

Pathophysiology and pharmacology of G protein-coupled receptors in the heart

Alyssa Grogan ¹, Emilio Y. Lucero ¹, Haoran Jiang ¹, and Howard A. Rockman ^{1,2,*}

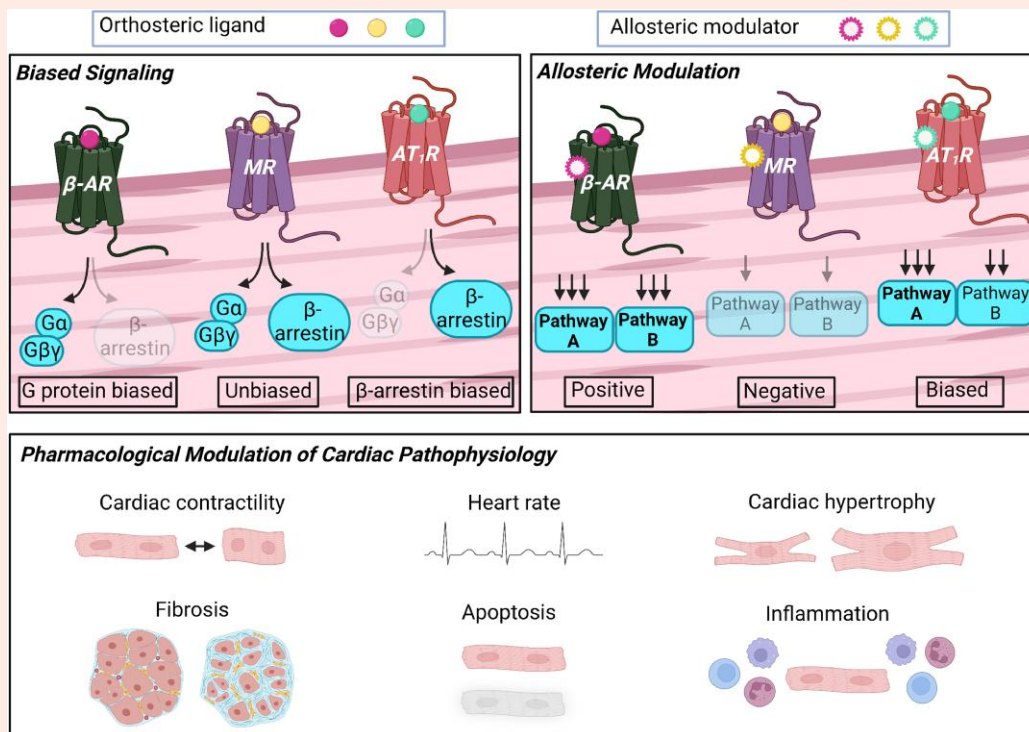
¹Department of Medicine, Duke University Medical Center, DUMC 3104, 226 CARL Building, Durham, NC 27710, USA; and ²Cell Biology, Duke University Medical Center, DUMC 3104, 226 CARL Building, 12 Durham, NC 27710, USA

Received 19 August 2022; revised 30 September 2022; accepted 6 October 2022; online publish-ahead-of-print 22 November 2022

Abstract

G protein-coupled receptors (GPCRs), comprising the largest superfamily of cell surface receptors, serve as fundamental modulators of cardiac health and disease owing to their key roles in the regulation of heart rate, contractile dynamics, and cardiac function. Accordingly, GPCRs are heavily pursued as drug targets for a wide variety of cardiovascular diseases ranging from heart failure, cardiomyopathy, and arrhythmia to hypertension and coronary artery disease. Recent advancements in understanding the signalling mechanisms, regulation, and pharmacological properties of GPCRs have provided valuable insights that will guide the development of novel therapeutics. Herein, we review the cellular signalling mechanisms, pathophysiological roles, and pharmacological developments of the major GPCRs in the heart, highlighting the β -adrenergic, muscarinic, and angiotensin receptors as exemplar subfamilies.

Graphical Abstract



Keywords

G protein-coupled receptors • Heart failure • Biased signalling • Allosteric modulators

* Corresponding author. Tel: +919 668 2520; fax: +919 668 2524, E-mail: h.rockman@duke.edu

1. Introduction

G protein-coupled receptors (GPCRs), also known as 7 transmembrane domain receptors (7TMRs), encompass the largest and most extensively studied superfamily of cell surface receptors.¹ GPCRs are activated by a diverse array of ligands including hormones, peptides, and neurotransmitters, and serve as key regulators of a variety of cellular responses. Given their involvement in many different physiological processes, GPCRs are highly pursued pharmacologically and represent the primary targets of ~35% of all small molecule drugs currently approved by the Food and Drug Administration (FDA).² Of the nearly 800 different human GPCR genes,³ more than 200 are expressed in the heart alone,⁴ underscoring their prominent role in regulating cardiac function and highlighting their potential as therapeutic targets in heart disease.

1.1 GPCR signalling mechanisms

Canonically, GPCRs are activated via the binding of an agonist to the orthosteric ligand binding site on the extracellular surface (Figure 1). Agonist-bound

GPCRs subsequently recruit heterotrimeric G proteins, consisting of $G\alpha$, $G\beta$, and $G\gamma$ subunits, and induce the exchange of guanosine diphosphate (GDP) for guanosine triphosphate (GTP) on $G\alpha$. This exchange stimulates the dissociation of $G\alpha$ from $G\beta\gamma$, generating two activated G protein units that independently transduce signals to downstream effectors.⁵

The $G\alpha$ subunit is divided into four major classes including $G\alpha_s$, $G\alpha_i$, $G\alpha_q/11$, and $G\alpha_{12/13}$, which each generate their own unique cellular responses (Figure 1).⁵ The $G\alpha_s$ subunit, a stimulatory $G\alpha$ protein, activates adenylyl cyclase (AC) to generate second messenger cyclic adenosine monophosphate (cAMP) which results in the activation of protein kinase A (PKA). In the heart, PKA phosphorylates sarcomeric proteins, calcium regulators, and ion channels resulting in enhanced contractile and calcium cycling dynamics thereby mediating positive inotropic and chronotropic responses.⁶ In contrast, receptors that are coupled to $G\alpha_i$ inhibit AC, which leads to decreased production of cAMP and reduced PKA activity. $G\alpha_q$ -coupled receptors promote the activation of phospholipase C (PLC), which generates the second messengers, namely diacylglycerol (DAG) and inositol triphosphate (IP_3) that stimulate protein kinase C (PKC) and calcium influx, respectively. Finally, $G\alpha_{12/13}$ activates the small GTPase Rho.

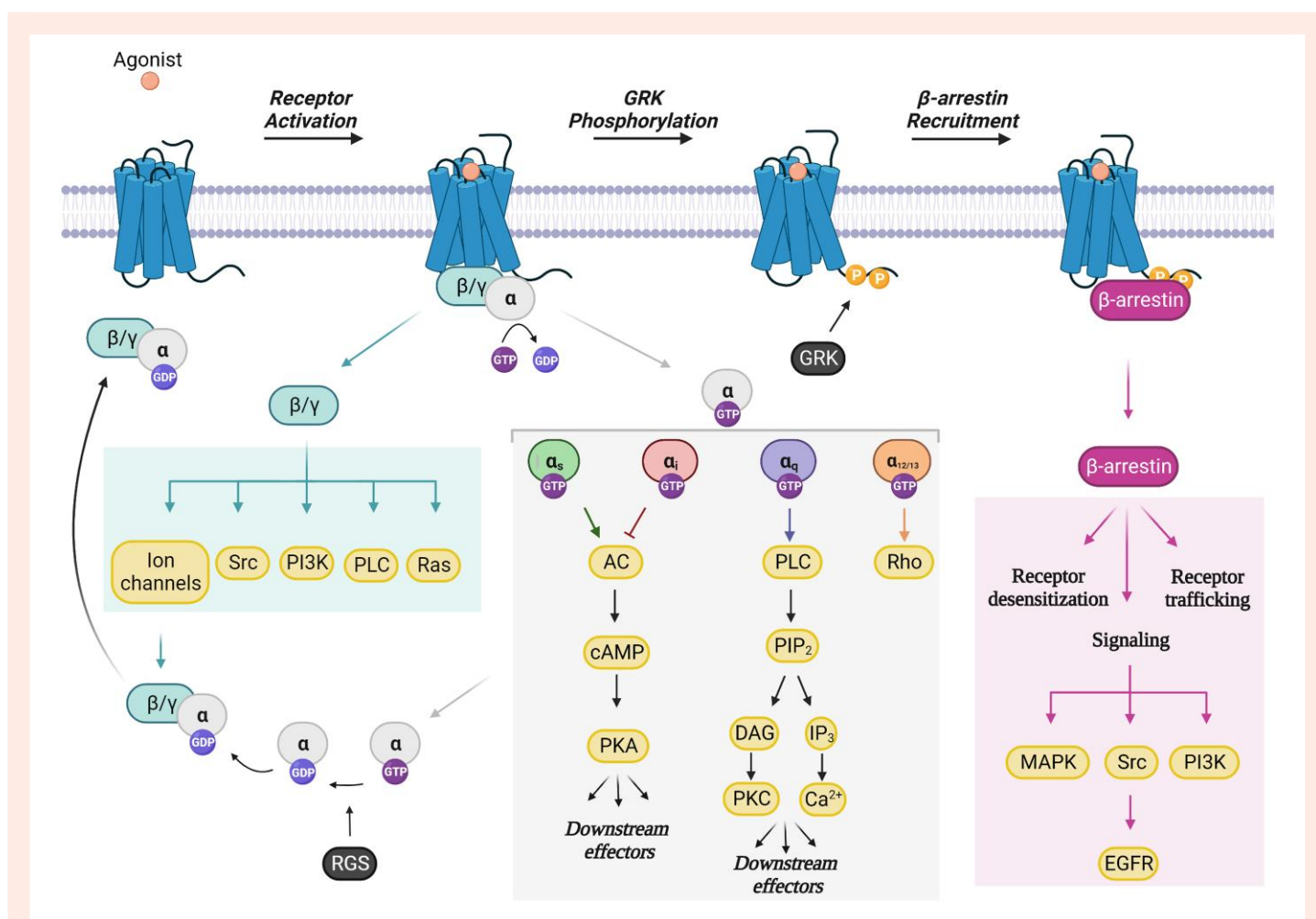


Figure 1 GPCR activation and downstream signalling. Following agonist binding, GPCRs recruit heterotrimeric G proteins ($G\alpha$, $G\beta$, $G\gamma$) and induce the exchange of GDP for GTP on $G\alpha$. The $G\alpha$ and $G\beta\gamma$ subunits subsequently dissociate forming two activated G protein units that independently transduce signals to downstream effectors. The four families of $G\alpha$ ($G\alpha_s$, $G\alpha_i$, $G\alpha_q$, and $G\alpha_{12/13}$) activate distinct signalling cascades, thereby generating unique cellular responses. RGS proteins accelerate the GTPase activity of $G\alpha$, thereby leading to the inactivation of G protein signalling. GDP, guanosine diphosphate; GTP, guanosine triphosphate; PI3K, phosphoinositide 3-kinase; PLC, phospholipase C; AC, adenylyl cyclase; cAMP, cyclic adenosine monophosphate; PKA, protein kinase A; PIP₂, phosphatidylinositol 4,5-bisphosphate; DAG, diacylglycerol; PKC, protein kinase C; IP₃, inositol triphosphate; MAPK, mitogen-activated protein kinase; EGFR, epidermal growth factor receptor; GRK, GPCR kinase; RGS, regulators of G protein signalling. Figure generated with Biorender.com.

To mediate signal termination, regulators of G protein signalling (RGS) and GPCR kinases (GRKs) are recruited to GTP-bound $G\alpha$ subunits or agonist-activated GPCRs, respectively. RGS proteins serve to accelerate $G\alpha$ GTP hydrolysis activity, causing the reassociation of $G\alpha$ with $G\beta\gamma$ and the inactivation of downstream G protein signalling (Figure 1).⁷ In contrast, GRKs are recruited to agonist-activated GPCRs to phosphorylate the COOH-terminal tail of the receptor (Figure 1).⁸ This in turn, facilitates the recruitment of β -arrestin, a multifunctional adaptor protein, which promotes receptor desensitization via sterically uncoupling G proteins from activated GPCRs.⁵ β -arrestins also serve as scaffolds for cyclic nucleotide phosphodiesterases (PDEs) and diacylglycerol kinases (DGKs) that degrade second messengers cAMP and DAG, respectively, providing another mechanism to dampen signalling.^{9,10} In addition to their desensitization function, β -arrestins facilitate receptor internalization and trafficking to endosomes via binding clathrin and its adaptor protein AP2.¹¹ Importantly, β -arrestins function as signal transducers via scaffolding major signalling complexes such as mitogen-activated protein kinases (MAPKs), Src, and phosphoinositide 3-kinase (PI3K).¹²

The discovery that β -arrestins serve as functional signal mediators has led to the emergence of a fundamental concept in GPCR biology termed biased agonism. Compared to the cognate endogenous ligands for each GPCR which activate multiple signalling pathways in an unbiased fashion, biased ligands stabilize distinct receptor conformations to preferentially activate either G protein or β -arrestin pathways (Figure 2A).¹³ The concept of biased agonism provides important implications in the development of therapeutics since drugs can be developed to target specific signalling pathways that will generate more beneficial physiological outcomes and/or reduce adverse side effects.

1.2 Allosteric modulation of GPCR function

Aside from the orthosteric binding region, ligands can target allosteric sites of the receptor (i.e. regions that are topographically distinct from the orthosteric binding pocket). These ligands, termed allosteric modulators, most often do not possess intrinsic activity on their own, but can enhance (positive allosteric modulator) or reduce (negative allosteric modulator) the activity of receptors stimulated by orthosteric ligands (Figure 2B).¹⁴ The identification of allosteric modulators that possess intrinsic activity in the absence of an orthosteric ligand has additionally established 'ago-allosteric modulators' as an entity.¹⁵ Importantly, both allosteric and ago-allosteric modulators can potentially act in a biased fashion by enhancing the activation of a particular signalling pathway while either having no effect or antagonizing alternative pathways (Figure 2B).¹⁶ Given that the orthosteric binding pocket is typically highly conserved among GPCR subtypes, while allosteric regions exhibit greater sequence and structural diversity, allosteric modulators have a higher potential for being subtype-specific and/or generating biased cellular effects.¹⁴

Herein, we review the cellular signalling mechanisms, physiological functions, clinical implications, and current pharmacological developments of the major subtypes of GPCRs that regulate cardiac physiology. Given the large number of different GPCRs expressed in the heart,⁴ we chose to focus on the receptors that have shown the greatest pharmacological potential and/or highest efficacy in the treatment of heart disease, including the β -adrenergic, muscarinic, and angiotensin receptor families.

