

REVIEW ARTICLE OPEN



Towards a multilevel model of major depression: genes, immuno-metabolic function, and cortico-striatal signaling

Elisabeth R. Paul ^{1,2}, Lars Östman^{1,3}, Markus Heilig ^{1,3}, Helen S. Mayberg ⁴ and J. Paul Hamilton ⁵✉

© The Author(s) 2023

Biological assay and imaging techniques have made visible a great deal of the machinery of mental illness. Over fifty years of investigation of mood disorders using these technologies has identified several biological regularities in these disorders. Here we present a narrative connecting genetic, cytokine, neurotransmitter, and neural-systems-level findings in major depressive disorder (MDD). Specifically, we connect recent genome-wide findings in MDD to metabolic and immunological disturbance in this disorder and then detail links between immunological abnormalities and dopaminergic signaling within cortico-striatal circuitry. Following this, we discuss implications of reduced dopaminergic tone for cortico-striatal signal conduction in MDD. Finally, we specify some of the flaws in the current model and propose ways forward for advancing multilevel formulations of MDD most efficiently.

Translational Psychiatry (2023)13:171 | <https://doi.org/10.1038/s41398-023-02466-7>

Over the last half-century, in-vivo imaging and biological assay techniques have developed at a rapid pace. These techniques have made visible in mood disorders a broad variety of biological processes that were previously inaccessible. While a rough theoretical scaffolding has guided biological inquiry into mood disorders, most of this research has been exploratory given the complexity of the systems in play. More recently, and with many investigations of mood disorders conducted, several important regularities have emerged across biological levels including genetics, endocrine and cytokine signaling, and metabolomics, in addition to the neural-systems level as assessed with molecular and functional neuroimaging. In this review, we present the status of a developing multilevel biological perspective on major depressive disorder (MDD). We show that it is now possible to present an integrative thread from genetics to cytokines to neurotransmitters up to the neural-systems level in MDD. In presenting this formulation, we hope to show that cross-level integration in MDD is necessary in the conceptualization of this disorder.

Developing a biologically coherent understanding of major depression must first take into account the marked heterogeneity of MDD, given that putative depressive subtypes have shown distinct biological profiles [1]. Subtyping of MDD is both challenging and still in development [2, 3], with further progress entailing iterative refinement through interactions between the domains of behavior, biology, and nosology. The DSM-IV and DSM-5 have specifiers for melancholic and atypical subtypes of depression with the DSM-5 including a new specifier for depression “with anxious distress” [4, 5]. Importantly, severity of depression is emerging as an important dimension in the subtyping of MDD, with severe depression having a unique genetic signature [6] and clinical course, regardless of melancholic or atypical features [7]. To make this review tractable, we will focus on presenting a multilevel

biological model that tends more toward atypical depression, which is distinct from melancholic and anxious depression in that increased appetite, hypersomnia, and rejection sensitivity occur in this subtype of MDD [4]. Among currently available diagnostic categories of depression, we use atypical MDD as the point of departure for our biological narrative both because atypical depression is the most heritable [8] and because it is closest to an emerging immuno-metabolic conceptualization of MDD [9]. Importantly, in developing this account, we include investigations both of atypical MDD and MDD more broadly construed when variables examined have high relevance to atypical depression (e.g., metabolomic and immunological studies).

GENETICS OF DEPRESSION

In spite of evidence from twin-concordance studies for moderate (33–42%) heritability of MDD [10], early genome-wide association studies (GWAS) of depression did not identify genome-wide-significant single nucleotide polymorphisms (SNPs) associated with this disorder [11]. As the size of case and control samples grew, the power of GWAS studies improved and significant variants were detected [12–15]. Correspondence between detected SNPs across these studies, however, was low with only one common depression-associated SNP identified across any pair of studies [12–15]. A recent meta-analysis of GWAS studies of depression, however, identified stronger and more consistent findings, with 102 genome-wide significant variants detected, 56 of which were found to be genome-wide significant in an independent replication cohort [16].

This most recent MDD GWAS identified a strong genetic link between MDD and metabolic dysfunction. This link is most readily seen from the results of genetic correlation analysis for estimating the degree of genetic correspondence between depression and a

¹Center for Social and Affective Neuroscience, Department of Biomedical and Clinical Sciences, Linköping University, Linköping, Sweden. ²Center for Medical Imaging and Visualization, Linköping University, Linköping, Sweden. ³Department of Psychiatry, Region Östergötland, Linköping, Sweden. ⁴Icahn School of Medicine at Mount Sinai, New York, NY, USA. ⁵Department of Biological and Medical Psychology, University of Bergen, Bergen, Norway. ✉email: paul.hamilton@uib.no

Received: 10 August 2021 Revised: 24 April 2023 Accepted: 3 May 2023

Published online: 19 May 2023

variety of traits. Among 31 traits found to have a significant, positive genetic correlation with MDD, nine were from the domain of metabolic disturbance (obesity class 3, body fat, triglycerides, coronary artery disease, waist-to-hip ratio, overweight, waist circumference, obesity class 1, and body mass index).

METABOLIC DISTURBANCE AND DEPRESSION

The link between the genetics of MDD and metabolic disturbance is not surprising given consistent evidence connecting MDD and metabolic syndrome, a syndrome characterized by abdominal obesity, increased fasting-glucose levels, high blood pressure, high triglyceride levels, and decreased high-density lipoprotein cholesterol [17]. For example, a meta-analysis of 29 epidemiological cross-sectional and prospective longitudinal cohort studies showed a bidirectional relation between MDD and metabolic syndrome such that depression predicts metabolic syndrome and vice versa [18]. While the potentially many pathophysiological mechanisms connecting metabolic syndrome and depression have not been extensively investigated, one promising candidate is insulin resistance. While the brain was long considered an insulin-independent organ, more recent work has shown that insulin receptors are distributed abundantly in the brain [19] and that pancreas-derived insulin binds to epithelial cells on the blood-brain barrier and can then enter the brain via receptor-mediated transcytosis [20]. Importantly, brain cells expressing insulin receptors also show insulin insensitivity [21]. The implications for neural-level insulin resistance and MDD are clear considering findings that obese humans show neural unresponsiveness to exogenous insulin [22] and fail to show mood elevating effects [23] observed in normal-weight humans resulting from a course of intranasal insulin administration [24].

In the context of metabolic disturbance in MDD, it is important to consider a potential role for the gut microbiome in the pathophysiology of depression. Several putative links between MDD and gut-microbe composition are under consideration [25]. One intriguing possibility extends from microbial degradation of dietary fibers in the gut, a function which human intestinal enzymes lack. Some of the byproducts of this degradation, namely short chain fatty acids (SCFAs), are believed to be bioactive, with effects on metabolism [26]. A double-blind study assessing the metabolic and behavioral effects of dietary fiber supplementation found a lipid-lowering effect, weight loss, and increased satiety, which was associated with increases in fecal SCFAs [27]. This is a promising finding considering the increasing attention SCFAs have garnered recently as a possible biomarker, or pathophysiological substrate, of MDD [28].

Importantly, in the context of MDD, metabolic disturbances and inflammation are consistently linked. A large cross-sectional study examined waking cortisol response, inflammatory cytokine levels, and metabolic syndrome in melancholic MDD and atypical MDD. This investigation showed that metabolic syndrome and increased inflammation, but not heightened cortisol response, co-occur in atypical MDD whereas exaggerated waking cortisol response occurs in melancholic MDD in the absence of elevated inflammation and metabolic syndrome [1]. This association between atypical MDD, inflammation, and metabolic syndrome has been replicated in a large, twin-registry cohort study of metabolic syndrome [29]. Thus, extant data reveal strong links between the genetics of depression, metabolic disturbance, and inflammation.

