



Nutritional profile of the land snail *Cyclophorus saturnus*, a rich-in-nutrients food item from Thailand

Supap Nontasan^a, Rachanee Nammatra^b, Eakapol Wangkahart^{a,*}

^a Laboratory of Fish Immunology and Nutrigenomics, Applied Animal and Aquatic Sciences Research Unit, Division of Fisheries, Faculty of Technology, Maharakham University, Kantharawichai District, Maha Sarakham 44150, Thailand

^b Walai Rukhvej Botanical Research Institute, Maharakham University, Kantharawichai District, Maha Sarakham 44150, Thailand

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ABSTRACT

Cyclophorus saturnus is an edible land snail traditionally harvested for human food, yet little is known about its nutritional value, especially in Thailand. This study aimed to investigate its nutritional potential as an alternative food resource. In the present study, proximate composition, essential mineral content, amino acid, and lipid profiles of the meat were evaluated. Proximate analysis showed that *C. saturnus* contained 80.04% moisture, 11.88% protein, 6.04% carbohydrate, and 0.93% fat, with 80.01 kcal/100 g fresh matter. For minerals, calcium was the most abundant element in the meat. Its protein contained glutamic and aspartic as the major amino acids, while it was not a good source of tryptophan and methionine but was considered a very rich source of other essential amino acids (amino acid scores greater than 100). Its lipid fraction showed a higher proportion of mono and polyunsaturated fatty acids (MUFA and PUFA, 67.69%) and a lower proportion of saturated fatty acids (SFA) (32.31%). The PUFA/SFA ratio (1.56), hypocholesterolemic/hypercholesterolemic ratio (HH; 5.58), atherogenicity index (AI; 0.48), and thrombogenicity index (TI; 0.20) are considered nutritionally healthy for humans. Overall, this study demonstrates the nutritional potential of *C. saturnus* to serve as a nutritious part of the human diet and as an alternative ingredient in food systems; therefore, its production and consumption should be more extensively promoted.

1. Introduction

Land snails are a specialized food item that has long been prized by numerous countries, particularly Thailand, China, Taiwan, and Western European nations [1]. In recent years, snail farming has grown significantly worldwide due to the economic importance of land snails and growing demand for edible use and cosmetics [2]. According to estimates, 43,000 tons of snails were consumed globally in 2016, and 50,000 tons are expected to be consumed by the end of 2025 [3]. Snail meat has been reported to be rich in protein and have a low fat and cholesterol content that make it meet the demands of modern nutrition [4–6]. It has also been reported that snail meat is a valuable source of essential fatty acids, amino acids, and minerals [4,7]. Due to its nutritional benefits, snail meat has been considered a healthy alternative source of dietary protein for human consumption in many parts of the world [8].

In Thailand, land snails especially *Cyclophorus* genus are commonly used as an ingredient for regional cuisine such as Larb and Tom Yum Hoi [9]. The land snails of the genus *Cyclophorus* are predominantly active during the wet season and widely distributed in many

* Corresponding author.

E-mail address: eakapol.w@msu.ac.th (E. Wangkahart).

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regions of South East Asia including Thailand, particularly in humid regions of mountainous areas [10]. Typically, they are harvested by hand-picking during the rainy season. *Cyclophorus* snails have an operculum and a thick, massive pyramidal or turbinated shell with stripe patterns of brown, white and black colors on the surface [11]. The maximum size range of the snails can be between 20 and 25 mm in height and 30–40 mm in width. An adult female snail typically lays eggs (40–150 eggs per hatching) in moist soil, decaying logs, or dead leaves, and 5–8 months are required for larvae to develop into adults after hatching [9]. Some of their foods include fungi, lichen, decaying plants as well as fresh plants such as vegetables, tubers and flowers [11]. Among the various species of edible *Cyclophorus* snails, *Cyclophorus saturnus*, locally known as the “Hoihom or Fragrant snail,” is one of the most common species that is plentiful and consumed in Thailand because its meat has a unique aroma and good flavor when cooked [12,13].

As an alternative protein source for human beings, the nutritional potential of edible snails has recently received more attention. Some reports are available on the nutritional value of different species of land snails such as *Helix pomatia* [7,14], *Cornu aspersum*, *Eobania vermiculata* [4], *Theba pisana* [5]. However, to the best of our knowledge, no data are available on the nutritional properties of *C. saturnus*. Understanding their nutritional characteristics would make it easier to identify the components necessary for their marketability and promotion that could give it considerable economic value in the future. Therefore, the aim of this work was to investigate the nutritional potential of *C. saturnus* including fatty acids and amino acids as well as proximate and mineral compositions in the edible part of the snails collected from Thailand. Such snail would allow the development of many value added products to address the modern consumers who are increasingly looking for a nutritionally interesting material and reveal a rich-in-nutrients food item.

2. Materials and methods

2.1. Snail sample preparation and ethics statement

Snail samples (*C. saturnus*) with mixed sex were collected from Phu Phan Mountain range, northeastern Thailand. Snails were kept in plastic bags with ice during transport to laboratory and then stored at -20°C until analysis. The samples were prepared according to the modified procedures of Galluzzo et al. [4]. A total of 200 specimens with mean individual weights of 5.09 ± 0.50 g without considering snail gender was used in this study. The edible portion of snail muscle was taken from the shell, homogenized, and used for chemical analyses with three replications performed for each analysis. The Institute of Animals for Scientific Development (IAD) of Thailand's guidelines for the use of animals in research were followed strictly during the study. The handling of land snails in this study was authorized by the Mahasarakham University ethics council (IACUC-MSU-21/2023).

2.2. Proximate and energy value analysis

The moisture content was analyzed by drying until reaching a constant weight at 105°C in an oven, according to the methods specified by the Association of Official Analytical Chemists [15]. Then the loss of weight on drying was used to calculate the amount of moisture in snail samples and presented as the % of fresh matter (FM). Protein assay was performed using an auto Kjeldahl System (Buchi B-324/I-437, Uster, Switzerland) following the Kjeldahl procedure [15]. Protein content was calculated using a determined nitrogen content with a conversion factor of 6.25 ($\text{N} \times 6.25$). The lipid in snail samples was quantified using the Soxhlet extraction apparatus (Gerhardt GmbH, Königswinter, Germany) using hexane as an extraction solvent [15]. Ash was measured by the incineration of samples at 550°C in a muffle furnace for 7 h [15]. The total carbohydrate content was obtained by calculation using a simple subtraction method, while the energy value of the snail samples was calculated using energy conversion factors and presented as kcal/100 g FM as the sum of the percentage composition of lipids, protein, and total carbohydrate, multiplying by factors of 9, 4, and 4, respectively, as described by Gomot [16].

