Understanding COPD Etiology, Pathophysiology, and Definition

Jeffrey L Curtis

Introduction COPD Etiology and Pathology Multiple Pathways to COPD Status Exposures Leading to COPD as Currently Defined Small Airways Disease Is a Prominent Early COPD Pathology Initiated in Part by Loss of Local Innate Immune Defenses Immune Inflammation Can Drive SAD Progression to Emphysema SAD Can Be Detected by Imaging Abnormalities That Correlate to Histopathological Changes Additional Pathologic Processes Leading to COPD Rationale for a Revised Definition of COPD Is Airway Narrowing or Air Flow Limitation Universal in Those With COPD Risk Factors? The Dilemma of Redefining COPD Summary

COPD, one of the leading worldwide health problems, currently lacks truly disease-modifying medical therapies applicable to most patients. Developing such novel therapies has been hampered by the marked heterogeneity of phenotypes between individuals with COPD. Such heterogeneity suggests that, rather than a single cause (particularly just direct inhalation of tobacco products), development and progression of COPD likely involve both complex gene-by-environment interactions to multiple inhalational exposures and a variety of molecular pathways. However, there has been considerable recent progress toward understanding how specific pathological processes can lead to discrete COPD phenotypes, particularly that of small airways disease. Advances in imaging techniques that correlate to specific types of histological damage, and in the immunological mechanisms of lung damage in COPD, hold promise for development of personalized therapies. At the same time, there is growing recognition that the current diagnostic criteria for COPD, based solely on spirometry, exclude large numbers of individuals with very similar disease manifestations. This concise review summarizes current understanding of the etiology and pathophysiology of COPD and provides background explaining the increasing calls to expand the diagnostic criteria used to diagnose **COPD** and some challenges in doing so. Key words: COPD; pathophysiology; small airways disease; nosology. [Respir Care 2023;68(7):859-870. © 2023 Daedalus Enterprises]

Introduction

The goals of this review are 2-fold: to summarize succinctly the current understanding of the etiology and pathophysiology of the condition presently recognized as COPD and to explain the increasing calls for an expanded redefinition of COPD not based purely on spirometry.¹⁻⁵ The focus is predominately on human data, with selected supplementation on mechanisms from experimental studies. I draw upon a variety of evidence, in particular from 2 ongoing multi-center observational cohort studies, COPDGene⁶ and SPIROMICS.⁷

COPD Etiology and Pathology

Multiple Pathways to COPD Status

COPD is a highly prevalent, heterogeneous, variably progressive spectrum of conditions resulting primarily from lung injury and repair following prolonged oxidative inhalational exposures.^{4,8-13} COPD has a long latency and is greatly underdiagnosed;14,15 it can progress even when inciting exposures are removed,¹⁶ suggesting that once its underlying pathological processes attain momentum they become self-sustaining. As a disease currently defined by abnormal lung function, it is unsurprising but important to appreciate that COPD can be reached not only by accelerated loss of lung function, long postulated to be the primary pathway,¹⁷ but also by failure to achieve normal lung function in early adulthood with subsequent normal age-related decline¹⁸ or by both.¹⁹ Accordingly, early childhood exposures^{20,21} and genetic causes of impaired lung growth²² may account for as much of half of COPD cases, based on accepted spirometry criteria, but many of those individuals will not progress to severe disease.^{18,20} Inclusion of such individuals with late mild COPD in clinical trials may dilute the effect of potential disease-modifying agents targeting specific molecular pathways.

Exposures Leading to COPD as Currently Defined

In industrialized nations, direct inhalation of tobacco products has been long recognized as the principal causative exposure leading to COPD. In developing nations, by contrast, air pollution (both indoor due to biomass fuel use and outdoor) is a major contributor.^{23,24} Unfortunately, almost all data on lung pathology in COPD come from analysis of smoking-related disease in regions of high sociodemographic index (SDI). Far more work is needed on

DOI: 10.4187/respcare.10873

other exposures as potential etiological agents before extrapolation to the Global South can be assured.

However, this etiological dichotomy based on SDI is overly simplistic to explain disease status and outcomes in individuals. Risks of COPD are not uniformly distributed among residents of one high-SDI nation, the United States, but instead reflect a complex mix of differences in race/ethnicity, social class, and geographic location.²⁵⁻²⁷ These factors are doubtless applicable worldwide and will probably worsen due to climate change.²⁸⁻³⁰ COPD is also common among never smokers in the industrialized world, with apparent sex-based differences; eg, in the population-based Rotterdam study, smoking history was absent in far more females than males with COPD (27.2% vs 7.3%).³¹ Importantly, different oxidant inhalation exposures can interact to drive adverse COPD outcomes.^{32,33} Collectively, these findings imply the urgent need to reduce or eliminate all toxic inhalational exposures.

Small Airways Disease Is a Prominent Early COPD Pathology Initiated in Part by Loss of Local Innate Immune Defenses

Small airways are defined as those < 2 mm in diameter; in adults, they are stated to be distributed between the fourth-14th (mean eighth)³⁴ or eighth-22nd (mean 14th)³⁵ generation of airway branching. This region is distal to the point of maximal airway resistance and is characterized by diminishing individual airway diameters but a rapidly expanding total cross-sectional airway area. These distal airways are uniquely susceptible to damage induced by inhaled oxidative stresses such as smoking and air pollution for several reasons. The small airways are the region where gas transport switches from laminar or turbulent flow to diffusion, which facilitates deposition of fine particles and may enhance time of exposure to gas-phase irritants. Additionally, as considered below, small airways are much more dependent than the proximal airways on innate immune defenses that are altered by smoking.

Changes in small airways are the best studied pathology in COPD and likely the primary smoking-induced lesion in most of those who progress to significant air flow obstruction.³⁶ Elegant retrograde catheter studies in the 1960s demonstrated that small airways are the major site air flow obstruction in the human lung, with their contribution to total peripheral resistance increased 4–40-fold in advanced emphysema.³⁷ A recent study using archival paraffinembedded lung sections also found damage and loss of terminal and transitional bronchioles in spirometrically mildmoderate COPD. Loss of small airways in Global Initiative for Chronic Obstructive Lung Disease (GOLD) 1–2 patients was significant in regions without emphysema, with a 40% reduction in small airway numbers in those with GOLD stage 1 obstruction.³⁸ These studies indicate the importance

Dr Curtis is affiliated with Medical Service, VA Ann Arbor Healthcare System, Ann Arbor, Michigan; Division of Pulmonary and Critical Care Medicine, Michigan Medicine, Ann Arbor, Michigan; and Graduate Program in Immunology, University of Michigan, Ann Arbor, Michigan.

Dr Curtis discloses relationships with AstraZeneca PLC, Novartis AG, and CSL Behring LLC.

This work was supported by R01 HL144718, R01 HL144849, and U01 HL137880 from National Institutes of Health, National Heart, Lung, and Blood Institute; and I01 CX002377 and I01 CX001969 from the Department of Veterans Affairs. The opinions expressed are solely those of the author and do not reflect the official positions of the Department of Veterans Affairs or the Department of Health and Human Services.

Correspondence: Jeffrey L Curtis MD, Pulmonary and Critical Care Medicine Section (506/111G), VA Ann Arbor Healthcare System, 2215 Fuller Road, Ann Arbor, MI 48105-2303. E-mail: jlcurtis@umich.edu.