2. β -Adrenergic receptors

Adrenergic receptors were initially subclassified into two distinct receptor families (α and β) by Ahlquist in 1948.¹⁷ Within the β -adrenergic (β AR) family, there are three major subtypes that include the β_1 AR, β_2 AR, and β_3 AR, which are encoded by the genes *ADRB1*, *ADRB2*, and *ADRB3*, respectively. All three β ARs are endogenously activated by the catecholamine hormones epinephrine and norepinephrine (Figure 3). For the β_1 AR, norepinephrine has a slightly higher potency than epinephrine, while the agonist potency is reversed for the β_2 AR.¹⁸

2.1 Tissue expression and localization

The β_1 AR is the most common subtype in the heart, as it comprises about 80% of total β ARs while the remaining 20% is mainly the β_2 AR.¹⁹ The β_3 AR, in contrast, is the least characterized to date and only constitutes about 3% of all cardiac β ARs.²⁰ Consistent with its increased abundance, the β_1 AR subtype displays a more widespread cellular distribution as it localizes to both the outer plasma membrane and T-tubule membranes of cardiomyocytes,²¹ whereas β_2 AR and β_3 AR are confined to the T-tubular network.^{21,22} In contrast to the healthy heart, failing human hearts exhibit a substantial reduction in β AR receptor density due to selective downregulation of β_1 AR.¹⁹ As a consequence, this shifts the β_1 AR: β_2 AR ratio from 80:20 to 60:40.¹⁹

2.2 Cellular signalling mechanisms

Under normal physiological conditions, the β_1 AR couples to $G\alpha_s$ as the signalling transducer, while the β_2 AR couples to both $G\alpha_s$ and $G\alpha_i$, and the β_3 AR predominantly signals via $G\alpha_i$.^{23,24} In addition to PKA activation via $G\alpha_s$, stimulation of β_1 ARs increases calcium/calmodulin-dependent protein kinase II (CaMKII) activity in a β -arrestin-dependent manner.²⁵ β_2 AR coupling to $G\alpha_i$ in the heart can activate cell survival signalling via $G\beta\gamma$ -mediated activation of PI3K and Akt.²³ Furthermore, the dissociated $G\beta\gamma$ subunit from $G\alpha_i$ can also activate the Src-family tyrosine kinases and G protein Ras.^{26,27}

Although β_3 ARs canonically signal via $G\alpha_i$, functioning to inhibit activation of AC and PKA, administration of the β_3 AR-selective agonist, L755-507, generated positive inotropic responses in transgenic mice overexpressing human β_3 AR.²⁸ Moreover, stimulation with L755-507 resulted in increased AC activity in a pertussis-toxin insensitive fashion, indicating overexpressed β_3 ARs couple to $G\alpha_s$.²⁸ Unlike β_1 ARs and β_2 ARs that have rich serine/threonine residues that can be phosphorylated by GRKs, β_3 ARs lack these residues and therefore are more resistant to agonist-induced desensitization.²⁴

Aside from their desensitization and internalization functions, β -arrestins also serve as signal transducers following β AR activation. Specifically, in response to an agonist, β -arrestin recruits c-Src to β AR, which subsequently induces the activation of extracellular signal-regulated kinases (ERK).²⁹ Furthermore, β -arrestin-mediated c-Src stimulation leads to the activation of matrix metalloproteases (MMPs), which in turn, initiate epidermal growth factor receptor (EGFR) transactivation.³⁰ Importantly, *in vivo*, β AR-mediated EGFR transactivation confers cardioprotection by preventing the development of ventricular dilation and contractile impairment under conditions of chronic catecholamine stimulation.³⁰

2.3 Physiological functions and contribution to disease

Increased sympathetic nervous system activation and the adrenergic response are essential in maintaining circulatory support during acute heart failure, yet are associated with worse survival.³¹ Indeed, prolonged activation of catecholamine signalling is detrimental to the heart since this causes β AR desensitization, excessive CaMKII activation, and cardiomyocyte hypertrophy and apoptosis.^{25,32-34} Consistent with this, transgenic mice overexpressing β_1 AR exhibit depressed cardiac function, progressive hypertrophy, and fibrosis.^{35,36}

Similarly, mice with cardiac-specific overexpression of β_2 AR display severe left ventricular dysfunction following aortic stenosis,³⁷ and spontaneous tachyarrhythmia related to increased interstitial fibrosis.³⁸ Furthermore, β_2 AR overexpressing mice are more susceptible to ischaemic injury than their wild-type counterparts, which is further exacerbated by pertussis-toxin treatment suggesting that the pathological manifestations of β_2 AR overexpression are mediated primarily by $G\alpha_s$ rather than $G\alpha_i$.³⁹ Indeed, β_2 AR activation is considered anti-apoptotic through its ability to couple to $G\alpha_i$.^{40,41} Therefore, β_2 AR signalling may be either protective or deleterious in the heart depending on transducer coupling.

In addition to the evaluation of β AR overexpression, the generation of different β AR knockout mouse models revealed that the absence of β AR is also pathogenic. Particularly, β_1 AR knockout mice (β_1 AR^{-/-}) die

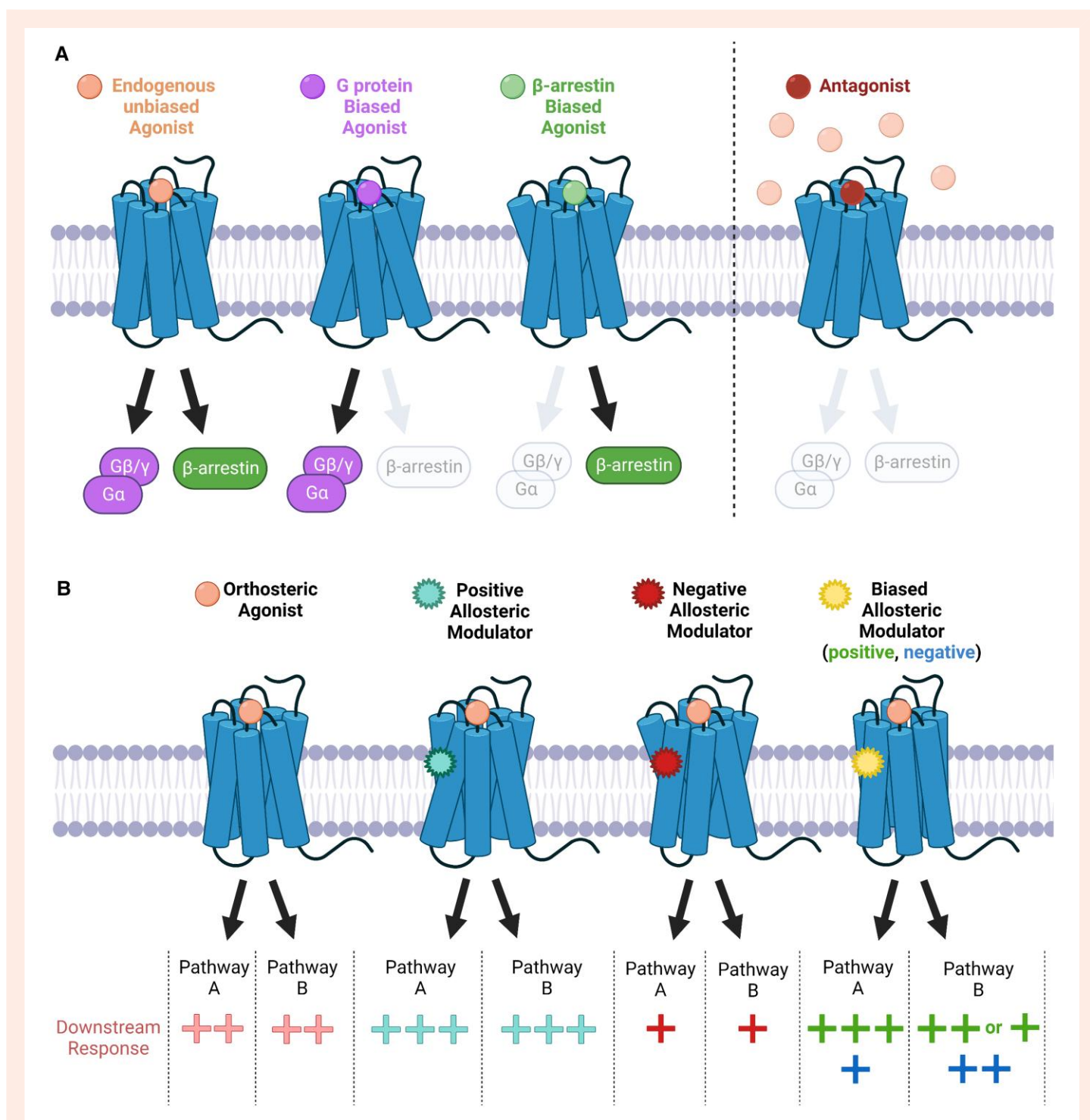


Figure 2 Biased agonism and allosteric modulation of GPCR function. (A) Unbiased orthosteric agonists non-selectively activate G protein and β -arrestin signalling pathways, while biased orthosteric ligands preferentially activate one pathway over the other. Conversely, antagonists competitively inhibit agonist binding and thereby inactivate both G protein and β -arrestin signalling in a balanced fashion. (B) Allosteric modulators bind to sites that are topographically distinct from the orthosteric binding pocket of the receptor. Functionally, positive allosteric modulators enhance (+++) the activity of an orthosteric agonist, while negative allosteric modulators reduce (+) orthosteric activity. Biased allosteric modulators can preferentially enhance (positive; +++) or reduce (negative; +) the activity of a particular pathway (i.e. pathway A) while having no effect (++) and/or antagonizing (+) an alternative pathway (i.e. pathway B) in the presence of an orthosteric agonist. Figure generated with Biorender.com.

prenatally, and those that do survive lack chronotropic or inotropic responses after administration of isoproterenol.⁴² Although endogenous β_2 ARs are present in β_1 AR^{-/-} hearts, they do not functionally compensate

for the absence of β_1 ARs, suggesting that the β_1 AR subtype predominantly mediates the chronotropic/inotropic response in the heart.⁴² As further evidence for this, while β_1 AR/ β_2 AR double knockout and β_1 AR^{-/-} mice

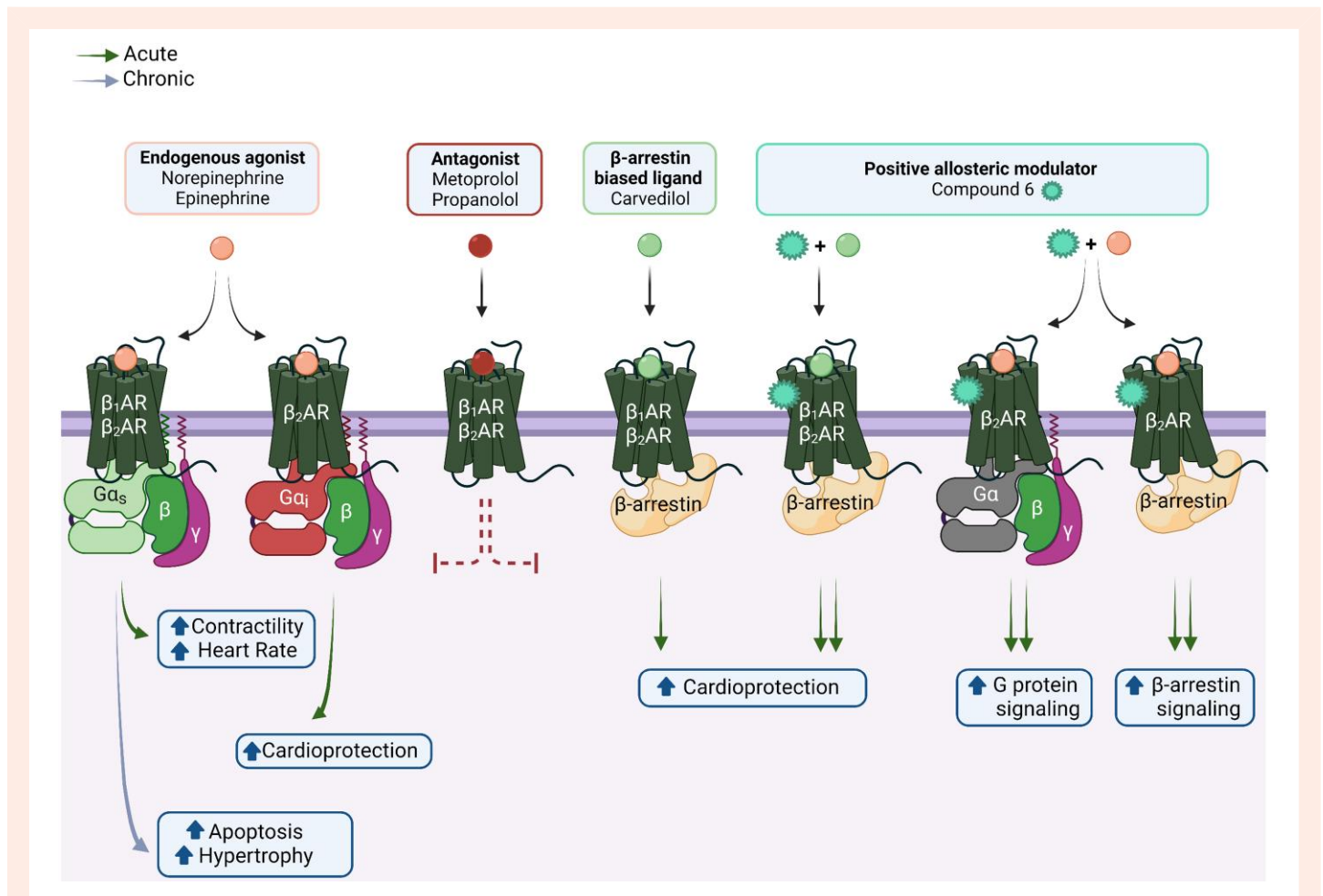


Figure 3 Physiological and pathological effects of cardiac β AR signalling. Norepinephrine and epinephrine, the endogenous β AR agonists, activate both β_1 AR and β_2 AR subtypes, albeit with different potencies. In general, β_1 AR is coupled to $G\alpha_s$ and β_2 AR is coupled to both $G\alpha_s$ and $G\alpha_i$. In the short term, the activation of β_1 AR/ β_2 AR through $G\alpha_s$ generates positive chronotropic and inotropic responses; however, chronic activation through $G\alpha_s$ can induce cardiomyocyte apoptosis and hypertrophy. In contrast, $G\alpha_i$ -mediated signalling via β_2 AR is thought to be cardioprotective due to its anti-apoptotic and anti-fibrotic effects. Furthermore, both receptor subtypes can couple to β -arrestin, which also induces cardioprotective signalling cascades in the heart. Carvedilol, a clinically utilized β -arrestin-biased ligand, preferentially stimulates β -arrestin signalling while antagonizing G protein signalling. Notably, carvedilol-mediated β -arrestin-dependent cardioprotection is potentiated by Compound 6, a positive allosteric modulator of the β_1 AR and β_2 AR. Additionally, in response to agonist, Compound 6 potentiates G protein and β -arrestin signalling mediated by β_2 AR, but not β_1 AR. As expected, traditional antagonists such as metoprolol or propranolol, block β AR signalling via both G protein and β -arrestin. Figure generated with biorender.com.

lack heart rate and contractile responses to catecholamine infusion,^{43,44} adult β_2 AR^{-/-} mice respond normally.^{43,45} Moreover, progressive cardiac dysfunction following myocardial infarction is attenuated in β_1 AR^{-/-} and β_1 AR/ β_2 AR double knockout mice, but not β_2 AR^{-/-} mice, demonstrating that regulation of cardiac function in normal hearts and following injury is predominantly mediated through β_1 ARs.⁴³

On the other hand, the role of β_3 AR in cardiac function is relatively unclear. Several studies have reported that pharmacological activation and/or overexpression of β_3 ARs reduced contractility in healthy hearts,^{46–48} while others have suggested that β_3 AR overexpression will augment contractility.²⁸ Moreover, overexpression or selective stimulation of β_3 AR by BRL 37 344 is protective against cardiac remodelling and dysfunction in response to pressure overload, myocardial infarction, or neurohormonal stimulation through a nitric oxide synthase (NOS)-mediated pathway.^{49–51} However, other studies have revealed that β_3 AR antagonism via administration of L-748 337 improved the cardiac function and exercise performance in a canine model of pacing-induced heart failure.⁵² Therefore, additional studies are needed to further interrogate the potential protective or detrimental impacts of β_3 AR in the heart.