2.3. Mineral composition analysis and estimated daily intakes (EDIs)

Snail meat samples were assessed for various mineral elements including calcium (Ca), magnesium (Mg), sodium (Na), iron (Fe), manganese (Mn), zinc (Zn), and copper (Cu) using atomic absorption spectrophotometry (AAS, ZEEnit 700 P, Analytik Jena, Germany), as described by Nkansah et al. [6] with some modifications. Briefly, snail meat sample was dried at 105°C . Ten milliliters of nitric acid (HNO_3) were pipetted into a digestion flask containing 1 g of dried sample and heated in a fume hood until the fumes disappeared. The cooled mixture was then mixed with 4 mL of 70% perchloric acid (HClO_4) and heated again to obtain a clear liquid before it was cooled and diluted to 50 mL with distilled water. A clear aliquot was taken for absorbance measurement using AAS. The calibration curve of each mineral standard was generated, and the mineral content in sample was calculated, with results expressed as mg/100 g of dried sample (DM). Colorimetric determination of total phosphorus (P) by the molybdate method was carried out after calcination and incineration of snail meat sample using UV/Vis Spectrophotometer (SPECORD 210 PLUS, Analytik Jena, Germany), according to the AOAC procedure [15].

The body weight of the consumers, the daily snail consumption rate, and the element concentration all contribute to estimated daily intakes (EDIs) of trace elements from snail consumption. Estimation was done using Equation (1) [17].

$$\text{EDIs} = \text{C metal} \times \text{C factor} \times \text{D food intake} / \text{body weight} \quad (1)$$

Where C metal, C factor and D food intake are the mean essential metal concentration in the various snail species (mg/100 g), the

Table 1
Proximate composition and energy value of *C. saturnus*.

Composition	% (FM)	% (DM)
Moisture	80.04 ± 0.39	–
Protein	11.88 ± 0.03	59.51 ± 0.15
Fat	0.93 ± 0.08	4.66 ± 0.40
Ash	1.12 ± 0.02	5.61 ± 0.10
Carbohydrate	6.04 ± 0.30	30.26 ± 1.50
Energy (kcal/100 g)	80.01 ± 1.36	400.85 ± 6.81

Values are expressed as mean ± standard error of the mean (SEM).

FM = fresh matter basis, DM = dry matter basis.

conversion factor (0.085), the daily consumption rate of snail (kg/day), respectively. In this investigation, the body weight was estimated at 70 kg.

2.4. Fatty acid composition analysis

Snail meat samples' fatty acid profiles were determined following a modified version of the previous methods [18,19]. Fatty acid methyl esters (FAMES) were prepared from the lipid extract of snail sample using saponification followed by methylation procedures. The extract (1 mL) was saponified with 4 mL of 0.5 M NaOH in methanol at 80 °C for 60 min, and then, the methylation of fatty acids was performed by the addition of 5 mL of methanolic BF₃ reagent at 80 °C for 20 min. The obtained mixture was cooled and added to saturated NaCl (5 mL). FAMES were subsequently extracted with 2 mL of iso-octane (2, 2, 4-trimethylpentane) and transferred into an amber glass autosampler vial before analysis of the fatty acid composition using gas chromatography (430-GC model, Bruker, Billerica, MA, USA) equipped with a flame ionization detector (GC-FID). Chromatographic separation was performed on a fused silica Rt-2560 GC column (100 m × 0.25 mm ID, 0.50 μm phase; Restek Corporation, Bellefonte PA, USA) with a 1 mL/min flow rate, using nitrogen as the carrier gas. Individual fatty acids were identified and calculated based on a comparison of their retention times to the FAME standards and reported as a percentage of fatty acids in lipid fraction as well as mg of individual fatty acids per 100 g of fresh snail meat. The nutritional quality indexes of lipids, including the total saturated fatty acids (ΣSFA), total monounsaturated fatty acids (ΣMUFA), total polyunsaturated fatty acids (ΣPUFA), total n3 PUFA (Σn3), total n6 PUFA (Σn6), ΣPUFA/ΣSFA, and n3 PUFA/n6 PUFA ratio, were calculated based on the fatty acid composition, while the hypocholesterolemic/hypercholesterolemic (HH) ratio, atherogenicity index (AI), and thrombogenicity index (TI) were estimated using Equations (2)–(4) [20].

$$HH = (\text{cis} - C18:1 + \Sigma PUFA) / (C12:0 + C14:0 + C16:0) \quad (2)$$

$$AI = [C12:0 + (4 \times C14:0) + C16:0] / \Sigma MUFA \quad (3)$$

$$TI = (C14:0 + C16:0 + C18:0) / [(0.5 \times \Sigma MUFA) + (0.5 \times \Sigma n6) + (3 \times \Sigma n3) + (n3/n6)] \quad (4)$$

2.5. Amino acid profile analysis

The amino acid composition of the snail meat samples was determined using an automatic amino acid analyzer (ARACUS, MembraPure, Hennigsdorf, Germany) following the method of Wangkahart et al. [21] with minor modifications. Snail samples were hydrolyzed at 110 °C for 24 h with 6 M HCl containing 0.5% 2-mercaptoethanol, followed by using a nitrogen blower to eliminate HCl from the digested solution. The residual amount was then redissolved with 0.02 M HCl and subsequently passed through a 0.45-μm syringe filter prior to analysis of the amino acid composition. A separate study was used to determine the tryptophan content. The weighted samples were hydrolyzed for 20 h at 110 °C in 4 M LiOH containing 95 mM ascorbic acid. The hydrolysate was then neutralized with 6 M HCl, centrifuged, and the supernatant was then passed through a 0.45-μm syringe filter and examined via an automatic amino acid analyzer.

