

Saccharomyces genome database update: server architecture, pan-genome nomenclature, and external resources

Edith D. Wong, Stuart R. Miyasato, Suzi Aleksander, Kalpana Karra, Robert S. Nash, Marek S. Skrzypek, Shuai Weng, Stacia R. Engel,* J. Michael Cherry*

Department of Genetics, Stanford University, Stanford, CA 94305, USA

*Corresponding author: Department of Genetics, School of Medicine, Stanford University, 3165 Porter Drive, Palo Alto, CA 94304-5477, USA. Email: stacia@stanford.edu;

*Corresponding author. Email: cherry@stanford.edu

Abstract

As one of the first model organism knowledgebases, *Saccharomyces* Genome Database (SGD) has been supporting the scientific research community since 1993. As technologies and research evolve, so does SGD: from updates in software architecture, to curation of novel data types, to incorporation of data from, and collaboration with, other knowledgebases. We are continuing to make steps toward providing the community with an *S. cerevisiae* pan-genome. Here, we describe software upgrades, a new nomenclature system for genes not found in the reference strain, and additions to gene pages. With these improvements, we aim to remain a leading resource for students, researchers, and the broader scientific community.

Keywords: *Saccharomyces cerevisiae*, budding yeast, SGD, model organism database, knowledgebase, MOD

Introduction

The *Saccharomyces* Genome Database (SGD; www.yeastgenome.org) is a knowledgebase with a purpose to collect, organize, and annotate scientific data about the yeast *Saccharomyces cerevisiae*. A knowledgebase is an online resource that integrates, curates, and organizes published results into an online resource that provides easy access and reusability of these information. SGD's content is managed by a group of dedicated, PhD-trained scientists who screen published research articles, manually curate the information, and connect it to other relevant resources. The curation team works closely with our expert software developers to create a robust website that presents yeast genetic and cellular biology research in a manner that is useful for both experimental and computational scientists. In recent years, software technologies have advanced to provide services and data more quickly. To streamline SGD's framework, we continue to update our website architecture to make use of cloud services and move away from managing physical servers toward serverless compute engines.

Since its creation, SGD has been the authority for the *S. cerevisiae* reference genome (http://sgd-archive.yeastgenome.org/sequence/S288C_reference/; [Cherry et al. 2012](#)), including gene annotation and nomenclature ([Cherry 1995](#)). We have recently updated the reference genome annotation, expanded our collection of allele information, and are developing the Alliance of Genome Resources in partnership with five other model organism knowledgebases and the Gene Ontology (GO) Consortium ([Engel et al. 2022](#)). Although S288C is the strain from which the yeast

reference genome sequence is derived, many publications use other strain backgrounds that contain genes not found in S288C ([Engel et al. 2016](#); [Song et al. 2016](#)). To accommodate these genes so that we can fully serve and support the yeast research community, we are moving toward providing an *S. cerevisiae* pan-genome. We have recently established a new systematic nomenclature for these genes not found in S288C, which we describe below.

Outreach to the community and working with researchers to incorporate published data is another important part of SGD's work ([MacPherson et al. 2017](#)). In recent years, we have collaborated with the ComplexPortal ([Meldal et al. 2022](#)) to curate and to incorporate macromolecular complex data into SGD, resulting in new complex pages covering the complete yeast complexome ([Wong et al. 2019](#)). We have also worked directly with researchers to incorporate genome-wide protein abundance data into SGD protein pages ([Nash et al. 2020](#)). We continue to work with the community to incorporate new connections to external databases, including the recent addition of RNA secondary structures, described below.

Updating server architecture

As new web and cloud technologies develop, we evaluate which would be most appropriate for SGD and of most benefit to our users. SGD has migrated our server infrastructure to utilize Docker containers (<https://www.docker.com/resources/what-container/>) running under the Amazon Web Services (AWS) Fargate (<https://aws.amazon.com/fargate/>) service (Fig. 1).

