Pulmonary Surfactant Therapy

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Surfactant replacement therapy is now an integral part of the care of neonates since several clinical trials of natural surfactant extracts and synthetic preparations have shown efficacy in the treatment of infants with hyaline membrane disease. In these studies, early treatment with exogenous surfactant substantially reduced mortality and the incidence of air leak, although it did not appear to reduce the incidence of other complications, in particular bronchopulmonary dysplasia. Early reports of exogenous surfactant therapy in patients with the adult respiratory distress syndrome, although promising, remain limited in number. More research is needed to improve on current modes of therapy and to investigate the possible role of surfactant in other lung diseases of both newborns and adults.

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The possible role of surface forces in the lungs was first considered in 1929 by the Swiss scientist Kurt von Neergaard.¹ Two and a half decades later, Pattle² and Clements³ independently demonstrated the existence of surface-active material in the alveoli of animal lungs. Soon after, in 1959, Avery and Mead established the relationship between this surface-active material, lung surfactant, and hyaline membrane disease of human neonates.⁴ Since then, a great deal of effort has been devoted to studying the biologic properties of lung surfactant and its potential as a human therapy. The first report of the beneficial use of surfactant therapy in neonates with the respiratory distress syndrome (RDS) was published in 1980.5 Several ensuing trials demonstrated the efficacy of exogenous surfactant in reducing the mortality and morbidity attributable to RDS. These results led the United States Food and Drug Administration (FDA) to approve the use of surfactants in neonates in 1990.

In this review, we summarize some biologic aspects of pulmonary surfactant, outline conclusions drawn from clinical trials using surfactant preparations in neonates, and discuss some of the questions that remain unanswered.*

Surfactant Composition

Lung surfactant is a complex lipoprotein assembled and secreted into the alveolar spaces by alveolar epithelial type II cells. Its composition, being fairly constant

*See also the editorial by R. J. Mason, MD, "Surfactant Replacement Therapy—Room for Improvement," on pages 74-76 of this issue.

among mammalian species,⁶ consists of about 90% lipids and 10% proteins. The main lipid fraction is the saturated lecithin dipalmitoyl phosphatidylcholine (DPPC). Negatively charged phospholipids are present in high proportion (10% to 15%), as is free cholesterol (about 10%). A variety of other phospholipids and neutral lipids make up the remainder of the lipids. Three lipid-associated apolipoproteins have been isolated from lung surfactant and designated SP-A, SP-B, and SP-C.7 Two of these, SP-B and SP-C, are small hydrophobic proteins that co-isolate with lipids. SP-A, on the other hand, more easily dissociates from lipids and can be solubilized in water.⁸ A fourth protein, SP-D, has recently been purified from lung lavage and does not appear to directly interact with lipids.9 SP-D bears striking structural homology to SP-A, and along with mannose-binding protein and serum conglutinin, belongs to the newly described "collectin" family.10 This complex lipoprotein has unique functional properties.

Biologic Properties of Lung Surfactant

The primary function of lung surfactant is to lower surface tension at the air-water interface of the lung alveoli, thereby stabilizing lung volume at low transpulmonary pressure. Adsorption of DPPC by itself to the liquid surface is poor,¹¹ but it is greatly enhanced by surfactant apoproteins.¹²

Attention has recently been directed to the immune properties of surfactant. Apoproteins SP-A and SP-D appear to have specific immune properties. Both proteins bind to bacteria^{13,14}; SP-A enhances the

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ABBREVIATIONS USED IN TEXT

CPHT = colfosceril palmitate, hexadecanol, and tyloxapol DPPC = dipalmitoyl phosphatidylcholine FDA = United States Food and Drug Administration RDS = respiratory distress syndrome

phagocytosis of bacteria¹³ and viruses¹⁵ and promotes the chemotaxis of phagocytic cells.¹⁶

Other properties of surfactant may include inhibiting pulmonary edema formation¹⁷ and enhancing fluid dispersal¹⁸ and ciliary transport in small airways.¹⁹

Surfactant Metabolism

The synthesis of lung surfactant appears to be restricted to alveolar epithelial type II cells, although nonciliated bronchiolar cells have been shown in several species to make surfactant apoproteins.²⁰ Once assembled, the components of surfactant are stored in secretory organelles, lamellar bodies, before their secretion into the alveolar liquid subphase where they undergo structural transformations that lead to the formation of the surface layer (Figure 1). Labeling studies show that surfactant is in a state of continuous flux and that type II cells are capable of regulated endocytosis and recycling of the various surfactant components.²¹

Surfactant Replacement Therapy

Before it was approved by the FDA for human use, exogenous surfactant underwent extensive testing in a great number of well-designed, carefully controlled, and properly randomized clinical studies that have set a standard of scientific excellence and productive collaboration in the neonatal community.