The composition was then computed by comparison with the retention time and peak areas of the standards and expressed as mg/g fresh meat and mg/g of protein (DM). The essential amino acid score (as a percentage of adequacy) of the snail meat sample was calculated based on the reference amino acid pattern for adults [22] using the following equation:

$$\text{Amino acid score} = \text{Amino acid content in sample} \times 100 / \text{Reference amino acid pattern} \quad (5)$$

3. Results and discussion

3.1. Proximate composition and energy value

The data from the proximate analysis and energy value of the *C. saturnus* sample, reported on both a fresh and dry matter basis, are

shown in Table 1.

3.1.1. Moisture content and dry matter

The amount of water is one of the key factors indicating the sensory quality of the flesh in terms of texture, tenderness and juiciness [23–25]. Moisture was the main ingredient in fresh *C. saturnus* meat, 80.04% of the content, and 19.96% dry matter was identified. The moisture value of the *C. saturnus* meat sample was in the range of the moisture content reported for different species of land snails (75.20–84.91% FM) by previous studies, according to Pissia, Matsakidou, and Kiosseoglou [8]. The observed value of moisture content in *C. saturnus* meat was close to the result previously reported in land snail *H. pomatia* from Turkey (80.80% FM) [14] and *C. haughtoni* from Thailand (81.85%) [9], while the lower levels were recorded in *Archachatina Marginata* (73.67% FM) and *Achatina* from Nigeria (75.28% FM) [26]. In other fresh meats were reported to contain water as the main component such as beef (73.1%), and lamb (72.9%) but lower than that found in *C. saturnus* meat.

3.1.2. Protein content

The *C. saturnus* sample was found to contain 11.88% protein FM, which could make it a good source of protein compared with the protein values found in cereal grains, 7–18% FM [27]. Previous research reported that different species of land snails collected from different locations contained protein values varying between 10.08% and 20.76% FM [8,26,28]. This is the first report on the content of protein in *C. saturnus* snails from Thailand, and the amount is slightly lower than that reported for *C. haughtoni* (14.07% FM), a land snail in the same genus found in Thailand [9] and *Helix aspersa* (12.87% FM), from Turkey [29]. Milinsk et al. [30] also reported a content of protein ranging from 9.50% to 12.56% FM in *H. aspersa*, a farmed snail fed a different feed formula. The protein content of *C. saturnus* was found to be lower than the levels found in conventional meats, including pork, beef, and chicken (17.3–24.1% FM) [31], and fish (15–20% FM) [32].

3.1.3. Fat content

In this study, the content of fat of *C. saturnus* (0.93% FM) was close to the fat value reported by Babalola and Akinsoyinu [26] for *Limicolaria* species (1.05% FM), a common edible land snail found in Nigeria. However, previous studies showed that the fat content in edible snail meat could be as low as 0.38% FM, as recorded for *C. haughtoni* collected from Thailand [9] and 0.41% FM for *H. aspersa* collected from Turkey [29]; in addition, the content was reported as 2.44% FM in *Archachatina marginata* obtained from Nigeria by Babalola and Akinsoyinu [26]. The fat content in *C. saturnus* was considered as low as 0.93% when compared to the lean portion in some raw meats such as pork loin (4.7% FM), beef loin (3.3–7.6% FM), duck meat (6.2% FM) and chicken breast (1.2–8.9% FM) [31]; therefore, it can be an alternative meat for people with fat-related diseases.

3.1.4. Ash content

The mean total ash detected in *C. saturnus* was 1.12% FM, which is within the range of ash content (between 1.07% and 2.33% FM) previously described in the literature for several species of edible land snails collected from different sites [8].

3.1.5. Total carbohydrate content

The percentage of total carbohydrates found in *C. saturnus* was 6.04% FM, as shown in Table 1. This value was slightly lower than the content of total carbohydrate reported by Babalola and Akinsoyinu [26] (6.91% FM) for *Limicolaria* sp., a land snail obtained from a natural area in Oyo state, Nigeria. Caetano et al. [5] demonstrated that snails contained a variety of carbohydrate types, including monomers like glucose and reserve polysaccharides, primarily glycogen. Different values of total carbohydrate, estimated indirectly, were observed for different land snail species. For *C. haughtoni*, the land snail species of the same *Cyclophorus* genus that was collected from Thailand showed a lower value of total carbohydrate [9]. Babalola and Akinsoyinu [26] reported levels of total carbohydrates in wild snail meat of 1.8% FM for *A. marginata* and 7.25% FM for *Achatina fulica*, while Milinsk et al. [30] reported that between 7.07% and 10.01% FM of total carbohydrates were observed in the meat of farmed snails (*H. aspersa maxima*) fed different diets. Moniruzzaman et al. [33] demonstrated that freshwater snails and clams from Bangladesh contained carbohydrates in amounts of 30.2–57.3% DM, higher than *C. saturnus* (30.26% DM). However, very low values of total carbohydrates were also reported by Fagbuaro et al. [34], with less than 1% FM for several land snail species including *A. marginata ovum*, *A. marginata saturalis*, *Achatina*, and *Limicolaria* spp. Variation in the amount of total carbohydrate in land snails could be related to several factors including the snail's species, diet, and location [5].

3.1.6. Energy value

The energy value obtained for *C. saturnus* meat was 80.01 kcal/100 g FM, as shown in Table 1. This value was higher than that reported in the literature for *H. aspersa maxima* meat (66 kcal/100 g FM) by Gomot [16], while it was within the range of energy value (76.92–87.08 kcal/100 g FM) observed for the same species of snail meat by Milinsk et al. [30]. The energy value of *C. saturnus* meat was in the range of the values recorded for freshwater mollusk species, between 346 and 436 kcal/100 g DM [33]. However, *C. saturnus* meat was low in calories even compared with other lean cuts of meat such as chicken breast (108–176 kcal/100 g FM), pork loin (131 kcal/100 g FM), and beef loin (114 kcal/100 g FM) [31]. Therefore, due to the reasonable amount of protein, the consumption of snail meat could help to reduce excessive intake of carbohydrates and fats.

Table 2
Mineral composition and estimated daily intakes of trace elements from the intake of *C. saturnus*.