Received: November 16, 2022. **Accepted:** December 21, 2022

© The Author(s) 2023. Published by Oxford University Press on behalf of the Genetics Society of America.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

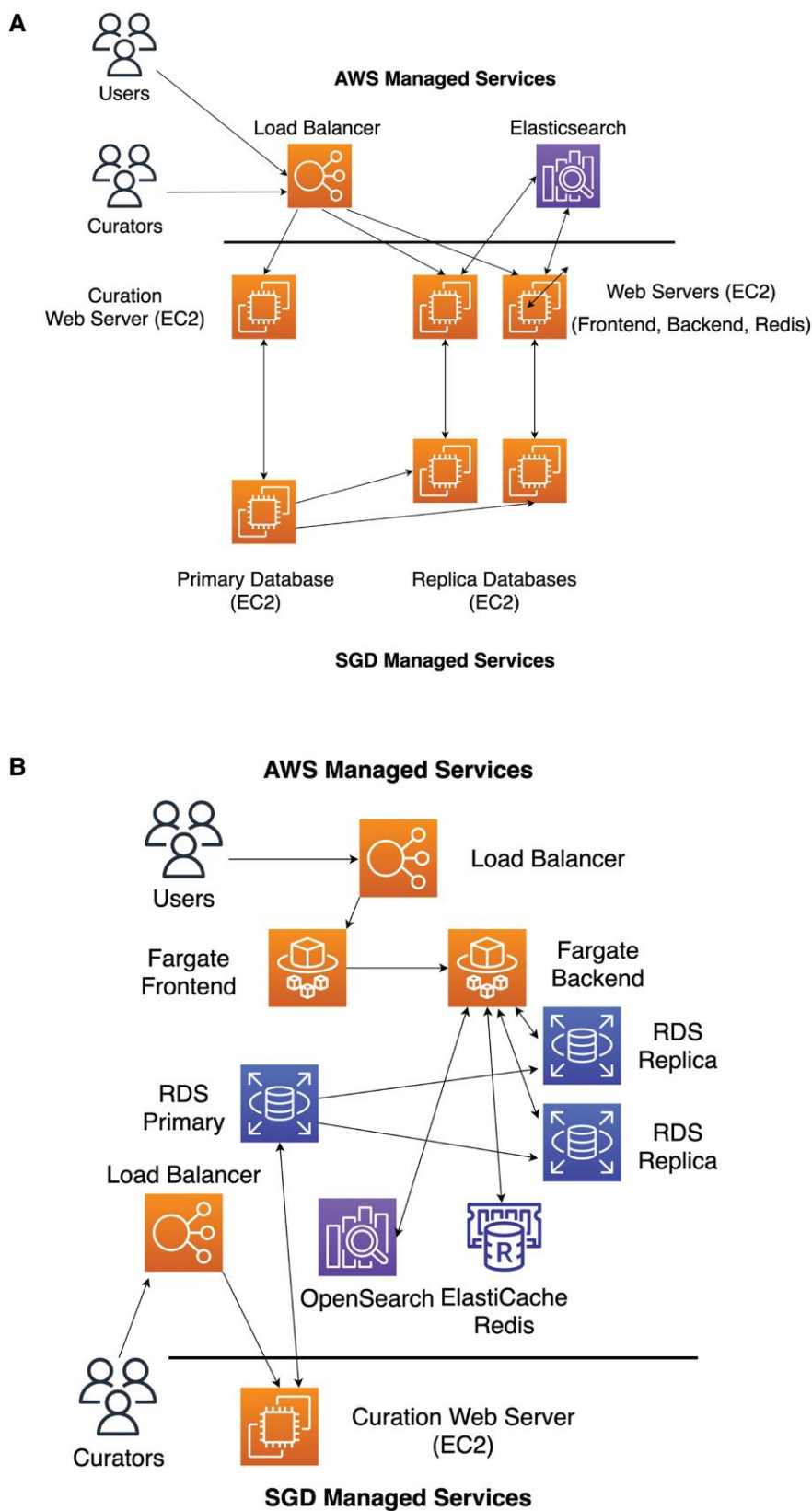


Fig. 1. Streamlining SGD's backend with Fargate and Docker. AWS managed services are above the horizontal line, SGD managed services are below. a) Previous architecture of SGD. AWS managed services are limited to load balancer and elasticsearch; all other services are SGD managed. b) Current architecture, with SGD managed services reduced to the curation web server.

