# **Aberrant expression of** *NPPB* **through YAP1 and TAZ activation in mesothelioma with Hippo pathway gene alterations**



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#### **Abstract**

**Background:** Mesothelioma is a neoplastic disease associated with asbestos exposure. It is highly malignant and has a poor prognosis; thus, early detection is desirable. Recent whole-genome analysis has revealed that mesothelioma is characterized by a high frequency of mutations in a set of genes involved in the Hippo pathway, such as *NF2* and *LATS2*. However, a rapid, simple, and precise method for finding mesothelioma with these mutations has not yet been established.

**Methods:** Clustering of Hippo pathway gene alteration groups and the differential expression of each gene in mesothelioma patients were analyzed using The Cancer Genome Atlas database. Gene expression levels in various tumors and normal tissues were analyzed using public databases. Knockdown or transient expression of *YAP1* or *TAZ* was performed to evaluate the regulation of gene expression by these genes. NT-proBNP was measured in the pleural effusions of 18 patients and was compared with NF2 expression in five cases where cell lines had been successfully established.

**Results:** *NPPB* mRNA expression was markedly higher in the group of mesothelioma patients with Hippo pathway gene mutations than in the group without them. *NPPB* expression was low in all normal tissues except heart, and was highest in mesothelioma. Mesothelioma patients in the high *NPPB* expression group had a significantly worse prognosis than those in the low *NPPB* expression group. *NPPB* expression was suppressed by knockdown of *YAP1* or *TAZ*. NT-proBNP was abundant in the effusions of mesothelioma patients and was particularly high in those with impaired NF2 expression.

**Conclusions:** *NPPB*, whose levels can be measured in pleural effusions of mesothelioma patients, has the potential to act as a biomarker to detect *NF2*-Hippo pathway gene alterations and/or predict patient prognosis. Additionally, it may provide useful molecular insights for a better understanding of mesothelioma pathogenesis and for the development of novel therapies.

#### **KEYWORDS**

BNP, Hippo pathway, mesothelioma, *NPPB*

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### **1** | **INTRODUCTION**

Mesothelioma is a highly aggressive tumor arising from the surface of serous membranes, such as the pleura, peritoneum, and pericardium. Asbestos exposure is a major risk factor for its development. Patients at advanced stages of the disease have a very poor prognosis, with a mean survival of about 12months, even after cisplatin/pemetrexed combination therapy.<sup>1</sup> Recent dual therapy with immune checkpoint inhibitors has also shown promise for improving patient outcomes,<sup>2</sup> however this is not yet conclusive. Although the use of asbestos has already been banned in many countries, including Japan, it is still used in developing countries, and in some countries there are only restrictions on its use, not bans.<sup>3</sup> Mesothelioma diagnosis utilizes immunohistochemical detection of mesothelioma-related markers, such as WT-1 and calretinin, in combination with cancer-related markers, such as CEA.<sup>3,4</sup> However, for anatomical reasons, and in the absence of specific clinical manifestations of mesothelioma, patients are often diagnosed only at advanced stages.

Genome-wide analyses have identified *CDKN2A*, *BAP1*, and *NF2* as genes frequently mutated in mesothelioma.[5,6](#page-11-1) Recent studies have reported that the detection of deletions at the 9p21 locus, where *CDKN2A* is located, by fluorescence in situ hybridization (FISH) and the detection of defective BAP1 expression by immunohistochemical staining can provide a diagnosis of mesothelioma with very high specificity. $4,7-10$  However, these techniques have not been fully proved for detecting *NF2* gene mutations and protein expression defects.<sup>[11](#page-11-3)</sup>

*NF2* functions as a tumor suppressor by positively regulating the Hippo signaling pathway[.12–15](#page-11-4) In addition to *NF2*, genetic alterations in *LATS2*, which encodes a component of the Hippo pathway, have been detected in mesothelioma, strongly supporting the idea that disruption of the Hippo pathway induces the development of mesothelioma. $12,16$ Molecularly, the Hippo pathway plays a role in promoting the nuclear export and degradation of the transcriptional coactivators YAP1 and TAZ, whereas disruption of the Hippo pathway increases the nuclear accumulation of these proteins. YAP1 and TAZ in the nucleus bind to TEAD family DNA-binding proteins (TEAD1–4) and activate the transcription of tumor-promoting genes. Previous studies have demonstrated that shRNA-mediated silencing of YAP1 or TAZ strongly suppresses cell growth in mesothelioma cells with *NF2* mutations, while expression of activating mutants of YAP1 or TAZ promotes tumorigenesis in normal immortalized mesothelial cells.<sup>17-19</sup> In recent years, several TEAD inhibitors that block YAP1 signaling have been successively developed. $20-22$  Experiments using mouse models have reported that TEAD inhibitors dramatically inhibit the growth of mesothelioma cells with Hippo pathway gene mutations.<sup>20</sup> Given this situation, it is necessary to develop

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easier and more reliable methods to assess Hippo pathway inactivation and YAP1 and TAZ activation in actual patient pleural fluid and tumor samples in order to appropriately administer these new drugs to mesothelioma patients.

