RESEARCH ARTICLE

Impact of essential metals on insulin sensitivity and fetuin in obesity‑related type 2 diabetes pathogenesis

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Abstract

Purpose Essential metals may be crucial in obesity and type 2 diabetes (T2DM); diabesity pathogenesis and consequences. This study aimed to determine the metal levels in obese and non-obese patients with and without T2DM and their relationships with fetuin-A(Fet-A) levels, insulin sensitivity, and insulin resistance.

Methods A total of 314 participants were enrolled, with 160 newly diagnosed T2DM patients and 154 non-T2DM subjects categorized into diabetic obese $(n=57)$, diabetic non-obese $(n=103)$, non-diabetic obese $(n=48)$, and non-diabetic nonobese (*n*=106) subgroups. Fet-A, insulin sensitivity (QUCKI)/resistance (HOMA-IR), fasting glucose, and body mass index (BMI) were assessed. The essential metals were measured using inductively coupled plasma mass spectroscopy (ICP-MS). **Results** Fet-A levels were 3-fold higher (1391.4 \pm 839.8 ng/ml) in T2DM patients than in non-T2DM (2165.6 \pm 651.9 vs. 424.3±219.1 ng/ml, *p*<0.0001). Fet-A levels were 2.3-fold higher in the diabetic obese group than in the diabetic non-obese group $(p<0.0001)$. Fet-A levels were 2.0-fold higher in the diabetic non-obese group than in the non-diabetic obese group $(p<0.0001)$. Fet-A levels were positively correlated with insulin resistance (HOMA-IR) ($r=0.34$, $p<0.0001$) and negatively correlated with insulin sensitivity (QUIKI) (r=−0.41*, p*<0.0001).

Cu, Se, Zn, and Fe levels were significantly lower in diabetic patients than in non-diabetic patients $(p<0.05)$. Se and Zn were significantly correlated with Fet-A (r = −0.41, *p* = 0.049 and r = −0.42, *p* = 0.001, respectively). Se and Zn were also correlated with insulin resistance (HOMA-IR) (r=−0.45, *p*=0.049 and r=−0.36, *p*=0.012, respectively) and insulin sensitivity (QUIKI) ($r=0.49$, $p=0.042$ and $r=0.30$, $p=0.003$, respectively). Similarly, Fe was negatively correlated with insulin levels (r=−0.33, *p*=0.04) and insulin sensitivity (r=−0.34, *p*=0.30). However, Mn was signifcantly correlated with Fet-A ($r=0.37$, $p=0.001$) and insulin resistance/sensitivity ($r=0.24$, $p=0.026$ and $r=-0.24$, $p=0.041$) respectively in the diabetic obese group. Mg was an independent predictor of diabesity.

Conclusions Mg play a signifcant role in obesity-related T2DM pathogenesis and complications via Fet-A, insulin sensitivity, and resistance modifcations.

Keywords Fetuin-A · Insulin resistance · Obesity · Obesity-related type 2 diabetes mellitus · Non-obese diabetes

Introduction

Obesity and Type-2 diabetes mellitus (T2DM) are lifestyle disorders that are afected by ethnicity and geographical diversity. Hyperglycemia, insulin resistance/sensitivity, and relative insulin insufficiency characterize T2DM $[1, 2]$ $[1, 2]$ $[1, 2]$ $[1, 2]$.

 \boxtimes Vandana Tiwari drvandana2166@yahoo.com Obesity is a major epidemic in modern society and a link between obesity and diabetes (diabesity) is emerging. Data from the registry revealed that diabetes medication was less efective when body mass index was increased. Furthermore, altered adipokine secretion has been suggested to represent a basic link between obesity and T2DM [\[3](#page-8-2)].

Fet-A, an endogenous inhibitor of insulin receptor tyrosine kinase in the liver and muscles, efectively prevents obesity [\[4\]](#page-8-3). In humans, elevated Fet-A levels are an independent risk factor for T2DM [\[5\]](#page-8-4) and are strongly related to insulin resistance and fat storage [\[6](#page-8-5)]. Body mass index (BMI) and waist-to-hip ratio (WHR) were signifcantly associated with Fet-A levels and increased in insulin-resistant obesity

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compared to insulin-sensitive obesity [\[7](#page-8-6)]. Moreover, relevant mediators such as fasting glucose, HbA1c, and C-peptide levels increase and low insulin levels are associated with the pathogenesis of T2DM in individuals with higher Fet-A levels [\[8](#page-8-7)]. Various essential metals have been implicated in the etiology of T2DM and their potential roles have been discussed for decades [[9\]](#page-8-8). Essential metals may afect the onset and pathophysiology of diabetes in various ways. For example, metal imbalances may disrupt normal glucose and insulin metabolism, leading to insulin resistance/sensitivity and the development of diabetic complications [[10,](#page-8-9) [11\]](#page-8-10). Previous studies on the possible functions of essential metals in diabetes have primarily focused on zinc and iron.

