

Encephalitis

Review Article

pISSN 2765-4559 eISSN 2734-1461 encephalitis | Vol. 1, No. 2 | March 25, 2021 https://doi.org/10.47936/encephalitis.2021.00017

Symptomatologic pathomechanism of N-methyl D-aspartate receptor encephalitis

Woo-Jin Lee

Department of Neurology, Seoul National University Hospital, Seoul, Korea

N-methyl D-aspartate receptor (NMDAR) encephalitis is a well-characterized clinical syndrome. The main molecular mechanism of NMDAR encephalitis is autoantibody-mediated NMDAR hypofunction in the neuronal synapse. Several pathomechanistic hypotheses might explain how NMDAR hypofunction causes the typical symptoms and prognosis of NMDAR encephalitis. Suppression of NMDAR-dependent gamma-aminobutyric acid interneurons provokes an accelerated activation of the positive feedback loops of the dorsolateral prefrontal cortex/subiculum-nucleus accumbens circuit in the striatum, the ventral tegmental area (VTA), and the nucleus reuniens in the thalamus-hippocampus-VTA loop. Dysregulated activation of the VTA and cortex via those positive feedback loops may explain the rapid clinical deterioration at acute stages of the disease and the well-characterized syndrome that includes limbic system dysfunction, intractable seizures, dyskinesia, coma, and the characteristic extreme delta brush. Progressive cerebellar atrophy is correlated with cumulative disease burden and is associated with worse long-term outcomes, which might be explained by the NMDAR-dependent pathways required to maintain neuronal survival. Those pathomechanistic hypotheses for NMDAR encephalitis support the rationale for the early introduction of combination immunotherapy and the use of adjuvant immunotherapy in patients with persisting symptoms in chronic disease phases.

Keywords; N-methyl D-aspartate receptor encephalitis, Hashimoto's encephalitis, Pathophysiology

Introduction

N-methyl D-aspartate receptor (NMDAR) encephalitis is the most common antibody-mediated autoimmune encephalitis that frequently affects young females [1-4]. Ovarian teratoma, herpes simplex encephalitis, or other autoimmune diseases such as neuromyelitis optica spectrum disorder are known to trigger the production of immunoglobulin G1 (IgG1) and promote autoantibody binding to the NR1 subunit of NMDAR, resulting in the hypofunction of NMDA signals in neuronal synapses [5-8].

NMDAR encephalitis is a well-characterized clinical syndrome [9,10]. After the prodromal manifestation of fever and headache, typical symptoms occur that involve the limbic system, including memory dysfunction, psychiatric symptoms, language dis-

turbance, and seizures. Once developed, encephalitis tends to rapidly deteriorate neural function, with patients exhibiting intractable seizures, dyskinesia, dysautonomia, central hypoventilation, and coma [4,11]. Although NMDAR encephalitis has a monophasic course with a near-complete recovery of symptoms in most cases, long-term sequelae such as executive dysfunction, psychomotor slowing, disinhibition, and disturbed sleep remain in some severe and intractable cases [9-14]. However, the mechanism for the anti-NMDAR autoantibody-related development of these characteristic clinical features and patterns of progression has not been fully addressed.

Here, we briefly introduce several pathomechanistic hypotheses that explain how NMDAR hypofunction causes the typical symptoms and prognosis of NMDAR encephalitis.

Received: February 1, 2021 Revised: February 15, 2021 Accepted: February 16, 2021

Correspondence: Woo-Jin Lee

Department of Neurology, Seoul National University Hospital, 101 Daehak-ro, Jongno-gu, Seoul 03080, Korea

E-mail: eoozin85@snu.ac.kr

ORCID: https://orcid.org/0000-0001-6090-7921

Copyright © 2021 by The Korean Encephalitis and Neuroinflammation Society

This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

NMDAR and Its Function

NMDAR is one of the three major glutamate-gated cation channels, along with α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor and kainate receptor. NMDAR is a tetramer comprised of two NR1 and two NR2 or NR3 subunits [15]. Alternative splicing of the GRIN1 gene results in eight isoforms of the NR1 subunit; NR2 and NR3 are also classified into four (NR2A-NR2D) and two (NR3A-NR3B) isoforms, respectively. The combination of these isoforms makes up various subtypes of NMDARs with different spatial distributions and molecular functions [15,16].

Binding of glycine/D-serine to the NR1 subunit or binding of glutamate/NMDA to the NR2/3 subunit activates NMDARs and results in the flow of Na⁺ and Ca²⁺ into neurons and K⁺ out of neurons. By means of Ca2+ influx, NMDAR mediates various intra-neuronal signaling pathways [6,9,17,18]. NMDAR is also involved in synaptic plasticity, which is crucial for learning and memory, and a certain level of NMDA signaling is also critical for maintaining neuronal survival [19-24].