INFLAMMATION, DOPAMINE, AND DEPRESSION

Inflammation is a protective response of the body to pathogens or injury. Researchers have noted the resemblance between depressive symptoms and the sickness-syndrome induced by inflammation, which is characterized by increased sleep, inactivity, and social withdrawal [30]. This, in addition to the finding that immune-activating therapies—such as interferon-alpha treatment

for hepatitis C [31] and autoimmune disorders [32]—lead to elevated levels of depressive symptoms has motivated investigations of the role of inflammation in MDD. Indeed, a significant body of evidence for immune system involvement in MDD has amassed over the past four decades. These data indicate a general dysregulation of the inflammatory response in MDD with abnormalities in both pro- and anti-inflammatory chemical messengers [33–35]. While it is uncommon to treat MDD with anti-inflammatory drugs alone, augmentation of standard antidepressants with anti-inflammatories is effective in comparison to placebo augmentation [36]. Moreover, it has been proposed that the rapid-acting antidepressant effects of the psychoactive drug ketamine could be anti-inflammatory in nature given the effects of this drug on both peripheral and central inflammation [37].

Inflammation in both the peripheral and central nervous system can lead to the activation of the brain's microglia and dendritic cells, prompting an immune response that alters neural functioning [38]. Administration of the pro-inflammatory cytokine interferon-alpha to rats, for example, decreases central levels of dopamine and tetrahydrobiopterin [39], where tetrahydrobiopterin is an enzyme co-factor for tyrosine hydroxylase, the rate-limiting enzyme in dopamine synthesis. Additionally, activation of mitogen-activated protein kinase pathways by inflammatory challenge upregulates dopamine-transporter activity [40], reducing post-synaptic availability of dopamine. Further, results from mouse models show that inflammation-activated microglia produce prostaglandins which modulate striatal-neuron activation resulting in negative affect [41], potentially by inhibition of dopaminergic cells [42]. Similar effects have been observed in humans where increased anhedonia and depressive symptoms in response to inflammatory stimuli have been reliably related to decreased dopamine release and striatal activation [43]. Importantly, dopamine is not the only neurotransmitter implicated in the inflammation-induced signaling cascade. Inflammation has also been linked to reduced availability of serotonin [44] in addition to neuro-structural alterations [45] through the kynurenine pathway.

KYNURENE PATHWAY AND DEPRESSION

Of many genes previously predicted to associate with MDD, one of the few found to do so is the KYNU gene [16] which codes for kynureninase, an enzyme critically involved in the kynurenine pathway. See Fig. 1 for a schematic overview of this pathway. Along this pathway, tryptophan is converted to kynurenine (KYN) by the rate-limiting enzymes indoleamine 2,3-dioxygenase 1 (IDO1) and tryptophan dioxygenase 2 (TDO2). Pro-inflammatory cytokines, such as interferon-gamma, activate IDO1 and TDO2 [46], reducing the bioavailability of tryptophan to convert to serotonin. KYN can then be further metabolized by kynurenine aminotransferases I-IV, which catalyze the synthesis of kynurenic acid (KYNA) in astrocytes [47]. Alternatively, KYN can be metabolized by kynurenine 3 monooxygenase (KMO) to 3-hydroxykynurenine (3-HK) in microglia [46]. Here in the kynurenine pathway, the KYNU gene comes into play by coding for kynureninase, which catalyzes KYN and 3-HK to anthranilic and 3-hydroxyanthranilic acids, respectively. In subsequent kynurenine pathway steps, these acids can either convert in microglia [48] to quinolinic acid (QUIN)—which can then become nicotinamide—or to picolinic acid (PIC). Importantly, the functional implications of having the KYNU-gene risk allele have not been well characterized in the brain or elsewhere. One recent study assessed KYNU expression in the anterior cingulate in postmortem brain tissue of 24 depressed patients and 12 never-disordered controls, finding no differences in KYNU expression between the groups [49].

Several kynurenine pathway metabolites have neuroactive effects, some of which are neurotoxic and others which are neuroprotective or neurotrophic. 3-HK is considered neurotoxic by virtue of being a generator of free radicals, whereas QUIN is excitotoxic through

