



Effects of Hydrosalpinx on Endometrial Implantation Failures: Evaluating Salpingectomy in Women Undergoing in vitro fertilization

Efeitos do Hydrosalpinx no Falho de Implantação Endometrial: Avaliar a salpingectomia nas mulheres em curso de Fertilização in vitro

Antonio Palagiano¹ Mauro Cozzolino^{2,3,4} Filippo Maria Ubaldi⁵ Chiara Palagiano⁶
Maria Elisabetta Coccia⁷

¹ Department of General and Specialized Surgery for Women and Children, Università degli Studi della Campania Luigi Vanvitelli, Napoli, Italy

² Department of Obstetrics, Gynecology and Reproductive Sciences, Yale School of Medicine, New Haven, Connecticut, United States

³ Universidad Rey Juan Carlos, Madrid, Spain

⁴ IVIRMA, IVI Foundation, Valencia, Spain

⁵ GENERA Centre for Reproductive Medicine, Clinica Valle Giulia, Roma, Italy

⁶ Università Campus Bio-Medico, Romq, Italy

⁷ Department of Biomedical, Experimental and Clinical Sciences, Division of Obstetrics and Gynecology, Università degli Studi di Firenze, Azienda Ospedaliero Universitaria Careggi, Firenze, Italy

Address for correspondence Mauro Cozzolino, MD, IVIRMA, IVI Foundation, Avenida Fernando Abril Martorell, 106 - Torre A, Planta 1ª, 46026, Valencia, Spain (e-mail: mauro.cozzolino@ivirma.com).

Rev Bras Ginecol Obstet 2021;43(4):304–310.

Abstract

Hydrosalpinx is a disease characterized by the obstruction of the salpinx, with progressive accumulation in the shape of a fluid-filled sac at the distal part of the tuba uterina, and closed to the ovary. Women with hydrosalpinges have lower implantation and pregnancy rates due to a combination of mechanical and chemical factors thought to disrupt the endometrial environment. Evidence suggests that the presence of hydrosalpinx reduces the rate of pregnancy with assisted reproductive technology. The main aim of the present is review to make an overview of the possible effects of hydrosalpinx on in vitro fertilization (IVF). We conducted a literature search on the PubMed, Ovid MEDLINE, and Google Scholar data bases regarding hydrosalpinx and IVF outcomes. Hydrosalpinx probably has a direct toxic effect on sperm motility and on the embryos. In addition, the increasing liquid inside the salpinges could alter the mechanisms of endometrial receptivity. The window of endometrial receptivity is essential in the implantation of blastocysts, and it triggers multiple reactions arising from the endometrium as well as the blastocysts. Hydrosalpinx could influence the expression of *homeobox A10* (*HOXA10*) gene, which plays an essential role in directing embryonic development and implantation. Salpingectomy restores the endometrial expression of *HOXA10*; therefore, it may be one mechanism by which tubal

Keywords

- ▶ hydrosalpinx
- ▶ salpingectomy
- ▶ endometrial receptivity
- ▶ hydrosalpinx fluid

received

June 17, 2020

accepted

October 23, 2020

published online

February 18, 2021

DOI <https://doi.org/10.1055/s-0040-1722155>.
ISSN 0100-7203.

© 2021. Federação Brasileira de Ginecologia e Obstetrícia. All rights reserved.

This is an open access article published by Thieme under the terms of the Creative Commons Attribution License, permitting unrestricted use, distribution, and reproduction so long as the original work is properly cited. (<https://creativecommons.org/licenses/by/4.0/>)

Thieme Revinter Publicações Ltda., Rua do Matoso 170, Rio de Janeiro, RJ, CEP 20270-135, Brazil

removal could result in improved implantation rates in IVF. In addition, salpingectomy does not affect the ovarian response, nor reduces the antral follicle count. Further studies are needed to establish the therapeutic value of fluid aspiration under ultrasonographic guidance, during or after oocyte retrieval, in terms of pregnancy rate and ongoing pregnancy.

Introduction

Hydrosalpinx is the accumulation of fluid within the ampullar lumen as a result of occlusion of the infundibulum. It is a common condition among women of reproductive age, and it is related to diminished pregnancy rates (PRs)¹⁻³ with assisted reproductive technology (ART). The incidence of hydrosalpinx diagnosed by ultrasound among infertile women is between 10% and 13%, and it may increase to 30% with the use of hysterosalpingography or laparoscopy.⁴

The diagnosis of hydrosalpinx is commonly made by ultrasonography (US) or hysterosalpingography, the latter representing the gold standard. However, under certain conditions, US is unable to detect small amounts of fluid inside the oviduct, which could consequently grow during ovarian stimulation, producing a deleterious effect on embryo implantation.^{5,6} The exact mechanisms behind the adverse effects observed on pregnancy are not yet well understood. Hydrosalpinx fluid (HF) is known to be embryotoxic, and contains growth-factor inhibitors. Moreover, it seems that the liquid contained in the hydrosalpinx could reduce sperm motility and the velocity of motile spermatozoa following 24 hours of incubation.⁷

The fallopian tube is distended during exogenous hormone administration, and this emphasizes that steroids affect the endoluminal ampullar secretion.⁶ The aim of the present review was to evaluate the effects of hydrosalpinx on reproductive outcomes: it has a negative effect either on the embryo or the endometrium. In addition, we assessed the different surgical managements of hydrosalpinx.

Methods

The present review included studies evaluating the effect of hydrosalpinx on IVF outcomes. The literature search was performed on the following databases: MEDLINE, EMBASE, Global Health, The Cochrane Library (Cochrane Database of Systematic Reviews, Cochrane Central Register of Controlled Trials, Cochrane Methodology Register), Health Technology Assessment Database, and Web of Science; we searched the entirety of those databases for studies published until July 2020. The literature search was conducted using the combination of the following Medical Subject Heading (MeSH) terms and any relevant keywords in different orders: "hydrosalpinx," "hydrosalpinges," "hydrosalpinx fluid," "embryo toxicity," "IVF," "ART outcomes," "implantation failure," "embryotoxic," "reproductive outcomes," "live birth," and "clinical pregnancy rate." The search was performed in English. The reference lists of the included studies were also

manually checked to look for studies that were not found in the electronic literature search.