Anti-NMDAR Autoantibody Mechanism of Action

The main mechanism of the NMDAR autoantibody is the anti-NMDAR autoantibody-provoked cross-linking of NMDARs which inhibits NMDAR's interaction with ephrin B2, a synaptic protein that stabilizes NMDAR for its placement in the synaptic structure [5,6,8,9,18,25,26]. Consequently, anti-NMDAR autoantibodies cause the NMDAR hypofunction by removing NMDAR from the synaptic surface by receptor internalization or by repositioning to the extrasynaptic surface [5,6,8,9,18,25]. Binding of IgG1 autoantibodies to NMDARs induces the transient NMDAR-mediated hyperactivation of neurons, which manifests as seizure, dyskinesia, and dystonia, and provokes excitotoxic damage to neurons. Additionally, IgG1 autoantibodies have a high affinity for the $Fc\gamma$ receptor, which enables the complement activation and provocation of T-cell mediated autoimmunity, resulting in permanent damage to neurons [6]. This mechanism might be an explanation for long-term neurological deficit after severe and refractory NMDAR encephalitis associated with diffuse cerebral atrophy and progressive cerebellar atrophy [9,10,12,13].

Pathomechanism of Rapidly Progressive Limbic Dysfunction

Given that the typical symptoms involving the limbic system in NMDAR encephalitis resemble the core symptoms of schizophrenia, the NMDA-mediated pathomechanism of schizophrenia might be successfully applied to understand the symptomatologic mechanism of NMDAR encephalitis [9,10,27]. In the schizophrenia model, parvalbumin or glutamate decarboxylase (GAD) positive inhibitory interneurons activated by NMDAR have a key mechanistic role. Parvalbumin-positive interneurons exert gamma-aminobutyric acid (GABA)-dependent inhibition on glutamatergic pyramidal neurons in the dorsolateral prefrontal cortex (DLPFC) and the subiculum of the hippocampus, which excites the nucleus accumbens (NA) of the striatum. NA exerts inhibitory regulation on the globus pallidus medialis (GPm), and the GPm also inhibits the ventral tegmental area (VTA) in the midbrain. Excitatory dopaminergic neurons in the VTA stimulate the DLPFC, NA, and hippocampus while receiving inhibitory regulation from GAD-positive interneurons [9,10,27].

Taken together, the DLPFC/subiculum, NA, and VTA form a closed loop of simulative interaction, and NMDAR-containing interneurons have a critical role as a restrainer of this positive feedback system [9,10,27]. Hypofunction of NMDAR in this system induces hyperactivation of the NA and VTA by disinhibition, resulting in dysregulated excitation of the DLPFC and subiculum (Figure 1). Given that DLPFC is the fundamental structure responsible for executive function and the integration of other cognitive functions and the subiculum is the main output structure of the hippocampus, dysregulated hyperactivation of these structures explains memory, language, and other cognitive dysfunctions [28-30]. Additionally, the VTA exerts widespread dopaminergic stimulatory projections to the cerebral cortex and limbic structures, which are called mesocortical and mesolimbic pathways, respectively [31]. Therefore, unregulated hyperfunction of the VTA might be responsible for psychiatric symptoms along with working memory deficit and global cognitive impairment in NMDAR encephalitis [27]. However, in contrast to NMDAR encephalitis, schizophrenia is a disorder with a high genetic predisposition, and its symptoms contain both positive and negative symptoms of dopaminergic dysregulation. Additionally, as well as NMDAR hypofunction, the disrupted balance between NMDA and AMPA receptors also has a significant role in the pathomechanism of schizophrenia [32]. The differences between these two diseases should be investigated to further elucidate the pathomechanism of NMDAR encephalitis.

The unregulated auto-activation of the DLPFC/subiculum-NA-VTA system also explains the rapidly progressing features of disease at the acute stages. Due to the loss of the NMDAR restrainer, the stimulatory interaction among DLPFC/subiculum, NA, and VTA strengthens by itself via this positive feedback loop, manifesting as an accelerated disease progression to the neurological nadir [9-11].

Extreme Delta Brush: Role of the Thalamus

Although not yet elucidated due to the high complexity of its involved circuits, thalamic structures might be critically involved in the pathogenesis of extreme delta brush, the core features of NMDAR encephalitis [33]. This hypothesis is supported by descriptions of abnormally enhanced delta and beta-gamma activities in both schizophrenia and NDMAR encephalitis [33-36] and research demonstrating that the thalamus is the main generator of the brain activities of those frequency bands during wakefulness [37-43].

The first pathologically important thalamic structure is the nucleus reuniens (nRE). The nRE is located in the medial thalamus and exerts excitatory signaling on the cornu ammonis 1 and the subiculum of the hippocampus [43,44]. Thus, the nRE facilitates activation of VTA via the subiculum-NA-VTA pathway [43,44]. As activation of dopaminergic cells in the VTA results in the reciprocal enhancement of thalamic bursting, the nRE-hippocampus-VTA constitutes another loop of simulative interaction.

