ORIGINAL ARTICLE



Association of rs2062323 in the TREM1 gene with Alzheimer's disease and cerebrospinal fluid-soluble TREM2

Zuo-Teng Wang^{1,2} | Yan Fu¹ | Shi-Dong Chen³ | Yu-Yuan Huang³ | Ya-Hui Ma¹ | Yan-Jiang Wang⁴ | Lan Tan^{1,2} | Jin-Tai Yu³ |

Correspondence

Jin-Tai Yu, Department of Neurology and Institute of Neurology, Huashan Hospital, Shanghai Medical College, Fudan University, 12th Wulumuqi Zhong Road, Shanghai 200040, China. Email: jintai_yu@fudan.edu.cn

Lan Tan, Department of Neurology, Qingdao Municipal Hospital, Qingdao University, Qingdao 266071, China. Email: dr.tanlan@163.com

Funding information

National Natural Science Foundation of China, Grant/Award Number: 81771148 and 82071201

Abstract

Introduction and aims: Genetic variations play a significant role in determining an individual's AD susceptibility. Research on the connection between AD and TREM1 gene polymorphisms (SNPs) remained lacking. We sought to examine the associations between TREM1 SNPs and AD.

Methods: Based on the 1000 Genomes Project data, linkage disequilibrium (LD) analyses were utilized to screen for candidate SNPs in the TREM1 gene. AD cases (1081) and healthy control subjects (870) were collected and genotyped, and the associations between candidate SNPs and AD risk were analyzed. We explored the associations between target SNP and AD biomarkers. Moreover, 842 individuals from ADNI were selected to verify these results. Linear mixed models were used to estimate associations between the target SNP and longitudinal cognitive changes.

Results: The rs2062323 was identified to be associated with AD risk in the Han population, and rs2062323T carriers had a lower AD risk (co-dominant model: OR, 0.67, 95% CI, 0.51-0.88, p = 0.0037; additive model: OR, 0.82, 95% CI, 0.72-0.94, p = 0.0032). Cerebrospinal fluid (CSF) sTREM2 levels were significantly increased in middle-aged rs2062323T carriers (additive model: $\beta = 0.18$, p = 0.0348). We also found significantly elevated levels of CSF sTREM2 in the ADNI. The rate of cognitive decline slowed down in rs2062323T carriers.

Conclusions: This study is the first to identify significant associations between TREM1 rs2062323 and AD risk. The rs2062323T may be involved in AD by regulating the expression of TREM1, TREML1, TREM2, and sTREM2. The TREM family is expected to be a potential therapeutic target for AD.

KEYWORDS

ADNI, Alzheimer's disease, polymorphism, sTREM2, TREM1

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. CNS Neuroscience & Therapeutics published by John Wiley & Sons Ltd.

¹Department of Neurology, Qingdao Municipal Hospital, Qingdao University, Qingdao, China

²Department of Neurology, Qingdao Municipal Hospital, College of Medicine and Pharmaceutics, Ocean University of China, Qingdao, China

³Department of Neurology and Institute of Neurology, Huashan Hospital, State Key Laboratory of Medical Neurobiology and MOE Frontiers Center for Brain Science, Shanghai Medical College, Fudan University, Shanghai, China

⁴Department of Neurology and Centre for Clinical Neuroscience, Daping Hospital, Third Military Medical University, Chongqing, China

1 | INTRODUCTION

Previous studies indicated that the worldwide prevalence of dementia will increase ninefold by 2050.¹ Alzheimer's disease (AD) is a chronic disease with a long preclinical period (about 20 years), and it is the most common type of dementia.^{2,3} About 95% of all AD patients worldwide belong to the late-onset AD (LOAD) subtype. However, its pathogenesis is unclear, and the development of effective diagnostic and therapeutic tools has been hindered. Genetic studies have confirmed that LOAD is a polygenic and heterogeneous disease. And 60–80% of its risk depends on genetic factors.¹ Therefore, screening and verifying LOAD susceptibility or pathogenic genes using genetic techniques is one of the research priorities in this field.

The triggering receptor expressed on the myeloid cells 1 (TREM1) gene is located in human chromosome 6p21.1, and immediately adjacent to the TREM2 gene. 4,5 Monocytes/macrophages and blood neutrophils are the principal effector cells of human TREM1 in innate responses.5 TREM1 and TREM2 activate myeloid cells by signaling through the adaptor protein DAP12.5 The TREMs family is not only involved in the regulation of microglia phagocytosis and amyloid clearance, but also plays an indispensable role in the neuroinflammatory response of AD.⁶⁻⁸ Replogle and colleagues reported one common variant of the TREM1 gene (rs6910730G) associated with AD pathology and aging-related cognitive decline. One study found a significant decrease in TREM1 expression levels on the surface of peripheral blood monocytes from rs6910730G carriers. 10 Phagocytosis of Aß pathology was also significantly diminished by peripheral blood monocytes from rs6910730G carriers. 10 In addition, the TREM1 can facilitate the microglial phagocytosis of AB, is associated with immune responses in AD. 10,111 These findings suggested that TREM1 can serve as a potential therapeutic target for AD.¹² However, one study investigated the association between rs6910730G and AD (17,008 AD cases and 37,154 controls) using the International Genomics of Alzheimer's Project (IGAP) dataset and did not find any significant association between rs6910730G and AD susceptibility. 13 Therefore, the association between TREM1 gene polymorphism and AD risk needs further investigation. Moreover, it is necessary to further explore the underlying mechanisms between TREM1 and AD susceptibility using human biological samples.