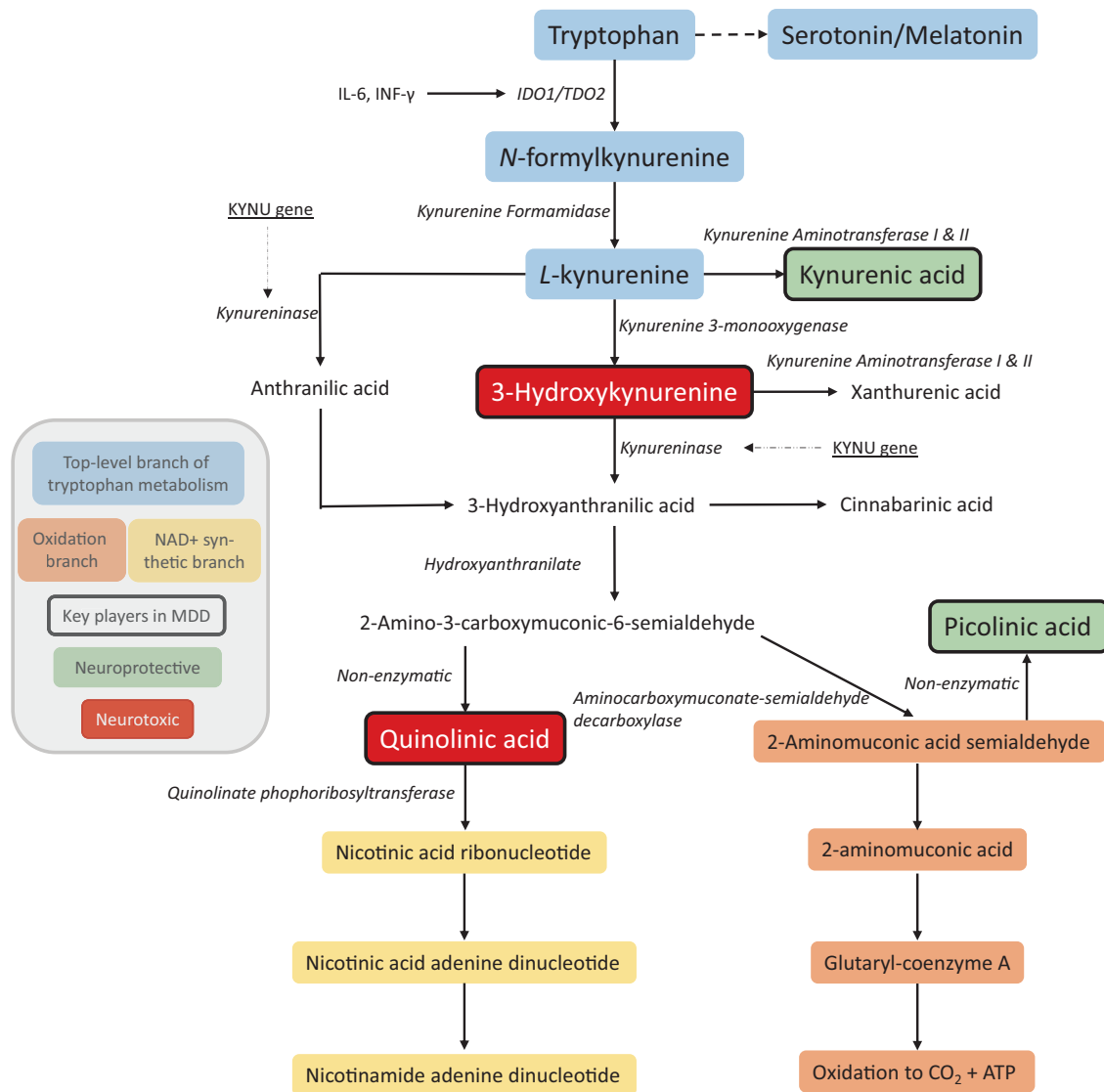


Fig. 1 Schematic overview of the kynurenine pathway and its significance in major depression. ATP adenosine triphosphate, IDO1 indoleamine 2,3-dioxygenase 1, IL interleukin, INF interferon, MDD major depressive disorder; NAD⁺ nicotinamide adenine dinucleotide, TDO2 tryptophan-2,3-dioxygenase 2. Blue fields belong to the top-level branch of the tryptophan metabolism pathway, orange fields are in the oxidative branch and fields within the NAD⁺ synthetic branch are shown in yellow. Metabolites shown in green are considered to have neuroprotective properties while metabolites in red are considered neurotoxic. Metabolites outlined in bold are key players along the kynurenine pathway in major depression.

agonism at N-methyl-D-aspartate (NMDA) receptors [50]. Among the neuroprotective kynurenine pathway metabolites are KYNA, which, like the fast-acting antidepressant ketamine, protects neurons through competitive antagonism at NMDA receptors [51, 52], and PIC, which blocks the neurotoxic effects of QUIN [53]. Increased neurotoxic [54] and decreased neuroprotective [55] kynurenine pathway metabolite levels, and decreased neuroprotective to neurotoxic metabolite ratios [56] have been observed in plasma in MDD. We recently extended these findings in the periphery by assessing kynurenine pathway metabolite levels in the cerebrospinal fluid (CSF) of MDD patients and healthy controls [57]. We found in CSF that neuroprotective PIC and PIC/QUIN ratios were reduced in MDD as well as inversely correlated with body mass index (BMI), which was significantly elevated in MDD. Importantly, weight gain is a cardinal symptom of atypical depression and has been considered important in conceptualizing atypical MDD in terms of metabolic and inflammatory disturbance [9]. This formulation is further supported by the finding that longitudinal changes in kynurenine

pathway metabolite levels and metabolic disturbances (diabetes, hypertension, and obesity) co-occur [58].

A shift from neuroprotective to neurotoxic activity in the kynurenine pathway could affect brain-structure volume in MDD. Brain areas that are reliably found to be altered in MDD, such as the hippocampus, amygdala, and striatum, are especially vulnerable to the effects of neurotoxic kynurenine pathway metabolites. Volumetric reductions in the hippocampus and amygdala, regions with a high expression of the GluN2B subunit on which QUIN acts [51], have been associated with a reduced KYNA/QUIN ratio in MDD [45]. Moreover, QUIN injection into the striatum leads to excitotoxic lesions in rodents, mostly due to loss of medium spiny neurons that predominate in this dopaminergically innervated region [59].

DOPAMINE AND DEPRESSION

As we detail above, inflammation impacts dopaminergic function. The first speculations about the involvement of dopaminergic

abnormalities in MDD conceptualized these abnormalities in terms of their relation to cognitive and behavioral stereotypes in depression, such as repetitive, unwanted focus on negative events and aspects of the self [60]. This later gave way to hypotheses of a dopaminergic deficit in MDD accounting for reduced psychomotor speed—which is especially evident in atypical depression—in addition to anhedonia and impaired concentration [61]. Enthusiasm for a dopamine-based hypothesis of depression subsequently waned, however, potentially as a response to inconsistent results from both postmortem and neuroimaging findings [61]. Nonetheless, recent data provide some cause for renewed interest in a role for dopamine in MDD.

As we have reported elsewhere [62], the striatal-dopamine neuroimaging literature in MDD is inconsistent, with the bulk of studies reporting no differences between depressed and healthy samples with respect to the binding potential of dopamine in the striatum. To bolster signal quality, most investigations conducted to date of the dopamine type-2 receptor (D_2)—most abundant in the striatum relative to other brain regions—have averaged across anatomically defined striatal regions-of-interest. Based on functional connectivity maps of the striatum, however, this region appears to be organized at a much finer spatial scale than its anatomical boundaries suggest [63]. If we therefore limit our consideration of the literature to the five studies that conducted voxel-wise, as opposed to coarser anatomical-region-of-interest comparisons of the binding potential of striatal D_2 receptors, a more consistent pattern of findings emerges. Three of these studies report increased binding potential in MDD [62, 64, 65], whereas one study reports reduced binding potential [66], with one study finding no difference between depressed and healthy samples [67]. It is important to consider that increased binding potential of striatal D_2 receptors could hypothetically result from reduced competition of radioligand with endogenous dopamine, or increased striatal D_2 receptor density, or a combination of these factors. Striatal D_2 receptor imaging can detect altered competition with endogenous ligand. For example, injection of amphetamine, which increases extra-synaptic striatal-dopamine levels via effects on monoamine transporters, results in reduced striatal D_2 receptor binding [68]. Conversely, in Parkinson's disease, where dopaminergic input to the striatum from the substantia nigra is reduced, increased radiotracer binding to striatal D_2 receptors is reliably observed [69]. Amphetamine challenge D_2 receptor binding studies using depressed samples—with the capability of disambiguating endogenous-competition versus receptor-density accounts in MDD—are lacking, with just one pilot study reporting equivocal effects [70].

While any assertions about the coherence in the dopamine neuroimaging literature in MDD might be preliminary, more consistent findings have been reported from a recent meta-analysis of studies assaying CSF for the dopamine metabolite homovanillic acid (HVA). Unlike serotonin and norepinephrine metabolites in CSF that do not consistently differ in depressed relative to control samples, HVA levels are reliably reduced, with 19 of 23 studies reporting lower HVA in the CSF in depression [71]. As CSF measures of neurotransmitter activity are necessarily coarse, this reliable finding raises additional questions about the dopaminergic regions implicated. Nonetheless, in the domain of pharmacotherapy, a recent meta-analysis provided support for the efficacy of partial dopamine agonists—a similarly broad tool—in the treatment of MDD in treatment-refractory patients [72]. Finally, single nucleotide polymorphisms identified in the most recent, large-scale GWAS of MDD showed a highly reliable gene-based hit ($p < 10E-13$) within the *DRD2* gene, which codes for the D_2 receptor. Interestingly, no significant gene-based hits were identified for genes involved in the synthesis, post-synaptic transport, or receptor expression for serotonin. Similarly, drug-gene interaction analyses identified several hits for dopaminergic drugs and only one for a drug with direct serotonergic properties—the non-selective serotonin reuptake inhibitor vilazodone [16].

THE CORTICO-STRIATAL-PALLIDO-THALAMIC CIRCUIT

The cortico-striatal-pallido-thalamic circuit (henceforth referred to as the cortico-striatal circuit) represents a macro-architectural facet of neural organization. This re-entrant circuit is a major pathway through which cortical and brain-stem functions are united. The first conceptualizations of the cortico-striatal circuit proposed that it had an aggregating function, funneling a broad set of inputs from all of cerebral cortex toward a behavioral output ultimately implemented in motor cortex [73]. Data collected subsequently, however, suggested that this circuit is divided into segregated, “closed-loop” sub-circuits, with a motor circuit from sensorimotor cortex to the putamen and an association loop coming from multimodal association regions and passing through the caudate [74]. This functional differentiation was later broadened to include at least five segregated cortico-striatal circuit subdivisions linked to motor, oculomotor, dorsolateral prefrontal (DLPFC), lateral orbitofrontal, and anterior cingulate cortices (ACC) [75].