Zn is an integral component of insulin, and is involved in its production, storage, and secretion. Many enzymes involved in glucose metabolism require Zn as a cofactor [[12](#page-8-11), [13\]](#page-8-12). Similarly, Fe has been postulated to affect the development of diabetes through numerous pathways, including the generation of insulin insufficiency and resistance as well as the cause of hepatic dysfunction [[14](#page-8-13)]. Most studies have concentrated on the relationship between a single essential element and T2DM, and have been confned to those who already have the disease. It is unclear whether these changes in essential element status occur before disease onset or as a result of disease. As a result, we intended to explore essential metal levels at the time of T2DM screening to probe variations in essential element levels in the early stages of the disease while controlling for possible confounders that were frequently overlooked in previous studies [[15–](#page-8-14)[17\]](#page-8-15). However, epidemiological data on the risk of metabolic dysfunction related to elementary metal levels in the North Indian population are scarce. Therefore, further studies are required to evaluate the reported associations and examine recent insights. Based on the above background context, the present study aimed to measure the levels of essential metals, including copper (Cu), magnesium (Mg), manganese (Mn), iron (Fe), selenium (Se), and zinc (Zn) , as well as their relationships with Fet-A levels, insulin sensitivity, and insulin resistance among T2DM patients with and without obesity.

Material and methods

Selection of subjects and sample collection

This case-control study was conducted at the Department of Biochemistry, Dr. Ram Manohar Lohia Institute of Medical Sciences, Lucknow, India. Patients newly diagnosed with T2DM with or without obesity based on the American Diabetes Association (ADA) and World Health Organization (WHO) criteria (2019) [\[18](#page-8-16)] and non-diabetic (control) subjects were recruited for the study.

Subjects with diabetes combined with acute complications such as ketoacidosis and hyperosmolar coma, acute or chronic infammation, severe hepatic and renal dysfunction, malignant tumors, and other endocrine and/or metabolic disorders were excluded from the study.

Anthropometric measurements

Age, sex, body weight, height, waist circumference, and hip circumference (WC) were recorded. Body mass index (BMI) was calculated as the weight (kg) divided by the square of height (m). Obesity was defned as BMI≥30 kg/m2 based on the characteristics of the Asian population (ADA criteria 2019). WC was measured at the midpoint between the lower borders of the rib cage and iliac crest. WHR was calculated as waist circumference divided by hip circumference (WHO criteria 1997). Individuals with BMI \geq 30 kg/m² were considered obese, while those with BMI $<$ 25 kg/m² and WHR more than 0.90 for men and 0.85 for females were considered obese [[19\]](#page-8-17).

Experimental group

A total of 314 subjects were enrolled in the study, of whom 160 were naïve-diagnosed T2DM patients and 154 were non-T2DM subjects. According to BMI and fasting blood sugar (FBS) levels, the study participants were subdivided into

Group-1: Diabetic Obese (DO; *n*=57) **Group-2:** Diabetic Non-Obese (DNO; *n*=103) **Group-3:** Nondiabetic Obese (NDO; *n*=48) **Group-4:** Non-Diabetic Non-Obese (NDNO; n=106)

Sampling

Venous blood samples from all study subjects were collected in the morning (after 12 h of fasting) in a fuoride, ethylenediaminetetraacetic acid (EDTA), and plain vacutainer. Serum samples were collected and stored at -80 °C until use. Plasma samples from fuoride vials were analyzed for fasting plasma glucose (FPG) levels, and serum samples from plain vials were analyzed for lipid and metal profles.

Estimation of biochemical parameters

Fasting blood sugar (FBS) and lipid profles were analyzed using commercial reagents on an automated analyzer (Beckman Coulter), and HbA1c was analyzed from samples collected in an EDTA vial on a BIORAD (Variant Turbo). Serum insulin concentrations were measured using electrochemiluminescence (ECL).

