A Multiplex Biomarker Assay Improves the Prediction of Survival in Epithelial Ovarian Cancer

ARTURAS DOBILAS¹, ANNA ÅKESSON², PIA LEANDERSSON³ and CHRISTER BORGFELDT¹

¹Department of Obstetrics and Gynecology, Skåne University Hospital, Lund, Sweden; ²Clinical Studies Sweden–Forum South, Skåne University Hospital, Lund, Sweden; ³Department of Reproductive Medicine Center, Skåne University Hospital, Malmo, Sweden

Abstract. Background/Aim: Epithelial ovarian cancer (EOC) is usually diagnosed in advanced stages and has a high mortality rate. In this study, we used the proximity extension assay from Olink Proteomics to search for new plasma protein biomarkers to predict overall survival (OS) in patients with EOC. Materials and Methods: Peripheral blood samples were obtained preoperatively from 116 EOC patients undergoing primary debulking surgery: 28 early EOC cases (FIGO stage I-II) and 88 advanced EOC cases (FIGO stage III-IV). Proteins were measured using the Olink Oncology II and Inflammation panels. In total, 177 unique protein biomarkers were analysed. Cross-validation and LASSO regression were combined to select prediction models for OS. Results: The model including age and the three-biomarker combination of neurotrophin-3 (NT-3)+transmembrane glycoprotein NMB (GPNMB)+mesothelin (MSLN) predicted worse OS with AUC=0.79 (p=0.004). Adding cancer antigen 125 (CA125) and human epididymis protein 4 (HE4) to the model further improved performance (AUC=0.83; p=0.003). In a postoperative model including age and stage (III+IV vs. I+II), the three-biomarker panel of chemokine (C-C motif) ligand 28 (CCL28)+T-cell leukaemia/lymphoma protein 1A (TCL1A)+GPNMB improved the prediction of OS (from AUC=0.83 to AUC=0.90; p=0.05). In the postoperative model including age and dichotomized stage (III vs. I+II), the biomarkers CCL28 and GPNMB1 improved the prediction of OS (AUC=0.86; p<0.001). The combination of high levels of

Correspondence to: Arturas Dobilas, MD, Department of Obstetrics and Gynecology, Skåne University Hospital, Klinikgatan 12, 221 85 Lund, Sweden. Tel: +46 40332167, Mob.: +46 768950984, e-mail: arturas.dobilas@med.lu.se

Key Words: Epithelial ovarian cancer, overall survival, biomarker.



This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY-NC-ND) 4.0 international license (https://creativecommons.org/licenses/by-nc-nd/4.0).

both CA125 and HE4 predicted worse survival (p=0.05). Conclusion: In this explorative study evaluating the performance of plasma protein biomarkers in predicting OS, we found that adding biomarkers, especially NT-3, to the panel improved the prediction of OS.

Epithelial ovarian cancer (EOC) is considered a silent carcinoma because it is usually diagnosed in advanced stages. Patients with borderline ovarian tumours have an excellent prognosis, but the prognosis is poor for patients with advanced EOC. Despite advances in ovarian cancer therapy, half of patients will die within five years (1, 2). Various biomarkers and their combinations have been tested for the detection of EOC. CA125 has commonly been used since the early 1990s, but it has limitations. A biomarker panel consisting of HE4, CA125 and age can improve discrimination between malignant and benign ovarian tumours (3). HE4 has been found to be an independent marker for shorter progression-free survival and shorter overall survival (4, 5). However, a gold standard has not been found. In the exploration for new biomarker panels, we chose to use the Olink® Oncology II and Inflammation panels, which are broad and well-established protein panels that use very small amounts of plasma.

A series of studies has shown that immunological components play a key role in cancer development. Ovarian cancer can create a complex tumour microenvironment with ascites consisting of a mixture of various immunosuppressive cells that impairs the ability of the patient's immune system to fight the disease (6). A variety of cytokines, chemokines and growth factors are present in EOC (7-9). The aim of the study was to search for new protein biomarkers and biomarker panels.

Materials and Methods

Patients and samples. Peripheral blood samples were obtained preoperatively from 180 women with an adnexal mass admitted for surgery at the Department of Obstetrics and Gynecology, Skåne

Table I. Characteristics of included patients.