More recent work has challenged the assertion that the sub-circuits of the cortico-striatal circuit architecture are segregated. This work has been fueled by the claim that communication among the functionally specialized circuits is necessary for implementing complex, goal-directed behaviors. Consistent with this assertion, it was observed that in the striatum there are subregions of convergence between terminals from anterior cingulate, dorsolateral prefrontal, and motor regions [76]. Further, it was found that direct dopaminergic projections from the substantia nigra to cortical regions are diffuse, with individual cells in the substantia nigra sending collateral axons to parts of cortex that were putatively part of distinct cortico-striatal sub-circuits [77].

The cortico-striatal circuit has a “logic-board” configuration that determines whether signal conduction through this circuit is facilitated or inhibited. See Fig. 2 for a schematic. The so-called direct pathway is facilitatory such that cortical stimulation of the striatum blocks (via GABA-ergic striato-pallidal connections) tonic inhibition of the thalamus by the internal segment of the pallidum and substantia nigra pars reticulata, thereby enabling thalamo-cortical signal transfer. The inhibitory, indirect pathway is more complex and includes two additional terminals. Via this pathway, the cortically excited striatum sends inhibitory, GABA-ergic projections to the external pallidum which then dampens tonic pallidal inhibition of the subthalamic nucleus. The activated subthalamic nucleus then sends excitatory input to the internal pallidum/substantia nigra pars reticulata thereby facilitating its tonic inhibition of the thalamus [78].

The most important development in recent years with respect to our knowledge of the cortico-striatal architecture is that its subdivisions are not just a connected set of re-entrant, go/no-go circuits, but rather are connected in an orderly way that constitutes a “motivation-to-movement interface.” [79] The primary mechanism of this aspect of the cortico-striatal architecture is a change in bidirectional dopaminergic innervation between striatal and midbrain structures that runs along a ventral-to-dorsal gradient. Ventrally, striatum-to-midbrain dopamine innervation is maximized and midbrain-to-striatum dopaminergic connections are minimized. Proceeding dorsally, this asymmetry gradually reverses such that dopaminergic connectivity from the striatum to the midbrain diminishes and midbrain-to-striatum dopamine innervation is increased.

First, the ventral striatum—which is strongly connected with affective and reward-related regions such as the amygdala, hippocampus, orbitofrontal cortex, and anterior cingulate cortex—receives limited, and highly localized dopaminergic input from neurons in the ventral tegmental area extending to the substantia nigra pars compacta. Dopaminergic input from the ventral striatum back to midbrain regions, however, is much more topographically diffuse and includes parts of the midbrain that send dopaminergic projections to the central striatum, which receives glutamatergic projections from cortical regions implicated in higher cognitive

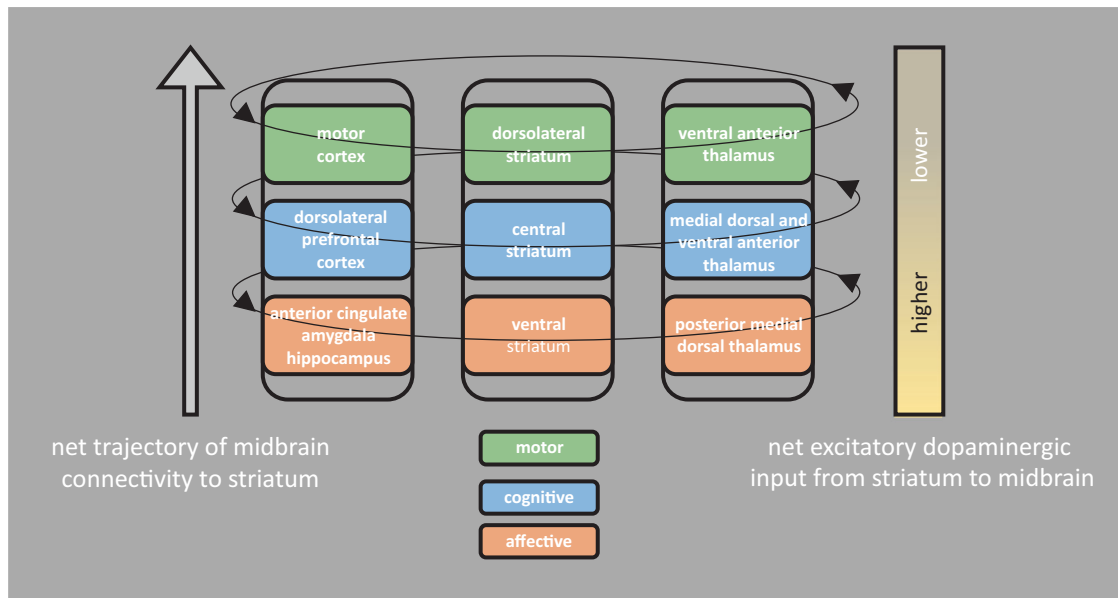


Fig. 2 The cortico-striatal circuit. Primary components of the dopaminergically mediated, ascending spiral, cortico-striatal-thalamic circuit.

functions. The central striatum, in turn, sends dopaminergic projections back to parts of the midbrain with dopaminergic connectivity to the dorsolateral aspect of the striatum. Finally, the dorsolateral striatum receives glutamatergic connections with motor regions and projects only limited dopaminergic connections to the midbrain. By virtue of this pattern of dopaminergic influence along an ascending spiral starting with the ventral striatum and ending with motor cortex, affective, cognitive, and motor processing are integrated [79]. See Fig. 2.

The interactions between the core cortico-striatal architecture and striatum-midbrain-striatum connections described here are not well characterized. There is evidence, however, that the dopaminergic projections from the substantia nigra pars compacta to the striatum reinforce cortical activation of cortico-striatal sub-circuits via contrasting actions on direct and indirect cortico-striatal pathways [78]. These nigrostriatal inputs appear to have a net excitatory effect on striatal neurons providing GABA-ergic inhibition of the external pallidum (facilitatory, direct pathway) and a net inhibitory effect on striatal neurons with GABA-ergic projections to the internal pallidum/substantia nigra pars reticulata (inhibitory, indirect pathway) [80–83]. Relatedly, the conventional account of the D_2 receptor is that this G-protein coupled receptor (GPCR) is relatively fast acting and inhibitory where $G_{\alpha i}$ subunits bind to and block adenylyl cyclases, preventing production of cyclic AMP [84]. Nonetheless, recently there has been growing acknowledgement that D_2 receptors can signal through beta-arrestin-2-dependent pathways and that this mode of action is complementary to canonical GPCR signaling, acting over the longer term to facilitate activity in D_2 receptor-expressing neurons [85]. Consistent with this account, mutant D_2 receptors that recruit arrestin but lack G-protein activity restore locomotor activity in D_2 receptor knockout mice when expressed virally in the ventral striatum [86].