Docker is a platform that enables developers to build, deploy, and run software as immutable “containers,” which are uniform, executable components that comprise the entire application runtime environment including code, libraries, and dependencies. Because each container is “complete,” the software can be written a single time and, like a virtual computer, can run anywhere like desktops, servers, and the cloud, which reduces the need for time-consuming refactoring. A Docker application can therefore be deployed with greater speed, security, reliability, and flexibility than traditional software deployments. As an example, SGD utilizes the same Docker image across all four of our major computing environments: development, QA, staging, and production. Deployment to any environment can be performed with just a few clicks from a management console or by running a simple command-line script. SGD also shares our Docker images to any interested user via the Amazon Elastic Container Registry (ECR, <https://aws.amazon.com/ecr/>) Public Repository (<https://gallery.ecr.aws/yeastgenome/>). If users wish to customize their own Docker image, SGD includes a modifiable Dockerfile as part of our source code repositories located at <https://github.com/yeastgenome/>.

AWS Fargate is a “serverless” environment, managed and operated by AWS. By using Fargate, SGD is no longer responsible for server management tasks that are necessary to maintain normal server operations, but which do not add direct value to the website end user. Performing data backups, upgrading operating system software, and managing security patches are responsibilities now handled by AWS (Fig. 1b). By transferring this nondifferentiated work to AWS, SGD staff can direct attention away from systems management and instead focus on adding data and features that directly benefit our end users. This increased efficiency adds value and resiliency to the site, which ultimately reduces costs for SGD in the long run as well.

Fargate detects when a container has become unresponsive and will automatically replace that container with a newly launched one. Fargate also implements auto-scaling, meaning it will launch new containers to handle spikes in demand as needed, as well as deactivate containers when the additional compute capacity is no longer needed. By leveraging such “elasticity,” SGD optimizes cost-effectiveness by deploying only the amount of compute resources that are needed to handle user demand at any given moment.

Initial SGD deployment of Docker containers in Fargate has resulted in modest cost savings. SGD expects these savings to increase over time as more applications are migrated from Elastic Compute Cloud (EC2) instances to Fargate, as SGD realizes increased economy of scale due to differences in data storage models. In the future, SGD also expects to maximize cost-efficiency by migrating some Docker containers to Lambda (<https://aws.amazon.com/lambda/>), the AWS serverless, event-driven service that charges only for compute and memory resources actually used when a job is invoked. Unlike EC2 and Fargate, Lambda charges do not accrue while awaiting the arrival of user requests.

Toward a pan-genome: new systematic names for genes not found in S288C

A number of *S. cerevisiae* genes are found in alternative background strains, but not in the reference genome of S288C. As these features are not in the reference genome sequence, systematic chromosomal locations are not defined. Historically, SGD has cataloged these genes using only their gene names. As gene names can change and are sometimes ambiguous, we have put into place

a new systematic nomenclature that accommodates all genes in the yeast pan-genome that are not found in S288C. This new systematic nomenclature is similar to, but distinct from, those used for ORFs (Cherry 1995; Cherry et al. 1998) and for ncRNAs (Engel et al. 2022). This new nomenclature is for those genes not found in S288C. For those genes that are found in S288C, the established systematic nomenclature remains the same. New genes that are identified in both S288C and non-S288C strains will also be given systematic gene names following the current, established systematic nomenclature when the reference genome is updated (Engel et al. 2022).

Nonreference genes are designated by a symbol consisting of three uppercase letters and a four-digit number, as follows: Y for “Yeast,” SC for “*Saccharomyces cerevisiae*,” and a four-digit number corresponding to the sequential order in which the gene was added to SGD. We currently have 55 of these genes in SGD, some of which are old favorites like maltose permease (MAL21/YSC0004) and the mating-type locus (MATA/YSC0046), while others are more recent additions like xylitol dehydrogenase (XDH1/YSC0051; Table 1). Going forward, as researchers publish evidence pointing to other *S. cerevisiae* genes not present in the S288C reference genome, they will be added to the annotation using the next sequential number available. Already, we have an additional 15 of these YSC names reserved by researchers and awaiting publication.