We will briefly describe the preparations that were tested (Table 1). Surfactant-TA (Surfacten, Tokoyo Tanabe, Tokyo, Japan) and beractant (Survanta, Abbott Laboratories, Chicago, Illinois) are made from minced lungs and modified by the addition of phospholipids and fatty acids. Infasurf (Forrest Laboratories, New York, NY) and calf lung surfactant extract are made from calf lung lavage extracts, and SF-RI 1 (Alveofact, Boehringer, Ingelheim, Germany) is from cow lung lavage extracts. Curosurf (Chiesi Pharmaceuticals, Parma, Italy) is derived from minced pig lung extracts. Amniotic fluid surfactant is the only human preparation

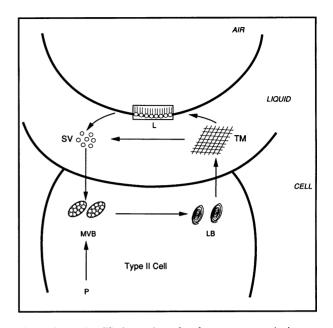


Figure 1.—A simplified overview of surfactant turnover is shown: Type II cells synthesize and assemble precursors (P) of surfactant into multivesicular bodies (MVB) and secrete lamellar body (LB) contents into the alveolar liquid lining where they undergo structural transformations: tubular myelin (TM) feeds the surface layer (L) and small vesicles (SV) are recycled into type II cell multivesicular bodies.

characterized and tested. The two synthetic preparations that were the most extensively tested are the artificial lung-expanding compound (ALEC, Britannia Pharmaceuticals, United Kingdom) and colfosceril palmitate, hexadecanol, and tyloxapol (CPHT; Exosurf Neonatal, Burroughs Wellcome Company, Research Triangle Park, North Carolina), but others were or are being developed.

Analyzing in detail each of the trials is beyond the scope of this review, and our intent is to summarize the conclusions that can be drawn from published results.

Clinical Trials of Exogenous Surfactant in Neonatal Respiratory Distress Syndrome

Trials that followed the report by Fujiwara and colleagues⁵ help answer the following questions:

Surfactant Type	Material	Proteins	
Bovine	Modified minced lung extracts (surfactant TA [Surfacten])	SP-B, SP-C	
Bovine	Modified minced lung extracts (beractant [Survanta])	SP-B, SP-C	
Bovine	Calf lung lavage extracts (CLSE, Infasurf)	SP-B, SP-C	
Bovine	Cow lung lavage extracts (SF-RI 1 [Alveofact])	SP-B, SP-C	
Porcine	Minced lung extracts (Curosurf)	SP-B, SP-C	
Human	Human amniotic fluid extract	SP-A, SP-B, SP-C	
Synthetic	DPPC-PG (artificial lung-expanding compound)	None	
Synthetic	DPPC, CPHT (Exosurf Neonatal)	None	

Trial		Dosage	Infants		
	Preparation		Age, wk (Weight, grams)	No.	Outcome
Enhorning, et al, 1985 ²⁴	CLSE	Single	< 30	72	↓Death, ↓IVH
Kwong et al, 1985 ²⁵	CLSE	Single	24-28	27	No difference
Kendig et al, 1988≊	CLSE	Single	25-29	65	↓Pneumothorax
Merritt et al, 1986 ²⁷	Human	Multiple	24-29	60	↓Death, ↓pneumothorax
Soll et al, 1990 ²⁸	Beractant	Single	24-30 (750-1,250)	156	↓Pneumothorax, ↑NEC
loekstra et al, 1991 ²⁹	Beractant	Multiple	23-29 (600-1,250)	430	\downarrow Death, \downarrow pneumothorax
Ten Centre Study Group, 1987 ³⁰	ALEC	Multiple	25-29	328	↓Death
Morley et al, 1988 ³¹	ALEC	Multiple	< 34	341	↓Death, ↓BPD, ↓IVH in < 30 wk
Phibbs et al, 1991 ³²	CPHT	Single	(700-1,350)	74	No difference
Bose et al, 1990 ³³	CPHT	Single	(700-1,350)	385	↓PIE
Stevenson et al, 1992 ³⁴	CPHT	Single	(500-699)	215	↓Pneumothorax, ↑pulmonary hemorrhage
Corbet et al, 1991 ³⁵	CPHT	Single	(700-1,100)	446	\downarrow Death, \downarrow pneumothorax

• Can the prophylactic administration of exogenous surfactant prevent RDS and its complications?

The prophylactic administration of surfactant aims at instituting surfactant in the airway as soon as the infant is delivered and before breathing has even begun. Possible advantages of this approach are indicated by experiments in prematurely delivered animals that showed that exogenous materials distribute better when instilled in a partially or totally fluid-filled airway²² and that, in the absence of surfactant, damage to the airway occurs early (in the first hour) after the initiation of mechanical ventilation.²³ Results from trials testing different preparations (Table 2)^{24.35} were subjected to meta-analyses.^{36,37}

From these analyses, it was concluded that the prophylactic administration of surfactant decreases neonatal mortality and the incidence of air leak. Although these effects did not reach statistical significance in all trials, they appeared consistent across populations and with all surfactant preparations tested. Despite initial hopes to the contrary, however, the incidence of bronchopulmonary dysplasia, intraventricular hemorrhage, and other complications of prematurity remained essentially unchanged. Because systematic prophylactic treatment of neonates at risk for RDS would result in a large number of them being subjected to unnecessary treatment, investigators were led to test exogenous surfactant therapy in infants with established RDS.

• Can surfactant therapy decrease the incidence and severity of RDS and its complications?

In the "rescue" mode of surfactant therapy (also referred to as "treatment"), infants are treated after the diagnosis of RDS has been established by clinical and radiologic criteria. Several investigators tested this approach in randomized trials (Table 3).³⁸⁻⁴⁹ Again, metaanalyses of the results showed a decrease in mortality and air leak similar to that found in prophylactic trials.^{36,37} Moreover, there was again little or no effect of surfactant therapy on sequelae associated with RDS. In particular, there was no notable effect on the incidence of bronchopulmonary dysplasia in survivors, although there was variability in the results among differentweight groups within and between studies. Direct comparative trials of the two approaches (prophylactic versus rescue) were also done.