In this study, we identified *NPPB* as a gene that was significantly upregulated in the samples from mesothelioma patients with Hippo pathway alterations. The transcript of *NPPB* is the brain natriuretic peptide (BNP) precursor, proBNP, which is further cleaved to produce BNP and NTproBNP. The *NPPB* gene product is known as BNP, which is one of the natriuretic peptides (NPs), including atrial NP (ANP, *NPPA* gene product) and C-type NPs (CNP, *NPPC* gene product). $23-26$  There are three known receptors for NP: NP receptor A (NPR-A, *NPR1* gene product), NP receptor B (NPR-B, *NPR2* gene product), and NP receptor C (NPR-C, *NPR3* gene product). BNP binds to NPR-A, but not NPR-B, and regulates various intracellular signals to exert its biological effects. NPR-C can bind to multiple NPs including BNP and functions as a clearance receptor.<sup>27,28</sup> NT-proBNP can be used as an alternative to BNP for measuring blood levels because it has no physiological activity but has a long half-life. BNP and NT-proBNP play a central role in the regulation of heart failure and are biomarkers that can be used to assess the severity of heart failure. $29-31$  They have also been found useful in the diagnosis and prognosis of acute coronary syndromes and stable ischemic heart disease.<sup>32,33</sup> Although high levels of BNP have been reported in pleural effusions from mesothelioma patients, $34$  little is known about why BNP is highly expressed and how it relates to clinical characteristics.

### **2** | **METHODS**

#### **2.1** | **Cell lines and pleural fluids**

HOMC-D4 cell lines established in our laboratory $35$  were used for assays at passages 10–20. They were deposited and are now available from RIKEN BioResource Center. NCI-H2052 and NCI-H2373 cells were purchased from the American Type Culture Collection. All cell lines were cultured in RPMI-1640 medium supplemented with  $10\%$  (v/v) fetal bovine serum (FBS) and 1% (v/v) antimicrobials at 37°C in a humidified incubator with  $5\%$  CO<sub>2</sub>. Pleural fluid samples from patients were collected and centrifuged at 3000rpm for 10min, and the supernatant was stored at −80°C until use. The research protocol was approved by the Ethics Committee of Aichi Cancer Center Research Institute.

### **2.2** | **Antibodies and reagents**

Anti-YAP1 (#14074), anti-phospho-YAP1 (Ser127) (#13008), anti-TAZ (#4883), and anti-TEAD1 (#12292) antibodies **13588 WILEY-Cancer Medicine** *CONSIDERAL* **EXATO ET AL.** 

were purchased from Cell Signaling Technology. FLAG-M2 and anti-β-actin (#A5441) antibodies were sourced from Sigma. K-975 was synthesized and purified to >99% purity by Namiki Shoji Co. Ltd.

### **2.3** | **Public databases**

The mRNA and protein expression levels and clinical data of patients with mesothelioma and other cancers were analyzed from The Cancer Genome Atlas (TCGA) dataset on the cBioportal website [\(https://www.cbiop](https://www.cbioportal.org/) [ortal.org/\)](https://www.cbioportal.org/). The mRNA expression levels of various cancer cell lines were analyzed from DepMap Public 22Q1 dataset from DepMap portal website [\(https://depmap.](https://depmap.org/portal/) [org/portal/\)](https://depmap.org/portal/), and the *NPPB* mRNA expression levels of each normal organ were analyzed from GTEx Analysis Release V8 in the GTEx data portal website ([https://](https://gtexportal.org/home/) [gtexportal.org/home/](https://gtexportal.org/home/)).

### **2.4** | **qRT-PCR**

Total RNA was prepared using an RNeasy Plus RNA extraction kit (Qiagen) according to the manufacturer's protocol. Random-primed, first-strand cDNA was synthesized from total RNA using the ReverTra Ace qPCR RT Kit (Toyobo). qRT-PCR analyses were performed in triplicate using the KAPA SYBR Fast qPCR kit (KAPA Biosystems) in the QuantStudio 3 system (Applied Biosystems). Relative gene expression normalized to the expression of β-actin as a reference gene was calculated using the  $\Delta\Delta$ Ct method with efficiency correction. The primer sequences used in this study were as follows: β-actin, 5′-CCAACCGCGAGAAG ATGA-3′ and 5′-CCAGAGGCGTACAGGGATAG-3′; *NPPB*, 5′-TCTGGCTGCTTTGGGAGGAAGA-3′ and 5′-CC TTGTGGAATCAGAAGCAGGTG-3′; *ANKRD1*, 5′-GTGG AAAAGCGAGAAACAACGA-3′ and 5′-TCAGGTTCTGG TTCCTTTACAACT-3′.

### **2.5** | **Western blotting**

Cells were lysed with  $1 \times$  SDS sample buffer (3% (w/v) SDS, 5% (v/v) glycerol, 62mM Tris–HCl [pH6.8], 5% (v/v) 2-mercaptoethanol) and incubated at 95°C for 5min. The proteins were resolved by a 5%–12% gradient polyacrylamide gel (#192-15201; Fujifilm), transferred onto a polyvinylidene fluoride membrane (#IPVH00010; Merck) and probed with a 1000-fold dilution of antibodies and horseradish peroxidase-conjugated secondary antibodies (Cell Signaling Technologies) in 5% (w/v) skim milk. Protein bands were detected using ECL Prime Detection Reagent (#RPN2236; GE Healthcare) and Amersham Imager 680 (GE Healthcare).

### **2.6** | **Enzyme-linked immunosorbent assay (ELISA)**

Cell culture medium was collected 48h after seeding the cells. The amount of BNP secreted from the cells into the culture medium was quantified using a Human BNP ELISA Kit (#ab193694, Abcam) according to the manufacturer's instructions. The absorbance was measured using SpectraMax iD3 (Molecular Devices LLC).

### **2.7** | **NT-proBNP measurement**

The amount of NT-proBNP in the supernatant of the pleural fluid was measured by a clinical laboratory (BML Inc.).

#### **2.8** | **Statistical analysis**

Comparisons between two groups of data obtained via qRT-PCR or ELISA analysis were performed using Student's *t*-test. The experiments were repeated at least three times. Statistical analysis of TCGA clinical data were performed using GraphPad Prism 9.