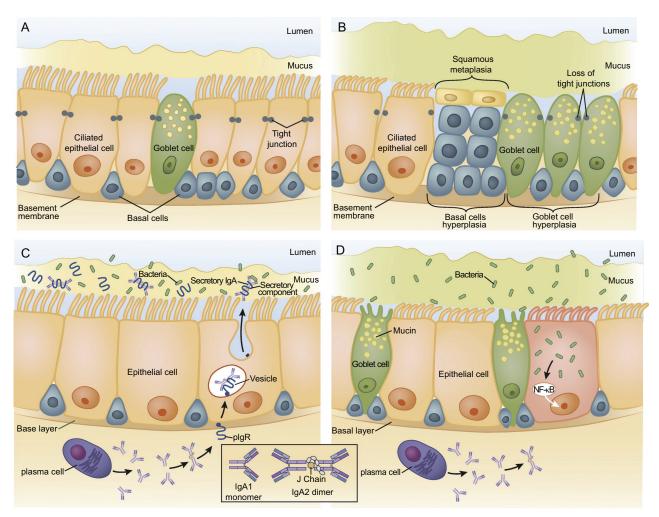


Fig. 1. Epigenetic changes induced by smoking lead to progressive small airway damage and inflammation in early COPD. (A and C) Normal small airways; (B and D) small airways in early COPD. A: Normal distal epithelium contains self-renewing basal cells, which differentiate into ciliated, mucus-producing goblet, and secretory (club) cells, joined by tight junctions that form an impermeable barrier. Mucus is separated from the epithelial surface by a robust aqueous periciliary layer. B: Smoking induces hyperplasia of basal and goblet cells, squamous metaplasia, loss of club and ciliated cells, decrease in the periciliary layer and ciliary damage and crowding, and junctional barrier loss. C: In normal small airways, dimeric immunoglobulin A (IgA) (structure shown in inset) is transcytosed by the polymeric immunoglobulin receptor (plgR) into the mucosal lumen. plgR cleavage at the luminal surface liberates secretory IgA, which prevents bacterial invasion. D: Smoking reduces plgR expression, leading to localized secretory IgA deficiency in small airways, allowing bacteria to invade and induce sustained airway inflammation. Illustration by Patricia Ferrer Beals. From Reference 40, with permission. NF-KB = nuclear factor kappa B; plgR = polymeric immunoglobulin A.

of identifying small airway disease (SAD) early to develop therapies to stop its progression. Clinical detection of SAD by pulmonary function testing has been difficult, as the process has minimal effect on FEV₁, and even sensitive techniques such as forced expiratory flow during the middle half of the FVC maneuver (FEF_{25–75%}) are highly variable. Nevertheless, use of spirometry on large populations can overcome that limitation, and results demonstrate the high global prevalence of SAD.³⁹ Alternative physiological methods to determine SAD include oscillometry, determination of residual volume, profiling the washout of inert gases, and machine learning approaches to analyze expiratory flow patterns. The proximate cause of SAD is smoking-induced reprograming of airway epithelium in multiple ways that collectively reduce small airway immune defenses.^{40,41} Healthy airway epithelium comprises a diverse community of cell types that all derive from basal progenitor cells (Fig. 1A). Smoking epigenetically reprograms these progenitors, causing them to undergo not only hyperplasia but also skewed maturation, with goblet cell expansion and loss of club cells and ciliated cells.^{42,43} Hyperplastic goblet cells produce mucus in greater amounts and with altered physical properties, which the reduced numbers of ciliated cells have difficulty clearing⁴⁴ (Fig. 1B). The adverse effect of this change is shown by the association of total sputum mucin concentrations with multiple adverse COPD outcomes, including annualized exacerbation rates, and with imaging or spirometric measures of SAD.⁴⁵ Visual scoring of mucus plugs detectable by computed tomography (CT), predominately in subsegmental airways, is associated with FEV₁ and hypoxemia, independently of emphysema and with a similar effect size.⁴³ Hence, reversing mucus hypersecretion could be one means of halting COPD progression.

Equally or perhaps more important to SAD pathogenesis is the local loss of host defenses resulting from epithelial reprograming. CC16, a secretoglobin family member that is the most abundant product of club cells, is reduced in smokers and to even a greater degree in mild and severe COPD.⁴⁶ In a transgenic murine cigarette smoke exposure model, CC16 protected lungs by reducing lung nuclear factor kappa B (NF-KB) activation.⁴⁶ Reprogramming also causes loss of translocation of dimeric secretory immunoglobulin A (sIgA) into small airway lumens.^{47,48} In health, sIgA binds bacteria and their products, leading to their elimination without inducing inflammation. In small airways, sIgA must be translocated by the polymeric Ig receptor (pIgR), which is specifically downregulated by smoking^{48,49} (Fig. 1C). This process is crucial, as it is now well recognized that even healthy lungs are not sterile but instead are repeatedly exposed to oropharyngeal bacteria.⁵⁰ In the absence of luminal sIgA, these bacteria can adhere and invade the epithelium, inducing NF-KB activation, which generates chemotactic molecules that attract inflammatory cells49,51 (Fig. 1D). Bacterial clearance from the lungs is further compromised by smoking and, to a greater degree, COPD development due to reduced host defense capabilities of the principal resident lung phagocytes.52-54 lung macrophages.55

These factors are directly relevant to the issue of why only some individuals with identical oxidant inhalational exposures develop COPD and why its manifestations are heterogeneous in those who do. One plausible explanation is focally stochastic interactions between epithelial-invasive bacteria and weakened local immunity. In other words, regional variation in host defenses, typified by focal loss of CC16 and pIgR and exaggerated production of abnormal mucus, sets the stage for variability in airway damage, both between individuals and between different lung regions in the same individual.⁵⁶⁻⁵⁸ Inheritable differences in bronchial anatomy that affect deposition of inhaled or aspirated microparticles are also likely a component of variability.^{59,60} That differences between individuals in the community composition of their oropharyngeal bacterial microbiome, the source of microaspirated organisms, are another factor is plausible but not entirely confirmed.⁶¹⁻⁶⁴ It is clear that advanced COPD is characterized by substantial changes in the community composition of the lung bacterial microbiome. However, whether changes in that community structure precede and drive the development of lower respiratory tract damage remains controversial.61-65

Immune Inflammation Can Drive SAD Progression to Emphysema

The Hogg lab recently provided additional evidence on how inflammation in terminal airways could lead to adjacent emphysema.⁶⁶ Using micro-CT, they demonstrated that loss of terminal bronchioles in COPD occurs in regions of microscopic emphysematous destruction (average air space size between 500–1,000 μ m). Analysis of gene expression there showed enrichment for interferon-gamma and for the chemokines CXL9-11 that it induces. By the CIBERSORT technique,⁶⁷ they also identified enrichment of CD8+ and CD4+ T cells and B cells.⁶⁶

These findings extend a significant body of studies collectively implying that adaptive immune mechanisms amplify lung damage initiated by defective lung host defense.^{10,68-71} Evidence is particularly strong for a role for B cells in emphysema pathogenesis,^{69,72-83} possibly facilitated by appearance within distal lung parenchyma of organized lymphoid follicles,^{84,85} which contain germinal centers that permit production of high-affinity antibodies. This process is even seen in panlobular emphysema due to alpha-1 anti-protease deficiency (AA1PD), the quintessential evidence for the protease-anti-protease imbalance hypothesis of COPD pathology. Explants from patients with AA1PD had notable CD4+ and CD8+ T cell infiltration and lymphoid follicles that were even more prominent that those from patients with usual emphysema.⁸⁶ Thus, auto-aggressive adaptive immune mechanisms likely amplify distal lung damage initiated as SAD into frank emphysema.

SAD Can Be Detected by Imaging Abnormalities That Correlate to Histopathological Changes

Noninvasive detection of SAD in living individuals has become possible by the imaging technique known as parametric response mapping (PRM)⁸⁷ and related techniques.^{88,89} Detailed cryo-CT analyses of human lung tissue frozen in standardized inflation after resection confirm that, at least in advanced COPD, the metric of functional small airways disease, PRM^{fSAD} (ie, persistently hyperinflated, non-emphysematous lung regions), actually denotes small airway pathology.90 This work has been extended by analyzing longitudinal change in PRM^{fSAD} and emphysema over time in the COPDGene cohort.⁹¹ Results showed evolution of initial areas of SAD into later centrilobular emphysema (Fig. 2). These findings suggest that imaging might be used as the outcome in clinical trials of agents designed to arrest SAD, as has been used successfully to prevent emphysema in AA1PD by augmentation therapy.92 Abnormal PRMfSAD is not specific for the SAD of smokers, as it has also been observed in never smokers with histologically confirmed constrictive bronchiolitis resulting from exposures during military deployment to Southwest Asia.⁹³

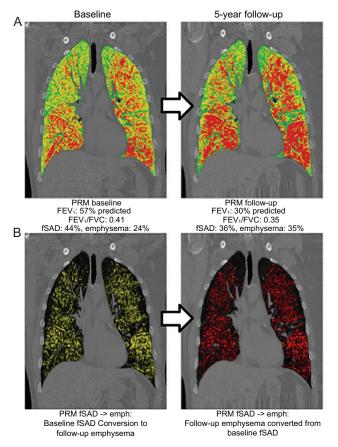


Fig. 2. Illustration of changes in parametric response mapping (PRM) metrics in a representative male patient with COPD. (A, B) Representative coronal computed tomography sections of the same individual at baseline (left) and after 5-y follow-up (right). A: All PRM metric values are depicted as normal lung parenchyma (green), functional small airway disease (fSAD, yellow) and emphysema (red). B: Only those individual voxels that were classified as fSAD at baseline (yellow) and which became emphysematous are shown. From Reference 91, with permission. PRM = parametric response mapping; fSAD = functional small airway disease; emph = emphysema.

