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Original research

In-depth characterisation of a cohort of individuals with missense and loss-of-function variants disrupting *FOXP2*

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ABSTRACT

Background Heterozygous disruptions of *FOXP2* were the first identified molecular cause for severe speech disorder: childhood apraxia of speech (CAS), and yet few cases have been reported, limiting knowledge of the condition.

Methods Here we phenotyped 28 individuals from 17 families with pathogenic *FOXP2*-only variants (12 loss-of-function, five missense variants; 14 males; aged 2 to 62 years). Health and development (cognitive, motor, social domains) were examined, including speech and language outcomes with the first cross-linguistic analysis of English and German.

Results Speech disorders were prevalent (23/25, 92%) and CAS was most common (22/25, 88%), with similar speech presentations across English and German. Speech was still impaired in adulthood, and some speech sounds (eg, 'th', 'r', 'ch', 'j') were never acquired. Language impairments (21/25, 84%) ranged from mild to severe. Comorbidities included feeding difficulties in infancy (10/26, 38%), fine (13/26, 50%) and gross (13/26, 50%) motor impairment, anxiety (5/27, 19%), depression (6/27, 22%) and sleep disturbance (10/24, 42%). Physical features were common (22/27, 81%) but with no consistent pattern. Cognition ranged from average to mildly impaired and was incongruent with language ability; for example, seven participants with severe language disorder had average non-verbal cognition.

Conclusions Although we identify an increased prevalence of conditions like anxiety, depression and sleep disturbance, we confirm that the consequences of *FOXP2* dysfunction remain relatively specific to speech disorder, as compared with other recently identified monogenic conditions associated with CAS. Thus, our findings reinforce that *FOXP2* provides a valuable entry point for examining the neurobiological bases of speech disorder.

INTRODUCTION

FOXP2 was the first gene implicated in a developmental speech and language disorder in the absence of intellectual disability.¹ A private heterozygous missense variant in *FOXP2* was found to

WHAT IS ALREADY KNOWN ON THIS TOPIC

⇒ Heterozygous disruptions of *FOXP2* were the first identified molecular cause for severe speech disorder: childhood apraxia of speech (CAS), and yet few cases have been reported, limiting knowledge of the condition.

WHAT THIS STUDY ADDS

⇒ Here we provide the most comprehensive characterisation of individuals with pathogenic *FOXP2* variants, almost doubling the number of published families to date. We provide the first cross-linguistic analysis of speech and language across German and English. We show that the phenotype for pathogenic *FOXP2* variants remains relatively specific to speech disorder, compared with phenotypes associated with other monogenic conditions involving CAS.

HOW THIS STUDY MIGHT AFFECT RESEARCH, PRACTICE OR POLICY

⇒ This study guides the identification of cases with a *FOXP2*-related disorder for a clinical genetic diagnosis, improves prognostic counselling and leads to a better targeted clinical management.

cosegregate with childhood apraxia of speech (CAS) in 15 members of the multigenerational British 'KE family', while being absent from all unaffected relatives and healthy controls.¹ The study also characterised a balanced chromosomal translocation with a 7q31.2 breakpoint disrupting *FOXP2* in an unrelated proband with similar speech deficits.¹ CAS is a disorder of speech motor planning and programming that manifests in impaired sequencing of speech sounds into syllables and words with the correct prosody.² The condition is associated with delayed and protracted speech development.

Pathogenic single-nucleotide variants (SNVs) and intragenic indels that disrupt *FOXP2* are rare. To our knowledge, there have been a dozen of these SNVs/indels reported in the literature to date: the



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original missense variant in the large KE family,¹ a non-sense (stop-gain) variant in two siblings and their mother,³ a frameshift in a sporadic patient⁴ and eight variants across eight small families (intragenic deletions, non-sense, missense and frameshift variants)⁵ (online supplemental table 1), each occurring in a heterozygous state. The limited number of cases reported may in part be due to the relatively mild speech-focused phenotype associated with pathogenic SNVs/indels of *FOXP2*, compared with other neurogenetic childhood disorders. While debilitating for affected probands and families, a CAS phenotype does not often lead to clinical genetic testing or to ascertainment in gene discovery cohorts for other neurodevelopmental disorders such as autism spectrum disorder (ASD) or intellectual disability. All probands with pathogenic *FOXP2* variants reported to date share a severe speech disorder presentation, most commonly CAS. Yet there is preliminary evidence that, in some cases, SNVs or indels of *FOXP2* may cause a broader phenotype including subtle dysmorphology and co-occurring neurodevelopmental features such as ASD.⁵

There are other variants that affect *FOXP2* that are not SNVs or indels. A large deletion downstream of *FOXP2* was hypothesised to have a position effect on expression.⁶ There have also been case series of large heterozygous 7q31 deletions or reciprocal-balanced translocations associated with more complex phenotypes involving disruptions of *FOXP2* in addition to neighbouring genes.^{7–15} Such phenotypes are now clinically defined as having a *FOXP2*-plus-related disorder.¹⁶

A systematic prospective cohort study of the phenotype(s) associated with *FOXP2* variants is warranted to guide better identification of cases for clinical genetic diagnosis, improve prognostic counselling and lead to better targeted intervention. Here we examine speech, language, health and broader developmental phenotypes associated with pathogenic *FOXP2* SNVs/indels in a cohort of 28 probands from 17 unrelated families (7 previously published but not deeply characterised for speech and language^{3–5} and 10 novel) expanding the genetic and clinical spectrum of the disorder. For the first time in any phenotypic study of *FOXP2*, the specificity of a linguistic phenotype, relative to broader neurodevelopmental skills (eg, communication ability compared with domains of social, motor and daily living skills), was examined using standardised tests. A novel cross-linguistic comparison of speech diagnoses in German-speaking versus English-speaking participants was also conducted to determine homogeneity of the speech phenotype across languages.

METHODS

Participants

Inclusion criterion was a molecular diagnosis of pathogenic variants (SNVs or intragenic deletions/duplications) in *FOXP2*, in individuals aged ≥ 6 months. Participants were recruited via clinical genetics colleagues or family self-referral in the Netherlands, France, Britain, Germany, the USA, the UK and Australia. Adult participants and caregivers of child participants provided informed consent to participate in the study. The assessment battery was tailored to cover a wide range of ages and languages.

Health and development

Health and medical information, including developmental milestones and existing diagnoses of neurodevelopmental conditions, were collected via an established direct (adult) or caregiver survey.^{17–19} Health professional reports and consults confirmed caregiver survey responses. Feeding (Child Oral and Motor

Proficiency Scale²⁰) and drooling (Drooling Impact Scale²¹) measures were collected where age appropriate.

Speech

In English-speaking participants, phonology and articulation were assessed using standardised tools (Diagnostic Evaluation or Articulation and Phonology²²) or Goldman Fristoe Test of Articulation—Second Edition.²³ Phonological delay versus disorder was delineated. Both for English-speaking (authors LDM, ATM) and German-speaking (author EM) participants, phonological and articulation errors were also analysed from a phonetic transcription of a 5 min conversational speech sample. Across both languages, CAS features were rated across three core diagnostic criteria^{17 18 24}: inconsistent speech errors, lengthened and disrupted coarticulatory transitions and impaired prosody. These three criteria were further operationalised into rateable speech errors (see online supplemental table 2). Similarly, dysarthria was assessed using the Mayo Clinic Dysarthria Classification System rating scale²⁵ and evaluating oral motor structure and function.²⁶ The Intelligibility in Context Scale²⁷ documented how well an individual was understood by conversational partners, with a five-point scale ranging from ‘never’ to ‘always’ understood.