2.4 Pharmacological perspectives

β -Blockers competitively antagonize the binding of endogenous catecholamines to the orthosteric site of β ARs and are a mainstay in the treatment of chronic heart failure. Among the many different clinically utilized β -blockers,⁵³ carvedilol possesses unique properties by virtue of its ability to recruit and activate cardioprotective β -arrestin signalling at both the β_1 AR and β_2 AR.^{54–57} Specifically, carvedilol stabilizes β ARs in a particular conformation that initiates β -arrestin-mediated signalling while remaining uncoupled from $G\alpha_s$.⁵⁴ While the evidence that carvedilol is more effective as a therapeutic agent comes exclusively from experimental studies, a clinical trial revealed that carvedilol was superior to metoprolol in reducing all-cause mortality in patients with heart failure.⁵⁸

Although β -blockers effectively reduce mortality, patients often experience side effects including fatigue and reduced functional capacity, which may limit their maximal effectiveness.⁵⁹ This has led to an effort to identify new β AR ligands, particularly β -arrestin-biased ligands⁶⁰ or allosteric modulators, aimed at preferentially enhancing the beneficial effects and minimizing the adverse effects of overall β -blockade.⁶¹ Recently, a positive allosteric

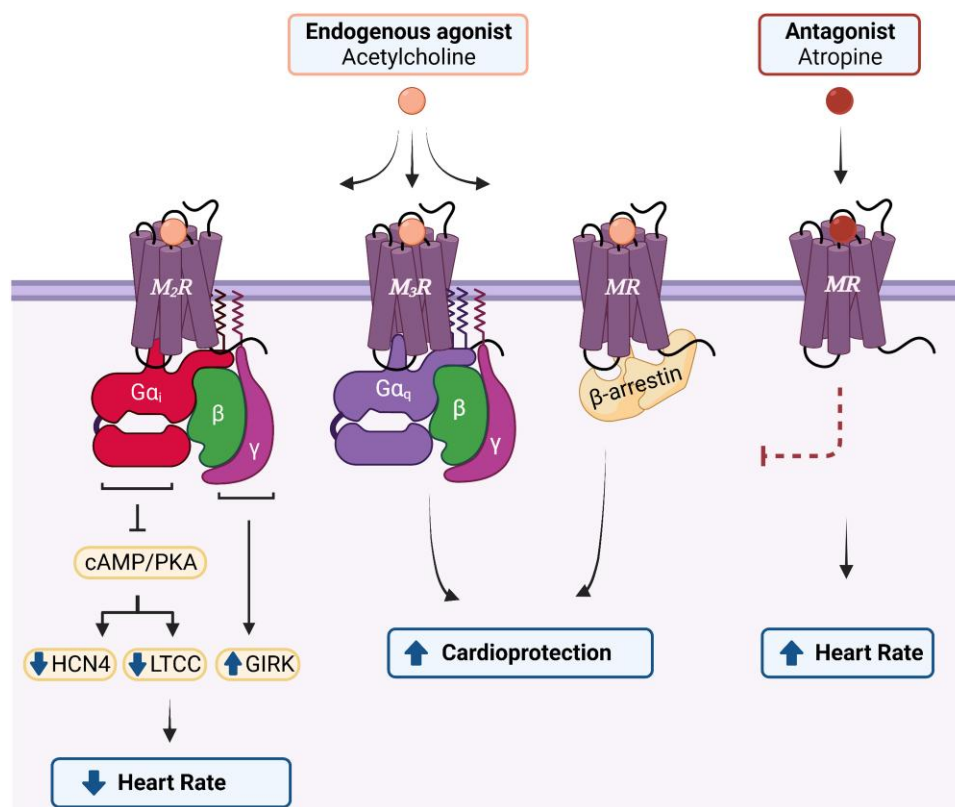


Figure 4 Physiological effects of cardiac MR signalling. The M_2R and M_3R , depicted herein as representative MR subtypes given that their physiological roles in the heart are the most extensively characterized, signal via distinct G protein classes. The M_2R , coupled to $G\alpha_i$, is the most abundant cardiac subtype and negatively regulates heart rate in response to the endogenous agonist, acetylcholine. In contrast, activation of M_3R , coupled with $G\alpha_q$, is considered cardioprotective due to its anti-arrhythmic/anti-fibrotic properties and protection against ischaemic injury. All MR subtypes couple to β -arrestin following agonist binding which initiates cardioprotective signalling cascades in the heart. In response to an antagonist, such as atropine, MR signalling is blocked and heart rate is increased. HCN4, hyperpolarization-activated cyclic nucleotide-gated 4 channel; LTCC, L-type Ca^{2+} channel; PKA, protein kinase A; GIRK, G protein-coupled inward rectifying K^+ channel. Figure generated with biorender.com.

modulator, Compound 6, was identified through a DNA-encoded small molecule library screen against the β_2AR .⁶² Strikingly, Compound 6 increases the binding affinity of carvedilol for β_1ARs ⁵⁷ and β_2ARs ,⁶³ potentiates carvedilol-stimulated β -arrestin-dependent signalling,^{57,63} and provides enhanced cardioprotection to ischaemic injury.⁵⁷

3. Muscarinic receptors

Muscarinic receptors (MRs) comprise a family of GPCRs that are activated by the principal neurotransmitter of the parasympathetic nervous system, acetylcholine (Figure 4). The five major subtypes, M_1R through M_5R , encoded by genes *CHRM-1* to *CHRM-5*, respectively, range in size from ~460–590 amino acids in length and exhibit significant sequence homology to each other.^{64,65} Given their ubiquitous expression across all organs,⁶⁶ they are implicated in a wide array of physiological processes ranging from the regulation of cognitive, behavioural, sensory, motor, and autonomic functions mediated by MRs in the central nervous system (CNS), to the regulation of heart rate, smooth-muscle contraction, and glandular secretion mediated by MRs in peripheral tissues.^{67,68}

3.1 Tissue expression and localization

The M_2R is the predominant subtype in the heart⁶⁶ where its expression is almost 2.5-fold higher in the human atrium than in ventricles.⁶⁹ The other subtypes are present in the heart as well, although to a lesser extent than

M_2R . While mRNA encoding all five subtypes has been reported in both atrial and ventricular human myocardium, immunoblotting experiments have only confirmed the expression of M_1R , M_2R , M_3R , and M_5R , but not M_4R , at the protein level.⁷⁰ Consistently, antibodies to all subtypes besides M_4R positively stain the sarcolemma of human ventricular cardiomyocytes, with M_3R and M_5R additionally exhibiting strong localization to the intercalated disc.⁷⁰ Although M_4R protein expression has not been reported in the human heart, both functional and binding assays have revealed its presence in atrial myocardium from canines.⁷¹ This suggests that all five subtypes might be physiologically relevant in the heart but emphasizes that their functional significance may be species-dependent.

Evaluation of MR abundance in human patients with heart disease, on the other hand, has generated conflicting results. Early studies found a moderate, but not statistically significant, upregulation of M_2R in the left ventricles of patients with end-stage heart failure.⁷² Corroborating this, positron emission tomography with radiolabeled 11C-MQNB, a muscarinic antagonist, revealed elevated MR levels in patients with idiopathic dilated cardiomyopathy perhaps as a compensatory mechanism to β -adrenergic signalling.⁷³ However, recent studies measuring the levels of MRs in atrial tissues from patients with chronic atrial fibrillation reported no change in M_2R expression, but an upregulation of the M_1R subtype.⁷⁴ This suggests that different cardiac disease states, chamber-specific differences, and/or receptor subtype-specific differences are important factors to consider when evaluating the impact of MRs in cardiac disease.

3.2 Cellular signalling mechanisms

Belonging to the Class A (rhodopsin-like) subtype of GPCRs, MRs undergo a stereotypical activation mechanism wherein agonist binding induces an outward movement in transmembrane helix 6 (TM6) that exposes a G protein binding cavity on the cytoplasmic surface.⁷⁵ Molecular dynamics simulations of different agonist-occupied receptor conformations⁷⁶ and cryo-EM structures of either M₁R or M₂R in complex with distinct heterotrimeric G protein families⁷⁷ further revealed that conformational rearrangements in TM5, 6, and 7 correlate with the selective coupling of MRs to different G protein classes. Specifically, M₂R and M₄R, mediate signalling via G α_i , whereas M₁R, M₃R, and M₅R predominantly couple to G α_q .⁶⁶

The M₂R and M₄R receptors are coupled to the G α_i signal to inhibit AC leading to reduced PKA activity, reduced L-type calcium channel current, and suppression of the hyperpolarization-activated cyclic nucleotide-gated 4 (HCN4) channel. The HCN4 channel is a non-selective cation channel permeable to Na⁺/K⁺ that mediates the pacemaker or 'funny' current (I_f) in nodal cells and conduction tissues.⁷⁸ Together, these mechanisms collectively counteract the positive chronotropic effects mediated by β -adrenergic-G α_s signalling, thereby modulating the heart rate response. In contrast, G α_q -coupled receptors M₁R, M₃R, and M₅R promote the activation of PKC and stimulate calcium influx via second messengers DAG and IP₃, respectively.

In addition to the signalling mechanisms mediated by G α , the G $\beta\gamma$ subunit of the G α complex directly couples and activates the G protein-coupled inward rectifying K⁺ (GIRK) channel leading to K⁺ influx, hyperpolarization, and slowed heart rate in response to MR activation.⁷⁹ Using atrial-specific or ventricle-specific GIRK knockout mice, Lee and colleagues demonstrated that MR-dependent regulation of heart rate is primarily mediated via atrial GIRK activation, rather than GIRK channels expressed in the ventricle.⁸⁰ Of note, this mechanism can be activated by the G $\beta\gamma$ subunits originating from G α_q -coupled M₁R stimulation as well.⁷⁴ Finally, like other GPCRs, MRs are phosphorylated by GRKs in an agonist-dependent fashion resulting in the recruitment of β -arrestins which facilitate receptor desensitization, internalization, and downstream signalling.⁸¹

3.3 Physiological functions and contribution to disease

The lack of subtype-specific MR ligands and their heterogenous distribution across tissues has complicated the investigation of the individual roles of each member of the MR family. In order to study the unique function of each receptor subtype, mice constitutively deficient in each of the MRs have been generated via gene targeting.⁶⁷ In addition to a wide array of cognitive and behavioural phenotypes mainly mediated through the loss of MRs in the CNS,⁶⁷ mice deficient in MRs also exhibit profound abnormalities in cardiac autonomic regulation. In particular, while the application of carbachol, a muscarinic agonist, induced bradycardia in isolated atrial cells from wild-type mice, atrial cells from *CHRM2*^{-/-} (M₂R knockout) mice remained unaffected.⁸² In contrast, cardiomyocytes from M₄R knockout mice, *CHRM4*^{-/-}, show a very modest decrease in carbachol-induced bradycardia, suggesting that the M₄R could instead participate in maximizing the bradycardic effects of carbachol stimulation, at least in the species where it is expressed.⁸² Although all other subtypes are detected to some level in the heart, the lack of compensatory effects mediated by the remaining subtypes in *CHRM2*^{-/-} mice suggests that the M₂R subtype predominantly mediates the negative chronotropic response in the heart.^{67,82}

A significant functional role has been established for the G α_q -coupled M₃R in the heart.^{83–88} In particular, pharmacological activation and/or overexpression of the M₃R have been associated with reduced cardiac hypertrophy and myocardial injury induced by angiotensin II,⁸³ reduced frequency of ventricular arrhythmias, cardiac fibrosis and electrical remodeling following chronic cardiac pressure overload,^{84–87} and is protective against cardiac damage following ischaemic injury.⁸⁸ In contrast, the physiological roles of the remaining G α_q -coupled receptors in the muscarinic family, M₁R and M₅R, remain largely unstudied in the heart.

Notably, several recent studies have investigated the protective potential of cardiac MR stimulation and downstream signalling. For example, modulating MR activity could be clinically useful in the treatment of doxorubicin-induced cardiotoxicity. Cancer patients being treated with anthracycline-based chemotherapies (such as doxorubicin) are susceptible to developing left ventricular systolic dysfunction and arrhythmia, likely arising from imbalanced cardiac autonomic signalling.^{89,90} Notably, treatment with muscarinic agonist bethanechol improved overall cardiac function in a rat model of doxorubicin-induced cardiotoxicity by enhancing mitochondrial function, and reducing mitochondrial oxidative damage, cardiomyocyte apoptosis, and myocardial inflammation.⁸⁹ In addition, recent studies conducted in zebrafish embryos demonstrated that the MR antagonist, tolterodine, initiates transcriptional changes in the heart that promote pacemaker cell fate. These findings proposed a new role for MRs in cardiac development and underscore the potential utility of MR antagonists as a therapy for developmentally related cardiac conduction system disorders.⁹¹

3.4 Pharmacological perspectives

Excessive sympathetic and attenuated parasympathetic activity manifest early in the development of cardiac disease. In fact, depressed parasympathetic activity can occur as early as three days after the development of cardiac dysfunction and precedes the upregulation of sympathetic activity.⁹² Most current pharmacological interventions aim to suppress the over-active sympathetic nervous system (i.e. beta blockers). However, targeted modulation of parasympathetic pathways (i.e. via MR ligands) remains relatively under-exploited.