DEPRESSION, DOPAMINE, AND THE CORTICO-STRIATAL CIRCUIT

In a meta-analysis of task-based functional neuroimaging studies of depression, we found reliably increased activation in MDD of the amygdala and ACC and reduced response of DLPFC to negative affective challenge. Moreover, we found reduced striatal response in MDD to both negative and positive affective provocation. We interpreted these findings in the context of the

dopaminergically mediated, ascending spiral cortico-striatal circuit model detailed above. Specifically, we proposed that in MDD amygdalar and ACC inputs to the “bottom,” affective subdivision of the cortico-striatal spiral were exaggerated (potentially by increased tonic input to these regions by the pulvinar nucleus of the thalamus) and that signal to higher cortical regions was dampened due to reductions in striatal dopaminergic activity facilitating signal conduction up the cortico-striatal spiral [87]. Embedded in this interpretation was the novel, testable hypothesis that striatal regions showing reductions in dopaminergic input in MDD should also show reduced functional connectivity with their cortical targets and that this should scale with the degree of the reduction in dopaminergic tone. We tested this hypothesis by concurrently imaging D_2 receptor binding potential using positron emission tomography (PET) and neural activity using resting functional magnetic resonance imaging in depressed and healthy comparison samples. Consistent with our hypotheses, we observed increased D_2 receptor binding potential in depressed participants in regions of the ventral and dorsal striatum. Moreover, we saw that as binding potential increased in ventral striatum and dorsal striatum in MDD, connectivity with the respective default-mode and posterior salience network targets of these striatal regions decreased [62]. This finding provided confirmation of the ascending spiral, cortico-striatal model in that dopaminergic input to the ventral striatum predicted connectivity only with its default-mode network target (hypothesized to subservise cognitive functioning) whereas dopaminergic input to dorsal striatum predicted connectivity primarily with motoric regions of the posterior salience network. From an affective standpoint, we propose that these data indicate that in MDD the emotional aspects of negative stimuli are experienced more intensely and that layers of mitigating, contextual information provided by higher cortical regions [88] are diminished due to reduced signal propagation to these regions up the cortico-striatal spiral. Imagine, for example, seeing an image of a snake on a television and your brain over-accentuating the snake and under-representing the televised medium.

In the continued development of cortico-striatal-thalamic models in MDD it will be important to integrate habenular function. The lateral habenula exerts an inhibitory influence on dopaminergic neurons of the ventral tegmental area and substantia nigra [89], which are considered important for learning

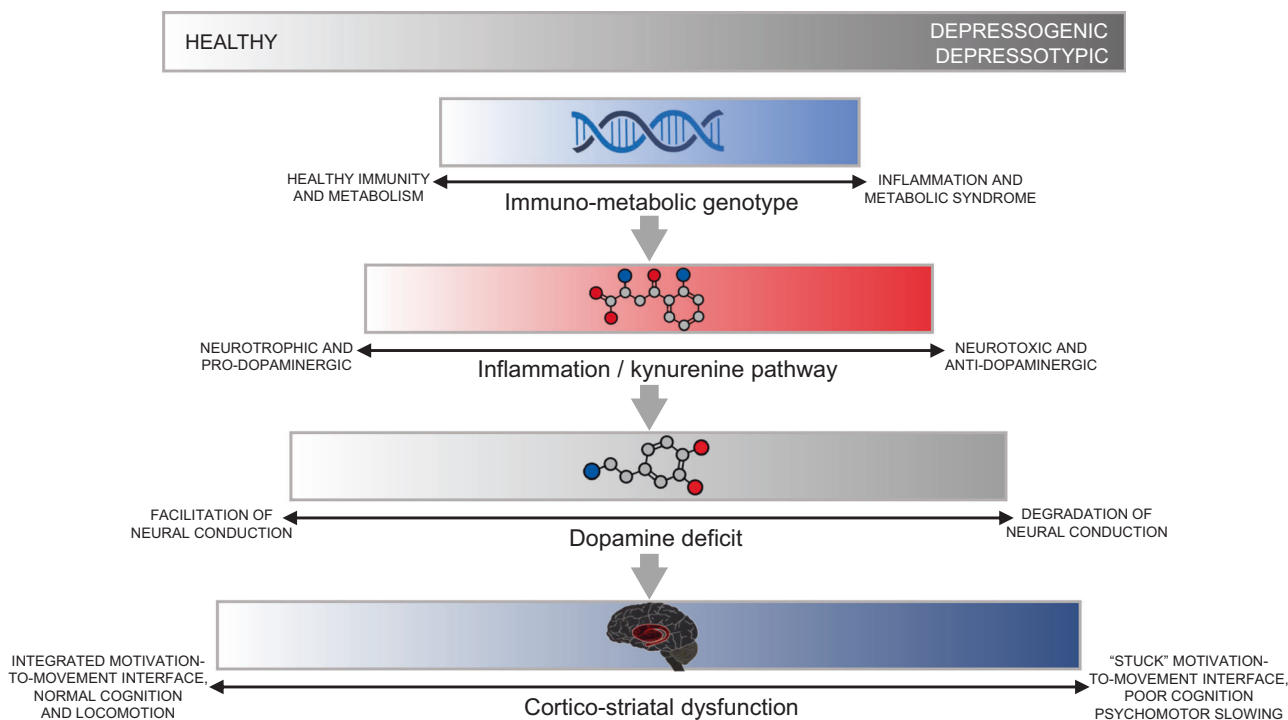


Fig. 3 Summary schematic. Our multilevel model of major depression from genetic to cytokine signaling to neurotransmitter to neural-system levels.

by negative reinforcement [90]. Consistent with this functional role of the habenula, aberrant habenula volume [91] and functioning [92, 93] have been observed in MDD.

SUMMARY, LIMITATIONS, AND FUTURE DIRECTIONS

Here we present a narrative connecting abnormalities from genetics to metabolic and cytokine and neurotransmitter signaling to the neural-systems level in major depression. See Fig. 3 for a summary schematic of our synthesis. The connections between these levels derive from large-scale and/or replicated investigations as well as from theoretical architectures. While the potential for such cross-level integration is encouraging, the account we present is far from comprehensive and final. For example, the connections we detail between the polygenetic signature of MDD and metabolic and immunological dysfunction extend from genetic correlations between these domains. We lack a stepwise, functional-genomics description of this link detailing the genetic-proteomic pathways involved. Moreover, much of the formulation we present connecting immunological abnormalities to dopaminergic signaling is based on pre-clinical models and awaits confirmation in humans. Further, the literature linking dopaminergic abnormalities in MDD with neural-system-level dysfunction is scant and awaits replication. Finally, while we have been careful to avoid unwarranted assertions of causality here, a hierarchical, multilevel account such as this one cannot help but contain some implied causality along the gene-to-whole-brain trajectory.

The account we present suggests at least as many testable hypotheses as it addresses in the development of a multilevel account of major depression. As we state above, formulations relating immunological alterations and dopamine signaling in MDD require more and better testing. In this context, multimodal radioligand imaging for relating immunological and neurotransmitter levels—tandem translocator protein and raclopride PET imaging, for example—in a case-control design could be useful. Further, our assertions that inflammation causes changes in dopaminergic signaling in the brain are derived only from animal-

model studies. To test this causal hypothesis in humans it would be feasible to conduct an experimental medicine study assessing dopaminergic neurotransmission in individuals during inflammation challenge (e.g., with lipopolysaccharide [94]) versus placebo. Finally, while the immuno-metabolic characterization of depression has been useful, immune and metabolic factors are not completely synonymous [9] and causal investigations of the latter are largely absent. Another potentially valuable line of investigation, therefore, could focus on the development of metabolic challenge paradigms with emphasis on their neural-level effects.