The second thalamic structure of interest is the nucleus reticularis thalami (nRT) of the dorsal thalamus, a crucial structure that regulates cortical wakefulness [37-39]. The nRT is composed of parvalbumin-positive GABAergic neurons activated by NMDARs. GABAergic neurons in the nRT exert a strong inhibitory regulation on thalamocortical neurons, and their activation results in strong cortical deactivation [40]. The inhibitory effect of nRT on thalamocortical neurons might is justified given that the nRT is the main inhibitor of the nRE, and thereby regulates general activation of the cortex via suppressing the nRE-hippocampus-VTA loop [43,45]. Due to these network properties, the nRT is regarded as the thalamic pacemaker of the cortex [40-42]. The infralimbic subdivision of the medial prefrontal cortex also contributes to the balance between nRE-mediated cortical activation and nRT-mediated depression by exerting stimulatory signals on both the nRT and nRE [43,45].

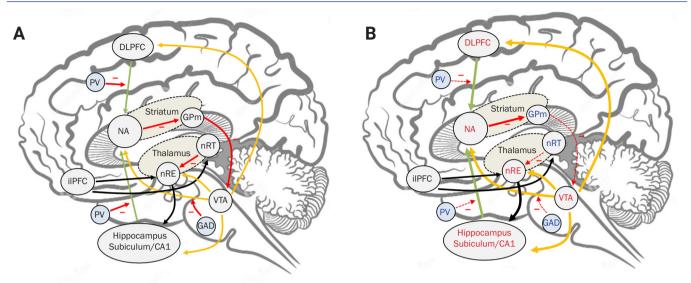
As the nRT's GABAergic inhibition on the nRE is mediated by NMDAR, the nRE is the most highly activated thalamic structure by NMDAR antagonism [34,36]. Loss of NMDAR-mediated inhibition from nRT can enhance the burst activation of nRE by the nRE-hippocampus-VTA positive feedback loop, resulting in the formation of a rhythmic, synchronized activity of nRE (Figure 1) [34,36]. Accordingly, the cortex might be synchronized with the rhythmic activity of nRE via thalamocortical connections and projections from the VTA [34,36,40-42]. Synchronized low-frequency rhythmic activity in the cortex might appear as a frontal dominant rhythmic delta activity, a core electroencephalography feature of severe NMDAR encephalitis [33]. Fast activities with beta to gamma frequency bands superimposed with rhythmic delta activities, constituting another major feature of extreme delta brush, might represent the increased rhythmic cortical activity (Figure 2) [33,35,46,47]. This hypothesis is consistent with a recent study which demonstrated that pharmacological inhibition of the nRT resulted in the marked increment of cortical, delta, and gamma activity during wakefulness [36].

Pathomechanism of Consciousness Decrement, Seizure, Dyskinesia, and **Psychosis**

The thalamic hypothesis might also be applied to explain other major symptoms of NMDAR encephalitis. Multiple neurotransmitter systems involving the brainstem, hypothalamus, thalamus, and basal forebrain form a complex interaction to regulate cortical wakefulness. Among them, the nRT is a key structure for maintaining wakefulness [38,42]. Although activation of the nRT induces local suppression of cortical activity, global hypofunction of the NMDAR-dependent nRT provokes an unregulated burst activation of the nRE-hippocampus-VTA loop [38,42]. This results in the predominate delta rhythm throughout the cortex, which may critically contribute to the decrement of consciousness [38,41,42].

The major pathomechanism of intractable seizure development in NMDAR encephalitis is the shutdown of NMDR-dependent, parvalbumin- or GAD-positive inhibitory interneurons in the cortex and the thalamus [6,9,27]. Synchronized rhythmic cortical activation resulting from unregulated activation of the DLP-FC/subiculum-NA-VTA and nRE-hippocampus-VTA positive feedback loops maximizes the susceptibility to seizure development [9,33,48] and can fulfill the definition of a seizure in itself [49]. Additionally, binding of autoantibodies to NMDAR might provoke a transient activation of NMDAR, which provokes the flow of Na⁺ and Ca²⁺ into neurons and K⁺ out of neurons [6,17,50]. This antibody binding-mediated neuronal excitation might partially contribute to the development of seizures at very acute stages of disease.