### **3** | **RESULTS**

### **3.1** | **Characteristics of mesothelioma with mutations in the Hippo pathway gene**

We first examined the clinical characteristics of mesothelioma with mutations in the Hippo pathway genes using data from 87 samples from the TCGA mesothelioma (TCGA-MESO) cohort. Since mutations in the *NF2*, *LATS1*, and *LATS2* genes are relatively frequent among all the genes related to the Hippo pathway in mesothelioma, $6$ patients with genetic alterations such as mutations, deletions, or gene fusions in these genes were designated as the Altered group (Hippo pathway gene alteration group) and analyzed in comparison with the other group (Unaltered group). 43% (35/82) of the patients were classified as the Altered group and had no significant differences with the other group in terms of cancer progression, sex, or total number of mutations (Figure [1A](#page-3-0)). Besides, increased *ANKRD1*, a well-known gene downstream of YAP1, was detected in the Altered group (Figure [1B\)](#page-3-0). Alterations in Hippo pathway genes are known to increase the amount of unphosphorylated YAP. Still, the results of our analysis

<span id="page-3-0"></span>**FIGURE 1** Characteristics of the Hippo pathway gene alteration group in The Cancer Genome Atlasmesothelioma cohort. (A) Oncoprint of genetic alterations in 87 patients with mesothelioma; the group of patients with alterations in either *NF2*, *LATS1*, or *LATS2* was classified as Altered group, and the other group as Unaltered group. In parentheses, the number of patients. Color differences indicate different types of genetic alterations. (B, C) Box plot showing gene expression levels of *ANKRD1* (B), protein phosphorylation of YAP1 at serine 127 and total YAP1 protein expression levels (C) in Altered and Unaltered groups. (D, E) Kaplan–Meier curves of overall survival in Altered and Unaltered groups (D) or in *ANKRD1* highexpressing and low-expressing groups (E).



did not reach statistical significance despite a trend toward a decrease in phosphorylated YAP in samples with alterations in Hippo pathway genes ( $p=0.0288$ ,  $q=0.754$ ). YAP1 and TAZ expression levels were not significantly different between the two groups, and there were no available data on phosphorylated TAZ (Figure [1C](#page-3-0)). Regarding patient prognosis, the median overall survival (OS) of the Altered and Unaltered groups was 13.4 and 23.3months, respectively (Figure [1D](#page-3-0)). There was no significant difference  $(p=0.0952)$  in OS between the two groups; still, there was a trend toward poorer prognosis in the Altered group. We further compared the OS of patient groups according to the expression levels of ANKRD1. Kaplan–Meier plot analysis results showed that the ANKRD1-High group had a significantly worse prognosis  $(p=0.0147)$  than the ANKRD1-Low group (Figure [1E](#page-3-0)). The discrepancy in the

results of these survival analyses indicates the difficulty of understanding the effects on the Hippo pathway based on only a few known genetic changes or single gene expression levels.

### **3.2** | **NPPB is highly expressed in mesothelioma with alterations in Hippo pathway genes**

Using the gene expression data obtained from the RNA-seq analysis, we further searched for genes whose expression levels differed between the Altered and Unaltered groups. A total of 68 genes were significantly upregulated in the Altered group (Figure [2A,](#page-3-1) Table [S1](#page-12-0)). These genes included *ANKRD1* and *CCND1*, [19](#page-11-14) which are known to be induced

<span id="page-3-1"></span>**FIGURE 2** High expression of *NPPB* in mesothelioma patients with Hippo gene alterations. (A) Volcano plot showing differentially expressed genes between Altered and Unaltered groups. Blue dots, genes with *q*-value >0.05. (B, C) Box plot showing gene expression levels of *NPPB* (B) and *NPR3* (C) in the Altered and Unaltered groups. (D) Box plot showing the expression levels of *NPPB* in 32 cancer types in TCGA database. Dots indicate outliers. (E) Violin plot showing the expression levels of *NPPB* in each of the human tissue types in the GTEx database. Outliers were eliminated.







<span id="page-6-0"></span>**FIGURE 3** Characteristics of groups divided according to *NPPB* expression levels in The Cancer Genome Atlas-mesothelioma (TCGA-MESO) cohort. (A) Oncoprint showing the genetic alterations of the patients in TCGA-MESO cohort with high and low *NPPB* expression. Color differences indicate different types of genetic alterations. (B–D) Dot plots showing the correlation between *NPPB* and *CLDN15* (B), *VIM* (C), or *CLDN15/VIM* (D) (C/V) gene expression values in TCGA-MESO cohort data. Red lines represent regression lines. (E, F) Kaplan– Meier curves of overall survival (E) and progression-free survival (F) in *NPPB* High and *NPPB* Low groups.

by YAP1 activation. Notably, expression of *NPPB*, which encodes a BNP, a powerful biomarker for heart failure diagnosis, was more than 13-fold higher in the Altered group (Figure [2A,B](#page-3-1)). In addition, levels of *NPR3*, which encodes the NP clearance receptor, were 5-fold higher in the Altered group (Figure [2A,C](#page-3-1)). No differences in expression levels of genes encoding other NPs or NP receptors were found between the two groups (Figure [S1\)](#page-12-1). To investigate whether *NPPB* and *NPR3* are upregulated in other types of cancer besides mesothelioma, we analyzed data from 9279 TCGA tumors across 32 cancer types. *NPPB* expression was strongly suppressed in most cancer types, but strongly expressed in mesothelioma (Figure [2D\)](#page-3-1), while *NPR3* was highly expressed in other cancer types, including medulloblastoma (Figure [S2\)](#page-12-1). Consistently, cell lines derived from mesothelioma cell patients had higher *NPPB* expression levels than those derived from other cancer types (Figure [S3\)](#page-12-1). Notably, *NPPB* is rarely expressed in any normal tissues except the heart (Figure [2E\)](#page-3-1). Therefore, we conducted further studies focusing on the changes in *NPPB* expression.