2 | METHODS

2.1 | Study population

A total of 1951 participants who were Han Chinese were gathered from Qingdao Municipal Hospital, Chongqing Daping Hospital, and Huashan Hospital. All eligible participants were required to be of Han Chinese origin and aged from 40 to 90 years old. The AD diagnosis was based on the criteria of the National Institute of Neurological and Communicative Diseases and Stroke and the Alzheimer's Disease and Related Disorders Association (NINCDS-ADRDA).¹⁴

The criteria for exclusion were: (1) family history of genetic diseases, (2) major neurological disorders, epilepsy, infection of the central nervous system, head trauma, or other neurodegenerative diseases (e.g., Parkinson's disease), (3) major psychological disorders, (4) severe systemic diseases (e.g., malignant tumors). Neuropsychological evaluations were performed on each subject using a structured questionnaire and an electronic medical record system. All physicians involved in the assessment have undergone standardized training. Blood samples from all participants were collected for testing. Cerebrospinal fluid (CSF) samples from 503 participants were collected for AD biomarker testing. Written informed consent was obtained from all participants or authorized representatives. The study's design was approved by the Institutional Review Board of all participating institutions (KY2020-1161, Medical Research Ethics-2021-No.51, and 2017-Temporary Audit-No.26-Fast).

In addition, 842 individuals (non-Hispanic white) from Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni. loni.usc.edu) were selected to verify the associations between the target SNP and biomarkers. ADNI's goal is to investigate clinical, imaging, genetic, and biochemical AD biomarkers. Adult participants range in age from 55 to 90. The website http://www.adniinfo.org has comprehensive information. 15-17

2.2 | Construction of linkage disequilibrium (LD) blocks and selection of tag-SNPs

The 1000 Genomes Project is a transnational cooperation project that contains whole-genome sequencing information for 26 populations from the Americas, Europe, South Asia, East Asia, and Africa. We selected 1000 Genomes Project Han Chinese in Bejing Data (reference: Human Genome grch37) and used haploview 4.1 (Hardy–Weinberg equilibrium p-value > 0.05, MAF > 0.1, LD threshold r^2 > 0.8) to analyze the SNPs locus on *TREM1* gene (6:41267385-41286692). Finally, 10 tag-SNPs inside *TREM1* were chosen based on the collected SNP information and LD blocks created by Haploview software (Table S2 and Figure S1). In addition, we also performed LD analysis using the Southern Chinese Han Chinese data (Figure S2) and European data (Figure S3) to verify the selected SNPs.

2.3 | Genotyping

Fasting peripheral venous blood (4–5 mL) was collected using anti-coagulant tubes. The QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany) was utilized to extract DNA from blood samples in the Han Chinese population. Participants' DNA was subjected to genotyping using Infinium Asian Screening Array. ADNI samples were genotyped with the Human 610-Quad BeadChip, Illumina Human Omni Express BeadChip, and Illumina Omni 2.5 M BeadChip. One SNP in *TREM1*, rs2062323, was analyzed for this study. *APOE* and genome-wide genotyping data were obtained from this database

(adni.loni.usc.edu). The APOE genotypes were determined using rs429358 and rs7412.

2.4 Measurement of AD core biomarkers

CSF was collected from participants in a standardized manner via lumbar puncture. 19 Within 2h, the CSF was transported to the lab and centrifuged at 2000g for 10 min. Prior to testing, the thaw/ freeze cycle was limited to 2 cycles. Enzyme-linked immunosorbent assay (ELISA) was used to quantify CSF Aβ42 (INNOTEST®; Fujirebio, Ghent, Belgium), T-tau (INNOTEST®; Fujirebio, Ghent, Belgium), p-tau (INNOTEST®; Fujirebio, Ghent, Belgium), and soluble TREM2 (sTREM2) (Abcam, no. Ab224881). All ELISA measurements are performed by experienced technicians who strictly follow the manufacturer's instructions. Clinical information is not disclosed to testers. Duplicate measurements were taken on samples and standards, and the average of the duplicate measurements was used for statistical analysis. All antibodies and plates are from a single batch to exclude variability between batches. In addition, the intra-batch coefficient of variation (CV) was <5%, and the inter-batch CV was <15%.

Alzheimer's Disease Neuroimaging Initiative provides sTREM2 data based on two platforms. One of the sTREM2 data is based on the MSD platform and has been comprehensively described in previous publications. ^{20–22} The corrected values were used and are available in the ADNI database as variables "MSD_sTREM2corrected". Furthermore, partial sTREM2 was tested at Washington University in St Louis. (developed in-house: WU platform, Piccio group). The corrected values were used and are available in the ADNI database as variables "WU_sTREM2corrected." The raw values are provided as pg/mL. The intraplate CV for CSF sTREM2 was <10%.

2.5 | Statistical analysis

The baseline demographic characteristics (age, gender, APOE4 carriage status, and MMSE score) were compared using ANOVA and chi-square tests. Logistic regression was utilized to analyze the relationship between target SNP and AD risk. Three genetic models, including co-dominant, dominant, and additive models, were applied. All risk analysis models were corrected for age, sex, and APOE4 carriage status. Finally, Bonferroni correction was employed in the setting of multiple comparisons: *p*-value < 0.005 was considered significant.

All continuous variables were normalized using the Box-Cox transformations (R software "car" package) and standardized using the z-scale transformations (R software "scale" package). We used multiple linear regression models to analyze the association between the target SNP and AD CSF biomarkers. The analysis used three genetic models (co-dominant, dominant, and additive). All models were corrected for age, sex, and APOE4 status. In addition,

we investigated the interaction of age, sex, and APOE4 carriage status on the association of rs2062323 with AD CSF biomarkers. Based on the results of the interaction, we further performed a subgroup analysis.

We included only non-Hispanic white individuals in the ADNI replication cohort. The CSF sTREM2 (MSD and WU platform) data were depolarized (mean ± 3-fold standard deviation), and the data were normalized and standardized using the R software "car" and "scale" packages. We used multiple linear regression models to analyze the association between the target SNP and CSF sTREM2. Three genetic models (co-dominant, dominant, and additive) were utilized in the analysis. All models were corrected for age, sex, and APOE4 status. In addition, we investigated the interaction of age, sex, and APOE4 carrying status on the relationship between rs2062323 and sTREM2. Finally, we used linear mixed models to explore the effects of target SNP on longitudinal changes in cognition.