Language and literacy

Receptive vocabulary and expressive vocabulary were assessed using clinician-administered standardised tools dependent on an individual’s age and language (see table 1 for assessments). Likewise, caregiver-administered standardised tools were used to measure speech and language skills. Assessment tools used were dependent on an individuals’ communicative ability, age and language (table 1). Literacy abilities were documented by direct (adult) or caregiver reports, the Vineland Adaptive Behaviour Scales—Third Edition²⁸ written communication subdomain and academic records.

Adaptive behaviour and intellectual ability

The Vineland Adaptive Behaviour Scale, Third Edition (VABS-3) provided scores across language, socialisation, self-care, daily living, motor skills and a composite total adaptive behaviour score.²⁸ A non-parametric Kruskal-Wallis test examined the relative involvement of VABS-3 subdomain scores, and a Wilcoxon signed-rank test was performed between VABS-3 receptive and expressive language scores to highlight any differences between these domains.

General intellectual abilities were assessed with the age-appropriate Wechsler assessment (see table 1 for assessments). Where Full-Scale Intelligence Quotient (FSIQ) could not be obtained, Perceptual Reasoning Index and Matrix Reasoning subtest scores were calculated from the relevant Wechsler assessment. A further three assessment tools were used to assess intellectual abilities in four children (table 1). Diagnoses of neurodevelopmental disorders (eg, autism) were identified by caregiver report and confirmed with clinical records.

RESULTS

Participants

Twenty-eight participants with pathogenic *FOXP2* variants were recruited from 17 families, comprising 10 unreported families (families 1–3, 6, 12–17) and 7 that were previously reported but not deeply characterised for speech and language abilities

Table 1 Language skills and cognition in this cohort with pathogenic missense/loss-of-function variants disrupting *FOXP2*

Case	Age at assessment (year range)	Receptive language		Expressive language		Intellectual abilities	Literacy impairment	
		Vocabulary	Grammar	Vocabulary	Grammar		Spelling	Reading
1a	6–8	Mild*	Moderate†	Moderate‡	Severe††	Average¶ PRI 100	Y	Y
1b	36–38	Mild*	–	–	–	–	Y	Y
1c	30–32	Average*	Average†	–	–	Average¶ PRI 100	Y	Y
1d	60–62	Average*	Moderate†	–	–	Average¶ MR 40	Y	N
2	6–8	Severe*	Severe**	Severe**	Severe††	Borderline‡‡ FSIQ 78	Y	Y
3a	3–5	–	Severe§§	–	Severe§§	Borderline¶¶ FSIQ 73	Y	Y
3b	39–41	–	–	–	–	–	N	N
4a	18–20	Severe*	Moderate†	–	–	Mild*** FSIQ 67	Y	Y
4b	18–20	Moderate*	Severe†	–	–	Mild*** FSIQ 65	Y	Y
4c	15–17	Average*	Average†	–	–	Mild¶ FSIQ 62	Y	Y
5	6–8	–	Severe§§	–	Severe§§	Average¶ PRI 94 Borderline¶ FSIQ 73	Y	Y
6	12–14	Moderate†††	Moderate†	Severe‡	Severe††	Borderline¶ PRI 79	Y	Y
7a	15–17	Severe*	Moderate†	Mild	Below average‡‡‡	Average¶¶ MR 10	Y	Y
7b	15–17	Average*	Moderate†	Severe¶¶	Below average‡‡‡	Average¶¶ MR 8	Y	Y
8a	36–38	Average*	Severe †	Mild***	–	Moderate*** MR 4	N	N
8b	15–17	Moderate*	Severe†	Severe¶¶	Below average‡‡‡	Average¶¶ MR 10	N	N
9	18–20	Severe*	Severe†	–	Severe‡‡‡	Severe¶¶ MR 2 Mild§§§ FSIQ 67	Y	Y
10	9–11	Average§§§	Average†	Mild¶¶	Below average‡‡‡	Average¶¶ MR 17	N	N
11a	39–41	Average*	Average†	Average***	Average‡‡‡	Moderate*** MR 5	N	N
11b	6–8	Average§§§	Severe†	Average¶¶	Severe‡‡‡	Average¶¶ FSIQ 92	Y	Y
12a	3–5	Severe*	–	Severe****	Severe****	Average¶¶ FSIQ 85	NA	NA
12b	33–35	–	–	–	–	–	N	N
13	0–2	Average††††	–	Mild‡‡‡‡	–	–	NA	NA
14	3–5	Severe††††	–	Severe‡‡‡‡	–	–	NA	NA
15	3–5	–	Average§§§§	Severe‡	Severe††	–	Y	Y
16a	42–44	–	–	–	–	–	N	Y
16b	9–11	Mild§§	–	Mild§§	–	Borderline¶¶ FSIQ 78	Y	Y
17	0–2	Average††††	–	Average‡‡‡‡	–	–	NA	NA

Average=−1 < SD; mild=−1 to −1.5 SD; moderate= −1.5 to −2 SD; severe=< −2 SD. Below average=a qualitative descriptor based on the analysis of a transcribed conversation speech sample.
*Peabody Picture Vocabulary Test.⁵⁹
†Test for Reception of Grammar, Second and German Editions.^{60 61}
‡Children's Communication Checklist, Second Edition Semantics subdomain.⁶¹
§Wechsler Abbreviated Scale of Intelligence, Second Edition.⁶²
¶Clinical Evaluation of Language Fundamentals Preschool, Second Edition.⁶³
**Children's Communication Checklist, Second Edition syntax subdomain.
††Kaufman Assessment Battery for Children, Second Edition.⁶⁴
‡‡Clinical Evaluation of Language Fundamentals, Fourth and Fifth Editions.^{65 66}
§§Wechsler Intelligence Scale for Children, Fourth and Fifth Editions.^{67 68}
¶¶Wechsler Adult Intelligence Scale, Second and Fourth Editions.^{62 69}
***Receptive One-Word Picture Vocabulary Test, Fourth Edition.⁷⁰
†††Analysis of transcribed conversation speech sample.
‡‡‡Kaufman Assessment Battery for Children, Second Edition.⁶⁴
§§§Snijders-Oomen Non-verbal Intelligence Test.⁷¹
¶¶¶Schlichting Test for Language Production.
****Communication and Symbolic Behaviour Scales Development Profile Infant-Toddler Checklist Understanding subdomain.⁷²
††††Communication and Symbolic Behaviour Scales Development Profile Infant-Toddler Checklist Words subdomain.
‡‡‡‡Vineland Adaptive Behaviour Scales, Third Edition, Expressive and Receptive subdomain scaled scores.²⁸
§§§§Wechsler Preschool and Primary Scale of Intelligence, Fourth Edition.⁷³
¶¶¶¶Universal Non-verbal Intelligence Test, Second Edition.⁷⁴
-, not assessed; FSIQ, Full-Scale Intelligence Quotient; MR, matrix reasoning; N, no; PRI, perceptual reasoning index; Y, yes.

(families 4, 5, 7–11). Participants had a median age of 16 years 4 months, range 2 years 7 months to 62 years 7 months, and 14 (50%) were male (table 2). Most participants were referred for genetic testing due to the proband's striking speech and language impairment, except for family 3 (n=2) and participant 17. Family 3 underwent genetic testing as part of research testing for preterm children (ID 3a), and subsequently a variant was also

identified in the father (ID 3b). Participant 17 was referred due to microcephaly and dysmorphic facial features.