### **3.3** | **Patients with mesothelioma expressing high levels of NPPB have a poor prognosis**

To gain a better understanding of the characteristics of the high-*NPPB* group, we divided the patients enrolled in TCGA-MESO cohort into two groups (High and Low) according to their *NPPB* expression levels (Figure [3A\)](#page-6-0). The genetic data showed that patients with alterations in the Hippo pathway or *CDKN2A* genes were significantly more common in the High group than in the Low group  $(p=0.0003$  and 0.0002, respectively; Table [1](#page-6-1)). On the other hand, patients with *BAP1* mutations were not statistically different between the High and Low groups.

There were no significant differences between the two groups in terms of clinical characteristics, including sex, pathology, and cancer progression (Table [1](#page-6-1)). Both sarcomatoid tumors in the histological classification were part of the *NPPB* High group, but this showed no statistically significant difference owing to there being too few cases. Remarkably, however, *CLDN15* expression, which is high in epithelioid mesothelioma, was significantly negatively correlated with *NPPB* expression (Figure [3B\)](#page-6-0). Furthermore, *NPPB* expression showed a significant positive correlation with that of *VIM*, which is upregulated <span id="page-6-1"></span>**TABLE 1** Clinical characteristics of groups divided according to *NPPB* expression levels in The Cancer Genome Atlasmesothelioma cohort.



Hippo gene mutation indicates genetic alterations in one or more of the NF2, LATS1, or LATS2 genes. The *p*-value for statistically significant term is indicated in bold with an asterisk.

in sarcomatoid mesothelioma (Figure [3C\)](#page-6-0). *CLDN15*/*VIM* expression values, which show a gradient in epithelial to sarcomatoid subtypes, also showed a significant negative correlation with *NPPB* expression (Figure [3D](#page-6-0)). Kaplan– Meier analysis of patient survival showed that the median OS in the Low *NPPB* group was 26.1months (95% CI, 19.4–31.2) whereas that in the high *NPPB* group was 12.7months (95% CI, 9.3–15.1; Figure [3E](#page-6-0)). Similarly, median PFS in the low *NPPB* group was 16.7months (95% CI, 13.3–26.2) whereas that in the high *NPPB* group was 10.3months (95% CI, 6.6–14.7) (Figure [3F](#page-6-0)).

### **3.4** | **NPPB transcription is promoted by the YAP1-TEAD complex**

We investigated whether high *NPPB* expression has a direct effect on tumor cell proliferation. We found that *NPPB* knockdown did not suppress proliferation in cells with either *NF2* (NCI-H2052, NCI-H2373) or *LATS2* (Y-MESO-27) gene alterations (Figure [S4A\)](#page-12-1). However, gene ontology analysis revealed numerous genes related to cell division and mitotic nuclei expressed differently between the High and Low *NPPB* expression groups (Figure [S4B\)](#page-12-1), suggesting that these variations may promote tumor growth.

In mesothelioma, cells with mutations in Hippo pathway genes, including *NF2*, YAP1, and TAZ are constitutively activated to enhance the expression of a series of

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genes involved in cell proliferation. Therefore, we performed knockdown experiments to determine whether *NPPB* is regulated by YAP1 and/or TAZ. Silencing YAP1 or TAZ expression in NCI-H2373 cells was performed by expressing shRNA against each gene (shYAP1 or shTAZ), and was confirmed via western blotting (Figure [4A,E\)](#page-7-0). Knockdown of *YAP1* or *TAZ* resulted in decreased *ANKRD1* mRNA expression compared to control cells (Figure [4B,F\)](#page-7-0). *NPPB* expression also decreased in YAP1 and TAZ knockdown cells (Figure [4C,G\)](#page-7-0). Furthermore, BNP secretion in the cell culture medium also decreased in knockdown cells compared to control cells (Figure [4D,H](#page-7-0)).

To further investigate whether YAP1 activation is sufficient to increase *NPPB* expression, FLAG-tagged wildtype YAP1 or an activated YAP1 mutant (YAP1 S127A) was overexpressed in HOMC-D4 normal immortalized mesothelial cells. Each YAP1 protein was expressed to the same extent, as confirmed with western blotting using FLAG antibody (Figure [5A](#page-8-0)). Phosphorylated YAP1, an indicator of YAP1 inactivation, was also detected, confirming the activation of the S127A mutant. Wild-type YAP1 and activated YAP1 mutants significantly increased mRNA levels of both *ANKRD1* and *NPPB* (Figure [5B,C\)](#page-8-0). Also, YAP1 S127A promoted extracellular BNP secretion more strongly than YAP1 WT (Figure [5D](#page-8-0)). These results indicate that, in mesothelioma cells, *NPPB* expression is regulated by YAP1 and TAZ.



<span id="page-7-0"></span>**FIGURE 4** Reduced *NPPB* expression following YAP1 or TAZ knockdown. (A) NCI-H2373 cells expressing shRNA targeting YAP1 (shYAP1 #1 and #2) or Scramble shRNA (shScr) as a control were generated using lentivirus and subjected to western blotting. (B, C) The expression ratio of *ANKRD1* (B) or *NPPB* (C) in each cell was measured via qRT-PCR. (D) The culture medium of NCI-H2373 cells expressing shYAP1 or shScr was subjected to enzyme-linked immunosorbent assay (ELISA) to assess the amount of secreted brain natriuretic peptide (BNP). (E–H) Using NCI-H2373 cells expressing shTAZ (shTAZ #1, #2) or shScr, the same experiments as described in A–D were performed.