• Is prophylactic better or worse than rescue therapy in decreasing mortality and morbidity associated with RDS?

Despite published results from five comparative human trials addressing this question (Table 4),⁵⁰⁻⁵⁴ the answer remains uncertain. Two trials found no significant differences between the two modes of administration.^{53,54} One study found increased oxygen requirement at 28 days after birth in the prophylactic group in contrast with decreased oxygen requirement at 36 weeks postconception in the rescue group.⁵⁰ On the contrary, a pronounced decrease in mortality and air leak was found in the group born at or before 26 weeks' gestation that received prophylactic therapy.⁵¹ It was also found that prophylactic therapy decreased mortality when compared with rescue administration in the population of infants (29 to 32 weeks' gestation) studied.⁵²

Although prophylactic treatment may lead to a delay in stabilizing a neonate, rescue treatment can be given as soon as the diagnosis of RDS is established, earlier than at the times tested in trials. The OSIRIS trial found a decreased incidence of "death or oxygen dependence at the expected date of delivery" and of air leak in infants treated early, before 2 hours of age, when compared with those treated later,⁵⁵ and a recent report from Japan found a decreased incidence of bronchopulmonary dysplasia in infants treated before 30 minutes of age compared with those treated at 6 hours of age.⁵⁶

• Do additional doses of surfactant provide further benefits to patients?

Two trials showed benefit from repeated doses,^{57,58} although the effects on mortality and the incidence of air

Trial			Infant Features				
	Preparation	Dosage	Weight, grams	No.	Lung Physiology	Hours After Birth	Outcome
Gitlin et al, 1987 ³⁸	Surfactant TA	Single	1,000-1,500	41	Fio ₂ >0.4	<8	↓Pneumothorax
Raju et al, 1987 ³⁹	Surfactant TA	Single	751-1,750	30	Fio ₂ >0.5	<6	↓Death and ↓pneumothorax;↑PDA
Fujiwara et al, 1990 [™]	Surfactant TA	Single	750-1,750	100	Fio ₂ >0.4		↓Pneumothorax, ↓IVH
Hallman et al, 1985 ⁴¹	Human	Multiple	<1,500	45	Fio ₂ >0.6	<10	↓Pneumothorax
Lang et al, 1990 ⁴²	Human	Multiple	24-32*; 1,500	59	Fio ₂ >0.8	<12	↓Pneumothorax
Horbar et al, 1989 ⁴³	Beractant	Single	750-1,750	159	Fio ₂ >0.4	3-6	↓Pneumothorax
Horbar et al, 19904	Beractant	Single	750-1,750	106	Fio ₂ >0.4	3-6	1îVH
Liechty et al, 1991 ⁴⁵	Beractant	Multiple	600-1,750	798	Fio ₂ >0.4	<8	↓Death, ↓pneumothora
Gortner et al, 1990*	SF-RI 1	Multiple	25-30*	69		<1	No difference
European Collaborative	. .				-		
Group, 1988 ⁴⁷	Porcine	Single	700-2,000	146	Fio ₂ > 0.6		↓Death, ↓pneumothora
Phibbs et al, 1991 ³²	CPHT	Single	>650	104	Fio ₂ >0.4-0.5		No difference
Long et al, 1991 ⁴⁸	CPHT	Multiple	700-1,350	419	a/A <0.22		↓Death, ↓pneumothora and PIE
Long et al, 1991 [,]	CPHT	Multiple	>1,250	1,237	a/A <0.22		↓Death; ↓BPD and pneumothorax; ↓IVH and ↓PDA

*Age in weeks.

leak reached significance only in the larger study.⁵⁸ It was reported in one abstract that infants treated with multiple doses of the synthetic surfactant CPHT had reduced neonatal mortality and a reduced need for oxygen and ventilatory support.⁵⁹ A later report of a large, albeit not placebo-controlled or blinded, international study indicated that more than two doses of CPHT appear to provide little, if any, additional benefit.⁵⁵

• Is surfactant instillation safe?

The most common adverse reaction seen after the tracheal instillation of surfactant was a transient deterioration of gas exchange, which responded well to and often could be prevented by an adjustment of the ventilator settings.^{60,61} Airway obstruction occurred with an incidence of about 3 patients per 1,000 in trials testing the use of CPHT.⁶² The incidence of such events when using other surfactant preparations has not been precisely ascertained.

An increase in the incidence of pulmonary hemorrhage was reported in a trial examining the effects of prophylactic CPHT in infants weighing 500 to 699 grams at birth.³⁴ On review of retrospective^{63,64} and autopsy analyses,⁶⁵ however, the relative risk for pulmonary hemorrhage with any surfactant therapy is estimated to be no greater than 1.5.⁶⁴

A report of an increase in the incidence of intraventricular hemorrhage in treated infants⁴⁴ was not confirmed by other trials and appears to have been an isolated incidence.⁶⁶

Administering human or animal-derived extracts to infants raised concerns over the infectious risk posed to patients and the potential for their sensitization to the protein fraction contained in the preparations. Isolated reports of increased incidence of bacterial sepsis did not reach statistical significance,²⁹ and to date no other report of serious infectious complication has appeared. Two studies failed to detect immune complexes⁶⁷ or circulating antibodies⁶⁸ in the serum of infants who received natural preparations, but one showed a suppression of the mitogen-induced proliferation of cord blood lymphocytes by the natural surfactant SF-RI 1.⁶⁸

Although the long-term follow-up of infants treated with surfactants has been limited both in the number of subjects and the duration, it has not shown adverse developmental effects of treatment.⁶⁹⁻⁷⁴ Increased survival of infants with disabilities might be expected, however, and several studies indicated such a trend.^{69,71,72}

• Are any of the available preparations better than the others?

