

Invasive candidiasis: current clinical challenges and unmet needs in adult populations

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Invasive candidiasis (IC) is a serious infection caused by several *Candida* species, and the most common fungal disease in hospitals in high-income countries. Despite overall improvements in health systems and ICU care in the last few decades, as well as the development of different antifungals and microbiological techniques, mortality rates in IC have not substantially improved. The aim of this review is to summarize the main issues underlying the management of adults affected by IC, focusing on specific forms of the infection: IC developed by ICU patients, IC observed in haematological patients, breakthrough candidaemia, sanctuary site candidiasis, intra-abdominal infections and other challenging infections.

Several key challenges need to be tackled to improve the clinical management and outcomes of IC patients. These include the lack of global epidemiological data for IC, the limitations of the diagnostic tests and risk scoring tools currently available, the absence of standardized effectiveness outcomes and long-term data for IC, the timing for the initiation of antifungal therapy and the limited recommendations on the optimal step-down therapy from echinocandins to azoles or the total duration of therapy.

The availability of new compounds may overcome some of the challenges identified and increase the existing options for management of chronic *Candida* infections and ambulant patient treatments. However, early identification of patients that require antifungal therapy and treatment of sanctuary site infections remain a challenge and will require further innovations.

Introduction

Invasive candidiasis (IC) is a serious infection caused by several *Candida* species (spp.) and the most common fungal disease in hospitals in high-income countries, with a worldwide prevalence ranging from 250 000 to approximately 700 000 people per year, an incidence rate of 2–14 cases per 100 000 persons and mortality rates ranging between 40% and 55%.^{1–3} IC includes both candidaemia (i.e. bloodstream infections) and deep-seated tissue candidiasis, which arises from dissemination of *Candida* spp. to a sterile body site (e.g. abdomen, peritoneum or bone).^{4–6} Most IC infections are caused by five pathogens: *C. albicans*, *Nakaseomyces glabrata* (previously known as *C. glabrata*), *C. tropicalis*, *C. parapsilosis* and *Pichia kudriavzevii* (previously known as *C. krusei*). *C. albicans* is the most common species, but non-*albicans* species are increasing, being responsible for more than 50% of cases in some series.⁷ However, this trend appears limited to specific continents (e.g. Europe, mainly due to the rise of *N. glabrata*) and a high heterogeneity is observed between

studies, overall but also in subgroups by continents.⁸ *C. auris* is a novel pathogen that has emerged in 2009, triggering global outbreaks.⁹

Candida spp. are commensal organisms present in the gut and skin of 50%–70% of healthy individuals in low numbers due to competition within the microbiome (i.e. the gut mycobiome represents only around 0.1% of the total gut microbes¹⁰). Several factors can lead to overgrowth of *Candida* spp., in particular exposure to antibiotic treatment, immunosuppression and corticosteroid treatment.¹¹ Moreover, *Candida* translocation from the gut into the bloodstream can be facilitated by increased permeability of the gut epithelia (for example, due to mucositis in onco-haematological patients or patients with inflammatory bowel disease),^{12–15} or breaches in the intestinal barrier following abdominal surgery,¹⁶ all of which significantly increase the risk of candidaemia. Additionally, the ability of *Candida* spp. to form biofilms on inert surfaces makes the presence of prosthetic material a risk factor for developing IC.¹⁷ Once candidaemia has developed, it can disseminate and generate deep-seated secondary

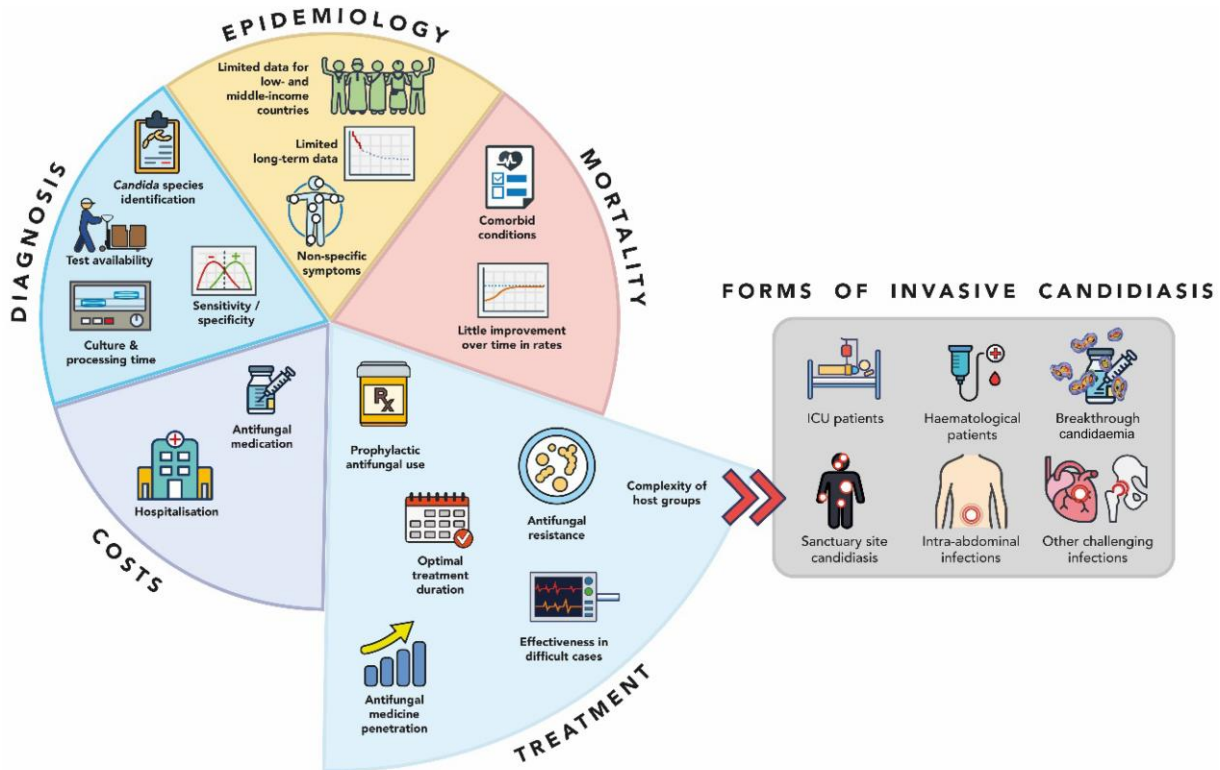


Figure 1. Current clinical challenges of IC.

infections in organs such as the lungs, liver, heart, eye, brain and bones.¹

Despite overall improvements in health systems and ICU care in the last few decades, as well as the development of different antifungals and microbiological techniques, mortality rates in IC have not substantially improved.¹⁸ Several challenges hinder the clinical management of patients with IC (Figure 1). Firstly, early diagnosis of candidaemia and deep-seated candidiasis remain a challenge due to the prolonged time to positivity of blood cultures, which can take up to 5 days to become positive, and due to the low yield of culture diagnostic tests for deep-seated candidiasis (~50%).¹⁹ Biomarkers [e.g. (1-3)- β -D-glucan (BDG), mannan (Mn)/anti-Mn antibodies] could aid earlier diagnosis; however, their role has not yet been clearly defined.¹⁹ Second, a worldwide shift to multidrug-resistant species (including *C. auris*, *N. glabrata* and *P. kudriavzevii*) has been observed.²⁰ While guidelines exist to guide the choice of antifungal therapy, patients affected by IC may require a tailored approach due to heterogeneous host factors and significant geographical variation in species distribution and antifungal drugs resistance rates.²¹⁻²³ Moreover, the value of different treatment strategies remains to be clarified.

In addition to increasing the risk of mortality, IC is associated with significant economic burden, mainly arising from the prolonged length of hospital stay, although the economic impact of IC is difficult to measure due to comorbidities.^{24,25}

The aim of this review is to summarize the main issues underlying the management of adults affected by IC (Figure 1), focusing

on specific forms of the infection: IC developed by ICU patients, IC observed in haematological patients, breakthrough candidaemia, sanctuary site candidiasis, intra-abdominal infections and other challenging infections. The impact of such issues on the clinical and economic burden of the disease is key to understanding the unmet needs of these patients. To maximize its reliability and quality, this narrative review followed the methodological recommendations from the Scale for the Assessment of Narrative Review Articles,²⁶ and eligible literature was identified through citation chasing of key references on IC and candidaemia and input from clinical experts (Supplementary Materials, available as Supplementary data at JAC Online).²⁷

Global incidence, epidemiological shifts and economic burden of *Candida* species

Global data indicates that the incidence of IC and candidaemia is increasing, with large studies reporting an incidence rate of 3–5 per 100 000 persons in the general population, 1%–2% of all ICU admissions²⁸ and a global annual incidence estimated to be ~750 000 cases/year.³ Within clinical settings, almost three-quarters of the cases were reported in the ICU (60%) and cancer and transplant units (13%).^{3,29,30} *Candida* spp. are the aetiology of 17% of all ICU infections in culture-positive patients.^{3,6,31,32} Pooled data for European countries reiterated the high ICU incidence rate, extrapolating that approximately 79 cases are diagnosed daily,³² with a cumulative incidence of IC of 7.07 episodes

per 1000 ICU admissions and a crude mortality rate of 42%.³³ The current incidence rate may be higher due to emerging risk groups, such as patients with severe COVID-19.^{32,34}