Parameter	mg/100 g FM	mg/100 g DM	EDIs
Macro mineral			
Calcium (Ca)	172.65 ± 0.98	865.00 ± 5.00	5.25E-3
Phosphorus (P)	100.80 ± 0.98	505.00 ± 5.00	3.06E-3
Magnesium (Mg)	36.93 ± 0.98	185.00 ± 5.00	6.07E-6
Trace mineral			
Sodium (Na)	22.98 ± 0.03	115.15 ± 0.15	6.99E-4
Iron (Fe)	8.97 ± 0.04	44.96 ± 0.22	6.06E-6
Zinc (Zn)	3.32 ± 0.01	16.63 ± 0.04	1.01E-4
Copper (Cu)	1.06 ± 0.01	5.29 ± 0.04	3.21E-5
Manganese (Mn)	0.42 ± 0.01	2.09 ± 0.02	1.26E-5

Values are expressed as mean ± standard error of the mean (SEM).

FM = fresh matter, DM = dry matter, EDIs = Estimated daily intakes.

3.2. Macro mineral compositions

The macro mineral compositions on a fresh and dry matter basis and the EDIs of Ca, P, and Mg are presented in Table 2. Among all the minerals measured, Ca was the most abundant in *C. saturnus* meat, (865 mg/100 g DM or 172.65 mg/100 g FM); these results agree with the findings of earlier studies on the major elements in other land snail species [5]. Nkansah et al. [6] also reported the content of Ca, the most plentiful mineral, in the meat of three snail species from Ghana, *A. marginata* (701.79 mg/100 g DM), *A. achatina* (656.9 mg/100 g DM), and *A. fulica* (402.09 mg/100 g DM), while Kalio and Etela [35] reported values in the range of 144–175 mg/100 g FM in *A. marginata*. Additionally, *C. saturnus* meat was an excellent source of Ca compared to some food products (on a fresh matter basis) such as fish (9–103 mg/100 g FM) [36], whole milk (124 mg/100 g FM), chicken eggs (64 mg/100 g FM) [37], chicken (12 mg/100 g FM), pork (6 mg/100 g FM), and beef (15 mg/100 g FM) [38]. Ca estimated EDIs were low when compared to their acceptable daily intake levels [39,40].

P was the next most abundant mineral element in *C. saturnus* meat. Both elements are essential minerals involved in several physiological processes in the human body, including bone formation [41]. In this study, the content of P (505 mg/100 g DM; 100.8 mg/100 g FM) was comparable to that of the wild Roman snail, *H. pomatia* (104.52 mg/100 g FM) [14], and higher than the values of 268.53, 241.9, and 61.29 mg/100 g DM reported in the meats of *A. marginata*, *A. achatina*, and *A. fulica*, respectively [6]. However, a higher concentration of P was previously reported by Fagbuaro et al. [34] for different species of giant land snail meat, in the range of 123.23–153.89 mg/100 g FM, which was higher than the value recorded in this study on the basis of fresh matter (Table 2). *C. saturnus* meat contains P at a lower level than traditional meats such as chicken, beef, and pork, with values in the range of 173–229 mg/100 g FM [38]. P estimated EDIs were low when compared to their acceptable daily intake levels [39,40].

Mg is also important to reduce blood pressure levels and improve cardiovascular health [42]. The result obtained in this study was 185 mg/100 g DM (36.93 mg/100 g FM), lower than that in the literature for *H. pomatia* (54.05 mg/100 g FM) [14]. Comparing the result of Mg on a fresh basis obtained in this study with other meats such as fresh fish of different species (16–109 mg/100 g FM) [36], lamb (19 mg/100 g FM), chicken (25 mg/100 g FM), and pork (25 mg/100 g FM) [38], it can be suggested that *C. saturnus* meat is a rich source of Mg. Finally, Mg estimated EDIs were low when compared to their acceptable daily intake levels [39,40].

3.3. Trace mineral elements

The trace mineral elements on a fresh and dry matter basis and the EDIs of Na, Fe, Zn, Cu and Mn are presented in Table 2. Considering the content of Na on a wet basis in snail meat, *C. saturnus* provided a lower concentration of Na (22.57 mg/100 g FM) than that found for other species of land snails (50.8–60.94 mg/100 g FM) [34] and for *Otala lactea* (60.1 mg/100 g FM) and *T. pisana* (92.2 mg/100 g FM) species [5]. The content obtained was also low when compared to traditional meats such as chicken, pork, and lamb, which contain 77, 55, and 46 mg/100 g FM, respectively [38].

Additionally, *C. saturnus* meat contained a considerable amount of Fe (44.96 mg/100 g DM; 8.97 mg/100 g FM) when compared to the content found in Roman land snail species (*H. pomatia*) (1.71 mg/100 g FM) [14] and the most popular snail species in Africa, including *A. marginata* (6.33 mg/100 g DM), *A. achatina* (5.75 mg/100 g DM), and *A. fulica* (26.64 mg/100 g DM) [6]. However, Gomot [16] observed a high content of Fe of the deshelled part in different species of reared snails, ranging between 30.4 and 78 mg/100 g DM. Fe is a crucial mineral involved in a range of metabolic activities in humans, including oxygen transport, hemoglobin formation, DNA synthesis, immune response, and energy production [43]. Flesh foods are rich in absorbable iron, recommended to prevent iron deficiency, particularly for children, teenagers, and women of reproductive age [43]. The presence of Fe in *C. saturnus* meat (8.81 mg/100 g on a fresh matter basis) was also high when compared with that in other fresh meats such as lamb (1.64 mg/100 g), chicken (0.89 mg/100 g), and pork (1.01 mg/100 g) [38], indicating that *C. saturnus* meat is an excellent source of Fe.

Meat products have also been reported as good sources of Zn, with the trace mineral playing numerous biological roles in human metabolism such as a cofactor of several enzymes [28]. The mean percentage of Zn obtained in this study was 3.32 mg/100 g FM, which was higher than the values reported by Adeyeye [28] for *A. marginata* (1.44 mg/100 g FM), *Archatina* sp. (1.87 mg/100 g FM), and *Limicolaria* sp. (1.23 mg/100 g FM), land snail species collected from Nigeria, whereas Kalio and Etela [35] reported a higher Zn

Table 3
Fatty acid composition and concentration obtained from the edible part of *C. saturnus*.

