

RESEARCH ARTICLE



Cite this: *RSC Med. Chem.*, 2023, 14, 957

Biological evaluation of novel amidino substituted coumarin-benzazole hybrids as promising therapeutic agents†

Anja Beč, ^a Livio Racané, ^b Lucija Žonja,^a Leentje Persoons, ^c
Dirk Daelemans, ^c Kristina Starčević,^d Robert Vianello ^e and Marijana Hranjec ^{*a}

Herein we present the design and the synthesis of novel substituted coumarin-benzimidazole/benzothiazole hybrids bearing a cyclic amidino group on the benzazole core as biologically active agents. All prepared compounds were evaluated for their *in vitro* antiviral and antioxidative activity as well as for their *in vitro* antiproliferative activity against a panel of several human cancer cell lines. Coumarin-benzimidazole hybrid **10** (EC₅₀ 9.0–43.8 μM) displayed the most promising broad spectrum antiviral activity, while two other coumarin-benzimidazole hybrids **13** and **14** showed the highest antioxidative capacity in the ABTS assay, superior to the reference standard BHT (IC₅₀ 0.17 and 0.11 mM, respectively). Computational analysis supported these results and demonstrated that these hybrids benefit from the high C–H hydrogen atom releasing tendency of the cationic amidine unit, and the pronounced ease with which they can liberate an electron, promoted by the electron-donating diethylamine group on the coumarin core. The coumarin ring substitution at position 7 with a *N,N*-diethylamino group also caused a significant enhancement of the antiproliferative activity, with the most active compounds being derivatives with a 2-imidazolyl amidine group **13** (IC₅₀ 0.3–1.9 μM) and benzothiazole derivative with a hexacyclic amidine group **18** (IC₅₀ 1.3–2.0 μM).

Received 2nd February 2023,
Accepted 27th March 2023

DOI: 10.1039/d3md00055a

rsc.li/medchem

Introduction

Molecular hybridization and the synthesis of hybrid molecules has been a major focus of medicinal chemists for over two decades.¹ The main purpose of combining two or more structurally different and biologically active scaffolds is to improve the activity and the affinity of newly synthesized compounds, to overcome drug resistance, to reduce side effects or to optimize various physico-chemical parameters including ADMET properties.² Usually these hybrid molecules contain a naturally active fragment combined with a

synthetically prepared pharmacophoric group giving rise to new molecules with great and diverse biological potential.³ Among all known heterocyclic derivatives, nitrogen heterocycles have been widely used in the rational design of novel molecules with improved biological properties.^{4a,b,5a,b} The most important benzazole representatives, benzimidazole and benzothiazole derivatives, have been well established as principal structural motifs in medicinal and pharmaceutical chemistry.^{6a,b,7,8} Numerous publications and reviews have described the biological potential and versatile properties of suchlike derivatives displaying prominent antitumoral, antimicrobial, antiviral, antioxidative, anti-inflammatory and antihistaminic activities.^{9a,b,10,11} We have previously reported that the biological activity could significantly be improved by the incorporation of an amidine group in the cationic form, usually placed at the termini of the molecules, on the benzazole scaffold.^{12–14} Consequently, the presence of this amidine group could improve the interaction with the putative biological target since the complex between the molecule and the target is further stabilized through both H-bonding and electrostatic interactions.^{15,16} On the other hand, the flavonoid coumarin and its derivatives comprise an extensive class of both naturally occurring and synthetic compounds that also exhibit remarkable pharmacological activities combined with generally low toxicity.^{17a,b,18a,b}

^a Department of Organic Chemistry, Faculty of Chemical Engineering and Technology, University of Zagreb, Marulićev trg 19, HR-10000 Zagreb, Croatia. E-mail: mhranjec@fkit.hr

^b Department of Applied Chemistry, Faculty of Textile Technology, University of Zagreb, Prilaz baruna Filipovića 28a, 10000 Zagreb, Croatia

^c KU Leuven, Department of Microbiology, Immunology and Transplantation, Laboratory of Virology and Chemotherapy, Rega Institute, Leuven, Belgium

^d Department of Chemistry and Biochemistry, Faculty of Veterinary Medicine, University of Zagreb, Heinzelova 55, HR-10000 Zagreb, Croatia

^e Laboratory for the Computational Design and Synthesis of Functional Materials, Division of Organic Chemistry and Biochemistry, Ruđer Bošković Institute, Zagreb, Croatia

† Electronic supplementary information (ESI) available: Experimental biology and computational details, 1H and 13C NMR spectra. See DOI: <https://doi.org/10.1039/d3md00055a>

Countless coumarin derivatives have been reported, and they still remain one of the most important substructures in rational drug design and discovery.^{19a,b,20,21} It is well established that coumarins readily interact with diverse biological targets and active sites through the formation of noncovalent interactions, such as hydrophobic and electrostatic interactions, π - π interactions and/or by forming hydrogen or van der Waals bonds.^{22,23} Their interesting chemical, physical and spectroscopic properties due to the structural planarity and the reactivity of both the benzene and the pyrone ring make coumarins compelling in a diverse array of research fields.^{24,25}

Recently, several studies were published describing the design, synthesis and biological evaluation of coumarin-benzazole hybrids. A group of authors synthesized coumarin-benzimidazole hybrids, namely 3-(1*H*-benzo[*d*]imidazol-2-yl)-7-(substituted amino)-2*H*-chromen-2-one derivatives, which were tested for their *in vitro* antitumor activity against a panel of 60 cancer cell lines.²⁶ The 7-ethanolamino substituted derivative showed promising activity towards leukemia, colon and breast cancer cell lines. Coumarin-benzimidazole hybrids consisting of two series, namely derivatives with benzimidazole nuclei directly attached to the coumarin ring as well as derivatives with an amide bond connecting the coumarin and benzimidazole nuclei were synthesized to explore their antioxidative and anti-inflammatory potential.²⁷ Another paper described the synthesis of a coumarin-benzothiazole-chlorambucil conjugate for photocontrolled release of the anticancer drug chlorambucil. The study confirmed that this conjugate displayed improved cytotoxicity in comparison to unconjugated chlorambucil due to effective intracellular release of the drug.²⁸ A coumarin-benzothiazole ratiometric ATP probe has been designed and synthesized in order to detect ATP in aqueous solutions. This sensor showed high selectivity over other nucleotide polyphosphate (NPP) anions, which is promising for the development and application of suchlike molecules for various biological assays.²⁹ Benzothiazolyl substituted coumarin derivatives were tested for their *in vitro* antiproliferative activity against the MCF-7 cell line, while molecular docking was performed on receptor tyrosine kinases (RTKs) as a possible molecular target.³⁰ Another group of authors designed and synthesized benzimidazole-coumarin conjugates with a -SCH₂- linkage between these two heterocyclic moieties, with several conjugates displaying anti-Hepatitis C virus activity.³¹

Recently, we designed and synthesized coumarin-benzazole conjugates which were evaluated for their antiproliferative activity against several cancer cell lines as well as for their antioxidative activity.³² The *N,N*-diethylamino substituted benzimidazole/imidazo[4,5-*b*]pyridine-coumarin hybrids (Fig. 1a) was the most promising derivative with potent and selective activity against CEM acute lymphoblastic leukemia cells. All synthesized compounds also showed moderate antioxidative potential. Encouraged by these findings, we have now designed prepared and evaluated novel coumarin-benzazole hybrids (Fig. 1b).

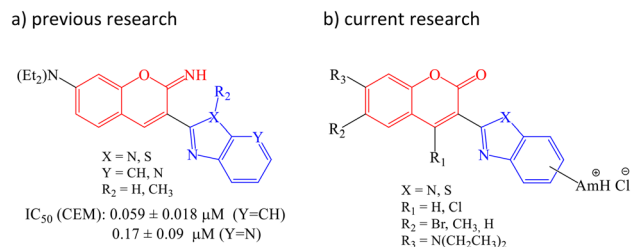


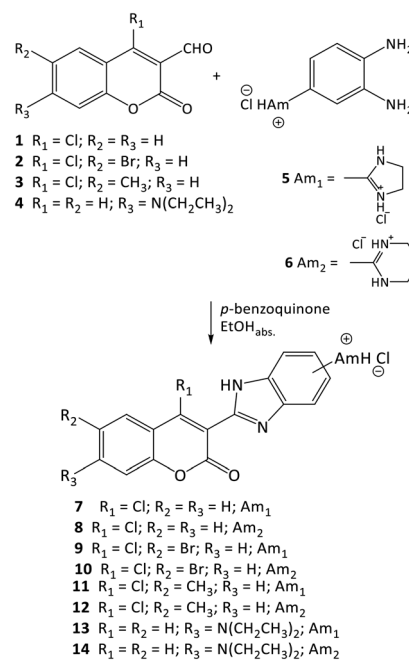
Fig. 1 Synthesized benzazole-coumarin conjugates in previous research (a) and current research (b).

The newly prepared derivatives were substituted at position 3, 4, 6 and/or 7 of the coumarin ring with benzazole (benzimidazole or benzothiazole) nuclei bearing amidine moieties. All newly synthesized compounds were evaluated for their *in vitro* antiviral activity, as well as for their antioxidative potential and antiproliferative activity on a diverse panel of cancer cell lines.

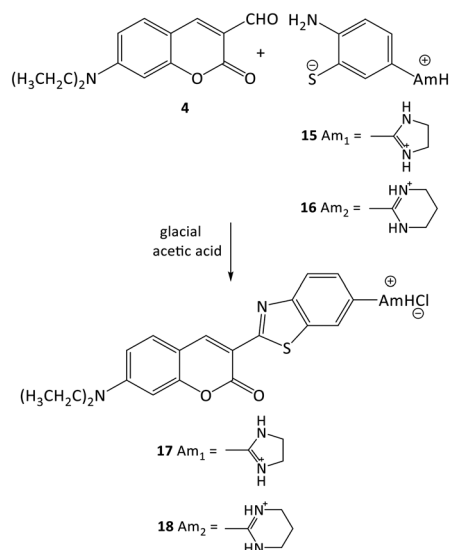
Results and discussion

Chemistry

Coumarin-benzimidazole/benzothiazole hybrids were synthesized by experimental procedures presented in Schemes 1 and 2. The main precursors necessary for the synthesis of the targeted amidino substituted benzimidazole derivatives, namely amidino substituted 1,2-phenylenediamines 5–6 and amidino substituted benzenethiolates 15–16 were synthesized according to previously published procedures that were optimized by our research group. Amidino substituted precursors 5–6 were prepared from the corresponding cyano substituted anilines,



Scheme 1 Synthesis of amidino-substituted coumarin-benzimidazole hybrids 7–14.