Additional Pathologic Processes Leading to COPD

Thus, the processes leading to SAD are sufficient to induce centrilobular emphysema and may be the major pathway in many individuals. However, SAD cannot explain the complete heterogeneity of COPD phenotypes and is unlikely even to be the only pathway to centrilobular emphysema (Table 1). An alternative, not mutually exclusive, process is loss of pulmonary microcirculation, which is supported by evidence from both human pathological specimens⁹⁴ and in vitro and experimental animal models.⁹⁵⁻¹⁰⁰ Using dynamic contrast-enhanced magnetic resonance imaging, the Multi-Ethnic Study of Atherosclerosis COPD Study found 30% reductions in pulmonary microvascular blood flow even in mild COPD; correlations were strongest with emphysema and were independent of measures of SAD.¹⁰¹

Table 1.	Pathological	Processes	Leading to	Emphysema
----------	--------------	-----------	------------	-----------

loss of p	ulmonary microvasculature
Direct ap	optosis of airway epithelial cells
Epithelial	cell death deriving from loss of matrix attachments

Additionally, alveolar destruction might result from induction of epithelial cell apoptosis, either following loss of matrix attachments due to remodeling¹⁰² or from the direct action of natural killer cells, which show enhanced cytotoxicity of autologous epithelial cells, relative to never smokers or non-obstructed smokers both in vitro^{103,104} and in situ.¹⁰⁵ Hence, the ultimate phenotype of any individual with COPD almost certainly results from one or more pathological processes that will require tailored interventions.

Rationale for a Revised Definition of COPD

Is Airway Narrowing or Air Flow Limitation Universal in Those With COPD Risk Factors?

Given this diversity of lung pathologies associated with smoking and spirometrically mild COPD, the question arises: do such exposures always induce airway narrowing or air flow limitation? The answer is a resounding no, as demonstrated in several groups of individuals with smoking histories. The first is those with emphysema despite normal spirometry. An analysis of the COPDGene cohort using visual reads according to Fleischner Society criteria identified some degree of emphysema in 44% of GOLD 0 participants.¹⁰⁶ In a 5-y follow-up study from a larger group in the same cohort, those who initially exhibited visually evident emphysema had significantly greater changes in multiple parameters characteristic of COPD, including FEV1/FVC.107 Nevertheless, these individuals would not currently be classified as having COPD based on spirometric criteria. In addition to radiographic detection of emphysema, considerable data support the detrimental effect of low values of diffusing capacity for carbon monoxide (D_{LCO}) , which is a risk factor for death independent of the degree of air flow limitation.^{108,109} The recent availability of reliable portable systems to measure both spirometry and $\dot{D}_{LCO}{}^{110}$ should permit broader use of this technique in cohort studies. These findings collectively argue strongly for including in the spectrum of COPD those with significant emphysema regardless of spirometry.

Additionally, in those with sufficient lifetime smoking history (10–20 or more pack-years), absence of air flow obstruction can still be associated with significant respiratory symptoms characteristic of COPD, including respiratory events compatible with exacerbations and empiric treatment using bronchodilators, as described in both the SPIROMICS¹¹⁰ and COPDGene¹¹² cohorts.

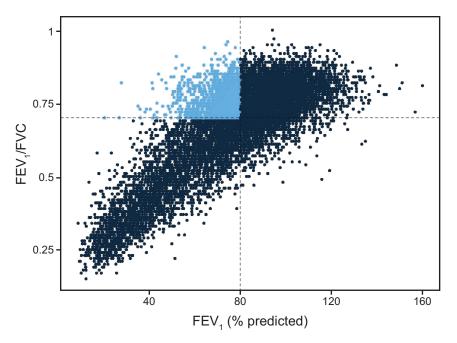


Fig. 3. Distribution of spirometry in the COPDGene cohort, which led to description of the preserved ratio but impaired spirometry (PRISm) phenotype. FEV₁% predicted is plotted on the x axis while FEV₁/FVC is plotted on the y axis. Dashed lines represent fixed-threshold criteria used to delineate PRISm individuals (highlighted in blue upper-left quadrant), individuals with normal lung function (upper-right quadrant), those with mild (lower-right quadrant), and moderate to severe COPD (lower-left quadrant). From Reference 112, with permission.

A third group currently excluded from the definition of COPD merits consideration: those with preserved ratio but impaired spirometry (PRISm)¹¹³ also termed restrictive spirometry.¹¹⁴ This group is typically defined as $FEV_1/FVC > 0.7$ and FEV₁% predicted < 80% (Fig. 3, upper left quadrant), although results are similar using lower limit of normal criteria. This graph also illustrates how rigid cutoffs based on spirometric values can misclassify; eg, some individuals with disproportionally high FVC values relative to their FEV_1 (also still in the normal range) will be diagnosed by GOLD criteria as having mild COPD (Fig. 3, lower right quadrant) rather than being normal variants. This issue is related strongly to age. Relative to the fixed-ratio definition, a cutoff based on lower limit of normal values for FEV₁/FVC diagnoses COPD less frequently in the elderly^{115,116} and more frequently in those ≤ 45 v old.¹¹⁵

PRISm was common at enrollment in the COPDGene cohort (12.3%); many in this category initially appeared to have a clinical phenotype similar to COPD, and many do transition over time to frank COPD, frequently at GOLD stage 2 or greater. Survival in those with PRISm was less than that of GOLD 0 subjects, that is, smokers without air flow obstruction.¹¹⁷ Congruent results were recently published from the Rotterdam cohort,¹¹⁸ which showed that the survival of a similarly defined PRISm group was much closer to that of patients with COPD on average and worse than that of GOLD 1 subjects. Some individuals with PRISm in a separate cohort also showed significant emphysema.¹¹⁹ Because the presence of significant interstitial lung abnormalities was exclusionary in most analysis of

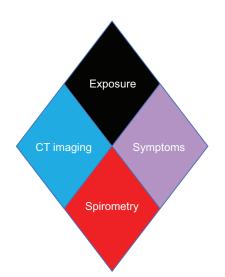


Fig. 4. Features used to define COPD in the COPD Genetic Epidemiology Study (COPDGene). Exposure in the COPDGene study includes individuals with a total of \geq 10 pack-years smoking. Computed tomography (CT) imaging includes individuals with quantitative assessment showing \geq 5% emphysema, a square root of airway wall area for a standardized airway of 10 mm internal diameter \geq 2.5 mm, or \geq 15% gas trapping. Symptoms include individuals with a Modified Medical Research Council dyspnea scale \geq 2 or chronic bronchitis. Spirometry includes individuals with FEV₁ < 80% predicted or FEV₁/FVC < 0.7. From Reference 2, with permission. CT = computed tomography.