Figure 1 Schematic demonstration of altered interactions among the structures involved in the pathomechanism of N-methyl D-aspartate receptor (NMDAR) encephalitis

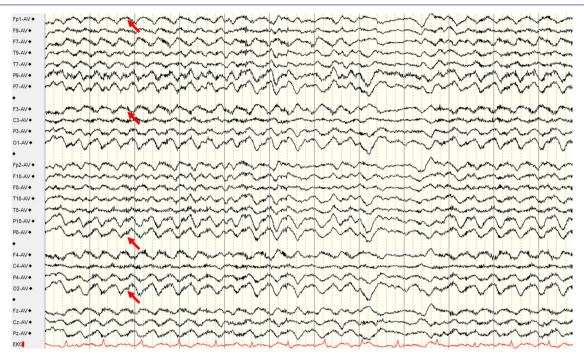


Panel A demonstrates the normal interaction, and panel B demonstrates altered interactions among the structures involved in the pathomechanism of NMDAR encephalitis. Red arrows indicate gamma-aminobutyric acid-mediated (GABAergic) inhibitory action, black arrows represent excitatory action, green arrows indicate glutamatergic excitatory action, and yellow arrows indicate dopaminergic excitatory action. In panel A, parvalbumin (PV)positive interneurons exert GABAergic inhibition on glutamatergic pyramidal neurons in the dorsolateral prefrontal cortex (DLPFC) and the subiculum of the hippocampus. Those pyramidal neurons, the nucleus accumbens (NA) in the striatum, and the ventral tegmental area (VTA) form a closed loop of simulative interaction, and NMDAR-containing interneurons are restrainers of this positive feedback system. The nucleus reuniens (nRE) in the medial thalamus exerts excitatory signaling on the cornu ammonis 1 (CA1) and subiculum and facilitates activation of the VTA via the subiculum-NA-VTA pathway. As activation of VTA results in the reciprocal enhancement of thalamic bursting, the nerd-hippocampus-VTA constitutes another loop of simulative interaction. GABAergic neurons in the nucleus reticularis thalami (nRT) in the dorsal thalamus are the main inhibitors of the nRE, and also inhibit activation of the cortex via suppressing the nRE-hippocampus-VTA loop. The infralimbic subdivision of the medial prefrontal cortex (iIPFC) contributes to the balance between nRE-mediated cortical activation and nRT-mediated depression. In panel B, hypofunction of NMDAR in the DLPFC/ subiculum-NA-VTA system induces hyperactivation of the NA and VTA, resulting in excitation of the DLPFC and subiculum. Loss of NMDAR-mediated inhibition of the nRT on the nRE can enhance the burst activation of the nRE, resulting in the formation of synchronized rhythmic activities in the nRE and cortex. Thick arrows indicate enhanced interactions, and finely dotted arrows indicate weakened interactions. Structures named in red indicate activated structures, whereas structures named in blue indicate inhibited structures. GPm, globus pallidus medialis; GAD, glutamate decarboxylase.

Dyskinesia is a frequently encountered symptom of NMDAR encephalitis, especially among severe cases [4]. Although the underlying mechanisms of dyskinesia are heterogeneous and largely unrevealed, a recent study using a rat model of NMDAR encephalitis suggested that levodopa-induced dyskinesia is strongly associated with predominant gamma-band oscillation in the primary motor cortex [51,52]. The use of a dopamine antagonist suppressed this gamma-band oscillation along and improved dyskinesia [51,52]. The authors suggested that prominent gamma oscillation represents VPA-mediated primary motor cortex hyperactivation, which might be responsible for the development of dyskinesia [51,52]. The hypothesis of dopamine-mediated cortical hyperactivation can also explain the pathomechanism of dyskinesia in NMDAR encephalitis, as the unregulated activation of both the DLPFC/subiculum-NA-VTA and the nRE-hippocampus-VTA results in enhancement of dopaminergic cortex activation by VTA [35,46].

In the NMDA model of schizophrenia, it is speculated that individuals susceptible to the disease have partially compromised NMDAR function [10,34,44]. A triggering event such as acute stress stimulates dopaminergic activity, which can turn on the DLPFC/subiculum-NA-VTA and the nRE-hippocampus-VTA systems to an auto-reactivated status by means of a positive feedback loop [9,10,27,43,44]. Once activated, positive feedback loops contribute to the sustained activation of the VTA, resulting in the persisting symptoms of schizophrenia even after the initial trigger is removed [10,27,34,44]. This hypothesis can be applied to understand psychosis in NMDAR encephalitis, which is prominent at the initial disease presentation and at the recovering stages [9,10,14,53]. At those disease stages, the level and activity of anti-NMDAR antibody might not accumulate to reach a

Figure 2 Extreme delta brush



Electroencephalography of a patient with N-methyl D-aspartate receptor encephalitis exhibits a synchronized low-frequency rhythmic activity and superimposed fast activities with beta to gamma frequency (red arrows).

This figure was reprinted from Encephalitis: the textbook of encephalitis by Korean Encephalitis and Neuroinflammation Society [47] with permission.

fulminant shut down of NMDAR function, but sufficiently diminish the NMDAR-mediated inhibition of the positive feedback systems to be activated by a certain trigger, resulting in delirium, disinhibition, and aggressive and compulsive behaviors [14,53].