<span id="page-8-0"></span>**FIGURE 5** Induction of *NPPB* expression through YAP1 activation. (A) NCI-H2373 cells or the cells expressing wild-type YAP1 (YAP1 WT) or constitutively active YAP1 mutant (YAP1 S127A) fused with FLAG tag were generated using lentivirus and subjected to western blotting. (B, C) The expression ratio of *ANKRD1* (B) or *NPPB* (C) in each cell was measured via qRT-PCR. (D) The culture medium of NCI-H2373 cells or the cells expressing YAP1 was subjected to enzyme-linked immunosorbent assay (ELISA) to assess the amount of secreted brain natriuretic peptide (BNP).

### **3.5** | **Measurement of NT-proBNP may be a useful biomarker for mesothelioma treatment**

Since YAP1 and TAZ are potential therapeutic targets, detection of BNP released from mesothelioma cells is expected to be a predictive marker of therapeutic response. Therefore, we measured BNP secretion as NT-proBNP levels in pleural effusions collected from 18 mesothelioma patients. NT-proBNP levels were widely distributed as 18– 4841pg/mL, with a median of 212.5pg/mL (Figure [6A](#page-9-0)). In addition, five mesothelioma cell lines had been established from these effusions: three with complete NF2 inactivation, one with relative NF2 downregulation with undetermined mechanism, and one with NF2 intact (Figure [6A\)](#page-9-0). In samples with normal NF2 expression, pleural fluid NTproBNP levels were below median; and in samples with low or undetectable NF2 expression, levels were above the median. Although statistical tests could not be performed due to the small number of cell lines established, the results suggest that mesotheliomas with Hippo pathway gene mutations may secrete high amounts of NT-proBNP in accordance with their gene expression levels.

Furthermore, we examined the possibility that increased *NPPB* expression may be a suitable biomarker for tumor detection and evaluation of therapeutic efficacy. The recently developed TEAD inhibitor K-975 suppresses the transcriptional activity of YAP1 and TAZ and potently inhibits the growth of mesothelioma with Hippo pathway gene mutations both in vitro and in vivo. To examine whether *NPPB* could serve as a marker to monitor the therapeutic response to K-975, we measured *NPPB* expression in mesothelioma cells before and after K-975 treatment. The expression of *NPPB* mRNA was more strongly suppressed than that of *ANKRD1* in cells treated with K-975 (Figure [6B,C](#page-9-0)), suggesting that monitoring the expression levels of *NPPB* or its gene products may be an indicator of inhibitor efficacy.

Finally, we searched for genes whose expressions correlated with *NPPB* to investigate the potential of *NPPB* for clinical use in mesothelioma patients. Among 19,782 genes in the TCGA-MESO database, *OXTR* (Spearman's correlation coefficient 0.66,  $p < 0.001$ ) was identified as the gene with the highest positive correlation (Figure [6D](#page-9-0)). *OXTR* is highly expressed in mesothelioma and reported as a potential therapeutic target. $36$  Among other genes with significant correlations, *AURKA* was also found (Spearman's correlation coefficient 0.49, *p*<0.001) (Figure [6E\)](#page-9-0). The *AURKA* transcript, aurora kinase, is a known target of the anticancer drug alisertib, which is currently in clinical trials for mesothelioma patients. Since the expression of aurora kinase is strongly correlated with that of *MYC*, [37](#page-11-16) it was suggested that *MYC* copy number could be a predictor of susceptibility to alisertib. However, Phase II clinical trials have recently revealed that *MYC* copy number increase or gene amplification does not correlate with response rate or improved patient outcome. $38$  Our data present the possibility of *NPPB* or NT-proBNP replacing *MYC* as a new marker to predict sensitivity to aurora kinase inhibitors.

#### **4** | **DISCUSSION**

In this study, we found that *NPPB* is most highly expressed in mesothelioma among 32 cancer types, and that its expression is induced in the mesothelioma (Altered) group with *NF2* or *LATS1/2* gene mutations. These results are consistent with previous studies reporting significantly higher BNP levels in pleural fluid from mesothelioma patients than in pleural fluid from other cancer patients such as lung cancer patients. $34$  A previous ChIP analysis

<span id="page-9-0"></span>**FIGURE 6** Exploring the clinical value of *NPPB* expression levels. (A) NT-proBNP levels were measured using pleural fluid collected from 18 patients. The NF2 expression ratios established from the pleural fluid in five cell lines are shown in color. (B, C) After purification of mRNA from cells treated with or without K-975, the expression ratios of *ANKRD1* (B) and *NPPB* (C) were measured via qRT-PCR. (D, E) Dot plot showing the expression correlations of the indicated genes using The Cancer Genome Atlasmesothelioma cohort data. Red lines represent regression lines.



using glioblastoma cells suggested transcriptional activation by YAP1 as a molecular mechanism for *NPPB* high expression. $39$  In the present study, we also showed that YAP1 and TAZ enhance *NPPB* transcription in *NF2* deficient cells. These findings will shed new light on the development of detection methods and therapeutics for mesothelioma.