PRISm, the risk of it progressing to frank interstitial lung disease remains uncertain to my knowledge and worthy of

Category	Odds of Change >350 mL in FEV ₁ (95% CI)ª	Hazard Ratio for All-Cause Mortality (95% CI) ^b	COPDGene 2019 Classification
А	1.0 (ref.)	1.0 (ref.)	No COPD
В	1.31 (1.04-1.65)	1.05 (0.76-1.44)	
С	1.42 (1.07-1.88)	1.55 (1.09-2.19)	Possible COPD
D	0.92 (0.64-1.30)	1.48 (1.03-2.12)	
E	1.74 (1.28-2.36)	1.90 (1.33-2.71)	
F	1.02 (0.66-1.60)	2.62 (1.84-3.72)	Probable COPD
G	2.11 (1.66-2.68)	1.76 (1.36-2.27)	
Н	2.82 (2.18-3.66)	5.18 (4.15-6.48)	Definite COPD

Fig. 5. Logistic regression and Cox regression models for FEV1 progression and all-cause mortality, respectively, with the proposed COPDGene 2019 classification of categories. (A) Change in FEV₁ assessment was done on n = 4,925 participants who returned for phase 2 clinical follow-up. Adjusted for age at first visit, sex, race, pack-years, and current smoking. Bolded numbers indicate categories where the 95% CI did not include 1.0. Symbols as in Figure 4. From Reference 2, with permission.

investigation. PRISm has always been recognized to be a highly heterogeneous group, but recent follow-up in COPDGene¹²⁰ indicates that it is a highly fluid category, with subsequent changes either to normal spirometry or obstruction. The usefulness of the PRISm category has been questioned,¹²¹ but it does appear to be a prevalent group at risk for poor outcomes and with largely unstudied responses to standard COPD treatments.

Importantly, all 3 of these common groups with lifetime smoking exposures (emphysema with little to no obstruction, symptomatic individuals with preserved spirometry, and PRISm) highlight the shortcomings of basing the diagnosis of COPD on spirometric cutoffs, whether fixed ratio or lower limit of normal based. Based on these groups who might fit under an expanded definition of the chronic lung conditions resulting from the multitude of oxidative inhalational exposures, it might be tempting to drop the "O" from the acronym. However, such a move would lose the name recognition that this long-neglected condition is finally gaining. It is recognized that the current spirometric diagnosis of COPD has considerable specificity, but low sensitivity, which has led to the proposal to recognize a broader group of exposed individuals as having pre-COPD.¹²²

The Dilemma of Redefining COPD

Thus, the current definition of COPD is increasingly recognized to be inadequate. The dilemma is how to replace it. An analysis of the COPDGene cohort attempted to capture COPD heterogeneity using spirometry and combinations of CT imaging and symptoms. Based on logistic and Cox regression models of actual outcomes, the classification proposes a range of categories, including possible and probable COPD (Figs. 4, 5).² Efforts are already underway to update and expand this classification.

However, one can imagine devising very different definitions, depending on whether the goal is to define homogeneous endotypic groups for clinical trials versus an easy-toapply algorithm to aid disease management in primary care. Similar tweaks would be needed for disability determination and potentially other uses. Hence, an all-encompassing redefinition appears elusive in the present state of pathological understanding. An alternative viewpoint is to cease considering COPD a unitary disease and instead accept it as a clinical syndrome.^{4,123,124} In the meanwhile, every effort should be made to continue to dissect the heterogeneity of this syndrome to identify true biological mechanism–defined endotypes that will be susceptible to precision therapies.

Summary

Understanding of the biological and molecular pathways leading to lung damage in COPD has increased greatly in the last decade, but there is still much to be learned. Research is urgently needed on the pathological correlates of inhalational exposures other than tobacco smoking that lead to impaired lung function and respiratory symptoms. Research on smokers should embrace the diversity of lifetime lung function trajectories and the heterogeneity of COPD by focusing on subgroups with chronologically early disease and those at increased risk of rapid loss of lung function. Advanced thoracic imaging¹²⁵ and the combination of genetic and multiomics risk scores^{126,127} promise to be the most efficient means to define targetable pathological pathways for personalized therapies. Finally, the current purely spirometric definition of COPD is inadequate, but revisions will likely require different definitions for disparate use cases.

REFERENCES

- Celli B, Fabbri L, Criner G, Martinez FJ, Mannino D, Vogelmeier C, et al. Definition and nomenclature of chronic obstructive pulmonary disease: time for its revision. Am J Respir Crit Care Med 2022;206(11):1317-1325.
- Lowe KE, Regan EA, Anzueto A, Austin E, Austin JHM, Beaty TH, et al. COPDGene 2019: redefining the diagnosis of chronic obstructive pulmonary disease. Chronic Obstr Pulm Dis 2019;6(5):384-399.
- Barnes PJ, Vestbo J, Calverley PM. The pressing need to redefine "COPD." Chronic Obstr Pulm Dis 2019;6(5):380-383.
- Agustí A, Melén E, DeMeo DL, Breyer-Kohansal R, Faner R. Pathogenesis of chronic obstructive pulmonary disease: understanding the contributions of gene-environment interactions across the life span. Lancet Respir Med 2022;10(5):512-524.
- Baugh AD, Shiboski S, Hansel NN, Ortega V, Barjaktarevic I, Barr RG, et al. Reconsidering the utility of race-specific lung function prediction equations. Am J Respir Crit Care Med 2022;205(7):819-829.
- Regan EA, Hokanson JE, Murphy JR, Make B, Lynch DA, Beaty TH, et al. Genetic epidemiology of COPD (COPDGene) study design. COPD 2010;7(1):32-43.
- Couper D, LaVange LM, Han M, Barr RG, Bleecker E, Hoffman EA, et al; SPIROMICS Research Group. Design of the subpopulations and intermediate outcomes in COPD Study (SPIROMICS). Thorax 2014;69(5):491-494.
- Han MK, Agusti A, Calverley PM, Celli BR, Criner G, Curtis JL, et al. Chronic obstructive pulmonary disease phenotypes: the future of COPD. Am J Respir Crit Care Med 2010;182(5):598-604.
- Castaldi PJ, Benet M, Petersen H, Rafaels N, Finigan J, Paoletti M, et al. Do COPD subtypes really exist? COPD heterogeneity and clustering in 10 independent cohorts. Thorax 2017;72(11):998-1006.
- Curtis JL, Freeman CM, Hogg JC. The immunopathogenesis of chronic obstructive pulmonary disease: insights from recent research. Proc Am Thorac Soc 2007;4(7):512-521.
- Hogg JC, Timens W. The pathology of chronic obstructive pulmonary disease. Annu Rev Pathol 2009;4:435-459.
- Tuder RM, Petrache I. Pathogenesis of chronic obstructive pulmonary disease. J Clin Invest 2012;122(8):2749-2755.
- Agustí A, Hogg JC. Update on the pathogenesis of chronic obstructive pulmonary disease. N Engl J Med 2019;381(13):1248-1256.
- 14. Labonte LE, Tan WC, Li PZ, Mancino P, Aaron SD, Benedetti A, et al; CanCOLD Collaborative Research Group. Undiagnosed chronic obstructive pulmonary disease contributes to the burden of health care use. Data from the CanCOLD Study. Am J Respir Crit Care Med 2016;194(3):285-298.
- Ho T, Cusack RP, Chaudhary N, Satia I, Kurmi OP. Under- and overdiagnosis of COPD: a global perspective. Breathe (Sheff) 2019;15 (1):24-35.
- Kohansal R, Martinez-Camblor P, Agusti A, Buist AS, Mannino DM, Soriano JB. The natural history of chronic airflow obstruction

revisited: an analysis of the Framingham offspring cohort. Am J Respir Crit Care Med 2009;180(1):3-10.