Aside from atropine, a muscarinic antagonist that is clinically utilized to acutely treat marked symptomatic bradycardia,⁹³ the majority of muscarinic agonists or antagonists characterized to date have been mainly developed for a variety of non-cardiac conditions such as glaucoma, asthma, smooth-muscle disorders, chronic obstructive pulmonary disease (COPD), peptic ulcer, Sjogren's syndrome, cancer, and various neurological disorders.⁶⁸ Due to the lack of subtype-selective MR ligands, and the ubiquitous expression of MR subtypes across different tissues, the clinical utility of the current therapies is limited due to the presence of non-selective side effects.⁶⁸ For instance, the use of tiotropium, one of the six licensed MR antagonists used as a bronchodilator agent for the treatment of COPD,⁹⁴ has the potential for cardiovascular risk. As evidence for this notion, recent studies have revealed that perfusion of tiotropium in the rat heart leads to cardiomyocyte cell damage via pathological calcium signalling.⁹⁵

A major barrier in the development of subtype-selective MR drugs is the high degree of conservation of the amino acids forming the orthosteric binding pocket across the different subtypes.⁶⁴ In particular, the five subtypes share 64–82% sequence identity overall and 82–92% identity within the transmembrane region which harbours the orthosteric binding pocket.^{64,77} Importantly, the crystal structure of the M₃R revealed the presence of a relatively large cavity separate from the orthosteric binding pocket.⁹⁶ Although the structural feature is generally conserved among all subtypes, the amino acids surrounding this region are more diverse, which potentially makes it a target for subtype-selective allosteric modulators.⁹⁷ In line with this, allosteric modulators for the M₁R, M₄R, and M₅R receptors have been developed that are predicted to bind to this region.⁹⁸ To date, many subtype-specific allosteric modulators have been identified for all five MRs, however, they have predominantly been pursued for the treatment of CNS disorders.⁹⁹ This warrants the further discovery and characterization of subtype-selective muscarinic allosteric modulators that might be clinically useful in the treatment of cardiovascular disease as well.

4. Angiotensin II receptors

Angiotensin II type 1 (AT₁R) and Type 2 (AT₂R) receptors, endogenously activated by the peptide hormone, Angiotensin II (Ang II), are additional members of the GPCR superfamily that are fundamental for the regulation of cardiovascular physiology (Figure 5). Although the role of AT₁R is well

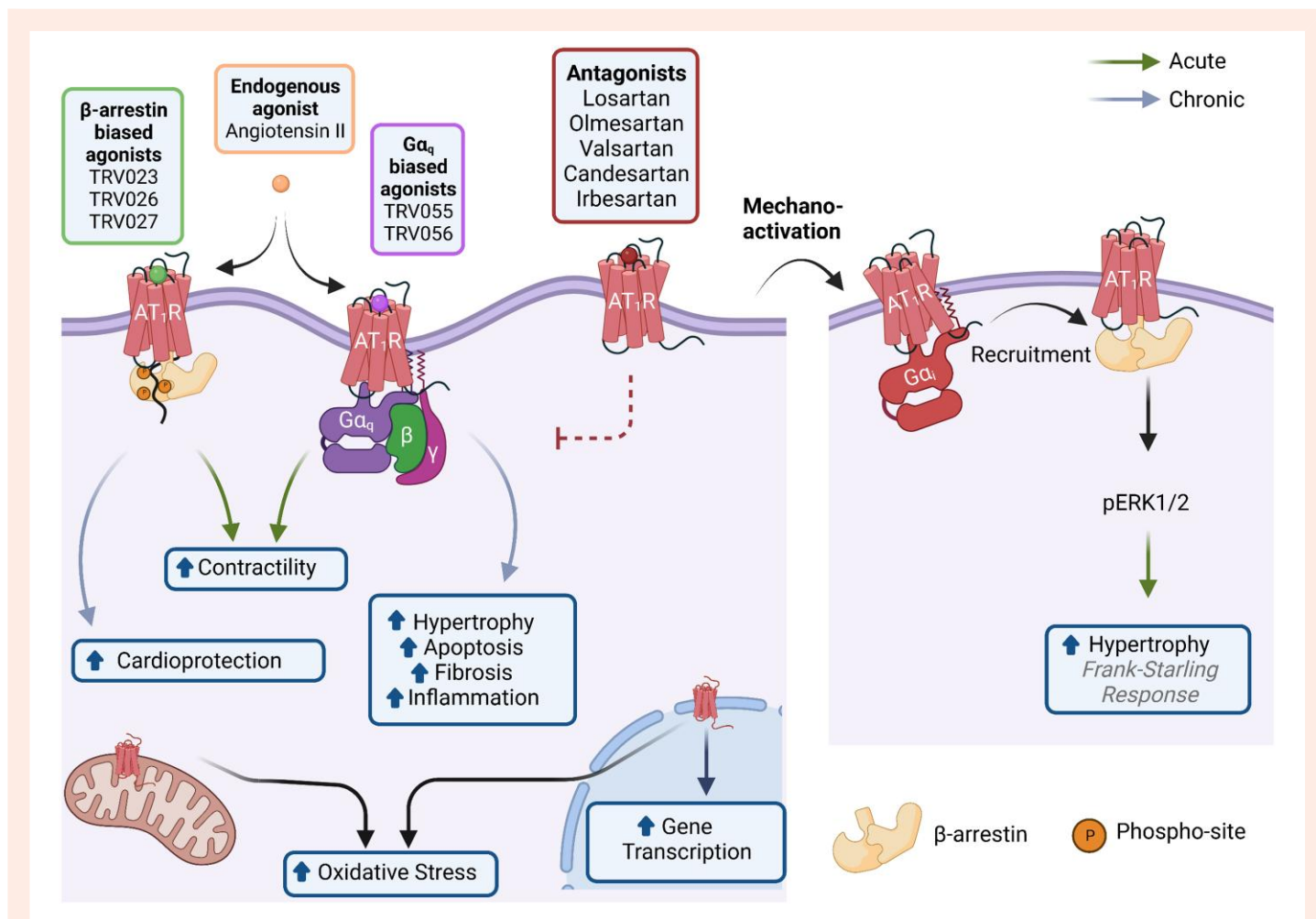


Figure 5 Physiological and pathological effects of cardiac AT₁R signalling. The AT₁R is a pleiotropic receptor that localizes to the cellular, mitochondrial, and nuclear membranes of cells. Ligands that preferentially stabilize the AT₁R/ β -arrestin complex (TRV023, TRV026, TRV027) increase the contractility of cardiomyocytes and have cardioprotective effects under long-term use. Likewise, ligands that preferentially stabilize the AT₁R/G α_q complex (TRV055 and TRV056) increase the contractility of cardiomyocytes. However, chronic activation of this signalling pathway promotes cardiac hypertrophy, apoptosis, fibrosis, and inflammation. The endogenous ligand for the AT₁R, Ang II, is unbiased towards both signalling pathways. AT₁R antagonists such as Losartan, Olmesartan, and Valsartan, among others, stabilize the receptor in an inactive conformation thereby blocking the activation of any signalling cascade. Another activation mechanism of the AT₁R is via membrane stretch. In turn, β -arrestin is recruited in a process mediated by G α_q , and activates ERK1/2 phosphorylation regulating the Frank–Starling mechanism and inducing hypertrophy. Finally, the activation of the AT₁R at mitochondrial or nuclear membranes induces oxidative stress and activates gene transcription. Figure generated with Biorender.com.

established in the heart, the significance of AT₂R remains highly controversial since altering the expression of AT₂R does not affect cardiac function^{100,101} and findings regarding the role of AT₂R in cardiac pathology appear contradictory. Therefore, we mainly focus on the AT₁R while providing a summary of the most relevant aspects of the AT₂R in cardiac pathophysiology.

The translation of human AT₁R comes from a single gene, *AGTR1*, which shares ~95% homology to bovine and rat AT₁Rs.¹⁰² Comparatively, two separate AT₁R genes are found in mice and rats, *Agtr1a* and *Agtr1b*, that encode for the AT_{1A}R and AT_{1B}R, respectively. Both murine receptors share ~95% homology and are functionally identical, yet are expressed in a tissue-specific manner.¹⁰³ The AT_{1A}R is the predominant isoform in rodent hearts and is therefore the main subtype addressed in this review when discussing AT₁R in rodent models. Multiple reviews with a specific focus on the structure, trafficking, function, and/or pathophysiology of the AT₁R are also available for complementary information.^{4,104–106}

4.1 Tissue expression and localization

The AT₁R is expressed in all cell types of the heart including the endothelial cells,¹⁰⁷ vascular smooth-muscle cells,¹⁰⁸ fibroblasts,¹⁰⁹ myocardial cells,¹¹⁰ and immune cells.¹¹¹ The AT₁R represents 59% of the total AT receptors in human ventricles, compared to its homologue subtype, the AT₂R, which comprises the remaining 41%.¹¹² Conversely, the human atrium shows an inverted relationship where the AT₁R:AT₂R ratio is 30:70.¹¹³ Acute heart injury such as myocardial infarction upregulates AT₁R expression, whereas chronic cardiac damage such as dilated cardiomyopathy downregulates its expression.¹¹² Thus, the temporal nature of the cardiac disease is a key regulator of AT₁R levels. Beyond the cellular membrane, AT₁R further localizes to nuclear and mitochondrial membranes in the heart where it activates gene transcription and induces oxidative stress, respectively.^{114,115} This suggests that the localization within the cell is also an important factor for the signal transduction and function of AT₁R.

4.2 Cellular signalling mechanisms

The ubiquitous expression of AT₁R along with the endocrine, paracrine, autocrine, and/or intracrine effects of its endogenous ligand, Ang II, are main contributors to the multiple cellular effects (i.e. growth, contractility, inflammation, fibrosis, and apoptosis, among others) regulating the cardiovascular system.¹⁰⁴ Independent from Ang II, AT₁R can also be activated by the mechanical forces exerted within the cardiovascular system.^{116,117} Moreover, the AT₁R can couple to multiple intracellular transducers including several G protein subtypes (i.e. G_{α_{q/11}}, G_{α_s}, G_{α_{i/o}}, and G_{α_{12/13}}) and β arrestins to initiate different cellular responses.^{118,119} The best-characterized signalling pathways for the AT₁R in physiological conditions are the canonical G_{α_q} pathway and the β arrestin pathway.

Like other GPCRs, the dynamic structure of the AT₁R permits multiple active conformations that preferentially interact with distinct transducers and can be further stabilized by biased ligands.^{120,121} Importantly, the development of biased ligands for the AT₁R has allowed for a more in-depth exploration of the signalling pathways downstream of the AT₁R and their biological consequences. This is clearly illustrated in a recent proteomics study utilizing proximity ligation assay to identify more than a thousand functional and structural proteins proximal to the AT₁R after activation with unbiased or biased ligands for the G_{α_q} or the β-arrestin pathway.¹²² Understanding the effects of these ligands in the heart will enable more precise modulation of AT₁R activity and will allow for the development of novel strategies to preferentially target beneficial signalling cascades in disease states.

4.3 Physiological functions and contributions to disease

Upon activation, Ang II binds to the AT₁R and increases cardiomyocyte contractility by coupling to the canonical G_{α_q} pathway as well as β-arrestin.¹²³ While the latter observation was first shown *in vitro* using [Sar1, Ile4, Ile8]-Angiotensin II (SII), the first β-arrestin-biased ligand available for the AT₁R,¹²³ this concept was later demonstrated *in vivo* using the β-arrestin-biased AT₁R ligands TRV023 and TRV027.^{124,125} While the regulation of calcium is central to the contractile response induced by both Ang II and β-arrestin-biased ligands (TRV023 and TRV 027), the mechanisms by which they regulate intracellular calcium are distinct. Specifically, stimulation with Ang II leads to increased intracellular calcium concentration via enhanced calcium release through the AT₁R-G_{α_q}-IP₃/PKC signalling axis.¹²⁶ In contrast, β-arrestin-biased activation of the AT₁R increases myofilament calcium sensitivity without altering the global intracellular calcium transient, in part through modifying the phosphorylation status of myofilament proteins.^{127–129}

In contrast to acute activation, chronic AT₁R stimulation by direct stimulation with Ang II,^{130,131} overexpression of AT₁R, or expression of constitutively active mutants of the AT₁R^{132,133} induces cardiac hypertrophy. Similarly, increasing the activity of the local renin angiotensin system (RAS) of the heart by angiotensinogen overexpression, the main precursor of Ang II, also induces cardiac hypertrophy.¹³⁴ The hypertrophic response induced by chronic AT₁R stimulation is largely mediated by G_{α_q}, since *in vivo* overexpression of G_{α_q} or *in vitro* transfection of a constitutively activated G_{α_q} mutant induces hypertrophy and apoptosis in cardiomyocytes.¹³⁵ Moreover, overexpression of an inhibitor of G_{α_q} blocks the development of cardiac hypertrophy induced by pressure overload.¹³⁶ The alternative β-arrestin pathway might also contribute to this effect since cardiac-specific overexpression of the AT₁R mutant lacking G_{α_q}/G_{α_i} coupling also induces hypertrophy.^{131,137} Nevertheless, numerous studies have demonstrated that β-arrestin-biased activation of AT₁R is cardioprotective via promoting cardiomyocyte cell survival.^{116,125,138}

Although the hypertrophic response mediated by chronic AT₁R stimulation is well established, it has not been consistently observed. Recent studies demonstrate that mice overexpressing AT₁R or a constitutively active AT₁R mutant do not undergo hypertrophy despite developing fibrosis and/or ventricular dysfunction.^{139,140} Moreover, AT₁R knockout mice develop cardiac hypertrophy following pressure overload or myocardial

infarction, suggesting that the AT₁R may not be critical for the development of hypertrophy in heart failure.^{141,142} Accordingly, chronic stimulation with Ang II in mice lacking AT₁R in the kidney, yet expressing physiological levels of AT₁R in the heart, does not develop cardiac hypertrophy.¹⁴³ In line with this, selective expression of the AT₁R in the kidney or in resistance vessels mimics the cardiac hypertrophy induced by chronic Ang II treatment observed in wild-type mice.^{143,144} Taken together, the development of cardiac hypertrophy following the manipulation of AT₁R could be secondary to the increased peripheral resistance that induces pressure overload in the heart.