The above data were analyzed using SPSS software (version 22.0) and R software (version 3.6.1). We used the International Genomics of Alzheimer's Project (IGAP) to query the information related to our target SNP. In addition, expression quantitative trait loci (eQTL) analyses were conducted using multiple publicly available datasets (Genotype-Tissue Expression (GTEx), https://gtexportal.org/home/snp/rs2062323) in human brain tissues and the whole blood.²³

3 | RESULTS

3.1 | Characteristics of the study population

We included 870 cognitively normal individuals and 1081AD patients to study the association between candidate SNPs and AD risk. Table 1 shows the demographic characteristics of these Han Chinese subjects. We studied the relationship between target SNP loci and AD biomarkers in 503 cognitively normal Han Chinese populations. The demographic characteristics of these 503 individuals are presented in Table 3. In addition, we included 676 non-demented and 166AD participants in the ADNI replication cohort. The demographic characteristics are shown in Tables S5 and S7.

TABLE 1 Baseline demographic characteristics of participants included.

Characteristic	HC (n = 870)	AD (n = 1081)	p
Age (years, mean \pm SD)	64.41 ± 10.96	65.49 ± 9.95	0.025
Sex (female, %)	455 (52.3)	626 (57.9)	0.013
APOE ε 4 (yes, %)	152 (17.5)	454 (42.0)	< 0.001
MMSE score (mean ± SD)	27.22 ± 2.96	15.43 ± 7.30	< 0.001

Note: ANOVA, non-parametric Kruskal-Wallis H test, and Chi-squared test were used to compare the baseline demographic and clinical characteristics.

Abbreviations: AD, Alzheimer's disease; MMSE, Mini-Mental State Exam; SD, standard deviation.

TABLE 2 Associations between tag-SNPs and the risk of sporadic Alzheimer's disease.

poradic Alzheir				
Tag-SNPs	Genetic modelsa ^a	OR value	95% CI	p-Value ^b
rs1471743	Codominant	0.94	0.75-1.19	0.6136
		1.68	0.80-3.70	0.1769
	Dominant	0.98	0.79-1.23	0.8838
	Additive	1.03	0.84-1.26	0.7975
rs11755124	Codominant	1.18	0.96-1.43	0.1107
		1.42	1.05-1.93	0.0243
	Dominant	1.22	1.01-1.48	0.0368
	Additive	1.19	1.03-1.36	0.0157
rs2062323	Codominant	0.78	0.62-0.97	0.0257
		0.67	0.51-0.88	0.0037
	Dominant	0.74	0.60-0.92	0.0058
	Additive	0.82	0.72-0.94	0.0032
rs4714448	Codominant	0.96	0.76-1.20	0.6986
		2.33	1.13-5.17	0.0279
	Dominant	1.02	0.82-1.28	0.8287
	Additive	1.09	0.89-1.33	0.3948
rs34353646	Codominant	0.98	0.78-1.24	0.8630
		2.34	1.13-5.20	0.0269
	Dominant	1.05	0.84-1.32	0.6656
	Additive	1.11	0.91-1.36	0.2960
rs6914090	Codominant	1.06	0.88-1.29	0.5382
		1.41	0.98-2.05	0.0693
	Dominant	1.11	0.92-1.34	0.2693
	Additive	1.13	0.97-1.31	0.1109
rs2234242	Codominant	1.21	0.99-1.47	0.0550
		0.77	0.53-1.11	0.1613
	Dominant	1.13	0.94-1.36	0.2048
	Additive	1.02	0.88-1.18	0.8106
rs6909482	Codominant	0.97	0.78-1.21	0.7967
		0.80	0.61-1.04	0.0987
	Dominant	0.92	0.75-1.13	0.4064
	Additive	0.90	0.79-1.03	0.1181
rs6910301	Codominant	1.22	1.00-1.48	0.0486
		0.84	0.59-1.19	0.3269
	Dominant	1.14	0.95-1.38	0.1556
	Additive	1.04	0.90-1.20	0.6293
rs6939973	Codominant	0.93	0.74-1.18	0.5529
		2.34	1.09-5.42	0.0353
	Dominant	1.00	0.80-1.25	0.9845
	Additive	1.06	0.87-1.30	0.5475

Note: All models were adjusted by age, sex, and APOE ϵ 4. Abbreviations: CI, confidence interval; OR, odds ratio.

TABLE 3 Baseline demographic characteristics of participants included.

Characteristic	HC (n = 503)
Age (years, mean ± SD)	62.84±9.39
Sex (female, %)	266 (52.9)
Education (years, mean \pm SD)	9.36 ± 4.45
APOE ε 4 (yes, %)	73 (14.5)
MMSE score (mean \pm SD)	27.73 ± 2.38
CSF sTREM2 (pg/mL)	18688.99 ± 6924.02
CSF Aβ42 (pg/mL)	267.04 ± 142.37
CSF tau (pg/mL)	197.96 ± 94.52
CSF ptau (pg/mL)	43.17 ± 13.63

Abbreviations: AD, Alzheimer's disease; ADAS, Alzheimer's Disease Assessment Scale; ADNI_EF, Alzheimer Disease Neuroimaging Initiative-executive function; ADNI_MEM, Alzheimer Disease Neuroimaging Initiative-memory; CSF, cerebrospinal fluid; MMSE, Mini-Mental State Exam; NFL, Neurofilament Light; NPI, Neuropsychiatric Inventory; SD, standard deviation.

3.2 | Allele frequencies and genotype distributions of the tag-SNPs

The genotype distribution of the target SNPs is shown in Tables S1 and S2. We identified a separate genetic locus rs2062323 on the *TREM1* that was significantly associated with AD risk in the Han Chinese population.

3.3 | Association between tag-SNPs within the TREM1 gene and the risk of AD

People carrying the rs2062323T allele had a significantly lower risk of AD (Table 2). *p*-values < 0.005 after Bonferroni correction was considered significant. No statistically significant results were found at other SNP loci. There was no statistical significance for other SNPs.