Insulin resistance (HOMA‑IR) and insulin sensitivity (QUICKI) calculations

Insulin resistance was calculated by modifed homeostasis model assessment of insulin resistance [[20\]](#page-8-18):

HOMA – IR = fasting insulin (U/mL) \times FPG (mmol/L)/22.5

The Quantitative Insulin Sensitivity Check Index (QUICKI) was calculated as follows [[21](#page-8-19)]:

 $QUICKI = 1 / log (fasting insulin) + log (fasting glucose)$

Estimation of fetuin‑A levels

Fet-A levels were measured using a quantitative sandwich enzyme-linked immunosorbent assay (ELISA) method (Quantikine, R&D Systems Inc., Minneapolis, MN, USA) according to the manufacturer's protocol. Briefy, serum samples were diluted to 1:10 in dilution bufer. The test was performed within 5 h. Absorbance was measured at a wavelength of 450 nm. The average detectable Fet-A level was 303–671 μg/ml. The intra-assay variability was 7.2%, and the inter-assay variability was 6.4%.

Estimation of essential metals

Essential metals (Cu, Mg, Mn, Fe, Se, Cr, and Zn) were measured in serum samples using inductively coupled plasma mass spectrometry (ICP-MS) (Perkin Elmer). Before analysis, 100 μL of serum samples were mixed with 100 μL of internal standard (IS), then made up to 5 mL by adding diluent; v/v 15 mL methanol +0.005% v/v Triton X solution +10 mL nitric acid, making it up to 1000 mL by adding double-distilled water. The metals were then analyzed by comparison with the standard using a linearity curve.

Statistical analysis

Quantitative variables are presented as Mean \pm SD. Student's t test was used to compare continuous data, while categorical data were analyzed using the χ 2 test to assess the difference in biochemical parameters between the two groups. One-way analysis of variance (ANOVA) was used to compare anthropometric and biochemical parameters between the diabetic and obese groups. Pearson's correlation coefficients were calculated to determine the correlation between fetuin-A levels and FBG, HbA1c, lipid profle, insulin resistance (HOMA-IR), insulin sensitivity (QUICKI), metals, and C-peptide levels. Univariate and multivariate linear regression analyses were used to evaluate the independent serum essential metal predictors in the diferent diabetes groups. All analyses were conducted using SPSS 16.0 (Chicago, Inc. USA). Statistical significance was set at $p < 0.05$.

Ethics

This study was approved by the Ethics Committee (Ref. code: IEC-24/18) of the Dr. Ram Manohar Lohia Institute of Medical Sciences, Lucknow, India. Written informed consent was obtained from all participants prior to enrollment. A structured pro forma was used to collect anthropometric and personal information.

Result

Anthropometric, biochemicals and elements levels in T2DM and non‑T2DM

There were no signifcant diferences in age and sex between patients (T2DM) and controls (non-T2DM), indicating adequate matching $(p=0.22 \text{ and } p=0.86, \text{ respectively})$. BMI and WHR were signifcantly higher in T2DM patients than in non-T2DM patients ($P = 0.006$ and $P = 0.011$, respectively). The T2DM group showed signifcantly higher levels of diabetes-associated markers such as FBS, HbA1c, insulin, insulin resistance (HOMA-IR), insulin sensitivity (QUICKI), and C-peptide $(p < 0.05)$. The lipid profile (TC, TG, LDL-C, and VLDL-C) levels were signifcantly higher in the diabetic group than in the control group. Fet-A levels were 3-fold higher $(1391.4 \pm 839.8 \text{ ng/ml})$ in T2DM patients than in non-T2DM $(444.9 \pm 203.3 \text{ ng/ml})$. Cu, Se, Zn, and Fe levels were signifcantly lower in diabetic patients than in non-diabetic patients $(p < 0.05)$ (Table [1](#page-3-0)).

Fet‑A and essential elements levels associated with diabesity

Among the subjects, 57 (18.16%) were obese-diabetic, 103 (32.80%) were non-obese-diabetic, while in the non-diabetic group, 48 (15.29%) were non-diabetic obese, and 106 (33.75%) were non-diabetic non-obese (Table [2\)](#page-4-0).