Cerebellar Atrophy in NMDAR Encephalitis and Its Implication on Long-term Sequelae

NMDAR encephalitis exhibits two types of brain atrophy; diffuse cerebral atrophy and cerebellar atrophy [54-57]. Although diffuse cerebral atrophy is reversible and has an unclear association with long-term outcomes, cerebellar atrophy is progressive, irreversible, and associated with poor long-term clinical outcomes [54-57]. In our long-term cohort of NMDAR encephalitis patients with recurrent magnetic resonance imaging evaluations, we observed a significant reduction in both cerebellum and cerebrum volume, although the degree was higher in the cerebellum. Cerebellar atrophy was identified in 23.3% of the total study population and followed a progressive course once developed. The degree of cerebellar volume reduction was correlated with poorer scores of long-term global neurological outcomes and major functional domains such as memory, language, and psychiatric symptoms, which indicate that the degree of cerebellar atrophy is associated with the extent of permanent neurological deficits. The degree of cerebellar volume reduction was also associated with an increased cumulative disease burden (unpublished data).

High degrees of cerebellar atrophy suggest that the cerebellum might be especially vulnerable to brain atrophy in NMDAR encephalitis [19,58]. Previous studies demonstrated that a physiological level of NMDARs is required to maintain neuronal survival due to NMDAR-dependent Ca²⁺-mediated signaling. The cAMP response element-binding protein, extracellular signal-regulated kinase, calcium/calmodulin-dependent kinase II, and nuclear factor kappa-B pathways are required for the downstream activation of brain-derived neurotrophic factor [19-24]. Therefore, autoantibody-mediated depletion of NMDAR below a certain level might provoke neuronal degeneration. Considering that cerebellar granule cells, comprising more than 50% of the neurons in the brain, are highly abundant with NMDAR, the cerebellum might be the most highly affected by NMDAR depletion [59].

The progressive features of cerebellar atrophy imply that a longterm mechanism of neuronal degeneration might persist. A previous study demonstrated chronic persistence of autoantibodies in cerebrospinal fluid, even in patients with favorable recovery, and a slighter decrease of autoantibody was correlated with worse outcomes [60]. This finding suggests that the degree of cerebellar atrophy represents the cumulative effect of long-term NMDAR depletion by persisting autoantibodies [6,9,18]. In this regard, cerebellar volume reduction might be utilized as a surrogate marker for the cumulative burden of disease, which is also associated with long-term outcomes. The association of cerebellar volume reduction with functional outcomes might also be explained by the complex interconnection of the cerebellum with the limbic areas and neocortex [59,61-63].

Therapeutic Implications

Accelerated activation of the DLPFC/subiculum-NA-VTA and the nRE-hippocampus-VTA systems via positive feedback explains intractability at the fulminant status and the rapid progression of disease to the neurological nadir at acute stages of disease. In this regard, the use of combination immunotherapy at early disease stages might be rationalized, especially in cases with high clinical severity or rapid progression, to restore the NMDAR-mediated regulatory system and effectively interrupt the self-accelerated activation of these systems [11]. Furthermore, given that unregulated activation of the VTA-mediated dopaminergic pathway is critically involved in consciousness decrement, seizure, dyskinesia, and psychosis in NMDAR encephalitis, drugs affecting the dopaminergic system should be used with special attention to their adverse effects.

The irreversible and progressive feature of cerebellar atrophy, of which its degree is correlated with long-term neurological deficits, also supports the benefit of the early introduction of combination immunotherapy over the conventional stepwise use of immunotherapy, as it promotes early control of the disease and therefore can minimize the cumulative disease burden [11,64,65]. Additionally, in patients with persisting neurological symptoms in the chronic phase, adjuvant immunotherapy might be rationalized to interrupt the ongoing disease activity, halt the progression of cerebellar atrophy, and prevent longterm sequelae [11,64-68].

Conclusion

The main molecular mechanism of NMDAR encephalitis is autoantibody-mediated NMDAR hypofunction in the neuronal synapse. This results in the suppression of the NMDAR-dependent GABAergic interneurons, provoking accelerated activation of the DLPFC/subiculum-NA-VTA and the nRE-hippocampus-VTA systems via positive feedback which explains the rapidly deteriorating clinical course to the neurological nadir. Dysregulated activation of the VTA and cortex via those positive feedback loops might explain the well-characterized clinical syndrome of NMDAR encephalitis that includes limbic system dysfunction, intractable seizures, dyskinesia, coma, and the characteristic extreme delta brush. Progressive cerebellar atrophy is identified in severe and intractable cases, and its degree is correlated with the cumulative disease burden and worse outcomes, which might be explained by the NMDAR-dependent pathways for maintaining the cerebellar granule cell survival. Those pathomechanistic hypotheses of NMDAR encephalitis support the rationale for the early introduction of combination immunotherapy and the use of adjuvant immunotherapy in patients with persisting symptoms in chronic phases.