Genetic results

The 17 families each had a unique *FOXP2* variant, comprising 12 loss-of function and 5 missense variants (table 2). The 12

Table 2 Genotypes and phenotypes of this cohort

Case	cDNA *	Protein	Variant type	ACMG criteria	ACMG classification	Inheritance	Country	Sex	Age, years†	Sleep	Motor	NDD
1a	c.1191dupA	p.Glu398Argfs*31	Loss of function (frameshift)	PVS1, PM2, PP1	Pathogenic	Maternal	Aus	M	6–8	–	Fine and gross	–
1b						Maternal	Aus	F	36–38	–	–	–
1c						Maternal	Aus	F	30–32	+	Fine and gross	ASD, ADHD
1d						NA	Aus	F	60–62	+	–	–
2	(114254407–114308861)×3		Loss of function (intragenic duplication)			De novo	Aus	M	6–8	+	–	ASD, impulsivity, hyperactive, borderline IQ
3a	(114036269–114347379)×1		Loss of function (intragenic deletion)			Paternal	Aus	M	3–5	–	Fine	Borderline IQ, sensory and attention difficulties
3b						De novo	Aus	M	39–41	–	Fine	–
4a†	c.982C>T	p.Arg328†	Loss of function (non-sense)	PVS1, PS2, PM2, PP1	Pathogenic	Maternal	Aus	M	18–20	+	Fine	Tourette's syndrome, mild ID, ASD
4b†						Maternal	Aus	F	18–20	+	Fine	Mild ID, ASD
4c						Maternal	Aus	F	15–17	+	Fine	Mild ID, ASD
5†	c.1168_1169delCA	p.Gln415Valfs*7	Loss of function (frameshift)	PVS1, PS2, PM2	Pathogenic	De novo	Aus	M	6–8	–	Mild tremor	Borderline IQ
6	c.1666C>T	p.Leu556Phe	Missense	PS2, PM1, PM2, PP3, PP4	Pathogenic	De novo	UK	F	12–14	+	Fine and gross	–
7a†	(114296590–114310602)×1		Loss of function (intragenic deletion)			De novo	Ger	F – twin	15–17	–	Gross	–
7b†						De novo	Ger	F – twin	15–17	–	Gross	–
8a†	c.982C>T	p.Arg328†	Loss of function (nonsense)	PVS1, PS2, PM2, PP1	Pathogenic	NA	Ger	F	36–38	–	–	NA
8b†						Maternal	Ger	M	15–17	–	–	–
9†	c.1607G>C	p.Arg536Pro	Missense	PS1, PS2, PM1, PM2, PP3, PP4	Pathogenic	Paternal (? mosaic Fa.)	Ger	F	18–20	NA	Gross	Autistic features, mild ID
10†	c.1432C>T	p.Arg478†	Loss of function (nonsense)	PVS1, PS2, PM2	Pathogenic	De novo	Ger	F	9–11	NA	Broad-based gait	–
11a†	c.1514C>T	p.Pro505Leu	Missense	PS1, PS2, PM1, PM2, PP3, PP4	Pathogenic	NA	Ger	M	39–41	NA	NA	–
11b†						Paternal	Ger	M	6–8	NA	NA	Borderline IQ
12a	c.1385C>G	p.Ser462†	Loss of function (non-sense)	PVS1, PS2, PM2, PP1	Pathogenic	Maternal	NL	M	3–5	–	Fine and gross	SPD
12b						NA	NL	F	33–35	–	–	DCD, ASD
13	c.1513C>A	p.Pro505Thr	Missense	PS1, PS2, PM1, PM2, PP3, PP4	Pathogenic	De novo	NL	M	0–2	–	Fine and gross	–

Continued

Table 2 Continued

Case	cDNA *	Protein	Variant type	ACMG criteria	ACMG classification	Inheritance	Country	Sex	Age, years†	Sleep	Motor	NDD
14	c.1658 G>A	p.Arg553His	Missense	PVS1, PS2, PM2	Pathogenic	De novo	USA	M	3–5	+	Fine and gross	Hyperactive, attention deficit, restricted interests and behaviour
15	c.1428del	p.Lys417Asnfs#7	Loss of function (frameshift)	PVS1, PS2, PM2	Pathogenic	De novo	USA	M	3–5	–	–	Sensory-seeking behaviours
16a	(113844506–114106056)×1		Loss of function (intragenic deletion)			De novo	USA	F	42–44	+	Gross	–
16b						Maternal	USA	F	9–11	+	Gross	Dyslexia, ASD, borderline IQ
17	c.1A>G	p.M1?	Translational start-site variant	PM1, PM2, PM4, PP3, PP4	Likely pathogenic	NA	USA	M	0–2	–	Fine and gross	–

*NM_014491, all deletions and the microduplication are [arr\[hg19\]7q31.1](#).
 †Ages in 2-year age ranges.
 ‡Previously published.
 ††ADHD, attention-deficit hyperactive disorder; ASD, autism spectrum disorder; Aus, Australia; DCD, developmental coordination disorder; F, female; Ger, Germany; M, male; NA, not assessed; NDD, neurodevelopmental disorder; NL, Netherlands; OCD, obsessive-compulsive disorder; SPD, sensory processing disorder; UK, United Kingdom; USA, United States of America.

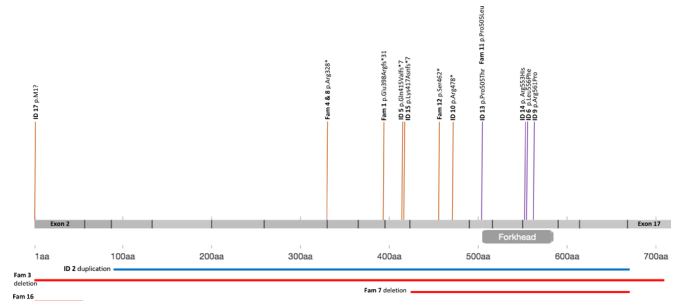


Figure 1 Schematic representation of 17 pathogenic *FOXP2* variants in this cohort from 28 individuals in 17 families (NM_014491).

loss-of-function variants included 3 frameshifts, 4 stop-gain/non-sense variants, 1 variant abolishing the translational start site, 3 intragenic deletions and 1 intragenic duplication. All missense variants were located in the forkhead-box DNA-binding domain of the encoded protein (figure 1). Eleven of the 28 participants had confirmed de novo variants, 12 inherited their variant from a parent, and for 5 participants the inheritance status was unknown (table 2; online supplemental figure 1). Participant 9 was previously reported to have inherited the *FOXP2* variant from their father who had the same variant in a mosaic state, and who was unavailable to take part in the present study. Deletions and sequence variants were submitted to Decipher (<https://decipher.sanger.ac.uk/>).

Health and development

Over a third of the assessed participants had feeding difficulties in infancy (10/26), and some had excessive drooling (5/21) in the early years of life (online supplemental table 3; table 2). Gross motor impairments during early development (13/26) and fine motor impairment (13/26) were also present, with a subset of participants having both fine and gross motor impairment (7/26). Of those with fine motor impairment, participant 12b had a formal diagnosis of developmental coordination disorder. In a small number of individuals, hypotonia (IDs 6, 12a) or microcephaly (ID 17) was also noted. Two participants had hearing impairment: mild, conductive hearing loss (25-39dBHL, ID 15) and moderate mixed hearing loss (40-69dBHL, ID 1d). Sleep disturbances were relatively common (10/24), mostly characterised as difficulty falling asleep (5/24) or frequent waking (4/24). Visual impairments (8/26) were present (online supplemental table 3). Physical features were reported in most participants (22/27, four previously reported (online supplemental table 3)), but with no distinct morphological profile across families. Recorded physical features involved the nose (upturned nose: ID 1a; prominent nose: IDs 1b, 1c; hypoplastic alae nasae: ID 4b, 4c; high nasal root: ID 8b; rounded, fleshy or prominent nasal tip: IDs 1b, 1c, 3a, 5, 8b), philtrum (short/flat philtrum: IDs 2, 17), ears (prominent/protruding ears: IDs 1a, 1b, 12b; anteverted ears: ID 14), eyes (periorbital fullness: IDs 3a, 5, 13; prominent eyes: ID 16b), jaw (retrognathia: IDs 1a, 1b, 3a, 13) and lips (full lips IDs: 1a, 1b, 1c, 4a, 4b, 13; thin upper lip: 3a). In individual cases, mild finger pads (ID 10), tapering fingers (ID 8b), single palmar crease (ID 17) and clinodactyly (ID 12a) were also noted.