Other paracrine, stretch, and transactivation mechanisms of the AT₁R are also contributors to cardiomyocyte growth. Indeed, Ang II-stimulated cardiac fibroblasts secrete exosomes and multiple cytokines such as transforming growth factor (TGF-1β) and interleukin-6 that stimulate cardiomyocytes to increase local Ang II production, AT₁R expression, and induce hypertrophy.^{145–148} In addition, AT₁R can be activated by mechanical stretch independent from Ang II, leading to hypertrophic growth.¹¹⁷ This mechano-activation mechanism appears to be dependent on G_{α_i} and β-arrestin pathways.^{116,149,150} Mechanical stretch also increases the expression of the AT₁R and other components of the RAS, while Ang II decreases AT₁R expression in the heart.¹⁵¹ Therefore, mechano-activation of the AT₁R behaves as a positive feedback loop for the cardiac RAS. Notably, this mechanism underlies the Frank–Starling response which describes the length-dependent activation of contractility wherein increased cardiac filling, and thus increased sarcomere length, enhances the force of contraction.¹²⁷ Finally, AT₁R stimulation transactivates EGFR through multiple intracellular mechanisms including β-arrestin recruitment and direct association/dimerization between both receptors ultimately inducing hypertrophy.¹⁵² Aside from cardiomyocyte growth, AT₁R activation also leads to apoptosis, hyperplasia, fibrosis, and oxidative stress, which further contribute to disease.^{130,132,133,139}

4.4 Pharmacological perspectives

Eight AT₁R blockers (ARBs), which stabilize the receptor in an inactive state, have been approved by the FDA and are clinically utilized for the treatment of multiple cardiovascular diseases including heart failure.^{153,154} The AT₁R β-arrestin-biased ligand TRV027 was also recently proposed as pharmacotherapy for acute heart failure. However, TRV027 failed to improve the clinical status of patients with this condition in clinical trials.¹⁵⁵ This unexpected outcome might be related to the short duration of the treatment (1 month). Accordingly, Ryba and colleagues showed that a 3-month treatment with the β-arrestin-biased ligand TRV067 improves the cardiac function of mice with dilated cardiomyopathy.¹⁵⁶ Alternatively, the developmental stage might also modify the efficacy of the treatment, since activation of the AT₁R-β-arrestin2 pathway in neonatal or immature cardiomyocytes induces sustained cardiac contractility¹⁵⁷ whereas this effect is short lasting in adult hearts of mice or rats.^{124,158} Thus, refocusing these novel compounds to treat heart failure for a longer period or in the paediatric population might prove therapeutic.

4.5 Angiotensin II Type 2 receptor

The AT₂R subtype, only ~34% identical to the AT₁R,¹⁵⁹ exhibits unique transducer coupling properties and complex pathophysiological effects. The first crystal structure available of the AT₂R showed a non-canonical position of helix 8 that blocked the binding sites of G proteins and β-arrestins.¹⁶⁰ This finding is consistent with multiple studies that reported a lack of signalling through G proteins or internalization.¹⁶¹ The recently crystalized AT₂R bound to Ang II shows a rather canonical outward position of helix 8, raising the possibility of conformational selection by different ligands to induce multiple responses.¹⁶² Besides Ang II, new endogenous ligands derived from Ang II, such as Ang 1–7 and Ang 1–9, have been described as agonists for the AT₂R that may be beneficial in the heart.^{163,164}

In the human heart, AT₂R has been detected in cardiomyocytes,¹⁶⁵ fibroblasts,¹¹² and coronary arteries.¹⁶⁶ During heart failure or dilated cardiomyopathy, AT₂R levels are downregulated or upregulated, respectively,

suggesting that the regulation of the AT₂R depends on the aetiology of the disease, or perhaps reflects alterations in the relative proportions of different cell types during the pathological process of adverse cardiac remodelling.^{112,165} Moreover, studies evaluating the contribution of AT₂R to cardiac disease show conflicting results. Some claim that AT₂R activation is associated with decreased hypertrophy and fibrosis.^{167–169} Conversely, others report that AT₂R overexpression results in constitutive hypertrophy in neonatal rat cardiomyocytes, fibrosis, and heart failure,^{170,171} whereas deletion of the AT₂R decreases hypertrophy following chronic Ang II administration or myocardial infarction.^{172,173} These discrepancies might be explained by the expression levels of the AT₂R during the cardiac insult.¹⁷⁴ Thus, pharmacological activation of AT₂R during disease states when there are high expression levels of AT₂R might prove therapeutic. Indeed, multiple recent studies using AT₂R agonists in models of heart disease have shown cardioprotective effects in rodents.^{164,175,176} In summary, the AT₂R is a potential therapeutic target in cardiac disease that needs further examination to better understand its pathophysiological significance.

5. Conclusion

GPCRs serve as excellent therapeutic targets for the identification of novel heart failure treatments given their diverse and essential roles in cardiac health and disease. The β -adrenergic, muscarinic, and angiotensin receptor families exemplify only 3 of the >200 types of GPCRs in the heart that fundamentally regulate cardiac function and demonstrate great pharmacological potential. Importantly, recent advancements in understanding the mechanisms of biased signalling coupled with the identification of novel allosteric modulators have enabled the discovery of more selective ligands possessing enhanced cardioprotective effects with reduced unwanted side effects. Nonetheless, the heterogeneous distribution of GPCRs across multiple cell types in the heart,^{4,177} combined with their overlapping signalling networks, and localization to cellular compartments aside from the cell surface,^{177–180} remain essential elements to consider in evaluating the physiological effects of these novel drugs. Further investigation of the biased signalling pathways, physiological functions, disease mechanisms, and pharmacological features of cardiac GPCRs will therefore prove impactful in designing novel therapeutics.

Conflict of interest: H.A.R. is a scientific cofounder of Trevena Inc., a company that is developing new GPCR ligands.

Funding

This work was supported by the National Institutes of Health Grant HL056687 to H.A.R.

References

- Pierce KL, Premont RT, Lefkowitz RJ. Seven-transmembrane receptors. *Nat Rev Mol Cell Biol* 2002;**3**:639–650.
- Sriram K, Insel PA. G protein-coupled receptors as targets for approved drugs: how many targets and how many drugs? *Mol Pharmacol* 2018;**93**:251–258.
- Bjarnadottir TK, Gloriam DE, Hellstrand SH, Kristiansson H, Fredriksson R, Schiöth HB. Comprehensive repertoire and phylogenetic analysis of the G protein-coupled receptors in human and mouse. *Genomics* 2006;**88**:263–273.
- Wang J, Gareri C, Rockman HA. G-Protein-Coupled receptors in heart disease. *Circ Res* 2018;**123**:716–735.
- Jiang H, Galtes D, Wang J, Rockman HA. G protein-coupled receptor signaling: transducers and effectors. *Am J Physiol Cell Physiol* 2022;**323**:C731–C748.
- Liu Y, Chen J, Fontes SK, Bautista EN, Cheng Z. Physiological and pathological roles of protein kinase A in the heart. *Cardiovasc Res* 2022;**118**:386–398.
- Zhang P, Mende U. Regulators of G-protein signalling in the heart and their potential as therapeutic targets. *Circ Res* 2011;**109**:320–333.
- Pitcher JA, Freedman NJ, Lefkowitz RJ. G protein-coupled receptor kinases. *Annu Rev Biochem* 1998;**67**:653–692.
- Perry SJ, Baillie GS, Kohout TA, McPhee I, Magiera MM, Ang KL, Miller WE, McLean AJ, Conti M, Houslay MD, Lefkowitz RJ. Targeting of cyclic AMP degradation to beta 2-adrenergic receptors by beta-arrestins. *Science* 2002;**298**:834–836.
- Nelson CD, Perry SJ, Regier DS, Prescott SM, Topham MK, Lefkowitz RJ. Targeting of diacylglycerol degradation to M1 muscarinic receptors by beta-arrestins. *Science* 2007;**315**:663–666.
- Laporte SA, Oakley RH, Zhang J, Holt JA, Ferguson SS, Caron MG, Barak LS. The beta2-adrenergic receptor/betaarrestin complex recruits the clathrin adaptor AP-2 during endocytosis. *Proc Natl Acad Sci U S A* 1999;**96**:3712–3717.
- Jean-Charles PY, Kaur S, Shenoy SK. G protein-coupled receptor signaling through beta-arrestin-dependent mechanisms. *J Cardiovasc Pharmacol* 2017;**70**:142–158.
- Smith JS, Lefkowitz RJ, Rajagopal S. Biased signalling: from simple switches to allosteric microprocessors. *Nat Rev Drug Discov* 2018;**17**:243–260.
- Christopoulos A. Allosteric binding sites on cell-surface receptors: novel targets for drug discovery. *Nat Rev Drug Discov* 2002;**1**:198–210.
- Schwartz TW, Holst B. Allosteric enhancers, allosteric agonists and ago-allosteric modulators: where do they bind and how do they act? *Trends Pharmacol Sci* 2007;**28**:366–373.
- Slosky LM, Caron MG, Barak LS. Biased allosteric modulators: new frontiers in GPCR drug discovery. *Trends Pharmacol Sci* 2021;**42**:283–299.
- Ahliquist RP. A study of the adrenotropic receptors. *Am J Physiol* 1948;**153**:586–600.
- Lefkowitz RJ. Editorial: selectivity in beta-adrenergic responses: clinical implications. *Circulation* 1974;**49**:783–786.
- Bristow MR, Ginsburg R, Umans V, Fowler M, Minobe W, Rasmussen R, Zera P, Menlove R, Shah P, Jamieson S. Beta 1- and beta 2-adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective beta 1-receptor down-regulation in heart failure. *Circ Res* 1986;**59**:297–309.
- Michel LYM, Farah C, Balligand JL. The Beta3 adrenergic receptor in healthy and pathological cardiovascular tissues. *Cells* 2020;**9**:2584.
- Bathe-Peters M, Gmach P, Boltz HH, Einsiedel J, Gotthardt M, Hubner H, Gmeiner P, Lohse MJ, Annibale P. Visualization of beta-adrenergic receptor dynamics and differential localization in cardiomyocytes. *Proc Natl Acad Sci U S A* 2021;**118**:e2101119118.
- Schobesberger S, Wright PT, Poulet C, Sanchez Alonso Mardones JL, Mansfield C, Friebe A, Harding SE, Balligand JL, Nikolaev VO, Gorelik J. beta3-Adrenoceptor redistribution impairs NO/cGMP/PDE2 signalling in failing cardiomyocytes. *Elife* 2020;**9**:e52221.
- Xiao RP. Beta-adrenergic signaling in the heart: dual coupling of the beta2-adrenergic receptor to G(s) and G(i) proteins. *Sci STKE* 2001;**2001**:re15.
- Cannavo A, Koch WJ. Targeting beta3-adrenergic receptors in the heart: selective agonism and beta-blockade. *J Cardiovasc Pharmacol* 2017;**69**:71–78.
- Mangmool S, Shukla AK, Rockman HA. Beta-Arrestin-dependent activation of ca(2+)/calmodulin kinase II after beta(1)-adrenergic receptor stimulation. *J Cell Biol* 2010;**189**:573–587.
- Luttrell LM, Hawes BE, van Biesen T, Luttrell DK, Lansing TJ, Lefkowitz RJ. Role of c-src tyrosine kinase in G protein-coupled receptor- and Gbetagamma subunit-mediated activation of mitogen-activated protein kinases. *J Biol Chem* 1996;**271**:19443–19450.
- Hordijk PL, Verlaan I, van Corven EJ, Moolenaar WH. Protein tyrosine phosphorylation induced by lysophosphatidic acid in rat-1 fibroblasts. Evidence that phosphorylation of map kinase is mediated by the gi-p21ras pathway. *J Biol Chem* 1994;**269**:645–651.
- Kohout TA, Takaoka H, McDonald PH, Perry SJ, Mao L, Lefkowitz RJ, Rockman HA. Augmentation of cardiac contractility mediated by the human beta(3)-adrenergic receptor overexpressed in the hearts of transgenic mice. *Circulation* 2001;**104**:2485–2491.
- Luttrell LM, Ferguson SS, Daaka Y, Miller WE, Maudsley S, Della Rocca GJ, Lin F, Kawakatsu H, Owada K, Luttrell DK, Caron MG, Lefkowitz RJ. Beta-arrestin-dependent formation of beta2 adrenergic receptor-src protein kinase complexes. *Science* 1999;**283**:655–661.
- Noma T, Lemaire A, Naga Prasad SV, Barki-Harrington L, Tilley DG, Chen J, Le Corvoisier P, Violin JD, Wei H, Lefkowitz RJ, Rockman HA. Beta-arrestin-mediated beta1-adrenergic receptor transactivation of the EGFR confers cardioprotection. *J Clin Invest* 2007;**117**:2445–2458.
- Cohn JN, Levine TB, Olivari MT, Garberg V, Lura D, Francis GS, Simon AB, Rector T. Plasma norepinephrine as a guide to prognosis in patients with chronic congestive heart failure. *N Engl J Med* 1984;**311**:819–823.
- Bristow MR, Hershberger RE, Port JD, Minobe W, Rasmussen R. Beta 1- and beta 2-adrenergic receptor-mediated adenylate cyclase stimulation in nonfailing and failing human ventricular myocardium. *Mol Pharmacol* 1989;**35**:295–303.
- Ungerer M, Bohm M, Elce JS, Erdmann E, Lohse MJ. Altered expression of beta-adrenergic receptor kinase and beta 1-adrenergic receptors in the failing human heart. *Circulation* 1993;**87**:454–463.
- Scheuer J. Catecholamines in cardiac hypertrophy. *Am J Cardiol* 1999;**83**:70H–74H.
- Engelhardt S, Hein L, Wiesmann F, Lohse MJ. Progressive hypertrophy and heart failure in beta1-adrenergic receptor transgenic mice. *Proc Natl Acad Sci U S A* 1999;**96**:7059–7064.
- Bisognano JD, Weinberger HD, Bohlmeier TJ, Pende A, Reynolds MV, Sastravaha A, Roden R, Asano K, Blaxall BC, Wu SC, Communal C, Singh K, Colucci W, Bristow MR, Port DJ. Myocardial-directed overexpression of the human beta(1)-adrenergic receptor in transgenic mice. *J Mol Cell Cardiol* 2000;**32**:817–830.
- Du XJ, Autelitano DJ, Dilley RJ, Wang B, Dart AM, Woodcock EA. Beta(2)-adrenergic receptor overexpression exacerbates development of heart failure after aortic stenosis. *Circulation* 2000;**101**:71–77.
- Nguyen MN, Kiriazis H, Ruggiero D, Gao XM, Su Y, Jian A, Han LP, McMullen JR, Du XJ. Spontaneous ventricular tachyarrhythmias in beta2-adrenoceptor transgenic mice in relation to cardiac interstitial fibrosis. *Am J Physiol Heart Circ Physiol* 2015;**309**:H946–H957.
- Cross HR, Steenbergen C, Lefkowitz RJ, Koch WJ, Murphy E. Overexpression of the cardiac beta(2)-adrenergic receptor and expression of a beta-adrenergic receptor kinase-1 (betaARK1) inhibitor both increase myocardial contractility but have differential effects on susceptibility to ischemic injury. *Circ Res* 1999;**85**:1077–1084.