It is worth noting that pathway-based approaches have their limitations: the molecules involved in the Hippo signaling pathway and their regulators are not yet well defined and thus, it is difficult to fully understand the impact of known genetic variants on the pathway. Protein array data may also not fully capture pathway activity due to the limited number of molecules analyzed and incomplete pathway coverage compared to that of other high-throughput technologies. In this regard, expression of BNP and NT-proBNP is expected to be a potentially useful additional tool estimating YAP1 and TAZ activation in tumors. Although no effective drugs have yet been developed for mesothelioma patients with Hippo pathway mutations or YAP1/TAZ hyperactivation, a number of pan-TEAD inhibitors that block YAP1/TAZ activity and compounds that act only on specific TEAD proteins have recently been reported. $20,40,41$ Furthermore, a Phase 1, first-in-human clinical trial has been initiated to investigate the efficacy of these agents in mesothelioma patients with *NF2* mutations ([Clini](http://clinicaltrials.gov) [calTrials.gov](http://clinicaltrials.gov) Identifier: NCT04665206, NCT05228015). Thus, although it will need to be evaluated in future clinical trials, measurement of BNP and NT-proBNP expression is expected to be a suitable biomarker for predicting the response to these drugs.

It should be noted, that increased expression of BNP and NT-proBNP is observed in heart failure; however, in the present study, we were unable to collect sufficient clinical information on cardiovascular function from the patients. Previous reports have shown that BNP levels are elevated in plasma and pleural fluid in mesothelioma patients. However, there are differences between mesothelioma and heart failure patients. Specifically, in heart failure patients, BNP levels in plasma are several times higher than in pleural fluid; conversely, in mesothelioma patients, BNP levels are higher in pleural fluid than in plasma. $34$  This difference may be caused by local tumor BNP production in mesothelioma tissue. Still, the biological consequences of BNP entering the blood from the pleural fluid need to be carefully examined in future studies.

Analysis of the TCGA-MESO dataset in this study revealed for the first time that mesothelioma patients with high *NPPB* expression had a much poorer prognosis than those with low *NPPB* expression. On the other hand, when mesothelioma patients were divided into two groups (with and without Hippo pathway gene alterations), there was no significant difference in prognosis between them. This suggests that *NPPB* may play an important function in tumorigenesis and malignant transformation independently of *NF2* or *LATS1/2* gene alterations. However, our experiments did not confirm that *NPPB* promotes mesothelioma cell proliferation directly. In this regard, it is possible that *NPPB* expression exerts a coordinated effect not only on its own cells but also on surrounding cancer cells. The *NPPB* gene product, BNP, has been reported to exert paracrine effects on

the proliferation of surrounding fibroblasts after being secreted out of cells. $42$  Since the tumor microenvironment has important prognostic implications for mesothelioma patients, $43$  the effects of BNP released outside mesothelioma cells on immune cells and peritumoral cells should be investigated in the future. In addition, since the *CDKN2A* mutation in mesothelioma patients is known to show a worse prognosis, *CDKN2A* inactivation may also be associated with *NPPB* expression.

Immunohistochemistry (IHC) is the most common method to detect aberrant expression of a particular protein. However, reliable detection of disturbed NF2 expression in mesothelioma using this technique is still under development.<sup>11</sup> Instead, the FISH method, which detects gene deletions, has been reported to show good sensitivity and specificity in detecting loss of *NF2* in mesothelioma.<sup>44</sup> Nevertheless, FISH still has some technical challenges, such as an ambiguous cutoff value used to evaluate the signal. The mechanisms of gene inactivation in mesothelioma are diverse, including genetic and epigenetic events.[45](#page-12-7) Detection of the *NPPB* gene product using IHC may help detect mesothelioma cells that have genetic alterations in Hippo pathway genes, including *NF2* and *LATS1/2*, or that have YAP1 and TAZ activated by these mutations.

It should also be noted that in the present analysis, it is still unclear whether changes in Hippo pathway genes or *NPPB* mRNA expression correlate with the intracellular synthesis or extracellular secretion of NP-proBNP in pleural effusions. In Figure [6,](#page-9-0) we show that NT-proBNP in the original pleural fluid was elevated in four of the five established cell lines with *NF2* alterations, suggesting that *NF2* alterations may result in elevated NT-proBNP, albeit in a small number of cases. Accordingly, we have purchased several commercial antibodies and tried to quantify BNP via IHC; however, we are yet to find one capable of estimating BNP expression levels. Developing suitable antibodies for IHC and validating their clinical usefulness will be one of the next objectives of our research.

## **5** | **CONCLUSIONS**

We found that *NPPB* transcription and extracellular secretion of its gene products, BNP and NT-proBNP, were enhanced in mesothelioma patients with Hippo pathway gene alterations. Thus, our findings might contribute to the prediction of the efficacy of new therapeutic strategies that target Hippo signaling alteration. High expression of *NPPB* was also associated with poor patient prognosis. Further studies are needed to determine the impact of *NPPB* on mesothelioma, which will shed further light on

the molecular mechanisms of mesothelioma pathogenesis and malignant transformation.

### **AUTHOR CONTRIBUTIONS**

**Tatsuhiro Sato:** Conceptualization (lead); data curation (lead); formal analysis (lead); funding acquisition (equal); investigation (lead); project administration (lead); visualization (lead); writing – original draft (lead); writing – review and editing (lead). **Ken Akao:** Investigation (supporting); validation (supporting). **Ayuko Sato:** Investigation (supporting); writing – review and editing (supporting). **Tohru Tsujimura:** Investigation (supporting); writing – review and editing (supporting). **Satomi Mukai:** Investigation (supporting); writing – review and editing (supporting). **Yoshitaka Sekido:** Funding acquisition (equal); project administration (supporting); resources (lead); supervision (lead); writing – review and editing (supporting).

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#### **CONFLICT OF INTEREST STATEMENT**

The authors declare that they have no competing interests.

#### **DATA AVAILABILITY STATEMENT**

Data supporting the results of this study are available from the corresponding author upon reasonable request.

#### **ETHICS STATEMENT**

All the protocols for research were approved by the Aichi Cancer Center Ethics Committee.