- Fletcher C, Peto R. The natural history of chronic airflow obstruction. Br Med J 1977;1(6077):1645-1648.
- Lange P, Celli B, Agusti A, Boje Jensen G, Divo M, Faner R, et al. Lung-function trajectories leading to chronic obstructive pulmonary disease. N Engl J Med 2015;373(2):111-122.
- Allinson JP, Hardy R, Donaldson GC, Shaheen SO, Kuh D, Wedzicha JA. Combined impact of smoking and early life exposures on adult lung function trajectories. Am J Respir Crit Care Med 2017;196(8):1021-1030.
- Rennard SI, Drummond MB. Early chronic obstructive pulmonary disease: definition, assessment, and prevention. Lancet 2015;385 (9979):1778-1788.
- Martinez FD. Early-life origins of chronic obstructive pulmonary disease. N Engl J Med 2016;375(9):871-878.
- 22. John C, Soler Artigas M, Hui J, Nielsen SF, Rafaels N, Pare PD, et al. Genetic variants affecting cross-sectional lung function in adults show little or no effect on longitudinal lung function decline. Thorax 2017;72(5):400-408.
- Yang IA, Jenkins CR, Salvi SS. Chronic obstructive pulmonary disease in never smokers: risk factors, pathogenesis, and implications for prevention and treatment. Lancet Respir Med 2022;10(5):497-511.
- Manisalidis I, Stavropoulou E, Stavropoulos A, Bezirtzoglou E. Environmental and health impacts of air pollution: a review. Front Public Health 2020;8:14.
- Ejike CO, Woo H, Galiatsatos P, Paulin LM, Krishnan JA, Cooper CB, et al. Contribution of individual and neighborhood factors to racial disparities in respiratory outcomes. Am J Respir Crit Care Med 2021;203 (8):987-997.
- 26. Woo H, Brigham EP, Allbright K, Ejike C, Galiatsatos P, Jones MR, et al. Racial segregation and respiratory outcomes among urban black residents with and at risk of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2021;204(5):536-545.
- Raju S, Keet CA, Paulin LM, Matsui EC, Peng RD, Hansel NN, et al. Rural residence and poverty are independent risk factors for chronic obstructive pulmonary disease in the United States. Am J Respir Crit Care Med 2019;199(8):961-969.
- Hansel NN, McCormack MC, Kim V. The effects of air pollution and temperature on COPD. COPD 2016;13(3):372-379.
- 29. Thilakaratne R, Hoshiko S, Rosenberg A, Hayashi T, Buckman JR, Rappold AG. Wildfires and the changing landscape of air pollution– related health burden in California. Am J Respir Crit Care Med 2023 in press, 207(7):887-898.
- Schweitzer MD, Calzadilla AS, Salamo O, Sharifi A, Kumar N, Holt G, et al. Lung health in era of climate change and dust storms. Environ Res 2018;163:36-42.
- Terzikhan N, Verhamme KM, Hofman A, Stricker BH, Brusselle GG, Lahousse L. Prevalence and incidence of COPD in smokers and nonsmokers: the Rotterdam Study. Eur J Epidemiol 2016;31(8):785-792.
- 32. Paulin LM, Gassett AJ, Alexis NE, Kirwa K, Kanner RE, Peters S, et al; for SPIROMICS investigators. Association of long-term ambient ozone exposure with respiratory morbidity in smokers. JAMA Intern Med 2020;180(1):106-115.
- 33. Paulin LM, Diette GB, Blanc PD, Putcha N, Eisner MD, Kanner RE, et al; SPIROMICS Research Group. Occupational exposures are associated with worse morbidity in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2015;191(5):557-565.
- 34. Weibel ER. Morphometry of the human lung. New York: Academic Press; 1963.
- Horsfield K, Cumming G. Morphology of the bronchial tree in man. J Appl Physiol 1968;24(3):373-383.
- Hogg JC, Pare PD, Hackett TL. The contribution of small airway obstruction to the pathogenesis of chronic obstructive pulmonary disease. Physiol Rev 2017;97(2):529-552.

- Hogg JC, Macklem PT, Thurlbeck WM. Site and nature of airway obstruction in chronic obstructive lung disease. N Engl J Med 1968;278(25):1355-1360.
- Koo HK, Vasilescu DM, Booth S, Hsieh A, Katsamenis OL, Fishbane N, et al. Small airways disease in mild and moderate chronic obstructive pulmonary disease: a cross-sectional study. Lancet Respir Med 2018;6(8):591-602.
- 39. Knox-Brown B, Patel J, Potts J, Ahmed R, Aquart-Stewart A, Cherkaski HH, et al; BOLD Collaborative Research Group. Small airways obstruction and its risk factors in the Burden of Obstructive Lung Disease (BOLD) study: a multinational cross-sectional study. Lancet Glob Health 2023;11(1):e69-e82.
- Martinez FJ, Han MK, Allinson JP, Barr RG, Boucher RC, Calverley P, et al. At the root: defining and halting progression of early chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2018;197 (12):1540-1551.
- Amatngalim GD, Schrumpf JA, Dishchekenian F, Mertens TCJ, Ninaber DK, van der Linden AC, et al. Aberrant epithelial differentiation by cigarette smoke dysregulates respiratory host defense. Eur Respir J 2018;51(4):1701009.
- 42. Shaykhiev R, Crystal RG. Early events in the pathogenesis of chronic obstructive pulmonary disease. Smoking-induced reprogramming of airway epithelial basal progenitor cells. Ann Am Thorac Soc 2014;11 Suppl 5(Suppl 5):S252-S258.
- Yang J, Zuo WL, Fukui T, Chao I, Gomi K, Lee B, et al. Smokingdependent distal-to-proximal repatterning of the adult human small airway epithelium. Am J Respir Crit Care Med 2017;196(3):340-352.
- Crystal RG. Airway basal cells. The "smoking gun" of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2014;190(12):1355-1362.
- 45. Kesimer M, Ford AA, Ceppe A, Radicioni G, Cao R, Davis CW, et al. Airway mucin concentration as a marker of chronic bronchitis. N Engl J Med 2017;377(10):911-922.
- Laucho-Contreras ME, Polverino F, Gupta K, Taylor KL, Kelly E, Pinto-Plata V, et al. Protective role for club cell secretory protein-16 (CC16) in the development of COPD. Eur Respir J 2015;45(6):1544-1556.
- 47. Polosukhin VV, Cates JM, Lawson WE, Zaynagetdinov R, Milstone AP, Massion PP, et al. Bronchial secretory immunoglobulin a deficiency correlates with airway inflammation and progression of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2011;184(3):317-327.
- 48. Gohy ST, Detry BR, Lecocq M, Bouzin C, Weynand BA, Amatngalim GD, et al. Polymeric immunoglobulin receptor downregulation in chronic obstructive pulmonary disease. Persistence in the cultured epithelium and role of transforming growth factor-beta. Am J Respir Crit Care Med 2014;190(5):509-521.
- 49. Polosukhin VV, Richmond BW, Du RH, Cates JM, Wu P, Nian H, et al. Secretory IgA deficiency in individual small airways is associated with persistent inflammation and remodeling. Am J Respir Crit Care Med 2017;195(8):1010-1021.
- Yagi K, Huffnagle GB, Lukacs NW, Asai N. The lung microbiome during health and disease. IJMS 2021;22(19):10872.
- Curtis JL. A hairline crack in the levee: focal secretory IgA deficiency as a first step toward emphysema. Am J Respir Crit Care Med 2017;195(8):970-973.
- Berenson CS, Kruzel RL, Eberhardt E, Sethi S. Phagocytic dysfunction of human alveolar macrophages and severity of chronic obstructive pulmonary disease. J Infect Dis 2013;208(12):2036-2045.
- 53. Bewley MA, Preston JA, Mohasin M, Marriott HM, Budd RC, Swales J, et al. Impaired mitochondrial microbicidal responses in chronic obstructive pulmonary disease macrophages. Am J Respir Crit Care Med 2017;196(7):845-855.
- Bewley MA, Budd RC, Ryan E, Cole J, Collini P, Marshall J, et al; COPDMAP. Opsonic phagocytosis in chronic obstructive pulmonary disease is enhanced by Nrf2 agonists. Am J Respir Crit Care Med 2018;198(6):739-750.