		Serous carcinoma	Mucinous carcinoma	Clear cell carcinoma	Endometroid carcinoma	Total
Stage	I	7	4	2	8	21 (18.1%)
	II	6	0	0	1	7 (6.0%)
	III	65	2	1	2	70 (60.4%)
	IV	16	0	0	2	18 (15.5%)
Total		94	6	3	13	116 (100%)

Table II. Biomarker model based on LASSO regression when the reference model included age.

Reference model	AUC (95%)	p-Value*	Sensitivity at 95% specificity	Specificity at 95% sensitivity	Specificity at best point	Sensitivity at best point
Age Additional marker combinations	0.624 (0.516-0.732)	-	0.253 (0.024-0.410)	0.091 (0.000-0.242)	0.879 (0.394-1.000)	0.422 (0.217-0.855)
NT-3+GPNMB+MSLN NT-3+GPNMB NT-3	0.792 (0.707-0.878) 0.768 (0.705-0.870) 0.730 (0.632-0.828)	0.004 0.008 0.054	0.410 (0.253-0.578)	,	0.879 (0.455-1.000)	0.699 (0.410-0.964) 0.615 (0.374-0.976) 0.699 (0.301-0.868)

LASSO: Least absolute shrinkage and selection operator. *Comparison of the reference model including age with the models with added biomarker(s).

University Hospital Lund, Sweden, from 2005 to 2012. Blood was collected in citrate tubes and centrifuged, and the plasma was stored at -20°C until analysed. All diagnoses were verified by histopathological examination. The disease was staged, and morphology was analysed according to the International Federation of Gynecology and Obstetrics (FIGO). No cancer patient had received neoadjuvant chemotherapy. The patient cohort included 28 early EOC cases (FIGO stage I-II) and 88 advanced EOC cases (FIGO stage III-IV). The frozen plasma samples were sent to Olink Proteomics AB, Uppsala, Sweden, for analyses.

Proximity extension assay. Proteins were measured using the Olink Oncology II and Inflammation panels (Olink Proteomics AB, Uppsala, Sweden) according to the manufacturer's instructions. Included biomarkers in each panel have been listed in Leandersson et al. (10). Proximity extension assay (PEA) technology has been described previously (11). The analyses were performed at Olink Proteomics AB in Uppsala, Sweden. The technicians performing the analyses were unaware of the patient disease status. Samples were randomized on the plates and run-in duplicates. Data were quality controlled and normalized using an internal extension control as well as an interplate control to adjust for intra- and internal variation. All assay validation data are available on the manufacturer's website.

Six EOC cases did not pass internal quality control in the PEA analyses and were excluded from statistical analyses due to either large intracorrelation variance or inability to read one of the duplicate samples.

Statistical analyses. To investigate whether combinations of proteins were associated with survival at 60 months, a combination of crossvalidation and LASSO regression was employed. Overall survival at 60 months was chosen since many studies report 5-year survival. We split the data randomly into a training set and a test set. The shrinkage parameter (λ) was estimated using k-fold cross-validation in the training set. To perform variable selection, the estimated shrinkage parameter $\lambda_{\rm CV}$ was then used in the test set. The selected variables and the absolute value of the coefficients were saved, and the process was repeated 10 times. The variables were then ordered by the number of times they were selected and the sum of their estimated coefficients. The lowest ranked variable was removed, and the entire process was repeated until a final model was selected. This method has also been described by Leandersson et al. (2020) (10). The final models were estimated with logistic regression and predicted probabilities from this model were used to assess the model's discriminatory abilities. Receiver operator curves (ROCs) were constructed, and the area under the curve (AUC) was calculated with 95% confidence intervals using the nonparametric bootstrap procedure. Cross-validation and LASSO regression analyses were carried out using R v 4.1.0 (12).

Availability of data and materials. All data were obtained according to the Swedish Act concerning the Ethical Review of Research Involving Humans to ensure confidentiality and are available on reasonable request.