There is currently one published trial comparing the efficacy of CPHT and beractant administered in a rescue mode.⁷⁵ Although there were small differences favoring the use of beractant in the effects on ventilatory settings needed to maintain comparable early gas exchange in the two groups, there was no observed difference in the mortality or incidence of any of the complications between the two groups. Studies comparing different preparations with those already tested are ongoing, but results have yet to be published.

Surfactant Therapy for Other Pulmonary Diseases

The main measurable effect of exogenous surfactant on lung mechanics of infants with RDS is an increase in functional residual capacity, possibly through stabilizing partially ventilated alveolar units and recruiting previ-

Trial	Preparation	Dosage				
			Age, wk	No.	Lung Physiology	Outcome
Dunn et al, 1991 ^{so}	CLSE	Multiple	< 30	182	<6 hr*	↓Pneumothorax and ↓BPD in treatment group
Kendig et al, 1991 ³¹	CLSE	Multiple	24-29	479	Fio ₂ >0.4	↓Death and ↓pneumo- thorax in prophylaxis group
Kattwinkel et al, 1993 ^{sz}	CLSE	Multiple	29-32	1,398	Fio ₂ > 0.3	JDeath in prophylaxis group
Merritt et al, 1991 ³³	Human	Multiple	24-29	203	Fio₂ >0.5; 2-12 hr*	No difference
Egberts et al, 1993⁵	Porcine	Multiple	26-29	147	Fio ₂ >0.6; 6-24 hr*	No difference

ously nonventilated ones.⁷⁶ Tidal volume and lung compliance, however, remain unchanged or decrease slightly.⁷⁶

Diseases of the lungs other than RDS may also be ameliorated with surfactant therapy. Studies in animals have suggested benefits of using exogenous surfactant in a variety of conditions, including pneumonia (bacterial,⁷⁷ aspiration⁷⁸), asthma,⁷⁹ and after lung transplantation.⁸⁰ In humans, exogenous surfactant improved lung mechanics and gas exchange in neonates with the meconium aspiration syndrome⁸¹ and in infants with respiratory failure requiring extracorporeal membrane oxygenation.⁸² The use of surfactant replacement therapy was also tested in adults and children with adult RDS. Preliminary reports have been encouraging,^{83,84} but controlled trials are clearly called for.

Remaining Questions, Future Prospects

Despite the success of surfactant replacement therapy in neonates with RDS, several questions pertaining to its use in clinical practice remain unanswered. In most trials a small fraction of the infants had little or no response to therapy.^{85,86} The proportion of these "nonresponders" varied with the population studied, but was as high as 24% in one study.⁸⁶ Subsequent analyses suggested that the presence of prenatal asphyxia increases the risk of a poor response,⁸⁷ as do those of large patent ductus arteriosus and high fluid intake.⁸⁸ A number of other factors could also contribute to limit the response to surfactant therapy:

• The dose of surfactant administered may be insufficient. The amount of surfactant administered in trials varied from 60 to 200 mg of phospholipids per kilogram. These doses were chosen largely on the basis of theoretical considerations, but little experimental work has been done to establish the correct dosing and administration regimen. Two trials specifically addressed the effects of the initial dosage. One found that administering 120 mg per kg of surfactant TA was more effective than half that dose.⁸⁹ Another showed that 67.5 mg per kg of CPHT was more effective than half and as effective as double that dose.⁹⁰

• The administration of surfactant may be ineffective because surfactant fails to reach the respiratory units or it is inactivated in situ, for instance by extravasated serum proteins.⁹¹ The distribution of exogenous surfactant in the lungs may also be affected by its mode of delivery, that is, bolus versus slow instillation.⁹² Aerosol delivery of surfactants is being tested in the treatment of patients with adult RDS,⁸⁴ but it has the serious disadvantage that, with current methods, administering an adequate dose takes hours or even days.

• Ventilation strategies may affect the response to therapy.⁸⁸ Preparations with different physical characteristics are expected to affect the patient's lung mechanics differently,⁷⁶ and ventilator settings may need to be adjusted accordingly, as has been shown in animals.⁹³

• Surfactant therapy is only a palliative treatment, limited in time, and patients ultimately depend on an effective endogenous surfactant production for full recovery. This point is illustrated by the report of a patient who died of neonatal RDS refractory to repeated exogenous surfactant administration and who was found at autopsy to lack the normal production of surfactant apoprotein B because of a genetic defect.^{94,95} Other genetic abnormalities are likely to be discovered that alter the normal production or functional properties of surfactant components.

• The administration of antenatal steroids is an important element in determining outcome in infants, as shown by several investigators. One report showed recently that infants whose mothers had received steroids antenatally had lower requirements for surfactant treatment or ventilator support and a lower incidence of intraventricular hemorrhage.⁹⁶ A retrospective analysis of trials using beractant showed that the prenatal administration of steroids reduced neonatal mortality to the same extent as did the postnatal administration of surfactant together have additive effects.⁹⁷ These observations confirm the results of an earlier analysis⁹⁸ and of animal experiments showing structural maturation of the lung after prenatal steroid therapy.⁹⁹

Finally, several laboratories are working to improve currently available preparations. The use of recombinant and synthetic peptides that attempt to mimic the properties of hydrophobic surfactant-associated proteins is a promising avenue of approach.¹⁰⁰⁻¹⁰²

Conclusions

In early 1995, have the closing lines in the fascinating story of surfactant biology been written? To the contrary, several important questions remain about pulmonary surfactant, its biologic processes, and its role in lung homeostasis and disease states. Furthermore, investigation of other modes of therapy, such as partial¹⁰³ or total¹⁰⁴ liquid ventilation, could lead to alternative treatments for special situations in which exogenous surfactant may prove not to be optimal therapy. Although we should not underestimate the achievements of recent years in this field, it is clear that much more research will be needed to complete our understanding of the lung surfactant system and to enable us to manipulate it to best advantage for patients with pulmonary diseases.