3.4 | Association between tag-SNP and CSF AD biomarkers

In the total population, no significant associations between the rs2062323 and AD CSF biomarkers were found in either the codominant, dominant, or additive models (Table S3). In addition, interaction analysis revealed an interaction between the rs2062323 and CSF sTREM2 with age (β = -0.0308, p = 0.0436) (Table S4). CSF sTREM2 levels were significantly higher in middle-aged (\le 65 years) Han Chinese carrying rs2062323T compared to non-carriers in age-based subgroup analyses (Figure 1 and Table 4). We found no significant correlations in the elderly population (>65 years) (Table 4).

^aAssuming M represents major allele and m represents minor allele, each genetic model can be described as follows: codominant: M/m vs M/M and m/m vs M/M, two OR values were listed from top to bottom in corresponding columns; dominant: (m/m+M/m) vs M/M; additive: m/m and M/m were weighed 2 and 1 respectively to M/M.

^bThe given *p* values were not corrected by Bonferroni correction. Figures in bold indicate the retained association after Bonferroni correction.

TABLE 4 Multiple regression results for associations of CSF sTREM2 with rs2062323 stratified by age.

	CSF sTRI	CSF sTREM2 (Mid-life)			CSF sTREM2 (Late-life)		
Genetic models ^a	n	Mean ± SD	β, p-value	n	Mean <u>+</u> SD	β, p-value	
Codominant							
rs2062323 ^{CC}	61	17009.38 ± 6511.57	reference	35	19961.54 ± 6749.44	reference	
rs2062323 ^{CT}	134	18367.54 ± 6571.81	0.30, 0.0390	85	20051.28 ± 7439.57	-0.03, 0.900	
rs2062323 ^{TT}	61	18765.97 ± 7184.54	0.36, 0.0340	45	18255.69 ± 6979.32	-0.31, 0.1740	
Dominant							
rs2062323 ^{CC}	61	17009.38 ± 6511.57	reference	35	19961.54 ± 6749.44	reference	
rs2062323 ^{CT/TT}	195	18492.18 ± 6753.31	0.32, 0.0209	130	19429.73 ± 7306.85	-0.13, 0.5110	
Additive							
rs2062323 ^{CC} (0)	61	17009.38 ± 6511.565	0.18, 0.0348	35	19961.54 ± 6749.444	-0.16, 0.1510	
rs2062323 ^{CT} (1)	134	18367.54 ± 6571.808		85	20051.28 ± 7439.574		
rs2062323 ^{TT} (2)	61	18765.97 ± 7184.54		45	18255.69 ± 6979.324		

Note: All models were adjusted by age, sex, APOE ε4, education (years), and MMSE score. Later-life (>65 y) versus Mid-life (≤65 y).

Abbreviations: $A\beta$, amyloid-beta; AD, Alzheimer's disease; AV45, 18F-AV45 amyloid-PET; CSF, cerebrospinal fluid; SD, standard deviation; SNP, Single nucleotide polymorphism; sTREM2, soluble TREM2.

p values that are statistically significant are shown in bold.

TABLE 5 Multiple regressions results for associations of CSF sTREM2 with rs2062323 in ADNI.

	CSF sTRI	CSF sTREM2 (non-demented)			CSF sTREM2 (AD)		
Genetic modelsa ^a	n	Mean ± SD	β, p-value	n	Mean ± SD	β, p-value	
Codominant							
rs2062323 ^{CC}	337	3867.45 ± 1783.10	reference	70	3932.31 ± 1876.94	reference	
rs2062323 ^{CT}	288	4190.77 ± 1963.62	0.19, 0.0136	53	4466.13 ± 2096.72	0.26, 0.1040	
rs2062323 ^{TT}	51	4152.04 ± 2084.20	0.15, 0.3150	12	5234.24 ± 2503.34	0.53, 0.0539	
Dominant							
rs2062323 ^{CC}	337	3867.45 ± 1783.10	reference	70	3932.31 ± 1876.94	reference	
rs2062323 ^{CT/TT}	339	4184.95 ± 1979.09	0.19, 0.0131	65	4607.39 ± 2181.96	0.33, 0.0448	
Additive							
rs2062323 ^{CC} (0)	337	3867.45 ± 1783.10	0.13, 0.0323	70	3932.31 ± 1876.94	0.26, 0.0252	
rs2062323 ^{CT} (1)	288	4190.77 ± 1963.62		53	4466.13 ± 2096.72		
rs2062323 ^{TT} (2)	51	4152.04 ± 2084.20		12	5234.24 ± 2503.34		

Note: All models were adjusted by age, sex, APOE ϵ 4, education (years) and MMSE score. Later-life (>65 years) versus Mid-life (≤65 years). Abbreviations: A β , amyloid-beta; AD, Alzheimer's disease; AV45, 18F-AV45 amyloid-PET; CSF, cerebrospinal fluid; sTREM2, soluble TREM2; SD, standard deviation; SNP, Single nucleotide polymorphism.

p values that are statistically significant are shown in bold.

3.5 | Replication in the ADNI cohort

We found significantly higher CSF sTREM2 (MSD and WU platform) levels in people carrying rs2062323T in the ADNI database. For MSD CSF sTREM2, significant associations existed in the non-demented population (Figure 2A, Figure 2B, and Table 5) and AD patients (Figure 2C,

Figure 2D, and Table 5). For WU CSF sTREM2, significant associations existed both in the non-demented population and AD patients (Table S8).

In addition, we performed interaction analyses in both diagnostic groups. Age, sex, and APOE4 genotype had no significant effect on the relationship between the rs2062323 and CSF sTREM2

^aAssuming M represents major allele and m represents minor allele, each genetic model can be described as follows: codominant: M/m vs M/M and m/m versus M/M, two OR values were listed from top to bottom in corresponding columns; dominant: (m/m + M/m) vs M/M; additive: m/m and M/m were weighed 2 and 1, respectively, to M/M.