Additional visuals and external links: structure and homology

SGD continues to incorporate relevant information from external databases (Wong et al. 2019; Nash et al. 2020). We have recently added secondary structures for RNAs from RNAcentral (<https://rnacentral.org/>; RNAcentral Consortium 2021). Secondary structure images can be found on both the Summary and Sequence pages for RNA genes (Fig. 2a). The Sequence pages have a more detailed view of the secondary structures (Fig. 2b). Additionally, RNAcentral IDs are provided as links directly to the corresponding RNAcentral page for more information.

Another way to connect users to external structural and homology databases is through the Resources section at the bottom of the Gene and Complex pages. Within the Resource section of the Summary, Sequence, Protein, and Homology pages, we have added links to the corresponding entry at the AlphaFold Protein Structure Database (<https://alphafold.ebi.ac.uk/>). Similarly, in the Resource section on Complex pages, we added links to ModelArchive (<https://www.modelarchive.org/>), a database of predicted 3-dimensional structures of macromolecular complexes. A newly developed database, AnalogYeast (<https://www.weizmann.ac.il/molgen/AnalogYeast/>), predicts homologous proteins by sequence similarity in other organisms, including humans and other model organisms such as *Caenorhabditis elegans* and *Arabidopsis thaliana* (Cohen et al. 2022). We added links to AnalogYeast’s search results within the Resource sections of the Protein and Homology pages to enable researchers to explore homologs of disease-associated genes.

Continuing directions

As SGD enters its fourth decade, we continue to evolve our database, website architecture, and user interface, keeping it sustainable and current to ensure the website and all curated data are easily accessible for both users and computers. As we progress toward the *S. cerevisiae* pan-genome, equipped with the new

Table 1. New systematic gene names for genes not found in the reference genome.