Ethics statement. Written informed consent was obtained from all study participants. Ethical approval was granted by the Ethical

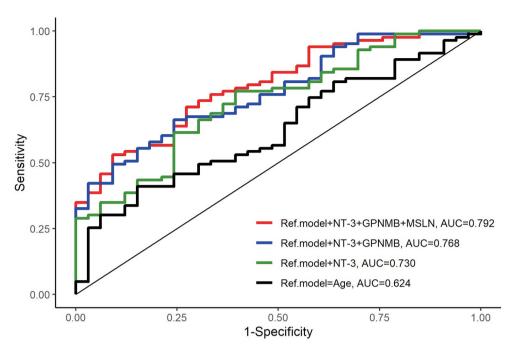


Figure 1. Comparison of the reference model and models with added biomarkers. A three-biomarker model including NT-3, GPNMB, and MSLN with age as the reference model showed the highest prediction of survival (AUC=0.792, p=0.004).

Table III. Biomarker models based on LASSO regression when the reference model includes age, CA125 and HE4.

Reference model	AUC (95%)	<i>p</i> -Value*	Sensitivity at 95% specificity	Specificity at 95% sensitivity	Specificity at best point	Sensitivity at best point
Age+CA125+HE4 Additional marker combinations	0.654 (0.544-0.764)	-	0.169 (0.000-0.349)	0.091 (0.000-0.333)	0.758 (0.363-0.970)	0.578 (0.301-0.916)
NT-3+GPNMB+MSLN NT-3+GPNMB NT-3	0.825 (0.748-0.902) 0.819 (0.742-0.895) 0.779 (0.694-0.864)	0.003 0.003 0.012	0.506 (0.349-0.711)	0.394 (0.091-0.576) 0.273 (0.061-0.556) 0.181 (0.030-0.455)	0.909 (0.727-1.000)	0.735 (0.482-0.926) 0.687 (0.518-0.843) 0.651 (0.386-0.855)

LASSO: Least absolute shrinkage and selection operator. *Comparison of the reference model including age+CA125+HE4 with the models with added biomarker(s).

Review Board at the Faculty of Medicine, Lund University, Sweden. Dnr 495 2016 (amendment to Dnr 558–2004 and 94–2006).

Results

Plasma from 116 EOC patients with different histopathological morphologies was included in the analyses (Table I). The diagnostic performance of a reference model including age only [AUC=0.624 (0.516-0.732)] for discrimination of survival was poor. The biomarker NT-3 alone and in combination with GPNMB and MSLN improved the diagnostic performance, with the best results for the model including age and the three-biomarker panel combination of NT-3+GPNMB+MSLN [AUC=0.792 (0.707-0.878); p=0.004] (Table II, Figure 1).

Adding CA125 and HE4 to age in the reference model improved the results [AUC=0.654 (0.544-0.764)], and the performance of CA125 and HE4 was further improved by the addition of NT-3+GPNMB+MSLN [AUC=0.825 (0.748-0.902); p=0.003] (Table III, Figure 2).

A third reference model was tested including age and stage [early (stage I+II) or late (stage III+IV), AUC=0.830 (0.748-0.913)]. The addition of the CCL28+TCL1A+GPNMB biomarkers to this reference model was found to create the best model for predicting OS [AUC=0.899 (0.831-0.966); p=0.048] (Table IV, Figure 3). When one or both biomarkers (GPNMB and TCL1A) were removed from this model, no statistical significance was observed (p=0.058 and p=0.258).

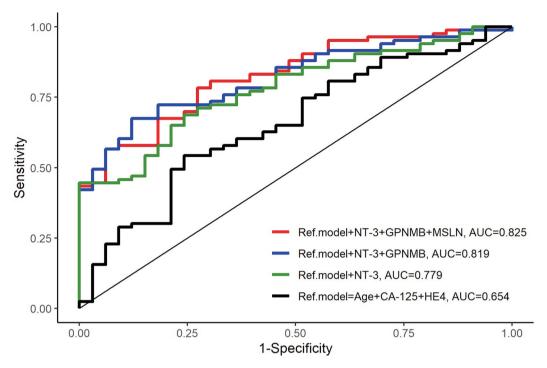


Figure 2. Comparison of the reference model and models with added biomarkers. A three-biomarker model including NT-3+GPNMB+MSLN with age+CA-125+HE4 as the reference model showed the highest prediction of survival (AUC=0.825, p=0.003).

Table IV. Biomarker models based on LASSO regression when the reference model includes age and stage (I-IV).