^aAssuming M represents major allele and m represents minor allele, each genetic model can be described as follows: codominant: M/m vs M/M and m/m vs M/M, two OR values were listed from top to bottom in corresponding columns; dominant: (m/m + M/m) vs M/M; additive: m/m and M/m were weighed 2 and 1, respectively, to M/M.

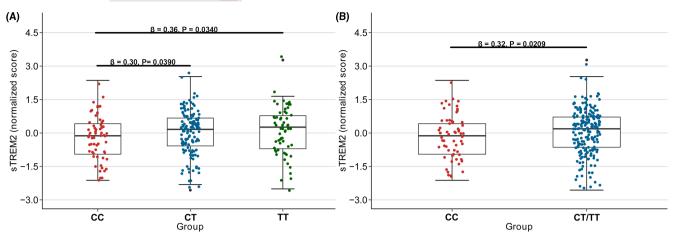


FIGURE 1 Associations of rs2062323 genotypes with CSF sTREM2 in middle-aged (≤65 years) Han Chinese population. We categorized the rs2062323 into four subgroups: CC, CT, TT, and CT/TT. We found that CSF sTREM2 was significantly higher in rs2062323T carriers.

(Tables S6 and S9). We also investigated longitudinal cognitive changes in a non-demented population. The results showed that the rate of cognitive decline was significantly slower in rs2062323T carriers compared to non-carriers (Figure 3).

3.6 | Functional annotation of rs2062323

The IGAP did a meta-analysis using the existing Genome-Wide Association Studies (GWAS) dataset in 2013. The stage 1 meta-analysis included 17,008 AD cases and 37,154 controls of European ancestry. Using these summary results, we identified that the rs2062323 was significantly associated with AD risk (effect allele, T, $\beta = -0.0381$, p = 0.03086). Moreover, multi-tissue eQTL analyses showed that rs2062323 could significantly influence *TREM1*, *TREM2*, and *TREML1* expression in the whole blood and specific brain regions (including basal ganglia, hypothalamus, and hippocampus) (Figures S4–S6). *TREM1*, *TREM2*, and *TREML1* expression levels are elevated in rs2062323T carriers (Figures S4–S6).

4 | DISCUSSION

In conclusion, we identified a genetic variant in *TREM1* is associated with the risk of AD in the Han Chinese population. The rs2062323T carriers had a significantly reduced risk of AD. In addition, the protective effect of rs2062323T on AD also applies to the European population. Cognitive decline was significantly slower in rs2062323T carriers in ADNI. Compared with the non-carriers, the rs2062323T carriers showed significantly increased CSF sTREM2 levels.

Of note, the moderating effects of age on the association between rs2062323 and CSF sTREM2 were only detected in the Han Chinese population. The frequency of the rs2062323T varies between ethnic groups in the 1000 Genomes Project. The frequency is around 50% in the Asian populations, and about 30% in the US and European populations (http://feb2014.archive.ensembl.org/Homo_sapiens/

Variation/Population?db=core;r=6:41239144-41240144;v=rs206 2323;vdb=variation;vf=1633078), and this difference may affect the association between rs2062323 and sTREM2. In addition, associations between rs2062323T and sTREM2 were significant in AD and non-AD populations, suggesting that these associations may not be affected by diagnostic status.

The GTEx databases showed that rs2062323T was associated with high expression of the TREM1, TREM2, and TREML1 genes in parietal brain regions. The TREM1, TREM2, and TREML1 belong to the TREM receptor family and can regulate inflammation by magnifying or inhibiting the toll-like receptor-induced signaling. ²⁵ Human TREM1 was an amplifier of acute inflammation and can link innate and adaptive immunity. TREM family genes were located at 6p21.1 in humans. Previous studies suggested that the 6p21.1 region is closely related to AD.²⁶ Neuroinflammation is one of the critical pathogenic mechanisms of AD and is associated with selective neuronal vulnerability. 27,28 Microglia are immune cells residing in the central nervous system and play a critical regulatory role in maintaining brain homeostasis. Microglia (namely macrophages) are important mediators of neuroinflammation in AD.²⁹ Previous studies have revealed associations between the TREM family genes (mainly TREM1 and TREM2) and AD risk. Given the homology among the TREM family genes, the TREM region likely contains a number of distinct variations and genes that affect various aspects of AD risk. Previous research has demonstrated that the adaptor protein DAP12 facilitates the signaling of TREM1 and TREM2, which in turn activates myeloid cells.⁵

The TREM2 protein is present throughout the brain's white matter and is more abundant in the neocortex and hippocampus, but is absent from the cerebellum.³⁰ Additionally, the TREM2 protein is primarily expressed in myeloid cells, including microglia, dendritic cells generated from monocytes, osteoblasts, and macrophages produced from bone marrow.⁵ Previous studies have shown that mutations and polymorphisms in the *TREM2* gene (rs75932628, rs143332484, rs142232675) are significantly associated with the risk of developing AD.^{31,32} These AD-associated risk polymorphisms have pathogenic consequences due to reduced TREM2 function.³⁰

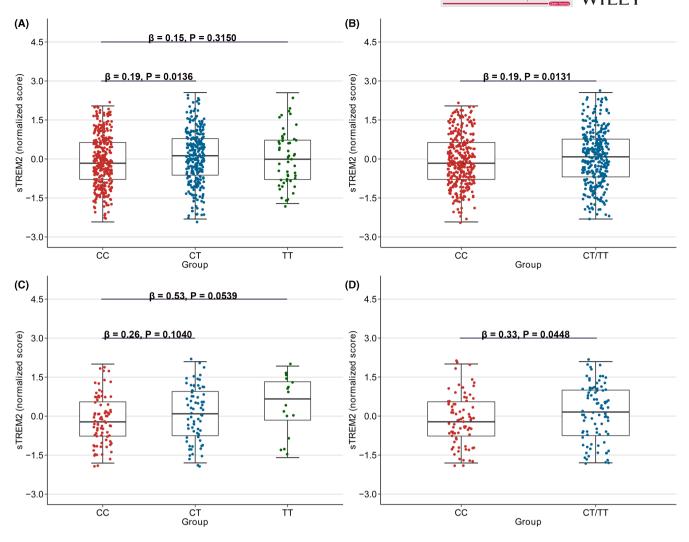


FIGURE 2 Associations of rs2062323 genotypes with CSF sTREM2 in ADNI. We categorized the rs2062323 into four subgroups: CC, CT, TT, and CT/TT. We found that CSF sTREM2 (MSD platform) were significantly higher in CT and CT/TT groups in the non-demented population (A and B). In addition, CSF sTREM2 was significantly higher in CT/TT groups in AD patients (C and D).