Intra-abdominal candidiasis may include *Candida* involvement of peritoneum or intra-abdominal abscess,^{35,36} and is relatively common among specific high-risk groups with prevalence ranging between 5% and 30%.¹⁹ A review on the global burden of IC reported data from 29 countries worldwide, estimating a global averaged incidence rate of 1.15 cases per 100000 for *Candida* peritonitis or intra-abdominal candidiasis, being associated with approximately half of the total cases of IC in ICU patients.³

Patients with haematological malignancies are also prone to developing IC, due to their compromised immune system and chemotherapy-induced mucositis, which results in the translocation of *Candida* into the bloodstream.^{6,15,37} A US prospective surveillance study of invasive fungal infections in haematopoietic stem cell transplant recipients conducted in 2001–2006 found *Candida* was responsible for 28% of invasive infections (mostly *N. glabrata*).¹⁴ A study involving 11802 Italian patients with haematological malignancies identified 175 cases of candidaemia (1.5%),¹³ whereas a Greek study on 27864 candidaemia patients reported an incidence rate of 1.4 cases/1000 admissions among haematology patients (versus 0.83/1000 in non-haematology patients); candidaemia was caused predominantly by non-*albicans* species.¹²

Candida spp. distribution also differs geographically; *C. albicans* is the most prevalent species in most regions of the world, but in the past decade an increase in non-*albicans* diagnosis has been observed.^{3,32,38} The second most prevalent species in the USA, north-western Europe and Canada is *N. glabrata*, particularly among elderly patients and solid organ transplant recipients. *C. parapsilosis* and *C. tropicalis* are more common in Southern Europe, South America, India and Pakistan, while *P. kudriavzevii*, the least common among the five main species, is more frequent in patients with severe immunodeficiency (e.g. haematological malignancies).^{39,40} Other less frequent species usually present in specific hosts rather than geographically (e.g. *C. dubliniensis* is more common in HIV-infected patients⁴¹). It is important to note that current IC epidemiology is highly determined by antifungal selection pressure, which is influenced by both prophylaxis and treatment.⁴² Widespread use of antifungals has driven the shift to non-*albicans* and more frequently resistant *Candida* spp. Additionally, *C. auris* has emerged as a global threat causing outbreaks in all continents. *C. auris*, known to survive on human skin and environmental surfaces for several weeks thereby facilitating its transmission,⁹ is highly resistant to azole and polyene antifungals and can be resistant to some commonly used disinfectants.^{43,44}

The collection and comparison of global data are hindered by specific challenges, including the lack of specific criteria for an incidence rate denominator.²⁰ Moreover, there are no available data for some low- and middle-income countries due to the absence of hospital infrastructures for blood culture analyses.³ These differences between studies constrain to what extent global incidence rates can be established and comparisons across countries can be made, and point towards an underestimation of the burden of disease. Large longitudinal studies, alongside regional and local surveillance studies,⁴⁵ are required to

understand epidemiological trends and shifts and to collect data to guide and support antifungal therapy.⁴⁶

Systematic analysis of global evidence reported that costs associated with IC are mainly driven by bed day costs, incremental hospitalization and antifungal drug expenditure.^{24,25} Survival and age influences costs, with both neonatal and older patients incurring higher costs, due to higher morbidity.⁴⁷

Mortality rates over time and factors affecting mortality

Nosocomial candidiasis has the highest rate of mortality for hospital-acquired infections, with 30-day post-diagnosis mortality estimated to range between 40% and 55%.^{1,2} Risk factors affecting mortality rates include older age, severity of the condition, use of immunosuppressive drugs, comorbidities, venous catheter retention and specific antifungal treatment.^{1,33} A large retrospective study conducted in nine European countries extrapolated the 30-day mortality rate to be 29 patients out of the 79 cases diagnosed per day.³² There have been limited changes to the mortality rate associated with IC in the past two decades (Figure 2), even though there are now several extended-spectrum triazole and echinocandin agents available for antifungal therapy, which have superior safety and potency than those antifungals agents available two decades ago.^{18,45,48}

Diagnostics

Culture and nonculture diagnostics

Diagnostic tests for IC should be able to accurately detect the infection and differentiate between the presence of candidaemia, deep-seated candidiasis or a combination of both.^{4,5,19} Identifying deep-seated candidiasis is important as patients may require longer therapy or surgical debridement.⁵ An early diagnosis is paramount for a timely treatment, and any delays increase the odds of mortality and healthcare associated costs.^{46,49}

Cultures from blood and sterile sites are currently the gold standard for diagnosis of IC.⁵⁰ Light microscopy using fluorescent brightener stains is also often used for the detection of *Candida* spp.,⁶ and may provide additional diagnostic benefit.⁵¹ It has been estimated that blood cultures have a sensitivity of between 63% and 83% for candidaemia in the absence of deep-seated candidiasis, and that their sensitivity is lower when deep-seated candidiasis is present,⁴⁶ ranging between 21% and 71%,⁵² with an overall sensitivity of approximately 50%. Sensitivity can also vary according to the *Candida* spp., and the use of prior antifungals.²² In addition, slow turnaround times, with a median time to positivity of 2–3 days, which may be even longer for *N. glabrata*,⁵³ may delay the start of the adequate antifungal therapy.^{22,46,49} Diagnosing IC can be further constrained by the absence of a specific clinical presentation.⁵⁴

These shortcomings may be complemented by nonculture diagnostic tests for *Candida*, such as mannan and anti-mannan antibody detection,^{19,21,52} BDG detection,¹⁹ *C. albicans* germ tube antibody (CAGTA) detection,⁵⁵ PCR detection of *Candida* DNA⁵⁶ and the T2 magnetic resonance (T2MR) *Candida* test.^{4,49} The nonculture diagnostic tests have a varying degree of sensitivity and there have been recent calls for studies to further assess the role of combination testing.¹⁹ For example, the mannan and

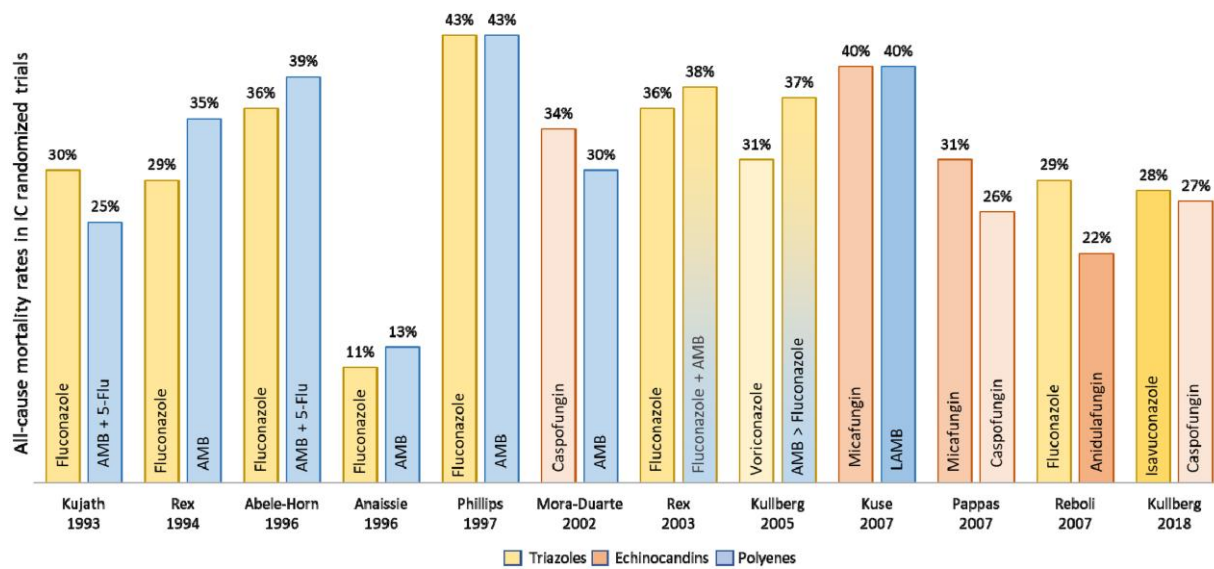


Figure 2. All-cause mortality rates in IC randomized trials (based on data reported by Demir).¹⁸ The figure does not account for differences in study design, namely number of patients randomized, and only includes antifungals currently reimbursed. AMB, amphotericin B; 5-Flu, 5-fluorocytosine; LAMB, lipid formulation of amphotericin B.

anti-mannan assays combined were shown to achieve both sensitivity and specificity $\geq 80\%$ for patients with *C. albicans*, *N. glabrata* or *C. tropicalis* infections.¹⁹ It has also been suggested that the combined use of biomarkers could be used as a complementary decision-support tool for the diagnosis and management of IC.^{19,52} For example, a combination of (1,3)- β -D-glucan, mannan and anti-mannan serum assays significantly shortened the duration of antifungal treatment in ICU patients with suspected IC, with no negative impact on outcome.⁵⁷ However, nonculture diagnostic tests also have limitations (Table 1) and ideally should be used as a complement to culture tests, while also taking into account patient characteristics, including the specific host group and severity of the clinical scenario to culture tests.^{6,50,54,60}

Role of candidaemia risk scores

Due to the underlying limitations of diagnostic tests, prediction rules or scoring systems have been proposed as early diagnostic tools to assess the risk of IC early in patients admitted to ICU, such as the *Candida* Colonization Index (CCI)⁶¹ and the *Candida* score.⁶² There are concerns that scoring systems for IC have a high negative predictive value but a low positive predictive value, which means that they may be more useful to rule out patients who do not have or will not develop IC.^{20,45,46,62,63} This is partially explained by the poor specificity of risk factors for developing IC, aligned with the low prevalence of IC in most clinical settings,¹⁹ which would make many patients eligible for antifungal therapy even when their real risk is low,⁶⁴ thus increasing its use and consequently the risk of selecting resistant strains.⁶⁵ The use of the *Candida* score is largely debated, and not validated for all populations. While this risk score was shown to have sensitivity and specificity for invasive candidiasis of 81% and 74%, respectively, the population tested mostly included surgical ICU patients, with only 35% of admissions for medical reasons.⁶² Thus, this tool may be less reliable for patients with nonsurgical reasons for ICU admission.