Co-occurring diagnoses

A quarter of participants had a diagnosis of ASD (7/27; 2 diagnosed in adulthood) and one had autistic traits but did not meet the criteria. Participant 1c had a diagnosis of attention-deficit

hyperactivity disorder (ADHD). Hyperactivity (2/27), attention difficulties (2/27) and restricted interests and behaviour (2/27) were also noted in further participants without formal ASD or ADHD diagnoses. Mental health conditions such as anxiety (5/27), depression (6/27) and obsessive-compulsive disorder (2/27) were reported in five adults and one adolescent.

Most participants (school aged or older) with pathogenic *FOXP2* variants attended mainstream schools (14/23); seven attended special education schools and two attended a mix of special education and mainstream schools. Learning support (eg, teaching aide, individualised learning plan) was common (15/23) across all settings. All five preschool participants attended specialist preschool settings for children with additional learning needs. All caregivers of school-aged children and adolescents reported that their child's academic progress had been most impacted by their speech and language impairments.

Communication development

Speech development was characterised by limited babbling and a reduced phonetic (sound) inventory relative to peers across the first 7 years of life when a full inventory is typically acquired.

Some developed first words around the typical age of development (12–15 months, 9/25), whereas others were slightly (15–18 months, 1/26) or more significantly (>18 months, 14/26) delayed (table 3). One participant (ID 14) had not said their first words yet in early childhood (3–5 years old). Eight participants had not yet mastered combining words (IDs 1a, 6, 9, 12a, 13, 14, 15, 17). Only three participants combined words in line with the typical development milestone of 2–3 years of age. The remaining participants combined words between 4 to 5 years (4/22), 6 to 7 years (2/22) and 8 years or older (5/22), representing protracted development relative to the typical developmental milestone of 2–3 years.

Speech

CAS was the most common speech diagnosis (88%, 22/25) (figure 2, table 3), with features including frequent sound omissions, the same consonant or vowel being produced differently across different words, impaired sequencing of phonemes and syllables, voicing errors, syllable segregation, difficulty achieving initial articulatory configurations, equal stress, altered suprasegmental features and slow rate (online supplemental table 3).

Table 3 Speech features and educational placement of individuals with pathogenic SNVs/indels disrupting *FOXP2*

Case	Age first words	Age first sentences	CAS	Phonological errors	Dysarthria	Oral motor impairment	Schooling	Support*
1a	>18 mo	NYA	+	Disorder	–	+	MS PS	+
1b	12–15 mo	2–3 years	+	Delay	–	+	MS SC and diploma	–
1c	2–3 years	>8 years	+	Delay	–	+	MS SC and diploma	+
1d	<12 mo	2–3 years	+	–	–	+	MS PS and diploma	–
2	>18 mo	4–5 years	+	Disorder	–	+	Mixed MS and specialist	+
3a	>15 mo	2–3 years	–	Disorder	–	+	MS PS	+
3b	NA	NA	–	–	–	–	MS SC and diploma	–
4a	>18 mo	>8 years	+	Disorder	+	+	MS SC	+
4b	>18 mo	>8 years	+	Disorder	–	+	MS SC	+
4c	>18 mo	>5 years	+	Disorder	–	+	MS SC	+
5	12–15 mo	6–7 years	+	Disorder	+	+	MS SC	+
6	>18 mo	NYA	+	Disorder	–	+	Specialist SC	+
7a	4–5 years	7–8 years	+	–	–	NA	Specialist	–
7b	4–5 years	7–8 years	+	–	–	NA	Specialist	–
8a	NA	NA	+	–	–	NA	MS SC	–
8b	4–5 years	NA	+	–	–	NA	Specialist	–
9	4–5 years	NYA	+	NA	NA	+	Specialist	–
10	12–15 mo	NA	+	–	–	NA	MS school	+
11a	NR	NA	–	–	–	NA	School for speech and language disorders, advanced technical college	–
11b	12–15 mo	NA	+	Disorder	–	NA	Pre for speech and language disorders	–
12a	>18 mo	NYA	+	Disorder	–	NA	Specialist	+
12b	>18 mo	6–7 years	NA	NA	NA	NA	MS and specialist, higher vocational education	–
13	NYA	NYA	+	Disorder	–	NA	Specialist pre	+
14	NYA	NYA	NA	NA	NA	NA	Specialist pre	+
15	12–15 mo	NYA	+	Disorder	–	NA	Specialist pre	+
16a	12–15 mo	4–5 years	NA	NA	NA	NA	MS SC and degree	–
16b	12–15 mo	4–5 years	+	Disorder	–	NA	MS PS	+
17	12–15 mo	NYA	+	Disorder	–	+	Specialist pre	–

+ = feature present, – = feature absent

*Support in the form of support staff in the classroom and/or individualised education plans.

CAS, childhood apraxia of speech; mo, months; MS, mainstream; NA, not assessed; NYA, not yet achieved; OCD, obsessive-compulsive disorder; Pre, preschool; PS, primary/elementary school; SC, secondary school; SNVs, single-nucleotide variants; wks, weeks.

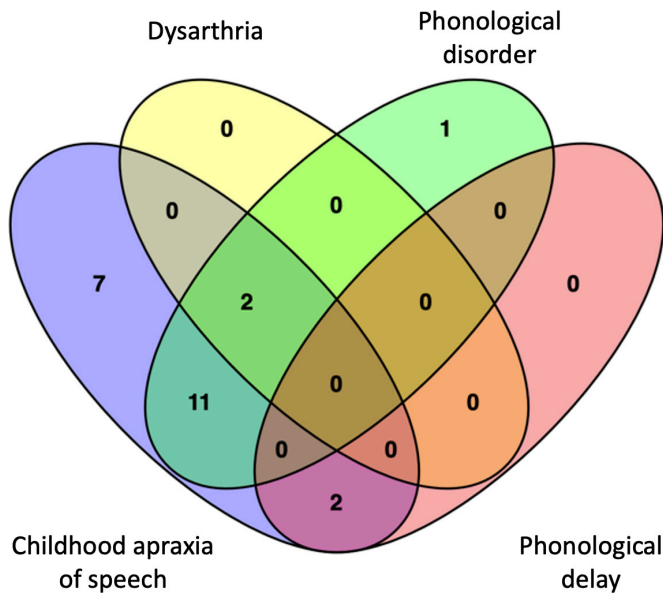


Figure 2 Speech disorders of participants with pathogenic missense/loss-of-function variants disrupting *FOXP2* (n=25, two participants had no speech disorder).

CAS was present in most English (86.7%, 13/15) and German (87.5%, 7/8) speakers.

Some participants had multiple co-occurring speech sound disorders (figure 2). Dysarthria (8.3%, 2/24) was infrequent and mild in severity, characterised as mixed nasality and harsh vocal quality (IDs 4a, 5). Phonological disorder was common (58.3%, 14/24), especially in children (<18 years); although for some (IDs 4a, 4b) this persisted into adulthood (table 3). A further two adults had a severe phonological delay, that is, typical phonological error patterns that appear in the speech of younger individuals but that should have resolved by 7 years of age (online supplemental table 4a; IDs 1b, 1c). Disordered oral motor movements were present both on speech (eg, say ‘pataka’) and non-speech tasks (eg, bite then blow) (92.9%, 13/14).