Results

Effects of Hydrosalpinx on Reproductive Outcome

The hydrosalpinges are associated with the presence of fluid in the uterine cavity, modifications in endometrial blood flow, the leukemia inhibitory factor, the inflammatory response, and *HOXA10* expression,^{8,9} in the joint, which could explain the mechanism leading to decreasing PRs and increasing miscarriage rates in women undergoing ART. Many retrospective studies have shown an impaired outcome of IVF in the presence of hydrosalpinx, and the meta-analyses have demonstrated that the probability of achieving pregnancy in the presence of hydrosalpinx halved.¹⁰ The miscarriage rate is increased in women with hydrosalpinx.¹¹⁻¹³ Theoretically, a good embryo could be implanted anywhere, as happens in the case of hysterectomized women,¹⁴ however, perfect integrity of the endometrial cavity is required to maximize embryo implantation.

In recent years, the increased success rates of treatments using ART have as key factors embryo quality, endometrial receptivity, and embryo transfer as key factors.¹⁵

The PR in most cases of IVF is below 40%; therefore, undiscovered pathologies in the uterus, endometrium and fallopian tubes could play a crucial role in implantation failure. Furthermore, some conditions, such as the presence of fluid within the uterine cavity,^{16,17} could have a detrimental effect on endometrial receptivity. In particular, HF plays a role in reducing the PR in in-vitro fertilization programs.¹¹ Kassabji et al.¹⁸ Demonstrated the deleterious effects of HF on IVF outcome, and, a few years later, a meta-analysis by Camus et al.³ demonstrated that the probability of achieving pregnancy in the presence of hydrosalpinx is halved, whereas the incidence of miscarriage is doubled. It is not well-known how the HF affects the implantation. It may be due to embryotoxicity, or to a direct effect on the endometrium, resulting in HF formation, or, finally, poor endometrial receptivity caused by alterations in the expression of markers or key molecules in the endometrium.

Direct Embryo Toxicity

Beyler et al.¹⁹ suggested a potential deleterious effect of the HF on in-utero embryo development. They evaluated in-vitro mouse embryo development in the presence of HF collected from 10 infertile women. The HF had a detrimental effect on the development of mouse embryos. Therefore, the poor PR in women with hydrosalpinx could be explained, in part, by

the reflux of a lipophilic embryotoxic factor into the uterine cavity.¹⁹ The HF may contain toxins that are potentially teratogenic. Morphological scores and the diameter of the yolk sac were significantly lower in embryos exposed to HF.²⁰

Roberts et al.²¹ evaluated the effects of different concentrations of HF, with and without lactate supplementation, on the preimplantation development and implantation of murine models. The rates of development of blastocysts were respectively of 45%, 55%, 12.5% and 17.5% depending on the concentration of HF, and, with lactate supplementation, the rates were of 35.0%, 52.5%, 12.5%; without lactate supplementation, the rate was of 5.0%, while in the control group, it was of 63.8%.

The implantation rates for the 0.1% and 1.0% groups with lactate supplementation were of 43.0% and 25.0% respectively, and the rates for the groups with lactate supplementation were of 50.6% and 61.8%, while in the control group the implantation rate was of 65.5%. Hence, these results made it possible to hypothesize that the HF has a concentration-dependent decline in in-vitro murine embryo development, with a minimal effect on the implantation rates. Furthermore, lactate supplementation did not significantly influence the implantation rate at any HF concentration.

However, the embryotoxic effect of the HF has not been demonstrated in human embryos.^{22,23} The analysis of the HF demonstrated the absence of bacteria, normal electrolyte concentrations with lower amounts of total protein and albumin, with a composition that does not differ from that of the normal tubal fluid.²² All fertilized eggs showed physiological segmentation, and the embryonal development rate was not impaired by the HF. Even the granulosa cells incubated in HF showed the same steroidogenic capacity as those incubated in its absence.

In the same way, Strandell et al.²³ showed no negative effect on human embryonic growth: donated frozen embryos were incubated at concentrations of 50% and 100% of HF, but the HF had effects on blastocyst development.

In all likelihood, the low implantation rate in IVF patients with hydrosalpinx may not be due to an embryotoxic effect, and the differences regarding mouse embryos may be related to inter-species differences. Furthermore, the HF has a negative effect on sperm motility and survival after 24 hours of incubation, and the motility reduction depends on the concentration of HF.⁷ Hydrosalpinx epithelial cells may be producing a fluid milieu which is hostile to sperm and early embryo development.²⁴

Effects of Fluid Formation

Hydrosalpinx is associated with the presence of fluid within the endometrial cavity that may have a direct effect on the endometrium. The origin of the fluid is a point of controversy, but a leakage from the ampulla to the uterine cavity is the most accepted theory. Some authors have suggested that the reflux could have a “flushing” effect on the embryos, attributing to a mechanical interference with the implantation failure.^{25–27} Before the invasion of the trophoblast, there is a progressive reabsorption of intraluminal uterine allowing the interactions between the embryo and the epithelial.^{28,29}

The HF has a direct, cytotoxic effect on developing mouse embryos, but not on human embryos,^{19,21–23} in whom it reduces the implantation and pregnancy rates. The peaks of reabsorption of intraluminal uterine fluid occur at the expected time of implantation in rodents,^{30,31} and are controlled by ovarian hormone secretions, local ion and water channels. The mechanisms underlying the fluid formation in the hydrosalpinx are poorly-known or not consistent.³²

The involvement of epithelial transporters and ion channels of the fallopian tube is a possibility, particularly the cystic fibrosis transmembrane conductance regulator (CFTR). In pelvic inflammatory disease (PID), which is mainly caused by infection by *Chlamydia trachomatis*, CFTR-mediated events may culminate in hydrosalpinx formation,^{33,34} and there is evidence³⁵ available that shows that ~ 20% of woman affected by chlamydial lower-genital-tract infection develop PID. A shift between the chlamydial and endometrial infections was reported in a study by Jones et al.,³⁶ in which *C. trachomatis* was recovered from uterine and fallopian tubes in women affected by acute salpingitis.³⁷ In both cases, there was a higher incidence of miscarriage rates³⁸ and higher levels of chlamydial heat shock proteins (HSPs), which seem to be responsible for the local immune response followed by an inflammatory reaction and poor embryo implantation.³⁹ Spandorfer et al.⁴⁰ demonstrated that HSP antibodies were more prevalent in women with hydrosalpinx and tubal occlusion than in patients with male factor infertility.