In AD patients, microglia processing of A β is an expansion and activation process, with activated microglia gathering near A β plaques and subsequently clearing A β . Thus, TREM2 deficiency can result in a significant decrease in the number of microglia surrounding the plaque. TREM2 does not bind to A β by itself. Notably, TREM2-mediated microglia polarization, processing, and plaque encapsulation depend on the lipidation of A β . TREM2 can be activated by the lipid component of the lipoprotein complex. This specific mechanism gives TREM2 the ability to sense its surroundings and mediate the phagocytosis of dead neurons, myelin, and amyloid plaques. Decreased transcript levels of the *TREM2* gene may affect microglia activation and thus diminish the processing of dead neurons, myelin, and amyloid plaques, and maybe a potential pathway for TREM1 involvement in AD progression.

The sTREM2 is generated either by the proteolysis of TREM2 or by a selective shear of TREM2 lacking a transmembrane domain. The role and importance of sTREM2 are not yet fully understood. It has been shown that sTREM2 may be promoting microglial survival and stimulate the production of innate immune factors as well asTREM2

protein. 34 Suárez-Calvet et al. showed that CSF sTREM2 levels were significantly elevated in the early stages of AD, which may reflect a corresponding change in microglia activation status after neuronal degeneration.²² Heslegrave et al.' study also revealed significantly elevated levels of CSF sTREM2 in AD, which may imply that CSF sTREM2 can be utilized to quantify the activation of glial cells.³⁵ Notably, as AD progresses, the level of sTREM2 varies dynamically. CSF sTREM2 was correlated with markers of neuronal damage (CSF T-tau and CSF p-tau), suggesting that it could be used as a biomarker of neurodegeneration. In addition, sTREM2 was found to co-localize with neurons and plaques in vivo, and sTREM2 may play a chemoattractant role in recruiting microglia to the vicinity of plaques. 36,37 The sTREM2 can also trigger microglia activation and induce an inflammatory response to protect microglia from apoptosis.³⁸ In addition, sTREM2 has a protective effect against amyloid pathology and associated toxicity. The sTREM2 not only reduces a load of amyloid plaques to rescue functional defects in spatial memory, but also enhances microglia proliferation, migration, aggregation, and uptake and degradation of A β in the vicinity of amyloid plaques. ^{36,38} The

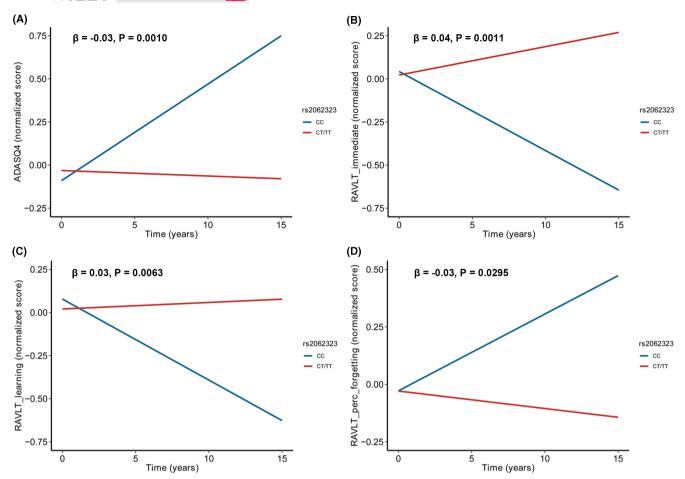


FIGURE 3 Associations of rs2062323 genotypes with longitudinal cognitive changes in non-demented population in ADNI. The rate of cognitive decline was significantly slower in rs2062323T carriers compared to non-carriers. ADAS_Q4: $\beta = -0.03$, p = 0.0010 (A); RAVLT_immediate: $\beta = 0.04$, p = 0.0011 (B); RAVLT_learning: $\beta = 0.03$, p = 0.0063 (C); RAVLT_Percent_Forgetting: $\beta = -0.03$, p = 0.0295 (D).

TREM1 gene intron variant rs2062323T is related to higher levels of sTREM2, suggesting that this variant may lessen the risk of AD through sTREM2-mediated protection against amyloid pathology and associated toxicity.

Previous studies have shown that TREM2 is an important target for drug development. In 2019, Alector/abbvie claimed that they developed a monoclonal antibody (AL002) that specifically binds and activates TREM2 and thus improves AD pathology. In addition, a study from the University of Washington School of Medicine further validated the potential therapeutic effects of AL002c, a variant of AL002. Their results showed that longterm administration of AL002c reduced filamentous plaques and neurodystrophy and alleviated microglia-mediated inflammatory responses, which in turn improving symptoms.³⁹ Given that rs2062323T is associated with high expression of TREM2 and sTREM2 genes, overexpression of rs2062323T by ligands or small molecules may be a potential route for drug discovery. In addition, the variant rs9357347C at 46kb upstream of the TREM2 gene (between TREM2 and TREML2 genes) is associated with elevated expression levels of TREML1 and TREM2 and reduces the risk of developing AD. 40 TREML1 plays a vital role in promoting vascular homeostasis and limiting inflammation. 41,42 The rs2062323T was also associated with higher levels of TREML1 and TREM2 expression, and this locus may have a protective effect on AD by enhancing vascular homeostasis and limiting neuroinflammation.