Reference model	AUC (95%)	<i>p</i> -Value*	Sensitivity at 95% specificity	Specificity at 95% sensitivity	Specificity at best point	Sensitivity at best point
Stage I+II vs. III+IV+age Additional marker combinations	0.830 (0.748-0.913)	-	0.398 (0.253-0.566)	0.455 (0.091-0.758)	0.727 (0.546-0.909)	0.892 (0.675-0.976)
CCL28+TCL1A+ GPNMB	0.899 (0.831-0.966)	0.048	0.657 (0.060-0.888)	0.485 (0.212-0.849)	0.909 (0.758-1.000)	0.855 (0.747-0.40)
CCL28+TCL1A CCL28	0.885 (0.816-0.955) 0.862 (0.782-0.942)	0.058 0.258	0.651 (0.072-0.843) 0.325 (0.024-0.795)	0.546 (0.091-0.788) 0.515 (0.030-0.788)	0.909 (0.727-1.000) 0.879 (0.667-1.000)	0.819 (0.651-0.952) 0.807 (0.675-0.964)

LASSO: Least absolute shrinkage and selection operator. *Comparison of the reference model including age+stage with the models with added biomarker(s).

Finally, starting from a reference model using age only, the addition of stage (early stage I+II vs. late stage III) and CCL28+GPNMB biomarkers improved the prediction of OS and was statistically significant in all combinations, with the best performance for the model including age, stage and CCL28+GPNMB [AUC=0.864 (0.783-0.946); p<0.00] (Table V, Figure 4).

The combination of high levels of both CA125 and HE4 (both biomarkers divided at median) predicted poorer survival (*p*=0.05) (Figure 5).

Discussion

In this explorative study evaluating the performance of plasma protein biomarkers for predicting OS in EOC patients, we found that biomarker panels can improve the prediction of survival. The model including age and the combination of NT-3, GPNMB, MSLN, CA125 and HE4 was the best model to predict overall survival.

Recently, neurotrophins such as NT-3 have been shown to regulate angiogenesis through direct and indirect mechanisms

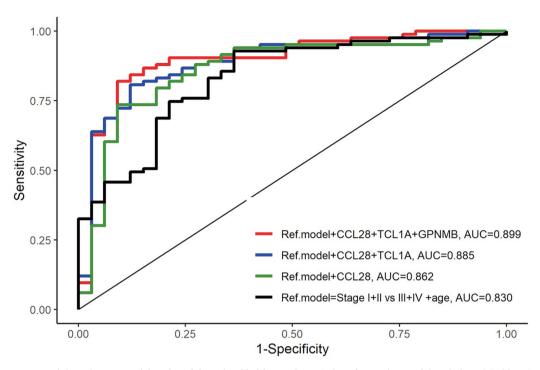


Figure 3. Comparison of the reference model and models with added biomarkers. A three-biomarker model including CCL28+TCL1A+GPNMB with Stage I+II vs. III+IV+age as the reference model showed the highest prediction of survival (AUC=0.899, p=0.048).

Table V. Biomarker models based on LASSO regression when the reference model only includes age (and stage IV is excluded).

Reference model	AUC (95%)	p-Value*	Sensitivity at 95% specificity	Specificity at 95% sensitivity	Specificity at best point	Sensitivity at best point
Age Additional marker combinations	0.633 (0.519-0.746)	-	0.258 (0.015-0.424)	0.063 (0.000-0.250)	0.889 (0.394-1.000)	0.422 (0.217-0.855)
Stage I+II vs. III+CCL28+GPNMB1	0.864 (0.783-0.946)	<0.001	0.364 (0.030-0.788)	0.406 (0.188-0.750)	0.844 (0.656-1.000)	0.818 (0.621-0.955)
Stage I+II vs. III+CCL28	0.856 (0.770-0.942)	0.001	0.318 (0.000-0.803)	0.281 (0.031-0.781)	0.875 (0.688-0.969)	0.803 (0.667-0.955)
Stage I+II vs. III	0.834 (0.750-0.918)	< 0.001	0.439 (0.258-0.621)	0.375 (0.031-0.750)	0.781 (0.563-0.969)	0.849 (0.606-0.970)

LASSO: Least absolute shrinkage and selection operator. *Comparison of the reference model including age with the models with added biomarker(s).