Furthermore, Witkin et al.³⁹ reported that the immunoglobulin A (IgA) antibodies against Chlamydia detected in the uterine cervix dramatically correlated to lower PRs in women undergoing IVF and embryo transfer (IVF-ET).

Although the connection between PID, Chlamydia infection and hydrosalpinx is largely recognized, the sequence of events that leads to the formation of fluid is still debated and not clearly demonstrated (► Fig. 1).

It is known that the movement of water in the oviducts, which does not depend on active transport, may represent

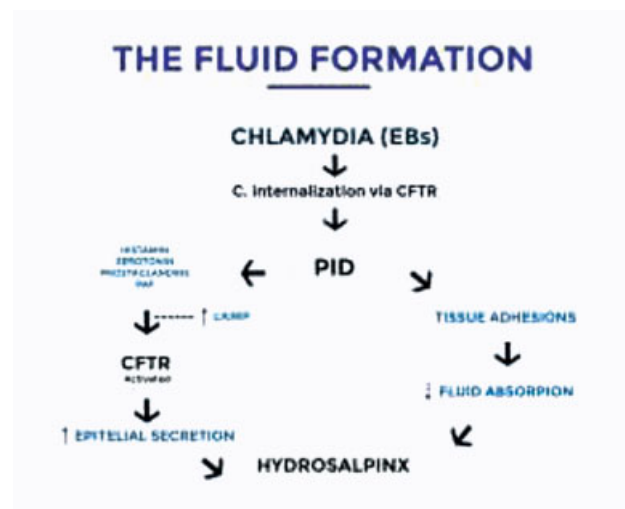


Fig. 1 Mechanisms responsible for the formation of the hydrosalpinx fluid.

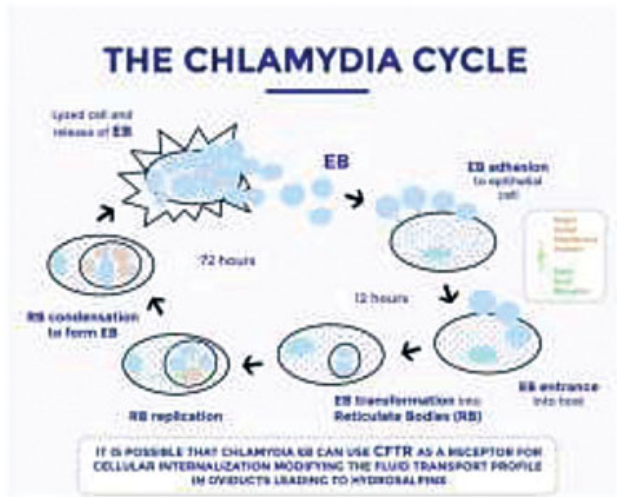


Fig. 2 Mechanisms through which the chlamydia cycle occurs within infected cells.

the response to osmotic gradients due to the transportation of ions.⁴¹ In particular, chloride ions move actively in the direction of the oviduct lumen, from the tubal serosa to the tubal mucosa,^{42,43} and generate a transepithelial potential difference across cultured cells.^{44,45}

The inner mucosa of the fallopian tubes is lined with ciliated columnar epithelial cells and peg cells (non-ciliated secretory cells). The loss of membrane polarity could generate HF after PID. Sodium/hydrogen exchangers (NHEs) and anion $\text{Cl}^-/\text{HCO}_3^-$ (AE), well-known plasma membrane transporters in human epithelial cells, are involved in this process. The CFTR has a well-recognized function as a cyclic adenosine 3',5'-monophosphate (cAMP)-activated chloride channel. The increases in the levels of cAMP are followed by an increase in the transepithelial potential difference in current, leading to enhancements in fluid secretion.⁴⁶ The CFTR is a transporter of chloride ions; furthermore, it acts as a receptor for some bacteria in the epithelial cells. *Pseudomonas aeruginosa* binds to the CFTR in pulmonary epithelial cells, whereas *Salmonella typhi* and *Cholera vibro* bind to the CFTR in gastrointestinal cells, increasing the serosa-to-mucosa fluxes of sodium and chloride, with subsequent diarrhea.^{47,48}

It is possible that Chlamydia elementary bodies (EBs) can use the CFTR as a receptor for cellular internalization in female infections of the reproductive, modifying the profile of the fluid transport in the oviducts, leading to hydrosalpinx (► Fig. 2). Downing et al.⁴⁹ studied the effects of inflammatory mediators, which may increase microvascular permeability, leading to fluid formation by the tubal epithelium. Histamine, serotonin, prostaglandins and the platelet-activating factor (PAF) are involved in this process. In particular, histamine, in cultured epithelial cells, influences ion movements and tubal secretions of fluid. In the presence of fimbrial adhesions, which are the consequence of infection and inflammatory disease, the epithelial secretion is blocked within the oviduct, leading to the formation of hydrosalpinx.⁴⁹

The tumor necrosis factor (TNF) may be involved in the pathogenesis of acute tubal injury associated with Chlamydia

infection. In fact, in women who are culture-positive for *C. trachomatis*, the TNF was identified only in those fallopian tubes with visual evidence of the disease, while the fluids obtained from morphologically normal tubes were negative. Therefore, localized cell-mediated activation of the immune system, which is identified by TNF production, appears to be a typical component of salpingitis.³⁸