There are limitations to this study. First, because our study was an observational one, the causal relationship between rs2062323 and CSF sTREM2 needs to be confirmed further. Second, given that the level of sTREM2 varies dynamically, the protective effect of rs2062323T may only exist in a particular age group. Third, our AD diagnosis was made based on the NINCDS-ADRDA criteria, and the results may be more accurate if combined with biomarkers. Fourth, this study was conducted with modest sample sizes, and the generalizability of our conclusions might be restricted by the sources of studied populations. Further large-scale community-based longitudinal studies are warranted to validate these associations.

5 | CONCLUSION

In conclusion, *TREM1* rs2062323 is an AD-protective SNP that is linked to higher levels of CSF sTREM2. The rs2062323 may be involved in AD by regulating the expression of TREM1, TREML1,

TREM2, and sTREM2. In future studies, using systems biology concepts to clarify the associations among TREM family members (TREM1, TREM2, and TREML1) in AD vulnerability may help to develop more effective therapeutic strategies.

AUTHOR CONTRIBUTIONS

JTY and LT had full access to all of the data in the study and take responsibility for the integrity of the data and accuracy of the data analysis. Concept and design: JTY and LT. Acquisition, analysis, or interpretation of data: all authors. Drafting of the manuscript: ZTW, YF, SDC, YYH, and YHM. Critical revision of the manuscript for important intellectual content: All authors. Statistical analysis: ZTW and YF. Obtained funding: JTY and LT. Administrative, technical, or material support: YJW, JTY, and LT. All authors read and approved the final manuscript.

ACKNOWLEDGMENTS

This study was supported by grants from the National Natural Science Foundation of China (81771148, 82071201). Shanghai Municipal Science and Technology Major Project (No.2018SHZDZX01), Research Start-up Fund of Huashan Hospital (2022QD002), Excellence 2025 Talent Cultivation Program at Fudan University (3030277001), and ZHANGJIANG LAB, Tianqiao and Chrissy Chen Institute, and the State Key Laboratory of Neurobiology and Frontiers Center for Brain Science of Ministry of Education, Fudan University.

Data collection and sharing for the ADNI data section was funded by the National Institutes of Health (Grant No. U01 AG024904) and Department of Defense ADNI (Grant No. W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; Eurolmmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

FUNDING INFORMATION

This study was supported by grants from the National Natural Science Foundation of China (81771148, 82071201).

CONFLICT OF INTEREST STATEMENT

The authors declare that they have no conflict of interest, financial, or otherwise.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ETHICS APPROVAL

All data sources used in this study received approval from an ethics standards committee on human experimentation and the procedures used in this study adhere to the tenets of the Declaration of Helsinki. Written consent for genetic screening were obtained from all participants or their legal representatives. Their confidentiality was preserved according to the guidelines for studies of human subjects. About ADNI, data involved in the study came from the publicly open Alzheimer's disease neuroimaging initiative (ADNI) database.

CONSENT TO PARTICIPATE

Informed written consent was obtained for all participants.

ORCID

Yan-Jiang Wang https://orcid.org/0000-0002-6227-6112 *Jin-Tai Yu* https://orcid.org/0000-0002-7686-0547

REFERENCES

- Scheltens P, De Strooper B, Kivipelto M, et al. Alzheimer's disease. Lancet. 2021;397(10284):1577-1590.
- Isaacson RS, Ganzer CA, Hristov H, et al. The clinical practice of risk reduction for Alzheimer's disease: a precision medicine approach. Alzheimers Dement. 2018;14(12):1663-1673.
- Mortamais M, Ash JA, Harrison J, et al. Detecting cognitive changes in preclinical Alzheimer's disease: a review of its feasibility. Alzheimers Dement. 2017;13(4):468-492.
- Carmona S, Zahs K, Wu E, Dakin K, Bras J, Guerreiro R. The role of TREM2 in Alzheimer's disease and other neurodegenerative disorders. *Lancet Neurol.* 2018;17(8):721-730.
- Colonna M. TREMs in the immune system and beyond. Nat Rev Immunol. 2003;3(6):445-453.
- Wang Y, Cella M, Mallinson K, et al. TREM2 lipid sensing sustains the microglial response in an Alzheimer's disease model. *Cell*. 2015;160(6):1061-1071.
- Roussos P, Katsel P, Fam P, Tan W, Purohit DP, Haroutunian V. The triggering receptor expressed on myeloid cells 2 (TREM2) is associated with enhanced inflammation, neuropathological lesions and increased risk for Alzheimer's dementia. Alzheimers Dement. 2015;11(10):1163-1170.
- Jiang T, Yu JT, Zhu XC, et al. Triggering receptor expressed on myeloid cells 2 knockdown exacerbates aging-related neuroinflammation and cognitive deficiency in senescence-accelerated mouse prone 8 mice. Neurobiol Aging. 2014;35(6):1243-1251.
- Replogle JM, Chan G, White CC, et al. A TREM1 variant alters the accumulation of Alzheimer-related amyloid pathology. *Ann Neurol*. 2015;77(3):469-477.