Fet-A levels were 2.3-fold higher in the diabetic obese group than in the non-diabetic obese group $(p < 0.0001)$. Furthermore, Fet-A levels were 2.0-fold higher in the diabetic non-obese group than in the non-diabetic obese group (*P*<0.0001). In contrast, the obese-non-diabetic and control groups (without diabetes or obesity) showed no signifcant difference $(p=0.065)$ in Fet-A levels (Table [2](#page-4-0)).

Zn was significantly decreased in the diabetic obese group compared to the non-diabetic non-obese group $(69.1 \pm 18.2 \text{ vs. } 83.4 \pm 21.8, p < 0.0001)$. The nondiabetic obese group Zn was significantly decreased

Table 1 Clinical characteristics, biochemical, and Essential metals of the Study Population

Abbreviations: *T2DM* Type-2 Diabetes mellitus, *BMI* body mass index, *WHR* Waist-to-Hip Ratio, *HOMA-IR* Homeostasis model assessment-estimated insulin resistance, *QUICKI* Quantitative insulin sensitivity check index, *FBS* Fasting Blood Sugar, *TC* Total Cholesterol, *TG* Triglyceride, *HDL* High-density Lipoprotein, *LDL* Low-density Lipoprotein, *VLDL* Very low-density Lipoprotein. Student *T*-test was used to compare groups. $*$ *p* value <0.05 is considered statistically significant

compared to the non-diabetic non-obese (control) group (78.4 \pm 20.4, vs. 83.4 \pm 21.8, $p = 0.015$). Mg was significantly increased in the diabetic obese group as compared to the non-diabetic obese group $(2.4 \pm 0.3 \text{ vs.})$ 2.2 ± 0.3 , $p < 0.0001$).

In contrast, Cr was significantly increased in diabetic obese patients than in non-obese and non-diabetic patients (20.26 ± 13.45 vs.11.36 ± 6.64, *p* < 0.0001). Cu and Se were also significantly reduced in the diabetic obese group as compared to the non-diabetic non-obese group (Cu; 99.24 ± 3.39 vs. 119.7 ± 40.3, *p* = 0.0002; Se;133.7 \pm 36.7 vs. 150.9 \pm 49.7, $p = 0.022$).

Essential elements serum level correlated with obesity and diabetic‑associated markers

Fet-A levels were negatively correlated with Cu, Se, Zn, and Mg ($r = -0.31$, $p = 0.031$; $r = -0.41$, $p = 0.049$; $r = -0.22$, $p = 0.001$; $r = -0.42$, $p = 0.046$, respectively), while HbA1c levels were negatively correlated with Cu and Zn (r=−0.30, *p*=0.038; r=−0.32, *p*=0.001, respectively). A significant positive correlation was found between QUICKI and Zn levels $(r = 0.30, p = 0.003)$. $Cu²⁺$ also showed a negative correlation with insulin and C-peptide levels $(r = -0.21, p = 0.04$ for both) (Table [3](#page-4-1)).

Table 2 Comparison of biochemical variables and essential metals within T2DM Obese, T2DM Non-Obese, Non-T2DM Obese, Non-Diabetic Obese, and Non-Diabetic Non-Obese