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REFERENCES

1. von Neergaard K: Neue Auffassungen über einen Grundbegriff der Atemmechanik: Retracktionskraft der Lunge, Abhängig von der berflächenspunnung in den Alveolen. Z Gesamte Exp Med 1929; 66:373-394

2. Pattle RE: Properties, function and origin of the alveolar lining layer. Nature 1955; 175:1125-1126

3. Clements J: Dependence of pressure-volume characteristics of lungs on intrinsic surface active material. Am J Physiol 1956; 187:592

4. Avery M, Mead J: Surface properties in relation to atelectasis and hyaline membrane disease. Am J Dis Child 1959; 97:517-523

5. Fujiwara T, Maeta H, Chida S, Morita T, Watabe Y, Abe T: Artificial surfactant therapy in hyaline-membrane disease. Lancet 1980; 1:55-59

6. King RJ: Pulmonary surfactant. J Appl Physiol 1982; 53:1-8

7. Possmayer F: A proposed nomenclature for pulmonary surfactant-associated proteins. Am Rev Respir Dis 1988; 138:990-998

8. King RJ, MacBeth MC: Physiochemical properties of dipalmitoyl phosphatidylcholine after interaction with an apolipoprotein of pulmonary surfactant. Biochim Biophys Acta 1979; 557:86-101

9. Persson A, Chang D, Rust K, Moxley M, Longmore W, Crouch E: Purification and biochemical characterization of CP4 (SP-D), a collagenous surfactant-associated protein. Biochemistry 1989; 28:6361-6367

10. Malhotra R, Haurum J, Thiel S, Sim RB: Interaction of C1q receptor with lung surfactant protein A. Eur J Immunol 1992; 22:1437-1445

11. Vilallonga F: Surface chemistry of $L-\alpha$ -dipalmitoyl lecithin at the air-water interface. Biochim Biophys Acta 1968; 163:290-300

12. Hawgood S, Benson BJ, Schilling J, Damm D, Clements JA, White RT: Nucleotide and amino acid sequences of pulmonary surfactant protein SP 18 and evidence for cooperation between SP 18 and SP 28-36 in surfactant lipid adsorption. Proc Natl Acad Sci USA 1987; 84:66-70

13. van Iwaarden F, Welmers B, Verhoef J, Haagsman HP, van Golde LMG: Pulmonary surfactant protein A enhances the host-defense mechanism of rat alveolar macrophages. Am J Respir Cell Mol Biol 1990; 2:91-98

14. Kuan SF, Rust K, Crouch E: Interactions of surfactant protein D with bacterial lipopolysaccharides—Surfactant protein D is an *Esch*-

erichia coli-binding protein in bronchoalveolar lavage. J Clin Invest 1992; 90:97-106

15. van Iwaarden JF, van Strijp JA, Ebskamp MJ, Welmers AC, Verhoef J, van Golde LM: Surfactant protein A is opsonin in phagocytosis of herpes simplex virus type 1 by rat alveolar macrophages. Am J Physiol 1991; 261(pt1):L204-L209

16. Wright JR, Youmans DC: Pulmonary surfactant protein A stimulates chemotaxis of alveolar macrophage. Am J Physiol 1993; 264:L338-L344

17. Nieman GF, Bredenberg CE: High surface tension pulmonary edema induced by detergent aerosol. J Appl Physiol 1985; 58:129-136

18. Liu MY, Wang LM, Li E, Enhorning G: Pulmonary surfactant will secure free airflow through a narrow tube. J Appl Physiol 1991; 71:742-748

19. Morgenroth K, Bolz J: Morphological features of the interaction between mucus and surfactant on the bronchial mucosa. Respiration 1985; 47:225-231

20. Weaver TE, Whitsett JA: Function and regulation of expression of pulmonary surfactant-associated proteins. Biochem J 1991; 273:249-264

21. Wright JR, Dobbs LG: Regulation of pulmonary surfactant secretion and clearance. Annu Rev Physiol 1991; 53:395-414

22. Jobe A, Ikegami M, Jacobs H, Jones S: Surfactant and pulmonary blood flow distributions following treatment of premature lambs with natural surfactant. J Clin Invest 1984; 73:848-856

23. Nilsson R, Grossmann G, Robertson B: Lung surfactant and the pathogenesis of neonatal bronchiolar lesions induced by artificial ventilation. Pediatr Res 1978; 12(pt1):249-255

24. Enhorning G, Shennan A, Possmayer F, Dunn M, Chen CP, Milligan J: Prevention of neonatal respiratory distress syndrome by tracheal instillation of surfactant: A randomized clinical trial. Pediatrics 1985; 76:145-153

25. Kwong MS, Egan EA, Notter RH, Shapiro DL: Double-blind clinical trial of calf lung surfactant extract for the prevention of hyaline membrane disease in extremely premature infants. Pediatrics 1985; 76:585-592

26. Kendig JW, Notter RH, Cox C, et al: Surfactant replacement therapy at birth: Final analysis of a clinical trial and comparisons with similar trials. Pediatrics 1988; 82:756-762