- Morales-Nebreda L, Misharin AV, Perlman H, Budinger GR. The heterogeneity of lung macrophages in the susceptibility to disease. Eur Respir Rev 2015;24(137):505-509.
- 56. Erb-Downward JR, Thompson DL, Han MK, Freeman CM, McCloskey L, Schmidt LA, et al. Analysis of the lung microbiome in the "healthy" smoker and in COPD. PLoS One 2011;6(2):e16384.
- 57. Dickson RP, Erb-Downward JR, Freeman CM, McCloskey L, Beck JM, Huffnagle GB, et al. Spatial variation in the healthy human lung microbiome and the adapted island model of lung biogeography. Ann Am Thorac Soc 2015;12(6):821-830.
- Dickson RP, Erb-Downward JR, Freeman CM, McCloskey L, Falkowski NR, Huffnagle GB, et al. Bacterial topography of the healthy human lower respiratory tract. mBio 2017;8(1):2150-7511.
- Smith BM, Traboulsi H, Austin JHM, Manichaikul A, Hoffman EA, Bleecker ER, et al. Human airway branch variation and chronic obstructive pulmonary disease. Proc Natl Acad Sci U S A 2018;115(5):E974-E981.
- 60. Smith BM, Kirby M, Hoffman EA, Kronmal RA, Aaron SD, Allen NB, et al; MESA Lung, CanCOLD, and SPIROMICS Investigators. Association of dysanapsis with chronic obstructive pulmonary disease among older adults. JAMA 2020;323(22):2268-2280.
- Segal LN, Clemente JC, Tsay JC, Koralov SB, Keller BC, Wu BG, et al. Enrichment of the lung microbiome with oral taxa is associated with lung inflammation of a Th17 phenotype. Nat Microbiol 2016;1: 16031. doi: 10.1038/nmicrobiol.2016.31.
- 62. Opron K, Begley LA, Erb-Downward JR, Freeman C, Madapoosi S, Alexis NE, et al. Lung microbiota associations with clinical features of COPD in the SPIROMICS cohort. NPJ Biofilms Microbiomes 2021;7(1):14. doi: 10.1038/s41522-021-00185-9.
- 63. Madapoosi SS, Cruickshank-Quinn C, Opron K, Erb-Downward JR, Begley LA, Li G, et al; SPIROMICS Research Group. Lung microbiota and metabolites collectively associate with clinical outcomes in milder stage chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2022;206(4):427-439.
- 64. Huang YJ, Erb-Downward JR, Dickson RP, Curtis JL, Huffnagle GB, Han MK. Understanding the role of the microbiome in chronic obstructive pulmonary disease: principles, challenges, and future directions. Transl Res 2017;179:71-83.
- Whiteside SA, McGinniss JE, Collman RG. The lung microbiome: progress and promise. J Clin Invest 2021;131(15).
- 66. Xu F, Vasilescu DM, Kinose D, Tanabe N, Ng KW, Coxson HO, et al. The molecular and cellular mechanisms associated with the destruction of terminal bronchioles in COPD. Eur Respir J 2022;59(5):2101411.
- Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, et al. Robust enumeration of cell subsets from tissue expression profiles. Nat Methods 2015;12(5):453-457.
- 68. Grumelli S, Corry DB, Song LZ, Song L, Green L, Huh J, et al. An immune basis for lung parenchymal destruction in chronic obstructive pulmonary disease and emphysema. PLOS Med 2004;1(1):e8.
- Lee SH, Goswami S, Grudo A, Song LZ, Bandi V, Goodnight-White S, et al. Anti-elastin autoimmunity in tobacco smoking–induced emphysema. Nat Med 2007;13(5):567-569.
- Shan M, Cheng HF, Song LZ, Roberts L, Green L, Hacken-Bitar J, et al. Lung myeloid dendritic cells coordinately induce T_H1 and T_H17 responses in human emphysema. Sci Transl Med 2009;1(4):4ra10.
- Brusselle GG, Joos GF, Bracke KR. New insights into the immunology of chronic obstructive pulmonary disease. Lancet 2011;378(9795):1015-1026.
- Faner R, Cruz T, Casserras T, Lopez-Giraldo A, Noell G, Coca I, et al. Network analysis of lung transcriptomics reveals a distinct B-cell signature in emphysema. Am J Respir Crit Care Med 2016;193(11):1242-1253.
- 73. Obeidat M, Nie Y, Fishbane N, Li X, Bosse Y, Joubert P, et al. Integrative genomics of emphysema-associated genes reveals potential disease biomarkers. Am J Respir Cell Mol Biol 2017;57(4):411-418.

- Brandsma CA, Timens W, Geerlings M, Jekel H, Postma DS, Hylkema MN, et al. Induction of autoantibodies against lung matrix proteins and smoke-induced inflammation in mice. BMC Pulm Med 2010;10:64.
- John-Schuster G, Hager K, Conlon TM, Irmler M, Beckers J, Eickelberg O, et al. Cigarette smoke-induced iBALT mediates macrophage activation in a B cell–dependent manner in COPD. Am J Physiol Lung Cell Mol Physiol 2014;307(9):L692-706.
- Hu JY, Liu BB, Du YP, Zhang Y, Zhang YW, Zhang YY, et al. Increased circulating beta2-adrenergic receptor autoantibodies are associated with smoking-related emphysema. Sci Rep 2017;7:43962.
- Gadgil A, Zhu X, Sciurba FC, Duncan SR. Altered T-cell phenotypes in chronic obstructive pulmonary disease. Proc Am Thorac Soc 2006;3(6):487-488.
- Feghali-Bostwick CA, Gadgil AS, Otterbein LE, Pilewski JM, Stoner MW, Csizmadia E, et al. Autoantibodies in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2008;177(2):156-163.
- Cosio MG, Saetta M, Agusti A. Immunologic aspects of chronic obstructive pulmonary disease. N Engl J Med 2009;360(23):2445-2454.
- Leidinger P, Keller A, Heisel S, Ludwig N, Rheinheimer S, Klein V, et al. Novel autoantigens immunogenic in COPD patients. Respir Res 2009;10(1):20.
- Karayama M, Inui N, Suda T, Nakamura Y, Nakamura H, Chida K. Anti-endothelial cell antibodies in patients With COPD. Chest 2010;138(6):1303-1308.
- Kuo YB, Chang CA, Wu YK, Hsieh MJ, Tsai CH, Chen KT, et al. Identification and clinical association of anti-cytokeratin 18 autoantibody in COPD. Immunol Lett 2010;128(2):131-136.
- 83. Xiong Y, Gao S, Luo G, Cheng G, Huang W, Jiang R, et al. Increased circulating autoantibodies levels of IgG, IgA, IgM against cytokeratin 18 and cytokeratin 19 in chronic obstructive pulmonary disease. Arch Med Res 2017;48(1):79-87.
- Hogg JC, Chu F, Utokaparch S, Woods R, Elliott WM, Buzatu L, et al. The nature of small airway obstruction in chronic obstructive pulmonary disease. N Engl J Med 2004;350(26):2645-2653.
- Ruddle NH. Lymphatic vessels and tertiary lymphoid organs. J Clin Invest 2014;124(3):953-959.
- Baraldo S, Turato G, Lunardi F, Bazzan E, Schiavon M, Ferrarotti I, et al. Immune activation in alpha1-antitrypsin-deficiency emphysema. Beyond the protease-antiprotease paradigm. Am J Respir Crit Care Med 2015;191(4):402-409.
- Galbàn CJ, Han MK, Boes JL, Chughtai KA, Meyer CR, Johnson TD, et al. Computed tomography–based biomarker provides unique signature for diagnosis of COPD phenotypes and disease progression. Nat Med 2012;18(11):1711-1715.
- Kirby M, Yin Y, Tschirren J, Tan WC, Leipsic J, Hague CJ, et al; CanCOLD Collaborative Research Group and the Canadian Respiratory Research Network. A novel method of estimating small airway disease using inspiratory-to-expiratory computed tomography. Respiration 2017;94(4):336-345.
- 89. Ostridge K, Gove K, Paas KHW, Burke H, Freeman A, Harden S, et al. Using novel computed tomography analysis to describe the contribution and distribution of emphysema and small airways disease in chronic obstructive pulmonary disease. Ann Am Thorac Soc 2019;16(8):990-997.
- Vasilescu DM, Martinez FJ, Marchetti D, Galbán CJ, Hatt C, Meldrum CA, et al. Noninvasive imaging biomarker identifies small airway damage in severe COPD. Am J Respir Crit Care Med 2019;200(5):575-581.
- Labaki WW, Gu T, Murray S, Hatt CR, Galban CJ, Ross BD, et al. Voxel-wise longitudinal parametric response mapping analysis of chest computed tomography in smokers. Acad Radiol 2019;26(2):217-223.
- 92. Edgar RG, Patel M, Bayliss S, Crossley D, Sapey E, Turner AM. Treatment of lung disease in alpha-1 antitrypsin deficiency: a systematic review. Int J Chron Obstruct Pulmon Dis 2017;12:1295-1308.
- 93. Davis CW, Lopez CL, Bell AJ, Miller RF, Rabin AS, Murray S, et al. The severity of functional small airways disease in military personnel

with constrictive bronchiolitis as measured by quantitative CT. Am J Respir Crit Care Med 2022;206:786-789.