Variables	Sub-groups					p Value			
	$(A)DO (n=57)$	(B)DNO $(n=103)$ (C)NDO $(n=48)$		(D)NDNO $(n=106)$ A vs. C		A vs D	A vs. B	C vs. D	
BMI (kg/m ²)	33.6 ± 3.5	24.7 ± 3.9	33.5 ± 2.8	24.3 ± 3.7	0.873	$<0.0001*$	$<0.0001*$	$< 0.0001*$	
WHR (cm)	0.96 ± 0.06	0.80 ± 0.03	0.95 ± 0.05	0.84 ± 0.02	0.361	$<0.0001*$	$< 0.0001*$	$< 0.0001*$	
TC (mg/dl)	181.5 ± 43.6	180.9 ± 46.7	177.0 ± 39.3	156.5 ± 39.9	0.582	$0.0003*$	0.936	$0.003*$	
$TG \, (mg/dl)$	181.6 ± 97.6	170.9 ± 89.8	158.4 ± 74.9	135.4 ± 64.7	0.183	$0.0004*$	0.489	0.053	
$HDL-C$ (mg/dl)	45.4 ± 15.0	43.2 ± 14.9	42.5 ± 9.6	42.3 ± 10.1	0.250	0.118	0.373	0.908	
$LDL-C$ (mg/dl)	110.4 ± 36.3	117.1 ± 42.5	110.0 ± 32.4	96.1 ± 34.4	0.953	$0.014*$	0.316	$0.019*$	
$VLDL-C$ (mg/dl)	36.2 ± 19.6	34.3 ± 17.9	32.0 ± 14.8	27.1 ± 12.9	0.225	$0.0005*$	0.535	$0.038*$	
FBS (mg/dl)	150.3 ± 43.8	174.7 ± 74.1	101.0 ± 8.1	98.9 ± 9.2	$< 0.0001*$	$<0.0001*$	$0.024*$	0.175	
HbA1c $(\%)$	6.9 ± 1.5	8.1 ± 2.6	5.3 ± 0.5	5.2 ± 0.4	$< 0.0001*$	$<0.0001*$	$0.001*$	0.159	
Insulin $(\mu I U/ml)$	19.1 ± 13.7	15.6 ± 12.7	17.1 ± 12.5	9.6 ± 6.3	0.446	$< 0.0001*$	0.109	$< 0.0001*$	
C -Peptide (ng/ml)	3.0 ± 1.3	2.5 ± 1.4	2.6 ± 1.2	2.3 ± 1.5	0.083	$0.003*$	$0.028*$	0.224	
Fetuin-A (ng/mL)	2165.6 ± 651.9	941 ± 564.3	489.4 ± 157.1	424.3 ± 219.1	$< 0.0001*$	$<0.0001*$	$< 0.0001*$	0.065	
Mg^{2+} (mg/dL)	2.4 ± 0.3	2.4 ± 0.5	2.7 ± 0.7	2.2 ± 0.3	$0.004*$	$0.0001*$	1.00	$< 0.0001*$	
$Cu^{2+} (\mu g/dL)$	99.24 ± 3.39	106.1 ± 47.5	103.6 ± 23.2	119.7 ± 40.3	0.163	$0.0002*$	0.278	$0.010*$	
$Se^{2+}(\mu g/L)$	133.7 ± 36.7	137.81 ± 33.1	136.9 ± 40.9	150.9 ± 49.7	0.673	$0.022*$	0.470	0.089	
Zn^{2+} (µg/dL)	69.1 ± 18.2	75.73 ± 21.1	78.4 ± 20.4	83.4 ± 21.8	$0.015*$	$<0.0001*$	$0.047*$	0.180	
$Cr^{2+} (\mu g/L)$	20.26 ± 13.45	17.50 ± 12.35	11.36 ± 6.64	34.29 ± 10.82	$0.001*$	$<0.0001*$	0.191	$< 0.001*$	
Mn^{2+} (µg/L)	2.55 ± 1.92	2.61 ± 2.44	2.99 ± 2.10	2.24 ± 2.01	0.265	0.341	0.192	$0.036*$	
Fe^{2+} (µg/dL)	133.77 ± 89.58	140.93 ± 63.51	155.42 ± 110.36	146.38 ± 94.11	0.269	0.408	0.206	0.602	

Abbreviations: *DO* Diabetic Obese, *DNO* Diabetic Non-Obese, *DNO* Diabetic Non-Obese, *NDNO* Non-Diabetic Non-Obese, *BMI* body mass index, *WHR* Waist-to-Hip Ratio, *HOMA-IR* Homeostasis model assessment-estimated insulin resistance, *QUICKI* Quantitative insulin sensitivity check index, *FBS* Fasting Blood Sugar, *TC* Total Cholesterol, *TG* Triglyceride, *HDL* High-density Lipoprotein, *LDL* Low-density Lipoprotein, *VLDL* Very low-density Lipoprotein. Student t-test was used to compare groups. The data has been represented in mean \pm SD. * *p* value <0.05 is considered statistically signifcant