- Jiang T, Zhang YD, Gao Q, et al. TREM1 facilitates microglial phagocytosis of amyloid beta. Acta Neuropathol. 2016;132(5):667-683.
- Sao T, Yoshino Y, Yamazaki K, et al. TREM1 mRNA expression in leukocytes and cognitive function in Japanese patients with Alzheimer's disease. J Alzheimers Dis. 2018;64(4):1275-1284.
- Saadipour K. TREM1: a potential therapeutic target for Alzheimer's disease. Neurotox Res. 2017;32(1):14-16.
- 13. Liu G. No association of TREM1 rs6910730 and TREM2 rs7759295 with Alzheimer disease. *Ann Neurol*. 2015;78(4):659.
- McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA work group under the auspices of Department of Health and Human Services Task Force on Alzheimer's disease. Neurology. 1984;34(7):939-944.
- Petersen RC, Aisen PS, Beckett LA, et al. Alzheimer's disease neuroimaging initiative (ADNI): clinical characterization. *Neurology*. 2010:74(3):201-209.
- Trojanowski JQ, Vandeerstichele H, Korecka M, et al. Update on the biomarker core of the Alzheimer's disease neuroimaging initiative subjects. Alzheimers Dement. 2010;6(3):230-238.
- 17. Weiner MW, Aisen PS, Jack CR Jr, et al. The Alzheimer's disease neuroimaging initiative: progress report and future plans. *Alzheimers Dement*. 2010;6(3):202-211 e207.
- Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics*. 2005;21(2):263-265.
- del Campo M, Mollenhauer B, Bertolotto A, et al. Recommendations to standardize preanalytical confounding factors in Alzheimer's and Parkinson's disease cerebrospinal fluid biomarkers: an update. Biomark Med. 2012;6(4):419-430.
- Kleinberger G, Yamanishi Y, Suarez-Calvet M, et al. TREM2 mutations implicated in neurodegeneration impair cell surface transport and phagocytosis. Sci Transl Med. 2014;6(243):243ra286.
- Suarez-Calvet M, Araque Caballero MA, Kleinberger G, et al. Early changes in CSF sTREM2 in dominantly inherited Alzheimer's disease occur after amyloid deposition and neuronal injury. Sci Transl Med. 2016;8(369):369ra178.
- Suarez-Calvet M, Kleinberger G, Araque Caballero MA, et al. sTREM2 cerebrospinal fluid levels are a potential biomarker for microglia activity in early-stage Alzheimer's disease and associate with neuronal injury markers. EMBO Mol Med. 2016;8(5):466-476.
- 23. Consortium GT. Human genomics. The genotype-tissue expression (GTEx) pilot analysis: multitissue gene regulation in humans. *Science*. 2015;348(6235):648-660.
- Lambert JC, Ibrahim-Verbaas CA, Harold D, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nat Genet. 2013;45(12):1452-1458.
- Ford JW, McVicar DW. TREM and TREM-like receptors in inflammation and disease. Curr Opin Immunol. 2009;21(1):38-46.
- Cruchaga C, Kauwe JS, Harari O, et al. GWAS of cerebrospinal fluid tau levels identifies risk variants for Alzheimer's disease. *Neuron*. 2013;78(2):256-268.
- 27. Leng F, Edison P. Neuroinflammation and microglial activation in Alzheimer disease: where do we go from here? *Nat Rev Neurol*. 2021;17(3):157-172.
- Wang ZT, Zhang C, Wang YJ, Dong Q, Tan L, Yu JT. Selective neuronal vulnerability in Alzheimer's disease. Ageing Res Rev. 2020:62:101114.

- Lue LF, Beach TG, Walker DG. Alzheimer's disease research using human microglia. Cell. 2019;8(8):838.
- Guerreiro R, Wojtas A, Bras J, et al. TREM2 variants in Alzheimer's disease. N Engl J Med. 2013;368(2):117-127.
- 31. Jonsson T, Stefansson H, Steinberg S, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med*. 2013;368(2):107-116.
- 32. Jin SC, Benitez BA, Karch CM, et al. Coding variants in TREM2 increase risk for Alzheimer's disease. *Hum Mol Genet*. 2014:23(21):5838-5846.
- 33. Ulrich JD, Ulland TK, Colonna M, Holtzman DM. Elucidating the role of TREM2 in Alzheimer's disease. *Neuron*. 2017;94(2):237-248.
- Bekris LM, Khrestian M, Dyne E, et al. Soluble TREM2 and biomarkers of central and peripheral inflammation in neurodegenerative disease. J Neuroimmunol. 2018;319:19-27.
- 35. Heslegrave A, Heywood W, Paterson R, et al. Increased cerebrospinal fluid soluble TREM2 concentration in Alzheimer's disease. *Mol Neurodegener*. 2016;11:3.
- Zhong L, Xu Y, Zhuo R, et al. Soluble TREM2 ameliorates pathological phenotypes by modulating microglial functions in an Alzheimer's disease model. Nat Commun. 2019;10(1):1365.
- Song WM, Joshita S, Zhou Y, Ulland TK, Gilfillan S, Colonna M. Humanized TREM2 mice reveal microglia-intrinsic and -extrinsic effects of R47H polymorphism. J Exp Med. 2018;215(3):745-760.
- Zhong L, Chen XF, Wang T, et al. Soluble TREM2 induces inflammatory responses and enhances microglial survival. J Exp Med. 2017;214(3):597-607.
- Wang S, Mustafa M, Yuede CM, et al. Anti-human TREM2 induces microglia proliferation and reduces pathology in an Alzheimer's disease model. J Exp Med. 2020;217(9):e20200785.
- Carrasquillo MM, Allen M, Burgess JD, et al. A candidate regulatory variant at the TREM gene cluster associates with decreased Alzheimer's disease risk and increased TREML1 and TREM2 brain gene expression. Alzheimers Dement. 2017;13(6):663-673.
- Washington AV, Gibot S, Acevedo I, et al. TREM-like transcript-1 protects against inflammation-associated hemorrhage by facilitating platelet aggregation in mice and humans. *J Clin Invest*. 2009;119(6):1489-1501.
- Derive M, Bouazza Y, Sennoun N, et al. Soluble TREM-like transcript-1 regulates leukocyte activation and controls microbial sepsis. J Immunol. 2012;188(11):5585-5592.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Wang Z-T, Fu Y, Chen S-D, et al. Association of rs2062323 in the *TREM1* gene with Alzheimer's disease and cerebrospinal fluid-soluble TREM2. *CNS Neurosci Ther.* 2023;29:1657-1666. doi:10.1111/cns.14129