

Neurotoxic effects of n-hexane on the human central nervous system: evoked potential abnormalities in n-hexane polyneuropathy

YANG-CHYUAN CHANG

From the Department of Neurology, National Taiwan University Hospital, Taipei, Taiwan

SUMMARY An outbreak of n-hexane polyneuropathy as a result of industrial exposure occurred in printing factories in Taipei area from December 1983 to February 1985. Multimodality evoked potentials study was performed on 22 of the polyneuropathy cases, five of the subclinical cases, and seven of the unaffected workers. The absolute and interpeak latencies of patterned visual evoked potential (pVEP) in both the polyneuropathy and subclinical groups were longer than in the normal controls. The pVEP interpeak amplitude was also decreased in the polyneuropathy cases. Brainstem auditory evoked potentials (BAEP), showed no difference of wave I latency between factory workers and normal controls, but prolongation of the wave I-V interpeak latencies was noted, corresponding with the severity of the polyneuropathy. In somatosensory evoked potentials (SEPs), both the absolute latencies and central conduction time (CCT) were longer in subclinical and polyneuropathy cases than in the unaffected workers and normal controls. From this evoked potentials study, chronic toxic effects of n-hexane on the central nervous system were shown.

Outbreaks of toxic neuropathy attributable to n-hexane have been described in industrial workers in Italy, Japan, Morocco, France and the United States.^{1,2} The industrial cases commonly occur in factories with inadequate ventilation and high air levels of n-hexane.¹ Clinically, the patients present with chronic or subacute sensorimotor polyneuropathy.

In human sural nerve biopsies^{1,2} and experimental animals,¹ it has been found that n-hexane can induce axonal swelling with myelin changes in the peripheral nerves. Similar pathological changes involving the terminal portion of axons in the central nervous system has been reported in experimental animals.³⁻⁵ Clinical evidence, however, suggesting chronic neurotoxic effect of n-hexane on the central nervous system has rarely been mentioned in patients with n-hexane polyneuropathy.^{1,5,6-8} In the present study,

evoked potential abnormalities in patients with n-hexane polyneuropathy are reported, the electrophysiological abnormalities being considered as related to the chronic neurotoxic effects of n-hexane on the central nervous system.

Materials and methods

Materials

From December 1983 to February 1985, an outbreak of toxic polyneuropathy was found in the printing industry near the Taipei area in Taiwan. After detailed epidemiological studies including analysis of bulk cleaning solvents used and one-hour personal air samples, questionnaire interviews on the working circumstances, and medical examinations on these patients had been carefully performed, it was found that the endemic neurological abnormalities were attributable to chronic n-hexane poisoning.⁹ Use of solvents with high contents of n-hexane, poor ventilation, and the habit of sleeping in the factories between shifts were important aetiological factors.⁹

There were 28 polyneuropathy patients. Clinically, they presented with progressive weakness and muscle atrophy, symmetrically involving the distal part of the lower extremities. In severe cases, the motor effects also involved the proximal portion of the lower limbs and the upper limbs. Subjective sensory symptoms such as numbness and paraesthesia were noted in one-third of the patients.

Among the 28 polyneuropathy cases, 22 patients under-

Presented in part at the 13th World Congress of Neurology (5 September 1985, Hamburg).

Address for reprint requests: Yang-Chyuan Chang, MD, Department of Neurology, National Taiwan University Hospital, Taipei, Taiwan 100.