| Systematic name | Standard gene name | Gene name description | Gene description |
|-----------------|--------------------|--|---|
| YSC0001 | ENS2 | ENdo.SceI | Mitochondrially encoded 50 kDa subunit of Endo.SceI |
| YSC0002 | KHR1 | Killer of heat resistant | Killer toxin |
| YSC0003 | MAL2 | MALtose fermentation | Multigene complex, polymeric locus for maltose fermentation |
| YSC0004 | MAL21 | MALtose fermentation | |
| YSC0005 | MAL22 | MALtose fermentation | Maltose permease |
| YSC0006 | MAL23 | MALtose fermentation | Maltase (alpha-D-glucosidase) |
| YSC0007 | MAL34 | MALtose fermentation | MAL-activator protein |
| YSC0008 | MAL4 | MALtose fermentation | ORF with sequence and positional similarity to the MAL64 gene |
| YSC0009 | MAL41 | MALtose fermentation | Multigene complex, polymeric locus for maltose fermentation |
| YSC0010 | MAL42 | MALtose fermentation | Maltose permease |
| YSC0011 | MAL43 | MALtose fermentation | Maltase (alpha-D-glucosidase) |
| YSC0012 | MAL6 | MALtose fermentation | MAL-activator protein |
| YSC0013 | MAL61 | MALtose fermentation | Multigene complex, polymeric locus for maltose fermentation |
| YSC0014 | MAL62 | MALtose fermentation | High-affinity maltose transporter |
| YSC0015 | MAL63 | MALtose fermentation | Maltase (alpha-D-glucosidase) |
| YSC0016 | MAL64 | MALtose fermentation | MAL-activator protein |
| YSC0017 | MATA1 | MATing type | MAL64 is a nonfunctional homolog of the MAL63 trans-activator |
| YSC0018 | MATA2 | MATing type | Expressed copy (at MATA) of a1 |
| YSC0019 | MEL1 | MELibiose | Protein of unknown function |
| YSC0020 | MEL10 | MELibiose | Secreted alpha-galactosidase |
| YSC0021 | MEL2 | MELibiose | Secreted alpha-galactosidase |
| YSC0022 | MEL3 | MELibiose | Secreted alpha-galactosidase |
| YSC0023 | MEL4 | MELibiose | Secreted alpha-galactosidase |
| YSC0024 | MEL5 | MELibiose | Secreted alpha-galactosidase |
| YSC0025 | MEL6 | MELibiose | Secreted alpha-galactosidase |
| YSC0026 | MEL7 | MELibiose | Secreted alpha-galactosidase |
| YSC0027 | MEL8 | MELibiose | Secreted alpha-galactosidase |
| YSC0028 | MEL9 | MELibiose | Secreted alpha-galactosidase |
| YSC0029 | MPR1 | sigMa 1278b gene for L-proline analog resistance | L-azetidine-2-carboxylic acid acetyltransferase |
| YSC0030 | NTS1-1 | Nontranscribed spacer | Nontranscribed region of the rDNA repeat between the 3'-ETS and RDN5 |
| YSC0031 | RF2 | Reading frame 2 | Maturase-like coding sequence downstream of COX3/Q0275 |
| YSC0032 | RTM1 | Resistance to toxic molasses | Member of the lipid-translocating exporter family |
| YSC0033 | STA1 | STArch | Glucoamylase (glucan 1,4-alpha-glucosidase) |
| YSC0034 | STA2 | STArch | Glucoamylase (glucan 1,4-alpha-glucosidase) |
| YSC0035 | STA3 | STArch | Starch hydrolysis |
| YSC0036 | STRP | <i>S. cerevisiae</i> TM30nm related protein | Transmembrane protein with a highly basic C-terminal region |
| YSC0037 | SUC1 | SUCrose | Invertase |
| YSC0038 | SUC3 | SUCrose | Invertase |
| YSC0039 | SUC4 | SUCrose | Invertase |
| YSC0040 | SUC5 | SUCrose | Invertase |
| YSC0041 | SUC7 | SUCrose | Invertase |
| YSC0042 | AWA1 | AWA (Japanese for "foam") | Putative GPI-anchored protein |
| YSC0043 | BIO6 | BIOtin | Putative 7-keto-8-aminopelargonic acid (KAPA) synthetase |
| YSC0044 | KHS1 | Killer of heat sensitive | Thermolabile killer toxin encoded on the right arm of Chromosome V |
| YSC0045 | TAT3 | Tyrosine amino acid transporter | Permease identified in lager brewing yeast strain Weihenstephan Nr.34 |
| YSC0046 | MATA | MATing type | Mating-type locus |
| YSC0047 | BIO1 | BIOtin | Putative pimeloyl-CoA synthetase involved in biotin biosynthesis |
| YSC0048 | BIO8 | BIOtin | Putative KAPA synthetase |
| YSC0049 | BIO7 | BIOtin | Putative pimeloyl-CoA synthetase involved in biotin biosynthesis |
| YSC0050 | ENA6 | Exitus NATru (Latin, "exit sodium") | Plasma membrane sodium-pumping ATPase |
| YSC0051 | XDH1 | Xylitol dehydrogenase | Xylitol dehydrogenase |
| YSC0052 | SUC8 | SUCrose | Invertase |
| YSC0053 | FPG1 | Foam-promoting gene | Putative cell wall mannoprotein involved in foam formation |
| YSC0054 | IMI1 | Irr1-mediated interaction | Protein involved in maintaining mitochondrial integrity and glutathione homeostasis |
| YSC0055 | MPR2 | sigMa1278b gene for L-proline analog resistance | L-azetidine-2-carboxylic acid acetyltransferase |

systematic nomenclature, we have the infrastructure to easily add new, published gene features that are not found in the S288C reference sequence. As founding members of the GO Consortium (<http://geneontology.org/>; Ashburner et al. 2000; Gene Ontology Consortium 2021) and the Alliance of Genome Resources (<https://www.alliancegenome.org/>; Alliance of Genome Resources Consortium 2022), as well as being the authority for *S. cerevisiae*

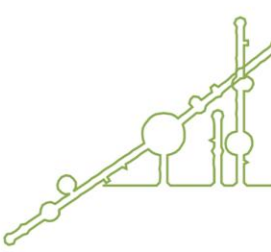
for GenBank, NCBI Gene, UniProtKB, and RNACentral, SGD has a long history of partnering with other databases. SGD will continue to actively participate in these collaborations, including working both with other databases and directly with researchers, to continue to connect fundamental yeast research with other research communities. We validate the accuracy of the external links as they are added to the site, as well as when an error is reported.