Scheme 2 Synthesis of amidino-substituted coumarin-benzothiazole hybrids 17–18.

while precursors 15–16 were both obtained through an acidic Pinner reaction from 6-cyanobenzothiazole in the form of zwitterions as monitored by IR spectroscopy. Targeted amidino substituted benzazole-coumarin conjugates were synthesized by using the reaction of cyclocondensation of corresponding precursors 5, 6, 15 and 16 with chosen commercially available substituted 2-oxo-2H-chromene-3-carbaldehydes 1–4. Benzimidazole-coumarin hybrids 7–14 were synthesized in the cyclocondensation performed in absolute ethanol and by using *p*-benzoquinone as oxidants, in moderate reaction yields. Benzothiazole-coumarin hybrids obtained within the cyclocondensation in refluxing acetic acid followed by quenching with hydrochloric acid in moderate reaction yields. This method is direct condensation of aldehydes with amidino substituted 2-aminothiophenols without using any catalyst or oxidant. Only 7-*N,N*-diethylamino substituted derivatives 17–18 were successfully synthesized while, the synthesis starting from halogeno substituted aldehydes 1–3 failed due to the competitive reaction of nucleophilic substitution.

All newly prepared amidino substituted coumarin-benzazole hybrids were structurally characterized with ¹H and ¹³C NMR spectroscopy. NMR analysis was based on the values of H–H coupling constants in the ¹H spectra as well as chemical shifts in both ¹H and ¹³C spectra, which were in consistence with the proposed structures. The signals for protons on N atoms on amidine groups were observed as bright signals in the region from 9–10 ppm. The signals for methylene protons of amidine group appeared in the aliphatic part of ¹H spectra as well as in the ¹³C NMR spectra. The successful cyclocondensation and formation of benzimidazole nuclei was confirmed with the signal related to the proton of NH group on benzimidazole nuclei in the region around 13 ppm. IR spectroscopy was used for the monitoring of Pinner reaction due to the synthesis of main precursors.

The successful formation of imino-ester was confirmed within the disappearance of the signal for a CN group at around 2200 cm⁻¹.

Cytotoxic and antiviral activity

All synthesized coumarin-benzimidazole 7–14 and coumarin-benzothiazole 17–18 hybrids were tested for their antiviral activity.

The results for the cytotoxic and antiviral activity evaluation of the newly synthesized derivatives are summarized in Table 1.

For clarity, only compounds showing cytotoxic or antiviral activity are depicted. The results are expressed as CC₅₀ (50% cytotoxic concentration) and EC₅₀ (50% effective concentration) values. According to the obtained results we can conclude that the most promising antiviral activity was noted for coumarin-benzimidazole hybrid 10, a 6-bromo-4-chloro substituted derivative bearing a hexacyclic amidine group (Fig. 2). This hybrid displayed broad spectrum antiviral activity with favorable EC₅₀ values ranging from 9.0 μM to 51.0 μM against all tested influenza viruses (subtypes H1N1, H3N2 and B), promising activity towards flaviviruses (EC₅₀ of 11.2 μM against YFV and 9.4 μM against zika virus), and more moderate activity against RSV (73.5 μM), sindbis virus (59.0 μM), and HCoV subtype OC43 (78.0 μM). Furthermore, hybrid 10 completely lacked cytotoxicity on all host cell lines, yielding encouraging selectivity indices. Benzimidazole derivatives 7–8 lacked antiviral activity, whereas the 6-bromo-4-chloro substituted derivative bearing a 2-imidazoliny group 9 showed selective activity towards influenza A virus subtype H1N1. Interestingly, the *N,N*-diethylamino substituted benzimidazole hybrid 13 did not show any antiviral activity but proved markedly cytotoxic on all host cell lines. Among the synthesized benzothiazole derivatives, the *N,N*-diethylamino substituted derivative 18 bearing a hexacyclic amidine group displayed interesting activity against several respiratory viruses (EC₅₀ values of 28.9 and 33.0 μM against HCoV OC43 and 229E, respectively, and 43.4 μM against RSV) as well as anti-HSV type 1 activity (25.5 μM), while the 2-imidazoliny substituted derivative 17 showed only moderate activity against two of these viruses.

Antioxidant activity *in vitro*

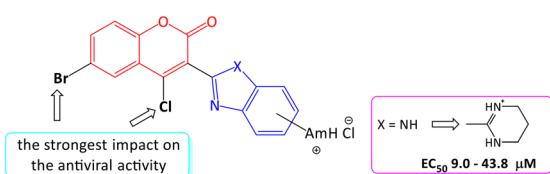
The antioxidant activity of the newly prepared coumarin-benzazole hybrids 7–14 and 17–18 is presented in Table 2. Reference compound BHT (butylated hydroxytoluene) was included in all assays and the results are presented as IC₅₀ values (for DPPH and ABTS) and mmolFe²⁺/mmolC (for FRAP) in Table 2. The antioxidant capacity is measured as the ability of pure compounds to decrease the assay color reacting directly with the ABTS^{•+} radicals which could be used to evaluate hydrophilic and lipophilic compounds as well.

Obtained results revealed that two coumarin-benzimidazole hybrids substituted with *N,N*-diethylamino group at the position 7 bearing either 2-imidazoliny 13 and

Table 1 Cytotoxicity and antiviral activity of hybrids 7–18

Cpd	Cytotoxicity (CC ₅₀ /μM)			Antiviral activity (EC ₅₀ /μM)											
	HEL			HCoV		HCoV		Influenza		Influenza		RSV		Zika	
	229	Huh7	MDCK	229E	OC43	NL63	H1N1	H3N2	Influenza	H3N2	Along	HSV-1 KOS	Yellow fever	Mr776	Sindbis
7	99.5	85	92.1	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
8	87.4	61.7	81.7	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
9	51.6	2.3	59.0	>100	>100	>100	16.2	>100	>100	>100	>100	>100	>100	>100	>100
10	>100	>100	>100	>100	78.0	>100	43.8	51.0	9.0	73.5	>100	11.2	9.4	59	
11	77	<0.8	10.0	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
12	>100	78.7	77.1	>100	74.2	>100	>100	>100	>100	64.1	>100	>100	>100	>100	>100
13	5.1	<0.8	<0.8	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100	>100
14	>100	>100	43.4	>100	>100	>100	>100	>100	>100	73.3	>100	72.6	77.7	>100	>100
17	>100	54.2	64.2	>100	69.7	>100	>100	>100	>100	>100	47.9	>100	>100	>100	>100
18	>100	1.8	4.2	33.0	28.9	>100	>100	>100	>100	43.4	25.5	>100	>100	>100	>100
Rem	>10	>10	—	0.06	0.06	0.03	—	—	—	0.03	—	6.2	0.7	>10	
Rib	>250	8.9	67.0	82.6	170.1	>250	10.5	4.0	2.8	10.8	—	>250	>250	148.1	
Zan	—	—	>100	—	—	—	0.1	16.8	0.05	—	—	—	—	—	—
Rim	—	—	>100	—	—	—	5.0	0.05	>100	—	—	—	—	—	—
BVDU	>100	—	—	—	—	—	—	—	—	—	0.05	—	—	—	—

hexacyclic amidine group **14** have shown the strongest antioxidative capacity, improved in comparison to the standard BHT (IC₅₀ 0.17 and 0.11 mM, respectively). All other benzimidazole derivatives have shown moderate antioxidative capacity. Coumarin-benzothiazole hybrids **17–18** have proven to be less active in comparison to the benzimidazole analogues with the most active derivative bearing hexacyclic amidine group **18**. The radical scavenging method using DPPH stable radical indicates the ability of tested species to donate proton/electron. From the presented results, it can be concluded that all tested coumarin-benzazole hybrids were less active when compared to the standard BHT. The best activity was displayed by 4-chloro substituted benzimidazole derivative bearing hexacyclic amidine group **8** (IC₅₀ 0.07 mM) as well as 6-bromine-4-chloro substituted benzimidazole derivative **13** bearing hexacyclic amidine group (IC₅₀ 0.11 mM). Benzothiazole derivatives **17–18** did not show any results within this assay due to precipitation. The reducing power of tested compounds refers to their antioxidant activity and FRAP assay was used for its determination. The FRAP values showed that all tested compounds are significantly less active relative to the standard BHT. The best activity in this assay was shown by 6-bromine-4-chloro substituted benzimidazole derivative bearing 2-imidazolyl group **9**. Benzothiazole derivatives **17–18** were less active when compared to the benzimidazole analogues. Thus, to conclude, the most promising antioxidative capacity (ABTS assay) was observed for benzimidazole derivatives **13** and **14**

**Fig. 2** Structure–activity relationship regarding the antiviral activity.

(Fig. 3), both substituted with *N,N*-diethylamino group at the position 7 being more active in comparison to BHT standard.

Scavenging the DPPH/ABTS/FRAP radicals by prepared compounds resulted in a decrease in absorption readings over time; the extent of decrease in DPPH/ABTS/FRAP absorption being proportional to the concentration of radicals that are being scavenged. It is well known that antioxidant capacity assays may be classified as electron transfer (SET) and hydrogen atom transfer (HAT)-based assays and that is difficult to distinguish between them. In most situations, these two reactions take place simultaneously. The results in our study emphasize that potential antioxidant capacity of all prepared compounds is based on predominantly on SET mechanism in all three assays.