Fatty acid	% of total fatty acids	mg/100 g fresh meat
Myristic acid (C14:0)	0.66 ± 0.04	6.11 ± 0.41
Pentadecylic acid (C15:0)	1.32 ± 0.08	12.26 ± 0.73
Palmitic acid (C16:0)	10.96 ± 0.38	101.96 ± 3.55
Margaric acid (C17:0)	4.21 ± 0.15	39.14 ± 1.35
Stearic acid (C18:0)	11.58 ± 0.03	107.68 ± 0.32
Arachidic acid (C20:0)	1.51 ± 0.15	14.05 ± 1.44
Heneicosylic acid (C21:0)	2.07 ± 0.22	19.27 ± 2.06
Total saturated fatty acids (Σ SFA)	32.31	300.47
Oleic acid (C18:1 n9c)	14.32 ± 0.26	133.19 ± 2.37
Gondoic acid (C20:1)	2.88 ± 0.01	26.77 ± 0.02
Total monounsaturated fatty acids (Σ MUFA)	17.20	159.96
Linoleic acid (C18:2 n6c)	13.60 ± 0.09	126.43 ± 0.85
γ -Linolenic acid (C18:3 n6)	6.53 ± 0.42	60.72 ± 3.86
Eicosadienoic acid (C20:2)	6.60 ± 0.05	61.34 ± 0.51
Eicosatrienoic acid (C20:3 n3)	3.24 ± 0.30	30.15 ± 2.79
Arachidonic acid (C20:4 n6)	17.86 ± 0.24	166.06 ± 1.58
Eicosapentaenoic acid (C20:5 n3; EPA)	2.67 ± 0.05	24.86 ± 0.51
Total polyunsaturated fatty acids (Σ PUFA)	50.49	469.56
PUFA/SFA ratio	1.56	
Σ n6	44.58	
Σ n3	5.92	
n6/n3 ratio	7.54	
HH	5.58	
TI	0.48	
AI	0.20	

Values are expressed as mean \pm standard error of the mean (SEM).

content in the range of 7.5–9.6 mg/100 g FM for *A. marginata*, a farmed snail fed different diets. Our study indicated that *C. saturnus* meat provided a Zn content close to that of raw beef and mutton at 3.6 and 3.8 mg/100 g FM; however, it was higher than that recorded in raw pork (1.6–2.7 mg/100 g FM), chicken (0.8–1 mg/100 g FM) [31], and fish (0.38–0.99 mg/100 g FM) [36].

Low levels of other minerals present in *C. saturnus* meat included Cu and Mn (Table 2). Both Cu and Mn are trace minerals required for numerous biological functions in the human body [6]. When compared to other land snail species such as *A. marginata* [35], with Cu and Mn values ranging from 0.56 to 0.76 and 0.15–0.25 mg/100 g FM, respectively, *C. saturnus* meat was superior in Cu (1.06 mg/100 g FM) and Mn (0.42 mg/100 g FM). According to data reported by Wright et al. [44], high concentrations of Cu and Mn were recorded in oysters (raw) in the ranges of 1.58–2.86 and 0.30–0.64 mg/100 g meat, respectively. They also reported lower levels than in conventional meats such as pork, chicken, and beef, with Cu content in the range of 0.05–0.09 mg/100 g fresh meat and Mn levels below the detection limit of assay (less than 0.01 mg/100 g) [44]. The level of Mn in *C. saturnus* (as reported in dry weight; Table 2) was considered close to the value in *Pomacea canaliculata* snails from a commercial farm in the Republic of Korea (2.0 mg/100 g DM), whereas *P. canaliculata* contained a higher concentration of Cu (7.1 mg/100 g DM) [1]. Overall, Na, Fe, Zn, Cu and Mn estimated EDIs were low when compared to their acceptable daily intake levels [39,40].

3.4. Fatty acid composition

The fatty acid profiles for *C. saturnus* meat consider three main types: saturated fatty acids (SFAs), monounsaturated fatty acids (MUFAs), and polyunsaturated fatty acids (PUFAs), as shown in Table 3. Fifteen fatty acids were detected in *C. saturnus* meat, including seven SFAs, two MUFAs, and six PUFAs. Interestingly, the most prevalent type of fatty acid in *C. saturnus* meat was PUFA (50.49% of lipid fraction or 469.56 mg/100 g fresh snail meat), followed by SFA (32.31% or 300.47 mg/100 g fresh snail meat) and MUFA (17.20% or 159.96 mg/100 g fresh snail meat), respectively. The highest proportion of PUFA was also reported by Galluzzo et al. [4] in other species of land snail (*T. pisana*, *C. aspersum*, and *E. vermiculata*), as up to 48% of the total fatty acids. In addition, Milinsk et al. [30] revealed that the meat of farmed snail *H. aspersa maxima* fed with different feeds had high proportions of PUFA in the range of 49.9–57.1%, while proportions of MUFA (20.7–23.8%) were not significantly different from those of SFA (20.2–26.6%). For meats from main animal species such as lamb, pork, beef, and chicken, the levels of total PUFA were 5.80, 11.86, 14.42, and 30.74%, respectively; however, these meats contained high levels of MUFA as well as SFA [38]. *C. saturnus* snail meat was rich in unsaturated fatty acids (UFA) and low in SFA, which is more desirable and nutritionally beneficial due to the potential health benefits, i.e., lowering the risk of various metabolic diseases [45].

Considering each type of fatty acid, stearic acid (C18:0) and palmitic acid (C16:0) were the major representatives of SFA, about 35.84% and 33.93% of the total SFA, respectively, while oleic acid (C18:1 n9c) was the most predominant MUFA (about 83% of the total MUFA). These findings are in agreement with previous studies on the predominance of these fatty acids in land snail meat such as *H. pomatia* [14], *H. aspersa* [29], *T. pisana*, *C. aspersum*, and *E. vermiculata* [4]. Evidence indicates that higher SFA intake—in particular, myristic, palmitic, and lauric acids—is associated with an elevation of low-density lipoprotein cholesterol levels and increased risk of cardiovascular disease, while stearic acid decreases such effects [46].

Table 4
Amino acid composition of the edible part of *C. saturnus*.