Two adults and one child did not have signs of CAS at the time of testing (IDs 3a, 3b, 11a). Participant 11a had a speech and language disorder in childhood for which they received therapy, but could not recall having CAS. The other adult participant (ID 3b) reported being a ‘quiet’ child but did not have speech therapy, and his son (ID 3a) had a phonological disorder without CAS.

Phonetic inventories were analysed for 12 English-speaking participants (online supplemental table 4a,b). Strikingly, 66.7% (8/12) of English-speaking participants did not have the affricate /dʒ/ (eg, ‘j’ in ‘jump’) and 58.3% (7/12) did not have its voiceless counterpart, /tʃ/ (eg, ‘ch’ in ‘chair’). Many were also missing the later developing sounds of /r/ (eg, ‘r’ in ‘rabbit’, 66.7%, 8/12), /θ/ (eg, voiceless ‘th’ in ‘thin’, 58.3%, 7/12) and /ð/ (eg, voiced ‘th’ in ‘this’, 58.3%, 7/12) (online supplemental table 4a,b). Other phonemes absent in some English speakers’ inventories were /ʃ/ (‘sh’, 41.7%, 5/12), /ŋ/ (‘ng’, 41.7%, 5/12), /l/ (27.3%, 3/11) and /s/ (27.3%, 3/11). The phonemes /dʒ/ and /tʃ/ were not present in most German participants (4/6).

Average intelligibility for children, assessed via the ICS, ranged from ‘never’ understood (20%, 2/10), to ‘rarely’ (33.3%, 3/10) to ‘sometimes’ understood (50%, 5/10). For adults, average intelligibility ranged from ‘sometimes’ (16.7%, 1/6) to ‘usually’ understood (66.7%, 4/6). Only one participant was ‘always’ understood (ID 3b).

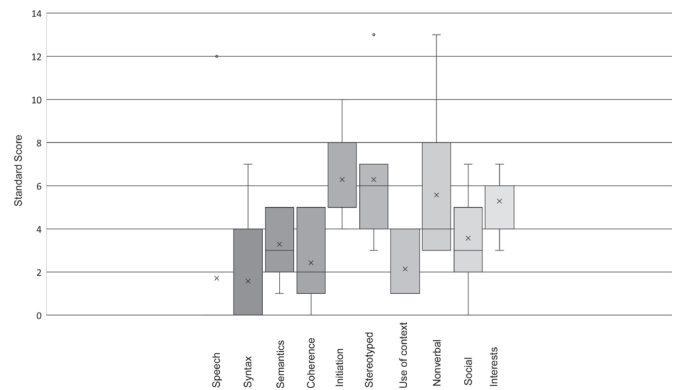


Figure 3 Children’s Communication Checklist subdomains⁶¹ in participants with pathogenic *FOXP2* variants (n=7, average=10, SD=3). Scores ≤6 are within normal limits, and scores <5 are low. Line=median, x=average, •=outlier.

Language and literacy

More than half of the cohort (56%, 14/25) had mild to severe receptive vocabulary impairment (table 3; online supplemental figure 2). Receptive grammar was commonly affected (72.2%, 13/18) ranging from moderate to severely impaired. Expressive vocabulary (83.3%, 15/18) and expressive grammar impairments were also common (86.7%, 13/15).

Speech was the most severely affected communication domain (mean score=1.7) for the Children’s Communication Checklist, Second Edition (CCC-2) (completed for IDs 1a, 2, 3a, 5, 6, 15, 16b; figure 3, normative mean=10, SD=3).

Five participants were minimally verbal (<30 words; IDs 9, 12a, 13, 14, 15; between 2 and 18 years old). Three of the five used high-tech Augmentative and Alternative Communication (AAC) devices (eg, a speech-generating application on a tablet), while the remaining two used some sign language.

Spelling (17/24) and reading (17/24) impairments were common (table 3). This was reflected in the results from the VABS-3 (written subdomain mean=9.25, normative mean=15, SD=2). Four participants were at the preliteracy stage (<5 years of age).

Adaptive behaviour and cognition

Receptive and expressive language performance was low in those assessed (n=11, mean=9.5 in both domains; online supplemental figure 3) and not significantly different between the two domains (p=0.79). Language performance (mean=70.73) was significantly different from (p=0.01) and higher than socialisation (mean=61.18) and daily living (mean=57.45). Motor skills were an area of relative strength, although impaired compared with norms (mean=77.18), although the sample size was small and results should be interpreted with caution. Further, normative data for motor skills are only available up to 9 years 11 months; however, none of the older participants reached the ceiling on motor skills.

Language skills were incongruent with intellectual skills for many, as seven participants with severe language impairment scored within the average range on non-verbal subtests (table 3). IQ was formally assessed in 20 participants (FSIQ n=10, non-verbal IQ n=10). In the non-verbal testing, seven performed in the average range, one was borderline and two were moderately impaired. For an FSIQ, two performed within the average range, four in the borderline range (70–85 IQ) and four had a mild intellectual disability (50–69 IQ).

DISCUSSION

We systematically delineated the speech and cognitive phenotype in 28 probands from 17 unrelated families (10 of which are novel) with heterozygous pathogenic missense/loss-of-function variants disrupting *FOXP2*, and completed the first cross-linguistic analysis of this disorder. Our data confirm aberrant speech and language development as a central feature. While speech presentation improves over time, with a reduction in CAS severity and improvement in phonological production, the disorder is characterised by impaired speech intelligibility that persists into adulthood, with most adults in our cohort being understood only ‘sometimes’ or ‘usually’ (rather than ‘always’) understood.

In terms of intellectual ability, scores ranged from below average to average in our cohort. For individuals with FSIQ data available, most (8/10) were scored as having borderline or mild intellectual disability, while for those who had only non-verbal IQ data available, most (7/10) were average. This range and profile are in line with previous findings for individuals with pathogenic SNVs/indels in *FOXP2*, with most individuals falling in the low average range and below for FSIQ and non-verbal IQ. The critical point of note here is that FSIQ takes into account the language metric of vocabulary knowledge, hence why FSIQ is generally more impaired than non-verbal IQ skills for individuals with this speech and language phenotype. At the same time, we also confirm observations from prior literature that the profile of this disorder differs from classic intellectual disability syndromes, in that severe speech and/or language impairments can occur against a background of non-verbal cognition within the normal range²⁹ as observed for seven of our probands with available data.

ASD has previously been reported in only a small number of individuals carrying pathogenic SNVs/indels of *FOXP2* (n=2/46; online supplemental table 1). The findings in our cohort indicate that there may be a higher prevalence of ASD in this disorder (25.9% of our cohort) than in the general population (~1%–2%),³⁰ although further research is needed to account for the discrepancy between our current findings and the prior literature. Of note, pathogenic variants in the closely related orthologue *FOXP1* are known to substantially increase the risk for ASD.³¹ Common non-coding polymorphisms in introns of *FOXP2* have shown associations with ADHD in large-scale genome-wide association studies, in the context of a multifactorial framework.³² The current study clearly shows that, by contrast, high-penetrance SNVs/indels disrupting this same locus do not yield elevated susceptibility to ADHD, with a prevalence in our cohort (1/27=4%) that is similar to that in the general population.³³

Sleep disturbances were common in our cohort and have been previously associated with idiopathic CAS³⁴ and other neurodevelopmental disorders, such as ASD, intellectual disability and ADHD.³⁵ The aetiology of sleep problems in such disorders is currently unknown, but they are posited to have biological and psychopathological causes. Although ASD, intellectual disability and ADHD were present here to varying degrees (as discussed above), within our cohort, sleep disturbance is also noted in children without those diagnoses.