It should be noted that the symptomatologic pathomechanisms introduced here are hypothetical. Given the abundant distribution of NMDAR throughout the brain and the high complexity of functional interactions among the structures addressed in this review, further investigations should be aimed to elucidate the exact pathomechanisms of NMDAR encephalitis symptoms by using *in vivo* models or by functional or perfusion imaging analyses matched to quantitative electroencephalography data.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

References

- 1. Lee SK, Lee ST. The laboratory diagnosis of autoimmune encephalitis. J Epilepsy Res 2016;6:45-50.
- 2. Dalmau J, Gleichman AJ, Hughes EG, et al. Anti-NMDA-receptor encephalitis: case series and analysis of the effects of antibodies. Lancet Neurol 2008;7:1091-1098.
- 3. Dalmau J, Tüzün E, Wu HY, et al. Paraneoplastic anti-N-methyl-D-aspartate receptor encephalitis associated with ovarian teratoma. Ann Neurol 2007:61:25-36.
- 4. Titulaer MJ, McCracken L, Gabilondo I, et al. Treatment and prognostic factors for long-term outcome in patients with anti-NMDA receptor encephalitis: an observational cohort study. Lancet Neurol 2013; 12:157-165.
- 5. Tüzün E, Zhou L, Baehring JM, Bannykh S, Rosenfeld MR, Dalmau J. Evidence for antibody-mediated pathogenesis in anti-NMDAR encephalitis associated with ovarian teratoma. Acta Neuropathol

- 2009;118:737-743.
- 6. Moscato EH, Peng X, Jain A, Parsons TD, Dalmau J, Balice-Gordon RJ. Acute mechanisms underlying antibody effects in anti-N-methyl-D-aspartate receptor encephalitis. Ann Neurol 2014;76:108-119.
- 7. Linnoila J, Pulli B, Armangué T, et al. Mouse model of anti-NMDA receptor post-herpes simplex encephalitis. Neurol Neuroimmunol Neuroinflamm 2018;6:e529.
- 8. Hughes EG, Peng X, Gleichman AJ, et al. Cellular and synaptic mechanisms of anti-NMDA receptor encephalitis. J Neurosci 2010;30:5866-5875.
- 9. Dalmau J, Armangué T, Planagumà J, et al. An update on anti-NMDA receptor encephalitis for neurologists and psychiatrists: mechanisms and models. Lancet Neurol 2019;18:1045-1057.
- 10. Kayser MS, Dalmau J. Anti-NMDA receptor encephalitis in psychiatry. Curr Psychiatry Rev 2011;7:189-193.
- 11. Lee WJ, Lee ST, Shin YW, et al. Teratoma removal, steroid, IVIG, rituximab and tocilizumab (T-SIRT) in anti-NMDAR encephalitis. Neurotherapeutics 2020 Sep 3 [Epub]. https://doi.org/10.1007/s13311-020-00921-7.
- 12. McKeon GL, Scott JG, Spooner DM, et al. Cognitive and social functioning deficits after anti-n-methyl-d-aspartate receptor encephalitis: an exploratory case series. J Int Neuropsychol Soc 2016;22:828-838.
- 13. Finke C, Kopp UA, Prüss H, Dalmau J, Wandinger KP, Ploner CJ. Cognitive deficits following anti-NMDA receptor encephalitis. J Neurol Neurosurg Psychiatry 2012;83:195-198.
- 14. de Bruijn MA, Aarsen FK, van Oosterhout MP, et al. Long-term neuropsychological outcome following pediatric anti-NMDAR encephalitis. Neurology 2018;90:e1997-e2005.
- 15. Paoletti P, Bellone C, Zhou Q. NMDA receptor subunit diversity: impact on receptor properties, synaptic plasticity and disease. Nat Rev Neurosci 2013;14:383-400.
- 16. Peer M, Prüss H, Ben-Dayan I, Paul F, Arzy S, Finke C. Functional connectivity of large-scale brain networks in patients with anti-NMDA receptor encephalitis: an observational study. Lancet Psychiatry 2017; 4:768-774.
- 17. Crisp SJ, Kullmann DM, Vincent A. Autoimmune synaptopathies. Nat Rev Neurosci 2016;17:103-117.
- 18. Dalmau J, Graus F. Antibody-mediated encephalitis. N Engl J Med 2018:378:840-851.
- 19. Marini AM, Paul SM. N-methyl-D-aspartate receptor-mediated neuroprotection in cerebellar granule cells requires new RNA and protein synthesis. Proc Natl Acad Sci USA 1992;89:6555-6559.
- 20. Shen K, Meyer T. Dynamic control of CaMKII translocation and localization in hippocampal neurons by NMDA receptor stimulation. Science 1999;284:162-166.
- 21. Zhu D, Wu X, Strauss KI, et al. N-methyl-D-aspartate and TrkB receptors protect neurons against glutamate excitotoxicity through an extra-

