (96.67)	12 (100)
Race, n (%)			
Asian	1 (2.50)	4 (13.33)	0
Black or African American	1 (2.50)	6 (20)	1 (8.33)
White	36 (90)	19 (63.33)	10 (83.33)
American Indian or Alaska Native	0	1 (3.33)	0
Other	2 (5)	0	1 (8.33)
Body weight, kg			
Mean (SD)	73.7 (11.2)	76.1 (10.9)	76.9 (11.8)
Median (min-max)	73.7 (52.3–101.6)	76.7 (58.4–96.5)	77.8 (57.1–95.5)

Abbreviations: FE, food-effect; MAD, multiple ascending dose; max, maximum; min, minimum; SAD, single ascending dose; SD, standard deviation.

Table 2. Summary of Plasma Pharmacokinetic Parameters of Utreloxastat Following a Single Oral Dose of Utreloxastat in Healthy Volunteers After a Medium-Fat Breakfast on Day 1 (Part 1 SAD)

Parameters ^a	100 mg	250 mg	500 mg	1000 mg
	Mean (SD) (N = 8) ^b	Mean (SD) (N = 8)	Mean (SD) (N = 8) ^b	Mean (SD) (N = 8)
C_{max} , ng/mL	264 (111)	670 (3.11)	2310 (922)	3930 (1680)
t_{max} , h	4.0 (1.0, 4.0)	4.0 (2.0, 6.0)	3.5 (2.0, 4.0)	4.0 (3.0, 6.0)
AUC_{0-t} , ng · h/mL	1160 (350)	3640 (748)	9960 (2380)	19 500 (5020)
AUC_{0-inf} , ng · h/mL	1310 (322)	3810 (748)	10 700 (2380)	20 600 (4960)
$t_{1/2}$, h	22.6 (4.9)	20.6 (3.7)	25.3 (8.8)	24.8 (10.1)
CL/F, L/h	81.9 (26.5)	67.5 (11.5)	49.4 (13.9)	51.0 (11.7)
C_{max}/D , ng/mL/mg	2.64 (1.11)	2.68 (1.24)	4.62 (1.84)	3.93 (1.68)
AUC_{0-t}/D , ng · h/mL/mg	11.6 (3.50)	14.6 (2.99)	19.9 (4.76)	19.5 (5.02)

Abbreviations: AUC_{0-inf} , area under the concentration-time curve from time 0 to infinity; AUC_{0-t} , area under the concentration-time curve from time 0 to time t, where t is the time of the last measured (or measurable) concentration; AUC_{0-t}/D , dose normalized AUC_{0-t} ; CL/F, total body clearance; C_{max} , the maximum observed plasma concentration; C_{max}/D , dose normalized C_{max} ; $t_{1/2}$, apparent terminal half-life.

^aData presented are mean (SD) for all parameters except for t_{max} , which are median (minimum-maximum).

^bOne subject who received 100 mg and 1 subject who received 500 mg had an adjusted R^2 value or AUC_{0-t}/AUC_{0-inf} ratio <0.80. Their CL/F, AUC_{0-inf} , and $t_{1/2}$ were not reportable and were not included in the summary statistics (therefore, N = 7 for these statistics).

respectively, following the power model, with 90% CIs of the slope parameter β lower bound just above the value of 1.

Multiple Dose. Subjects in the MAD 250-mg twice-daily cohort received only the morning dose on day 14 by mistake. Subjects in the 150-mg twice-daily/once-daily cohort received utreloxastat 150 mg twice daily from day 1 to day 6 and 150 mg once daily from day 7 to day 14. Following multiple doses of utreloxastat at doses of 250 mg twice daily, 150 mg twice daily/once

daily, and 500 mg once daily for 14 days (MAD), the absorption rate and mean t_{max} were similar to the first doses administered (between 2.5 and 3.5 hours on day 1 following the first dose, and between 2.0 and 4.0 hours on day 14) (Figure 4, Table 3). The $t_{1/2}$ was longer on day 14 with a mean value between 33.2 and 38.5 hours for evaluable subjects, who comprised less than half of the total number of subjects in the MAD study. The mean $t_{1/2}$ of the remaining subjects was \approx 70 hours; this statistic was not reported formally because the

Table 3. Summary of Plasma Pharmacokinetic Parameters of Utreloxastat Following Multiple Oral Doses of Utreloxastat (Part 2 MAD) After a Medium-Fat Breakfast (Days 1 and 14)