Computational analysis of the antioxidative activity

Computational analysis was employed to offer additional insights into the structure and properties of studied systems, and to elucidate relevant processes responsible for their antioxidant features. Given a large number of structurally similar systems inspected here, we decided to address a selected number of derivatives, involving **7**, **9** and the most potent **13** and **14**, together with several model systems **m1–m12** (Fig. 4). The latter are selected to allow enough structural and electronic information about the studied compounds, and aid in designing even more potent antioxidants based on the employed synthetic strategy. Data in Table 3 show that each system is characterized by its single-electron ionization energy (IE) and electron affinity (EA), together with the bond dissociation energy (BDE) required to homolytically cleave the hydrogen atom (H[•]) in the thermodynamically most favourable way. In some cases, BDE values for other bonds C–X (X = Cl, Br, NEt₂) are also computed, while all data are placed within a context by repeating the analysis for reference systems used in

Table 2 Antioxidative activity *in vitro* of tested compounds 7–14 and 17–18^a

Compound	ABTS (IC ₅₀ /mM)	DPPH (IC ₅₀ /mM)	FRAP (mmolFe ²⁺ /mmolL)
7	0.37 ± 0.04	0.44 ± 0.01	35.17 ± 0.88
8	0.29 ± 0.03	0.07 ± 0.01	38.37 ± 0.21
9	0.26 ± 0.01	0.13 ± 0.01	130.55 ± 3.54
10	0.36 ± 0.01	0.11 ± 0.00	95.25 ± 1.44
11	0.35 ± 0.02	0.19 ± 0.03	47.13 ± 0.23
12	0.46 ± 0.05	0.52 ± 0.08	47.87 ± 0.25
13	0.17 ± 0.01	0.14 ± 0.02	28.44 ± 0.75
14	0.11 ± 0.01	0.61 ± 0.04	28.83 ± 0.75
17	2.19 ± 0.26	—	2.84 ± 0.71
18	0.98 ± 0.09	—	3.29 ± 0.21
BHT	0.18 ± 0.02	0.025 ± 4.2	2089.34 ± 55.98

^a Values are presented as means ± standard deviation.

experimental assays, namely ABTS⁺, FRAP and DPPH[•] (Table 4). All molecules with the amidine unit were considered as protonated monocations (Fig. 5), based on a typically high basicity of amidine (p*K*_a ≈ 11–13), which surpasses the basicity of benzimidazole (p*K*_a ≈ 5–6), benzothiazole (p*K*_a ≈ 1–2), coumarin (p*K*_a < 0), and diethylaniline fragments (p*K*_a ≈ 6–7).³³

Starting with the reference compounds (Table 4), FRAP is seemingly the simplest system, where it is clear that its radical scavenging ability relies on the electron-accepting SET mechanism that reduces a ferric Fe(III) complex into an ferrous Fe(II) analogue. According to the literature and our data, FRAP is an excellent radical scavenger, evident in a very high electron affinity EA = 147.0 kcal mol⁻¹. The latter surpasses ionization energies of all investigated systems (Table 3), thereby supporting the thermodynamic feasibility of the electron-transfer process, and confirming its reported non-selective nature for a broad array of antioxidants.³⁴ The situation with other reagents, ABTS⁺ and DPPH[•], is less obvious, since their EA values cluster around 125 kcal mol⁻¹, which leaves only a modest (if any) thermodynamically favorable nature to exchange electrons with studied systems. Yet, we must emphasize that all of our systems have ionization energies that are either lower or close to 125 kcal mol⁻¹, which permits a certain feasibility of the SET mechanism even in these two assays. Moreover, when one relates the amount of the released energy following the attachment on the hydrogen atom onto the reference ABTS⁺

(–36.6 kcal mol⁻¹) and DPPH[•] (–66.7 kcal mol⁻¹) with the fact that all BDE values in 7, 9, 13 and 14 cluster around 80 kcal mol⁻¹, it leads to a conclusion that the HAT mechanism is highly unlikely with ABTS⁺, and only moderately feasible with DPPH[•]. We note in passing that the latter value appears in the right range, given that the value experimentally determined in the more polar aqueous solution is expectedly higher at –78.9 kcal mol⁻¹.³⁵ Therefore, in concluding this section we can emphasize that the SET mechanism likely dominates in determining the antioxidant capacity of our compounds in all three assays, with some relevance of the HAT mechanism within the DPPH[•] assay, in line with similar reports in the literature.³⁶

It is in this context that we will interpret the observed trends in the measured antioxidant capacities. Our designed systems consist of three distinct parts, namely coumarin, benzazole and amidine fragments. According to our analysis, each of them has its own role in governing the antioxidant features of the prepared hybrids. First of all, benzimidazole **m7** appears to be a better building block over benzothiazole **m8**, since it exhibits significantly lower IE (by 8 kcal mol⁻¹) and BDE values (by 16 kcal mol⁻¹), the latter pertaining to its ability to cleave the N–H bond relative to that from the C–H group. This is found in line with the fact that, in general, both benzothiazole hybrids 17 and 18 demonstrate significantly lower antioxidant features over their benzimidazole analogues 13 and 14, and thus are not considered further. On the other hand, amidines seem to worsen the electron-releasing tendency, since their IE values in **m9** and **m10** extend to around 150 kcal mol⁻¹, which stretches even beyond the electron-accepting ability of the FRAP assay, whose EA = 147 kcal mol⁻¹. As an illustration, the IE value is reduced by 8.5 kcal mol⁻¹ on going from IE = 122.2 kcal mol⁻¹ in **m11** to 113.7 kcal mol⁻¹, once the pentacyclic amidine is removed from the structure. Yet, their benefit is evident in their ability to facilitate the HAT mechanism, as the corresponding BDE values in **m9** and **m10** are lowered to 83.3 and 82.5 kcal mol⁻¹, respectively, which bears some significance for their response in the DPPH[•] assay. Interestingly, if one considers unionized amidines, it turns out that in the pentacyclic version, both IE and BDE values are further reduced to 108.0 and 75.0 kcal mol⁻¹.

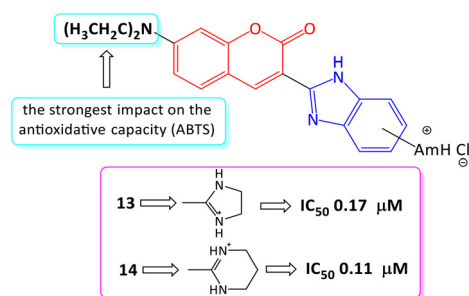


Fig. 3 Structure–activity relationship regarding the antioxidative capacity.

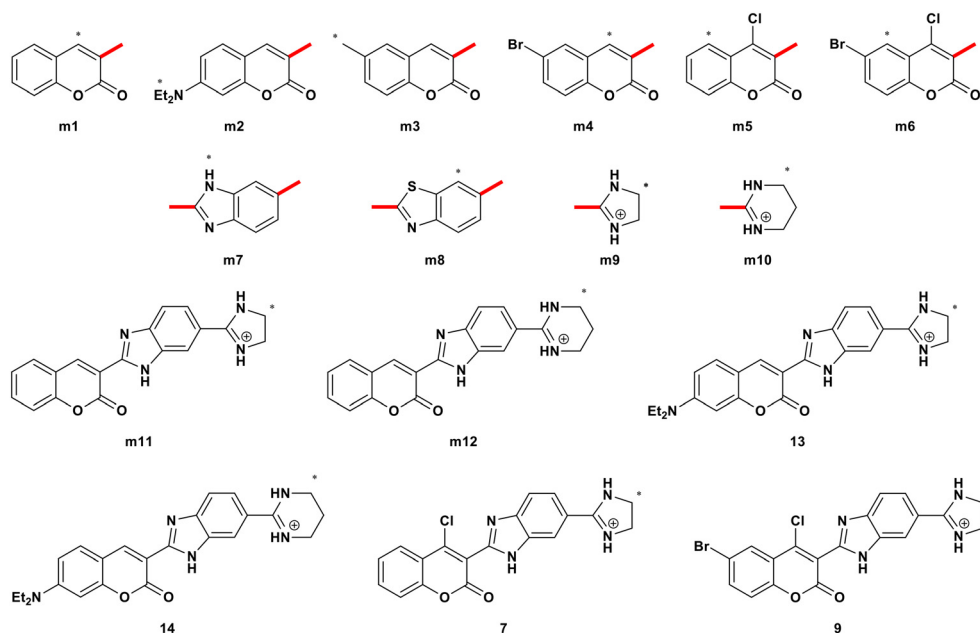


Fig. 4 Chemical structures of systems evaluated computationally. Asterisk (*) denotes thermodynamically most favourable hydrogen atom abstraction site elucidated from all X–H bonds except those from the junction methyl moieties marked in red.