Amino acid	% of total amino acids	mg/g of crude protein (DM)	mg/g fresh meat
Essential amino acid			
Arginine ^a	8.41 ± 0.17	74.78 ± 1.51	8.88 ± 0.18
Histidine	2.31 ± 0.01	20.56 ± 0.02	2.44 ± 0.01
Isoleucine	5.03 ± 0.01	44.70 ± 0.05	5.31 ± 0.01
Leucine	8.15 ± 0.02	72.50 ± 0.22	8.61 ± 0.03
Lysine	7.42 ± 0.07	66.00 ± 0.65	7.84 ± 0.08
Phenylalanine	5.04 ± 0.21	44.82 ± 1.85	5.32 ± 0.22
Threonine	6.80 ± 0.03	60.50 ± 0.28	7.19 ± 0.03
Valine	5.61 ± 0.02	49.93 ± 0.22	5.93 ± 0.03
Non-essential amino acid			
Alanine ^b	7.27 ± 0.04	64.62 ± 0.34	7.68 ± 0.04
Aspartic acid ^b	11.66 ± 0.11	103.65 ± 0.98	12.31 ± 0.12
Glutamic acid ^b	14.77 ± 0.18	131.31 ± 1.61	15.60 ± 0.19
Glycine ^b	6.89 ± 0.03	61.26 ± 0.30	7.28 ± 0.04
Hydroxyproline	0.93 ± 0.02	8.30 ± 0.16	0.99 ± 0.02
Proline	2.56 ± 0.13	22.74 ± 1.15	2.70 ± 0.14
Serine	3.23 ± 0.13	28.69 ± 1.13	3.41 ± 0.13
Tyrosine ^a	3.93 ± 0.02	34.95 ± 0.22	4.15 ± 0.03
ΣEAA	48.78	433.78	51.53
ΣNEAA	51.22	455.51	54.11
ΣAA	100	889.30	105.63

Values are expressed as mean ± standard error of the mean (SEM).

^a Conditionally essential amino acid.

^b Flavor-enhancing amino acids.

Arachidonic acid (C20:4 n6) and linoleic acid (C18:2 n6c) were recorded as the major PUFA, constituting up to 35.4% and 26.9% of the total PUFA present in *C. saturnus*, respectively. The predominance of linoleic and arachidonic acids was also reported in other snail species such as *H. aspersa* [29], *T. pisana* [4], and specifically *Lunella undulata*, a turban snail with a high content of arachidonic acid (34.37% of total PUFA) [47]. It has been well documented that linoleic and arachidonic acids are the dominant n6-PUFA present in the human diet, especially in animal source foods, and have a wide range of biological activities and health effects such as linoleic acid's reduction of blood cholesterol and LDL cholesterol concentration and arachidonic acid's role in brain development and function [38, 45,48]. In this study, eicosatrienoic acid (ETE) (C20:3 n3) and eicosapentaenoic acid (EPA) (C20:5 n3) were the major n3 PUFA found in *C. saturnus* meat in small amounts (5.92%) compared to the total n6 PUFA content (44.58%). This finding was consistent with that of other snail meat species containing a small amount of n3 PUFA such as *H. pomatia* (2.51%) [14] and *H. aspersa maxima* (5.51–6.19%) [30]. The dietary intake of long-chain n3 PUFAs such as EPA is linked to a reduced risk of noncommunicable diseases due to the essential role in various physiological mechanisms such as anti-inflammatory effects, neurologic and visual development, and lowering blood triacylglycerol levels [45].

Regarding a nutritional quality index, the ratio of n6/n3 PUFA is a classic index frequently used in the evaluation of dietary fat and oil quality, with optimal values recommended as a healthy diet ranging from 1 to 4 [49]. The ratio of n6/n3 in snail meat reported by Milinsk et al. [50] for farmed *H. aspersa maxima* ranged from 5.01 to 7.05. High n6/n3 ratios were also reported in edible land snails from different farms in Poland, with values between 6.12 and 10.20 for *C. aspersa maxima*, and between 6.56 and 10.26 for *C. aspersum* [7]. Additionally, the ratio of n6/n3 in *C. saturnus* meat was comparable to other traditional meats, such as some fish species (5.07–8.79) [51], skinless chicken (7.81) and pork (7.4) [52].

Additionally, the PUFA/SFA ratio is a typical value used for the evaluation of the nutritional quality of dietary foods; it is employed to estimate how food affects cardiovascular health, with higher values being associated with more beneficial health impacts [20]. It has been suggested that undesirable values of PUFA/SFA ratio are under 0.45 [53]; however, high values could lead to oxidative stress [54]. The favorable values of PUFA/SFA that lower the risk of cardiovascular diseases are in the range of 1–1.50, as suggested by Kang et al. [55]. The PUFA/SFA ratio of *C. saturnus* meat determined in the present study was 1.56, which was considered nutritionally healthy for humans. Szkucik et al. [7] reported these values in land snail species cultured on different farms, varying between 1.10 and 1.55 for *C. aspersa maxima* and 1.40 and 1.59 for *C. aspersum*. Higher PUFA/SFA values were reported in edible land snails collected from Italy, including *T. pisana* (2.32), *C. aspersum* (1.75), and *E. vermiculata* (1.82) [4]. However, a low PUFA/SFA ratio was recorded by Özogul et al. [14] for Roman land snails (*H. pomatia*), exhibiting a value of 0.68. For meats from other animal species, PUFA/SFA values ranged between 0.20 and 2.10 in different shellfish species, while the values in fish varied between 0.95 and 1.60 [56]. Pork, beef, lamb, and chicken showed lower PUFA/SFA values of 0.31, 0.35, 0.12, and 0.95, respectively [38].

The hypocholesterolemic/hypercholesterolemic (HH) ratio is one of the nutritional quality indices used to assess dietary lipid profiles; it is frequently used to evaluate the impact of particular fatty acids and hypercholesterolemic fatty acids on cholesterol metabolism [20]. Diets with a high ratio of HH are more nutritionally desirable and better for health since they contain a higher level of hypocholesterolemic fatty acids (oleic acid and PUFA) than hypercholesterolemic fatty acids (myristic, lauric, and palmitic acids) [56]. In this study, *C. saturnus* showed an index of 5.58, which was greater than the values found for shellfish (0.21–4.75) [56] and some fish species such as raw *Kutum* roach (3.59) [57] due to a lower proportion of C14:0 and C16:0 (Table 3).