Parameters	250 mg Twice Daily Mean (SD) ^a			500 mg Once Daily Mean (SD) ^a			150 mg Twice Daily/Once Daily ^b Mean (SD) ^a		
	Day 1 First Dose (N = 8)	Day 1 Second Dose (N = 8)	Day 14 (N = 8)	Day 1 First Dose (N = 8)	Day 14 (N = 7)	Day 14 (N = 7)	Day 1 First Dose (N = 8)	Day 1 Second Dose (N = 8)	Day 14 (N = 8)
C _{max} , ng/mL	788 (213)	876 (551)	1300 (299)	2270 (754)	2630 (773)	2630 (773)	569 (249)	588 (173)	603 (262)
t _{max} , h ^a	2.5 (2.0–4.0)	4.0 (2.0–6.0)	3.0 (1.0, 6.0)	2.50 (1.98–6.00)	2.0 (2.0–8.0)	2.0 (2.0–8.0)	3.5 (2.0–4.0)	2.0 (2.0–4.0)	4.0 (2.0–6.0)
C _{trough} , ng/mL	NA	NA	282 (73.9)	NA	162 (82.1)	162 (82.1)	NA	NA	49.4 (14.3)
AUC _{0-tau} , ng · h/mL	2650 (579)	3400 (1400)	5960 (1190)	9010 (1840)	12800 (3270)	12800 (3270)	1780 (544)	2290 (838)	3320 (969)
AUC ₀₋₂₄ , ng · h/mL	2650 (579)	2650 (579)	5960 (1190)	NC	NC	NC	1780 (544)	1780 (544)	2610 (753)
t _{1/2} , h	6070 (1860)	NA	NC	9010 (1840)	12,800 (3270)	12,800 (3270)	41.10 (1330)	41.10 (1330)	3320 (969)
R _{acc} ^f	NA	NA	35.5 (1.77) ^c	NA	38.5 (3.73) ^d	38.5 (3.73) ^d	NA	NA	33.2 (NA) ^e
t _{EHL} , h	NA	NA	2.32 (0.639)	NA	1.45 (0.289)	1.45 (0.289)	NA	NA	1.50 (0.335) ^g
	NA	NA	14.7 (5.46)	NA	14.1 (5.29)	14.1 (5.29)	NA	NA	16.4 (5.07)

Abbreviations: AUC₀₋₁₂, area under the concentration-time curve from time 0 to 12 hours; AUC₀₋₂₄, area under the concentration-time curve from time 0 to 24 hours; AUC_{0-inf}, area under the concentration-time curve from time 0 to infinity; AUC_{0-t}, area under the concentration-time curve from time 0 to time t, where t is the time of the last measured (or measurable) concentration; AUC_{0-tau}, area under the concentration-time curve within the dosing interval, calculated by linear up/log down trapezoidal method; C_{max}, maximum observed plasma concentration; C_{trough}, plasma concentration on day 14 before the morning dose; MAD, multiple ascending doses; NA, not applicable; NC, not calculated; R_{acc}, accumulation ratio; R², coefficient of determination; SD, standard deviation; t_{1/2}, apparent terminal half-life; t_{EHL}, effective half-life; t_{max}, time to reach C_{max}.

^aData presented are mean (SD) for all parameters except for t_{max}, which are medians (minimum-maximum).

^bFrom day 1 to day 6, subjects were dosed twice daily, and from day 7 to day 14, subjects were dosed once daily.

^cN = 3: There were 5 subjects whose adjusted R² value or AUC_{0-t}/AUC_{0-inf} ratio was <0.80; therefore, their t_{1/2} values were not reportable and were not included in the summary statistics.

^dN = 3: There were 4 subjects whose adjusted R² value or AUC_{0-t}/AUC_{0-inf} ratio was <0.80; therefore, their t_{1/2} values were not reportable and were not included in the summary statistics.

^eN = 1: There were 7 subjects whose adjusted R² value or AUC_{0-t}/AUC_{0-inf} ratio was <0.80; therefore, their t_{1/2} values were not reportable and were not included in the summary statistics.