Endometrial integrins may also modify endometrial receptivity. Meyer et al.⁵⁰ studied 103 patients affected by hydrosalpinx who underwent endometrial biopsies during the window of implantation, searching for three integrins as markers of endometrial receptivity: $\alpha_1\beta_1$, $\alpha_4\beta_1$, and $\alpha_v\beta_3$. Women with hydrosalpinx expressed significantly lower levels of $\alpha_v\beta_3$ compared with the control group, while there was no difference in the expression of the other two integrins. From a histological point of view, the endometrium with an absence of expression of $\alpha_v\beta_3$ was significantly out of phase compared with the control group. Furthermore, women biopsied after hydrosalpinx removal demonstrated an increase in $\alpha_v\beta_3$ expression in 70% of the cases. The present study confirms that a lower implantation rate in hydrosalpinx may be caused by poor endometrial receptivity, and that the tubal surgical removal in these patients can improve the IVF-ET outcome.

Poor Endometrial Receptivity

The window of endometrial receptivity is essential in the implantation of blastocysts, and it triggers multiple reactions from the endometrium as well as the blastocysts.⁵¹ The receptive endometrium and stimulated blastocysts function simultaneously to achieve blastocyst implantation, leading to pregnancy.⁵² Poor endometrial receptivity may be due to an abnormal expression of key molecules in the endometrium, cytokines, steroid hormones, peptides, growth factors and enzymes, which are essential for implantation.⁵³ The dialogue between the endometrium and the embryo is mediated by the expression of certain cytokines and other substances during the implantation window. The *HOXA10* is a protein-coding gene that plays an essential role in directing embryonic development and implantation.^{54,55} Its expression is necessary for endometrial receptivity.⁵⁵⁻⁵⁷

A targeted disruption or a target mutation of the *HOXA10* gene in mice causes implantation failure, but the same mice produce good embryos that normally implant in a wild-type surrogate. In contrast, wild-type embryos fail to implant in *HOXA10* deficient mice.⁵⁸ The *HOXA10* is expressed in the uterus during the menstrual cycle, and its expression increases during the mid-secretory phase of the menstrual cycle, corresponding to the time of implantation. In this "window," the levels of expression of the *HOXA10* increase dramatically in the glands, and even in the stromal cells.⁵⁹⁻⁶² In cultured endometrial cells, the expression of *HOXA10* was stimulated by estrogen or progesterone with a concentration-dependent dose, for progesterone.⁵⁶ Reduction of maternal *HOXA10* results in proportionally diminished windows of implantation. Furthermore, altered levels of *HOXA10* expression regulate the degree of endometrial receptivity.⁶³

The *HOXA10* gene influences embryo implantation through several mechanisms, such as pinopod development,⁶⁴ leukocyte infiltration, and stromal decidualization.^{62,65}

A defect in the endometrial expression of the *HOXA10* gene is present in several aberrations and pathologies involving the uterus, ovaries and salpinges, such as endometriosis, submucosal uterine leiomyomas, polycystic ovary syndrome (PCOS) and hydrosalpinges. In fact, optimal anatomy and physiology are necessary for normal implantation and placentation.⁸

The expression of the *HOXA10* was significantly lower in infertile patients with hydrosalpinges compared with controls with fertile women. Salpingectomy resulted in a statistically significant 15-fold increase in endometrial *HOXA10* expression.⁶⁶

The present research demonstrates that the decrease in *HOXA10* expression in response to HF may constitute a potential molecular mechanism for diminished implantation rates. Salpingectomy restores endometrial *HOXA10* expression, resulting in improved implantation rates in IVF-ET.

Management of Hydrosalpinx

Hydrosalpinx has a negative effect on the outcome of IVF; therefore, any surgical intervention performed on the damaged tube should increase the chances of a successful outcome.⁶⁷

Johnson et al.⁶⁸ published a complete review in 2010 that compared the surgical treatments on the fallopian tubes, prior to IVF, in women with hydrosalpinx. The authors included five randomized controlled studies involving 646 women, regarding salpingectomy, laparoscopic tubal occlusion, ultrasonographic aspiration of HF before or during the oocyte collection and, finally, salpingostomy. The results showed that the odds of ongoing pregnancy and clinical pregnancy increased after laparoscopic salpingectomy, prior to IVF, for patients with hydrosalpinx.⁶⁸ Tubal occlusion compared with salpingectomy did not show a significant advantage in terms of ongoing pregnancy or clinical pregnancy.

The Practice Committee of the American Society for Reproductive Medicine (ASRM), in collaboration with the Society of Reproductive Surgeons (SRS), published a study about IVF outcomes in women with unilateral or bilateral hydrosalpinges.⁶⁹ The PR in patients with hydrosalpinges was lower than that of the control group, with identical negative effects in both fresh and frozen embryo transfer cycles. Even the miscarriage rate was 2.3 times higher among women with hydrosalpinx than among those unaffected. Preliminary laparoscopic salpingectomy or proximal tubal occlusion, prior to IVF, improves pregnancy and live birth rates, whereas data was insufficient to evaluate the effectiveness of alternative treatments such as laparoscopic neosalpingostomy, transvaginal aspiration of HF, hysteroscopic tubal occlusion, or antibiotic treatment. Transvaginal aspiration of HF during egg collection is still controversial.^{70,71} The data in the literature is insufficient to determine whether hysteroscopic tubal occlusion could be considered helpful in preventing the deleterious effect of HSF. A recent study reports that the placement of Essure (Bayer AG, Leverkusen, Germany) is an alternative method for fallopian tube occlusion in hydrosalpinx before IVF when laparoscopy is contraindicated.⁷²