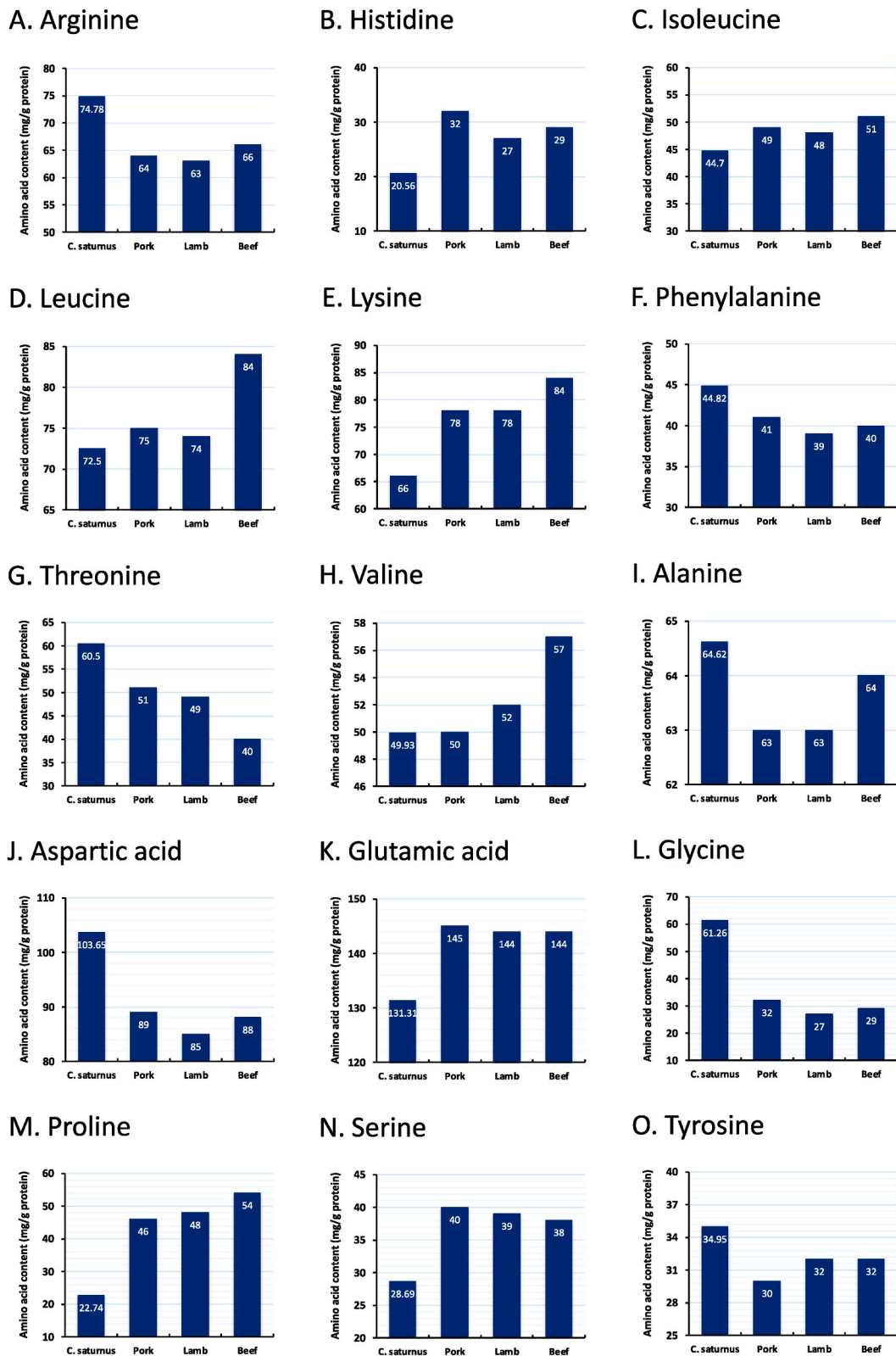


Fig. 1. Comparison of some amino acids among *C. saturnus* meat and conventional meats (Data adopted from Mazhangara et al. [70]). (A) Arginine. (B) Histidine. (C) Isoleucine. (D) Leucine. (E) Lysine. (F) Phenylalanine. (G) Threonine. (H) Valine. (I) Alanine. (J) Aspartic acid. (K) Glutamic acid. (L) Glycine. (M) Proline. (N) Serine. (O) Tyrosine.

Table 5

Amino acid scores of *C. saturnus* meat and comparison of essential amino acid composition between *C. saturnus* meat and amino acid reference pattern for adults [22].

Amino acid	Contents in sample (mg/g of protein)	Reference pattern	Amino acid score
Histidine	20.56	15	137.07
Isoleucine	44.70	30	149.00
Leucine	72.50	59	122.88
Lysine	66.00	45	146.67
Phenylalanine + Tyrosine	79.77	38	209.92
Threonine	60.50	23	263.04
Valine	49.93	39	128.03
Tryptophan	ND	6	–
Methionine	ND	16	–
Total AA with tyrosine	393.96		

ND = Not detected.

Atherogenicity index (AI) shows the link between the main classes of SFA and UFA, the former of which include myristic, lauric, and palmitic acids (but not stearic acid), which are regarded as pro-atherogenic, whereas UFA is considered anti-atherogenic [20]. Thrombogenicity index (TI) is also used to describe the thrombogenic potential (tendency for blood clot formation) of fatty acids by describing the interaction between prothrombogenic fatty acids (myristic, lauric, and palmitic acids) and antithrombogenic fatty acids (MUFA and PUFA) [20]. Diets with lower values of AI and TI are considered more nutritionally beneficial for cardiovascular health [1]. In this study, the AI and TI values of *C. saturnus* fat were 0.48 and 0.20, respectively—slightly lower than the values reported for farmed snail species such as *Pomacea canaliculata* from the Republic of Korea, which exhibited AI and TI values of 0.55 and 0.41, respectively [1]. For some species of shellfish such as scallops (*Chlamys farreri* and *Patinopecten yessoensis*), AI values ranged from 0.29 to 0.37, and TI values ranged from 0.09 to 0.17 [58]. For different fish species, the values of AI were reported to be in the range of 0.21–1.16, and the values of TI ranged from 0.14 to 0.87 [59–61].