(Garrido *et al.* 2019) (13). Neurotrophins include nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), and neurotrophins 3 and 4/5 (NT-3, NT 4/5), which have a high affinity for tropomyosin kinase receptors (TRKs) that regulate the development and plasticity of the nervous system and nonneuronal tissues, including reproductive organs such as the ovaries. Studies have shown that several types of cancer overexpress neurotrophins, which contribute to tumour progression and angiogenesis (13). The FDA and EMA have approved the use of pharmacologic inhibitors of pan-TRK receptors in patients with TRK fusion-positive

cancers; however, thus far, they have not been used in ovarian cancer treatment.

The low sensitivity and limited specificity of CA125 to detect early-stage EOC (50-62%) do not fulfil the requirements to use CA125 as a screening biomarker in asymptomatic women (14). However, CA125 in combination with other biomarkers or supplemental data, such as age or disease stage in our study, or ultrasound markers can improve performance. The ROMA and ADNEX algorithms, especially in premenopausal women (14), can help differentiate benign ovarian tumours from cancer (15, 16).

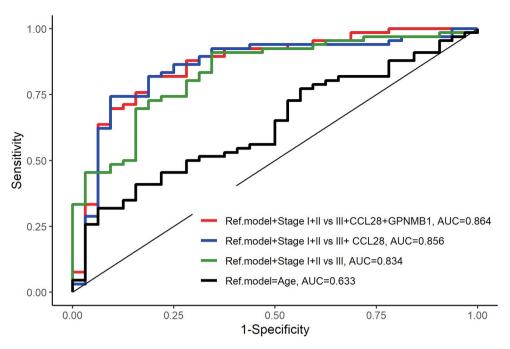


Figure 4. Comparison of the reference model and models with added biomarkers. The model including stage I+II vs. III+CCL28+GPNMB1 with age as the reference model showed the highest prediction of survival (AUC=0.864, p<0.001).

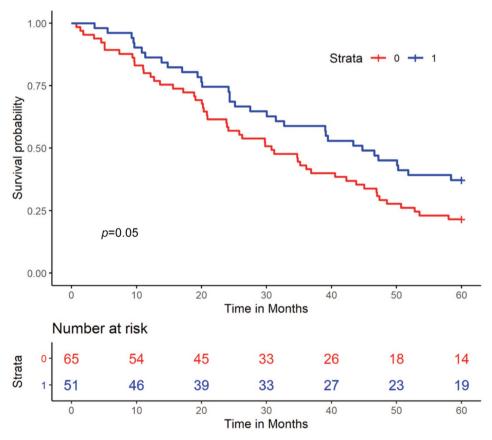


Figure 5. Kaplan-Meier analysis of overall survival in patients in terms of CA125 and HE4 levels. Patients with both values above the median are shown in red, and patients with one or both biomarkers below the median are shown in blue.

According to our results, the combination of high levels of CA125 and HE4 with the addition of NT-3 showed worse five-year OS.

The risk of EOC as well as other cancers increases with age (17, 18). Combining age with the already well-known markers CA125 and HE4 and the addition of NT-3 improved the prediction model of OS significantly. By adding GPNMB and MSLN, we expanded the study model, but no statistically significant results were found, which may have been influenced by the low numbers of patients in the study sample.

We found that increased levels of glycoprotein non-metastatic B (GPNMB) predicted unfavourable survival. GPNMB is also known as osteoactivin, a transmembrane protein overexpressed in several cancer tissues, such as breast cancer (19, 20), stomach cancer (21), colorectal cancer (22), hepatocellular cancer (23) and others. Higher GPNMB levels have been demonstrated to promote angiogenesis, migration, invasion, and metastasis of cancer cells (24, 25).

In our models, mesothelin (MSLN) was also a biomarker of worse survival. The expression of MSLN is dysregulated in several types of cancer, including pancreatic cancer (26) and ovarian cancer (27). Studies have shown that abnormal expression of MSLN plays an important role in tumour cell growth, invasion, and metastasis (28, 29). In ovarian cancer, the specific binding of MSLN and CA125 can mediate the adhesion of tumour cells, which promotes the implantation and metastasis of ovarian cancer in the pelvic and abdominal cavities (30).