The hysteroscopic proximal occlusion by intratubal devices improves the chance of achieving clinical pregnancy compared with no intervention. Nevertheless, the Essure implant is associated with a higher miscarriage rate compared with the other interventions.⁷³ Unfortunately, insertion of the Essure in the management of hydrosalpinx prior to IVF remains off-label, although the implant could increase fluid within the uterine cavity, with a consequent lower PR.⁷⁴ In fact, the hysteroscopic placement of Essure devices before IVF produces inferior pregnancy rates compared with the laparoscopic approach.⁷⁵ Therefore, salpingectomy should be the first option for women with hydrosalpinx before undergoing an IVF cycle. Salpingostomy should be considered to conserve the fallopian tubes, enabling natural conception.^{76,77}

Conclusion

Hydrosalpinx is associated with poor IVF outcomes, with reduced live birth rates and higher pregnancy loss. Therefore, accurate diagnosis of hydrosalpinx are critical, especially when a tiny amount of fluid inside the Fallopian tubes may be almost undetectable. The surgical removal of hydrosalpinx prior to IVF improves PRs and reproductive outcomes.⁷⁸ In addition, salpingectomy does not affect the ovarian response, nor reduces the antral follicle count.⁷⁹ Laparoscopic salpingectomy is the optimal surgical approach; therefore, it should be recommended. Further studies are needed to establish the therapeutic value of HF aspiration under ultrasonographic guidance, during or after oocyte retrieval, in terms of PR and ongoing pregnancy.

Conflict to Interests

The authors have no conflict of interests to declare.

References

- Schlaff WD. A reconsideration of salpingectomy for hydrosalpinx before in vitro fertilization: why bother? *Fertil Steril*. 2019;111(04):650–651. Doi: 10.1016/j.fertnstert.2019.02.014
- Zeyneloglu HB, Arici A, Olive DL. Adverse effects of hydrosalpinx on pregnancy rates after in vitro fertilization-embryo transfer. *Fertil Steril*. 1998;70(03):492–499. Doi: 10.1016/s0015-0282(98)00200-3
- Camus E, Poncet C, Goffinet F, Wainer B, Merlet F, Nisand I, Philippe HJ. Pregnancy rates after in-vitro fertilization in cases of tubal infertility with and without hydrosalpinx: a meta-analysis of published comparative studies. *Hum Reprod*. 1999;14(05):1243–1249. Doi: 10.1093/humrep/14.5.1243
- Savaris RF, Giudice LC. The influence of hydrosalpinx on markers of endometrial receptivity. *Semin Reprod Med*. 2007;25(06):476–482. Doi: 10.1055/s-2007-991045
- Hill GA, Herbert CM, Fleischer AC, Webster BW, Maxson WS, Wentz AC. Enlargement of hydrosalpinges during ovarian stimulation protocols for in vitro fertilization and embryo replacement. *Fertil Steril*. 1986;45(06):883–885. Doi: 10.1016/s0015-0282(16)49411-2
- Schiller VL, Tsuchiyama K. Development of hydrosalpinx during ovulation induction. *J Ultrasound Med*. 1995;14(11):799–803. Doi: 10.7863/jum.1995.14.11.799
- Ng EH, Ajonuma LC, Lau EY, Yeung WS, Ho PC. Adverse effects of hydrosalpinx fluid on sperm motility and survival. *Hum Reprod*. 2000;15(04):772–777. Doi: 10.1093/humrep/15.4.772
- de Ziegler D, Pirtea P, Galliano D, Cicinelli E, Meldrum D. Optimal uterine anatomy and physiology necessary for normal