- Majo J, Ghezzo H, Cosio MG. Lymphocyte population and apoptosis in the lungs of smokers and their relation to emphysema. Eur Respir J 2001;17(5):946-953.
- Kasahara Y, Tuder RM, Taraseviciene-Stewart L, Le Cras TD, Abman S, Hirth PK, et al. Inhibition of VEGF receptors causes lung cell apoptosis and emphysema. J Clin Invest 2000;106(11):1311-1319.
- 96. Kasahara Y, Tuder RM, Cool CD, Lynch DA, Flores SC, Voelkel NF. Endothelial cell death and decreased expression of vascular endothelial growth factor and vascular endothelial growth factor receptor 2 in emphysema. Am J Respir Crit Care Med 2001;163(3 Pt 1):737-744.
- Diab KJ, Adamowicz JJ, Kamocki K, Rush NI, Garrison J, Gu Y, et al. Stimulation of sphingosine 1-phosphate signaling as an alveolar cell survival strategy in emphysema. Am J Respir Crit Care Med 2010;181 (4):344-352.
- Schweitzer KS, Hatoum H, Brown MB, Gupta M, Justice MJ, Beteck B, et al. Mechanisms of lung endothelial barrier disruption induced by cigarette smoke: role of oxidative stress and ceramides. Am J Physiol Lung Cell Mol Physiol 2011;301(6):L836-L846.
- 99. Summers ME, Richmond BW, Kropski JA, Majka SA, Bastarache JA, Hatzopoulos AK, et al. Balanced Wnt/Dickkopf-1 signaling by mesenchymal vascular progenitor cells in the microvascular niche maintains distal lung structure and function. Am J Physiol Cell Physiol 2021;320(1):C119-C131.
- Hisata S, Racanelli AC, Kermani P, Schreiner R, Houghton S, Palikuqi B, et al. Reversal of emphysema by restoration of pulmonary endothelial cells. J Exp Med 2021;218(8):e20200938.
- 101. Hueper K, Vogel-Claussen J, Parikh MA, Austin JH, Bluemke DA, Carr J, et al. Pulmonary microvascular blood flow in mild chronic obstructive pulmonary disease and emphysema. The MESA COPD Study. Am J Respir Crit Care Med 2015;192(5):570-580.
- 102. Segura-Valdez L, Pardo A, Gaxiola M, Uhal BD, Becerril C, Selman M. Upregulation of gelatinases A and B, collagenases 1 and 2, and increased parenchymal cell death in COPD. Chest 2000;117(3):684-694.
- 103. Freeman CM, Stolberg VR, Crudgington S, Martinez FJ, Han MK, Chensue SW, et al. Human CD56+ cytotoxic lung lymphocytes kill autologous lung cells in chronic obstructive pulmonary disease. PLoS One 2014;9(7):e103840.
- 104. Finch DK, Stolberg VR, Ferguson J, Alikaj H, Kady MR, Richmond BW, et al. Lung dendritic cells drive NK cytotoxicity in chronic obstructive pulmonary disease via IL-15Ralpha. Am J Respir Crit Care Med 2018;198(9):1140-1150.
- 105. Pallazola AM, Rao JX, Mengistu DT, Morcos MS, Toma MS, Stolberg VR, et al. Human lung cDC1 drive increased perforin-mediated NK cytotoxicity in chronic obstructive pulmonary disease. Am J Physiol Lung Cell Mol Physiol 2021;321(6):L1183-11193.
- 106. Oh AS, Strand M, Pratte K, Regan EA, Humphries S, Crapo JD, et al; Genetic Epidemiology of COPDGene Investigators. Visual emphysema at chest CT in GOLD stage 0 cigarette smokers predicts disease progression: results from the COPDGene Study. Radiology 2020;296(3):641-649.
- 107. Lynch DA, Moore CM, Wilson C, Nevrekar D, Jennermann T, Humphries SM, et al; Genetic Epidemiology of COPD (COPDGene) Investigators. CT-based visual classification of emphysema: association with mortality in the COPDGene Study. Radiology 2018;288(3):859-866.
- Boutou AK, Shrikrishna D, Tanner RJ, Smith C, Kelly JL, Ward SP, et al. Lung function indices for predicting mortality in COPD. Eur Respir J 2013;42(3):616-625.
- 109. de-Torres JP, O'Donnell DE, Marín JM, Cabrera C, Casanova C, Marín M, et al. Clinical and prognostic impact of low diffusing capacity for carbon monoxide values in patients with global initiative for obstructive lung disease I COPD. Chest 2021;160(3):872-878.

- 110. Gochicoa-Rangel L, Pérez-Padilla R, Vázquez-García JC, Silva-Cerón M, Cid-Juárez S, Martínez-Briseño D, et al. Long-term stability of a portable carbon monoxide single-breath diffusing capacity instrument. Respir Care 2017;62(2):231-235.
- 111. Woodruff PG, Barr RG, Bleecker E, Christenson SA, Couper D, Curtis JL, et al; SPIROMICS Research Group. Clinical significance of symptoms in smokers with preserved pulmonary function. N Engl J Med 2016;374(19):1811-1821.
- 112. Regan EA, Lynch DA, Curran-Everett D, Curtis JL, Austin JH, Grenier PA, et al; Genetic Epidemiology of COPD (COPDGene) Investigators. Clinical and radiologic disease in smokers with normal spirometry. JAMA Intern Med 2015;175(9):1539-1549.
- 113. Wan ES, Castaldi PJ, Cho MH, Hokanson JE, Regan EA, Make BJ, et al; COPDGene Investigators. Epidemiology, genetics, and subtyping of preserved ratio impaired spirometry (PRISm) in COPDGene. Respir Res 2014;15(1):89-102.
- 114. Backman H, Eriksson B, Hedman L, Stridsman C, Jansson SA, Sovijärvi A, et al. Restrictive spirometric pattern in the general adult population: Methods of defining the condition and consequences on prevalence. Respir Med 2016;120:116-123.
- 115. Güder G, Brenner S, Angermann CE, Ertl G, Held M, Sachs AP, et al. "GOLD or lower limit of normal definition? A comparison with expert-based diagnosis of chronic obstructive pulmonary disease in a prospective cohort-study." Respir Res 2012;13(1):13.
- 116. van Dijk W, Tan W, Li P, Guo B, Li S, Benedetti A, et al; CanCOLD Study Group. Clinical relevance of fixed ratio vs lower limit of normal of FEV1/FVC in COPD: patient-reported outcomes from the CanCOLD cohort. Ann Fam Med 2015;13(1):41-48.
- 117. Wan ES, Fortis S, Regan EA, Hokanson J, Han MK, Casaburi R, et al; COPDGene Investigators. Longitudinal phenotypes and mortality in preserved ratio impaired spirometry in the COPDGene Study. Am J Respir Crit Care Med 2018;198(11):1397-1405.

Discussion

MacIntyre: I thought that was a wonderful overview. I find it fascinating that in 2022 we are, for the most part, still defining COPD by spirometry, a test first described in the mid-19th century, and we are rigidly adhering to that definition despite this enormous amount of literature showing us that the FEV_1 is missing an awful lot of chronic small airways inflammation and emphysema. I think this is of particular interest to our pharma friends who manufacture and market bronchodilators, drugs of only limited value in small airway disease and of no value in emphysema. We need a whole new array of therapeutics aimed at these different phenotypes. That wasn't really a question, more of a comment. I also find it interesting, as long as we're discussing FEV_1 , that the FEV₁ still remains in the FDA as the standard for approving drugs for COPD. And again, as Jeff has shown us very nicely, there's an awful lot of COPD out there that is not bronchodilator responsive. Any other thoughts?