Received 14 January 1986 and in revised form 4 June 1986.
Accepted 19 June 1986

Table 1 Neurological manifestations in *n*-hexane polyneuropathy

	Number
Peripheral nervous system manifestations (in 22 patients)	
Weakness in the lower extremities	22
Weakness in the upper extremities	11
Absent and/or decreased achilles reflex	22
Absent and/or decreased patellar reflex	22
Absent and/or decreased brachioradialis reflex	15
Absent and/or decreased biceps brachii reflex	8
Decreased vibration sense in the lower extremities	15
Hypalgesia in the lower extremities	7
Hypalgesia in the upper extremities	7
Optic nerve manifestations (44 eyes in 22 patients)	
Optic or retrobulbar neuritis	0
Optic atrophy	0
Deschromatopsia (by colour plate screening test)	2
Visual acuity at 20/50	1
Visual acuity at 20/33	2
Visual acuity at 20/25	12
Visual acuity at 20/20	16
Visual acuity at 20/15	13
Central nervous system manifestations (in 22 patients)	
Headache and/or dizziness	0
Mental changes	0
Sleep disorder	0
EEG abnormalities (examined in 6 patients)	0

went evoked potential study. Their neurological manifestations are listed in table 1. None of them complained of headache or dizziness. No mental changes were found on clinical mental status examination. Electroencephalography in six of 22 patients revealed normal tracings. Five subclinical cases and seven unaffected workers were also included in this study. They came from the same factories and performed similar work. In subclinical cases, there were neither neurological symptoms nor signs related to any peripheral nerve lesion, yet at least one abnormal nerve conduction velocity was seen in each case. Workers who had no symptoms and signs related to any peripheral nerve lesion, or abnormal nerve conduction velocities were classified as unaffected workers. The mean peripheral nerve conduction velocities in these groups are listed in table 2.

In the polyneuropathy cases, the age ranged from 17 to 34 years with a mean of 23.1 ± 5.6 years. The mean age in the subclinical cases was 23.2 ± 7.5 years, with a range from 18 to 32 years. A mean age of 28.3 ± 6.2 years and a range from 20 to 36 years were found in the unaffected workers. There was one female subject in each group.

Patterned visual evoked potentials (pVEPs)

Patterned visual evoked potentials were studied with a Nicolet Pathfinder II machine (Nicolet Biomedical Instrument), recorded from the scalp by silver/silver chlorided cup electrodes. The active electrode was placed at Oz with a reference at Cz in the 10–20 system. The low- and high-frequency filters of the amplifier were set at 1 and 100 Hz. The analysis time was 250 ms. Two hundred and fifty-six responses were averaged. For ensuring the reliability and reproducibility of the findings, the pVEP was performed twice at any given visual angle. Each eye was tested separately. The visual stimulation was generated through a NIC-1005 black-and-white checkerboard patterned reversal visual stimulator at a reversal rate of 1.88/s, with checks subtending 1° , $30'$, $15'$, and $7.5'$ of the visual angles. The full stimulus field size was 16° .

The N1 (N75), P1 (P100), and N2 (N135) latencies of the pVEPs obtained with a visual angle of $30'$ were measured. The amplitude measurements were referred to the N1–P1 and P1–N2 interpeak amplitudes.

The normal controls for patterned VEP consist of 22 males, with a mean age of 33.1 ± 7.9 years.

Table 2 Nerve conduction velocities (m/s) in workers exposed to *n*-hexane

	Laboratory norms	Unaffected workers (n = 14)	Subclinical cases (n = 10)	Polyneuropathy cases
Median MNCV	61.2 ± 5.8	57.0 ± 3.5	50.6 ± 6.1	43.3 ± 6.8 (n = 42)
Ulnar MNCV	59.9 ± 7.2	55.4 ± 5.0	45.6 ± 2.3	40.9 ± 5.0 (n = 42)
Peroneal MNCV	53.4 ± 6.1	49.1 ± 4.2	41.2 ± 5.2	34.0 ± 5.0 (n = 42)
Tibial MNCV	49.1 ± 5.2	46.2 ± 4.1	42.7 ± 3.8	35.9 ± 5.6 (n = 42)
Median SNCV	56.1 ± 4.8	56.0 ± 5.3	50.3 ± 3.3	47.7 ± 3.1 (n = 40)
Ulnar SNCV	52.4 ± 4.6	52.1 ± 4.7	45.7 ± 3.6	44.2 ± 4.6 (n = 37)
Sural SNCV	50.3 ± 4.7	48.9 ± 4.1	46.2 ± 2.8	43.1 ± 3.8 (n = 26)

Notes:

1. MNCV = motor nerve conduction velocity; SNCV = sensory nerve conduction velocity.
2. The results were calculated by combining the findings obtained from the nerves in the right side and those in the left side.
3. Absence of evoked muscle response or sensory action potential was excluded from calculation.