High levels of CCL28 in combination with TCL1A and GPNMB as well as EOC stage and age implied worse survival in our study. CCL28 has an important role in regulating the chemotaxis of cells (31).

The results of this study should be interpreted with caution due to the heterogeneity of the epithelial subtypes of histopathology, as only patients scheduled for primary upfront surgery were included in the study, and no patients scheduled for neoadjuvant chemotherapy were included. Cross-validation was used to validate the results. Despite the limitations of the study sample, we obtained positive results, which can provide an impetus for further research.

Conclusion

In summary, we identified biomarker panels predicting overall survival in EOC patients. The model including age and the combination of neurotrophin-3 (NT-3)+transmembrane glycoprotein NMB (GPNMB)+mesothelin (MSLN)+cancer antigen 125 (CA125) and human epididymis protein 4 (HE4) was the best model to predict overall survival. Future research is warranted to replicate our results and identify additional biomarkers in panels that can predict the prognosis of EOC patients.

Funding

The study was supported by funds from the Swedish Cancer Foundation and Regional Funds Region Skåne. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Conflicts of Interest

The Authors declare that they have no conflicts of interest.

Authors' Contributions

AD: Study conception and design; data acquisition, analysis, and interpretation; drafting and revising the manuscript. AÅ: Data acquisition and interpretation; drafting and revising the manuscript. PL: Data acquisition and revising the manuscript. CB: Study conception and design; data acquisition, analysis, and interpretation; revising the manuscript.

Acknowledgements

We are grateful to all the women who participated in this study; to the staff at the Department of Obstetrics and Gynecology, Lund Hospital, for assistance with collecting the blood samples; and to the laboratory staff at the Division of Oncology, Medicon Village, Lund University, for help with all practical issues concerning the preparation and shipping of samples.

References

- 1 Kalapotharakos G, Högberg T, Bergfeldt K and Borgfeldt C: Long-term survival in women with borderline ovarian tumors: a population-based survey of borderline ovarian tumors in Sweden 1960-2007. Acta Obstet Gynecol Scand 95(4): 473-479, 2016. PMID: 26714557. DOI: 10.1111/aogs.12846
- Webb PM and Jordan SJ: Epidemiology of epithelial ovarian cancer. Best Pract Res Clin Obstet Gynaecol 41: 3-14, 2017. PMID: 27743768. DOI: 10.1016/j.bpobgyn.2016.08.006
- 3 Leandersson P, Kalapotharakos G, Henic E, Borgfeldt H, Petzold M, Høyer-Hansen G and Borgfeldt C: A biomarker panel increases the diagnostic performance for epithelial ovarian cancer type I and II in young women. Anticancer Res 36(3): 957-965, 2016. PMID: 26976984.
- 4 Paek J, Lee SH, Yim GW, Lee M, Kim YJ, Nam EJ, Kim SW and Kim YT: Prognostic significance of human epididymis protein 4 in epithelial ovarian cancer. Eur J Obstet Gynecol Reprod Biol 158(2): 338-342, 2011. PMID: 21683503. DOI: 10.1016/j.ejogrb.2011.05.021
- 5 Kalapotharakos G, Asciutto C, Henic E, Casslén B and Borgfeldt C: High preoperative blood levels of HE4 predicts poor prognosis in patients with ovarian cancer. J Ovarian Res 5(1): 20, 2012. PMID: 22909379. DOI: 10.1186/1757-2215-5-20
- 6 Batchu RB, Gruzdyn OV, Kolli BK, Dachepalli R, Umar PS, Rai SK, Singh N, Tavva PS, Weaver DW and Gruber SA: IL-10 signaling in the tumor microenvironment of ovarian cancer. Adv Exp Med Biol 1290: 51-65, 2021. PMID: 33559854. DOI: 10.1007/978-3-030-55617-4
- 7 Lane D, Matte I, Garde-Granger P, Bessette P and Piché A: Ascites IL-10 promotes ovarian cancer cell migration. Cancer