^fR_{acc} was based on AUC₀₋₁₂ on day 14/AUC₀₋₁₂ on day 1 for MAD cohort 2:1 (250 mg twice daily) and cohort 2:3 (150 mg twice daily/once daily). R_{acc} was based on AUC_{0-tau} on day 14/AUC_{0-tau} on day 1 for MAD cohort 2:2 (500 mg once daily). AUC_{0-tau} = AUC₀₋₂₄ for cohort 2:2 on days 1 and 14.

^gN = 7: There was 1 subject whose t_{EHL} could not be estimated.

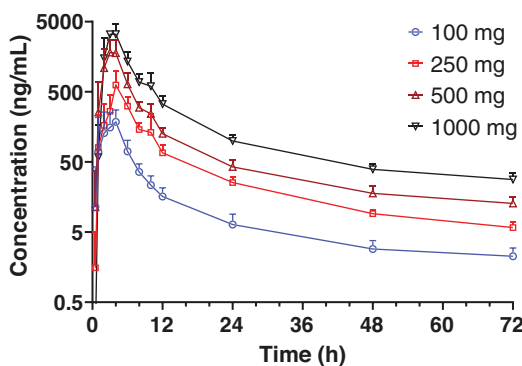


Figure 2. Mean (+SD) utreloxastat plasma concentration versus time following a single oral administration of utreloxastat with a medium-fat diet in healthy subjects (part I SAD). SAD, single ascending dose; SD, standard deviation.

AUC_{0-t}/AUC_{0-inf} ratio was greater than 0.80 for those subjects.

On day 1, utreloxastat exposures were assessed following administration of 250 or 150 mg utreloxastat with a medium-fat diet after both the morning dose (intensive PK sampling) and evening dose (reduced PK sampling due to overnight schedule).

Overall, no significant difference of exposure after the morning and evening doses was observed. Following the morning and evening dose of 250 mg of utreloxastat, the mean C_{max} was 788 and 876 ng/mL, respectively, and the mean AUC within the dosing interval (AUC_{0-tau}) was 2650 and 3400 hour \times ng/mL (Table 3). After the morning and evening dose of 150 mg of utreloxastat, the mean C_{max} was 569 and 588 ng/mL, respectively, and mean AUC_{0-tau} was 1780 and 2290 hour \times ng/mL, respectively. The slightly greater AUC_{0-tau} following evening dose could be due to reduced sampling and inadequate capture of the concentration-time profiles.

The predose concentrations increased only slightly from day 7 to day 14 in the 250-mg twice-daily and 500-mg once-daily dose groups, indicating that concentrations approached a steady state following 7 days of consecutive dosing. The plasma concentration before the morning dose on day 14 increased following dose escalation from 150 mg once daily to 500 mg once daily and 250 mg daily. The accumulation ratio of AUC_{0-tau} or the AUC from time 0 to 12 hours after dosing on day 14 to day 1 (R_{acc}) was ≈ 1.5 following 150 mg twice daily/once daily and 500 mg once daily and 2.32 following 250 mg twice daily. The effective half-life based on R_{acc} was ≈ 15 hours in the dose range following either twice-daily or once-daily dosing.

The clearance was reduced following multiple doses of utreloxastat over 14 days (Figure 4) for both the alpha phase of distribution and the terminal elimination

phase. This was also reflected in the much longer $t_{1/2}$ on day 14.

The amounts of unchanged utreloxastat excreted in urine on day 1 and day 14 following multiple doses were limited ($Fe < 0.23\%$), suggesting that the majority of absorbed utreloxastat was either eliminated by metabolism or biliary excretion.

Food Effect. The effect of food on utreloxastat was assessed in a crossover study with 12 subjects following a single dose of 500 mg of utreloxastat with a low- and high-fat diet (FE) (Figure 5). The effect of a medium-fat diet on utreloxastat from 8 subjects in the SAD study who received a single dose of 500 mg was also included in the analysis. Administration of utreloxastat with food increased the rate of absorption (C_{max}) and extent of absorption (AUC_{0-t} and AUC_{0-inf}) (Table 4).

Food did not alter the t_{max} or clearance of utreloxastat. The mean t_{max} was within the range of 2.5–3.5 hours under fasted and low-, medium-, and high-fat conditions. The geometric least square mean ratios (GMRs) in the low-fat diet condition versus the fasted condition for C_{max} , AUC_{0-t} , and AUC_{0-inf} were 180.25%, 148.66%, and 145.90%, respectively. With a high-fat diet, the GMRs for C_{max} , AUC_{0-t} , and AUC_{0-inf} were 241.30%, 182.45%, and 179.07%, respectively.