Other generic scoring systems that can be used to assess the risk of IC include the acute physiology and chronic health evaluation (APACHE II) score, which classifies disease severity and predict mortality in ICU patients,^{46,66–68} and the simplified acute physiology score (SAPS II), also a prognostic model for hospital mortality at ICU admission.^{69–71} However, the utility of these tests for IC is not clearly defined.

It has also been suggested that biomarkers of fungal infection, such as BDG, could be superior to candidaemia risk scores to support the decision to initiate treatment earlier.^{46,64,72} In one study including 95 patients with sepsis and >5 days in the ICU, a single negative BDG test at sepsis onset had a negative predictive value for candidaemia of 98.7%, and both negative and positive predictive values of such biomarkers were superior to the *Candida* score.⁷² These encouraging study results require further confirmation in high-quality studies.⁶³ Advances in genetic polymorphisms identification in the host have shown promising results in the identification of patients with a genetic predisposition to develop IC, which would put them at higher risk and more likely to benefit from antifungal treatment.⁷³ Using data from a prospective observational cohort study of 89 high-risk surgical ICU patients, the authors showed how one single-nucleotide polymorphism increased the susceptibility to intra-abdominal candidiasis infection.⁶⁹ Albeit promising, these results need to be confirmed in larger studies.⁷⁴ Finally, the use of risk scores (e.g. *Candida* score) could be combined with diagnostic tests with short turnaround, such as BDG detection, for a more accurate and time-efficient prediction of IC.⁷⁵

Critical factors for the management of IC

Treatment

The treatment of IC has evolved in the last three decades. Generally, treatment guidelines^{21,76} are more relevant for patients with candidaemia, as available IC evidence mostly came

Table 1. Overview of key diagnostics

Test	Advantages	Limitations
Microscopy	Fast turnaround ⁴⁶ High sensitivity when using fluorescent brightener staining ^{6,51}	Inability to identify species ⁶
Blood culture	Species identification ¹⁹ Susceptibility pattern	Slow turnaround ^{21,53} Timing of blood collection, during the course of infection ¹⁹ Necessary to culture a large blood volume (40 mL) in aerobic flasks ⁵³ When <i>Candida</i> density is low (<1 cfu/mL), blood cultures can result in false negatives Cultures may become negative after initiating antifungal therapy ⁵ May require invasive procedures ¹⁹ Cultures may become negative after initiating antifungal therapy ⁵ Long incubation required for optimal performance (3 days) ⁶ For intra-abdominal candidiasis, lack of specificity to differentiate infection from colonisation ³⁵
Sterile site cultures	Species identification Susceptibility pattern	Serial determinations required ²² Lower utility in immunosuppressed hosts ⁵² May not distinguish between past and acute infections ¹⁹ Sensitivity varies regarding <i>Candida</i> species (better for <i>C. albicans</i> , <i>N. glabrata</i> and <i>C. tropicalis</i>) ⁵⁴ Decreased specificity if <i>Candida</i> colonisation is present ⁵⁴ Low positive predictive value, potentially leading to antifungals overuse Limited by low serum concentrations and rapid bloodstream clearance ¹⁹ Not species-specific, requiring further tests to identify the fungus ⁴⁶ No data on susceptibility pattern Not approved by FDA ¹⁹ Not universally available
Mannan antigen/anti-mannan antibodies	Early detection ^{22,54} Useful to rule out infection ²²	Serial determinations required ²² Lower utility in immunosuppressed hosts ⁵² May not distinguish between past and acute infections ¹⁹ Sensitivity varies regarding <i>Candida</i> species (better for <i>C. albicans</i> , <i>N. glabrata</i> and <i>C. tropicalis</i>) ⁵⁴ Decreased specificity if <i>Candida</i> colonisation is present ⁵⁴ Low positive predictive value, potentially leading to antifungals overuse Limited by low serum concentrations and rapid bloodstream clearance ¹⁹ Not species-specific, requiring further tests to identify the fungus ⁴⁶ No data on susceptibility pattern Not approved by FDA ¹⁹ Not universally available
CAGTA	Fast turnaround and low cost ⁵ Could be used to detect whether candidaemia originated in a catheter or deep organs ⁵⁵	May not distinguish between past and acute infections ¹⁹ Limited by low serum concentrations and rapid bloodstream clearance ¹⁹ Sensitivity varies according to <i>Candida</i> species (lower for <i>C. tropicalis</i>) ^{5,19} Not species-specific, requiring further tests to identify the fungus ⁴⁶ Low positive predictive value, potentially leading to antifungals overuse No data on susceptibility pattern Not approved by FDA ¹⁹ Not universally available
BDG	Early detection ²² Useful to rule out infection ²²	Serial determinations required ²² Lower utility in patients with haematological disease ²² and immunosuppressed hosts ⁵² Sensitivity varies according to <i>Candida</i> species (lower for <i>C. parapsilosis</i>) ¹⁹ May not distinguish between past and acute infections ¹⁹ Not species-specific, requiring further tests to identify the fungus ⁴⁵ Low positive predictive value, potentially leading to antifungal overuse No data on susceptibility pattern Not universally available
Nucleic acid amplification-based methods	PCR	Early detection ⁵⁸ Monitoring of persistence or resolution of infection ⁴ Mostly developed in-house or commercially ¹⁹ Frequently performed in reference laboratories limiting the advantage of short turnaround time ⁴⁵ Data interpretation impaired by test heterogeneity ¹⁹ Not universally available
	T2Candida	Early detection ^{19,59} Automated molecular diagnosis ^{49,59} May detect candidaemia missed by cultures during Costs associated with the test ⁴⁶ Limited to some <i>Candida</i> species (<i>C. albicans</i> / <i>C. tropicalis</i> , <i>N. glabrata</i> / <i>P. kudriavzevii</i> , and <i>C. parapsilosis</i> , groupings that are based on typical antifungal susceptibility pattern) ^{49,59}

Continued

Table 1. *Continued*

Test	Advantages	Limitations
	empirical or pre-emptive AF therapy ⁴ Improved performance in neutropenic patients ⁴	No data on susceptibility pattern ⁵⁹ Not universally available

AF, antifungal; BDG, β -D-glucan; CAGTA, *C. albicans* germ tube antibody; cfu, colony forming units; FDA, Food and Drug Administration; PCR, polymerase chain reaction.

from trials enrolling patients with candidaemia, with fewer trials investigating deep-seated candidiasis.^{21,77–82} The rarer forms of IC have seldom been studied in prospective studies; hence, treatment regimens for these forms are based on anecdotal experience and retrospective case series.

The antifungal drugs available for the treatment of IC belong to three classes: echinocandins (anidulafungin, caspofungin or micafungin), azoles (fluconazole, voriconazole, itraconazole, posaconazole, isavuconazole) and amphotericin B-based regimens.^{23,76} The comparative effectiveness of these agents for the treatment of IC was recently reported by a network meta-analysis, which included data from 13 trials that randomized 3528 patients to one of the three antifungal classes.¹⁸ Results showed that echinocandins were associated with best clinical outcomes (i.e. response to antifungal therapy) when compared with the other two groups of agents. Moreover, a combined analysis of clinical studies involving almost 2000 patients showed that initial therapy with an echinocandin is a significant predictor of survival.⁸³ Overall, these agents have shown efficacy in 70%–75% of patients in randomized clinical trials.^{79–81,83–85} Accordingly, guidelines recommend echinocandins as a first-line treatment in most IC patients, without preference for a specific compound, mostly due to their broader spectrum of activity, higher fungicidal activity for most *Candida* species, low drug–drug interaction, rare acquired resistance and increased safety profile.^{18,76}

Azoles are generally well tolerated, but they have been shown to be about 15% less effective than echinocandins on average.⁸⁶ They are used instead of echinocandins as first-line therapy in some forms of deep-seated candidiasis, such as brain, intraocular and urinary tract infections, where echinocandins have lower penetration. In terms of formulations, echinocandins are approved for once-daily intravenous administration, and azoles can be administered intravenously or orally. Amphotericin B deoxycholate formulations have been associated with severe adverse events, such as nephrotoxicity and infusion-related adverse effects; hence, lipid formulations have been developed that present fewer, but still frequent, toxicities.^{78,87} These formulations are commonly used to treat patients who are intolerant or resistant to echinocandins and/or azoles, as well as in some deep-seated infections such as endocarditis, meningoencephalitis and endophthalmitis.⁷⁶