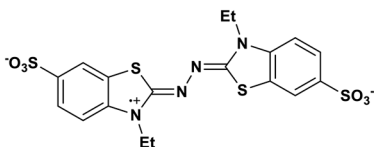
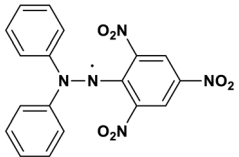
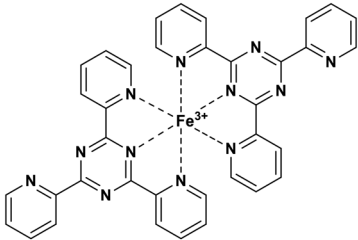
This indicates that all prepared compounds could demonstrate even better antioxidative features under, for example, more basic conditions or less polar environments, which would disfavour amidine protonation, thereby leaving it unionized and promoting radical scavenging properties. Finally, the essential coumarin core does not appear to be particularly suitable for designing potent antioxidants, as **m1** is linked with relatively unfavorable parameters, IE = 124.5 kcal mol⁻¹ and BDE = 102.7 kcal mol⁻¹. Still, due to its electron-conjugated nature, our synthetic strategy allows for a specific tuning of the antioxidant features by placing appropriate substituents at strategic places on the coumarin

Table 3 Ionization energies (IEs), electron affinities (EAs), and X–H bond dissociation energies (BDEs) in ethanol calculated by the (SMD)/B3LYP/6–31+G(d) model (in kcal mol⁻¹). The position of the X–H hydrogen atom abstraction is marked with an asterisk on Fig. 5

System	IE	EA	X–H BDE	Alternative BDE
m1	124.5	76.4	102.7	
m2	100.1	71.0	80.7	70.2 (C–NEt ₂ bond)
m3	121.8	76.3	80.6	87.4 (C–CH ₃ bond)
m4	127.2	79.5	103.2	78.0 (C–Br bond)
m5	128.6	80.9	103.2	75.2 (C–Cl bond)
m6	130.4	83.5	104.2	75.4 (C–Cl bond); 77.9 (C–Br bond)
m7	112.3	44.3	86.6	
m8	120.4	55.9	102.7	
m9	146.8	63.1	83.3	
m10	151.4	56.6	82.5	
m11	122.2	91.9	80.5	
m12	121.9	90.7	80.0	
7	129.1	91.3	78.5	70.5 (C–Cl bond)
9	125.9	97.0	80.2	70.7 (C–Cl bond); 77.8 (C–Br bond)
13	104.9	84.6	80.4	73.4 (C–NEt ₂ bond)
14	106.0	83.6	80.9	73.4 (C–NEt ₂ bond)

unit. This is only moderately observed with halogen atoms, as bromide in **m4** and chloride in **m5**, or their combination in **m6**, do not reveal any notable effect on the calculated parameters; yet only making these compounds even a bit less potent over **m1**. Still, a very interesting feature of all halogen-containing systems is that they can more easily liberate halogen radicals than any hydrogen atom within their structure. The matching BDEs for C–X (X = Cl, Br) bonds in **m4–m6** cluster between 75–78 kcal mol⁻¹, and are significantly lower than those for the most feasible hydrogen atom abstractions, for which BDEs exceed 100 kcal mol⁻¹. However, this is only one part of the picture, as one needs to look at the eventual ability of the reference assays to capture the liberated halogen radicals. Data in Table 4 reveal that neither ABTS^{•+} or DPPH[•] are able to efficiently bind Cl[•] and Br[•] radicals; instead only moderately stable non-bonding complexes are formed, which appears insufficient for the expected assay response. With this in mind, we can eliminate homolytic carbon–halogen bond cleavage as responsible for the observed antioxidant features in halogen-containing derivatives, which unfortunately opposes their favorable effect for the antiviral activity expounded earlier. In contrast, the electron-donating methyl (**m3**) and diethylamine in particular (**m2**), offer significant antioxidative improvements. They both reduce the BDE values to around 80 kcal mol⁻¹, thereby precisely matching the described effect of the amidine unit, while the –NEt₂ group additionally lowers the IE value to 100.1 kcal mol⁻¹ in **m2**. Because of that, the effect of the diethylamine moiety on the coumarin core appears as most efficient in promoting the potency of these compounds for both HAT and SET mechanisms, which is clearly seen in all three assays employed here. We note in passing that, due

Table 4 Ionization energies (IEs), electron affinities (EAs), and N-X bond association energies (BAEs) in ethanol solution for reference systems used in experimental assays as obtained by the (SMD)/B3LYP/6-31+G(d) model (in kcal mol⁻¹)

System	Structure	IE	EA	Bond	BAE	<i>d</i> (N-X)
ABTS ^{•+}		111.2	125.3	N-H N-Cl N-Br N-NEt ₂	-36.6 -16.4 (no bonding) -9.5 (no bonding) +31.6	1.03 Å 3.54 Å 3.36 Å 1.54 Å
DPPH [•]		104.5	124.5	N-H N-Cl N-Br N-NEt ₂	-66.7 -25.3 (no bonding) -19.5 (no bonding) +4.8	1.02 Å 4.85 Å 3.05 Å 1.40 Å
FRAP			147.0			

to the high stability of the N-centered radicals, such as, for example, DPPH[•], the energy required to cleave the C-NEt₂ bond in **m2** is very small, BDE = 70.2 kcal mol⁻¹, yet this turns irrelevant since none of the assays is able to bind the liberated Et₂N[•] radical in a thermodynamically favorable way (Table 4). When structures containing all three building blocks are considered (**m11** and **m12**), thereby more closely resembling the prepared hybrids, it appears that they retain the low electron-donating ability of the coumarin core (IEs ≈ 122 kcal mol⁻¹), and the high hydrogen atom donating tendency of the amidine groups (BDEs ≈ 80 kcal mol⁻¹); the latter making them very potent for the DPPH[•] assay. To further confirm that, let us mention that if **m11** is derived from the amidine moiety, its HAT ability is decreased to BDE = 102.8 kcal mol⁻¹, while its SET capacity is enhanced to IE = 113.7 kcal mol⁻¹. Yet, the full potential of our synthetic strategy becomes evident in compounds dressed up with the -NEt₂ group on the coumarin core, which does not impact

the favorable hydrogen atom releasing ability of a distant five (**13**, BDE = 80.4 kcal mol⁻¹) or six-membered amidine (**14**, BDE = 80.9 kcal mol⁻¹), while significantly improving the feasibility of the electron ejection to around 105 kcal mol⁻¹, being lowest of all examined systems. This feature is evident in all three employed assays, promoting systems **13-14** as starting points for further derivatization towards even more efficient antioxidants.

In vitro antiproliferative activity

All synthesized coumarin-benzimidazole **7-14** and coumarin-benzothiazole **17-18** hybrids were tested for their *in vitro* antiproliferative activity on the following panel of cancer cell lines: LN-229 – glioblastoma, Capan-1 – pancreatic adenocarcinoma, HCT-116 – colorectal carcinoma, NCI-H460 – lung carcinoma, DND-41 – acute lymphoblastic leukemia, HL-60 – acute myeloid leukemia, K-562 – chronic myeloid leukemia and Z-138 – non-Hodgkin lymphoma. Two standard anticancer drugs used in medical treatment, etoposide and nocodazole, were included as reference compounds. The obtained results are presented in Table 5 as IC₅₀ values (50% inhibitory concentration). Based on the obtained results, we can conclude that the majority of the tested hybrids showed low or moderate activity, while four derivatives displayed a pronounced antiproliferative effect on either a selection of cancer cell lines or on the whole panel. The *N,N*-diethylamino coumarin-benzimidazole hybrid bearing a cyclic 2-imidazolinyamidine group **13** was the most promising hybrid yielding IC₅₀ values ranging from 0.3 to 2.1 μM on all the different cancer types. Derivative **11**, a 4-chloro-6-methyl substituted benzimidazole hybrid substituted with a

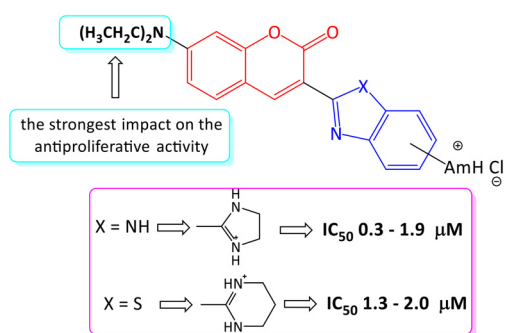
**Fig. 5** Structure-activity relationship regarding the antiproliferative activity.

Table 5 Antiproliferative activity *in vitro* of tested compounds 7–14 and 17–18 against a broad panel of cancer cell types

Cpd	IC ₅₀ /μM							
			Cell line					
	LN-229	Capan-1	HCT-116	NCI-H460	DND-41	HL-60	K-562	Z-138
7	>100	99.3	>100	>100	36.5	55.7	>100	50.5
8	>100	69.2	>100	>100	19.1	53.5	>100	44.5
9	40.9	29.9	50.2	>100	6.2	49.6	9.7	6.4
10	>100	54.2	>100	>100	49.2	>100	59.5	52.1
11	9.1	10.2	34.4	31.7	5.8	21.5	9.9	6.4
12	>100	71.9	>100	>100	16.9	64.2	>100	48.1
13	1.4	1.7	1.9	1.8	0.3	2.1	1.9	1.5
14	97.0	66.7	>100	>100	10.8	63.8	63.5	67.3
17	>100	80.6	>100	>100	10.2	>100	65.5	11.3
18	1.8	2.0	>100	>100	1.3	>100	>100	1.5
Etoposide	0.43	1.45	2.40	3.65	2.80	0.42	1.77	0.85
Nocodazole	0.10	0.13	0.29	0.25	0.47	0.10	0.07	0.31

2-imidazolyl amidine group, also showed marked antiproliferative activity with selectivity towards four cancer cell types (IC₅₀ 5.8–9.9 μM). The benzimidazole derivative **9** substituted with bromine, chlorine and a 2-imidazolyl group displayed a similar pattern with inhibitory concentrations in the range of 6.2 to 9.7 μM against three cancer cell types. When comparing all coumarin-benzimidazole hybrids, we can conclude that the presence of a pentacyclic 2-imidazolyl amidine group (derivatives **7**, **9**, **11** and **13**) significantly enhanced the antiproliferative activity when compared to derivatives bearing a hexacyclic amidine group. Additionally, the introduction of a methyl group instead of bromine at position 6 on the coumarin ring slightly improved the antiproliferative activity. The introduction of a *N,N*-diethylamino group placed at position 7 on the coumarin ring enhanced the antiproliferative activity to a greater extent when compared to a chlorine at position 4 or a bromine and methyl at position 6. Regarding the coumarin-benzothiazole hybrids, the *N,N*-diethyl-amino substituted benzothiazole derivative bearing a hexacyclic amidine group **18** proved most potent with favorable inhibitory concentrations ranging from 1.3 to 2.0 μM against four different cancer cell types.

In general, we might conclude that out of all synthesized coumarin-benzazoles the hybrids substituted with a *N,N*-diethylamino group at position 7 of the coumarin ring (Fig. 5) show the most promising antitumor profile.