REFERENCES

- Lamers F, Vogelzangs N, Merikangas K, De Jonge P, Beekman A, Penninx B. Evidence for a differential role of HPA-axis function, inflammation and metabolic syndrome in melancholic versus atypical depression. *Mol Psychiatry*. 2013;18:692–9.
- Musil R, Seemüller F, Meyer S, Spellmann I, Adli M, Bauer M, et al. Subtypes of depression and their overlap in a naturalistic inpatient sample of major depressive disorder. *Int J Methods Psychiatr Res*. 2018;27:e1569.
- Drysdale AT, Grosenick L, Downar J, Dunlop K, Mansouri F, Meng Y, et al. Resting-state connectivity biomarkers define neurophysiological subtypes of depression. *Nat Med*. 2017;23:28–38.
- American Psychiatric Association. Diagnostic and statistical manual of mental disorders (DSM-5®). American Psychiatric Pub, San Francisco, CA, USA, 2013.
- Fava M, Rush AJ, Alpert JE, Balasubramani G, Wisniewski SR, Carmin CN, et al. Difference in treatment outcome in outpatients with anxious versus nonanxious depression: a STAR* D report. *Am J Psychiatry*. 2008;165:342–51.
- Clements CC, Karlsson R, Lu Y, Juréus A, Rück C, Andersson E, et al. Genome-wide association study of patients with a severe major depressive episode treated with electroconvulsive therapy. *Mol Psychiatry*. 2021;26:2429–39.
- Lamers F, Beekman A, Van Hemert A, Schoevers R, Penninx B. Six-year longitudinal course and outcomes of subtypes of depression. *Br J Psychiatry*. 2016;208:62–68.
- Lamers F, Cui L, Hickie IB, Roca C, Machado-Vieira R, Zarate CA Jr, et al. Familial aggregation and heritability of the melancholic and atypical subtypes of depression. *J Affect Disord*. 2016;204:241–6.
- Milaneschi Y, Lamers F, Berk M, Penninx BW. Depression heterogeneity and its biological underpinnings: toward immunometabolic depression. *Biol Psychiatry*. 2020;88:369–80.