Although the use of echinocandins as first-line therapy has increased and the major role for fluconazole in the current management of IC is for step-down therapy,^{7,88} fluconazole is sometimes still used as first-line therapy, as shown by a

retrospective chart review of diagnostic and treatment decisions in patients with candidaemia conducted in six German hospitals.⁸⁹ This contrasts with ESCMID guidelines, but is in line with the Infectious Diseases Society of America (IDSA) guidelines accepting fluconazole as alternative for those not critically ill and without prior azole exposure.²¹ Further deviations from international guideline recommendations include the indication, dosage, route of administration and duration, with approximately half of the prescriptions being assessed as inappropriate.^{90,91}

Improving patient outcomes

Early initiation of antifungal therapy has been shown to reduce hospital mortality but requires starting antifungal therapy within 24 hours of taking blood cultures.⁹² As *Candida* spp. generally take longer than 24 hours to grow, the benefit from this time window is lost when relying on culture results. Most antifungal treatments are thus started empirically, when patients who are at high risk of developing IC are persistently febrile in the absence of microbiological evidence of infection.⁷⁶ Recent international guidelines for the management of sepsis and septic shock suggest that empirical antifungal therapy should be preferred for adults at high risk of fungal infection.⁹³ Empirical therapy has been associated with reduced overall mortality, although most evidence comes from uncontrolled studies.⁷⁶ Importantly, a Cochrane review reported that, on the basis of 19 studies that included 2374 non-neutropenic critically ill patients, empirical antifungal treatment reduced the risk of invasive fungal infection but did not reduce all-cause mortality.⁹⁴ Moreover, the broad use of empirical antifungals increases healthcare costs and is potentially linked with antifungal resistance.^{1,91,95,96}

Prompt source control (i.e. the elimination of the focus of infection) is also key in the management of IC. This may consist of the removal of contaminated intravascular catheters, infected prosthetic devices (for example, cardiac pacemaker leads), prosthetic joints or other devices, as well as the adequate drainage of infected material (such as peritoneal fluid, pleural fluid and/or abscess material) and surgical correction of the underlying pathology (e.g. perforation or leak).^{97,98} Source control is important due to the ability of *Candida* spp. to form biofilms on implanted medical devices representing a persistent nidus of infection.⁹⁹ In addition, biofilm formation is linked to the development of antifungal resistance due to decreased drug penetration and up-regulation of resistance mechanisms.^{100,101} A recent systematic review analyzed data from 34 prospective and retrospective cohort and case-control studies, finding that a central venous

catheter was associated with a significantly increased risk of developing IC (odds ratio 4.7, 95% confidence interval 2.7 to 8.1).⁶⁴ Retrospective studies of adult patients diagnosed with candidaemia show how intravascular catheters were a risk factor for infection⁵ and how catheter removal was associated with increased odds of survival.⁸⁹ Removal of indwelling intravascular catheters is therefore strongly recommended when candidaemia is present.^{51,76} If removal is not possible then treatment with a lipid-based amphotericin B formulation or an echinocandin is suggested,⁷⁶ as these drugs have shown activity against *Candida* biofilms.¹⁰² Catheter lock strategies, using high antifungal concentrations locally in the catheter lumen for hours or days, have also been performed. Lipid-based amphotericin B formulations and echinocandins are again the most commonly used antifungals, showing high efficacy to decrease biofilm formation but commonly failing to eradicate the *Candida* infection.¹⁰³ Novel strategies are being investigated including antifungal combination therapy, phototherapy, cationic peptides and even the use of plant-based therapies.^{104,105}

Diagnosis-driven drug management

The duration of antifungal treatment is often guided by the extent of organ involvement.⁷⁶ For candidaemia, the ESCMID recommends for treatment to be continued for 14 days after the last negative blood culture, whereas organ involvement may be screened with transoesophageal echocardiography and fundoscopy.⁷⁶ Rapid diagnostic tests can support early discontinuation of empirical antifungal therapy, with retrospective data showing that results from T2Candida panel performed better than BDG, when combined with blood cultures, decreasing the number of days critically ill patients were on empirical echinocandin therapy.¹⁰⁶ The decision to de-escalate treatment from intravenous echinocandins to oral fluconazole should take into account not only diagnostics but also patient's stability, tolerance of the administration route and species susceptibility.^{4,76,79,89,93,107}

The role of nonculture tests to guide treatment, including drug de-escalation, has also been explored. Evidence from a randomized trial of 234 critically ill non-immunocompromised patients admitted to ICU who were allocated to an echinocandin or placebo suggested that BDG monitoring could be used to decide when to de-escalate empirical therapy or when to withhold preemptive therapy,⁶⁰ whereas T2Candida has also been shown to improve management.^{4,108}

Therapeutic drug monitoring

Therapeutic drug monitoring (TDM) is used to measure antifungal drug levels to prevent over or underdosing. The IDSA and ESCMID recommend TDM when treating IC with voriconazole; ESCMID also advise TDM when prescribing 5-fluorocytosine and posaconazole.^{21,22}

Whenever antifungal absorption or excretion could be hindered [i.e. mucositis, administration via nasogastric tube or gastrostomy, or in critically ill patients undergoing continuous renal replacement therapy (CRRT)], TDM should also be considered.^{109,110} Finally, TDM may also be helpful in those cases of treatment failure, breakthrough infections, serious toxicity or

those *Candida* infections caused by species with high minimum inhibitory concentration (MIC).¹¹¹

Drug resistance

Candida infections resistant to one or multiple types of antifungals are increasingly being diagnosed, with prior antifungal therapy found to be the main driver for resistance selection.^{20,112,113} The widespread use of antifungals introduces a positive selection of *Candida* spp. that show some intrinsic or acquired resistance to treatment, including *N. glabrata* and *C. auris* (multidrug-resistant),^{114,115} as well as *P. kudriavzevii* (azole resistant). Data gathered from population-based or multicentre studies showed that the rates of azole resistance vary considerably depending on the setting and the *Candida* spp., ranging from 3% to 21%.²⁰

Acquired echinocandin resistance has also been reported, especially for *N. glabrata* and *C. tropicalis*. In the SENTRY antifungal surveillance programme, mutations in *FKS* gene hot spot regions were detected among echinocandin-resistant isolates, most of which were resistant to two or more echinocandins. Additionally, *C. parapsilosis* has an intrinsic polymorphism affecting also the *FKS1* gene, leading to decreased *in vitro* echinocandin susceptibility.¹¹⁶ However, the clinical impact of such reduced susceptibility remains controversial. *C. auris* was initially detected in Japan in 2009 and shows some resistance to all major antifungal treatments, including multi-drug-resistant isolates, with higher resistance for fluconazole, followed by amphotericin B and echinocandins.^{9,117} Hospital outbreaks of *C. auris* with rapid spread and high mortality have been since reported in Asia (India, Pakistan), Europe (the UK, Spain, Italy), Latin America (Colombia, Venezuela, Panama) and the USA.¹¹⁷⁻¹¹⁹

The rise in drug-resistant IC has highlighted a need for antifungal susceptibility testing, to achieve optimal treatment and to monitor the emergence of antifungal resistance. Several tests are currently available, including broth microdilution according to the CLSI¹²⁰ and the EUCAST,¹²¹ which represent gold standards for antifungal susceptibility testing. Alternative tests include disc diffusion, epsilometer tests, colorimetric broth microdilution and automated spectrophotometric systems.¹²² Generally, these methods can be time-consuming and/or technically complex, and the interpretation of results may be challenging.

Clinical challenges related to specific forms of IC

IC in ICU patients

Approximately 50% of episodes of IC occur in ICU, where the administration of antibiotics and immunosuppressive drugs, combined with the use of invasive procedures (e.g. the installation of central vascular catheters), total parenteral nutrition¹⁶ or intrabdominal surgery,^{36,38} significantly increase the risk of *Candida* infection.^{2,3,32} Length of ICU stay is consistently reported as increasing the risk of developing IC, although it is seldom possible to disentangle length of ICU stay from other confounding risk factors, as patients with longer ICU stays will have increased disease severity as well as more invasive therapies.^{50,64} All-cause mortality associated with candidaemia seems to be 2-fold higher for patients hospitalized in the ICU when compared with patients

Table 2. Antifungal dose adaptations during CRRT

Antifungal agent	Mechanism of action	Route of administration	Adverse effect	Elimination by CRRT	Recommended dose during CRRT
Lipid formulation of amphotericin B	Interacts with ergosterol in the fungal cell membrane	IV	Hepatic, renal and cardiovascular toxicity	Unaffected by CRRT	5 mg/kg/d
Fluconazole	Interacts with 14-demethylase in the fungal cell membrane	IV or oral	Hepatic toxicity	High elimination by CRRT	600 mg/12 h
Voriconazole	Reduces ergosterol synthesis	IV or oral	AKI toxicity with IV use, hepatic toxicity	Poor elimination of IV form by CRRT — No adaptations for CRRT	Loading dose: 6 mg/kg/12 h Maintenance dose: 4 mg/kg/12 h
Anidulafungin	Inhibits (1,3)- β -D-glucan synthetase	IV	Hepatic toxicity	Significant adsorption by CRRT adsorptive membranes	Loading dose: 200 mg/d Maintenance dose: 150 mg/d
Caspofungin	Interacts with 14-lanosterol demethylase in the fungal cell membrane and reduces ergosterol synthesis	IV	Severe hepatic toxicity	Unaffected by CRRT	Loading dose: 70 mg/d When BMI is >40 higher doses can be used (up to 140 mg/d) Maintenance dose: 50 mg/d (if >80 kg, maintenance with 70 mg/d is recommended)