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#### **REFERENCES**

- <span id="page-10-0"></span>1. Vogelzang NJ, Rusthoven JJ, Symanowski J, et al. Phase III study of pemetrexed in combination with cisplatin versus cisplatin alone in patients with malignant pleural mesothelioma. *J Clin Oncol*. 2003;21(14):2636-2644.
- <span id="page-10-1"></span>2. Baas P, Scherpereel A, Nowak AK, et al. First-line nivolumab plus ipilimumab in unresectable malignant pleural mesothelioma (CheckMate 743): a multicentre, randomised, open-label, phase 3 trial. *Lancet*. 2021;397(10272):375-386.

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- <span id="page-11-0"></span>3. Popat S, Baas P, Faivre-Finn C, et al. Malignant pleural mesothelioma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2022;33(2):129-142.
- <span id="page-11-2"></span>4. Husain AN, Colby TV, Ordonez NG, et al. Guidelines for pathologic diagnosis of malignant mesothelioma 2017 update of the consensus statement from the International Mesothelioma Interest Group. *Arch Pathol Lab Med*. 2018;142(1):89-108.
- <span id="page-11-1"></span>5. Bueno R, Stawiski EW, Goldstein LD, et al. Comprehensive genomic analysis of malignant pleural mesothelioma identifies recurrent mutations, gene fusions and splicing alterations. *Nat Genet*. 2016;48(4):407-416.
- <span id="page-11-13"></span>6. Hmeljak J, Sanchez-Vega F, Hoadley KA, et al. Integrative molecular characterization of malignant pleural mesothelioma. *Cancer Discov*. 2018;8(12):1548-1565.
- 7. Kinoshita Y, Hida T, Hamasaki M, et al. A combination of MTAP and BAP1 immunohistochemistry in pleural effusion cytology for the diagnosis of mesothelioma. *Cancer Cytopathol*. 2018;126(1):54-63.
- 8. Hida T, Hamasaki M, Matsumoto S, et al. BAP1 immunohistochemistry and p16 FISH results in combination provide higher confidence in malignant pleural mesothelioma diagnosis: ROC analysis of the two tests. *Pathol Int*. 2016;66(10):563-570.
- 9. Berg KB, Dacic S, Miller C, Cheung S, Churg A. Utility of methylthioadenosine phosphorylase compared with BAP1 immunohistochemistry, and CDKN2A and NF2 fluorescence In situ hybridization in separating reactive mesothelial proliferations from epithelioid malignant mesotheliomas. *Arch Pathol Lab Med*. 2018;142(12):1549-1553.
- 10. Sauter JL, Dacic S, Galateau-Salle F, et al. The 2021 WHO classification of tumors of the pleura: advances since the 2015 classification. *J Thorac Oncol*. 2022;17:608-622.
- <span id="page-11-3"></span>11. Sheffield BS, Lorette J, Shen Y, Marra MA, Churg A. Immunohistochemistry for NF2, LATS1/2, and YAP/TAZ fails to separate benign from malignant mesothelial proliferations. *Arch Pathol Lab Med*. 2016;140(5):391.
- <span id="page-11-4"></span>12. Sato T, Sekido Y. NF2/Merlin inactivation and potential therapeutic targets in mesothelioma. *Int J Mol Sci*. 2018;19(4):988.
- 13. Li W, Cooper J, Karajannis MA, Giancotti FG. Merlin: a tumour suppressor with functions at the cell cortex and in the nucleus. *EMBO Rep*. 2012;13(3):204-215.
- 14. Cooper J, Giancotti FG. Molecular insights into NF2/Merlin tumor suppressor function. *FEBS Lett*. 2014;588(16):2743-2752.
- 15. Yap TA, Aerts JG, Popat S, Fennell DA. Novel insights into mesothelioma biology and implications for therapy. *Nat Rev Cancer*. 2017;17(8):475-488.
- 16. Murakami H, Mizuno T, Taniguchi T, et al. LATS2 is a tumor suppressor gene of malignant mesothelioma. *Cancer Res*. 2011;71(3):873-883.
- <span id="page-11-5"></span>17. Kakiuchi T, Takahara T, Kasugai Y, et al. Modeling mesothelioma utilizing human mesothelial cells reveals involvement of phospholipase-C beta 4 in YAP-active mesothelioma cell proliferation. *Carcinogenesis*. 2016;37(11):1098-1109.
- 18. Matsushita A, Sato T, Mukai S, et al. TAZ activation by hippo pathway dysregulation induces cytokine gene expression and promotes mesothelial cell transformation. *Oncogene*. 2019;38(11):1966-1978.
- <span id="page-11-14"></span>19. Mizuno T, Murakami H, Fujii M, et al. YAP induces malignant mesothelioma cell proliferation by upregulating transcription of cell cycle-promoting genes. *Oncogene*. 2012;31(49): 5117-5122.
- <span id="page-11-6"></span>20. Kaneda A, Seike T, Danjo T, et al. The novel potent TEAD inhibitor, K-975, inhibits YAP1/TAZ-TEAD protein-protein interactions and exerts an anti-tumor effect on malignant pleural mesothelioma. *Am J Cancer Res*. 2020;10(12):4399-4415.
- 21. Tang TT, Konradi AW, Feng Y, et al. Small molecule inhibitors of TEAD auto-palmitoylation selectively inhibit proliferation and tumor growth of NF2-deficient mesothelioma. *Mol Cancer Ther*. 2021;20(6):986-998.
- 22. Holden JK, Crawford JJ, Noland CL, et al. Small molecule dysregulation of TEAD lipidation induces a dominant-negative inhibition of hippo pathway signaling. *Cell Rep*. 2020;31(12):107809.
- <span id="page-11-7"></span>23. Flynn TG, de Bold ML, de Bold AJ. The amino acid sequence of an atrial peptide with potent diuretic and natriuretic properties. *Biochem Biophys Res Commun*. 1983;117(3):859-865.
- 24. Kangawa K, Matsuo H. Purification and complete amino acid sequence of alpha-human atrial natriuretic polypeptide (α-hANP). *Biochem Biophys Res Commun*. 1984;118(1):131-139.
- 25. Kangawa K, Fukuda A, Matsuo H. Structural identification of beta- and gamma-human atrial natriuretic polypeptides. *Nature*. 1985;313(6001):397-400.
- 26. Sudoh T, Kangawa K, Minamino N, Matsuo H. A new natriuretic peptide in porcine brain. *Nature*. 1988;332(6159):78-81.
- <span id="page-11-8"></span>27. Potter LR. Domain analysis of human transmembrane guanylyl cyclase receptors: implications for regulation. *Front Biosci*. 2005;10:1205-1220.
- 28. Levin ER, Gardner DG, Samson WK. Natriuretic peptides. *N Engl J Med*. 1998;339(5):321-328.
- <span id="page-11-9"></span>29. Yasue H, Yoshimura M, Sumida H, et al. Localization and mechanism of secretion of B-type natriuretic peptide in comparison with those of A-type natriuretic peptide in normal subjects and patients with heart failure. *Circulation*. 1994;90(1):195-203.
- 30. Yamamoto K, Burnett JC Jr, Jougasaki M, et al. Superiority of brain natriuretic peptide as a hormonal marker of ventricular systolic and diastolic dysfunction and ventricular hypertrophy. *Hypertension*. 1996;28(6):988-994.
- 31. Vasan RS, Benjamin EJ, Larson MG, et al. Plasma natriuretic peptides for community screening for left ventricular hypertrophy and systolic dysfunction: the Framingham heart study. *JAMA*. 2002;288(10):1252-1259.
- <span id="page-11-10"></span>32. de Lemos JA, Morrow DA, Bentley JH, et al. The prognostic value of B-type natriuretic peptide in patients with acute coronary syndromes. *N Engl J Med*. 2001;345(14):1014-1021.
- 33. Kragelund C, Gronning B, Kober L, Hildebrandt P, Steffensen R. N-terminal pro-B-type natriuretic peptide and long-term mortality in stable coronary heart disease. *N Engl J Med*. 2005;352(7):666-675.
- <span id="page-11-11"></span>34. Tsolaki V, Zarogiannis S, Zygoulis P, et al. Malignant mesothelioma cells secrete natriuretic peptides: data and diagnostic clinical implications. *Respirology*. 2020;25(10):1060-1065.
- <span id="page-11-12"></span>35. Sato T, Nakanishi H, Akao K, et al. Three newly established immortalized mesothelial cell lines exhibit morphological phenotypes corresponding to malignant mesothelioma epithelioid, intermediate, and sarcomatoid types, respectively. *Cancer Cell Int*. 2021;21(1):546.
- <span id="page-11-15"></span>36. Kodama Y, Tanaka I, Sato T, et al. Oxytocin receptor is a promising therapeutic target of malignant mesothelioma. *Cancer Sci*. 2021;112(9):3520-3532.
- <span id="page-11-16"></span>37. Dauch D, Rudalska R, Cossa G, et al. A MYC-aurora kinase a protein complex represents an actionable drug target in p53 altered liver cancer. *Nat Med*. 2016;22(7):744-753.