40. Chesley A, Lundberg MS, Asai T, Xiao RP, Ohtani S, Lakatta EG, Crow MT. The beta(2)-adrenergic receptor delivers an antiapoptotic signal to cardiac myocytes through G(i)-dependent coupling to phosphatidylinositol 3'-kinase. *Circ Res* 2000;**87**:1172–1179.
41. Zhu WZ, Zheng M, Koch WJ, Lefkowitz RJ, Kobilka BK, Xiao RP. Dual modulation of cell survival and cell death by beta(2)-adrenergic signaling in adult mouse cardiac myocytes. *Proc Natl Acad Sci U S A* 2001;**98**:1607–1612.
42. Rohrer DK, Desai KH, Jasper JR, Stevens ME, Regula DP Jr, Barsh GS, Bernstein D, Kobilka BK. Targeted disruption of the mouse beta1-adrenergic receptor gene: developmental and cardiovascular effects. *Proc Natl Acad Sci U S A* 1996;**93**:7375–7380.
43. Yoo B, Lemaire A, Mangmool S, Wolf MJ, Curcio A, Mao L, Rockman HA. Beta1-adrenergic receptors stimulate cardiac contractility and CaMKII activation in vivo and enhance cardiac dysfunction following myocardial infarction. *Am J Physiol Heart Circ Physiol* 2009;**297**:H1377–H1386.
44. Rohrer DK, Chruscinski A, Schauble EH, Bernstein D, Kobilka BK. Cardiovascular and metabolic alterations in mice lacking both beta1- and beta2-adrenergic receptors. *J Biol Chem* 1999;**274**:16701–16708.
45. Chruscinski AJ, Rohrer DK, Schauble E, Desai KH, Bernstein D, Kobilka BK. Targeted disruption of the beta2 adrenergic receptor gene. *J Biol Chem* 1999;**274**:16694–16700.
46. Tavernier G, Toumaniantz G, Erfanian M, Heymann MF, Laurent K, Langin D, Gauthier C. Beta3-Adrenergic stimulation produces a decrease of cardiac contractility ex vivo in mice overexpressing the human beta3-adrenergic receptor. *Cardiovasc Res* 2003;**59**:288–296.
47. Gauthier C, Leblais V, Kobzik L, Trochu JN, Khandoudi N, Bril A, Balligand JL, Le Marec H. The negative inotropic effect of beta3-adrenoceptor stimulation is mediated by activation of a nitric oxide synthase pathway in human ventricle. *J Clin Invest* 1998;**102**:1377–1384.
48. Varghese P, Harrison RW, Lofthouse RA, Georgakopoulos D, Berkowitz DE, Hare JM. Beta(3)-adrenoceptor deficiency blocks nitric oxide-dependent inhibition of myocardial contractility. *J Clin Invest* 2000;**106**:697–703.
49. Belge C, Hammond J, Dubois-Deruy E, Manoury B, Hamelet J, Beauloye C, Markl A, Pouleur AC, Bertrand L, Esfahani H, Jnaoui K, Gotz KR, Nikolaev VO, Vanderper A, Herijgers P, Lobysheva I, Iaccarino G, Hilfiger-Kleiner D, Tavernier G, Langin D, Dessy C, Balligand JL. Enhanced expression of beta3-adrenoceptors in cardiac myocytes attenuates neurohormone-induced hypertrophic remodeling through nitric oxide synthase. *Circulation* 2014;**129**:451–462.
50. Niu X, Watts VL, Cingolani OH, Sivakumaran V, Leyton-Mange JS, Ellis CL, Miller KL, Vandegaer K, Bedja D, Gabrielson KL, Paolucci N, Kass DA, Barouch LA. Cardioprotective effect of beta-3 adrenergic receptor agonism: role of neuronal nitric oxide synthase. *J Am Coll Cardiol* 2012;**59**:1979–1987.
51. Niu X, Zhao L, Li X, Xue Y, Wang B, Lv Z, Chen J, Sun D, Zheng Q. Beta3-Adrenoreceptor stimulation protects against myocardial infarction injury via eNOS and nNOS activation. *PLoS One* 2014;**9**:e98713.
52. Masutani S, Cheng HJ, Morimoto A, Hasegawa H, Han QH, Little WC, Cheng CP. Beta3-Adrenergic receptor antagonist improves exercise performance in pacing-induced heart failure. *Am J Physiol Heart Circ Physiol* 2013;**305**:H923–H930.
53. Kotecha D, Flather MD, Altman DG, Holmes J, Rosano G, Wikstrand J, Packer M, Coats AJS, Manzano L, Bohm M, van Veldhuisen DJ, Andersson B, Wedel H, von Lueder TG, Rigby AS, Hjalmarson A, Kjekshus J, Cleland JGF; Beta-Blockers in Heart Failure Collaborative Group. Heart rate and rhythm and the benefit of Beta-blockers in patients with heart failure. *J Am Coll Cardiol* 2017;**69**:2885–2896.
54. Wisler JW, DeWire SM, Whalen EJ, Violin JD, Drake MT, Ahn S, Shenoy SK, Lefkowitz RJ. A unique mechanism of beta-blocker action: carvedilol stimulates beta-arrestin signaling. *Proc Natl Acad Sci U S A* 2007;**104**:16657–16662.
55. Kim IM, Tilley DG, Chen J, Salazar NC, Whalen EJ, Violin JD, Rockman HA. Beta-blockers alprenolol and carvedilol stimulate beta-arrestin-mediated EGFR transactivation. *Proc Natl Acad Sci U S A* 2008;**105**:14555–14560.
56. Kim IM, Wang Y, Park KM, Tang Y, Teoh JP, Vinson J, Traynham CJ, Pironti G, Mao L, Su H, Johnson JA, Koch WJ, Rockman HA. Beta-arrestin1-biased beta1-adrenergic receptor signaling regulates microRNA processing. *Circ Res* 2014;**114**:833–844.
57. Wang J, Pani B, Gokhan I, Xiong X, Kahsai AW, Jiang H, Ahn S, Lefkowitz RJ, Rockman HA. Beta-Arrestin-Biased allosteric modulator potentiates carvedilol-stimulated beta adrenergic receptor cardioprotection. *Mol Pharmacol* 2021;**100**:568–579.
58. Poole-Wilson PA, Swedberg K, Cleland JG, Di Lenarda A, Hanrath P, Komajda M, Lubsen J, Lutiger B, Metra M, Remme WJ, Torp-Pedersen C, Scherhag A, Skene A; Carvedilol Or Metoprolol European Trial Investigators. Comparison of carvedilol and metoprolol on clinical outcomes in patients with chronic heart failure in the carvedilol or metoprolol European trial (COMET): randomised controlled trial. *Lancet* 2003;**362**:7–13.
59. Bhatt AS, DeVore AD, DeWald TA, Swedberg K, Mentz RJ. Achieving a maximally tolerated beta-blocker dose in heart failure patients: is there room for improvement? *J Am Coll Cardiol* 2017;**69**:2542–2550.
60. Wang J, Hanada K, Staus DP, Makara MA, Dahal GR, Chen Q, Ahles A, Engelhardt S, Rockman HA. Galphai is required for carvedilol-induced beta1 adrenergic receptor beta-arrestin biased signaling. *Nat Commun* 2017;**8**:1706.
61. Wisler JW, Rockman HA, Lefkowitz RJ. Biased G protein-coupled receptor signaling: changing the paradigm of drug discovery. *Circulation* 2018;**137**:2315–2317.
62. Ahn S, Pani B, Kahsai AW, Olsen EK, Husemoen G, Vestergaard M, Jin L, Zhao S, Wingle LM, Rambarat PK, Simhal RK, Xu TT, Sun LD, Shim PJ, Staus DP, Huang LY, Franck T, Chen X, Lefkowitz RJ. Small-Molecule positive allosteric modulators of the beta2-adrenoceptor isolated from DNA-encoded libraries. *Mol Pharmacol* 2018;**94**:850–861.
63. Pani B, Ahn S, Rambarat PK, Vege S, Kahsai AW, Liu A, Valan BN, Staus DP, Costa T, Lefkowitz RJ. Unique positive cooperativity between the beta-arrestin-biased beta-blocker carvedilol and a small molecule positive allosteric modulator of the beta2-adrenergic receptor. *Mol Pharmacol* 2021;**100**:513–525.
64. Peralta EG, Ashkenazi A, Winslow JW, Smith DH, Ramachandran J, Capon DJ. Distinct primary structures, ligand-binding properties and tissue-specific expression of four human muscarinic acetylcholine receptors. *EMBO J* 1987;**6**:3923–3929.
65. Bonner TI, Young AC, Brann MR, Buckley NJ. Cloning and expression of the human and rat m5 muscarinic acetylcholine receptor genes. *Neuron* 1988;**1**:403–410.
66. Saternos HC, Almarghalani DA, Gibson HM, Meqdad MA, Antypas RB, Lingireddy A, AbouAlaiwi WA. Distribution and function of the muscarinic receptor subtypes in the cardiovascular system. *Physiol Genomics* 2018;**50**:1–9.
67. Wess J. Muscarinic acetylcholine receptor knockout mice: novel phenotypes and clinical implications. *Annu Rev Pharmacol Toxicol* 2004;**44**:423–450.
68. Wess J, Eglén RM, Gautam D. Muscarinic acetylcholine receptors: mutant mice provide new insights for drug development. *Nat Rev Drug Discov* 2007;**6**:721–733.
69. Deighton NM, Motomura S, Borquez D, Zerkowski HR, Doetsch N, Brodde OE. Muscarinic cholinergic receptors in the human heart: demonstration, subclassification, and distribution. *Naunyn-Schmiedeberg Arch Pharmacol* 1990;**341**:14–21.
70. Wang H, Han H, Zhang L, Shi H, Schram G, Nattel S, Wang Z. Expression of multiple subtypes of muscarinic receptors and cellular distribution in the human heart. *Mol Pharmacol* 2001;**59**:1029–1036.
71. Shi H, Wang H, Wang Z. Identification and characterization of multiple subtypes of muscarinic acetylcholine receptors and their physiological functions in canine hearts. *Mol Pharmacol* 1999;**55**:497–507.
72. Le Guludec D, Cohen-Solal A, Delforge J, Delahaye N, Syrota A, Merlet P. Increased myocardial muscarinic receptor density in idiopathic dilated cardiomyopathy: an in vivo PET study. *Circulation* 1997;**96**:3416–3422.
73. Glessler C, Dhein S, Ponick K, Brodde OE. Muscarinic receptors in the failing human heart. *Eur J Pharmacol* 1999;**375**:197–202.
74. Heijman J, Kirchner D, Kunze F, Chretien EM, Michel-Reher MB, Voigt N, Knaut M, Michel MC, Ravens U, Dobrev D. Muscarinic type-1 receptors contribute to IK, ACh in human atrial cardiomyocytes and are upregulated in patients with chronic atrial fibrillation. *Int J Cardiol* 2018;**255**:61–68.
75. Zhou Q, Yang D, Wu M, Guo Y, Guo W, Zhong L, Cai X, Dai A, Jang W, Shakhnovich EI, Liu ZJ, Stevens RC, Lambert NA, Babu MM, Wang MW, Zhao S. Common activation mechanism of class A GPCRs. *Elife* 2019;**8**:e50279.
76. Randakova A, Nelic D, Dolezal V, El-Fakahany EE, Boulos J, Jakubik J. Agonist-Specific conformations of the M2 muscarinic acetylcholine receptor assessed by molecular dynamics. *J Chem Inf Model* 2020;**60**:2325–2338.
77. Maeda S, Qu Q, Robertson MJ, Skiniotis G, Kobilka BK. Structures of the M1 and M2 muscarinic acetylcholine receptor/G-protein complexes. *Science* 2019;**364**:552–557.
78. Zaza A, Robinson RB, DiFrancesco D. Basal responses of the L-type Ca²⁺ and hyperpolarization-activated currents to autonomic agonists in the rabbit sino-atrial node. *J Physiol* 1996;**491**:347–355.
79. Touhara KK, MacKinnon R. Molecular basis of signaling specificity between GIRK channels and GPCRs. *Elife* 2018;**7**:e42908.
80. Lee SW, Anderson A, Guzman PA, Nakano A, Tolkacheva EG, Wickman K. Atrial GIRK channels mediate the effects of Vagus nerve stimulation on heart rate dynamics and arrhythmogenesis. *Front Physiol* 2018;**9**:943.
81. van Koppen CJ, Kaiser B. Regulation of muscarinic acetylcholine receptor signaling. *Pharmacol Ther* 2003;**98**:197–220.
82. Stengel PW, Gomez J, Wess J, Cohen ML. M(2) and M(4) receptor knockout mice: muscarinic receptor function in cardiac and smooth muscle in vitro. *J Pharmacol Exp Ther* 2000;**292**:877–885.
83. Liu Y, Wang S, Wang C, Song H, Han H, Hang P, Jiang Y, Wei L, Huo R, Sun L, Gao X, Lu Y, Du Z. Upregulation of M(3) muscarinic receptor inhibits cardiac hypertrophy induced by angiotensin II. *J Transl Med* 2013;**11**:209.
84. Liu Y, Sun L, Pan Z, Bai Y, Wang N, Zhao J, Xu C, Li Z, Li B, Du Z, Lu Y, Gao X, Yang B. Overexpression of M(3) muscarinic receptor is a novel strategy for preventing sudden cardiac death in transgenic mice. *Mol Med* 2011;**17**:1179–1187.
85. Liu Y, Sun HL, Li DL, Wang LY, Gao Y, Wang YP, Du ZM, Lu YJ, Yang BF. Choline produces antiarrhythmic actions in animal models by cardiac M3 receptors: improvement of intracellular Ca²⁺ handling as a common mechanism. *Can J Physiol Pharmacol* 2008;**86**:860–865.
86. Zhao L, Chen T, Hang P, Li W, Guo J, Pan Y, Du J, Zheng Y, Du Z. Choline attenuates cardiac fibrosis by inhibiting p38MAPK signaling possibly by acting on M3 muscarinic acetylcholine receptor. *Front Pharmacol* 2019;**10**:1386.
87. Chen X, Bai Y, Sun H, Su Z, Guo J, Sun C, Du Z. Overexpression of M3 muscarinic receptor suppressed adverse electrical remodeling in hypertrophic myocardium via increasing repolarizing K⁺ currents. *Cell Physiol Biochem* 2017;**43**:915–925.
88. Intachai K, Chattipakorn SC, Chattipakorn N, Shinlapawittayatorn K. Revisiting the cardioprotective effects of acetylcholine receptor activation against myocardial ischemia/reperfusion injury. *Int J Mol Sci* 2018;**19**:2466.
89. Prathumsap N, Ongnok B, Khuanjing T, Arinno A, Manechote C, Apajai N, Chunchai T, Arunsak B, Shinlapawittayatorn K, Chattipakorn SC, Chattipakorn N. Acetylcholine receptor agonists provide cardioprotection in doxorubicin-induced cardiotoxicity via modulating muscarinic M2 and alpha7 nicotinic receptor expression. *Transl Res* 2022;**243**:33–51.