AKI, acute kidney injury; CRRT, continuous renal replacement therapy; d, day; IV, intravenous.

in other hospital settings, although these results vary for different *Candida* spp.¹²³

In critically ill patients, severe comorbidities can alter the antifungal drugs' pharmacological profile. Drug distribution can be influenced by haemodynamic alterations, while hepatic and/or renal impairment can affect drug concentration in the bloodstream, metabolism and elimination. Additionally, hypoalbuminaemia can reduce the percentage of bound antifungal, increasing activity and potentially toxicity.¹²⁴

Dosing and PK/PD are also crucial during CRRT and extracorporeal membrane oxygenation (ECMO), which affect different classes of antifungals. Fluconazole is the most complex to dose in ICU during CRRT (Table 2) due to the high extracorporeal removal of fluconazole, which exceeds the normal renal clearance and necessitates a higher daily maintenance dose.¹²⁵ In line with available data and as long as hepatic function remains stable, a dose of fluconazole of 500–600 mg every 12 h is recommended in critically ill patients under CRRT.¹²⁶

The COVID-19 pandemic led to high ICU admissions and subsequently triggered an increase in the number of secondary invasive fungal diseases, including *Candida* and *Aspergillus*,³⁴ partially due to the high doses of corticosteroids given.¹²⁷ Aggregated data showed that by September 2020 the extent of candidiasis associated with COVID-19, both superficial and invasive, ranged between 0.7% and 23.5%.¹²⁸

Intra-abdominal infections

Intra-abdominal candidiasis is the most common type of deep-seated candidiasis.⁹⁸ A prospective study with 176 non-

neutropenic critically ill patients with severe abdominal conditions and a high prevalence of IC (18%) showed how BDG (cut-off value of 259 pg/mL) and CAGTA (positive versus negative) accurately discriminated between *Candida* spp. colonization and IC.¹²⁹ However, currently there are no tight criteria to distinguish abdominal *Candida* colonization from true *Candida* infection, and general criteria for sepsis and septic shock are often used to differentiate between abdominal colonization and infection.¹³⁰ This is an issue as most of the time sepsis and septic shock are due to causes other than true *Candida* infection, and thus many abdominal colonizations, especially in the post-operative period, are overtreated leading to more antifungal resistance. There is an urgent need to build an algorithm to differentiate abdominal *Candida* colonization from true infection.

The burden of intra-abdominal infections and its associated morbidity and mortality are higher in high-income countries, which may be partially explained by the widespread use of antibiotics and increased drug resistance.¹³¹

Clinical guidelines/expert consensus papers recommend prophylactic use of fluconazole for patients with abdominal surgery and recurrent gastrointestinal perforations or anastomotic leakages,^{35,76} with echinocandins or lipid formulation of amphotericin B being recommended as first-line antifungal therapy for critically ill patients or patients with previous exposure to azoles.³⁵ However, prophylactic use of fluconazole may be associated with an increase in fluconazole-resistant species.¹³²

The outstanding clinical challenges in intra-abdominal candidiasis thus include prophylactic use of fluconazole, need for source control and treatment selection and duration.

IC in patients with haematological malignancies

Owing to their compromised immune response, patients affected by haematological malignancies are prone to develop IC with high mortality risk, prolonged hospitalization and rising healthcare costs.^{15,37} This is a consequence of host defences being affected by cancer treatments, such as cytotoxic chemotherapy, ablative radiotherapy and immunosuppressive therapies. Other risk factors mentioned previously also apply to haematological cancer patients.^{12,37,133} Several studies have reported candidaemia-associated mortality rates in patients with haematological malignancies ranging between 29.5% and 45%.^{12,13,15,133}

To counteract the high incidence of candidaemia and its associated mortality, the use of fluconazole prophylaxis was introduced in the early 1990s.⁴² Clinical guidelines recommend anti-*Candida* prophylaxis for patients receiving allogeneic stem cell transplantation.²³ Prophylactic treatment with fluconazole has also been recommended for patients undergoing remission-induction¹³⁴ or salvage-induction¹³⁵ chemotherapy for acute myeloid leukaemia.

However, while prophylactic use of fluconazole succeeded in lowering the frequency of bloodstream infections caused by azole-sensitive *C. albicans*, it caused an increase in azole-resistant species such as *N. glabrata* and *P. kudriavzevii*, which now account for most candidaemia episodes in many cancer centres.¹³⁶ Additional concerns include drug-drug interactions, tolerability and breakthrough fungal infections,¹³⁴ as well as the non-specificity of signs and symptoms of IC in patients who frequently present with fever and sepsis. In a pooled analysis of 1271 patients with haematologic malignancies and patients undergoing haematopoietic stem cell transplant, echinocandins seemed to be marginally more effective than triazoles for prophylactic treatment.⁴⁸

Breakthrough candidaemia

Breakthrough candidaemia is defined as candidaemia that develops during systemic antifungal therapy administered as either prophylaxis, pre-emptive, empirical or targeted therapy.^{21,135} It has been linked to the emergence of drug resistance and poor outcomes,^{137–140} and several risk factors have been identified. A retrospective study conducted in Brazil from 2011 to 2016 identified 27 breakthrough episodes from 148 candidaemia episodes, with neutropenia and mucositis being independent risk factors and non-*albicans* species being more frequent among these patients.¹⁴¹ Similarly, in a multicentre study of hospitalized adults with candidaemia, *P. kudriavzevii* was more frequent and fluconazole-resistance was independently associated with risk of breakthrough episodes.¹⁴² Another study identified neutropenia, use of corticosteroids and heavy antibiotic exposure (previous use of two or more antibiotics for at least 14 days) as risk factors.¹⁴³ Additionally, a 3-year prospective study conducted in 567 consecutive cases of candidaemia recorded 37 cases of breakthrough candidaemia, 86% of which on fluconazole; breakthrough candidaemia was associated with gastrointestinal mucositis, graft-versus-host-disease, immunosuppression and parenteral nutrition, and non-*albicans Candida* were isolated in most breakthrough cases.¹⁴⁴ Overall, breakthrough candidiasis appears to be caused by drug-resistant, non-*albicans* species

selected by the use of antifungals and impairments in the immune response.

Sanctuary site candidiasis

CNS candidiasis

Candida CNS infections, mostly caused by *C. albicans*,¹⁴⁵ are rare but severe. They can arise from haematogenous spread, mostly in neonates due to blood-brain barrier immaturity, or in the presence of ventricular drainage devices or following neurosurgical procedures.¹⁴⁶ Increased risk in adults has been described for the immunocompromised patients^{145,147–149} and individuals who have a deficiency of the lectin receptor adaptor molecule CARD9.¹⁵⁰ Moreover, *Candida*-caused endocarditis are associated with CNS embolic complications in 12% to 22% of cases.^{148,151,152}

The most common manifestation of brain infection is overt meningitis, while in rarer cases chronic meningitis, brain abscesses, vasculitis with cerebral infarctions, spinal infections, ventriculitis and mycotic aneurysms can be observed.¹⁵³ A recent nationwide retrospective study conducted in France and covering the period between 2005 and 2018 identified 24 adult patients with CNS candidiasis. Mortality attributed to CNS candidiasis was 42%.¹⁵⁴

Treatment guidelines suggest the use of liposomal amphotericin B combined with flucytosine for CNS IC. Fluconazole may be used as a step-down therapy, while poor penetration of echinocandins limit their use in CNS. However, due to data scarcity, no strong recommendation is given.⁷⁶ A study reported the use of amphotericin B deoxycholate combined with flucytosine for >2 weeks in a series of HIV-infected patients, with four of five patients being treated successfully.¹⁴⁷ In two other series, 27 of 34 patients survived after similar treatments.^{155,156} Published data on voriconazole use in CNS candidiasis are sparse; efficacy may be limited by the variability of its concentration in the CSF.¹⁵⁷

Urinary candidiasis

Critically ill patients and those with urinary catheters are at high risk of developing candiduria.⁷³ In critically ill patients undergoing surgical procedures, the rates of concurrent candidaemia derived from a urinary source may reach up to 10%.¹⁵⁸ Additional risk factors for adult candiduria include advanced age, female gender, urinary tract anatomic abnormalities, abdominal surgery, multi-morbidity, broad-spectrum antibiotics therapies and diabetes mellitus.¹⁵⁸

If asymptomatic, then treatment is not recommended for candiduria, except in pre-operative patients who may be given fluconazole.⁷⁶ If possible, the urinary catheter should be removed to clear the infection,⁷⁶ and early urologic drainage procedure is also associated with improved outcomes.¹⁵⁹ The prophylactic antifungal treatment of *Candida* colonization in the urinary tract is a common inappropriate use of antifungal agents.⁴⁵ Different combinations of antifungal agents are available for symptomatic candiduria, including fluconazole or amphotericin B deoxycholate, with or without flucytosine; if fungus balls or casts are detected then surgical intervention is required.^{21,76} For the treatment of fluconazole-resistant strains, only amphotericin B and micafungin seem to reach adequate

urinary concentrations, although the clinical evidence is limited.^{160,161}

Endophthalmitis and chorioretinitis

In rare cases, candidaemia can give rise to two types of ocular infection: *Candida* chorioretinitis, restricted to the chorioretinal layers, and *Candida* endophthalmitis, usually extending into the vitreous body. The latter is associated with poor visual outcomes.¹⁶² Candidaemia-associated endophthalmitis and chorioretinitis can be treated with antifungals administered systemically or locally (i.e. via intravitreal injection). Systemic amphotericin B and echinocandins do not penetrate well in the vitreous humour, whereas fluconazole and voriconazole can reach therapeutic vitreous concentrations.^{163–165}