- cellular signal-regulated kinase pathway. J Neurosci Res 2005;80:104-113.
- 22. Lipsky RH, Xu K, Zhu D, et al. Nuclear factor kappaB is a critical determinant in N-methyl-D-aspartate receptor-mediated neuroprotection. J Neurochem 2001;78:254-264.
- 23. Bhave SV, Ghoda L, Hoffman PL. Brain-derived neurotrophic factor mediates the anti-apoptotic effect of NMDA in cerebellar granule neurons: signal transduction cascades and site of ethanol action. J Neurosci 1999;19:3277-3286.
- 24. Deisseroth K, Heist EK, Tsien RW. Translocation of calmodulin to the nucleus supports CREB phosphorylation in hippocampal neurons. Nature 1998;392:198-202.
- 25. Irani SR, Bera K, Waters P, et al. N-methyl-D-aspartate antibody encephalitis: temporal progression of clinical and paraclinical observations in a predominantly non-paraneoplastic disorder of both sexes. Brain 2010;133:1655-1667.
- 26. Planagumà J, Haselmann H, Mannara F, et al. Ephrin-B2 prevents N-methyl-D-aspartate receptor antibody effects on memory and neuroplasticity. Ann Neurol 2016;80:388-400.
- 27. Dalmau J. NMDA receptor encephalitis and other antibody-mediated disorders of the synapse: the 2016 Cotzias Lecture. Neurology 2016;87:2471-2482.
- 28. Levy R, Goldman-Rakic PS. Segregation of working memory functions within the dorsolateral prefrontal cortex. Exp Brain Res 2000;133:23-
- 29. Callicott JH, Bertolino A, Mattay VS, et al. Physiological dysfunction of the dorsolateral prefrontal cortex in schizophrenia revisited. Cereb Cortex 2000:10:1078-1092.
- 30. Aggleton JP, Christiansen K. The subiculum: the heart of the extended hippocampal system. Prog Brain Res 2015;219:65-82.
- 31. Settell ML, Testini P, Cho S, et al. Functional circuitry effect of ventral tegmental area deep brain stimulation: imaging and neurochemical evidence of mesocortical and mesolimbic pathway modulation. Front Neurosci 2017;11:104.
- 32. Owen MJ, Sawa A, Mortensen PB. Schizophrenia. Lancet 2016;388:86-
- 33. Schmitt SE, Pargeon K, Frechette ES, Hirsch LJ, Dalmau J, Friedman D. Extreme delta brush: a unique EEG pattern in adults with anti-NMDA receptor encephalitis. Neurology 2012;79:1094-1100.
- 34. Khlestova E, Johnson JW, Krystal JH, Lisman J. The role of GluN2C-containing NMDA receptors in Ketamine's psychotogenic action and in schizophrenia models. J Neurosci 2016;36:11151-11157.
- 35. Thuné H, Recasens M, Uhlhaas PJ. The 40-hz auditory steady-state response in patients with schizophrenia: a meta-analysis. JAMA Psychiatry 2016:73:1145-1153.
- 36. Thankachan S, Katsuki F, McKenna JT, et al. Thalamic reticular nucleus parvalbumin neurons regulate sleep spindles and electrophysio-

- logical aspects of schizophrenia in mice. Sci Rep 2019;9:3607.
- 37. Golmayo L, Nuñez A, Zaborszky L. Electrophysiological evidence for the existence of a posterior cortical-prefrontal-basal forebrain circuitry in modulating sensory responses in visual and somatosensory rat cortical areas. Neuroscience 2003;119:597-609.
- 38. Poulet JF, Crochet S. The cortical states of wakefulness. Front Syst Neurosci 2019;12:64.
- 39. Wimmer RD, Schmitt LI, Davidson TJ, Nakajima M, Deisseroth K, Halassa MM. Thalamic control of sensory selection in divided attention. Nature 2015;526:705-709.
- 40. Fuentealba P, Steriade M. The reticular nucleus revisited: intrinsic and network properties of a thalamic pacemaker. Prog Neurobiol 2005;75:125-141.
- 41. Herrera CG, Cadavieco MC, Jego S, Ponomarenko A, Korotkova T, Adamantidis A. Hypothalamic feedforward inhibition of thalamocortical network controls arousal and consciousness. Nat Neurosci 2016;19: 290-298.
- 42. Lewis LD, Voigts J, Flores FJ, et al. Thalamic reticular nucleus induces fast and local modulation of arousal state. Elife 2015;4:e08760.
- 43. Zimmerman EC, Grace AA. The nucleus reuniens of the midline thalamus gates prefrontal-hippocampal modulation of ventral tegmental area dopamine neuron activity. J Neurosci 2016;36:8977-8984.
- 44. Lisman JE, Pi HJ, Zhang Y, Otmakhova NA. A thalamo-hippocampal-ventral tegmental area loop may produce the positive feedback that underlies the psychotic break in schizophrenia. Biol Psychiatry 2010;68:17-24.
- 45. Zimmerman EC, Grace AA. Prefrontal cortex modulates firing pattern in the nucleus reuniens of the midline thalamus via distinct corticothalamic pathways. Eur J Neurosci 2018;48:3255-3272.
- 46. Mann EO, Mody I. Control of hippocampal gamma oscillation frequency by tonic inhibition and excitation of interneurons. Nat Neurosci 2010:13:205-212.
- 47. Korean Encephalitis and Neuroinflammation Society. Encephalitis: the textbook of encephalitis. Seoul: Koonja Publishing; 2021.
- 48. Wright S, Hashemi K, Stasiak L, et al. Epileptogenic effects of NMDAR antibodies in a passive transfer mouse model. Brain 2015;138:3159-3167.
- 49. Veciana M, Becerra JL, Fossas P, et al. EEG extreme delta brush: an ictal pattern in patients with anti-NMDA receptor encephalitis. Epilepsy Behav 2015;49:280-285.
- 50. Manto M, Dalmau J, Didelot A, Rogemond V, Honnorat J. Afferent facilitation of corticomotor responses is increased by IgGs of patients with NMDA-receptor antibodies. J Neurol 2011;258:27-33.
- 51. Halje P, Tamtè M, Richter U, Mohammed M, Cenci MA, Petersson P. Levodopa-induced dyskinesia is strongly associated with resonant cortical oscillations. J Neurosci 2012;32:16541-16551.
- 52. Güttler C, Altschüler J, Tanev K, et al. Levodopa-induced dyskinesia