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- <span id="page-12-2"></span>38. Gay CM, Zhou Y, Lee JJ, et al. A phase II trial of alisertib (MLN8237) in salvage malignant mesothelioma. *Oncologist*. 2020;25(10):e1457-e1463.
- <span id="page-12-3"></span>39. Stein C, Bardet AF, Roma G, et al. YAP1 exerts its transcriptional control via TEAD-mediated activation of enhancers. *PLoS Genet*. 2015;11(8):e1005465.
- 40. Bum-Erdene K, Zhou D, Gonzalez-Gutierrez G, et al. Smallmolecule covalent modification of conserved cysteine leads to allosteric inhibition of the TEAD Yap protein-protein interaction. *Cell Chem Biol*. 2019;26(3):378-389. e13.
- 41. Lu T, Li Y, Lu W, et al. Discovery of a subtype-selective, covalent inhibitor against palmitoylation pocket of TEAD3. *Acta Pharm Sin B*. 2021;11(10):3206-3219.
- <span id="page-12-4"></span>42. Cao L, Gardner DG. Natriuretic peptides inhibit DNA synthesis in cardiac fibroblasts. *Hypertension*. 1995;25(2):227-234.
- <span id="page-12-5"></span>43. Fusco N, Vaira V, Righi I, et al. Characterization of the immune microenvironment in malignant pleural mesothelioma reveals prognostic subgroups of patients. *Lung Cancer*. 2020;150:53-61.
- <span id="page-12-6"></span>44. Kinoshita Y, Hamasaki M, Yoshimura M, Matsumoto S, Iwasaki A, Nabeshima K. Hemizygous loss of NF2 detected by fluorescence in situ hybridization is useful for the diagnosis of malignant pleural mesothelioma. *Mod Pathol*. 2020;33(2):235-244.

<span id="page-12-7"></span>45. Christensen BC, Houseman EA, Godleski JJ, et al. Epigenetic profiles distinguish pleural mesothelioma from normal pleura and predict lung asbestos burden and clinical outcome. *Cancer Res*. 2009;69(1):227-234.

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<span id="page-12-1"></span>Additional supporting information can be found online in the Supporting Information section at the end of this article.

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