- implantation and placentation. *Fertil Steril*. 2016;105(04):844–854. Doi: 10.1016/j.fertnstert.2016.02.023
- 9 Li L, Xu BF, Chen QJ, Sun XX. Effects of hydrosalpinx on pinopodes, leukaemia inhibitory factor, integrin beta3 and MUC1 expression in the peri-implantation endometrium. *Eur J Obstet Gynecol Reprod Biol*. 2010;151(02):171–175. Doi: 10.1016/j.ejogrb.2010.04.024
 - 10 Cohen A, Almog B, Tulandi TJM. Hydrosalpinx sclerotherapy before in vitro fertilization: systematic review and meta-analysis. *Invasive Gynecol*. 2018;25(04):600–607. doi: 10.1016/j.jmig.2017.12.004
 - 11 Harb H, Al-Rshoud F, Karunakaran B, Gallos ID, Coomarasamy A. Hydrosalpinx and pregnancy loss: a systematic review and meta-analysis. *Reprod Biomed Online*. 2019;38(03):427–441. Doi: 10.1016/j.rbmo.2018.12.020
 - 12 Puttemans P, Campo R, Gordts S, Brosens I. Hydrosalpinx and ART: hydrosalpinx–functional surgery or salpingectomy? *Hum Reprod*. 2000;15(07):1427–1430. Doi: 10.1093/humrep/15.7.1427
 - 13 Puttemans PJ, Brosens IA. Salpingectomy improves in-vitro fertilization outcome in patients with a hydrosalpinx: blind victimization of the fallopian tube? *Hum Reprod*. 1996;11(10):2079–2081. Doi: 10.1093/oxfordjournals.humrep.a019052
 - 14 Fylstra DL. Ectopic pregnancy after hysterectomy: a review and insight into etiology and prevention. *Fertil Steril*. 2010;94(02):431–435. Doi: 10.1016/j.fertnstert.2009.03.039
 - 15 Cozzolino M, Diaz-Gimeno P, Pellicer A, Garrido N. Evaluation of the endometrial receptivity assay and the preimplantation genetic test for aneuploidy in overcoming recurrent implantation failure. *J Assist Reprod Genet*. 2020;37(12):2989–2997. doi: 10.1007/s10815-020-01948-7
 - 16 Griffiths AN, Watermeyer SR, Klentzeris LD. Fluid within the endometrial cavity in an IVF cycle—a novel approach to its management. *J Assist Reprod Genet*. 2002;19(06):298–301. Doi: 10.1023/a:1015785431828
 - 17 He RH, Gao HJ, Li YQ, Zhu XM. The associated factors to endometrial cavity fluid and the relevant impact on the IVF-ET outcome. *Reprod Biol Endocrinol*. 2010;8:46. Doi: 10.1186/1477-7827-8-46
 - 18 Kassabji M, Sims JA, Butler L, Muasher SJ. Reduced pregnancy outcome in patients with unilateral or bilateral hydrosalpinx after in vitro fertilization. *Eur J Obstet Gynecol Reprod Biol*. 1994;56(02):129–132. Doi: 10.1016/0028-2243(94)90269-0
 - 19 Beyler SA, James KP, Fritz MA, Meyer WR. Hydrosalpingeal fluid inhibits in-vitro embryonic development in a murine model. *Hum Reprod*. 1997;12(12):2724–2728. Doi: 10.1093/humrep/12.12.2724
 - 20 Chan LY, Chiu PY, Cheung LP, Haines CJ, Tung HF, Lau TK. A study of teratogenicity of hydrosalpinx fluid using a whole rat embryo culture model. *Hum Reprod*. 2003;18(05):955–958. Doi: 10.1093/humrep/deg189
 - 21 Roberts JE, Clarke HJ, Tulandi T, Tan SL. Effects of hydrosalpingeal fluid on murine embryo development and implantation. *J Assist Reprod Genet*. 1999;16(08):421–424. Doi: 10.1023/A:1020517524857
 - 22 Granot I, Dekel N, Segal I, Fieldust S, Shoham Z, Barash A. Is hydrosalpinx fluid cytotoxic? *Hum Reprod*. 1998;13(06):1620–1624. Doi: 10.1093/humrep/13.6.1620
 - 23 Strandell A, Sjögren A, Bentin-Ley U, Thorburn J, Hamberger L, Brännström M. Hydrosalpinx fluid does not adversely affect the normal development of human embryos and implantation in vitro. *Hum Reprod*. 1998;13(10):2921–2925. Doi: 10.1093/humrep/13.10.2921
 - 24 Ajonuma LC, Chan LN, Ng EH, Chow PH, Kung LS, Cheung ANY, et al. Characterization of epithelial cell culture from human hydrosalpinges and effects of its conditioned medium on embryo development and sperm motility. *Hum Reprod*. 2003;18(02):291–298. Doi: 10.1093/humrep/deg078
 - 25 Mansour RT, Aboufghar MA, Serour GI, Riad R. Fluid accumulation of the uterine cavity before embryo transfer: a possible hindrance for implantation. *J In Vitro Fert Embryo Transf*. 1991;8(03):157–159. Doi: 10.1007/BF01131707