In addition, 8 subjects in the SAD study were administered a single dose of 500 mg of utreloxastat with a medium-fat diet. The GMRs for C_{max} , AUC_{0-t} , and AUC_{0-inf} versus the 12 subjects in FE study under fasted condition were 169.71%, 161.01%, and 162.25%, respectively.

Sex Effect. Six men and 6 women were enrolled in the FE study. The median (min-max) body weight of male and female subjects was 86.1 (73.4–95.5) kg and 67.8 (57.5–79.6) kg, respectively. The median (min-max) age for male and female subjects was 40.0 (28.0–46.0) and 29.5 (23.0–42.0) years, respectively.

Overall, no apparent difference of exposure was seen between male and female subjects following a single oral dose of utreloxastat. The AUC_{0-t} and AUC_{0-inf} were comparable between male and female subjects under fasted, low-fat, and high-fat conditions (Figure 6). The C_{max} for male and female subjects was comparable in the fasted and high-fat conditions and lower for female subjects in the low-fat condition.

Safety Assessment. Utreloxastat showed no marked safety signal in healthy subjects following a single dose up to 1000 mg and multiple doses for 14 days at 500 mg once daily or 250 mg twice daily. All AEs reported were mild (grade 1) and had resolved by the end of the study. No deaths or treatment-emergent serious AEs or treatment-emergent AEs leading to discontinuation of utreloxastat occurred over the duration of the studies.

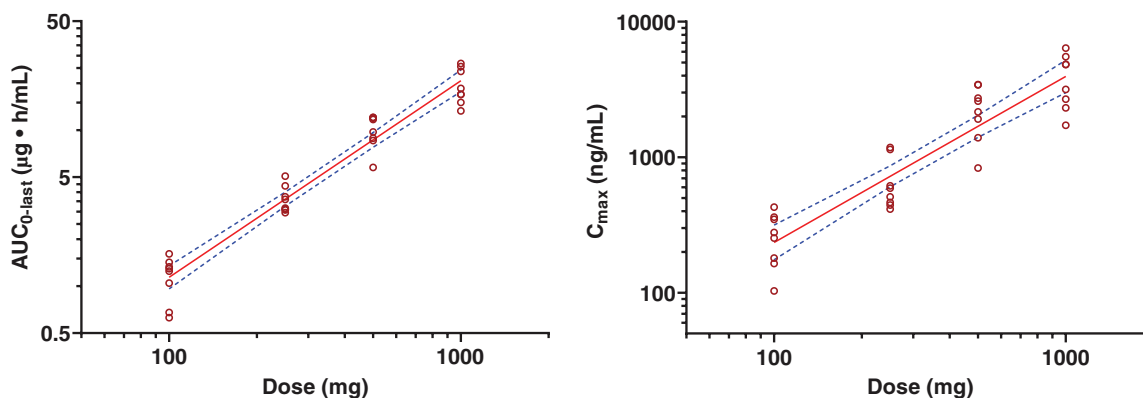


Figure 3. Dose linearity of utreloxastat plasma exposure following a single oral dose of utreloxastat (part I SAD). For AUC, $\beta = 1.26$ (1.15, 1.38), $R^2 = 0.9431$; for C_{max} , $\beta = 1.23$ (1.03, 1.42), $R^2 = 0.8403$. The dashed line represents the 90%CI. AUC, area under the concentration-time curve; AUC_{0-last} , area under the concentration-time curve from time 0 to the last observable time point; C_{max} , maximum observed plasma concentration; SAD, single ascending dose.