Subcellular distribution of compounds **13** and **18**

Since the newly synthesized derivatives **13** and **18** display strong autofluorescent properties, their subcellular distribution can be investigated by fluorescence microscopy. After an incubation period of 3 hours to allow for cellular uptake, imaging was performed on Hep-2 cells treated with 10 μM of compound **13** or **18** (Fig. 6). For both derivatives, marked localization towards the nucleoli and nuclear membrane is noted. Derivative **18** is also distributed throughout the cytoplasm in a pattern strongly resembling

ER localization as shown in Fig. 6. Hep-2 cells were obtained from ATCC (CCL-23).

Toxicity of compounds **13** and **18** to normal cells

The ratio between the concentrations of drug required for efficacy and the concentration that causes toxicity is referred to as the therapeutic window. At the highest dose tested (100 μM), both derivatives **13** and **18** show some toxicity towards normal PBMC from healthy donors (Fig. 7), although more pronounced for compound **13**. While 20 μM of derivative **13** yielded an average of 60% viability, all other tested concentrations for both compounds resulted in viability percentages above 80% relative to the untreated PBMC control. Since both molecules are active at low micromolar concentrations (average IC₅₀ 1.6 μM for **13** and 1.7 μM for **18**), we can conclude that there is good selectivity towards cancer cells (Fig. 7).

Conclusions

In this work, we have used molecular hybridization approach to design and synthesize amidino substituted coumarin-benzazole hybrids. All newly synthesized compounds were substituted at position 3 of the coumarin ring with benzazole nuclei bearing either an unsubstituted or a cyclic amidino group, as well as positions 4, 6 and 7 of the coumarin ring

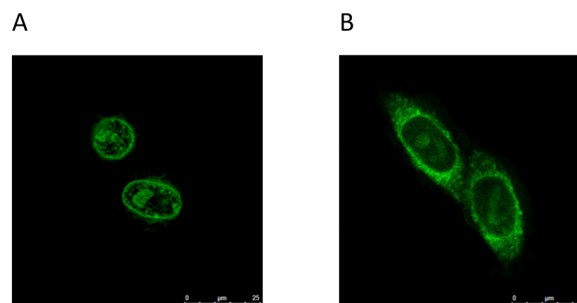


Fig. 6 Fluorescence imaging was performed to detect the subcellular localization of **13** (panel A) and **18** (panel B).

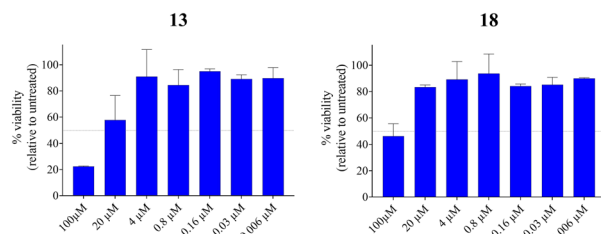


Fig. 7 Cell viability of PBMC treated with **13** and **18** for 72 hours (percentage relative to the untreated control).

with halogen, methyl of *N,N*-diethylamino groups. For the synthesis we have used previously published and well optimized synthetic procedures.

All newly synthesized compounds were evaluated for their biological activities including *in vitro* antiviral, antioxidative and antiproliferative activity. Antiviral activity was evaluated against a diverse panel of viruses and the obtained results revealed that the coumarin-benzimidazole hybrid **10** with a hexacyclic amidine group and bromine and chlorine at positions 6 and 4, respectively, displayed the most promising broad antiviral activity with EC_{50} values ranging from 9.0 to 43.8 μ M. Additionally, the 2-imidazolyl substituted benzimidazole derivative **9** showed selective activity towards influenza A virus subtype H1N1. Benzothiazole hybrids were less active in comparison to benzimidazoles. Regarding the antioxidative activities, the most promising results were observed for two coumarin-benzimidazole derivatives **13** and **14**, whose high radical-scavenging ability is evident in all three assays, while even surpassing that of the standard BHT in the ABTS assay with the IC_{50} values of 0.17 and 0.11 mM, respectively. Computational analysis supported these results and demonstrated that these hybrids benefit from the high C–H hydrogen atom releasing tendency of the cationic amidine unit, and the pronounced ease with which they can liberate an electron, promoted by the electron-donating diethylamine group on the coumarin core. Good antioxidative activity was also observed in DPPH assay with 4-chloro substituted benzimidazole derivative bearing a hexacyclic amidine group **7** being the most active one (IC_{50} 0.07 mM). In FRAP assay, the majority of the tested compounds were significantly less active in comparison to the included reference BHT. Antiproliferative activity was tested on a diverse panel of human cancer cell lines and the obtained results revealed that introducing a *N,N*-diethylamino group at position 7 of the coumarin ring significantly improved the antitumoral activity. The most pronounced activity was noted for the 2-imidazolyl substituted benzimidazole derivative **13** (IC_{50} 0.3–1.9 μ M) and the benzothiazole derivative with a hexacyclic amidine group **18** (IC_{50} 1.3–2.0 μ M). We can thus conclude that the pentacyclic 2-imidazolyl amidine group had the most significant impact on the enhancement of the antiproliferative activity. To summarize, the newly prepared coumarin-benzazole hybrids bearing an amidine group at the benzazole nuclei show considerable biological potential with pronounced antiproliferative, antiviral and antioxidant

activities, thus representing a promising scaffold for further design and optimization of promising therapeutic agents.

Experimental part

Chemistry

General methods. Melting points were recorded on SMP11 Bibby and Büchi 535 apparatus. 1H and ^{13}C NMR spectra were recorded on a Varian Gemini 300 or Varian Gemini 600 spectrophotometer using TMS as an internal standard in DMSO- d_6 . Chemical shifts are reported in ppm (δ) relative to TMS. All compounds were routinely checked by thin layer chromatography (TLC) using precoated Merck silica gel 60F-254 plates and the spots were detected under UV light (254 nm). Elemental analysis for C, H and N were performed on a Perkin-Elmer 2400 elemental analyzer where analyses are indicated only as symbols of elements, analytical results obtained are within 0.4% of the theoretical value.

Synthesis. Synthesis of main precursors necessary for the synthesis of targeted coumarin-benzothiazole and coumarin-benzimidazole hybrids, namely 2-(3,4-diaminophenyl)-4,5-dihydro-1*H*-imidazol-3-ium chloride,³⁵ 2-(3,4-diaminophenyl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride,^{37,38} 2-amino-5-(4,5-dihydro-1*H*-imidazol-3-ium-2-yl)-benzenethiolate hydrate¹⁴ or 2-amino-5-(3,4,5,6-tetrahydropyrimidin-1-ium-2-yl)-benzenethiolate¹⁴ was carried out according to the previously published experimental procedures.

General method for the synthesis of amidino-substituted coumarin-benzimidazole hybrids 7–14. A mixture of equivalent amounts of corresponding aldehyde, 2-(3,4-diaminophenyl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **5** or 2-(3,4-diaminophenyl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **6** in absolute ethanol and *p*-benzoquinone was stirred at reflux for 4 hours. The reaction mixture was cooled to room temperature and resulting product was filtered off and washed with diethyl-ether.

2-(2-(4-chloro-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **7**. The compound **7** was prepared following the general method from 4-chloro-2-oxo-2*H*-chromene-3-carbaldehyde **1** (0.10 g, 0.5 mmol), 2-(3,4-diaminophenyl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **5** (0.10 g, 0.5 mmol) and *p*-benzoquinone (0.05 g, 0.5 mmol) in absolute ethanol (3.5 mL) to obtain 0.09 g (45%) of brown powder. m.p. > 300 °C; 1H NMR (400 MHz, DMSO- d_6) (δ /ppm): 13.68 (bs, 1H, NH_{benz}), 13.63 (bs, 1H, NH_{benz}), 10.18 (s, 2H, $NH_{amidine}$), 8.11 (d, 1H, $J = 1.19$ Hz, H_{arom}), 8.07 (dd, 1H, $J_1 = 8.07, J_2 = 1.59$ Hz, H_{arom}), 7.94 (d, 1H, $J = 8.47$ Hz, H_{arom}), 7.72–7.67 (m, 2H, H_{arom}), 7.40–7.37 (m, 2H, H_{arom}), 2.11–1.92 (m, 4H, CH_2); ^{13}C NMR (151 MHz, DMSO- d_6) (δ /ppm): 176.1, 162.3, 159.5, 153.3, 150.9, 133.6, 133.0, 129.8, 125.2, 124.3, 123.9, 123.7, 120.5, 116.7, 115.6, 113.6, 113.0, 85.5, 17.7 (2C); anal. calcd. for $C_{20}H_{18}Cl_2N_4O_2$: C, 57.57; H, 4.35; Cl, 16.99; N, 13.43; O, 7.67. Found: C, 57.49; H, 4.27; Cl, 16.91; N, 13.39; O, 7.73%.

2-(2-(4-chloro-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **8**. The compound

8 was prepared following the general method from 4-chloro-2-oxo-2*H*-chromene-3-carbaldehyde **1** (0.10 g, 0.5 mmol), 2-(3,4-diaminophenyl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **6** (0.11 g, 0.5 mmol) and *p*-benzoquinone (0.05 g, 0.5 mmol) in absolute ethanol (2 mL) to obtain 0.14 g (64%) of green powder. m.p. > 300 °C; ¹H NMR (400 MHz, DMSO-*d*₆) (δ/ppm): 13.68 (bs, 1H, NH_{benz}), 13.62 (bs, 1H, NH_{benz}), 10.17 (s, 2H, NH_{amidine}), 8.11 (s, 1H, H_{arom}), 8.07 (dd, 1H, *J*₁ = 8.08, *J*₂ = 1.69 Hz, H_{arom}), 7.94 (d, 1H, *J* = 8.47 Hz, H_{arom}), 7.71–7.66 (m, 2H, H_{arom}), 7.40–7.36 (m, 2H, H_{arom}), 3.53 (s, 4H, CH₂), 2.05–1.99 (m, 2H, CH₂); ¹³C NMR (101 MHz, DMSO-*d*₆) (δ/ppm): 176.6, 162.8, 160.0, 153.8, 151.4, 150.2, 134.1, 133.5, 130.3, 125.7, 124.8, 124.3, 121.0, 117.2, 116.1, 114.1, 113.5, 86.0, 18.2 (2C); anal. calcd. for C₂₁H₂₀Cl₂N₄O₂: C, 58.48; H, 4.67; Cl, 16.44; N, 12.99; O, 7.42. Found: C, 58.49; H, 4.63; Cl, 16.49; N, 13.09; O, 7.50%.