 - 26 Andersen AN, Lindhard A, Loft A, Ziebe S, Andersen CY. The infertile patient with hydrosalpinges–IVF with or without salpingectomy? *Hum Reprod*. 1996;11(10):2081–2084. Doi: 10.1093/oxfordjournals.humrep.a019053
 - 27 Vandromme J, Chasse E, Lejeune B, Van Rysselberge M, Delvigne A, Leroy F. Hydrosalpinges in in-vitro fertilization: an unfavourable prognostic feature. *Hum Reprod*. 1995;10(03):576–579. Doi: 10.1093/oxfordjournals.humrep.a135992
 - 28 Chen Q, Zhang Y, Elad D, Jaffa AJ, Cao Y, Ye X, Duan E. Navigating the site for embryo implantation: biomechanical and molecular regulation of intrauterine embryo distribution. *Mol Aspects Med*. 2013;34(05):1024–1042. Doi: 10.1016/j.mam.2012.07.017
 - 29 Zhang S, Lin H, Kong S, Wang S, Wang H, Wang H, Armant DR. Physiological and molecular determinants of embryo implantation. *Mol Aspects Med*. 2013;34(05):939–980. Doi: 10.1016/j.mam.2012.12.011
 - 30 Sallee N, Baines DL, Naftalin RJ, Milligan SR. The hormonal control of uterine luminal fluid secretion and absorption. *J Membr Biol*. 2005;206(01):17–28. Doi: 10.1007/s00232-005-0770-7
 - 31 Naftalin RJ, Thiagarajah JR, Pedley KC, Pocock VJ, Milligan SR. Progesterone stimulation of fluid absorption by the rat uterine gland. *Reproduction*. 2002;123(05):633–638. Doi: 10.1530/rep.0.1230633
 - 32 Clemetson CA, Mallikarjuneswara VR, Moshfeghi MM, Carr JJ, Wilds JH. The effects of oestrogen and progesterone on the sodium and potassium concentrations of rat uterine fluid. *J Endocrinol*. 1970;47(03):309–319. Doi: 10.1677/joe.0.0470309
 - 33 Ajonuma LC, Ng EH, Chan HC. New insights into the mechanisms underlying hydrosalpinx fluid formation and its adverse effect on IVF outcome. *Hum Reprod Update*. 2002;8(03):255–264. Doi: 10.1093/humupd/8.3.255
 - 34 Ajonuma LC, Chan PK, Ng EH, et al. Involvement of cystic fibrosis transmembrane conductance regulator (CFTR) in the pathogenesis of hydrosalpinx induced by Chlamydia trachomatis infection. *J Obstet Gynaecol Res*. 2008;34(06):923–930. Doi: 10.1111/j.1447-0756.2008.00826.x
 - 35 Paavonen J, Eggert-Kruse W. Chlamydia trachomatis: impact on human reproduction. *Hum Reprod Update*. 1999;5(05):433–447. Doi: 10.1093/humupd/5.5.433
 - 36 Jones RB, Mammel JB, Shepard MK, Fisher RR. Recovery of Chlamydia trachomatis from the endometrium of women at risk for chlamydial infection. *Am J Obstet Gynecol*. 1986;155(01):35–39. Doi: 10.1016/0002-9378(86)90073-6
 - 37 Wølner-Hanssen P, Mårdh PA, Møller B, Weström L. Endometrial infection in women with Chlamydial salpingitis. *Sex Transm Dis*. 1982;9(02):84–88. Doi: 10.1097/00007435-198204000-00006
 - 38 Toth M, Jeremias J, Ledger WJ, Witkin SS. In vivo tumor necrosis factor production in women with salpingitis. *Surg Gynecol Obstet*. 1992;174(05):359–362
 - 39 Witkin SS, Sultan KM, Neal GS, Jeremias J, Grifo JA, Rosenwaks Z. Unsuspected Chlamydia trachomatis infection and in vitro fertilization outcome. *Am J Obstet Gynecol*. 1994;171(05):1208–1214. Doi: 10.1016/0002-9378(94)90134-1
 - 40 Spandorfer SD, Liu HC, Neuer A, Barmat LI, Davis O, Rosenwaks Z. The embryo toxicity of hydrosalpinx fluid is only apparent at high concentrations: an in vitro model that stimulates in vivo events. *Fertil Steril*. 1999;71(04):619–626. Doi: 10.1016/s0015-0282(98)00541-x
 - 41 Leese HJ. The formation and function of oviduct fluid. *J Reprod Fertil*. 1988;82(02):843–856. Doi: 10.1530/jrf.0.0820843
 - 42 Brunton WJ, Brinster RL. Active chloride transport in the isolated rabbit oviduct. *Am J Physiol*. 1971;221(02):658–661. Doi: 10.1152/ajplegacy.1971.221.2.658
 - 43 Brunton WJ. Beta-adrenergic stimulation of transmembrane potential and short circuit current of isolated rabbit oviduct. *Nat New Biol*. 1972;236(61):12–14. Doi: 10.1038/newbio236012a0
 - 44 Dickens CJ, Comer MT, Southgate J, Leese HJ. Human Fallopian tubal epithelial cells in vitro: establishment of polarity and potential role of intracellular calcium and extracellular ATP in fluid secretion. *Hum Reprod*. 1996;11(01):212–217. Doi: 10.1093/oxfordjournals.humrep.a019021