27. Merritt TA, Hallman M, Bloom BT, et al: Prophylactic treatment of very premature infants with human surfactant. N Engl J Med 1986; 315:785-790

28. Soll RF, Hoekstra RE, Fangman JJ, et al: Multicenter trial of single-dose modified bovine surfactant extract (Survanta) for prevention of respiratory distress syndrome. Pediatrics 1990; 85:1092-1102

29. Hoekstra RE, Jackson JC, Myers TF, et al: Improved neonatal survival following multiple doses of bovine surfactant in very premature neonates at risk for respiratory distress syndrome. Pediatrics 1991; 88:10-18

30. Ten Centre Study Group: Ten centre trial of artificial surfactant (artificial lung expanding compound) in very premature babies—Ten Centre Study Group. Br Med J (Clin Res) 1987; 294:991-996

31. Morley CJ, Greenough A, Miller NG, et al: Randomized trial of artificial surfactant (ALEC) given at birth to babies from 23 to 34 weeks gestation. Early Hum Dev 1988; 17:41-54

32. Phibbs RH, Ballard RA, Clements JA, et al: Initial clinical trial of EXOSURF, a protein-free synthetic surfactant, for the prophylaxis and early treatment of hyaline membrane disease. Pediatrics 1991; 88:1-9

33. Bose C, Corbet A, Bose G, et al: Improved outcome at 28 days of age for very low birth weight infants treated with a single dose of a synthetic surfactant. J Pediatr 1990; 117:947-953

34. Stevenson D, Walther F, Long W, et al: Controlled trial of a single dose of synthetic surfactant at birth in premature infants weighing 500 to 699 grams—The American Exosurf Neonatal Study Group I. J Pediatr 1992; 120(pt2):S3-S12 [erratum published in J Pediatr 1992; 120:762]

35. Corbet A, Bucciarelli R, Goldman S, Mammel M, Wold D, Long W: Decreased mortality rate among small premature infants treated at birth with a single dose of synthetic surfactant: A multicenter controlled trial. J Pediatr 1991; 118:277-284

36. Jobe AH: Pulmonary surfactant therapy. N Engl J Med 1993; $328{:}861{-}868$

37. Soll R, McQueen M: Respiratory distress syndrome, *In* Sinclair J, Bracken M (Eds): Effective Care of the Newborn Infant. New York, NY, Oxford University Press, 1992, pp 323-358

38. Gitlin JD, Soll RF, Parad RB, et al: Randomized controlled trial of exogenous surfactant for the treatment of hyaline membrane disease. Pediatrics 1987; 79:31-37

39. Raju TN, Vidyasagar D, Bhat R, et al: Double-blind controlled trial of single-dose treatment with bovine surfactant in severe hyaline membrane disease. Lancet 1987; 1:651-656

40. Fujiwara T, Konishi M, Chida S, et al: Surfactant replacement therapy with a single postventilatory dose of a reconstituted bovine surfactant in preterm neonates with respiratory distress syndrome: Final analysis of a multicenter, double-blind, randomized trial and comparison with similar trials. Pediatrics 1990; 86:753-764

41. Hallman M, Merritt TA, Jarvenpaa AL, et al: Exogenous human surfactant for treatment of severe respiratory distress syndrome: A randomized prospective clinical trial. J Pediatr 1985; 106:963-969

42. Lang MJ, Hall RT, Reddy NS, Kurth CG, Merritt TA: A controlled trial of human surfactant replacement therapy for severe respiratory distress syndrome in very low birth weight infants. J Pediatr 1990; 116:295-300

43. Horbar JD, Soll RF, Sutherland JM, et al: A multicenter randomized, placebo-controlled trial of surfactant therapy for respiratory distress syndrome. N Engl J Med 1989; 320:959-965

44. Horbar JD, Soll RF, Schachinger H, et al: A European multicenter randomized controlled trial of single dose surfactant therapy for idiopathic respiratory distress syndrome. Eur J Pediatr 1990; 149:416-423

45. Liechty EA, Donovan E, Purohit D, et al: Reduction of neonatal mortality after multiple doses of bovine surfactant in low birth weight neonates with respiratory distress syndrome. Pediatrics 1991; 88:19-28

46. Gortner L, Bernsau U, Hellwege HH, Hieronimi G, Jorch G, Reiter HL: A multicenter randomized controlled clinical trial of bovine surfactant for prevention of respiratory distress syndrome. Lung 1990; 168(suppl):864-869

47. Collaborative European Multicenter Study Group, Robertson B: Surfactant replacement therapy for severe neonatal respiratory distress syndrome. Pediatrics 1988; 82:683-691

48. Long W, Thompson T, Sundell H, Schumacher R, Volberg F, Guthrie R: Effects of two rescue doses of a synthetic surfactant on mortality rate and survival without bronchopulmonary dysplasia in 700- to 1350-gram infants with respiratory distress syndrome—The American Exosurf Neonatal Study Group I. J Pediatr 1991; 118(pt1):595-605

49. Long W, Corbet A, Cotton R, et al: A controlled trial of synthetic surfactant in infants weighing 1250 g or more with respiratory distress syndrome. N Engl J Med 1991; 325:1696-1703

50. Dunn MS, Shennan AT, Zayack D, Possmayer F: Bovine surfactant replacement therapy in neonates of less than 30 weeks' gestation: A randomized controlled trial of prophylaxis versus treatment. Pediatrics 1991; 87:377-386