Table 3 Correlation of diabesity markers with essential metals and fetuin-A

Variables	Fetuin-A (ng/ mL)	Insulin $(\mu I U)$ ml)	C-peptide (ng/dL)	HOMA-IR QUICKI			$HbA1c$ (%) FBS (mg/dL)	BMI (kg/m^2) WHR (cm)	
Cu^{2+} (µg/dL)	$r = -0.31$	$r = -0.21$	$r = -0.21$	$r = -0.11$	$r = 0.12$	$r = -0.30$	$r = -0.05$	$r = -0.20$	$r = -0.10$
	$p = 0.031*$	$p = 0.040*$	$p = 0.045*$	$p = 0.127$	$p = 0.136$	$p = 0.038*$	$p = 0.478$	$p = 0.047*$	$p = 0.160$
$Se^{2+} (\mu g/L)$	$r = -0.41$	$r = -0.12$	$r = -0.42$	$r = -0.35$	$r = 0.49$	$r = -0.11$	$r = -0.80$	$r = -0.45$	$r = -0.11$
	$p = 0.049*$	$p = 0.113$	$p = 0.513$		$p = 0.049^*$ $p = 0.042^*$	$p = 0.021$	$p = 0.203$	$p = 0.042*$	$p = 0.044$
Zn^{2+} (µg/dL)	$r = -0.42$	$r = -0.14$	$r = -0.13$	$r = -0.36$ $r = 0.30$		$r = -0.32$	$r = -0.13$	$r = -0.16$	$r = -0.17$
	$p = 0.001*$	$p = 0.033$	$p = 0.047$	$p = 0.012^*$	$p = 0.003*$	$p = 0.001*$	$p = 0.038$	$p = 0.011$	$p = 0.043$
$Cr^{2+} (\mu g/L)$	$r = 0.10$	$r = 0.03$	$r = -0.02$	$r = 0.04$	$r = -0.05$	$r = 0.05$	$r = -0.01$	$r = -0.03$	$r = -0.02$
	$p = 0.425$	$p = 0.653$	$p = 0.734$	$p = 0.475$	$p = 0.447$	$p = 0.393$	$p = 0.879$	$p = 0.685$	$p = 0.753$
Mn^{2+} (µg/L)	$r = 0.06$	$r = 0.01$	$r = -0.01$	$r = 0.01$	$r = -0.01$	$r = 0.01$	$r = -0.11$	$r = 0.16$	$r = 0.11$
	$p = 0.314$	$p = 0.943$	$p = 0.864$	$p = 0.813$	$p = 0.942$	$p = 0.924$	$p = 0.074$	$p = 0.040$	$p = 0.074$
Fe^{2+} (µg/dL)	$r = -0.09$	$r = -0.33$	$r = -0.11$	$r = 0.10$	$r = -0.34$	$r = -0.03$	$r = -0.12$	$r = -0.03$	$r = -0.01$
	$p = 0.129$	$p = 0.040*$	$p = 0.081$	$p = 0.106$	$p = 0.030*$	$p = 0.589$	$p = 0.073$	$p = 0.605$	$p = 0.821$
Mg^{2+} (mg/dL) $r = -0.42$		$r = -0.15$	$r = -0.02$	$r = -0.06$	$r = -0.06$	$r = -0.12$	$r = -0.12$	$r = 0.16$	$r = 0.11$
	$p = 0.046*$	$p = 0.820$	$p = 0.820$	$p = 0.371$	$p = 0.373$	$p = 0.560$	$p = 0.042*$	$p = 0.010$	$p = 0.074$
Fetuin-A $(ng/ 1)$		0.23	0.27	0.33	-0.40	0.40	0.38	0.33	0.19
mL)		${<}0.0001*$	${<}0.0001*$	${<}0.0001*$	${<}0.0001*$	${<}0.0001*$	${<}0.0001*$	${<}0.0001*$	${<}0.0001*$

Abbreviations: *r* Pearson correlation, *BMI* body mass index, *WHR* Waist-to-Hip Ratio, *FBS* Fasting Blood Sugar, *HbA1c* Glycosylated Hemoglobin, *HOMA-IR* Homeostasis model assessment-estimated insulin resistance, *QUICKI* Quantitative insulin sensitivity check index, *r* Correlation Coefficient. Pearson correlation was used to analyze correlation. Boldfaced *p value <0.05 is considered statistically significant

Fet‑A serum levels correlated with essential elements in study subgroups

A signifcant positive correlation between Fet-A, HOMA-IR, and Cu in the diabetic obese group $(r=0.30, p=0.021;$ $r = 0.38$, $p = 0.010$) and a negative correlation between QUICKI and Cu $(r=-0.33, p=0.026)$ was observed in the diabetic obese group. Fet-A and HOMA-IR were positively correlated with Mg ($r=0.37$; $p=0.001$; $r=0.25$, $p=0.026$) and a weak negative correlation was observed between QUICKI and Mn($r = -0.24$, $p = 0.041$) in the diabetic obese group. However, in the non-diabetic group, Fet-A was positively correlated with Mg $(r=0.461, p=0.004)$. In contrast, no correlation was found between Fet-A and the other essential metals (Table [4\)](#page-5-0).