Brainstem auditory evoked potentials (BAEPs)

Brainstem auditory evoked potential data were collected, averaged, and analysed on a Pathfinder II machine. The auditory stimulus in both condensation and rarefaction polarities consisted of 100 μ s clicks presented at a rate of 11.1/s. The intensity of the click stimulus was 60 dB above threshold for each subject, while a white masking noise at a level of -40 dB was applied on the contralateral non-stimulating ear. For each trial, 2048 responses were averaged. Each ear was tested separately and at least twice for ensuring reproducibility and reliability. Silver/silver chlorided cup electrodes were attached at vertex (Cz, as the positive electrode) and each mastoid (A1, A2). The electrode resistance was kept lower than 3 kohm. Filter settings were at 150 and 3000 Hz. The analysis time was 10 ms. All subjects were tested in the supine position on a bed.

Absolute latencies were measured from stimulus to the positive peaks. Out of the seven peaks, the absolute latencies of peaks I, III, and V were used. For determination of the central conduction time in the auditory pathway, interpeak latencies (IPLs) between peaks I-III, III-V, and I-V were measured.

In the normal controls, there were 25 males with the mean age of 32.8 \pm 9.5 years.

Somatosensory evoked potentials (SEPs)

Three averaging channels for somatosensory evoked potentials were used to record simultaneously scalp SEPs, neck SEPs, and brachial plexus potentials by stimulation of the median nerve. The analysis time was 50 ms.

For scalp SEPs, the active electrode was placed on either the left or right central region of the scalp (C3 or C4) contralateral to the side of median nerve stimulation. The band-pass was set at 1 and 3000 Hz. For neck SEPs the active electrode was attached at the 7th cervical spinous process and the filter settings were 5 and 3000 Hz. The brachial plexus potentials were recorded at the Erb's point ipsilateral to the side of stimulation, with low- and high-frequency filters at 5 and 3000 Hz. An electrode at Fz was used as the common reference point. With Pathfinder II, 1024 or 2048 responses for each channel were averaged.

Square wave pulse electric stimulation of 0.1 ms duration at a rate of 2.7 per second was applied to the median nerve at wrist. Stimulation intensity was adjusted to the minimum current for eliciting visible thumb twitching. Right and left median nerve were tested separately.

For measurement, the absolute latencies of N1 (N20) component of scalp SEPs and N13 peak of neck SEPs were recorded. Central conduction time (CCT) in the somato-

sensory pathway was defined as the time differences between N1 latency of the scalp SEP and N13 latency of the neck SEP.

Scalp SEPs by stimulation of the peroneal nerve at the ankle were recorded at the vertex (Cz), with a reference point at Fz. Filters were set at 1 and 3000 Hz and the analysis time was 200 ms. Electric stimulation of 0.2 ms duration was delivered at a rate of 2.1/s and its intensity was adjusted to induce visible twitching of the toes. Two hundred and fifty-six or 512 responses were averaged for each trial. Each leg was stimulated separately. For measurement, the absolute latency of P1 (P40) was selected.

In the normal controls, there were 25 males and 17 females, with a mean age of 36.9 \pm 9.2 years.

Results

All the results of evoked potentials presenting in the following tables were calculated by combining the findings obtained from the right side stimulation and those from the left side stimulation.

Patterned visual evoked potentials

Table 3 shows the absolute latencies of N1, P1, and N2 components of pVEPs. The N1-N2 interpeak latencies, the N1-P1 interpeak amplitude, and the P1-N2 interpeak amplitude are also listed in table 3. N1, P1, and N2 absolute latencies and N1-N2 interpeak latencies were significantly longer in the sub-clinical and polyneuropathy cases than in the normal controls and the unaffected workers. Significant decreases of N1-P1 and P1-N2 interpeak amplitudes were also found in polyneuropathy cases.