- Microenviron 11(2-3): 115-124, 2018. PMID: 30039195. DOI: 10.1007/s12307-018-0215-3
- 8 Matte I, Lane D, Laplante C, Rancourt C and Piché A: Profiling of cytokines in human epithelial ovarian cancer ascites. Am J Cancer Res 2(5): 566-580, 2012. PMID: 22957308.
- 9 Han GH, Yun H, Chung JY, Kim JH and Cho H: TMED9 expression level as a biomarker of epithelial ovarian cancer progression and prognosis. Cancer Genomics Proteomics 19(6): 692-702, 2022. PMID: 36316042. DOI: 10.21873/cgp.20352
- 10 Leandersson P, Åkesson A, Hedenfalk I, Malander S and Borgfeldt C: A multiplex biomarker assay improves the diagnostic performance of HE4 and CA125 in ovarian tumor patients. PLoS One 15(10): e0240418, 2020. PMID: 33075095. DOI: 10.1371/journal.pone.0240418
- 11 Assarsson E, Lundberg M, Holmquist G, Björkesten J, Thorsen SB, Ekman D, Eriksson A, Rennel Dickens E, Ohlsson S, Edfeldt G, Andersson AC, Lindstedt P, Stenvang J, Gullberg M and Fredriksson S: Homogenous 96-plex PEA immunoassay exhibiting high sensitivity, specificity, and excellent scalability. PLoS One 9(4): e95192, 2014. PMID: 24755770. DOI: 10.1371/journal.pone.0095192
- 12 The R Project: R project for statistical computing, 2022. Available at: https://www.r-project.org [Last accessed on February 19, 2023]
- 13 Garrido MP, Torres I, Vega M and Romero C: Angiogenesis in gynecological cancers: role of neurotrophins. Front Oncol 9: 913, 2019. PMID: 31608227. DOI: 10.3389/fonc.2019.00913
- 14 Sölétormos G, Duffy MJ, Othman Abu Hassan S, Verheijen RH, Tholander B, Bast RC Jr, Gaarenstroom KN, Sturgeon CM, Bonfrer JM, Petersen PH, Troonen H, CarloTorre G, Kanty Kulpa J, Tuxen MK and Molina R: Clinical use of cancer biomarkers in epithelial ovarian cancer: updated guidelines from the European Group on Tumor Markers. Int J Gynecol Cancer 26(1): 43-51, 2016. PMID: 26588231. DOI: 10.1097/IGC.000000000000000586
- 15 Bandiera E, Romani C, Specchia C, Zanotti L, Galli C, Ruggeri G, Tognon G, Bignotti E, Tassi RA, Odicino F, Caimi L, Sartori E, Santin AD, Pecorelli S and Ravaggi A: Serum human epididymis protein 4 and risk for ovarian malignancy algorithm as new diagnostic and prognostic tools for epithelial ovarian cancer management. Cancer Epidemiol Biomarkers Prev 20(12): 2496-2506, 2011. PMID: 22028406. DOI: 10.1158/1055-9965.EPI-11-0635
- 16 Furrer D, Grégoire J, Turcotte S, Plante M, Bachvarov D, Trudel D, Têtu B, Douville P and Bairati I: Performance of preoperative plasma tumor markers HE4 and CA125 in predicting ovarian cancer mortality in women with epithelial ovarian cancer. PLoS One 14(6): e0218621, 2019. PMID: 31220149. DOI: 10.1371/journal.pone.0218621
- 17 Rooth C: Ovarian cancer: risk factors, treatment and management. Br J Nurs 22(17): S23-S30, 2013. PMID: 24067270. DOI: 10.12968/bjon.2013.22.Sup17.S23
- 18 Doubeni CA, Doubeni AR and Myers AE: Diagnosis and management of ovarian cancer. Am Fam Physician 93(11): 937-944, 2016. PMID: 27281838.
- 19 Rose AA, Pepin F, Russo C, Abou Khalil JE, Hallett M and Siegel PM: Osteoactivin promotes breast cancer metastasis to bone. Mol Cancer Res 5(10): 1001-1014, 2007. PMID: 17951401. DOI: 10.1158/1541-7786.MCR-07-0119
- 20 Huang YH, Chu PY, Chen JL, Huang CT, Huang CC, Tsai YF, Wang YL, Lien PJ, Tseng LM and Liu CY: Expression pattern and prognostic impact of glycoprotein non-metastatic B (GPNMB) in triple-negative breast cancer. Sci Rep *11*(1): 12171, 2021. PMID: 34108545. DOI: 10.1038/s41598-021-91588-3