We provide novel insights into other clinical diagnoses of mental health conditions that might be associated with pathogenic *FOXP2* variants. In particular, anxiety (19%) and depression (22%) had a higher prevalence than in the general population (between 2% and 4%)³³ and than in other neurodevelopmental disorders which were also present in our cohort,

such as mild intellectual disability (~3%–4%).³⁶ Anxiety has previously been associated with idiopathic CAS,^{37,38} and speech and language disorders are known to have possible negative impacts on mental health.^{37–40} It is difficult to ascertain whether mental health disorders are part of the phenotypic spectrum due to pathogenic *FOXP2* variants, or occur as a secondary consequence of the communication deficits experienced by affected individuals, as is seen in other speech disorders, such as stuttering.⁴¹ All participants with anxiety and depression were older than 16 years old, perhaps indicating that these mental health conditions arise later in life due to the impact of the communication impairment.

Gross motor impairment is thought to be relatively uncommon in individuals with pathogenic SNVs/indels disrupting *FOXP2*.⁴² However, two-thirds of the assessed participants in this study indicated having difficulty with gross motor skills during development. *FOXP2* disruption therefore appears to impact brain circuits involved in fine as well as gross motor development. Gross motor skill learning deficits have been identified in knock-out animal models.⁴³

We did not find convincing evidence of a dysmorphism phenotype in individuals with pathogenic *FOXP2* variants. Although physical features were noted for 81% of participants, most were minor and only shared among individuals from the same family. There was no consistent pattern of morphology seen across multiple unrelated probands in the cohort.

Regardless of the associated developmental features noted here, CAS was the most striking and consistent phenotypic characteristic in the present cohort. Dysarthria was far less common than CAS, clarifying the role of *FOXP2* in the planning and programming of movement sequences, as supported by animal models.⁴³ Adult participants typically had more intact, although incomplete, phonetic inventories than younger participants. In our study, more than half of all English-speaking participants with pathogenic missense/loss-of-function variants in *FOXP2* were missing one or more of the phonemes: /tʃ, dʒ, ɹ, θ, ð/ from their inventory. Three of these phonemes (/ɹ, θ, ð/) are in the ‘late eight’ sounds of English speech development, while affricates /tʃ, dʒ/ sit within the ‘middle eight’, referring to whether they are acquired earlier or later during typical phoneme acquisition.⁴⁴

There may be a window for plasticity and acquisition of new phonemes. Children with speech sound disorders are more likely to have persistent speech error patterns if these are not resolved by 8½ years old.⁴⁴ We might speculate that *FOXP2* dysfunction has the greatest impact between 2 and 7 years old when most phonemes are acquired.^{45,46} Intriguingly, neural expression of orthologues of *FOXP2* in model organisms has been shown to vary during different periods of vocal development,^{47–49} for example being upregulated in parts of the brain of the male Zebra finch during a developmental window that is important for vocal learning.⁵⁰ Reduced expression of this gene in mice alters the development and continuing plasticity of neuronal networks,⁵¹ impairs synaptic plasticity in striatal and cerebellar circuits and affects the learning of motor skills.^{52,53} Perhaps, the lack of acquisition of ‘late eight’ sounds and affricates in children with *FOXP2* disruptions may relate to the closing of the relevant developmental window. Other theories to explain the lack of acquisition of these phonemes include reduced functional load and ambient frequency for these phonemes⁵⁴ or the motoric complexity of these sounds⁵⁵ which rely heavily on tongue coordination and movement, known to be impaired in children with CAS.^{55–57} Further research is required to disentangle these relationships.

Speech impairment in the three participants without CAS was minimal or even absent, contrasting with unaffected, previously reported cases of *FOXP2* disruption.¹³ The speech of participant 17 was also less impaired than that of other children, although he had a diagnosis of CAS. Participant 11a was not referred for genetic testing for his speech, but rather to determine whether the variant in his child was *de novo* or inherited. Of note, family 3 and participant 17 were the only participants who had not been referred for testing primarily on the basis of speech and learning impairments. Thus, it is possible that there is a broader range of speech phenotypes associated with pathogenic *FOXP2* variants, and that individuals with milder presentations are unlikely to be referred for genetic testing.

Although there was no statistically significant difference between receptive and expressive language as reported by caregivers, other standardised tests indicated receptive language was more intact than expressive language. Literacy skills were also low across the cohort, in line with the high rate of literacy challenges for individuals with idiopathic CAS.⁵⁸

The speech domain on the CCC-2 confirmed that speech was the most severely impaired form of communication in children. AAC systems should be considered for children with pathogenic *FOXP2* variants due to the protracted speech development and severe speech impairment which persists for many throughout their lifetime.

We were unable to identify any clear phenotype–genotype correlation in the present cohort as we did not have sufficient power due to too few cases of missense and loss-of-function variants. The severity of speech and language disorder differed even among individuals with the same *FOXP2* variant in the same family. Family 3 had a relatively mild presentation compared with the other individuals in the study, despite having an intragenic deletion encompassing all *FOXP2* exons. Participant 17 had a translational start-site variant and a mild phenotype, with a larger phonemic repertoire and expressive vocabulary than other participants of similar ages in the cohort. This variant may not cause a clear-cut loss of function since there are alternative transcription start sites, potentially leading to a shorter protein.

In conclusion, CAS and language impairments are the most discernable features associated with heterozygous pathogenic missense/loss-of-function variants disrupting *FOXP2*. We also provide the evidence of additional neurodevelopmental features in subsets of our cohort, such as mild intellectual disability, ASD, anxiety, depression and sleep disturbances. There appear to be no distinctive physical features consistently associated with *FOXP2* disruptions. The phenotype associated with pathogenic variants that directly disrupt *FOXP2* remains relatively specific to speech disorder, compared with phenotypes associated with other monogenic conditions involving CAS.⁵⁸ Thus, our findings demonstrate that *FOXP2* provides an especially valuable entry point for examining the neurobiological bases of speech disorder.

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Data availability statement Data are available on reasonable request. All data relevant to the study are included in the article or uploaded as supplementary information. The datasets generated and analysed during this study are not publicly available because participants have not given permission for data to be made public but may be requested from the corresponding author (AM) who could go back to the participants to request data sharing. Genotypic data were submitted to Decipher (<https://decipher.sanger.ac.uk/>).