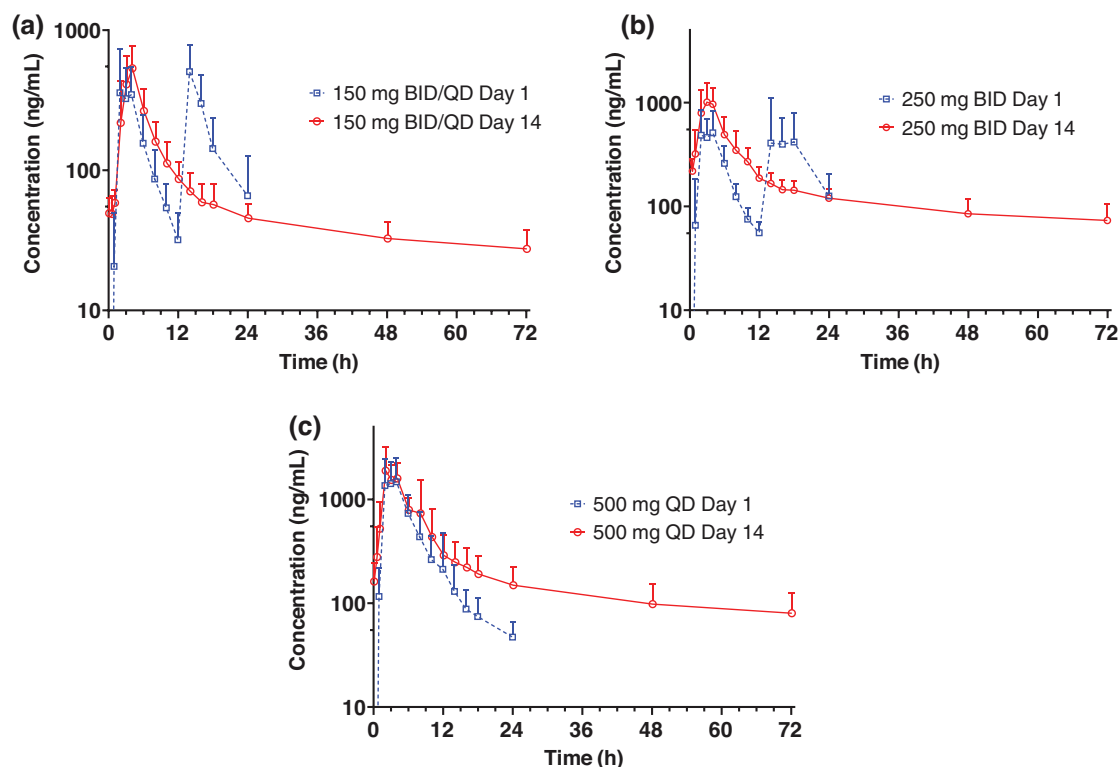


Figure 4. Mean (+SD) utreloxastat plasma concentration versus time following oral administration of utreloxastat at doses of (a) 150 mg, (b) 250 mg, or (c) 500 mg with a medium-fat diet in healthy subjects on days 1 and 14 (part 2 MAD). For subjects who received the 150-mg dose, days 1–6 were dosed twice daily (BID) and days 7–14 were dosed once daily (QD), with only the morning dose administered on day 14. For subjects who received the 250-mg dose BID, only the morning dose was administered on day 14. MAD, multiple ascending doses; SD, standard deviation.

ECGs were monitored in a supine position at screening, check-in, before dosing (before morning dose in twice daily), and 3, 6, and 8 hours after dosing (after morning dose in twice daily) on day 1 (SAD, MAD, and FE), and days 7 and 14 (MAD). No Fridericia corrected QT interval increases of ≥ 60 milliseconds were observed. An

exploratory exposure-response analysis indicated that there was no apparent effect of utreloxastat concentration on corrected QT interval elongation. Overall, there were no clinically abnormal ECG results.

During the MAD study, it was observed that high-density lipoprotein (HDL) and total cholesterol