2-(2-(6-bromo-4-chloro-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **9**. The compound **9** was prepared following the general method from 6-bromo-4-chloro-2-oxo-2*H*-chromene-3-carbaldehyde **2** (0.10 g, 0.3 mmol), 2-(3,4-diaminophenyl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **5** (0.07 g, 0.3 mmol) and *p*-benzoquinone (0.04 g, 0.3 mmol) in absolute ethanol (3 mL) to obtain 0.11 g (67%) of green powder. m.p. > 300 °C; ¹H NMR (600 MHz, DMSO-*d*₆) (δ/ppm): 13.67 (bs, 1H, H_{benz}), 13.61 (bs, 1H, H_{benz}), 10.15 (s, 2H, H_{amidine}), 8.10 (s, 1H, H_{arom}), 8.09 (d, 1H, *J* = 2.18 Hz, H_{arom}), 7.94 (d, 1H, *J* = 8.45 Hz, H_{arom}), 7.83 (dd, 1H, *J*₁ = 8.67, *J*₂ = 2.49 Hz, H_{arom}), 7.69 (d, 1H, *J* = 8.37 Hz, H_{arom}), 7.38 (d, 1H, *J* = 8.68 Hz, H_{arom}), 2.05–1.97 (m, 4H, H_{arom}); ¹³C NMR (151 MHz, DMSO-*d*₆) (δ/ppm): 174.6, 165.3, 161.8, 152.3, 150.9, 136.0, 134.0, 130.0, 127.2, 124.5, 122.3, 119.3, 117.8, 115.9, 115.6, 113.9, 85.7, 44.4 (2C); anal. calcd. for C₂₀H₁₇BrCl₂N₄O₂: C, 48.41; H, 3.45; Br, 16.10; Cl, 14.29; N, 11.29; O, 6.45. Found: C, 48.46; H, 3.40; Br, 16.05; Cl, 14.22; N, 11.23; O, 6.40%.

2-(2-(6-bromo-4-chloro-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **10**. The compound **10** was prepared following the general method from 6-bromo-4-chloro-2-oxo-2*H*-chromene-3-carbaldehyde **2** (0.10 g, 0.3 mmol), 2-(3,4-diaminophenyl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **6** (0.08 g, 0.3 mmol) and *p*-benzoquinone (0.04 g, 0.3 mmol) in absolute ethanol (3 mL) to obtain 0.09 g (54%) of green powder. m.p. > 300 °C; ¹H NMR (600 MHz, DMSO-*d*₆) (δ/ppm): 13.67 (bs, 1H, H_{benz}), 13.61 (bs, 1H, H_{benz}), 10.16 (s, 2H, H_{amidine}), 8.10 (s, 1H, H_{arom}), 8.09 (d, 1H, *J* = 2.48 Hz, H_{arom}), 7.94 (d, 1H, *J* = 8.49 Hz, H_{arom}), 7.83 (dd, 1H, *J*₁ = 8.69, *J*₂ = 2.48 Hz, H_{arom}), 7.69 (dd, 1H, *J*₁ = 8.48, *J*₂ = 1.45 Hz, H_{arom}), 7.38 (d, 1H, *J* = 8.68 Hz, H_{arom}), 3.52 (s, 4H, CH₂), 2.04–2.00 (m, 2H, CH₂); ¹³C NMR (151 MHz, DMSO-*d*₆) (δ/ppm): 162.8, 160.0, 153.8, 151.4, 150.2, 134.1, 133.5, 130.3, 125.7, 124.7, 124.3, 124.2, 121.0, 117.2, 116.1, 114.1, 113.5, 86.0, 18.2; anal. calcd. for C₂₁H₁₉BrCl₂N₄O₂: C, 49.44; H, 3.75; Br, 15.66; Cl, 13.90; N, 10.98; O, 6.27. Found: C, 49.46; H, 3.71; Br, 15.72; Cl, 13.81; N, 11.04; O, 6.21%.

2-(2-(4-chloro-6-methyl-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **11**. The

compound **11** was prepared following the general method from 4-chloro-6-methyl-2-oxo-2*H*-chromene-3-carbaldehyde **3** (0.10 g, 0.4 mmol), 2-(3,4-diaminophenyl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **5** (0.09 g, 0.4 mmol) and *p*-benzoquinone (0.05 g, 0.4 mmol) in absolute ethanol (2.5 mL) to obtain 0.12 g (69%) of green powder. m.p. > 300 °C; ¹H NMR (400 MHz, DMSO-*d*₆) (δ/ppm): 13.70 (bs, 1H, H_{benz}), 13.65 (bs, 1H, H_{benz}), 10.78 (s, 2H, H_{amidine}), 8.29 (s, 1H, H_{arom}), 7.96 (s, 2H, H_{arom}), 7.82 (d, 1H, *J* = 1.47 Hz, H_{arom}), 7.54 (d, 1H, *J* = 1.48 Hz, H_{arom}), 7.27 (d, 1H, *J* = 8.27 Hz, H_{arom}), 4.06–4.03 (m, 4H, CH₂), 2.40 (s, 3H, CH₃); ¹³C NMR (101 MHz, DMSO-*d*₆) (δ/ppm): 152.9, 151.9, 148.0, 147.5, 145.3, 134.9, 133.0, 132.4, 128.2, 126.9, 119.8, 119.5, 112.0 (2C), 92.8, 50.3 (2C), 39.9, 28.8, 20.2; anal. calcd. for C₂₁H₂₀Cl₂N₄O₂: C, 58.48; H, 4.67; Cl, 16.44; N, 12.99; O, 7.42. Found: C, 58.43; H, 4.71; Cl, 16.40; N, 12.94; O, 7.46%.

2-(2-(4-chloro-6-methyl-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **12**.

The compound **12** was prepared following the general method from 4-chloro-6-methyl-2-oxo-2*H*-chromene-3-carbaldehyde **3** (0.10 g, 0.4 mmol), 2-(3,4-diaminophenyl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **6** (0.11 g, 0.4 mmol) and *p*-benzoquinone (0.05 g, 0.4 mmol) in absolute ethanol (2.5 mL) to obtain 0.10 g (57%) of green powder. m.p. > 300 °C; ¹H NMR (400 MHz, DMSO-*d*₆) (δ/ppm): 13.66 (bs, 1H, H_{benz}), 13.60 (bs, 1H, H_{benz}), 10.18 (s, 2H, H_{amidine}), 8.11 (s, 1H, H_{arom}), 7.94 (d, 1H, *J* = 8.47 Hz, H_{arom}), 7.84 (s, 1H, H_{arom}), 7.70 (dd, 1H, *J*₁ = 8.49, *J*₂ = 1.67 Hz, H_{arom}), 7.49 (dd, 1H, *J*₁ = 8.37, *J*₂ = 2.00 Hz, H_{arom}), 7.27 (d, 1H, *J* = 8.28 Hz, H_{arom}), 3.53 (s, 4H, CH₂), 2.41 (s, 3H, CH₃), 2.04–1.99 (m, 2H, CH₂); ¹³C NMR (101 MHz, DMSO-*d*₆) (δ/ppm): 176.7, 166.9, 162.9, 160.0, 151.9, 151.5, 150.2, 134.9, 133.5, 130.3, 125.3, 124.7, 124.2, 120.7, 117.0, 116.10, 114.0, 113.5, 85.9, 29.5, 20.9, 18.5; anal. calcd. for C₂₂H₂₂Cl₂N₄O₂: C, 59.33; H, 4.98; Cl, 15.92; N, 12.58; O, 7.19. Found: C, 59.36; H, 4.92; Cl, 15.87; N, 12.53; O, 7.09%.

2-(2-(7-(diethylamino)-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **13**.

The compound **13** was prepared following the general method from 7-(*N,N*-diethylamino)-2-oxo-2*H*-chromene-3-carbaldehyde **4** (0.10 g, 0.4 mmol), 2-(3,4-diaminophenyl)-4,5-dihydro-1*H*-imidazol-3-ium chloride **5** (0.09 g, 0.4 mmol) and *p*-benzoquinone (0.04 g, 0.4 mmol) in absolute ethanol (2.5 mL) to obtain 0.16 g (90%) of orange powder. m.p. > 300 °C; ¹H NMR (400 MHz, DMSO-*d*₆) (δ/ppm): 12.90 (bs, 1H, H_{benz}), 12.77 (bs, 1H, H_{benz}), 10.64 (s, 2H, H_{amidine}), 8.64 (s, 1H, H_{arom}), 8.40–8.19 (m, 1H, H_{arom}), 8.88–7.78 (m, 2H, H_{arom}), 7.75 (d, 1H, *J* = 8.97 Hz, H_{arom}), 6.84 (dd, 1H, *J*₁ = 9.09, *J*₂ = 2.27 Hz, H_{arom}), 6.68 (d, 1H, *J* = 2.06 Hz, H_{arom}), 4.02 (s, 4H, CH₂), 3.53–3.49 (m, 4H, CH₂), 1.17 (d, 6H, *J* = 6.96 Hz, CH₃); ¹³C NMR (101 MHz, DMSO-*d*₆) (δ/ppm): 166.0, 160.6, 157.3, 152.6, 150.2 (2C), 131.7, 122.6, 119.7, 116.1 (2C), 110.6, 108.6 (2C), 107.1 (2C), 96.7, 44.9 (2C), 44.7 (2C), 12.8 (2C); anal. calcd. for C₂₄H₂₈ClN₅O₂: C, 63.50; H, 6.22; Cl, 7.81; N, 15.43; O, 7.05. Found: C, 63.45; H, 6.16; Cl, 7.74; N, 15.39; O, 7.12%.