51. Kendig JW, Notter RH, Cox C, et al: A comparison of surfactant as immediate prophylaxis and as rescue therapy in newborns of less than 30 weeks' gestation. N Engl J Med 1991; 324:865-871

52. Kattwinkel J, Bloom BT, Delmore P, et al: Prophylactic administration of calf lung surfactant extract is more effective than early treatment of respiratory distress syndrome in neonates of 29 through 32 weeks' gestation. Pediatrics 1993; 92:90-98

53. Merritt TA, Hallman M, Berry C, et al: Randomized, placebocontrolled trial of human surfactant given at birth versus rescue administration in very low birth weight infants with lung immaturity. J Pediatr 1991; 118(pt1):581-594

54. Egberts J, de Winter J, Sedin G, et al: Comparison of prophylaxis and rescue treatment with Curosurf in neonates less than 30 weeks' gestation: A randomized trial. Pediatrics 1993; 92:768-774

55. OSIRIS (Open Study of Infants at high risk of or with Respiratory Insufficiency—the role of Surfactant) Collaborative Group: Early versus delayed neonatal administration of a synthetic surfactant—The judgement of OSIRIS. Lancet 1992; 340:1363-1369

56. Konishi M, Fujiwara T, Chida S, et al: A prospective, randomized trial of early versus late administration of a single dose of surfactant-TA. Early Hum Dev 1992; 29:275-282

57. Dunn MS, Shennan AT, Possmayer F: Single- versus multipledose surfactant replacement therapy in neonates of 30 to 36 weeks' gestation with respiratory distress syndrome. Pediatrics 1990; 86:564-571

58. Speer CP, Robertson B, Curstedt T, et al: Randomized European multicenter trial of surfactant replacement therapy for severe neonatal respiratory distress syndrome: Single versus multiple doses of Curosurf. Pediatrics 1992; 89:13-20

59. Gerdes J, Cook L, Corbet A, Long W: Effects of three vs one prophylactic doses of Exosurf Neonatal in 700-1100 gm neonates (Abstr). Pediatr Res 1991; 29:1270A

60. Russell L, White A, Andrews E, et al: Observational study of synthetic surfactant in 11,455 infants (Abstr). Pediatr Res 1992; 31:100A

61. Zola EM, Overbach AM, Gunkel JH, et al: Treatment Investigational New Drug experience with Survanta (beractant). Pediatrics 1993; 91:546-551

62. Corbet A: Clinical trials of synthetic surfactant in the respiratory distress syndrome of premature infants. Clin Perinatol 1993; 20:737-760

63. Long W, Corbet A, Allen A, et al: Retrospective search for bleeding diathesis among premature newborn infants with pulmonary hemorrhage after synthetic surfactant treatment. J Pediatr 1992; 120:S45-S48

64. Raju TN, Langenberg P: Pulmonary hemorrhage and exogenous surfactant therapy: A meta-analysis. J Pediatr 1993; 123:603-610

65. van Houten J, Long W, Mullett M, et al: Pulmonary hemorrhage in premature infants after treatment with synthetic surfactant: An autopsy evaluation—The American Exosurf Neonatal Study Group I and the Canadian Exosurf Neonatal Study Group. J Pediatr 1992; 120(pt2):S40-S44

66. Gunkel JH, Banks PL: Surfactant therapy and intracranial hemorrhage: Review of the literature and results of new analyses. Pediatrics 1993; 92:775-786

67. Whitsett JA, Hull WM, Luse S: Failure to detect surfactant protein-specific antibodies in sera of premature infants treated with Survanta, a modified bovine surfactant. Pediatrics 1991; 87:505-510

68. Bartmann P, Bamberger U, Pohlandt F, Gortner L: Immunogenicity and immunomodulatory activity of bovine surfactant (SF-RI 1). Acta Paediatr 1992; 81:383-388

69. Dunn MS, Shennan AT, Hoskins EM, Lennox K, Enhorning G: Two-year follow-up of infants enrolled in a randomized trial of surfactant replacement therapy for prevention of neonatal respiratory distress syndrome. Pediatrics 1988; 82:543-547

70. Morley CJ, Morley R: Follow up of premature babies treated with artificial surfactant (ALEC). Arch Dis Child 1990; 65:667-669

71. Robertson B, Curstedt T, Tubman R, et al: A 2-year follow up of babies enrolled in a European multicentre trial of porcine surfactant replacement for severe neonatal respiratory distress syndrome. Eur J Pediatr 1992; 151:372-376

72. Vaucher YE, Harker L, Merritt TA, et al: Outcome at twelve months of adjusted age in very low birth weight infants with lung immaturity: A randomized, placebo-controlled trial of human surfactant. J Pediatr 1993; 122:126-132

73. Ware J, Taeusch HW, Soll RF, McCormick MC: Health and developmental outcomes of a surfactant controlled trial: Follow-up at 2 years. Pediatrics 1990; 85:1103-1107

74. Walter D, McGuinness G, Bose C, et al: Double blind one year follow-up in 1450 infants randomized to $Exosurf^{TM}$ Neonatal or air (Abstr). Pediatr Res 1991; 29:270A

75. Horbar JD, Wright LL, Soll RF, et al: A multicenter randomized trial comparing two surfactants for the treatment of neonatal respiratory distress syndrome. J Pediatr 1993; 123:757-766

76. Cotton RB, Olsson T, Law AB, et al: The physiologic effects of surfactant treatment on gas exchange in newborn premature infants with hyaline membrane disease. Pediatr Res 1993; 34:495-501