A randomized multicentre trial that compared voriconazole with amphotericin B followed by fluconazole for the treatment of candidaemia reported that ocular involvement occurred in 16% of patients with candidaemia, mostly manifesting as chorioretinitis, whereas endophthalmitis was uncommon (1.6%); treatment with either voriconazole or amphotericin B followed by fluconazole was successful for ocular candidiasis in most (65%) cases.¹⁶⁶ According to clinical guidelines, fluconazole or voriconazole are recommended as the drugs of choice for susceptible isolates, whereas liposomal amphotericin B either alone or combined with flucytosine is recommended when the susceptibility of the isolate is unknown. In the case of endophthalmitis, vitrectomy and intravitreal injection of amphotericin B are recommended in addition to systemic therapy.⁷⁶

Additional challenging infection sites

Candida endocarditis

Candida species cause <2% of all infective endocarditis cases and can arise on a native valve, a prosthetic valve or in the presence of pacemaker or other implanted material. *Candida* endocarditis might be considered a biofilm-related infection following earlier fungal bloodstream infection. Overall, prognosis is poor with 1-year mortality >50% and substantial relapse rates.^{149,167,168} *Candida* vegetations are typically larger and more friable than bacterial ones, harbouring a higher risk for embolic events, ophthalmologic complications and cutaneous lesions. Due to this high mortality, early diagnosis and initiation of antifungals is vital.^{167,168} Most cases are identified with transthoracic echocardiography, although transoesophageal echocardiography could help improve diagnosis. Although blood cultures are mostly positive in *Candida* endocarditis,¹⁶⁹ its yield can be lower in patients receiving prior antifungals and cultures can take several days to produce results. Hence, attempts should be made to get a tissue sample for diagnosis. Moreover, nonculture methods could have an important role in early diagnosis, such as BDG detection and PCR amplification, with a sensitivity of 89% and over 92% for fungal endocarditis, respectively, according to a recent systematic review.¹⁶⁹

A recent review of 140 cases of *Candida* endocarditis showed that surgery, effective antibiofilm treatment (defined in such study as antifungal treatment with liposomal amphotericin B or echinocandins) and chronic suppressive antifungal therapy were independently associated with improved prognosis.¹⁷⁰

Accordingly, international guidelines recommend surgery whenever feasible, as well as antifungal treatment with liposomal amphotericin B, which can be combined with flucytosine,⁷⁶ or high-dose echinocandins. Additionally, long-term suppressive therapy should be considered for those patients who cannot undergo valve replacement and in those cases of prosthetic valve endocarditis. Finally, risk factors for *Candida* endocarditis are un-specific and this entity should be considered in patients with relapsing or persistent candidaemia, especially in critically ill patients.¹⁷¹

Bone and joint candidiasis

These infections include osteomyelitis/spondylodiscitis, arthritis and prosthetic joint infection, commonly following haematogenous spread are also possible. The spine is the most common site of osteomyelitis involvement.¹⁷² Although fluconazole monotherapy has been classically recommended, it is plausible that a biofilm component exists. Thus, initial ‘induction’ treatment with an echinocandin or lipid-based amphotericin, followed by long-term fluconazole therapy (6–12 months), seems advisable.^{21,173,174} For septic arthritis, surgery is mandatory and in cases of prosthetic joint infections, removal of the joint prosthesis is advised. If this is not possible, lifelong fluconazole therapy is commonly indicated.⁷⁶ If reimplantation of the prosthesis is considered, guidelines recommend administering antifungal treatment for at least 12 weeks before and 6 weeks after prosthesis implantation.²¹ Again, biofilm-active antifungals are conceptually desirable, although their use is limited by the need for intravenous administrations. Moreover, the quality of evidence that supports these recommendations is very low, and this scenario is further complicated by the reported increase in non-*albicans* species with decreased azole susceptibility^{173,175,176} and frequent bacterial co-infection.

Candida pneumonia

While *Candida* species are often isolated from the respiratory tract of ICU intubated patients or patients with tracheostomies, the existence of *Candida* pneumonia has been largely debated.¹⁷⁷ Indeed, the true incidence of *Candida* pneumonia ranges from 0.23% to 0.4%, with increased risk linked to genetic predisposition and severe immunodeficiency.^{7,21,177} In this context, antifungal therapy should only be considered in immunocompromised patients on mechanical ventilation with biopsy-proven candidiasis and without an alternative aetiology.¹⁷⁷

Conclusions and perspectives

Candida is still one of the main fungal pathogens responsible for serious fungal disease, and non-*albicans* species as well as multidrug-resistant *Candida* infections are increasingly detected in clinical settings. Several key challenges need to be tackled to improve the clinical management and outcomes of IC patients. First, collecting and comparing global epidemiological data for IC is hindered by the absence of specific criteria for an incidence denominator and inconsistently collected data. Large longitudinal studies, alongside regional and local surveillance studies, are required to understand epidemiological trends and shifts and

to collect data to guide and support empirical antifungal therapy. Another key challenge concerns the currently available diagnostic tests and risk scoring tools, which have a limited capacity to inform appropriate treatment and improve clinical outcomes in a very heterogeneous population of patients. Additionally, the lack of standardized effectiveness outcomes and long-term data for IC limits the ability to capture downstream consequences of treatment pathways, namely treatment failures.¹⁷⁸ Understanding which elements of care, like survival or length of stay, are most likely to be affected by treatment, and standardizing those outcomes and their reporting across trials, would promote comparability across treatments.

The timing for the initiation of antifungal therapy is also a key aspect to improve patient outcomes, hindered by the diagnostics limitations. Empirical antifungal therapy is recommended for adults who are at high risk of fungal infection, alongside prompt source control to eliminate the focus of the infection, when possible. Finally, treatment-wise, echinocandins have shown promising efficacy and safety results¹⁸ and are recommended by guidelines as first-line treatment in most IC patients, but they carry higher costs of drug acquisition and administration compared with older-generation antifungal agents.^{179,180} Furthermore, available cost-effectiveness data for echinocandins in treating IC are limited, both quantitatively and qualitatively,¹⁷⁸ and inconsistently reported. In addition, few studies are available to support recommendations on the optimal step-down therapy from echinocandins to azoles or the total duration of therapy. Strategy studies should prospectively identify optimal step-down protocols as well as explore further limitations of the total duration of antifungal treatment in selected patient groups.

Several new drugs have shown promising efficacy against IC in recent phase 2 and 3 clinical trials. Rezafungin is a novel echinocandin with an extended half-life and prolonged therapeutic drug concentrations in peripheral tissues, allowing weekly versus daily administration compared to existing echinocandins. These pharmacodynamics properties simplify treatment of outpatients requiring extended therapy and make rezafungin a viable alternative for IC prophylaxis in haematological patients. A phase 2 trial has demonstrated that the efficacy and safety of rezafungin are comparable to those of the other echinocandins,⁸² while recently published results of the phase 3 trial ReSTORE have shown that rezafungin is non-inferior to caspofungin in patients with candidaemia or IC regarding day-14 global cure and 30-day all-cause mortality, with no differences in adverse events occurrence.¹⁸¹ Ibrexafungerp is an oral glucan synthase inhibitor with broad activity against *Candida* spp., including azole resistant (e.g. *C. auris*), and it has a comparable efficacy and safety with standard of care, according to a small phase 2 trial.^{182,183} An ongoing salvage study suggests it could be used for drug-resistant infections as an alternative to echinocandins (NCT03059992). Fosmanogepix is a guanosine monophosphate inhibitor with a broad activity against *Candida* species (except against *P. kudriavzevii*) and can be administered orally and parenterally twice daily.¹⁸⁴ A small phase 2 clinical trial has shown good efficacy and safety in fluconazole-resistant IC¹⁸⁵; a similar study on *C. auris*-candidaemia has recently successfully met its clinical objectives (NCT04148287). ATI-2307 mitochondrial inhibitor was found to have *in vitro* and *in vivo* activity against most *Candida* species and has potential to be used for drug-resistant *Candida*

infections.¹⁸⁶ However, no comparative clinical trials have been conducted so far.

In addition to new drug development, there is an increased interest in combination therapy to tackle the rising of drug resistance and the high mortality rates in IC. A recent systematic review found that, although the effect of combination treatments varied greatly across studies depending on *Candida* species, drug and methodology used, some combination regimens had a synergistic effect on difficult-to-treat species or had higher efficacy than monotherapy on the prevention/reduction of biofilms and the clearance of infected tissues.¹⁸⁷ However, these data are subject to substantial biases, and further data on combination therapy is needed.

The availability of new compounds and combination therapy overcome some of the challenges identified and increase our options for management of chronic *Candida* infections and ambulant patient treatments. However, early identification of patients that require antifungal therapy and treatment of sanctuary site infections remain a challenge and will require further innovations.

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Supplementary data

Available as [Supplementary data](#) at JAC Online.