- 21 Rho HW, Lee BC, Choi ES, Choi IJ, Lee YS and Goh SH: Identification of valid reference genes for gene expression studies of human stomach cancer by reverse transcription-qPCR. BMC Cancer 10: 240, 2010. PMID: 20507635. DOI: 10.1186/ 1471-2407-10-240
- 22 Eldai H, Periyasamy S, Al Qarni S, Al Rodayyan M, Muhammed Mustafa S, Deeb A, Al Sheikh E, Afzal M, Johani M, Yousef Z and Aziz MA: Novel genes associated with colorectal cancer are revealed by high resolution cytogenetic analysis in a patient specific manner. PLoS One 8(10): e76251, 2013. PMID: 24204606. DOI: 10.1371/journal.pone.0076251
- 23 Tian F, Liu C, Wu Q, Qu K, Wang R, Wei J, Meng F, Liu S and Chang H: Upregulation of glycoprotein nonmetastatic B by colonystimulating factor-1 and epithelial cell adhesion molecule in hepatocellular carcinoma cells. Oncol Res 20(8): 341-350, 2013. PMID: 23924854. DOI: 10.3727/096504013X13657689382851
- 24 Rose AA, Annis MG, Dong Z, Pepin F, Hallett M, Park M and Siegel PM: ADAM10 releases a soluble form of the GPNMB/Osteoactivin extracellular domain with angiogenic properties. PLoS One 5(8): e12093, 2010. PMID: 20711474. DOI: 10.1371/journal.pone.0012093
- 25 Rich JN, Shi Q, Hjelmeland M, Cummings TJ, Kuan CT, Bigner DD, Counter CM and Wang XF: Bone-related genes expressed in advanced malignancies induce invasion and metastasis in a genetically defined human cancer model. J Biol Chem 278(18): 15951-15957, 2003. PMID: 12590137. DOI: 10.1074/jbc.M211498200
- 26 Le K, Wang J, Zhang T, Guo Y, Chang H, Wang S and Zhu B: Overexpression of mesothelin in pancreatic ductal adenocarcinoma (PDAC). Int J Med Sci 17(4): 422-427, 2020. PMID: 32174772. DOI: 10.7150/ijms.39012
- 27 Li Y, Tian W, Zhang H, Zhang Z, Zhao Q, Chang L, Lei N and Zhang W: MSLN correlates with immune infiltration and chemoresistance as a prognostic biomarker in ovarian cancer. Front Oncol 12: 830570, 2022. PMID: 35692779. DOI: 10.3389/ fonc.2022.830570
- 28 Avula LR, Rudloff M, El-Behaedi S, Arons D, Albalawy R, Chen X, Zhang X and Alewine C: Mesothelin enhances tumor vascularity in newly forming pancreatic peritoneal metastases. Mol Cancer Res 18(2): 229-239, 2020. PMID: 31676721. DOI: 10.1158/1541-7786.MCR-19-0688
- 29 Inoue S, Tsunoda T, Riku M, Ito H, Inoko A, Murakami H, Ebi M, Ogasawara N, Pastan I, Kasugai K, Kasai K, Ikeda H and Inaguma S: Diffuse mesothelin expression leads to worse prognosis through enhanced cellular proliferation in colorectal cancer. Oncol Lett 19(3): 1741-1750, 2020. PMID: 32194667. DOI: 10.3892/ol.2020.11290
- 30 Coelho R, Marcos-Silva L, Ricardo S, Ponte F, Costa A, Lopes JM and David L: Peritoneal dissemination of ovarian cancer: role of MUC16-mesothelin interaction and implications for treatment. Expert Rev Anticancer Ther 18(2): 177-186, 2018. PMID: 29241375. DOI: 10.1080/14737140.2018.1418326
- 31 Rodriguez MW, Paquet AC, Yang YH and Erle DJ: Differential gene expression by integrin beta 7+ and beta 7- memory T helper cells. BMC Immunol *5*: 13, 2004. PMID: 15236665. DOI: 10.1186/1471-2172-5-13

Received January 15, 2023 Revised February 16, 2023 Accepted February 19, 2023