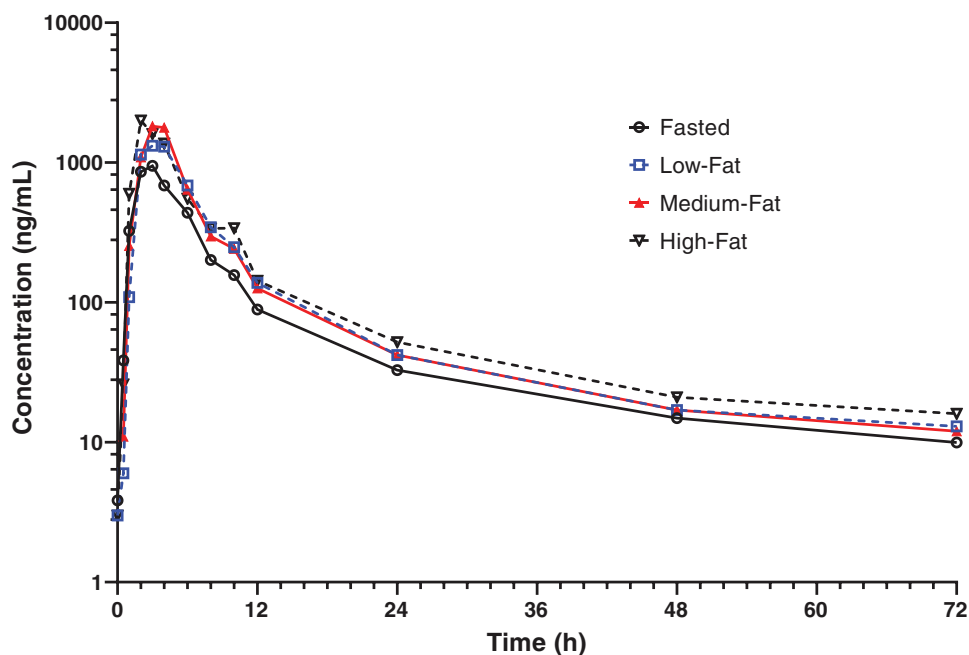


Figure 5. Mean utreloxastat plasma concentration versus time following a single oral administration of 500 mg of utreloxastat under various food conditions in healthy subjects (part 3, FE). FE, food effect.

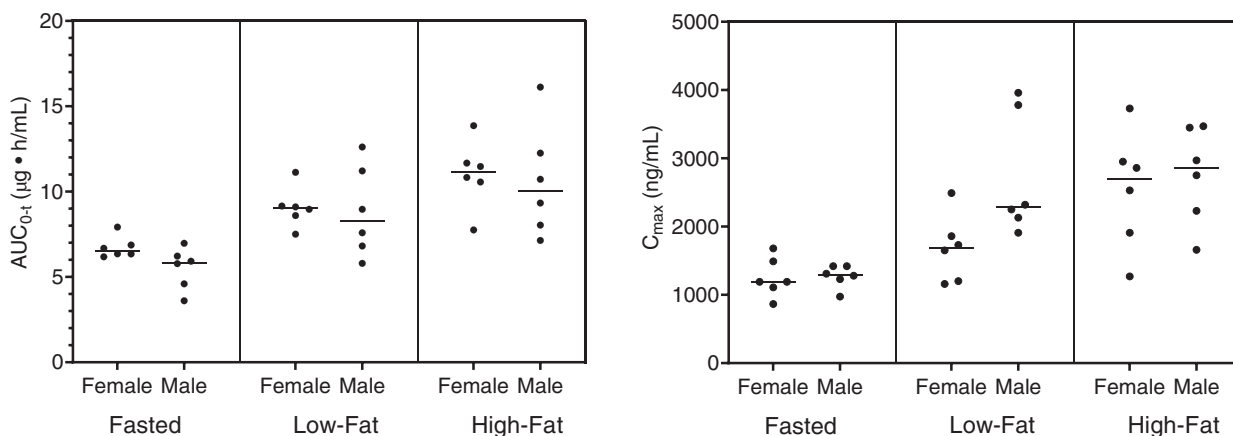


Figure 6. The effect of sex on utreloxastat exposure (AUC_{0-t}) following a single oral dose of 500 mg of utreloxastat. $N = 6$ each for female and male subjects. The bar represents the median value. AUC_{0-t} , area under the concentration-time curve from time 0 to time t , where t is the time of the last measured (or measurable) concentration; C_{max} , maximum plasma concentration.

decreased following multiple doses of utreloxastat. A greater reduction in HDL was observed with increasing utreloxastat dose. This decrease in HDL was fully reversible once utreloxastat administration was stopped. There were no AEs associated with these laboratory findings, which were not considered clinically significant.