2-(2-(7-(diethylamino)-2-oxo-2*H*-chromen-3-yl)-1*H*-benzo[d]imidazol-5-yl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **14**.

The compound **14** was prepared following the general method from 7-(*N,N*-diethylamino)-2-oxo-2*H*-chromene-3-carbaldehyde **4** (0.10 g, 0.4 mmol), 2-(3,4-diaminophenyl)-3,4,5,6-tetrahydropyrimidin-1-ium chloride **6** (0.09 g, 0.4 mmol) and *p*-benzoquinone (0.04 g, 0.4 mmol) in absolute ethanol (2.5 mL) to obtain 0.04 g (21%) of orange powder. m.p. > 300 °C; ¹H NMR (400 MHz, DMSO-*d*₆) (δ/ppm): 12.80 (bs, 1H, H_{benz}), 12.70 (bs, 1H, H_{benz}), 9.98 (s, 2H, H_{amidine}), 8.10–8.00 (m, 2H, H_{arom}), 7.79 (d, 2H, *J* = 8.45 Hz, H_{arom}), 7.76 (d, *J* = 8.96 Hz, H_{arom}), 7.55 (d, 1H, *J* = 7.08 Hz, H_{arom}), 6.85 (dd, 1H, *J*₁ = 9.00, *J*₂ = 2.10 Hz, H_{arom}), 6.69 (d, *J* = 1.79 Hz, H_{arom}), 3.54–3.60 (m, 8H, CH₂), 2.04–1.99 (m, 2H, CH₂), 1.18 (t, 6H, *J* = 6.91 Hz, CH₃); ¹³C NMR (101 MHz, DMSO-*d*₆) (δ/ppm): 160.6, 157.3, 152.5, 150.2, 131.7 (2C), 122.4 (2C), 121.6, 116.1, 110.6 (2C), 108.6 (2C), 107.3 (2C), 96.7, 44.8 (2C), 40.6, 40.4, 18.41, 12.8 (2C); anal. calcd. for C₂₅H₃₀ClN₅O₂: C, 64.16; H, 6.46; Cl, 7.57; N, 14.96; O, 6.84. Found: C, 64.22; H, 6.38; Cl, 7.63; N, 14.88; O, 6.77%.

General method for the synthesis of amidino-substituted benzothiazole derivatives 17–18. A mixture of 7-(*N,N*-diethylamino)-2-oxo-2*H*-chromene-3-carbaldehyde **4** (0.3 mmol) and 2-amino-5-(4,5-dihydro-1*H*-imidazol-3-ium-2-yl)-benzenethiolate hydrate (0.3 mmol) **15** or 2-amino-5-(3,4,5,6-tetrahydropyrimidin-1-ium-2-yl)-benzenethiolate **16** (0.3 mmol) in glacial acetic acid (5 ml) was stirred at reflux under nitrogen for 2 h, followed by the addition of diethyl-ether and the crude product was filtered off. Crude product was suspended in water and concentrated hydrochloric acid (0.02 mL, 0.2 mmol) was added to the solution. The reaction mixture was stirred at room temperature for 3 h, followed by addition of acetone and the resulting precipitate was filtered off and washed with diethyl-ether.

2-[2-(7-(*N,N*-Diethylamino)-2-oxo-2*H*-chromen-3-yl)benzothiazol-6-yl]-4,5-dihydro-1*H*-imidazol-3-ium chloride **17**. The compound **17** was prepared following the general method from 2-amino-5-(4,5-dihydro-1*H*-imidazol-3-ium-2-yl)-benzenethiolate hydrate **15** (0.06 g, 0.3 mmol) and 7-(*N,N*-diethylamino)-2-oxo-2*H*-chromene-3-carbaldehyde **4** (0.07 g, 0.3 mmol) to obtain of 0.03 g (47%) red powder. m.p. > 300 °C; ¹H NMR (300 MHz, DMSO-*d*₆) (δ/ppm): 10.77 (s, 2H, NH_{amidine}), 9.00 (s, 1H, H_{arom}), 8.80 (s, 1H, H_{arom}), 8.12 (s, 2H, H_{arom}), 7.78 (d, 1H, *J* = 8.97 Hz, H_{arom}), 6.86 (d, 1H, *J* = 8.88 Hz, H_{arom}), 6.66 (s, 1H, H_{arom}), 4.04 (s, 4H, CH₂), 3.52 (q, 4H, *J* = 6.92 Hz, CH₂), 1.17 (t, 6H, *J* = 6.86 Hz, CH₃); ¹³C NMR (DMSO-*d*₆, 151 MHz): δ/ppm = 166.1, 164.6, 160.3, 157.1, 155.6, 152.8, 143.5, 135.6, 132.1, 126.2, 123.3, 122.0, 117.6, 110.6, 109.6, 108.1, 96.2, 44.5 (2C), 44.5 (2C), 12.35 (2C); Anal. Calcd. for C₂₄H₂₇ClN₄O₂S: C, 61.20; H, 5.78; Cl, 7.53; N, 11.90; O, 6.79; S, 6.81. Found: C, 61.28; H, 5.86; Cl, 7.58; N, 11.97; O, 6.83; S, 6.86%.

2-[2-(7-(*N,N*-Diethylamino)-2-oxo-2*H*-chromen-3-yl)benzothiazol-6-yl]-3,4,5,6-tetra-hydropyrimidin-1-ium chloride **18**. The compound **18** was prepared following the general method from 2-amino-5-(3,4,5,6-tetrahydropyrimidin-1-ium-2-yl)-benzenethiolate **16** (0.06 g, 0.3 mmol) and 7-(*N,N*-diethylamino)-2-oxo-2*H*-chromene-3-carbaldehyde **4** (0.07 g, 0.3

mmol) to obtain of 0.04 g (54%) red powder. m.p. > 300 °C; ¹H NMR (300 MHz, DMSO-*d*₆) (δ/ppm): 10.16 (s, 2H, NH_{amidine}), 9.06 (s, 1H, H_{arom}), 8.57 (s, 1H, H_{arom}), 8.14 (d, 1H, *J* = 8.58 Hz, H_{arom}), 7.85 (dd, 1H, *J*₁ = 8.77 Hz, *J*₂ = 1.69 Hz, H_{arom}), 7.81 (d, 1H, *J* = 9.08 Hz, H_{arom}), 6.88 (dd, 1H, *J*₁ = 8.97 Hz, *J*₂ = 2.00 Hz, H_{arom}), 6.70 (s, 1H, H_{arom}), 4.05 (s, 8H, CH₂), 2.05–1.96 (m, 2H, CH₂), 1.17 (t, 6H, *J* = 6.87 Hz, CH₃); ¹³C NMR (DMSO-*d*₆, 75 MHz): δ/ppm = 165.7, 160.9, 159.6, 157.5, 155.3, 153.2, 143.9, 135.9, 132.5, 126.1, 124.6, 122.8, 122.3, 111.1, 110.2, 108.6, 96.7, 45.0 (2C), 18.2 (3C), 12.8 (2C); anal. calcd. for C₂₅H₂₉ClN₄O₂S: C, 61.91; H, 6.03; Cl, 7.31; N, 11.55; O, 6.60; S, 6.61. Found: C, 61.96; H, 6.08; Cl, 7.26; N, 11.48; O, 6.57; S, 6.66%.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

We greatly appreciate the financial support of the Croatian Science Foundation under the projects 4379 entitled *Exploring the antioxidative potential of benzazole scaffold in the design of novel antitumor agents*. R.V. wishes to thank the Zagreb University Computing Centre (SRCE) for granting computational resources on the ISABELLA cluster.

Notes and references

- L. F. Tietze, H. P. Bell and S. Chandrasekhar, Natural product hybrids as new leads for drug discovery, *Angew. Chem., Int. Ed.*, 2003, **42**, 3996–4028, DOI: [10.1002/anie.200200553](https://doi.org/10.1002/anie.200200553).
- Shaveta, S. Mishra and P. Singh, *Eur. J. Med. Chem.*, 2016, **124**, 500–536, DOI: [10.1016/j.ejmech.2016.08.039](https://doi.org/10.1016/j.ejmech.2016.08.039).
- R. B. Silverman, *The Organic Chemistry of Drug Design and Drug Action*, Elsevier Academic Press, 2nd edn, 2004.
- (a) P. C. Sharma, A. Sinhmar, A. Sharma, H. Rajak and D. P. Pathak, *J. Enzyme Inhib. Med. Chem.*, 2013, **28**, 240–266, DOI: [10.3109/14756366.2012.720572](https://doi.org/10.3109/14756366.2012.720572); (b) A. Kumar, L. P. Datta, S. Samanta, H. Arora and T. Govindaraju, *Isr. J. Chem.*, 2021, **61**, 222–230, DOI: [10.1002/ijch.202000098](https://doi.org/10.1002/ijch.202000098).
- (a) N. Shrivastava, M. J. Naim, M. J. Alam, F. Nawaz, S. Ahmed and O. Alam, *Arch. Pharm. Chem. Life Sci.*, 2017, **350**, e1700040, DOI: [10.1002/ardp.201700040](https://doi.org/10.1002/ardp.201700040); (b) K. Rajasekhar, N. Narayanaswamy, N. A. Murugan, G. Kuang, H. Ågren and T. Govindaraju, *Sci. Rep.*, 2016, **6**, 23668, DOI: [10.1038/srep23668](https://doi.org/10.1038/srep23668).
- (a) S. Tariq, P. Kamboj and M. Amir, *Arch. Pharm. Chem. Life Sci.*, 2019, **352**, e1800170, DOI: [10.1002/ardp.201800170](https://doi.org/10.1002/ardp.201800170); (b) E. A. A. El-Meguid, A. M. Naglah, G. O. Moustafa, H. M. Awad and A. M. El Kerdawy, *Bioorg. Med. Chem. Lett.*, 2022, **58**, 128529, DOI: [10.1016/j.bmcl.2022.128529](https://doi.org/10.1016/j.bmcl.2022.128529).
- J. Akhtar, A. A. Khan, Z. Ali, R. Haider and M. Shahar Yar, *Eur. J. Med. Chem.*, 2017, **125**, 143–189, DOI: [10.1016/j.ejmech.2016.09.023](https://doi.org/10.1016/j.ejmech.2016.09.023).
- G. Yadav and S. Ganguly, *Eur. J. Med. Chem.*, 2015, **97**, 419–443, DOI: [10.1016/j.ejmech.2014.11.053](https://doi.org/10.1016/j.ejmech.2014.11.053).