A

| | | | | | | |
|---------|----------|---------------|-----------|--------------|------------|------------|
| Summary | Sequence | Gene Ontology | Phenotype | Interactions | Regulation | Literature |
|---------|----------|---------------|-----------|--------------|------------|------------|

SNR42 / YNCK0020C Overview

Standard Name: SNR42¹
Systematic Name: YNCK0020C
SGD ID: SGD:S00006502
Aliases: snR42
Feature Type: snoRNA gene
Description: H/ACA box small nucleolar RNA (snoRNA); guides pseudouridylation of large subunit (LSU) rRNA at position U2975₂₃₄₅
Name: Small Nucleolar RNA¹
Description: URS00005FAA28
RNAcentral ID:
Secondary Structure:



Comparative Info: 

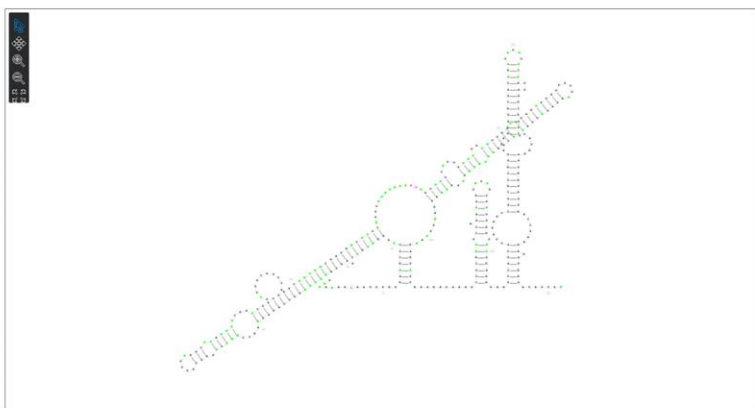
B

| | |
|-------------------|---|
| SNR42 / YNCK0020C | Secondary Structure [®] |
|-------------------|---|

Secondary structure

Generated by R2DT using the *RF01265* template provided by Rfam. [Learn more](#) →

Toggle colours
 Toggle numbers
 Save PNG
 Save SVG
 Copy dot-bracket notation



Colour legend

- Same as the template
- Modified compared to the template. **Tip:** Hover over green nucleotides for more details
- Inserted nucleotides
- Repositioned compared to the template

Tip: Hover over the nucleotides to see nucleotide numbers

Dot-bracket notation

```

.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....
(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....
(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....(((((((.....

```

Fig. 2. Secondary structures from RNAcentral are now available on SGD RNA gene pages. a) Thumbnail view of SNR42/YNCK0020C secondary structure on Locus Summary page and (b) detailed view of secondary structure of SNR42/YNCK0020C on Sequence page.

Additionally, we encourage and welcome community feedback through our helpdesk, sgd-helpdesk@lists.stanford.edu, on any enhancements or new features that we make.

Data availability

All information and materials provided by SGD are available with the Creative Commons Attribution 4.0 International (CC BY 4.0) license. This license allows others to distribute, remix, adapt, and build upon the information or materials, even commercially, as long as credit to the source is provided.

Funding

Research reported in this publication was supported by the National Human Genome Research Institute of the National Institutes of Health under Award Numbers U24HG001315, U24HG010859, and U24HG012212. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Conflicts of interest

None declared.