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REFERENCES

- Lai CS, Fisher SE, Hurst JA, Vargha-Khadem F, Monaco AP. A forkhead-domain gene is mutated in a severe speech and language disorder. *Nature* 2001;413:519–23.
- ASHA. Childhood apraxia of speech. secondary childhood apraxia of speech, 2007. Available: <https://www2.asha.org/articlesummary.aspx?id=8589947136>
- MacDermot KD, Bonora E, Sykes N, Coupe A-M, Lai CSL, Vernes SC, Vargha-Khadem F, McKenzie F, Smith RL, Monaco AP, Fisher SE. Identification of FOXP2 truncation as a novel cause of developmental speech and language deficits. *Am J Hum Genet* 2005;76:1074–80.
- Turner SJ, Hildebrand MS, Block S, Damiano J, Fahey M, Reilly S, Bahló M, Scheffer IE, Morgan AT. Small intragenic deletion in FOXP2 associated with childhood apraxia of speech and dysarthria. *Am J Med Genet A* 2013;161A:2321–6.
- Reuter MS, Riess A, Moog U, Briggs TA, Chandler KE, Rauch A, Stampfer M, Steindl K, Gläser D, Joset P, Krumbiegel M, Rabe H, Schulte-Mattler U, Bauer P, Beck-Wödl S, Kohlhasse J, Reis A, Zweier C, DDD Study. Foxp2 variants in 14 individuals with developmental speech and language disorders broaden the mutational and clinical spectrum. *J Med Genet* 2017;54:64–72.
- Rieger M, Krumbiegel M, Reuter MS, Schützenberger A, Reis A, Zweier C. 7q31.2q31.31 deletion downstream of FOXP2 segregating in a family with speech and language disorder. *Am J Med Genet A* 2020;182:2737–41.
- Feuk L, Kalervo A, Lipsanen-Nyman M, Skaug J, Nakabayashi K, Finucane B, Hartung D, Innes M, Kerem B, Nowaczyk MJ, Rivlin J, Roberts W, Senman L, Summers A, Szatmari P, Wong V, Vincent JB, Zeesman S, Osborne LR, Cardy JO, Kere J, Scherer SW, Hannula-Jouppi K. Absence of a paternally inherited FOXP2 gene in developmental verbal dyspraxia. *Am J Hum Genet* 2006;79:965–72.
- Palka C, Alfonsi M, Mohn A, Cerbo R, Guanciali Franchi P, Fantasia D, Morizio E, Stuppia L, Calabrese G, Zori R, Chiarelli F, Palka G. Mosaic 7q31 deletion involving FOXP2 gene associated with language impairment. *Pediatrics* 2012;129:e183–8.
- Rice GM, Raca G, Jakielski KJ, Laffin JJ, Iyama-Kurtycz CM, Hartley SL, Sprague RE, Heintzelman AT, Shriberg LD. Phenotype of FOXP2 haploinsufficiency in a mother and son. *Am J Med Genet A* 2012;158A:174–81.
- Shriberg LD, Ballard KJ, Tomblin JB, Duffy JR, Odell KH, Williams CA. Speech, prosody, and voice characteristics of a mother and daughter with a 7;13 translocation affecting FOXP2. *J Speech Lang Hear Res* 2006;49:500–25.
- Tomblin JB, O'Brien M, Shriberg LD, Williams C, Murray J, Patil S, Bjork J, Anderson S, Ballard K. Language Features in a Mother and Daughter of a Chromosome 7;13 Translocation Involving FOXP2. *J Speech Lang Hear Res* 2009;52:1157–74.
- Zeesman S, Nowaczyk MJM, Teshima I, Roberts W, Cardy JO, Brian J, Senman L, Feuk L, Osborne LR, Scherer SW. Speech and language impairment and oromotor dyspraxia due to deletion of 7q31 that involves FOXP2. *Am J Med Genet A* 2006;140:509–14.
- Žilina O, Reimand T, Zjablovskaja P, Männik K, Männamaa M, Traat A, Puusepp-Benazzouz H, Kurg A, Ounap K. Maternally and paternally inherited deletion of 7q31 involving the FOXP2 gene in two families. *Am J Med Genet A* 2012;158A:254–6.
- Lennon PA, Cooper ML, Peiffer DA, Gunderson KL, Patel A, Peters S, Cheung SW, Bacino CA. Deletion of 7q31.1 supports involvement of FOXP2 in language impairment: clinical report and review. *Am J Med Genet A* 2007;143A:791–8.
- Becker M, Devanna P, Fisher SE, Vernes SC. A chromosomal rearrangement in a child with severe speech and language disorder separates FOXP2 from a functional enhancer. *Mol Cytogenet* 2015;8:69.
- Morgan A, Fisher SE, Scheffer I, Hildebrand M. FOXP2-related speech and language disorders. In: Pagon RAA MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, Bird TD, Fong, C T, Mefford HC, Smith RJH, Stephens K, eds. *GeneReviews*[Internet]. Seattle, Washington, 2017.
- Braden RO, Amor DJ, Fisher SE, Mei C, Myers CT, Mefford H, Gill D, Srivastava S, Swanson LC, Goel H, Scheffer IE, Morgan AT. Severe speech impairment is a distinguishing feature of FOXP1-related disorder. *Dev Med Child Neurol* 2021;63:1417–26.
- Morgan A, Braden R, Wong MMK, Colin E, Amor D, Liégeois F, Srivastava S, Vogel A, Bizaoui V, Ranguin K, Fisher SE, van Bon BW. Speech and language deficits are central to SETBP1 haploinsufficiency disorder. *Eur J Hum Genet* 2021;29:1216–25.
- Morison LD, Braden RO, Amor DJ, Brignell A, van Bon BWM, Morgan AT. Social motivation a relative strength in DYRK1A syndrome on a background of significant speech and language impairments. *Eur J Hum Genet* 2022;30:800–11.
- Pados BF, Thoyre SM, Park J. Age-based norm-reference values for the Child Oral and Motor Proficiency Scale. *Acta Paediatr* 2018;107:1427–32.
- Reid SM, Johnson HM, Reddihough DS. The drooling impact scale: a measure of the impact of drooling in children with developmental disabilities. *Dev Med Child Neurol* 2010;52:e23–8.
- Dodd B, Zhu H, Crosbie S, Holm A, Ozanne A. *Diagnostic evaluation of articulation and phonology (DEAP)*. Psychology Corporation, 2002.
- Goldman R, Fristoe M. *Goldman-Fristoe Test of Articulation [kit]*. American Guidance Service, 2000.
- Mei C, Fedorenko E, Amor DJ, Boys A, Hoeflin C, Carew P, Burgess T, Fisher SE, Morgan AT. Deep phenotyping of speech and language skills in individuals with 16p11.2 deletion. *Eur J Hum Genet* 2018;26:676–86.
- Duffy JR. *Motor speech disorders e-book: substrates, differential diagnosis, and management*. Elsevier Health Sciences, 2019.
- Robbins J, Klee T. Clinical assessment of oropharyngeal motor development in young children. *J Speech Hear Disord* 1987;52:271–7.
- McLeod S. Intelligibility in context scale: a parent-report screening tool translated into 60 languages. *Journal of Clinical Practice in Speech-Language Pathology* 2015;17:9–14.
- Sparrow SS, Cicchetti DV, Saulnier CA. *Vineland-3: vineland adaptive behavior scales*. PsychCorp, 2016.
- Watkins KE, Dronkers NF, Vargha-Khadem F. Behavioural analysis of an inherited speech and language disorder: comparison with acquired aphasia. *Brain* 2002;125:452–64.
- Baron-Cohen S, Scott FJ, Allison C, Williams J, Bolton P, Matthews FE, Brayne C. Prevalence of autism-spectrum conditions: UK school-based population study. *Br J Psychiatry* 2009;194:500–9.
- Bacon C, Rappold GA. The distinct and overlapping phenotypic spectra of FoxP1 and FOXP2 in cognitive disorders. *Hum Genet* 2012;131:1687–98.
- Ridler C. First genetic risk loci for ADHD identified. *Nat Rev Neurol* 2019;15:4.
- Exchange GHD. GBD results tool. secondary GBD results tool, 2019. Available: <https://ghdx.healthdata.org/gbd-results-tool>
- Teverovsky EG, Bickel JO, Feldman HM. Functional characteristics of children diagnosed with childhood apraxia of speech. *Disabil Rehabil* 2009;31:94–102.
- Schreck KA, Richdale AL. Sleep problems, behavior, and psychopathology in autism: inter-relationships across the lifespan. *Curr Opin Psychol* 2020;34:105–11.
- Cooper S-A, Smiley E, Morrison J, Williamson A, Allan L. An epidemiological investigation of affective disorders with a population-based cohort of 1023 adults with intellectual disabilities. *Psychol Med* 2007;37:873–82.
- Lewis BA, Benckek P, Tag J, Miller G, Freebairn L, Taylor HG, Iyengar SK, Stein CM. Psychosocial comorbidities in adolescents with histories of childhood apraxia of speech. *Am J Speech Lang Pathol* 2021;30:2572–88.
- Cassar C, McCabe P, Cumming S. "I still have issues with pronunciation of words": A mixed methods investigation of the psychosocial and speech effects of Childhood Apraxia of Speech in adults. *Int J Speech Lang Pathol* 2022;1–13.
- Fujiki M, Brinton B, Hart CH, Olsen J, Coombs M. Using measurement invariance to study social withdrawal in children with developmental language disorders. *Lang Speech Hear Serv Sch* 2019;50:253–66.
- Durkin K, Toseeb U, Botting N, Pickles A, Conti-Ramsden G. Social confidence in early adulthood among young people with and without a history of language impairment. *J Speech Lang Hear Res* 2017;60:1635–47.
- Smith KA, Iverach L, O'Brian S, Kefalianos E, Reilly S. Anxiety of children and adolescents who stutter: a review. *J Fluency Disord* 2014;40:22–34.