- 9 (a) A. Irfan, F. Batool, S. A. Z. Naqvi, A. Islam, S. M. Osman, A. Nocentini, S. A. Alissa and C. T. Supuran, *J. Enzyme Inhib. Med. Chem.*, 2019, **35**, 265–279, DOI: [10.1080/14756366.2019.1698036](https://doi.org/10.1080/14756366.2019.1698036); (b) N. Narayanaswamy, S. Narra, R. R. Nair, D. K. Saini, P. Kondaihab and T. Govindaraju, *Chem. Sci.*, 2016, **7**, 2832, DOI: [10.1039/c5sc03488d](https://doi.org/10.1039/c5sc03488d).
- 10 G. Yadav and S. Ganguly, *Eur. J. Med. Chem.*, 2015, **97**, 419–443, DOI: [10.1016/j.ejmech.2014.11.053](https://doi.org/10.1016/j.ejmech.2014.11.053).
- 11 R. S. Keri, A. Hiremathad, S. Budagumpi and B. M. Nagaraja, *Chem. Biol. Drug Des.*, 2015, **86**, 19–65, DOI: [10.1111/cbdd.12462](https://doi.org/10.1111/cbdd.12462).
- 12 M. Hranjec, M. Kralj, I. Piantanida, M. Sedić, L. Šuman, K. Pavelić and G. Karminski-Zamola, *J. Med. Chem.*, 2007, **50**, 5696–5711, DOI: [10.1021/jm070876h](https://doi.org/10.1021/jm070876h).
- 13 L. Racane, V. Tralić-Kulenović, S. Kraljević Pavelić, I. Ratkaj, P. Peixoto, R. Nhili, S. Depauw, M. P. Hildebrand, M. H. David-Cordonnier, K. Pavelić and G. Karminski-Zamola, *J. Med. Chem.*, 2010, **53**, 2418–2432, DOI: [10.1021/jm901441b](https://doi.org/10.1021/jm901441b).
- 14 L. Racané, M. Cindrić, I. Zlatar, T. Kezele, A. Milić, K. Brajša and M. Hranjec, *J. Enzyme Inhib. Med. Chem.*, 2021, **36**, 163–174, DOI: [10.1080/14756366.2020.1850711](https://doi.org/10.1080/14756366.2020.1850711).
- 15 M. Cindrić, S. Jambon, A. Harej, S. Depauw, M. David-Cordonnier, S. Kraljević Pavelić, G. Karminski-Zamola and M. Hranjec, *Eur. J. Med. Chem.*, 2017, **136**, 468–479, DOI: [10.1016/j.ejmech.2017.05.014](https://doi.org/10.1016/j.ejmech.2017.05.014).
- 16 A. Carneiro, M. J. Matos, E. Uriarte and L. Santana, *Molecules*, 2021, **26**, 501, DOI: [10.3390/molecules26020501](https://doi.org/10.3390/molecules26020501).
- 17 (a) S. Emami and S. Dadashpour, *Eur. J. Med. Chem.*, 2015, **102**, 611–630, DOI: [10.1016/j.ejmech.2015.08.033](https://doi.org/10.1016/j.ejmech.2015.08.033); (b) K. Rajasekhar, C. J. Achar and T. Govindaraju, *Org. Biomol. Chem.*, 2017, **15**, 1584, DOI: [10.1039/c6ob02760a](https://doi.org/10.1039/c6ob02760a).
- 18 (a) A. Thakur, R. Singla and V. Jaitak, *Eur. J. Med. Chem.*, 2015, **101**, 476–495, DOI: [10.1016/j.ejmech.2015.07.010](https://doi.org/10.1016/j.ejmech.2015.07.010); (b) T. Abdizadeh, M. R. Kalani, K. Abnous, Z. Tayarani-Najaran, B. Z. Khashyarmansh, R. Abdizadeh, R. Ghodsi and F. Hadizadeh, *Eur. J. Med. Chem.*, 2017, **132**, 42–62, DOI: [10.1016/j.ejmech.2017.03.024](https://doi.org/10.1016/j.ejmech.2017.03.024).
- 19 (a) S. Sandhu, Y. Bansal, O. Silakari and G. Bansal, *Bioorg. Med. Chem.*, 2014, **22**, 3806–3814, DOI: [10.1016/j.bmc.2014.05.032](https://doi.org/10.1016/j.bmc.2014.05.032); (b) Y. V. Suseela, P. Satha, N. A. Murugan and T. Govindaraju, *Theranostics*, 2020, **10**, 10394–10414, DOI: [10.7150/thno.48675](https://doi.org/10.7150/thno.48675).
- 20 X. F. Song, J. Fan, L. Liu, X. F. Liu and F. Gao, *Arch. Pharm.*, 2020, **353**, e2000025, DOI: [10.1002/ardp.202000025](https://doi.org/10.1002/ardp.202000025).
- 21 J. S. Prusty and A. Kumar, *Mol. Diversity*, 2020, **24**, 1367–1383, DOI: [10.1007/s11030-019-09992-x](https://doi.org/10.1007/s11030-019-09992-x).
- 22 L. Zhang and Z. Xu, *Eur. J. Med. Chem.*, 2019, **181**, 111587, DOI: [10.1016/j.ejmech.2019.111587](https://doi.org/10.1016/j.ejmech.2019.111587).
- 23 A. Dorababu, *Eur. J. Med. Chem.*, 2021, **2**, 100006, DOI: [10.1016/j.ejmcr.2021.100006](https://doi.org/10.1016/j.ejmcr.2021.100006).
- 24 I. Boček, M. Hranjec and R. Vianello, *J. Mol. Liq.*, 2022, **355**, 118982, DOI: [10.1016/j.molliq.2022.118982](https://doi.org/10.1016/j.molliq.2022.118982).
- 25 H. Li, L. Cai and Z. Chen, *Advances in Chemical Sensors*, ed. W. Wang, InTech, 2011, pp. 121–150.
- 26 K. Paul, S. Bindal and V. Luxami, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 3667–3672, DOI: [10.1016/j.bmcl.2012.12.071](https://doi.org/10.1016/j.bmcl.2012.12.071).
- 27 R. K. Arora, N. Kaur, Y. Bansal and G. Bansal, *Acta Pharm. Sin. B*, 2014, **4**, 368–375, DOI: [10.1016/j.apsb.2014.07.001](https://doi.org/10.1016/j.apsb.2014.07.001).
- 28 S. Barman, S. K. Mukhopadhyay, M. Gangopadhyay, S. Biswas, S. Dey and N. D. Pradeep Singh, *J. Mater. Chem. B*, 2015, **3**, 3490, DOI: [10.1039/c4tb02081b](https://doi.org/10.1039/c4tb02081b).
- 29 M. T. Gabr, M. M. H. Ibrahim, A. Tripathi and C. Prabhakar, *Chemosensors*, 2019, **7**, 34, DOI: [10.3390/chemosensors7030034](https://doi.org/10.3390/chemosensors7030034).
- 30 S. G. Kini, S. Choudhary and M. Mubeen, *J. Comput. Methods Mol. Des.*, 2012, **2**, 51–60.
- 31 S. C. Tsay, S. Y. Lin, W. C. Huang, M. H. Hsu, K. C. Hwang, C. C. Lin, J. C. Horng, I. C. Chen, J. R. Hwu, F. K. Shieh, P. Leyssen and J. Neyts, *Molecules*, 2016, **21**, 228, DOI: [10.3390/molecules21020228](https://doi.org/10.3390/molecules21020228).
- 32 N. Perin, M. Cindrić, P. Vervaeke, S. Liekens, T. Mašek, K. Starčević and M. Hranjec, *Med. Chem.*, 2021, **17**, 13–20, DOI: [10.2174/1573406416666191218101427](https://doi.org/10.2174/1573406416666191218101427).
- 33 S. Tshepelevitsh, A. Kütt, M. Lõkov, I. Kaljurand, J. Saame, A. Heering, P. Plieger, R. Vianello and I. Leito, *Eur. J. Org. Chem.*, 2019, **2019**, 6735–6748, DOI: [10.1002/ejoc.201900956](https://doi.org/10.1002/ejoc.201900956).
- 34 N. B. Sadeer, D. Montesano, S. Albrizio, G. Zengin and M. F. Mahomoodally, *Antioxidants*, 2020, **9**, 709, DOI: [10.3390/antiox9080709](https://doi.org/10.3390/antiox9080709).
- 35 M. C. Foti and C. Daquino, *Chem. Commun.*, 2006, 3252–3254.
- 36 J. Chen, J. Yang, L. Ma, J. Li, N. Shahzad and C. K. Kim, *Sci. Rep.*, 2020, **10**, 2611.
- 37 K. Starčević, M. Kralj, K. Ester, I. Sabol, M. Grce, K. Pavelić and G. Karminski-Zamola, *Bioorg. Med. Chem.*, 2007, **15**, 4419–4426, DOI: [10.1016/j.bmc.2007.04.032](https://doi.org/10.1016/j.bmc.2007.04.032).
- 38 L. Racané, V. Tralić-Kulenović, Z. Mihalić, G. Pavlović and G. Karminski-Zamola, *Tetrahedron*, 2008, **64**, 11594–11602, DOI: [10.1016/j.tet.2008.10.026](https://doi.org/10.1016/j.tet.2008.10.026).