Brainstem auditory evoked potentials

Table 4 shows the absolute latencies of wave I, III, and V in the BAEP study. The I-III, III-V, and I-V interpeak latencies are also listed. There was no significant difference of wave I latency among the four groups ($F = 0.5326, \alpha > 0.13$). The absolute wave III and V latencies, and the I-III, III-V and I-V interpeak latencies were significantly prolonged in the polyneuropathy cases. The tendency for the longer interpeak latencies to be associated with the more severe the involvement of the peripheral nerves can be observed in Table 4.

Table 3 Findings of patterned visual evoked potentials at a visual angle of 30' in workers exposed to *n*-hexane

	Normal controls (n = 44)	Unaffected workers (n = 14)	Subclinical cases (n = 10)	Polyneuropathy cases (n = 44)	Statistically significant levels
N1 latency (ms)	72 \pm 4	71 \pm 2	75 \pm 5	75 \pm 5	t = 3.108, p < 0.005
P1 latency (ms)	96 \pm 3	96 \pm 3	102 \pm 7	101 \pm 7	t = 4.355, p < 0.0005
N2 latency (ms)	127 \pm 7	131 \pm 8	139 \pm 12	138 \pm 12	t = 5.252, p < 0.0005
N1-N2 interpeak latency (ms)	55 \pm 5	60 \pm 9	65 \pm 10	63 \pm 12	t = 4.082, p < 0.0005
N1-P1 interpeak amplitude (μ V)	8.00 \pm 3.49	7.72 \pm 2.08	7.51 \pm 1.35	5.70 \pm 2.85	t = 3.386, p < 0.005
P1-N2 interpeak amplitude (μ V)	9.36 \pm 2.85	9.52 \pm 3.84	9.03 \pm 2.52	5.58 \pm 2.37	t = 6.765, p < 0.0005

Statistically significant levels were obtained from Student's t test on "normal controls" versus "polyneuropathy cases".

Table 4 Findings of brainstem auditory evoked potentials in workers exposed to n-hexane

	Normal controls (n = 50)	Unaffected workers (n = 14)	Subclinical cases (n = 10)	Polyneuropathy cases (n = 44)	Statistically significant levels
Wave I latency (ms)	1.62 ± 0.13	1.66 ± 0.13	1.64 ± 0.15	1.65 ± 0.14	t = 1.077, p > 0.25
Wave III latency (ms)	3.70 ± 0.15	3.82 ± 0.15	3.90 ± 0.25	3.93 ± 0.26	t = 5.330, p < 0.0005
Wave V latency (ms)	5.55 ± 0.15	5.69 ± 0.30	5.87 ± 0.18	5.93 ± 0.24	t = 9.320, p < 0.0005
I-III interpeak latency (ms)	2.08 ± 0.12	2.16 ± 0.26	2.27 ± 0.16	2.29 ± 0.18	t = 6.726, p < 0.0005
III-V interpeak latency (ms)	1.85 ± 0.14	1.86 ± 0.23	1.98 ± 0.19	2.00 ± 0.21	t = 4.118, p < 0.0005
I-V interpeak latency (ms)	3.94 ± 0.16	4.02 ± 0.25	4.25 ± 0.30	4.28 ± 0.21	t = 8.888, p < 0.0005

Statistically significant levels were obtained from Student's t test on "normal controls" versus "polyneuropathy cases".