90. Caru M, Corbin D, Perie D, Lemay V, Delfrate J, Drouin S, Bertout L, Krajcinovic M, Laverdiere C, Andelfinger G, Sinnett D, Curnier D. Doxorubicin treatments induce significant changes on the cardiac autonomic nervous system in childhood acute lymphoblastic leukemia long-term survivors. *Clin Res Cardiol* 2019;**108**:1000–1008.
91. Burczyk MS, Burkhalter MD, Tena TC, Grisanti LA, Kauk M, Matysik S, Donow C, Kustermann M, Rothe M, Cui Y, Raad F, Laue S, Moretti A, Zimmermann WH, Wess J, Kuhl M, Hoffmann C, Tilley DG, Philipp M. Muscarinic receptors promote pacemaker fate at the expense of secondary conduction system tissue in zebrafish. *JCI Insight* 2019;**4**:e121971.
92. Ishise H, Asanoi H, Ishizaka S, Joho S, Kameyama T, Umeno K, Inoue H. Time course of sympathovagal imbalance and left ventricular dysfunction in conscious dogs with heart failure. *J Appl Physiol* (1985) 1998;**84**:1234–1241.
93. Kusumoto FM, Schoenfeld MH, Barrett C, Edgerton JR, Ellenbogen KA, Gold MR, Goldschlager NF, Hamilton RM, Joglar JA, Kim RJ, Lee R, Marine JE, McLeod CJ, Oken KR, Patton KK, Pellegrini CN, Selzman KA, Thompson A, Varosy PD. 2018 ACC/AHA/HRS guideline on the evaluation and management of patients with bradycardia and cardiac conduction delay: a report of the American college of cardiology/American heart association task force on clinical practice guidelines and the heart rhythm society. *Circulation* 2019;**140**:e382–e482.
94. Matera MG, Cazzola M. Muscarinic receptor antagonists. *Handb Exp Pharmacol* 2017;**237**: 41–62.
95. Cassambai S, Mee CJ, Renshaw D, Hussain A. Tiotropium bromide, a long acting muscarinic receptor antagonist triggers intracellular calcium signalling in the heart. *Toxicol Appl Pharmacol* 2019;**384**:114778.
96. Kruse AC, Hu J, Pan AC, Arlow DH, Rosenbaum DM, Rosemond E, Green HF, Liu T, Chae PS, Dror RO, Shaw DE, Weis WI, Wess J, Kobilka BK. Structure and dynamics of the M3 muscarinic acetylcholine receptor. *Nature* 2012;**482**:552–556.
97. Kruse AC, Li J, Hu J, Kobilka BK, Wess J. Novel insights into M3 muscarinic acetylcholine receptor physiology and structure. *J Mol Neurosci* 2014;**53**:316–323.
98. Digby GJ, Shirey JK, Conn PJ. Allosteric activators of muscarinic receptors as novel approaches for treatment of CNS disorders. *Mol Biosyst* 2010;**6**:1345–1354.
99. Bock A, Schrage R, Mohr K. Allosteric modulators targeting CNS muscarinic receptors. *Neuropharmacology* 2018;**136**:427–437.
100. Hein L, Barsh GS, Pratt RE, Dzau VJ, Kobilka BK. Behavioural and cardiovascular effects of disrupting the angiotensin II type-2 receptor in mice. *Nature* 1995;**377**:744–747.
101. Sugino H, Ozono R, Kurisu S, Matsuura H, Ishida M, Oshima T, Kambe M, Teranishi Y, Masaki H, Matsubara H. Apoptosis is not increased in myocardium overexpressing type 2 angiotensin II receptor in transgenic mice. *Hypertension* 2001;**37**:1394–1398.
102. Curnow KM, Pascoe L, White PC. Genetic analysis of the human type-1 angiotensin II receptor. *Mol Endocrinol* 1992;**6**:1113–1118.
103. de Gasparo M, Catt KJ, Inagami T, Wright JW, Unger T. International union of pharmacology. XXIII. The angiotensin II receptors. *Pharmacol Rev* 2000;**52**:415–472.
104. Forrester SJ, Booz GVV, Sigmund CD, Coffman TM, Kawai T, Rizzo V, Scalia R, Eguchi S. Angiotensin II signal transduction: an update on mechanisms of physiology and pathophysiology. *Physiol Rev* 2018;**98**:1627–1738.
105. Karnik SS, Unal H, Kemp JR, Tirupula KC, Eguchi S, Vanderheyden PM, Thomas WG. International union of basic and clinical pharmacology. XCIX. Angiotensin receptors: interpreters of pathophysiological angiotensinergic stimuli [corrected]. *Pharmacol Rev* 2015;**67**: 754–819.
106. Zhang H, Luginina A, Mishin A, Baidya M, Shukla AK, Cherezov V. Structural insights into ligand recognition and activation of angiotensin receptors. *Trends Pharmacol Sci* 2021;**42**: 577–587.
107. Jacques D, Abdel Malak NA, Sader S, Perreault C. Angiotensin II and its receptors in human endocardial endothelial cells: role in modulating intracellular calcium. *Can J Physiol Pharmacol* 2003;**81**:259–266.
108. Bkaily G, Sleiman S, Stephan J, Asselin C, Choufani S, Kamal M, Jacques D, Gobeil F Jr, D'Orleans-Juste P. Angiotensin II AT1 receptor internalization, translocation and de novo synthesis modulate cytosolic and nuclear calcium in human vascular smooth muscle cells. *Can J Physiol Pharmacol* 2003;**81**:274–287.
109. Kawano H, Do YS, Kawano Y, Starnes V, Barr M, Law RE, Hsueh WA. Angiotensin II has multiple profibrotic effects in human cardiac fibroblasts. *Circulation* 2000;**101**:1130–1137.
110. Wharton J, Morgan K, Rutherford RA, Catravas JD, Chester A, Whitehead BF, De Leval MR, Yacoub MH, Polak JM. Differential distribution of angiotensin AT2 receptors in the Normal and failing human heart. *J Pharmacol Exp Ther* 1998;**284**:323–336.
111. Nataraj C, Oliverio MI, Mannon RB, Mannon PJ, Audoly LP, Amuchastegui CS, Ruiz P, Smithies O, Coffman TM. Angiotensin II regulates cellular immune responses through a calcineurin-dependent pathway. *J Clin Invest* 1999;**104**:1693–1701.
112. Tsutsumi Y, Matsubara H, Ohkubo N, Mori Y, Nozawa M, Murasawa S, Kijima K, Maruyama K, Masaki H, Moriguchi Y, Shibasaki Y, Kamihata H, Inada M, Iwasaka T. Angiotensin II type 2 receptor is upregulated in human heart with interstitial fibrosis, and cardiac fibroblasts are the major cell type for its expression. *Circ Res* 1998;**83**:1035–1046.
113. Regitz-Zagrosek V, Friedel N, Heymann A, Bauer N, Neuss M, Rolfs A, Steffen C, Hildebrandt A, Hetzer R, Fleck E. Regulation, chamber localization, and subtype distribution of angiotensin II receptors in human hearts. *Circulation* 1995;**91**:1461–1471.
114. Abadir PM, Foster DB, Crow M, Cooke CA, Rucker JJ, Jain A, Smith BJ, Burks TN, Cohn RD, Fedarko NS, Carey RM, O'Rourke B, Walston JD. Identification and characterization of a functional mitochondrial angiotensin system. *Proc Natl Acad Sci U S A* 2011;**108**: 14849–14854.
115. Tadevosyan A, Maguy A, Villeneuve LR, Babin J, Bonnefoy A, Allen BG, Nattel S. Nuclear-delimited angiotensin receptor-mediated signaling regulates cardiomyocyte gene expression. *J Biol Chem* 2010;**285**:22338–22349.
116. Rakesh K, Yoo B, Kim IM, Salazar N, Kim KS, Rockman HA. Beta-arrestin-biased agonism of the angiotensin receptor induced by mechanical stress. *Sci Signal* 2010;**3**:ra46.
117. Zou Y, Akazawa H, Qin Y, Sano M, Takano H, Minamino T, Makita N, Iwanaga K, Zhu W, Kudoh S, Toko H, Tamura K, Kihara M, Nagai T, Fukamizu A, Umemura S, Iiri T, Fujita T, Komuro I. Mechanical stress activates angiotensin II type 1 receptor without the involvement of angiotensin II. *Nat Cell Biol* 2004;**6**:499–506.
118. Sauliere A, Bellot M, Paris H, Denis C, Finana F, Hansen JT, Altie MF, Seguelas MH, Pathak A, Hansen JL, Senard JM, Gales C. Deciphering biased-agonism complexity reveals a new active AT1 receptor entity. *Nat Chem Biol* 2012;**8**:622–630.
119. Wei H, Ahn S, Shenoy SK, Karnik SS, Hunyady L, Luttrell LM, Lefkowitz RJ. Independent beta-arrestin 2 and G protein-mediated pathways for angiotensin II activation of extracellular signal-regulated kinases 1 and 2. *Proc Natl Acad Sci U S A* 2003;**100**:10782–10787.
120. Winkler LM, Elgeti M, Hilger D, Latorraca NR, Lerch MT, Staus DP, Dror RO, Kobilka BK, Hubbell WL, Lefkowitz RJ. Angiotensin analogs with divergent bias stabilize distinct receptor conformations. *Cell* 2019;**176**:468–478 e411.
121. Winkler LM, Skiba MA, McMahon C, Staus DP, Kleinhenz ALW, Suomivuori CM, Latorraca NR, Dror RO, Lefkowitz RJ, Kruse AC. Angiotensin and biased analogs induce structurally distinct active conformations within a GPCR. *Science* 2020;**367**:888–892.
122. Pfeiffer CT, Wang J, Paulo JA, Jiang X, Gygi SP, Rockman HA. Mapping angiotensin II type 1 receptor-biased signaling using proximity labeling and proteomics identifies diverse actions of biased agonists. *J Proteome Res* 2021;**20**:3256–3267.
123. Rajagopal K, Whalen EJ, Violin JD, Stiber JA, Rosenberg PB, Premont RT, Coffman TM, Rockman HA, Lefkowitz RJ. Beta-arrestin2-mediated inotropic effects of the angiotensin II type 1A receptor in isolated cardiac myocytes. *Proc Natl Acad Sci U S A* 2006;**103**: 16284–16289.
124. Violin JD, DeWire SM, Yamashita D, Rominger DH, Nguyen L, Schiller K, Whalen EJ, Gowen M, Lark MW. Selectively engaging beta-arrestins at the angiotensin II type 1 receptor reduces blood pressure and increases cardiac performance. *J Pharmacol Exp Ther* 2010;**335**:572–579.
125. Kim KS, Abraham D, Williams B, Violin JD, Mao L, Rockman HA. Beta-Arrestin-biased AT1R stimulation promotes cell survival during acute cardiac injury. *Am J Physiol Heart Circ Physiol* 2012;**303**:H1001–H1010.
126. Freer RJ, Pappano AJ, Peach MJ, Bing KT, McLean MJ, Vogel S, Sperelakis N. Mechanism for the positive inotropic effect of angiotensin II on isolated cardiac muscle. *Circ Res* 1976;**39**: 178–183.
127. Abraham DM, Davis RT III, Warren CM, Mao L, Wolska BM, Solaro RJ, Rockman HA. Beta-Arrestin mediates the frank-starling mechanism of cardiac contractility. *Proc Natl Acad Sci U S A* 2016;**113**:14426–14431.
128. Monasky MM, Taglieri DM, Henze M, Warren CM, Utter MS, Soergel DG, Violin JD, Solaro RJ. The beta-arrestin-biased ligand TRV120023 inhibits angiotensin II-induced cardiac hypertrophy while preserving enhanced myofibrillar response to calcium. *Am J Physiol Heart Circ Physiol* 2013;**305**:H856–H866.
129. Tarigopula M, Davis RT III, Mungai PT, Ryba DM, Wieczorek DF, Cowan CL, Violin JD, Wolska BM, Solaro RJ. Cardiac myosin light chain phosphorylation and inotropic effects of a biased ligand, TRV120023, in a dilated cardiomyopathy model. *Cardiovasc Res* 2015;**107**:226–234.
130. Sadoshima J, Izumo S. Molecular characterization of angiotensin II-induced hypertrophy of cardiac myocytes and hyperplasia of cardiac fibroblasts. Critical role of the AT1 receptor subtype. *Circ Res* 1993;**73**:413–423.
131. Zhai P, Yamamoto M, Galeotti J, Liu J, Masurekar M, Thaisz J, Irie K, Holle E, Yu X, Kupersmidt S, Roden DM, Wagner T, Yatani A, Vatner DE, Vatner SF, Sadoshima J. Cardiac-specific overexpression of AT1 receptor mutant lacking G alpha q/G alpha i coupling causes hypertrophy and bradycardia in transgenic mice. *J Clin Invest* 2005;**115**: 3045–3056.
132. Ainscough JF, Drinkhill MJ, Sedo A, Turner NA, Brooke DA, Balmforth AJ, Ball SG. Angiotensin II type-1 receptor activation in the adult heart causes blood pressure-independent hypertrophy and cardiac dysfunction. *Cardiovasc Res* 2009;**81**: 592–600.
133. Paradis P, Dali-Youcef N, Paradis FW, Thibault G, Nemer M. Overexpression of angiotensin II type I receptor in cardiomyocytes induces cardiac hypertrophy and remodeling. *Proc Natl Acad Sci U S A* 2000;**97**:931–936.
134. Mazzolai L, Nussberger J, Aubert JF, Brunner DB, Gabbiani G, Brunner HR, Pedrazzini T. Blood pressure-independent cardiac hypertrophy induced by locally activated renin-angiotensin system. *Hypertension* 1998;**31**:1324–1330.
135. Adams JW, Sakata Y, Davis MG, Sah VP, Wang Y, Liggett SB, Chien KR, Brown JH, Dorn GW II. Enhanced galphaq signaling: a common pathway mediates cardiac hypertrophy and apoptotic heart failure. *Proc Natl Acad Sci U S A* 1998;**95**:10140–10145.
136. Akhter SA, Luttrell LM, Rockman HA, Iaccarino G, Lefkowitz RJ, Koch WJ. Targeting the receptor-gq interface to inhibit in vivo pressure overload myocardial hypertrophy. *Science* 1998;**280**:574–577.
137. Rajagopal K, Lefkowitz RJ, Rockman HA. When 7 transmembrane receptors are not G protein-coupled receptors. *J Clin Invest* 2005;**115**:2971–2974.
138. Revankar CM, Vines CM, Cimino DF, Prossnitz ER. Arrestins block G protein-coupled receptor-mediated apoptosis. *J Biol Chem* 2004;**279**:24578–24584.