Discussion and Conclusion

The current phase I study of utreloxastat in healthy volunteers indicated a predictable dose-PK relationship. Utreloxastat was readily absorbed and reached C_{max}

≈ 4 hours after dosing. The PK of utreloxastat followed a biphasic decay. A shoulder peak during the elimination phase around 10 hours after dosing was observed in some subjects during the SAD, MAD, and FE studies. The probability of this shoulder peak increased with high dose or high-fat diet. It was hypothesized that this could be attributed to enterohepatic circulations. In doses ranging from 100 to 1000 mg, exposures to utreloxastat C_{max} , AUC_{0-t} , and AUC_{0-inf} increased slightly over dose proportionally. No apparent utreloxastat exposure difference was observed between male and female subjects. The majority of absorbed

Table 4. Evaluation of the Effect of Food on the Pharmacokinetics of Utreloxastat Following a Single Oral Dose of 500 mg of Utreloxastat in the Fasted Condition (Part 3 FE) Versus Fed Condition (Part 3 FE and Part 1 SAD) in Healthy Subjects

PK Parameter ^a	Fasted Condition	Fed Condition	Fed Condition (Test)/Fasted Condition (reference)	
	GM	GM	GMR	90%CI
Low-fat				
C_{max} , ng/mL	1143.03	2060.29	180.25	147.09–220.88
AUC_{0-t} , ng · h/mL	5891.03	8757.77	148.66	138.76–159.27
AUC_{0-inf} , ng · h/mL	6329.74	9234.79	145.90	135.18–157.46
Medium-fat				
C_{max} , ng/mL	1244.40	2111.91 ^b	169.71	130.37–220.93
AUC_{0-t} , ng · h/mL	6011.19	9678.75 ^b	161.01	133.98–193.50
AUC_{0-inf} , ng · h/mL	6418.90	10,414.90 ^c	162.25	134.10–196.32
High-fat				
C_{max} , ng/mL	1143.03	2758.10	241.30	188.12–309.51
AUC_{0-t} , ng · h/mL	5891.03	10,748.37	182.45	167.68–198.52
AUC_{0-inf} , ng · h/mL	6329.74	11,334.88	179.07	163.10–196.61

Abbreviations: AUC_{0-inf} , area under the concentration-time curve from time 0 to infinity; AUC_{0-t} , area under the concentration-time curve from time 0 to time t, where t is the time of the last measured (or measurable) concentration; C_{max} , maximum plasma concentration; FE, food-effect; GM, geometric mean; GMR, geometric least squares mean ratio; PK, pharmacokinetic; R^2 , coefficient of determination; SAD, single ascending dose

Various diets in the fed condition included a medium-fat diet (part 1 SAD), low-fat diet (part 3 FE), and high-fat diet (part 3 FE).

^a PK parameters include back-transformed least squares mean and confidence intervals from an analysis of variance model performed on log-transformed values. Subjects who received the medium-fat diet (part 1 SAD) were compared to subjects under the fasted condition (part 3 FE).

^b Subjects included those from the SAD study (N = 8).

^c N = 7. There was one subject whose adjusted R^2 value or AUC_{0-t}/AUC_{0-inf} ratio was <0.80, and therefore the AUC_{0-inf} was not reportable and was not included in the summary statistics.

utrelxastat was eliminated either by metabolism or biliary excretion. The amount of unchanged utrelxastat detected in urine was limited (Fe <0.23%).

Administration of utrelxastat with food increased both the C_{max} and AUCs of utrelxastat, and maximum increase was observed with a high-fat diet. However, the difference between various diets (low-, medium-, and high-fat) was considered nonsignificant (GMRs between 170% and 241% for C_{max} , between 149% and 182% for AUC_{0-t} , and between 146% and 179% for AUC_{0-inf}) considering the intersubject variability (\approx 40% for C_{max} and 23% for AUC_{0-t} and AUC_{0-inf}).

Following repeated dosing for 14 days, accumulation of utrelxastat was observed (R_{acc} of 1.5 for once-daily and 2.32 for twice-daily dosing). The effective half-life was 15 hours following either once-daily or twice-daily dosing, and the apparent clearance was reduced. Both the alpha phase and the terminal phase elimination rates were reduced, and the $t_{1/2}$ was \geq 33 hours on day 14 compared to around 23 hours following the first dose.

The current study also indicated a favorable safety profile of utrelxastat in healthy subjects following a single dose up to 1000 mg and multiple doses for 14 days at 500 mg once daily or 250 mg twice daily. All AEs reported during the study were mild and were resolved by the end of the study.

These PK and safety data indicate that utrelxastat is an appropriate candidate for further development for the treatment of ALS. Utreloxastat inhibits 15-LO to reduce oxidative stress and prevent the depletion of glutathione. Specifically, it inhibits ferroptosis through the reduction of the nonheme iron in 15-LO from its active Fe³⁺ state to its inactive Fe²⁺ state. While ALS has several proposed mechanisms, oxidative stress is widely accepted as a factor in disease progression.