Table 5 Findings of somatosensory evoked potentials (SEPs) in workers exposed to n-hexane

	Normal controls (n = 84)	Unaffected workers (n = 14)	Subclinical cases (n = 10)	Polyneuropathy cases	Statistically significant levels
Scalp SEP latency by median N stim (ms)	18.44 ± 0.80	18.53 ± 0.57	19.40 ± 0.95	20.49 ± 0.90 (n = 42)	t = 12.444, p < 0.0005
Neck SEP latency by median N stim (ms)	12.73 ± 0.72	12.72 ± 0.71	13.48 ± 0.78	14.30 ± 0.78 (n = 42)	t = 11.221, p < 0.0005
Central conduction time by median N stim (ms)	5.70 ± 0.45	5.81 ± 0.49	5.92 ± 0.44	6.20 ± 0.55 (n = 42)	t = 5.451, p < 0.0005
Scalp SEP latency by peroneal N stim (ms)	39.13 ± 2.45	39.86 ± 1.68	42.32 ± 2.84	45.37 ± 3.68 (n = 40)	t = 11.199, p < 0.0005

Statistically significant levels were obtained from Student's t test on "normal controls" versus "polyneuropathy cases".

Somatosensory evoked potentials

In one polyneuropathy patient, no identifiable SEPs could be elicited by electric stimulation of either the median or the peroneal nerve. In another patient, scalp SEPs could not be obtained by peroneal nerve stimulation.

Table 5 shows the scalp and neck SEP latencies obtained by stimulation of the median nerve, and the scalp SEP latencies by stimulation of the peroneal nerve. Central somatosensory conduction time (CCT), which was calculated by subtracting the neck SEP latency from the scalp SEP latency by stimulation of the median nerve, is also listed. All the SEP latencies and CCT in subclinical and polyneuropathy cases were significantly longer than those in the normal controls. There was a tendency to have longer SEP latencies and CCT with increasing severity of peripheral nerves involvement.

Discussion

Acute exposure to high air concentration of n-hexane can induce nausea, headache, giddiness, euphoria, hallucination, and mild narcosis.¹ Sensorimotor polyneuropathy is the principal neurological manifestation in subacute or chronic exposure to n-hexane, although cranial neuropathies, blurred vision, and abnormal colour vision associated with macular changes have also been reported.^{6,8,10-12} During the initial neurological evaluation of the patients with n-hexane polyneuropathy, symptoms

and signs indicating CNS involvement were often not obvious. Sobue *et al*⁶ reported 35 n-hexane polyneuropathy patients among vinyl sandal manufacturers and found that except for irritability, dizziness, or sleeplessness, there was no evidence of CNS involvement. In a detailed clinical investigation of 93 patients with n-hexane polyneuropathy, Yamamura¹⁰ concluded that there were no symptoms and signs indicating lesions in CNS.

Pathologically, giant axonal swelling with dying back degeneration is the main finding in sural nerve biopsies from n-hexane polyneuropathy patients and in experimental hexacarbon polyneuropathy.¹ However, besides the peripheral nerves, axonal swelling, axonal degeneration, and secondary breakdown of myelin sheath was also observed in the CNS in experimental animals intoxicated with hexacarbon. In rats, Spencer and Schaumburg³ found that the long ascending and descending tracts of the spinal cord were affected first. The same pathological changes in the cerebellum and lateral geniculate body were also present in rats with advanced disease.³ In cats intoxicated by low-level but prolonged administration of 2,5-hexanedione (the putative neurotoxic metabolite of n-hexane), Schaumburg and Spencer⁴ demonstrated early axonal swellings in the mammillary bodies, the lateral geniculate body and distal optic tract, and the superior colliculi. In view of these pathological studies, it is not unreasonable to postulate that there may be simultaneous CNS lesions in n-hexane polyneuropathy patients.

Although there is an absence of clear clinical signs of CNS degeneration at the initial neurological evaluations in patients with n-hexane polyneuropathy, indirect evidence indicating CNS involvement has been mentioned on follow-up studies. After recovery of muscle strength and sensation, the patients may have residual spasticity and hyperactive tendon reflexes,^{1 5 8 13} suggesting irreparable damage to the long tracts of the spinal cord. Schaumburg and Spencer⁴ thought that weakness and sensory impairment associated with peripheral nerves damage would mask signs of CNS degeneration.