References

- 1 Kullberg BJ, Arendrup MC. Invasive candidiasis. *N Engl J Med* 2015; **373**: 1445–56. <https://doi.org/10.1056/NEJMra1315399>
- 2 Logan C, Martin-Loeches I, Bicanic T. Invasive candidiasis in critical care: challenges and future directions. *Intensive Care Med* 2020; **46**: 2001–14. <https://doi.org/10.1007/s00134-020-06240-x>

- 3** Bongomin F, Gago S, Oladele RO *et al.* Global and multi-national prevalence of fungal diseases—estimate precision. *J Fungi (Basel)* 2017; **3**: 57. <https://doi.org/10.3390/jof3040057>
- 4** Clancy CJ, Pappas PG, Vazquez J *et al.* Detecting infections rapidly and easily for candidemia trial, part 2 (DIRECT2): a prospective, multicenter study of the T2Candida panel. *Clin Infect Dis* 2018; **66**: 1678–86. <https://doi.org/10.1093/cid/cix1095>
- 5** Martinez-Jimenez MC, Munoz P, Guinea J *et al.* Potential role of *Candida albicans* germ tube antibody in the diagnosis of deep-seated candidemia. *Med Mycol* 2014; **52**: 270–5. <https://doi.org/10.1093/mmy/myt025>
- 6** Pappas PG, Lionakis MS, Arendrup MC *et al.* Invasive candidiasis. *Nat Rev Dis Primers* 2018; **4**: 18026. <https://doi.org/10.1038/nrdp.2018.26>
- 7** McCarty TP, White CM, Pappas PG. Candidemia and invasive candidiasis. *Infect Dis Clin North Am* 2021; **35**: 389–413. <https://doi.org/10.1016/j.idc.2021.03.007>
- 8** Giacobbe DR, Maraolo AE, Simeon V *et al.* Changes in the relative prevalence of candidaemia due to non-*albicans* *Candida* species in adult inpatients: a systematic review, meta-analysis and meta-regression. *Mycoses* 2020; **63**: 334–42. <https://doi.org/10.1111/myc.13054>
- 9** Du H, Bing J, Hu T *et al.* *Candida auris*: epidemiology, biology, antifungal resistance, and virulence. *PLoS Pathog* 2020; **16**: e1008921. <https://doi.org/10.1371/journal.ppat.1008921>
- 10** Zhang F, Aschenbrenner D, Yoo JI *et al.* The gut mycobiome in health, disease, and clinical applications in association with the gut bacterial microbiome assembly. *Lancet Microbe* 2022; **3**: e969–83. [https://doi.org/10.1016/S2666-5247\(22\)00203-8](https://doi.org/10.1016/S2666-5247(22)00203-8)
- 11** Musuuzza JS, Watson L, Parmasad V *et al.* Prevalence and outcomes of co-infection and superinfection with SARS-CoV-2 and other pathogens: a systematic review and meta-analysis. *PLoS ONE* 2021; **16**: e0251170. <https://doi.org/10.1371/journal.pone.0251170>
- 12** Gamaletsou MN, Walsh TJ, Zaoutis T *et al.* A prospective, cohort, multicentre study of candidaemia in hospitalized adult patients with haematological malignancies. *Clin Microbiol Infect* 2014; **20**: O50–7. <https://doi.org/10.1111/1469-0691.12312>
- 13** Pagano L, Caira M, Candoni A *et al.* The epidemiology of fungal infections in patients with hematological malignancies: the SEIFEM-2004 study. *Haematologica* 2006; **91**: 1068–75.
- 14** Kontoyiannis DP, Marr KA, Park BJ *et al.* Prospective surveillance for invasive fungal infections in hematopoietic stem cell transplant recipients, 2001–2006: overview of the transplant-associated infection surveillance network (TRANSNET) database. *Clin Infect Dis* 2010; **50**: 1091–100. <https://doi.org/10.1086/651263>
- 15** Sipsas NV, Lewis RE, Tarrand J *et al.* Candidemia in patients with hematologic malignancies in the era of new antifungal agents (2001–2007): stable incidence but changing epidemiology of a still frequently lethal infection. *Cancer* 2009; **115**: 4745–52. <https://doi.org/10.1002/cncr.24507>
- 16** Manolakaki D, Velmahos G, Kourkoumpetis T *et al.* *Candida* infection and colonization among trauma patients. *Virulence* 2010; **1**: 367–75. <https://doi.org/10.4161/viru.1.5.12796>
- 17** Mayer FL, Wilson D, Hube B. *Candida albicans* pathogenicity mechanisms. *Virulence* 2013; **4**: 119–28. <https://doi.org/10.4161/viru.22913>
- 18** Demir KK, Butler-Laporte G, Del Corpo O *et al.* Comparative effectiveness of amphotericin B, azoles and echinocandins in the treatment of candidemia and invasive candidiasis: a systematic review and network meta-analysis. *Mycoses* 2021; **64**: 1098–110. <https://doi.org/10.1111/myc.13290>
- 19** Clancy CJ, Nguyen MH. Diagnosing invasive candidiasis. *J Clin Microbiol* 2018; **56**: e01909–17. <https://doi.org/10.1128/JCM.01909-17>
- 20** Lamoth F, Lockhart SR, Berkow EL *et al.* Changes in the epidemiological landscape of invasive candidiasis. *J Antimicrob Chemother* 2018; **73**: i4–i13. <https://doi.org/10.1093/jac/dkx444>
- 21** Pappas PG, Kauffman CA, Andes DR *et al.* Clinical practice guideline for the management of candidiasis: 2016 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2016; **62**: e1–e50. <https://doi.org/10.1093/cid/civ933>
- 22** Cuenca-Estrella M, Verweij PE, Arendrup MC *et al.* ESCMID* guideline for the diagnosis and management of *Candida* diseases 2012: diagnostic procedures. *Clin Microbiol Infect* 2012; **18**(Suppl 7):9–18. <https://doi.org/10.1111/1469-0691.12038>
- 23** Ullmann AJ, Akova M, Herbrecht R *et al.* ESCMID* guideline for the diagnosis and management of *Candida* diseases 2012: adults with haematological malignancies and after haematopoietic stem cell transplantation (HCT). *Clin Microbiol Infect* 2012; **18**(Suppl 7):53–67. <https://doi.org/10.1111/1469-0691.12041>
- 24** Ismail WNAW, Jasmi N, Khan TM *et al.* The economic burden of candidemia and invasive candidiasis: a systematic review. *Value Health Reg Issues* 2020; **21**: 53–8. <https://doi.org/10.1016/j.vhri.2019.07.002>
- 25** Drgona L, Khachatryan A, Stephens J *et al.* Clinical and economic burden of invasive fungal diseases in Europe: focus on pre-emptive and empirical treatment of *Aspergillus* and *Candida* species. *Eur J Clin Microbiol Infect Dis* 2014; **33**: 7–21. <https://doi.org/10.1007/s10096-013-1944-3>
- 26** Baethge C, Goldbeck-Wood S, Mertens S. SANRA—a scale for the quality assessment of narrative review articles. *Res Integr Peer Rev* 2019; **4**: 5. <https://doi.org/10.1186/s41073-019-0064-8>
- 27** Cooper C, Booth A, Britten N *et al.* A comparison of results of empirical studies of supplementary search techniques and recommendations in review methodology handbooks: a methodological review. *Syst Rev* 2017; **6**: 234. <https://doi.org/10.1186/s13643-017-0625-1>
- 28** McCarty TP, Pappas PG. Invasive candidiasis. *Infect Dis Clin North Am* 2016; **30**: 103–24. <https://doi.org/10.1016/j.idc.2015.10.013>
- 29** Wisplinghoff H, Ebberts J, Geurtz L *et al.* Nosocomial bloodstream infections due to *Candida* spp. in the USA: species distribution, clinical features and antifungal susceptibilities. *Int J Antimicrob Agents* 2014; **43**: 78–81. <https://doi.org/10.1016/j.ijantimicag.2013.09.005>
- 30** Marchetti O, Bille J, Fluckiger U *et al.* Epidemiology of candidemia in Swiss tertiary care hospitals: secular trends, 1991–2000. *Clin Infect Dis* 2004; **38**: 311–20. <https://doi.org/10.1086/380637>
- 31** Vincent JL, Rello J, Marshall J *et al.* International study of the prevalence and outcomes of infection in intensive care units. *JAMA* 2009; **302**: 2323–9. <https://doi.org/10.1001/jama.2009.1754>
- 32** Koehler P, Stecher M, Cornely OA *et al.* Morbidity and mortality of candidaemia in Europe: an epidemiologic meta-analysis. *Clin Microbiol Infect* 2019; **25**: 1200–12. <https://doi.org/10.1016/j.cmi.2019.04.024>
- 33** Bassetti M, Giacobbe DR, Vena A *et al.* Incidence and outcome of invasive candidiasis in intensive care units (ICUs) in Europe: results of the EUCANDICU project. *Crit Care* 2019; **23**: 219. <https://doi.org/10.1186/s13054-019-2497-3>
- 34** White PL, Price JS, Backx M. Evaluation of the performance of the associates of Cape Cod STAT assay for the diagnosis of invasive fungal disease in critical-care patients with COVID-19. *J Clin Microbiol* 2021; **59**: e0086921. <https://doi.org/10.1128/JCM.00869-21>
- 35** Bassetti M, Marchetti M, Chakrabarti A *et al.* A research agenda on the management of intra-abdominal candidiasis: results from a consensus of multinational experts. *Intensive Care Med* 2013; **39**: 2092–106. <https://doi.org/10.1007/s00134-013-3109-3>
- 36** Bassetti M, Righi E, Ansaldo F *et al.* A multicenter multinational study of abdominal candidiasis: epidemiology, outcomes and predictors of mortality. *Intensive Care Med* 2015; **41**: 1601–10. <https://doi.org/10.1007/s00134-015-3866-2>