10. Sullivan PF, Neale MC, Kendler KS. Genetic epidemiology of major depression: review and meta-analysis. *Am J Psychiatry*. 2000;157:1552–62.
11. Sullivan PF, Daly MJ, Ripke S, Lewis CM, Lin DY, Wray NR, et al. A mega-analysis of genome-wide association studies for major depressive disorder. *Mol Psychiatry*. 2013;18:497–511.
12. Howard DM, Adams MJ, Shirali M, Clarke TK, Marioni RE, Davies G, et al. Genome-wide association study of depression phenotypes in UK Biobank identifies variants in excitatory synaptic pathways. *Nat Commun*. 2018;9:1470.
13. Hyde CL, Nagle MW, Tian C, Chen X, Paciga SA, Wendland JR, et al. Identification of 15 genetic loci associated with risk of major depression in individuals of European descent. *Nat Genet*. 2016;48:1031–6.
14. Wray NR, Ripke S, Mattheisen M, Trzaskowski M, Byrne EM, Abdellaoui A, et al. Genome-wide association analyses identify 44 risk variants and refine the genetic architecture of major depression. *Nat Genet*. 2018;50:668–81.
15. Cai N, Bigdeli TB, Kretschmar W, Li Y, Liang J, Song L, et al. Sparsewhole-genome sequencing identifies two loci for major depressive disorder. *Nature*. 2015;523:588.
16. Howard DM, Adams MJ, Clarke TK, Hafferty JD, Gibson J, Shirali M, et al. Genome-wide meta-analysis of depression identifies 102 independent variants and highlights the importance of the prefrontal brain regions. *Nat Neurosci*. 2019;22:343.
17. Reaven G. The metabolic syndrome or the insulin resistance syndrome? Different names, different concepts, and different goals. *Endocrinol Metab Clin N Am*. 2004;33:283–303.
18. Pan A, Keum N, Okereke OI, Sun Q, Kivimaki M, Rubin RR, et al. Bidirectional Association Between Depression and Metabolic Syndrome A systematic review and meta-analysis of epidemiological studies. *Diabetes Care*. 2012;35:1171–80.
19. HOUTEN MV, POSNER BI, KOPRIWA BM, BRAWER JR. Insulin-binding sites in the rat brain: in vivo localization to the circumventricular organs by quantitative radioautography. *Endocrinology*. 1979;105:666–73.
20. Begg DP. Insulin transport into the brain and cerebrospinal fluid. *Vitam Horm*. 2015;98:229–48.
21. Bruning JC, Gautam D, Burks DJ, Gillette J, Schubert M, Orban PC, et al. Role of brain insulin receptor in control of body weight and reproduction. *Science*. 2000;289:2122–5.
22. Tschritter O, Preissl H, Hennige AM, Stumvoll M, Porubska K, Frost R, et al. The cerebrocortical response to hyperinsulinemia is reduced in overweight humans: a magnetoencephalographic study. *Proc Natl Acad Sci USA*. 2006;103:12103–8.
23. Hallschmid M, Benedict C, Schultes B, Born J, Kern W. Obese men respond to cognitive but not to catabolic brain insulin signaling. *Int J Obes*. 2008;32:275–82.
24. Benedict C, Hallschmid M, Hatke A, Schultes B, Fehm HL, Born J, et al. Intranasal insulin improves memory in humans. *Psychoneuroendocrinology*. 2004;29:1326–34.
25. Cruz-Pereira JS, Rea K, Nolan YM, O'Leary OF, Dinan TG, Cryan JF. Depression's unholy trinity: dysregulated stress, immunity, and the microbiome. *Annu Rev Psychol*. 2020;71:49–78.
26. Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol*. 2015;11:577–91.
27. Fechner A, Kiehltopf M, Jahreis G. The formation of short-chain fatty acids is positively associated with the blood lipid-lowering effect of lupin kernel fiber in moderately hypercholesterolemic adults. *J Nutr*. 2014;144:599–607.
28. Milaneschi Y, Arnold M, Kastenmüller G, Dehkordi SM, Krishnan RR, Dunlop BW, et al. Genomics-based identification of a potential causal role for acylcarnitine metabolism in depression. *J Affect Disord*. 2022;307:254–63.
29. Capuron L, Su S, Miller AH, Bremner JD, Goldberg J, Vogt GJ, et al. Depressive symptoms and metabolic syndrome: is inflammation the underlying link? *Biol Psychiatry*. 2008;64:896–900.
30. Saper CB, Romanovsky AA, Scammell TE. Neural circuitry engaged by prostaglandins during the sickness syndrome. *Nat Neurosci*. 2012;15:1088–95.
31. Conversano C, Carmassi C, Carlini M, Casu G, Gremigni P, Dell'Osso L. Interferon alpha therapy in patients with chronic hepatitis C infection: quality of life and depression. *Hematol Rep*. 2015;7:5632.
32. Euesden J, Danese A, Lewis CM, Maughan B. A bidirectional relationship between depression and the autoimmune disorders - new perspectives from the National Child Development Study. *PLoS ONE*. 2017;12:e0173015.
33. Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK, et al. A meta-analysis of cytokines in major depression. *Biol Psychiatry*. 2010;67:446–57.
34. Enache D, Pariante CM, Mondelli V. Markers of central inflammation in major depressive disorder: a systematic review and meta-analysis of studies examining cerebrospinal fluid, positron emission tomography and post-mortem brain tissue. *Brain Behav Immun*. 2019;81:24–40.
35. Kohler CA, Freitas TH, Maes M, de Andrade NQ, Liu CS, Fernandes BS, et al. Peripheral cytokine and chemokine alterations in depression: a meta-analysis of 82 studies. *Acta Psychiatr Scand*. 2017;135:373–87.
36. Köhler O, Benros ME, Nordentoft M, Farkouh ME, Iyengar RL, Mors O, et al. Effect of anti-inflammatory treatment on depression, depressive symptoms, and adverse effects: a systematic review and meta-analysis of randomized clinical trials. *JAMA Psychiatry*. 2014;71:1381–91.
37. Nikkheslat N. Targeting inflammation in depression: ketamine as an anti-inflammatory antidepressant in psychiatric emergency. *Brain Behav Immun Health*. 2021;18:100383.
38. Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW. From inflammation to sickness and depression: when the immune system subjugates the brain. *Nat Rev Neurosci*. 2008;9:46–56.
39. Kitagami T, Yamada K, Miura H, Hashimoto R, Nabeshima T, Ohta T. Mechanism of systemically injected interferon-alpha impeding monoamine biosynthesis in rats: role of nitric oxide as a signal crossing the blood-brain barrier. *Brain Res*. 2003;978:104–14.
40. Morón JA, Zakharova I, Ferrer JV, Merrill GA, Hope B, Lafer EM, et al. Mitogen-activated protein kinase regulates dopamine transporter surface expression and dopamine transport capacity. *J Neurosci*. 2003;23:8480–8.
41. Klawonn AM, Fritz M, Castany S, Pignatelli M, Canal C, Simila F, et al. Microglial activation elicits a negative affective state through prostaglandin-mediated modulation of striatal neurons. *Immunity*. 2021;54:225–34 e226.
42. Fritz M, Klawonn AM, Nilsson A, Singh AK, Zajdel J, Wilhelms DB, et al. Prostaglandin-dependent modulation of dopaminergic neurotransmission elicits inflammation-induced aversion in mice. *J Clin Investig*. 2016;126:695–705.
43. Miller AH, Raison CL. The role of inflammation in depression: from evolutionary imperative to modern treatment target. *Nat Rev Immunol*. 2016;16:22–34.
44. Grohmann U, Fallarino F, Puccetti P. Tolerance, DCs and tryptophan: much ado about IDO. *Trends Immunol*. 2003;24:242–8.
45. Savitz J, Drevets WC, Smith CM, Victor TA, Wurfel BE, Bellgowan PS, et al. Putative neuroprotective and neurotoxic kynurenine pathway metabolites are associated with hippocampal and amygdalar volumes in subjects with major depressive disorder. *Neuropsychopharmacology*. 2015;40:463–71.
46. Schwarcz R, Bruno JP, Muchowski PJ, Wu HQ. Kynurenines in the mammalian brain: when physiology meets pathology. *Nat Rev Neurosci*. 2012;13:465–77.
47. Guillemin GJ, Kerr SJ, Smythe GA, Smith DG, Kapoor V, Armati PJ, et al. Kynurenine pathway metabolism in human astrocytes: a paradox for neuronal protection. *J Neurochem*. 2001;78:842–53.
48. Steiner J, Walter M, Gos T, Guillemin GJ, Bernstein HG, Sarnyai Z, et al. Severe depression is associated with increased microglial quinolinic acid in subregions of the anterior cingulate gyrus: evidence for an immune-modulated glutamatergic neurotransmission? *J Neuroinflammation*. 2011;8:94.
49. Brown SJ, Brown AM, Purves-Tyson TD, Huang X-F, Weickert CS, Newell KA. Alterations in the kynurenine pathway and excitatory amino acid transporter-2 in depression with and without psychosis: Evidence of a potential astrocyte pathology. *J Psychiatr Res*. 2022;147:203–11.
50. Guillemin GJ. Quinolinic acid, the inescapable neurotoxin. *FEBS J*. 2012;279:1356–65.
51. Stone TW. Development and therapeutic potential of kynurenine acid and kynurenine derivatives for neuroprotection. *Trends Pharmacol Sci*. 2000;21:149–54.
52. Anis NA, Berry SC, Burton NR, Lodge D. The dissociative anaesthetics, ketamine and phencyclidine, selectively reduce excitation of central mammalian neurones by N-methyl-aspartate. *Br J Pharmacol*. 1983;79:565–75.
53. Beninger RJ, Colton AM, Ingles JL, Jhamandas K, Boegman RJ. Picolinic acid blocks the neurotoxic but not the neuroexcitatory properties of quinolinic acid in the rat brain: evidence from turning behaviour and tyrosine hydroxylase immunohistochemistry. *Neuroscience*. 1994;61:603–12.
54. Doolin K, Allers KA, Pleiner S, Liesener A, Farrell C, Tozzi L, et al. Altered tryptophan catabolite concentrations in major depressive disorder and associated changes in hippocampal subfield volumes. *Psychoneuroendocrinology*. 2018;95:8–17.
55. Colle R, Masson P, Verstuyft C, Feve B, Werner E, Boursier-Neyret C, et al. Peripheral tryptophan, serotonin, kynurenine, and their metabolites in major depression: a case-control study. *Psychiatry Clin Neurosci*. 2020;74:112–7.
56. Savitz J, Drevets WC, Wurfel BE, Ford BN, Bellgowan PS, Victor TA, et al. Reduction of kynurenine acid to quinolinic acid ratio in both the depressed and remitted phases of major depressive disorder. *Brain Behav Immun*. 2015;46:55–59.
57. Paul ER, Schwieler L, Erhardt S, Boda S, Trepci A, Kampe R, et al. Peripheral and central kynurenine pathway abnormalities in major depression. *Brain Behav Immun*. 2022;101:136–45.
58. Oxenkrug GF. Metabolic syndrome, age-associated neuroendocrine disorders, and dysregulation of tryptophan-kynurenine metabolism. *Ann N Y Acad Sci*. 2010;1199:1–14.
59. Roberts RC, Ahn A, Swartz KJ, Beal MF, DiFiglia M. Intrastratial injections of quinolinic acid or kainic acid: differential patterns of cell survival and the effects of data analysis on outcome. *Exp Neurol*. 1993;124:274–82.
60. Randrup A, Munkvad I, Fog R. Mental and behavioural stereotypies elicited by stimulant drugs. relation to the dopamine hypothesis of schizo-phrenia, mania