139. Billet S, Bardin S, Verp S, Baudrie V, Michaud A, Conchon S, Muffat-Joly M, Escoubet B, Souil E, Hamard G, Bernstein KE, Gasc JM, Elghozi JL, Corvol P, Clauser E. Gain-of-function mutant of angiotensin II receptor, type 1A, causes hypertension and cardiovascular fibrosis in mice. *J Clin Invest* 2007;**117**:1914–1925.
140. Rivard K, Grandy SA, Douillette A, Paradis P, Nemer M, Allen BG, Fiset C. Overexpression of type 1 angiotensin II receptors impairs excitation-contraction coupling in the mouse heart. *Am J Physiol Heart Circ Physiol* 2011;**301**:H2018–H2027.
141. Bridgman P, Aronovitz MA, Kakkar R, Oliverio MI, Coffman TM, Rand WM, Konstam MA, Mendelsohn ME, Patten RD. Gender-specific patterns of left ventricular and myocyte remodeling following myocardial infarction in mice deficient in the angiotensin II type 1a receptor. *Am J Physiol Heart Circ Physiol* 2005;**289**:H586–H592.
142. Harada K, Komuro I, Shiojima I, Hayashi D, Kudoh S, Mizuno T, Kijima K, Matsubara H, Sugaya T, Murakami K, Yazaki Y. Pressure overload induces cardiac hypertrophy in angiotensin II type 1A receptor knockout mice. *Circulation* 1998;**97**:1952–1959.
143. Crowley SD, Gurley SB, Herrera MJ, Ruiz P, Griffiths R, Kumar AP, Kim HS, Smithies O, Le TH, Coffman TM. Angiotensin II causes hypertension and cardiac hypertrophy through its receptors in the kidney. *Proc Natl Acad Sci U S A* 2006;**103**:17985–17990.
144. Sparks MA, Rianto F, Diaz E, Revoori R, Hoang T, Bouknight L, Stegbauer J, Vivekanandan-Giri A, Ruiz P, Pennathur S, Abraham DM, Gurley SB, Crowley SD, Coffman TM. Direct actions of AT1 (type 1 angiotensin) receptors in cardiomyocytes do not contribute to cardiac hypertrophy. *Hypertension* 2021;**77**:393–404.
145. Campbell SE, Katwa LC. Angiotensin II stimulated expression of transforming growth factor-beta1 in cardiac fibroblasts and myofibroblasts. *J Mol Cell Cardiol* 1997;**29**:1947–1958.
146. Fredj S, Bescond J, Louault C, Delwail A, Lecron JC, Potreau D. Role of interleukin-6 in cardiomyocyte/cardiac fibroblast interactions during myocyte hypertrophy and fibroblast proliferation. *J Cell Physiol* 2005;**204**:428–436.
147. Lyu L, Wang H, Li B, Qin Q, Qi L, Nagarkatti M, Nagarkatti P, Janicki JS, Wang XL, Cui T. A critical role of cardiac fibroblast-derived exosomes in activating renin angiotensin system in cardiomyocytes. *J Mol Cell Cardiol* 2015;**89**:268–279.
148. Schultz Jel J, Witt SA, Glascock BJ, Nieman ML, Reiser PJ, Nix SL, Kimball TR, Doetschman T. TGF-beta1 mediates the hypertrophic cardiomyocyte growth induced by angiotensin II. *J Clin Invest* 2002;**109**:787–796.
149. Wang J, Hanada K, Gareri C, Rockman HA. Mechanoactivation of the angiotensin II type 1 receptor induces beta-arrestin-biased signaling through galpha coupling. *J Cell Biochem* 2018;**119**:3586–3597.
150. Tang W, Strachan RT, Lefkowitz RJ, Rockman HA. Allosteric modulation of beta-arrestin-biased angiotensin II type 1 receptor signaling by membrane stretch. *J Biol Chem* 2014;**289**:28271–28283.
151. Malhotra R, Sadoshima J, Brosius FC III, Izumo S. Mechanical stretch and angiotensin II differentially upregulate the renin-angiotensin system in cardiac myocytes in vitro. *Circ Res* 1999;**85**:137–146.
152. Kagiya S, Eguchi S, Frank GD, Inagami T, Zhang YC, Phillips MI. Angiotensin II-induced cardiac hypertrophy and hypertension are attenuated by epidermal growth factor receptor antisense. *Circulation* 2002;**106**:909–912.
153. Brunner HR, Gavras H, Laragh JH. Angiotensin-II blockade in man by sar1-ala8-angiotensin II for understanding and treatment of high blood-pressure. *Lancet* 1973;**2**:1045–1048.
154. Singh KD, Karnik SS. Angiotensin type 1 receptor blockers in heart failure. *Curr Drug Targets* 2020;**21**:125–131.
155. Pang PS, Butler J, Collins SP, Cotter G, Davison BA, Ezekowitz JA, Filippatos G, Levy PD, Metra M, Ponikowski P, Teerlink JR, Voors AA, Bharucha D, Goin K, Soergel DG, Felker GM. Biased ligand of the angiotensin II type 1 receptor in patients with acute heart failure: a randomized, double-blind, placebo-controlled, phase IIB, dose ranging trial (BLAST-AHF). *Eur Heart J* 2017;**38**:2364–2373.
156. Ryba DM, Li J, Cowan CL, Russell B, Wolska BM, Solaro RJ. Long-Term biased beta-arrestin signaling improves cardiac structure and function in dilated cardiomyopathy. *Circulation* 2017;**135**:1056–1070.
157. Kashiwara T, Kawagishi H, Nakada T, Numaga-Tomita T, Kadota S, Wolf EE, Du CK, Shiba Y, Morimoto S, Yamada M. Beta-Arrestin-Biased AT1 agonist TRV027 causes a neonatal-specific sustained positive inotropic effect without increasing heart rate. *JACC Basic Transl Sci* 2020;**5**:1057–1069.
158. Kashiwara T, Nakada T, Kojima K, Takeshita T, Yamada M. Angiotensin II activates CaV1.2 channels through beta-arrestin2 and casein kinase 2 in mouse immature cardiomyocytes. *J Physiol* 2017;**595**:4207–4225.
159. Nakajima M, Mukoyama M, Pratt RE, Horiuchi M, Dzau VJ. Cloning of cDNA and analysis of the gene for mouse angiotensin II type 2 receptor. *Biochem Biophys Res Commun* 1993;**197**:393–399.
160. Zhang H, Han GW, Balyuk A, Ishchenko A, White KL, Patel N, Sadybekov A, Zamylyny B, Rudd MT, Hollenstein K, Tolstikova A, White TA, Hunter MS, Weierstall U, Liu W, Babaoglu K, Moore EL, Katz RD, Shipman JM, Garcia-Calvo M, Sharma S, Sheth P, Soisson SM, Stevens RC, Katritch V, Cherezov V. Structural basis for selectivity and diversity in angiotensin II receptors. *Nature* 2017;**544**:327–332.
161. Porrello ER, Delbridge LM, Thomas WG. The angiotensin II type 2 (AT2) receptor: an enigmatic seven transmembrane receptor. *Front Biosci (Landmark Ed)* 2009;**14**:958–972.
162. Asada H, Inoue A, Ngako Kadji FM, Hirata K, Shiimura Y, Im D, Shimamura T, Nomura N, Iwanari H, Hamakubo T, Kusano-Arai O, Hisano H, Uemura T, Suno C, Aoki J, Iwata S. The crystal structure of angiotensin II type 2 receptor with endogenous peptide hormone. *Structure* 2020;**28**:418–425 e414.
163. Bosnyak S, Jones ES, Christopoulos A, Aguilar MI, Thomas WG, Widdop RE. Relative affinity of angiotensin peptides and novel ligands at AT1 and AT2 receptors. *Clin Sci (Lond)* 2011;**121**:297–303.
164. Mendoza-Torres E, Riquelme JA, Vielma A, Sagredo AR, Gabrielli L, Bravo-Sagua R, Jalil JE, Rothermel BA, Sanchez G, Ocaranza MP, Lavandero S. Protection of the myocardium against ischemia/reperfusion injury by angiotensin-(1–9) through an AT2R and akt-dependent mechanism. *Pharmacol Res* 2018;**135**:112–121.
165. Matsumoto T, Ozono R, Oshima T, Matsuura H, Sueda T, Kajiyama G, Kambe M. Type 2 angiotensin II receptor is downregulated in cardiomyocytes of patients with heart failure. *Cardiovasc Res* 2000;**46**:73–81.
166. Toedebusch R, Belenchia A, Pulakat L. Cell-Specific protective signaling induced by the novel AT2R-agonist NP-6A4 on human endothelial and smooth muscle cells. *Front Pharmacol* 2018;**9**:928.
167. Booz GW, Baker KM. Role of type 1 and type 2 angiotensin receptors in angiotensin II-induced cardiomyocyte hypertrophy. *Hypertension* 1996;**28**:635–640.
168. Blaw BL, Stewart JM, Bourassa E, Katovich MJ, Walter G, Speth RC, Summers C, Raizada MK. Angiotensin II type 2 receptor gene transfer elicits cardioprotective effects in an angiotensin II infusion rat model of hypertension. *Physiol Genomics* 2004;**19**:255–261.
169. Wu L, Iwai M, Nakagami H, Chen R, Suzuki J, Akishita M, de Gasparo M, Horiuchi M. Effect of angiotensin II type 1 receptor blockade on cardiac remodeling in angiotensin II type 2 receptor null mice. *Arterioscler Thromb Vasc Biol* 2002;**22**:49–54.
170. D'Amore A, Black MJ, Thomas WG. The angiotensin II type 2 receptor causes constitutive growth of cardiomyocytes and does not antagonize angiotensin II type 1 receptor-mediated hypertrophy. *Hypertension* 2005;**46**:1347–1354.
171. Yan X, Price RL, Nakayama M, Ito K, Schuldt AJ, Manning VJ, Sanbe A, Borg TK, Robbins J, Lorell BH. Ventricular-specific expression of angiotensin II type 2 receptors causes dilated cardiomyopathy and heart failure in transgenic mice. *Am J Physiol Heart Circ Physiol* 2003;**285**:H2179–H2187.
172. Brede M, Roell W, Ritter O, Wiesmann F, Jahns R, Haase A, Fleischmann BK, Hein L. Cardiac hypertrophy is associated with decreased eNOS expression in angiotensin AT2 receptor-deficient mice. *Hypertension* 2003;**42**:1177–1182.
173. Ichihara S, Senbonmatsu T, Price E Jr, Ichiki T, Gaffney FA, Inagami T. Angiotensin II type 2 receptor is essential for left ventricular hypertrophy and cardiac fibrosis in chronic angiotensin II-induced hypertension. *Circulation* 2001;**104**:346–351.
174. Xu J, Sun Y, Carretero OA, Zhu L, Harding P, Shesely EG, Dai X, Rhabale NE, Peterson E, Yang XP. Effects of cardiac overexpression of the angiotensin II type 2 receptor on remodeling and dysfunction in mice post-myocardial infarction. *Hypertension* 2014;**63**:1251–1259.
175. Lauer D, Slavic S, Sommerfeld M, Thone-Reineke C, Sharkovska Y, Hallberg A, Dahlof B, Kintscher U, Unger T, Steckelings UM, Kaschina E. Angiotensin type 2 receptor stimulation ameliorates left ventricular fibrosis and dysfunction via regulation of tissue inhibitor of matrix metalloproteinase 1/matrix metalloproteinase 9 axis and transforming growth factor beta1 in the rat heart. *Hypertension* 2014;**63**:e60–e67.
176. Ocaranza M P, Riquelme JA, Garcia L, Jalil JE, Chiong M, Santos RAS, Lavandero S. Counter-regulatory renin-angiotensin system in cardiovascular disease. *Nat Rev Cardiol* 2020;**17**:116–129.
177. Mazarura GR, Dallagnol JCC, Chatenet D, Allen BG, Hebert TE. The complicated lives of GPCRs in cardiac fibroblasts. *Am J Physiol Cell Physiol* 2022;**323**:C813–C822.
178. Irannejad R, Pessino V, Mika D, Huang B, Wedegaertner PB, Conti M, von Zastrow M. Functional selectivity of GPCR-directed drug action through location bias. *Nat Chem Biol* 2017;**13**:799–806.
179. Nguyen AH, Thomsen ARB, Cahill TJ III, Huang R, Huang LY, Marcink T, Clarke OB, Heissel S, Masoudi A, Ben-Hail D, Samaan F, Dandey VP, Tan YZ, Hong C, Mahoney JP, Triest S, Little J IV, Chen X, Sunahara R, Steyaert J, Molina H, Yu Z, des Georges A, Lefkowitz RJ. Structure of an endosomal signaling GPCR-G protein-beta-arrestin megacomplex. *Nat Struct Mol Biol* 2019;**26**:1123–1131.
180. Thomsen ARB, Plouffe B, Cahill TJ III, Shukla AK, Tarrasch JT, Dosey AM, Khsai AV, Strachan RT, Pani B, Mahoney JP, Huang L, Breton B, Heydenreich FM, Sunahara RK, Skiniotis G, Bouvier M, Lefkowitz RJ. GPCR-G Protein-beta-Arrestin super-Complex mediates sustained G protein signaling. *Cell* 2016;**166**:907–919.