Preclinical studies indicate that utrelxastat is more potent in inhibiting ferroptosis than either edaravone or riluzole. These studies found that utrelxastat inhibited ferroptosis with half maximal effective concentration values \leq 98 nM with little effect on cell viability and was 70 times more potent than edaravone at preventing ferroptotic death of human spinal astrocytes. In these experiments and >50 times more potent than riluzole.

ALS represents a high unmet medical need considering the limited treatment options currently available to patients and the severe symptoms and high fatality associated with the disease. Based on the safety profile and PK of utrelxastat, the dose of 250 mg twice daily to be administered with food was selected for further investigation for treatment of patients with ALS in a randomized, placebo-controlled phase 2 study to evaluate safety and efficacy.

Acknowledgements

The authors would like to thank Susie Bryan and Suzanne Keating for their editorial support.

Conflicts of Interest

This study is funded by PTC Therapeutics, Inc. All authors are employees and stock owners of PTC Therapeutics, Inc.

References

1. Kiernan MC, Vucic S, Cheah BC, et al. Amyotrophic lateral sclerosis. *Lancet (London, England)*. 2011;377(9769):942-955.
2. Park H, Yang E. Oxidative stress as a therapeutic target in amyotrophic lateral sclerosis: opportunities and limitations. *Diagnostics*. 2021;11(1546).
3. Talbott EO, Malek AM, Lacomis D. The epidemiology of amyotrophic lateral sclerosis. In: *Handbook of Clinical Neurology*. 3rd ed.: Elsevier; 2016:225-238.
4. Petrov D, Mansfield C, Moussy A, Hermine O. ALS clinical trials review: 20 years of failure. are we any closer to registering a new treatment? *Front Aging Neurosci*. 2017;9:68.
5. Lewerenz J, Ates G, Methner A, Conrad M, Maher P. Oxytosis/Ferroptosis—(Re-) emerging roles for oxidative stress-dependent non-apoptotic cell death in diseases of the central nervous system. *Front Neurosci*. 2018;12:214.
6. Snodgrass RG, Brüne B. Regulation and Functions of 15-Lipoxygenases in Human Macrophages. *Front Pharmacol*. 2019;10(719).
7. Wenzel SE, Tyurina YY, Zhao J, et al. PEBP1 wardens ferroptosis by enabling lipoxygenase generation of lipid death signals. *Cell*. 2017;171(3):628-641.
8. Ye L, Stockwell B. Transforming lipoxygenases: PE-specific enzymes in disguise. *Cell*. 2017;171:501-502.
9. Masaldan S, Bush AI, Devos D, Rolland AS, Moreau C. Striking while the iron is hot: iron metabolism and ferroptosis in neurodegeneration. *Free Radical Biol Med*. 2019;133:221-233.
10. Bogdanov M, Brown RH, Matson W, et al. Increased oxidative damage to DNA in ALS patients. *Free Radical Biol Med*. 2000;29(7):652-658.
11. Ihara Y, Nobukuni K, Takata H, Hayabara T. Oxidative stress and metal content in blood and cerebrospinal fluid of amyotrophic lateral sclerosis patients with and without a Cu, Zn-superoxide dismutase mutation. *Neuro Res*. 2005;27(1):105-108.
12. Perluigi M, Fai Poon H, Hensley K, et al. Proteomic analysis of 4-hydroxy-2-nonenal-modified proteins in G93A-SOD1 transgenic mice—a model of familial amyotrophic lateral sclerosis. *Free Radical Biol Med*. 2005;38(7):960-968.
13. Mitsumoto H, Santella RM, Liu X, et al. Oxidative stress biomarkers in sporadic ALS. *Amyotrophic Lateral Sclerosis*. 2008;9(3):177-183.
14. FDA. Guidance for Industry: Assessing the Effects of Food on Drugs in INDs and NDAs — Clinical Pharmacology Considerations. In: 2022.
15. Hummel J, McKendrick S, Brindley C, French R. Exploratory assessment of dose proportionality: review of current approaches and proposal for a practical criterion. *Pharm Stat*. 2009;8(1):38-49.

Supplemental Information

Additional supplemental information can be found by clicking the Supplements link in the PDF toolbar or the Supplemental Information section at the end of web-based version of this article.