Haynes: I would extend that to say that not only do we have this love affair with FEV_1 for diagnosis but also determining whether therapy works. There are a lot of data that show that bronchodilators do work even if it's not reflected in the FEV_1 .¹

Mike Hess: I think that really hits it on the head, the love affair with FEV_1 . And I say this as a fellow registered pulmonary function technologist, not only is it an old test, it can be inaccurate; it is highly dependent on technique and coaching. When I've spoken with a few different clinicians who are relatively new to the COPD world they said, "Well what do we do, then?"; and I say, "this is the best we've got. We don't have a blood marker; we don't have a lab test. We can look at some of the imaging and use that, but we are

- 118. Wijnant SRA, De Roos E, Kavousi M, Stricker BH, Terzikhan N, Lahousse L, et al. Trajectory and mortality of preserved ratio impaired spirometry: the Rotterdam Study. Eur Respir J 2020;55(1):1901217.
- 119. Wei X, Ding Q, Yu N, Mi J, Ren J, Li J, et al. Imaging features of chronic bronchitis with preserved ratio and impaired spirometry (PRISm). Lung 2018;196(6):649-658.
- 120. Wan ES, Hokanson JE, Regan EA, Young KA, Make BJ, DeMeo DL, et al. Significant spirometric transitions and preserved ratio impaired spirometry among ever smokers. Chest 2022;161(3):651-661.
- 121. Knox-Brown B, Amaral AF, Burney P. Concerns about PRISm. Lancet Respir Med 2022;10(6):e51-e52.
- 122. Han MK, Agusti A, Celli BR, Criner GJ, Halpin DMG, Roche N, et al. From GOLD 0 to pre-COPD. Am J Respir Crit Care Med 2021;203(4):414-423.
- 123. Agusti A, Bel E, Thomas M, Vogelmeier C, Brusselle G, Holgate S, et al. Treatable traits: toward precision medicine of chronic airway diseases. Eur Respir J 2016;47(2):410-419.
- 124. Sterk PJ. Chronic diseases like asthma and COPD: do they truly exist? Eur Respir J 2016;47(2):359-361.
- 125. Labaki WW, Martinez CH, Martinez FJ, Galban CJ, Ross BD, Washko GR, et al. The role of chest computed tomography in the evaluation and management of the patient with COPD. Am J Respir Crit Care Med 2017;196(11):1372-1379.
- 126. Moll M, Sakornsakolpat P, Shrine N, Hobbs BD, DeMeo DL, John C, et al; SpiroMeta Consortium. Chronic obstructive pulmonary disease and related phenotypes: polygenic risk scores in population-based and case-control cohorts. Lancet Respir Med 2020;8(7):696-708.
- 127. Moll M, Boueiz A, Ghosh AJ, Saferali A, Lee S, Xu Z, et al; HAPIN Investigators. Development of a blood-based transcriptional risk score for chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2022;205(2):161-170.

legally and regulatorily bound by this FEV_1 idea." I definitely think it's time for an expanded definition. It also reminds me that everything old is new again because in some reading I've done on chronic nonspecific lung disease back in the 1960s Dutch definition, are we going back to that and then developing phenotypes?

Carlin: I agree with everyone else here in the room that I think the definition certainly needs to be changed, but how can we make it simple enough for a care provider to make an accurate diagnosis given the extreme limitations of time for the patient-physician interactions? I liken it back to what Tom Petty MD as part of the National Lung Health Education Program (which he founded) said years ago, that in order to diagnose an illness some type of "numerical" foundation should be made. For example, no one would diagnose hypertension without a blood pressure measurement or diagnose hyperlipidemia without a lipid or cholesterol measurement. While spirometry may not be the absolute best way to diagnose COPD, it still stands as a cornerstone of the diagnosis.

Haynes: I would suggest that artificial intelligence (AI) is the way we get there. Expecting a primary care physician to calculate all this and consider all of these factors in the limited time they have is unrealistic, but we certainly have AI, and it would be easy to write software to *help* make that diagnosis.

MacIntyre: I assume we'll hear more about this later on with COPD assessment discussions, but it's obviously terribly impractical to require sophisticated imaging (eg, CT scanning) to detect emphysema. We need something simpler. Some things that come to mind is using AI or something like that to analyze the flows as the lung gets smaller and the airways get smaller. The FEF₂₅₋₇₅ is way too crude, but looking at the shape of the curve could yield valuable information. This is the so-called silent area of the spirometry tracing. Impulse oscillometry is perhaps another way of delving into the small airways in the lung. And then I'll put in a plug for one of my personal favorites, D_{LCO}. We think of diffusion capacity as a simple measurement of gas transport across the alveolar capillary membrane. However, ventilation distribution, intrapulmonary gas mixing, and the properties of the alveolar capillary membrane might all be things impacting D_{LCO} we could use to look for emphysema. Indeed, one of my favorite projects in COPDGene is looking at the relationship of D_{LCO} and CT to determine emphysema. There's clearly a relationship there; the r value is not huge, but it's

real. So there may be tools that are a little simpler than going into the CT scanner to get at these small airway abnormalities and emphysema.

Orr: I'd like to make a quick comment on PFTs and diagnosis and share a little concern about liberalizing the diagnostic criteria. What I tend to see clinically is that patients come to me without spirometry and they're already on triple therapy. And I think that as long as the PFT results are a reflex to bronchodilators, and that dyspnea symptoms that are uncontrolled are a reflex to more bronchodilators, we're in a difficult situation. I think there's a lot of medication overuse for things that aren't really responsive to these bronchodilator medications.

Mike Hess: That also crossed my mind. We'll have to thread that needle. We already see people where, "ok, you coughed, you smoked 20 years ago, ergo you have COPD," and now you're tagged with this forever, and you're going to be on this particular medication regimen, and that's it. There is, I think, a danger in oversimplifying.

Criner: I think one of the things, as Brian [Carlin] mentioned, it's not only the primary care physicians but the majority of patients with COPD, and it's probably going to increase in people who are in low- or middle-income countries. And you know, there are more radiologists on Longwood Avenue in Boston than there are on the whole continent of Africa. So some of these tools that we use like CT scans or other elite measurements are useful in characterizing the extent of the disease but are not practical for the clinical diagnosis or to

be used in definitions. I think one of the challenges is to be able, with an increasing worldwide prevalence of chronic obstructive lung diseases related to environmental changes and changes in the climate, is to be able to come up with a diagnosis that's established on physiological and clinical principles that are tied to mechanistic things that broaden the diagnostic abilities that we have and be able to triage patients to receive care based on that. Some of the papers that Jeff went through were SPIROMICS papers by Prescott Woodruff.^{2,3} Those subjects were treated because they were symptomatic. And for the most part, in any clinical trial, the clinicians get it right. So, there's much more to be gained by milking the clinical history and physical examination than we currently use, and I think we can do a better job of trying to hone those skills.

MacIntyre: I think this is a great beginning. These comments and questions are likely going to keep coming up over the next day and a half, and we'll be referring back to this first talk.

REFERENCES

- O'Donnell DE, Forkert L, Webb KA. Evaluation of bronchodilator responses in patients with "irreversible" emphysema. Eur Respir J 2001;18(6):914-920.
- Yee N, Markovic D, Buhr RG, Fortis S, Arjomandi M, Couper D, et al. Significance of FEV₃/FEV₆ in recognition of early airway disease in smokers at risk of development of COPD: analysis of the SPIROMICS cohort. Chest 2022;161(4):949-959.
- Woodruff PG, Barr RG, Bleecker E, Christenson SA, Couper D, Curtis JL, et al; SPIROMICS Research Group. Clinical significance of symptoms in smokers with preserved pulmonary function. N Engl J Med 2016;374(19):1811-1821.

This article is approved for Continuing Respiratory Care Education credit. For information and to obtain your CRCE (free to AARC members) visit www.rcjournal.com