- 42 Morgan A, Ttofari Eecen K, Pezic A, Brommeyer K, Mei C, Eadie P, Reilly S, Dodd B. Who to Refer for Speech Therapy at 4 Years of Age Versus Who to "Watch and Wait"? *J Pediatr* 2017;185:200–4.
- 43 Fisher SE, Scharff C. Foxp2 as a molecular window into speech and language. *Trends Genet* 2009;25:166–77.
- 44 Shriberg LD. Four new speech and prosody-voice measures for genetics research and other studies in developmental phonological disorders. *J Speech Hear Res* 1993;36:105–40.
- 45 Dodd B, Holm A, Hua Z, Crosbie S. Phonological development: a normative study of British English-speaking children. *Clin Linguist Phon* 2003;17:617–43.
- 46 Shriberg LD, Gruber FA, Kwiatkowski J. Developmental phonological disorders. III: long-term speech-sound normalization. *J Speech Hear Res* 1994;37:1151–77.
- 47 Schulz SB, Haesler S, Scharff C, Rochefort C. Knockdown of FOXP2 alters spine density in area X of the zebra finch. *Genes Brain Behav* 2010;9:732–40.
- 48 Lai CSL, Gerrelli D, Monaco AP, Fisher SE, Copp AJ. Foxp2 expression during brain development coincides with adult sites of pathology in a severe speech and language disorder. *Brain* 2003;126:2455–62.
- 49 Bolhuis JJ, Okanoya K, Scharff C. Twitter evolution: converging mechanisms in birdsong and human speech. *Nat Rev Neurosci* 2010;11:747–59.
- 50 Haesler S, Wada K, Nshdejan A, Morrisey EE, Lints T, Jarvis ED, Scharff C. Foxp2 expression in avian vocal learners and non-learners. *J Neurosci* 2004;24:3164–75.
- 51 Vernes SC, Oliver PL, Spiteri E, Lockstone HE, Puliyadi R, Taylor JM, Ho J, Mombereau C, Brewer A, Lowy E, Nicod J, Groszer M, Baban D, Sahgal N, Cazier J-B, Ragoussis J, Davies KE, Geschwind DH, Fisher SE. Foxp2 regulates gene networks implicated in neurite outgrowth in the developing brain. *PLoS Genet* 2011;7:e1002145.
- 52 Groszer M, Keays DA, Deacon RMJ, de Bono JP, Prasad-Mulcare S, Gaub S, Baum MG, French CA, Nicod J, Coventry JA, Enard W, Fray M, Brown SDM, Nolan PM, Pääbo S, Channon KM, Costa RM, Eilers J, Ehret G, Rawlins JNP, Fisher SE. Impaired synaptic plasticity and motor learning in mice with a point mutation implicated in human speech deficits. *Curr Biol* 2008;18:354–62.
- 53 French CA, Jin X, Campbell TG, Gerfen E, Groszer M, Fisher SE, Costa RM. An aetiological FOXP2 mutation causes aberrant striatal activity and alters plasticity during skill learning. *Mol Psychiatry* 2012;17:1077–85.
- 54 Stokes SF, Surendran D. Articulatory complexity, ambient frequency, and functional load as predictors of consonant development in children. *J Speech Lang Hear Res* 2005;48:577–91.
- 55 Murdoch BE, Attard MD, Ozanne AE, Stokes PD. Impaired tongue strength and endurance in developmental verbal dyspraxia: a physiological analysis. *Eur J Disord Commun* 1995;30:51–64.
- 56 Terband H, Maassen B, van Lieshout P, Nijland L. Stability and composition of functional synergies for speech movements in children with developmental speech disorders. *J Commun Disord* 2011;44:59–74.
- 57 Bradford A, Murdoch B, Thompson E, Stokes P. Lip and tongue function in children with developmental speech disorders: a preliminary investigation. *Clin Linguist Phon* 1997;11:363–87.
- 58 Hildebrand MS, Jackson VE, Scerri TS, Van Reyk O, Coleman M, Braden RO, Turner S, Rigby KA, Boys A, Barton S, Webster R, Fahey M, Saunders K, Parry-Fielder B, Paxton G, Hayman M, Coman D, Goel H, Baxter A, Ma A, Davis N, Reilly S, Delatycki M, Liègeois FJ, Connelly A, Gez J, Fisher SE, Amor DJ, Scheffer IE, Bahlo M, Morgan AT. Severe childhood speech disorder: gene discovery highlights transcriptional dysregulation. *Neurology* 2020;94:e2148–67.
- 59 Dunn LM, Dunn DM. *PPVT-4: Peabody picture vocabulary test*. Pearson Assessments, 2007.
- 60 Fox-Boyer AV. *TROG-D: test Zur Überprüfung des Grammatikverständnisses*. Schulz-Kirchner Verlag GmbH, 2020.
- 61 Bishop D. Children's Communication Checklist (CCC-2). In: Volkmar FR, ed. *Encyclopedia of autism spectrum disorders*. 2nd edition. New York, NY: Springer, 2003.
- 62 Wechsler D. *Wechsler abbreviated scale of intelligence*. 2nd edn (WASI-II). San Antonio, TX: Pearson, 2011.
- 63 Semel EW, Secord W. *Clinical evaluation of language fundamentals preschool*. 2nd edn. San Antonio, TX: The Psychological Corporation, 2004.
- 64 Dumont R, Willis JO. *Kaufman assessment battery for children*. 2nd edn. Encyclopedia of Special Education, 2008: 1196–7.
- 65 Semel EW, Secord W. *Clinical evaluation of language fundamentals, Australian standardised edition*. 4th edn. Marrickville: Harcourt Assessment, 2006.
- 66 Wiig EH, Secord WA, Semel E. *Clinical evaluation of language fundamentals: CELF-5*. Pearson, 2013.
- 67 Wechsler D. *Wechsler intelligence scale for children*. 4th edn: WISC-IV. Pearson, 2013.
- 68 Wechsler D. *Wechsler intelligence scale for children*. 5th edn: WISC-V. Pearson, 2013.
- 69 Wechsler D. *Wechsler adult intelligence scale*. 4th edn (WAIS-IV). San Antonio, TX: NCS Pearson, 2008: 22. 816–27.
- 70 Martin NA, Bronwell R. *Expressive One-Word picture vocabulary tests*. 4th edn: ROWPVT. Pearson, 2010.
- 71 Snijders JT, Snijders-Oomen AWM, Tellegen PJ. *SON-R 21/2-7: Snijders-Oomen niet-verbale intelligentietest: handleiding en verantwoording*. Swets Test Publishers, 1998.
- 72 Wetherby AM, Prizant BM. *Communication and symbolic behavior scales: developmental profile*. Paul H Brookes Publishing Co, 2002.
- 73 Wechsler D. *Wechsler preschool and primary scale of intelligence*. 4th edn. San Antonio, TX: The Psychological Corporation, 2012.
- 74 McCallum RS, Bracken BA. *A multidimensional nonverbal alternative for cognitive assessment*. 2nd edn. The Universal Nonverbal Intelligence Test, 2018.