A signifcant positive correlation between Fet-A and BMI ($r=0.34$, $p < 0.0001$) and a weak positive correlation between Fet-A and WHR $(r=0.2, p<0.0001)$ was observed (Fig. [1A](#page-6-0) and B). Fet-A was signifcantly negatively correlated with insulin resistance (HOMA-IR) $(r = 0.34)$, $p < 0.0001$) and positively correlated with insulin sensitivity (QUCKI) (r=−0.41, *p*<0.001) (Fig. [1C](#page-6-0) and D).

Analysis using univariate and multivariate linear regression with Fet‑A as the dependent variable

In univariate linear regression analysis, Cu showed a positive association with Fet-A (β = 0.266, *p* < 0.0001) in the diabetic obese group. Mg was negatively associated with diabetes $(\beta = -262, p < 0.0001)$. Similarly, Mn was negatively associated with Fet-A in both diabetic obese groups ($\beta = -0.248$, p <0.0001) and the non-diabetic obese group (β =−0.461, $p < 0.0001$) and was positively associated with diabetic nonobese ($β = 0.375$, $p < 0.0001$). Zn was negatively associated with Fet-A ($\beta = -0.299$, $p < 0.0001$) in the non-diabetic obese group.

In the Multivariate linear regression analysis, only Mn showed a positive association with Fet-A in the diabetic obese ($\beta = 0.507$, $p < 0.0001$) group. In contrast, a negative association was observed with Fet-A levels

Abbreviations: r Pearson correlation coefficient. Pearson correlation was analyzed for correlation. **p* value <0.05 is considered statistically signifcant

Table 4 Correlation of fetuin-A with essential metals in study subgroups

Fig. 1 Pearson Correlation between **A**. Fetuin-A and BMI, **B**. Fetuin-A and WHR, **C**. Between Fetuin-A and HOMA-IR, **D**. Between Fetuin-A and QUICKI in Diabetic Obese groups

in the non-diabetic group (β = -0.521 , *p* < 0.0001). The other essential metals were not associated with Fet-A (Table [5\)](#page-7-0).

Discussion

India's culture, language and cuisine are diverse. Indian cuisine combines the tastes of subcontinents. It emphasizes sugar and fat as dietary items, and an integral Indian way of life. A strong association was observed between caloric intake and obesity. In India, obesity is rising rapidly due to increased energy intake resulting from greater purchasing power and the availability of high-fat, energy-dense foods, as well as a decrease in energy expenditure due to urbanization and mechanization [[22,](#page-8-20) [23\]](#page-8-21). The incidence of T2DM has reached epidemic proportions in India (65 million), paralleling the increase in overweight and obesity (International Diabetes Federation, 2014) [\[18\]](#page-8-16). In the present study, we examined the relationship between essential metals (Cu, Mg, Mn, Fe, Se, Cr, and Zn) and insulin resistance (HOMA-IR, sensitivity (QUIKI], and Fet-A) among obese T2DM patients, non-obese T2DM patients and obese non T2DM.

Studies have demonstrated that Fet-A is directly involved in the concomitant development of insulin resistance and obesity [\[24](#page-8-22), [25](#page-8-23)]. A recent meta-analysis of cohort studies showed that T2DM patients exhibit higher Fet-A levels than non-diabetic individuals [\[26,](#page-8-24) [27\]](#page-8-25). The results of the present study revealed that T2DM patients had signifcantly higher serum levels of Fet-A than non-obese patients with T2DM (controls). Fet-A levels were associated with diabetic markers (insulin, HbA1c, HOMA-IR, QUIKI, and BMI). Furthermore, Fet-A was signifcantly positively correlated with BMI, WHR, fasting blood sugar, HbA1c, insulin, C-peptide, HOMA-IR, and QUICKI, consistent with previous studies [\[5](#page-8-4), [28](#page-9-0)]. Ismail et al. reported that Fet-A levels were elevated in obese adults and showed a signifcant correlation with obesity, consistent with our fndings [\[29\]](#page-9-1).