- are mediated by cortical gamma oscillations in experimental parkinsonism. Mov Disord 2020 Nov 28 [Epub]. https://doi.org/10.1002/ mds.28403.
- 53. Al-Diwani A, Handel A, Townsend L, et al. The psychopathology of NMDAR-antibody encephalitis in adults: a systematic review and phenotypic analysis of individual patient data. Lancet Psychiatry 2019;6:235-246.
- 54. lizuka T, Yoshii S, Kan S, et al. Reversible brain atrophy in anti-NMDA receptor encephalitis: a long-term observational study. J Neurol 2010; 257:1686-1691.
- 55. Kataoka H, Sawa N, Tonomura Y, Ueno S. Early progression of brain atrophy in patients with anti-N-methyl-D-aspartate receptor encephalitis: case reports. Medicine (Baltimore) 2017;96:e6776.
- 56. Laurikainen H, Isotupa I, Nyman M, et al. Longitudinal brain morphology in anti-NMDA receptor encephalitis: a case report with controls. BMC Psychiatry 2019;19:145.
- 57. lizuka T, Kaneko J, Tominaga N, et al. Association of progressive cerebellar atrophy with long-term outcome in patients with anti-N-methyl-d-aspartate receptor encephalitis. JAMA Neurol 2016;73:706-713.
- 58. Ciani E, Rizzi S, Paulsen RE, Contestabile A. Chronic pre-explant blockade of the NMDA receptor affects survival of cerebellar granule cells explanted in vitro. Brain Res Dev Brain Res 1997;99:112-117.
- 59. Lackey EP, Heck DH, Sillitoe RV. Recent advances in understanding the mechanisms of cerebellar granule cell development and function and their contribution to behavior. F1000Res 2018;7:F1000 Faculty Rev-1142.
- 60. Gresa-Arribas N, Titulaer MJ, Torrents A, et al. Antibody titres at diagnosis and during follow-up of anti-NMDA receptor encephalitis: a retrospective study. Lancet Neurol 2014;13:167-177.
- 61. Kleim JA, Freeman JH Jr, Bruneau R, et al. Synapse formation is associated with memory storage in the cerebellum. Proc Natl Acad Sci U S A 2002:99:13228-13231.
- 62. Mariën P, Ackermann H, Adamaszek M, et al. Consensus paper: language and the cerebellum: an ongoing enigma. Cerebellum 2014; 13:386-410.
- 63. Schmahmann JD, Sherman JC. The cerebellar cognitive affective syndrome. Brain 1998;121:561-579.
- 64. Lee WJ, Lee ST, Byun JI, et al. Rituximab treatment for autoimmune limbic encephalitis in an institutional cohort. Neurology 2016;86: 1683-1691.
- 65. Lee WJ, Lee ST, Moon J, et al. Tocilizumab in autoimmune encephalitis refractory to rituximab: an institutional cohort study. Neurotherapeutics 2016;13:824-832.
- 66. Shin YW, Lee ST, Park KI, et al. Treatment strategies for autoimmune encephalitis. Ther Adv Neurol Disord 2017;11:1756285617722347.
- 67. Shin YW, Lee ST, Kim TJ, Jun JS, Chu K. Bortezomib treatment for severe refractory anti-NMDA receptor encephalitis. Ann Clin Transl

Neurol 2018;5:598-605.

68. Lim JA, Lee ST, Moon J, et al. New feasible treatment for refractory au-

toimmune encephalitis: low-dose interleukin-2. J Neuroimmunol 2016;299:107-111.