- 45 Downing SJ, Maguiness SD, Watson A, Leese HJ. Electrophysiological basis of human fallopian tubal fluid formation. *J Reprod Fert.* 1997;111(01):29–34. Doi: 10.1530/jrf.0.1110029
- 46 Sheppard DN, Welsh MJ. Structure and function of the CFTR chloride channel. *Physiol Rev.* 1999;79(1, Suppl):S23–S45. Doi: 10.1152/physrev.1999.79.1.S23
- 47 Pier GB, Grout M, Zaidi T, Meluleni G, Mueschenborn SS, Banting G, et al. Salmonella typhi uses CFTR to enter intestinal epithelial cells. *Nature.* 1998;393(6680):79–82. Doi: 10.1038/30006
- 48 Gerçeker AA, Zaidi T, Marks P, Golan DE, Pier GB. Impact of heterogeneity within cultured cells on bacterial invasion: analysis of *Pseudomonas aeruginosa* and *Salmonella enterica* serovar typhi entry into MDCK cells by using a green fluorescent protein-labelled cystic fibrosis transmembrane conductance regulator receptor. *Infect Immun.* 2000;68(02):861–870. Doi: 10.1128/iai.68.2.861-870.2000
- 49 Downing SJ, Tay JI, Maguiness SD, Watson A, Leese HJ. Effect of inflammatory mediators on the physiology of the human Fallopian tube. *Hum Fert (Camb).* 2002;5(02):54–60. Doi: 10.1080/1464727022000198932
- 50 Meyer WR, Castelbaum AJ, Somkuti S, et al. Hydrosalpinges adversely affect markers of endometrial receptivity. *Hum Reprod.* 1997;12(07):1393–1398. Doi: 10.1093/humrep/12.7.1393
- 51 Yoshinaga K. Research on Blastocyst Implantation Essential Factors (BIEFs). *Am J Reprod Immunol.* 2010;63(06):413–424. Doi: 10.1111/j.1600-0897.2010.00853.x
- 52 Paulson RJ. Introduction: Endometrial receptivity: evaluation, induction and inhibition. *Fertil Steril.* 2019;111(04):609–610. Doi: 10.1016/j.fertnstert.2019.02.029
- 53 Aghajanova L, Hamilton AE, Giudice LC. Uterine receptivity to human embryonic implantation: histology, biomarkers, and transcriptomics. *Semin Cell Dev Biol.* 2008;19(02):204–211. Doi: 10.1016/j.semcdb.2007.10.008
- 54 Block K, Kardana A, Igarashi P, Taylor HS. In utero diethylstilbestrol (DES) exposure alters Hox gene expression in the developing müllerian system. *FASEB J.* 2000;14(09):1101–1108. Doi: 10.1096/fasebj.14.9.1101
- 55 Taylor HS, Daftary GS, Selam B. Endometrial HOXA10 expression after controlled ovarian hyperstimulation with recombinant follicle-stimulating hormone. *Fertil Steril.* 2003;80(Suppl 2):839–843. Doi: 10.1016/s0015-0282(03)00985-3
- 56 Taylor HS, Arici A, Olive D, Igarashi P. HOXA10 is expressed in response to sex steroids at the time of implantation in the human endometrium. *J Clin Invest.* 1998;101(07):1379–1384. Doi: 10.1172/JCI1057
- 57 Daftary GS, Taylor HS. Endocrine regulation of HOX genes. *Endocr Rev.* 2006;27(04):331–355. Doi: 10.1210/er.2005-0018
- 58 Satokata I, Benson G, Maas R. Sexually dimorphic sterility phenotypes in Hoxa10-deficient mice. *Nature.* 1995;374(6521):460–463. Doi: 10.1038/374460a0
- 59 Eun Kwon H, Taylor HS. The role of HOX genes in human implantation. *Ann N Y Acad Sci.* 2004;1034:1–18. Doi: 10.1196/annals.1335.001
- 60 Sarno JL, Kliman HJ, Taylor HS. HOXA10, Pbx2, and Meis1 protein expression in the human endometrium: formation of multimeric complexes on HOXA10 target genes. *J Clin Endocrinol Metab.* 2005;90(01):522–528. Doi: 10.1210/jc.2004-0817
- 61 Gui Y, Zhang J, Yuan L, Lessey BA. Regulation of HOXA-10 and its expression in normal and abnormal endometrium. *Mol Hum Reprod.* 1999;5(09):866–873. Doi: 10.1093/molehr/5.9.866
- 62 Daftary GS, Taylor HS. Pleiotropic effects of Hoxa10 on the functional development of peri-implantation endometrium. *Mol Reprod Dev.* 2004;67(01):8–14. Doi: 10.1002/mrd.20013
- 63 Bagot CN, Troy PJ, Taylor HS. Alteration of maternal Hoxa10 expression by in vivo gene transfection affects implantation. *Gene Ther.* 2000;7(16):1378–1384. Doi: 10.1038/sj.gt.3301245
- 64 Bagot CN, Kliman HJ, Taylor HS. Maternal Hoxa10 is required for pinopod formation in the development of mouse uterine receptivity to embryo implantation. *Dev Dyn.* 2001;222(03):538–544. Doi: 10.1002/dvdy.1209
- 65 Lim H, Ma L, Ma WG, Maas RL, Dey SK. Hoxa-10 regulates uterine stromal cell responsiveness to progesterone during implantation and decidualization in the mouse. *Mol Endocrinol.* 1999;13(06):1005–1017. Doi: 10.1210/mend.13.6.0284
- 66 Daftary GS, Kayisli U, Seli E, Bukulmez O, Arici A, Taylor HS. Salpingectomy increases peri-implantation endometrial HOXA10 expression in women with hydrosalpinx. *Fertil Steril.* 2007;87(02):367–372. Doi: 10.1016/j.fertnstert.2006.06.041
- 67 Strandell A, Lindhard A, Waldenström U, Thorburn J. Hydrosalpinx and IVF outcome: cumulative results after salpingectomy in a randomized controlled trial. *Hum Reprod.* 2001;16(11):2403–2410. Doi: 10.1093/humrep/16.11.2403
- 68 Johnson N, van Voorst S, Sowter MC, Strandell A, Mol BW. Surgical treatment for tubal disease in women due to undergo in vitro fertilisation. *Cochrane Database Syst Rev.* 2010;(01):CD002125. Doi: 10.1002/14651858.CD002125.pub3
- 69 Practice Committee of American Society for Reproductive Medicine in collaboration with Society of Reproductive Surgeons. Salpingectomy for hydrosalpinx prior to in vitro fertilization. *Fertil Steril.* 2008;90(5, Suppl):S66–S68. Doi: 10.1016/j.fertnstert.2008.08.089
- 70 Sowter MC, Akande VA, Williams JA, Hull MG. Is the outcome of in-vitro fertilization and embryo transfer treatment improved by spontaneous or surgical drainage of a hydrosalpinx? *Hum Reprod.* 1997;12(10):2147–2150. Doi: 10.1093/humrep/12.10.2147
- 71 Van Voorhis BJ, Sparks AE, Syrop CH, Stovall DW. Ultrasound-guided aspiration of hydrosalpinges is associated with improved pregnancy and implantation rates after in-vitro fertilization cycles. *Hum Reprod.* 1998;13(03):736–739. Doi: 10.1093/humrep/13.3.736
- 72 Lorente González J, Ríos Castillo JE, Pomares Toro E, Romero Nieto MI, Castelo-Branco C, Arjona Berral JE. Essure a novel option for the treatment of hydrosalpinx: a case series and literature review. *Gynecol Endocrinol.* 2016;32(02):166–170. Doi: 10.3109/09513590.2015.1103221
- 73 Barbosa MW, Sotiriadis A, Papatheodorou SI, Mijatovic V, Nastri CO, Martins WP. High miscarriage rate in women treated with Essure® for hydrosalpinx before embryo transfer: a systematic review and meta-analysis. *Ultrasound Obstet Gynecol.* 2016;48(05):556–565. Doi: 10.1002/uog.15960
- 74 Sills ES, Walsh DJ, Jones CA, Wood SH. Endometrial fluid associated with Essure implants placed before in vitro fertilization: Considerations for patient counseling and surgical management. *Clin Exp Reprod Med.* 2015;42(03):126–129. Doi: 10.5653/cepm.2015.42.3.126
- 75 Xu B, Zhang Q, Zhao J, Wang Y, Xu D, Li Y. Pregnancy outcome of in vitro fertilization after Essure and laparoscopic management of hydrosalpinx: a systematic review and meta-analysis. *Fertil Steril.* 2017;108(01):84–95.e5. Doi: 10.1016/j.fertnstert.2017.05.005
- 76 Hamilton CJ, Evers JL, Hoogland HJ. Ovulatory disorders and inflammatory adnexal damage: a neglected cause of the failure of fertility microsurgery. *Br J Obstet Gynaecol.* 1986;93(03):282–284. Doi: 10.1111/j.1471-0528.1986.tb07909.x
- 77 Gomet V, Wang I. Laparoscopic surgery for infertility therapy. *Curr Opin Obstet Gynecol.* 1994;6(02):141–148
- 78 Demir B, Bozdag G, Sengul O, Kahyaoglu I, Mumusoglu S, Zengin D. The impact of unilateral salpingectomy on antral follicle count and ovarian response in ICSI cycles: comparison of contralateral side. *Gynecol Endocrinol.* 2016;32(09):741–744. Doi: 10.3109/09513590.2016.1161741
- 79 Yoon SH, Lee JY, Kim SN, Chung HW, Park SY, Lee C. Does salpingectomy have a deleterious impact on ovarian response in in vitro fertilization cycles? *Fertil Steril.* 2016;106(05):1083–1092.e5. Doi: 10.1016/j.fertnstert.2016.05.030