In addition to bedside neurological examination, electrophysiological techniques have also been employed to evaluate the neurotoxic effects of n-hexane on the CNS. By studying H-reflex behaviour in patients with n-hexane polyneuropathy, Bravaccio *et al*¹³ found that an increased excitability of alpha motorneurons in their patients gave good evidence of spinal cord involvement. After analysis of scalp SEPs by stimulation of the median nerves, Mutti *et al*¹⁴ noted much flatter later SEP components in 15 women exposed to n-hexane than in the control group, suggesting some neurotoxic effects of n-hexane on the CNS. Seppalainen *et al*^{12 15} found abnormal amplitudes and latencies of flash visual evoked potentials in industrial workers exposed to n-hexane. They interpreted these changes as the result of cerebral dysfunction, probably conduction block in intracerebral axons.

In the present study, the N1, P1 and N2 absolute latencies, and N1–N2 interpeak latency of patterned VEP were longer in both the subclinical and polyneuropathy groups than in the normal controls and unaffected workers. Although maculopathy and colour discrimination defects are the main eye findings in industrial workers exposed to n-hexane,^{12 15} they usually do not induce latency abnormalities of the patterned VEP.¹⁶ Optic nerve atrophy and retrobulbar neuritis, which can cause abnormalities in patterned VEP and have been reported in patients with n-hexane polyneuropathy,¹⁰ were not found in the present study (table 2). Therefore, the pVEP abnormality shown in table 3 cannot be explained by the toxic effects of n-hexane on the retina or visual pathway peripheral to the geniculate bodies. As P1 of patterned VEP is generated in the primary visual cortex but probably does not represent the first cortical component,¹⁶ the latency abnormalities in the subclinical and polyneuropathy groups result from cerebral dysfunction. Cerebral dysfunction could also be responsible for the amplitude attenuation in the polyneuropathy patients, although the role of maculopathy in amplitude changes is not clear.

In the BAEP study, I–III, III–V, and I–V interpeak latencies reflect neural conduction in the correspond-

ing segments of the central auditory pathway in the brainstem. In the present study, the lack of difference of wave I latency between the normal controls and factory workers suggests that the auditory nerve itself was not severely affected in n-hexane intoxication. Prolongation of interpeak latencies in subclinical cases and polyneuropathy patients should be interpreted as neurotoxic effects of n-hexane on the brainstem.

Slow conduction in the peripheral nerves can partly explain the delay of the scalp SEP and neck SEP latencies in patients with n-hexane polyneuropathy. Prolongation of the central somatosensory conduction time as shown in the present study is the other crucial factor for the delay of SEP latencies. As the impulses for eliciting scalp SEPs are transmitted through the posterior columns of the spinal cord, and early pathological changes involving the rostral part of the gracile and cuneate tracts have been found in experimental rats intoxicated by n-hexane,^{1 3} it is not surprising that there is abnormally prolonged central somatosensory conduction time in patients with n-hexane polyneuropathy. This is electrophysiological evidence of neurotoxic effects of n-hexane on the spinal cord and/or the brainstem.

In analysing evoked potential abnormalities, subject variables which may influence the latency and amplitude of evoked potentials should be considered. Aging has often been reported to increase the absolute and interpeak latencies, and to decrease the amplitude.¹⁷ In the present study, the mean ages of each worker group were younger than that of normal controls, hence, there would be much more abnormality if the evoked potential results were age-adjusted. Sex differences have been mentioned in the absolute P1 latency of patterned VEPs and interpeak latencies of BAEPs,¹⁷ yet have not been observed in central somatosensory conduction time.¹⁸ Sex differences were considered negligible in the present study, for there was only one female subject in each worker group.

From the results of the evoked potential study, it is concluded that there are chronic neurotoxic effects of n-hexane on the central nervous system, including the cerebrum, the brainstem, and the spinal cord. These toxic effects exist in patients with clinical or subclinical polyneuropathy, and also possible in exposed workers without abnormal nerve conduction velocities in the peripheral nerves.