- and depression. In: *Recent advances in neuropsychopharmacology*. Elsevier, Göteborg, Sweden; 1981. p. 63–74.
61. Dunlop BW, Nemeroff CB. The role of dopamine in the pathophysiology of depression. *Arch Gen Psychiatry*. 2007;64:327–37.
 62. Hamilton JP, Sacchet MD, Hjørnevik T, Chin FT, Shen B, Kämpfe R, et al. Striatal dopamine deficits predict reductions in striatal functional connectivity in major depression: a concurrent 11 C-raclopride positron emission tomography and functional magnetic resonance imaging investigation. *Transl Psychiatry*. 2018;8:1–10.
 63. Choi EY, Yeo BT, Buckner RL. The organization of the human striatum estimated by intrinsic functional connectivity. *J Neurophysiol*. 2012;108:2242–63.
 64. Peciña M, Sikora M, Avery ET, Heffernan J, Peciña S, Mickey BJ, et al. Striatal dopamine D2/3 receptor-mediated neurotransmission in major depression: implications for anhedonia, anxiety and treatment response. *Eur Neuropsychopharmacol*. 2017;27:977–86.
 65. Meyer JH, McNeely HE, Sagrati S, Boovariwala A, Martin K, Verhoeff NPL, et al. Elevated putamen D2 receptor binding potential in major depression with motor retardation: an [11C] raclopride positron emission tomography study. *Am J Psychiatry*. 2006;163:1594–602.
 66. Montgomery AJ, Stokes P, Kitamura Y, Grasby PM. Extrastriatal D2 and striatal D2 receptors in depressive illness: pilot PET studies using [11C] FLB 457 and [11C] raclopride. *J Affect Disord*. 2007;101:113–22.
 67. Hirvonen J, Karlsson H, Kajander J, Markkula J, Rasi-Hakala H, Någren K, et al. Striatal dopamine D2 receptors in medication-naïve patients with major depressive disorder as assessed with [11C] raclopride PET. *Psychopharmacology*. 2008;197:581–90.
 68. Martinez D, Slifstein M, Broft A, Mawlawi O, Hwang D-R, Huang Y, et al. Imaging human mesolimbic dopamine transmission with positron emission tomography. Part II: amphetamine-induced dopamine release in the functional subdivisions of the striatum. *J Cereb Blood Flow Metab*. 2003;23:285–300.
 69. Kaasinen V, Vahlberg T, Stoessl AJ, Strafella AP, Antonini A. Dopamine receptors in Parkinson's disease: a meta-analysis of imaging studies. *Mov Disord*. 2021;36:1781–91.
 70. Parsey RV, Oquendo MA, Zea-Ponce Y, Rodenhiser J, Kegeles LS, Pratap M, et al. Dopamine D2 receptor availability and amphetamine-induced dopamine release in unipolar depression. *Biol Psychiatry*. 2001;50:313–22.
 71. Ogawa S, Tsuchimine S, Kunugi H. Cerebrospinal fluid monoamine metabolite concentrations in depressive disorder: A meta-analysis of historic evidence. *J Psychiatr Res*. 2018;105:137–46.
 72. Romeo B, Blecha L, Locatelli K, Benyamina A, Martelli C. Meta-analysis and review of dopamine agonists in acute episodes of mood disorder: efficacy and safety. *J Psychopharmacol*. 2018;32:385–96.
 73. Kemp JM, Powell TPS. The connexions of the striatum and globus pallidus: synthesis and speculation. *Philos Trans R Soc Lond B Biol Sci*. 1971;262:441–57.
 74. DeLong M, Georgopoulos A, Crutcher M. Cortico-basal ganglia relations and coding of motor performance. *Exp Brain Res*. 1983;49:30–40.
 75. Alexander GE, DeLong MR, Strick PL. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu Rev Neurosci*. 1986;9:357–81.
 76. Calzavara R, Maily P, Haber SN. Relationship between the corticostriatal terminals from areas 9 and 46, and those from area 8A, dorsal and rostral premotor cortex and area 24c: an anatomical substrate for cognition to action. *Eur J Neurosci*. 2007;26:2005–24.
 77. Gaspar P, Stepniowska I, Kaas J. Topography and collateralization of the dopaminergic projections to motor and lateral prefrontal cortex in owl monkeys. *J Comp Neurol*. 1992;325:1–21.
 78. Alexander GE, Crutcher MD. Functional architecture of basal ganglia circuits: neural substrates of parallel processing. *Trends Neurosci*. 1990;13:266–71.
 79. Haber SN. The place of dopamine in the cortico-basal ganglia circuit. *Neuroscience*. 2014;282:248–57.
 80. Bouras C, Schulz P, Constantinidis J, Tissot R. Differential effects of acute and chronic administration of haloperidol on substance P and enkephalins in diverse rat brain areas. *Neuropsychobiology*. 1986;16:169–74.
 81. Hong JS, Yoshikawa K, Kanamatsu T, Sabol SL. Modulation of striatal enkephalinergic neurons by antipsychotic drugs. *Fed Proc*. 1985;44:2535–9.
 82. Pan HS, Penney JB, Young AB. γ -Aminobutyric acid and benzodiazepine receptor changes induced by unilateral 6-hydroxydopamine lesions of the medial forebrain bundle. *J Neurochem*. 1985;45:1396–404.
 83. Young WS, Bonner TI, Brann MR. Mesencephalic dopamine neurons regulate the expression of neuropeptide mRNAs in the rat forebrain. *Proc Natl Acad Sci USA*. 1986;83:9827–31.
 84. Bonci A, Hopf FW. The dopamine D2 receptor: new surprises from an old friend. *Neuron*. 2005;47:335–8.
 85. Urs NM, Peterson SM, Caron MG. New concepts in dopamine D2 receptor biased signaling and implications for schizophrenia therapy. *Biol Psychiatry*. 2017;81:78–85.
 86. Donhamsetti P, Gallo EF, Buck DC, Stahl EL, Zhu Y, Lane JR, et al. Arrestin recruitment to dopamine D2 receptor mediates locomotion but not incentive motivation. *Mol Psychiatry*. 2020;25:2086–2100.
 87. Hamilton JP, Etkin A, Furman DJ, Lemus MG, Johnson RF, Gotlib IH. Functional neuroimaging of major depressive disorder: a meta-analysis and new integration of baseline activation and neural response data. *Am J Psychiatry*. 2012;169:693–703.
 88. Schwarz KA, Wieser MJ, Gerdes AB, Mühlberger A, Pauli P. Why are you looking like that? How the context influences evaluation and processing of human faces. *Soc Cogn Affect Neurosci*. 2013;8:438–45.
 89. Christoph GR, Leonzio RJ, Wilcox KS. Stimulation of the lateral habenula inhibits dopamine-containing neurons in the substantia nigra and ventral tegmental area of the rat. *J Neurosci*. 1986;6:613–9.
 90. Matsumoto M, Hikosaka O. Lateral habenula as a source of negative reward signals in dopamine neurons. *Nature*. 2007;447:1111–5.
 91. Savitz JB, Nugent AC, Bogers W, Roiser JP, Bain EE, Neumeister A, et al. Habenula volume in bipolar disorder and major depressive disorder: a high-resolution magnetic resonance imaging study. *Biol Psychiatry*. 2011;69:336–43.
 92. Furman DJ, Gotlib IH. Habenula responses to potential and actual loss in major depression: preliminary evidence for lateralized dysfunction. *Soc Cogn Affect Neurosci*. 2016;11:843–51.
 93. Lawson RP, Nord CL, Seymour B, Thomas DL, Dayan P, Pilling S, et al. Disrupted habenula function in major depression. *Mol Psychiatry*. 2017;22:202–8.
 94. Zielen S, Trischler J, Schubert R. Lipopolysaccharide challenge: immunological effects and safety in humans. *Expert Rev Clin Immunol*. 2015;11:409–18.

AUTHOR CONTRIBUTIONS

JPH and ERP constructed the primary narrative of the review. LÖ, MH, and HSB provided section-specific content and revision advice for the primary narrative.

FUNDING

Open access funding provided by University of Bergen.

COMPETING INTERESTS

HM consults to Abbott Labs, Blackrock Neurotech and Cogwear and receives intellectual licensing fees from Abbott Labs. MH has received consulting fees, research support or other compensation from Indivior, Camurus, BrainsWay, Aelis Farma, and Janssen Pharmaceuticals. Neither HM's nor MH's competing interests are relevant to the present work. ERP, LÖ, and JPH have no competing interests to report.

ADDITIONAL INFORMATION

Correspondence and requests for materials should be addressed to J. Paul Hamilton.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2023