Literature cited

- Alliance of Genome Resources Consortium. Harmonizing model organism data in the Alliance of Genome Resources. *Genetics*. 2022;220:iyac022. doi:[10.1093/genetics/iyac022](https://doi.org/10.1093/genetics/iyac022).
- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet*. 2000;25(1):25–29. doi:[10.1038/75556](https://doi.org/10.1038/75556).
- Cherry JM. Genetic nomenclature guide. *Saccharomyces cerevisiae*. *Trends Genet*. 1995;11–12.
- Cherry JM, Adler C, Ball C, Chervitz SA, Dwight SS, et al. SGD: *Saccharomyces* genome database. *Nucleic Acids Res*. 1998;26(1):73–79. doi:[10.1093/nar/26.1.73](https://doi.org/10.1093/nar/26.1.73).
- Cherry JM, Hong EL, Amundsen C, Balakrishnan R, Binkley G, et al. *Saccharomyces* genome database: the genomics resource of budding yeast. *Nucleic Acids Res*. 2012;40(D1):D700–D705. doi:[10.1093/nar/gkr1029](https://doi.org/10.1093/nar/gkr1029).
- Cohen N, Kahana A, Schuldiner M. A similarity-based method for predicting enzymatic functions in yeast uncovers a new AMP hydrolase. *J Mol Biol*. 2022;434(7):167478. doi:[10.1016/j.jmb.2022.167478](https://doi.org/10.1016/j.jmb.2022.167478).
- Engel SR, Weng S, Binkley G, Paskov K, Song G, et al. From one to many: expanding the *Saccharomyces cerevisiae* reference genome panel. *Database*. 2016;2016:baw020. doi:[10.1093/database/baw020](https://doi.org/10.1093/database/baw020).
- Engel SR, Wong ED, Nash RS, Aleksander S, Alexander M, et al. New data and collaborations at the *Saccharomyces* genome database: updated reference genome, alleles, and the Alliance of Genome Resources. *Genetics*. 2022;220(4):iyab224. doi:[10.1093/genetics/iyab224](https://doi.org/10.1093/genetics/iyab224).
- Gene Ontology Consortium. The Gene Ontology resource: enriching a Gold mine. *Nucleic Acids Res*. 2021;49(D1):D325–D334. doi:[10.1093/nar/gkaa1113](https://doi.org/10.1093/nar/gkaa1113).
- MacPherson KA, Starr B, Wong ED, Dalusag KS, Hellerstedt ST, et al. Outreach and online training services at the *Saccharomyces* genome database. *Database (Oxford)*. 2017;2017:bax002. doi:[10.1093/database/bax002](https://doi.org/10.1093/database/bax002).
- Meldal BHM, Perfetto L, Combe C, Lubiana T, Ferreira Cavalcante JV, et al. Complex Portal 2022: new curation frontiers. *Nucleic Acids Res*. 2022;50(D1):D578–D586. doi:[10.1093/nar/gkab991](https://doi.org/10.1093/nar/gkab991).
- Nash RS, Weng S, Karra K, Wong ED, Engel SR, et al. Incorporation of a unified protein abundance dataset into the *Saccharomyces* genome database. *Database (Oxford)*. 2020;2020:baaa008. doi:[10.1093/database/baaa008](https://doi.org/10.1093/database/baaa008).
- RNAcentral Consortium. RNAcentral 2021: secondary structure integration, improved sequence search and new member databases. *Nucleic Acids Res*. 2021;49(D1):D212–D220. doi:[10.1093/nar/gkaa921](https://doi.org/10.1093/nar/gkaa921).
- Song G, Balakrishnan R, Binkley G, Costanzo MC, Dalusag K, et al. Integration of new alternative reference strain genome sequences into the *Saccharomyces* genome database. *Database (Oxford)*. 2016;2016:baw074. doi:[10.1093/database/baw074](https://doi.org/10.1093/database/baw074).
- Wong ED, Skrzypek MS, Weng S, Binkley G, Meldal BHM, et al. Integration of macromolecular complex data into the *Saccharomyces* genome database. *Database (Oxford)*. 2019;2019:baz008. doi:[10.1093/database/baz008](https://doi.org/10.1093/database/baz008).

Communicating editor: S. Laceyfield