References

- 1 Spencer PS, Couri D, Schaumburg HH. n-Hexane and methy n-butyl ketone. In: Spencer PS, Schaumburg HH, eds. *Experimental and Clinical Neurotoxicology*. Baltimore: Williams & Wilkins, 1980:456–75.

- 2 Scelsi R, Poggi P, Fera L, Gonella G. Industrial neuropathy due to n-hexane—clinical and morphological findings in three cases. *Clin Toxicol* 1981;**18**:1387–93.
- 3 Spencer PS, Schaumburg HH. Ultrastructural studies of the dying-back process, IV. Differential vulnerability of PNS and CNS fibers in experimental central-peripheral distal axonopathies. *J Neuropathol Exp Neurol* 1977;**36**:300–20.
- 4 Schaumburg HH, Spencer PS. Environmental hydrocarbons produce degeneration in cat hypothalamus and optic tract. *Science* 1978;**199**:199–200.
- 5 Schaumburg HH, Spencer PS. Clinical and experimental studies of distal axonopathy—a frequent form of brain and nerve damage produced by environmental chemical hazards. *Ann NY Acad Sci* 1979;**329**:14–29.
- 6 Sobue I, Yamamura Y, Ando K, Iida M, Takayanagi T. n-Hexane polyneuropathy—outbreak among vinyl sandal manufacturers. *Clin Neurol (Jpn)* 1968;**8**:393–403.
- 7 Matsumura M, Inoue N, Ohnishi A, Santo T, Goto I. Toxic polyneuropathy due to glue-sniffing. *Clin Neurol (Jpn)* 1972;**12**:290–6.
- 8 Korobkin R, Asbury AK, Summer AJ, Nielsen SL. Glue-sniffing neuropathy. *Arch Neurol* 1975;**32**:158–62.
- 9 Wang JD, Chang YC, Kao KP, Huang CC, Lin CC, Yeh WY. An outbreak of n-hexane induced polyneuropathy among press proofing workers in Taipei. *Am J Industrial Med* 1986;**10**:111–8.
- 10 Yamamura Y. n-Hexane polyneuropathy. *Fol Psychiat Neurol (Jpn)* 1969;**23**:45–57.
- 11 Gonzalez E, Downey J. Polyneuropathy in a glue sniffer. *Arch Phys Med* 1972;**53**:333–7.
- 12 Seppalainen AM, Raitta C, Huuskonen MS. Nervous and visual effects of occupational n-hexane exposure. In: Lechner H, Aranibar A, eds. *EEG and Clinical Neurophysiology*. Amsterdam: Excerpta Medica 1980:656–61.
- 13 Bravaccio F, Ammendola A, Barruffo L, Carlomagno S. H-reflex behavior in glue (n-hexane) neuropathy. *Clin Toxicol* 1981;**18**:1369–75.
- 14 Mutti A, Ferri F, Lommi G, Lotta S, Lucertini S, Franchini I. n-Hexane-induced changes in nerve conduction velocities and somatosensory evoked potentials. *Int Arch Occup Environ Health* 1982;**51**:45–54.
- 15 Seppalainen AM, Raitta C, Huuskonen MS. n-Hexane-induced changes in visual evoked potentials and electroretinograms of industrial workers. *Electroencephalogr Clin Neurophysiol* 1979;**47**:492–8.
- 16 Lueders H, Lesser RP, Klem G. Pattern evoked potentials. In: Henry CE, ed. *Current Clinical Neurophysiology: Update on EEG and Evoked Potentials*. New York: Elsevier/North Holland 1980:467–525.
- 17 Spehlmann R. *Evoked Potential Primer*. Boston: Butterworth Publishers, 1985:98–9, 208 & 297.
- 18 Chang YC. Influences of age and body height on central somatosensory conduction time in normal subjects. *J Formosan Med Assoc* 1984;**83**:912–21.