Magnesium (Mg) has a role in glucose metabolism and enhances insulin activity and secretion [[30\]](#page-9-2). Insulin resistance and Mg reduction trigger the loss of the insulin resistance cycle and depletion of intracellular Mg, restricting its function in cellular processes $[31]$. Therefore, this deficiency might result in inadequate glucose regulation. A study demonstrated that patients with diabetes responded positively to orally administered Mn [\[32\]](#page-9-4). Parallel to the overhead lines, our data demonstrated that Mn levels moderately correlated with Fet-A levels, insulin resistance, and sensitivity in the diabetic obese group; however, obesity acts as a modulator.

In our study, Fe levels were decreased in patients with diabetes and had a signifcant but weak correlation with insulin sensitivity. Previous studies have shown increased insulin sensitivity and secretion with regular blood donation or iron chelation treatments [\[33](#page-9-5), [34](#page-9-6)]. However, the specifc mechanisms underlying iron-induced diabetes remain unclear. Similarly, Se was signifcantly lower in diabetic

Table 5 Univariate and multivariate linear regression model as fetuin-A dependent variables

Abbreviations: *DO* Diabetic Obese, *DNO* Diabetic Non-Obese, *DNO* Diabetic Non-Obese. Univariate and multivariate regression analysis was done to predict the independent marker. **p* value <0.05 is considered statistically signifcant

obese patients than in non-diabetic obese patients and was correlated with insulin resistance and sensitivity, respectively. Parrell's fndings also demonstrated insulin-mimetic efects on the expression of glucose-6-phosphate dehydrogenase (G6PD) in diabetic rats [[35\]](#page-9-7), which is noteworthy given that G6PD activity is considerably lower in diabetic rats than in controls [[36\]](#page-9-8).

Mg was signifcantly decreased in the diabetic group; in contrast, its levels were not infuenced by obesity, which is consistent with previous studies that revealed that cellular and extracellular magnesium depletion is a hallmark of T2DM [[37–](#page-9-9)[39\]](#page-9-10) and that insulin increases the movement of magnesium from the extracellular to the intracellular compartments of a cell, thus boosting the magnesium level available for energy production via magnesium-dependent adenosine triphosphate (ATP). However, in the present study, Mg was not correlated with insulin resistance; however, linear regression analysis revealed that Mg is responsible for obesity via Fet-A. This study demonstrated that Zn was decreased in the diabetic obese group; however, Zn was negatively associated with obesity. This fnding demonstrates that zinc defciency may alter the progression of T2DM. There is evidence of increased insulin secretion and, consequently, increased zinc levels in the early stages of the disease [[40\]](#page-9-11). The pancreas can generate more insulin, but cannot synthesize more zinc, resulting in zinc depletion.

Similarly, copper insufficiency has been associated with decreased glucose tolerance and increased lipid peroxidation owing to increased HbA1c levels [[41](#page-9-12), [42](#page-9-13)]. This suggests that Cu decreased in the diabetic group, was correlated with Fet-A and insulin resistance/sensitivity, and was positively associated with diabesity. However, the cause of this increase and its repercussions have not been fully explained, nor is it understood whether aberrations in copper metabolism result from the disease or whether they are crucial for its progression. Linear regression analysis showed that Mn was an independent predictive marker and was positively associated with diabesity.

However, these fndings have some limitations. First, it had a cross-sectional design and could not allow causal inferences. Second, because the number of subjects in this study was relatively small and eating habits and physical exercise were not recorded, the fndings were probably infuenced by other causative and genetic factors. Finally, as this study was conducted at a single center, and the study cohort was confned to one ethnic group, the data may not represent the whole population.

Conclusion

The present study suggests an association between Fet-A, diabetic markers and obesity, which may indicate its role in the pathogenesis of obesity-related T2DM. Cu, Mg, and Mn play signifcant roles in obesity-related T2DM, whereas Zn is only involved in obesity pathogenesis and complications via Fet-A, insulin sensitivity, and insulin resistance. Mg is an independent predictor for diabesity.

Authors' contribution Anumesh Pathak- Manuscript drafting and writing, data analysis, tables and graphs preparation.

Vandana Tiwari: conceptualization, funding acquisition, execution of the experiment, validation of data, and draft editing.

Manish Raj Kulshrestha- Manuscript preparation and editing.

Shivani Singh- Sample collection & data collection, help in tables preparation.

Shefali Singh- Sample collection & enrolled patients. Vikram Singh- Clinical history & patient details.

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