77. Lachman B: Surfactant therapy. Resuscitation 1989; 18:S37

78. Lamm WJ, Albert RK: Surfactant replacement improves lung recoil in rabbit lungs after acid aspiration. Am Rev Respir Dis 1990; 142:1279-1283

79. Richman PS, Batcher S, Catanzaro A: Pulmonary surfactant suppresses the immune lung injury response to inhaled antigen in guinea pigs. J Lab Clin Med 1990; 116:18-26

80. Christie N, Puskas J, Hirai T, et al: Surfactant improves gas exchange in canine lung allografts after extended hypothermic storage (Abstr). Am Rev Respir Dis 1992; 145:A306

81. Auten RL, Notter RH, Kendig JW, Davis JM, Shapiro DL: Surfactant treatment of full-term newborns with respiratory failure. Pediatrics 1991; 87:101-107

82. Lotze A, Knight GR, Martin GR, et al: Improved pulmonary outcome after exogenous surfactant therapy for respiratory failure in term infants requiring extracorporeal membrane oxygenation. J Pediatr 1993; 122:261-268

83. Richman P, Spragg R, Merritt T: Administration of porcine-lung surfactant to humans with ARDS: Initial experience (Abstr). Am Rev Respir Dis 1987; 135:A5

84. Weg J, Reines H, Balk R: Safety and efficacy of an aerosolized surfactant (Exosurf) in human sepsis induced ARDS. Chest 1991; 100:137S

85. Fujiwara T, Konishi M, Chida S, Maeta H: Factors Affecting Response to a Postnatal Single Dose of Exogenous Surfactant. Presented at the Conference Surfactant Treatment of Lung Diseases, Carefree, Arizona, 1987

86. Charon A, Taeusch W, Fitzgibbon C, Smith GB, Treves ST, Phelps DS: Factors associated with surfactant treatment response in infants with severe respiratory distress syndrome. Pediatrics 1989; 83:348-354

87. Mercier C, Soll R: Clinical trials of natural surfactant extract in respiratory distress syndrome. Clin Perinatol 1993; 20:711-736

88. Hallman M, Merritt TA, Bry K, Berry C: Association between neonatal care practices and efficacy of exogenous human surfactant: Results of a bicenter randomized trial. Pediatrics 1993; 91:552-560

89. Konishi M, Fujiwara T, Naito T, et al: Surfactant replacement therapy in neonatal respiratory distress syndrome—A multi-centre, ran-

domized clinical trial: Comparison of high- versus low-dose of surfactant TA. Eur J Pediatr 1988; 147:20-25

90. Berry D, Pramanik A, Philips J III, et al: Comparison of the effect of three doses of a synthetic surfactant on the alveolar-arterial oxygen gradient in infants weighing ≥1250 grams with respiratory distress syndrome. J Pediatr 1994; 124:294-301

91. Tierney DF, Johnson RP: Altered surface tension of lung extracts and lung mechanics. J Appl Physiol 1965; 20:1253-1260

92. Ueda T, Ikegami M, Rider E, Jobe A: Distribution of surfactant and ventilation in surfactant-treated preterm lambs. J Appl Physiol 1994; 76:45-55

93. Rider ED, Jobe AH, Ikegami M, Sun B: Different ventilation strategies alter surfactant responses in preterm rabbits. J Appl Physiol 1992; 73:2089-2096

94. Nogee ML, DeMello DE, Dehner LP, Colten HR: Brief report: Deficiency of pulmonary surfactant protein B in congenital alveolar proteinosis. N Engl J Med 1993; 328:406-410

95. Nogee L, Hamvas A, Murphy A, Dietz H: Molecular diagnosis of hereditary surfactant protein B (SP-B) deficiency (Abstr). Pediatr Res 1994; 35:1443A

96. Kari M, Hallman M, Eronen M, et al: Prenatal dexamethasone treatment in conjunction with rescue therapy of human surfactant: A randomized placebo-controlled multicenter study. Pediatrics 1994; 93:730-736

97. Jobe AH, Mitchell BR, Gunkel JH: Beneficial effects of the combined use of prenatal corticosteroids and postnatal surfactant on preterm infants. Am J Obstet Gynecol 1993; 168:508-513

98. Crowley P, Chalmers I, Keirse MJ: The effects of corticosteroid administration before preterm delivery: An overview of the evidence from controlled trials. Br J Obstet Gynaecol 1990; 97:11-25

99. Ikegami M, Berry D, elKady T, Pettenazzo A, Seidner S, Jobe A: Corticosteroids and surfactant change lung function and protein leaks in the lungs of ventilated premature rabbits. J Clin Invest 1987; 79:1371-1378

100. Cochrane CG, Revak SD: Pulmonary surfactant protein B (SP-B): Structure-function relationships. Science 1991; 254:566-568

101. Waring A, Taeusch W, Bruni R, et al: Synthetic amphipathic sequences of surfactant protein-B mimic several physiochemical and *in vivo* properties of native pulmonary surfactant proteins. Peptide Res 1989; 2:308-313

102. McLean LR, Krstenansky JL, Jackson RL, Hagaman KA, Olsen KA, Lewis JE: Mixtures of synthetic peptides and dipalmitoylphosphatidylcholine as lung surfactants. Am J Physiol 1992; 262:L292-300

103. Fuhrman BP, Paczan PR, DeFrancisis M: Perfluorocarbon-associated gas exchange. Crit Care Med 1991; 19:712-722

104. Shaffer TH, Wolfson MR, Clark LJ: Liquid ventilation. Pediatr Pulmonol 1992; 14:102-109