3.5. Amino acid composition

The amino acid composition in *C. saturnus* meat is presented as mg/g meat (on a fresh matter basis) and mg/g of crude protein (on a dry matter basis) in Table 4. The total amino acid content measured was 105.63 mg/g fresh meat (889.3 mg/g of crude protein, DM). The meat contained essential and nonessential amino acids with values of 51.53 and 54.11 mg/g fresh meat, respectively. One of the most significant indicators of protein's nutritional quality is the food's composition and proportion of essential amino acids. It was demonstrated that *C. saturnus* meat was a good source of essential amino acids (arginine, histidine, isoleucine, leucine, lysine, phenylalanine, threonine, and valine), which constituted 48.78% of the total amino acids detected. Arginine is classified as a conditionally essential amino acid for infants and especially for vulnerable adults experiencing pregnancy, stress, critical illness, or disorders [1,62]. Among the essential amino acids detected, arginine, lysine, and leucine were dominant in the snail meat sample, which corresponded to previous reports on the amino acid profile in the snails *Limicolaria* sp., *A. archatina*, *A. marginata* [63], and *P. canaliculata* [1]. However, the essential amino acids tryptophan and methionine were not detected in the *C. saturnus* sample. Both of these amino acids were not also found in another land snail species of genus *Cyclophorus* (*C. fulguratus*) collected from Thailand [11], while they were found at a marginal level in *P. canaliculata* [1]. Previous studies have indicated that methionine is the limiting amino acid in land snails [29,64]. Adeyeye and Afolabi [63] also reported that tryptophan was not found in land snails, *Limicolaria* sp., *A. archatina*, and *A. marginata*.

Interestingly, the land snail *C. saturnus* had a relatively high concentration of flavor-enhancing amino acids, particularly glutamic and aspartic acids (131.31 and 103.65 mg/g protein DM, respectively), which contributes to its pleasant umami taste [65]. Glutamic acid acts as a precursor for the production of glutathione and a neurotransmitter, gamma-aminobutyric acid (GABA), while aspartic acid has physiological functions as nucleic acid and nucleotide constituents [66]. The highest content of glutamic acid was also recorded by Adeyeye and Afolabi [63], for land snails *A. archatina* (111 mg/g protein DM) and *A. marginata* (144 mg/g protein DM), while the aspartic acid concentrations of these snails were 69.4 and 72.7 mg/g protein DM, respectively. Additionally, the presence of a high glutamic acid content is in accordance with the previous report for the snail *Cookia sulcata* (138–141 mg/g protein DM), while this species had lower aspartic acid (69–70 mg/g protein DM) than the value obtained in the present study [67]. In the present study, alanine and glycine were also found in high concentrations, which enhances the pleasant sweet–umami taste of *C. saturnus* meat [68]. It has also been reported that, because amino acids participate in Maillard reactions when heated during processing conditions, their high concentration may also affect flavor due to the generation of volatile compounds [69]. When compared to conventional meats such as beef, pork, and lamb [70] *C. saturnus* meat contained comparable amounts of some amino acids, especially arginine (Fig. 1A), phenylalanine (Fig. 1F), threonine (Fig. 1G), alanine (Fig. 1I), aspartic acid (Fig. 1J), glycine (Fig. 1L), and tyrosine (Fig. 1O).

The amino acid score, a method of evaluating the quality of protein of *C. saturnus* meat samples, is shown in Table 5. The amino acid scores were greater than 100 in all essential amino acids, but the level of tryptophan and sulfur-containing amino acid (methionine) was low and unable to detect. However, it appears to be a very valuable source of other essential amino acids, which are considered crucial for human consumption [22].

Table 6Nutritional properties of *C. saturnus* meat and respective percentages of the recommended daily intake (%RDI) provided by 100 g of *C. saturnus* meat.

Nutrients	Recommended value ^a	Content per 100 g fresh meat	%RDI
Energy (kcal)	2000	80.01	4.00
Protein (g)	50	11.88	23.76
Total fat (g)	65	0.93	6.20
Carbohydrate (g)	300	6.04	2.01
Calcium (mg)	800	172.65	21.58
Phosphorus (mg)	800	100.80	12.60
Magnesium (mg)	350	36.93	10.55
Sodium (mg)	2000	22.98	1.10
Iron (mg)	15	8.97	59.83
Zinc (mg)	15	3.32	22.13
Copper (mg)	2	1.06	52.79
Manganese (mg)	3.5	0.42	11.92

^a Thai Recommended Daily Intakes (Thai RDI) for population over 6 years of age based on a 2000 kcal daily energy requirement.

3.6. Overview of nutritional properties of *C. saturnus* meat

The key nutritional data for *C. saturnus* meat is summarized in Table 6. The respective percentage of recommended daily intake (Thai RDI) [71] for each parameter is also shown taking into account a portion of 100 g of the meat, simulating the official labeling of this food item. Findings indicated that an intake of 100 g of the snail meat could offer 80 kcal, which is equivalent to 4% of the total energy in a 2000 kcal daily diet, and this serving is a great source of protein and minerals including Ca, P, Mg, Fe, Zn, Cu, and Mn, contributing more than 10% of the RDI for each, whereas low contributions (less than 10%) of the Na, energy, fat, and carbohydrate RDI were achieved from this snail portion.

4. Conclusions

This is the first report on the nutritional composition of *C. saturnus* compared to other edible snail meats and conventional foods of animal origin. Nutritional elements present in the snail meat indicate that *C. saturnus* can be considered a valuable dietary source of various health-related nutrients. *C. saturnus* contains a reasonable amount of protein and provides fewer calories due to the lower fat and carbohydrate contents. It is a rich source of essential minerals such as Ca, Mg, Fe, Zn, Cu, and Mn. In terms of the fatty acid profile, *C. saturnus* contains a high level of UFA and a low content of SFA, and properly balanced PUFA/SFA, n6/n3, HH, AI, and TI ratios that are considered healthy for humans. It is also a good source of essential amino acids arginine, histidine, isoleucine, leucine, lysine, phenylalanine, threonine and valine, while methionine and tryptophan are the limiting amino acids. In addition, *C. saturnus* contains a high level of flavor-enhancing amino acids that could contribute to the delicious taste of the meat. To enrich sustainable food supplies, it is important to support and promote the production and consumption of this snail species worldwide, especially in developing countries.

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Author contribution statement

Supap Nontasan, Eakapol Wangkahart: Conceived and designed the experiments; Performed the experiments; Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

Rachanee Nammatra: Contributed reagents, materials, analysis tools or data; Wrote the paper.

Data availability statement

The data that has been used is confidential.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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