- 37** Zirkel J, Klinker H, Kuhn A *et al.* Epidemiology of *Candida* blood stream infections in patients with hematological malignancies or solid tumors. *Med Mycol* 2012; **50**: 50–5. <https://doi.org/10.3109/13693786.2011.587211>
- 38** Klingspor L, Tortorano AM, Peman J *et al.* Invasive *Candida* infections in surgical patients in intensive care units: a prospective, multicentre survey initiated by the European Confederation of Medical Mycology (ECMM) (2006–2008). *Clin Microbiol Infect* 2015; **21**: e1–e10. <https://doi.org/10.1016/j.cmi.2014.08.011>
- 39** Castanheira M, Messer SA, Rhomberg PR *et al.* Antifungal susceptibility patterns of a global collection of fungal isolates: results of the SENTRY antifungal surveillance program (2013). *Diagn Microbiol Infect Dis* 2016; **85**: 200–4. <https://doi.org/10.1016/j.diagmicrobio.2016.02.009>
- 40** Pfaller MA, Moet GJ, Messer SA *et al.* Geographic variations in species distribution and echinocandin and azole antifungal resistance rates among *Candida* bloodstream infection isolates: report from the SENTRY antimicrobial surveillance program (2008 to 2009). *J Clin Microbiol* 2011; **49**: 396–9. <https://doi.org/10.1128/JCM.01398-10>
- 41** Sullivan D, Coleman D. *Candida dubliniensis*: characteristics and identification. *J Clin Microbiol* 1998; **36**: 329–34. <https://doi.org/10.1128/JCM.36.2.329-334.1998>
- 42** Marr KA, Seidel K, White TC *et al.* Candidemia in allogeneic blood and marrow transplant recipients: evolution of risk factors after the adoption of prophylactic fluconazole. *J Infect Dis* 2000; **181**: 309–16. <https://doi.org/10.1086/315193>
- 43** Jeffery-Smith A, Taori SK, Schelenz S *et al.* *Candida auris*: a review of the literature. *Clin Microbiol Rev* 2018; **31**: e00029–17. <https://doi.org/10.1128/CMR.00029-17>
- 44** Spivak ES, Hanson KE. *Candida auris*: an emerging fungal pathogen. *J Clin Microbiol* 2018; **56**: e01588–17. <https://doi.org/10.1128/JCM.01588-17>
- 45** Pfaller MA, Castanheira M. Nosocomial candidiasis: antifungal stewardship and the importance of rapid diagnosis. *Med Mycol* 2016; **54**: 1–22.
- 46** Calandra T, Roberts JA, Antonelli M *et al.* Diagnosis and management of invasive candidiasis in the ICU: an updated approach to an old enemy. *Crit Care* 2016; **20**: 125. <https://doi.org/10.1186/s13054-016-1313-6>
- 47** Strollo S, Lionakis MS, Adjemian J *et al.* Epidemiology of hospitalizations associated with invasive candidiasis, United States, 2002–2012(1). *Emerg Infect Dis* 2016; **23**: 7–13. <https://doi.org/10.3201/eid2301.161198>
- 48** Wang JF, Xue Y, Zhu XB *et al.* Efficacy and safety of echinocandins versus triazoles for the prophylaxis and treatment of fungal infections: a meta-analysis of RCTs. *Eur J Clin Microbiol Infect Dis* 2015; **34**: 651–9. <https://doi.org/10.1007/s10096-014-2287-4>
- 49** Tang DL, Chen X, Zhu CG *et al.* Pooled analysis of T2 *Candida* for rapid diagnosis of candidiasis. *BMC Infect Dis* 2019; **19**: 798. <https://doi.org/10.1186/s12879-019-4419-z>
- 50** Lagunes L, Rello J. Invasive candidiasis: from mycobiome to infection, therapy, and prevention. *Eur J Clin Microbiol Infect Dis* 2016; **35**: 1221–6. <https://doi.org/10.1007/s10096-016-2658-0>
- 51** Schelenz S, Barnes RA, Barton RC *et al.* British Society for Medical Mycology best practice recommendations for the diagnosis of serious fungal diseases. *Lancet Infect Dis* 2015; **15**: 461–74. [https://doi.org/10.1016/S1473-3099\(15\)70006-X](https://doi.org/10.1016/S1473-3099(15)70006-X)
- 52** Clancy CJ, Nguyen MH. Finding the ‘missing 50%’ of invasive candidiasis: how nonculture diagnostics will improve understanding of disease spectrum and transform patient care. *Clin Infect Dis* 2013; **56**: 1284–92. <https://doi.org/10.1093/cid/cit006>
- 53** Cobos-Trigueros N, Morata L, Torres J *et al.* Usefulness of time-to-positivity in aerobic and anaerobic vials to predict the presence of *Candida glabrata* in patients with candidaemia. *J Antimicrob Chemother* 2013; **68**: 2839–41. <https://doi.org/10.1093/jac/dkt285>
- 54** Mikulska M, Calandra T, Sanguinetti M *et al.* The use of mannan antigen and anti-mannan antibodies in the diagnosis of invasive candidiasis: recommendations from the Third European Conference on Infections in Leukemia. *Crit Care* 2010; **14**: R222. <https://doi.org/10.1186/cc9365>
- 55** Wei S, Wu T, Wu Y *et al.* Diagnostic accuracy of *Candida albicans* germ tube antibody for invasive candidiasis: systematic review and meta-analysis. *Diagn Microbiol Infect Dis* 2019; **93**: 339–45. <https://doi.org/10.1016/j.diagmicrobio.2018.10.017>
- 56** Camp I, Spettel K, Willinger B. Molecular methods for the diagnosis of invasive candidiasis. *J Fungi (Basel)* 2020; **6**: 101. <https://doi.org/10.3390/jof6030101>
- 57** Rouzé A, Loridant S, Poissy J *et al.* Biomarker-based strategy for early discontinuation of empirical antifungal treatment in critically ill patients: a randomized controlled trial. *Intensive Care Med* 2017; **43**: 1668–77. <https://doi.org/10.1007/s00134-017-4932-8>
- 58** Avni T, Leibovici L, Paul M. PCR diagnosis of invasive candidiasis: systematic review and meta-analysis. *J Clin Microbiol* 2011; **49**: 665–70. <https://doi.org/10.1128/JCM.01602-10>
- 59** Honore PM, Redant S, Preseau T *et al.* T2MR can be used as a non-culture-based test together with biomarkers to improve detection of *Candida* in the bloodstream and reduce time delay in treating invasive candidiasis. *Expert Rev Anti Infect Ther* 2022; **20**: 327–9. <https://doi.org/10.1080/14787210.2021.1964954>
- 60** Dupuis C, Le Bihan C, Maubon D *et al.* Performance of repeated measures of (1-3)-beta-D-glucan, mannan antigen, and antimannan antibodies for the diagnosis of invasive candidiasis in ICU patients: a preplanned ancillary analysis of the EMPIRICUS randomized clinical trial. *Open Forum Infect Dis* 2021; **8**: ofab080. <https://doi.org/10.1093/ofid/ofab080>
- 61** Pittet D, Monod M, Suter PM *et al.* *Candida* colonization and subsequent infections in critically ill surgical patients. *Ann Surg* 1994; **220**: 751–8. <https://doi.org/10.1097/00000658-199412000-00008>
- 62** Leon C, Ruiz-Santana S, Saavedra P *et al.* A bedside scoring system (‘*Candida* score’) for early antifungal treatment in nonneutropenic critically ill patients with *Candida* colonization. *Crit Care Med* 2006; **34**: 730–7. <https://doi.org/10.1097/01.CCM.0000202208.37364.7D>
- 63** Ostrosky-Zeichner L, Sable C, Sobel J *et al.* Multicenter retrospective development and validation of a clinical prediction rule for nosocomial invasive candidiasis in the intensive care setting. *Eur J Clin Microbiol Infect Dis* 2007; **26**: 271–6. <https://doi.org/10.1007/s10096-007-0270-z>
- 64** Thomas-Ruddel D, Schlattmann P, Pletz M *et al.* Risk factors for invasive candida infection in critically ill patients—a systematic review and meta-analysis. *Chest* 2022; **161**: 345–55. <https://doi.org/10.1016/j.chest.2021.08.081>
- 65** Cortegiani A, Russotto V, Raineri SM *et al.* Should we continue to use prediction tools to identify patients at risk of *Candida* spp. infection? If yes, why? *Crit Care* 2016; **20**: 351. <https://doi.org/10.1186/s13054-016-1521-0>
- 66** Knaus WA, Draper EA, Wagner DP *et al.* APACHE II: a severity of disease classification system. *Crit Care Med* 1985; **13**: 818–29. <https://doi.org/10.1097/00003246-198510000-00009>
- 67** Knaus WA, Wagner DP, Draper EA *et al.* The APACHE III prognostic system. Risk prediction of hospital mortality for critically ill hospitalized adults. *Chest* 1991; **100**: 1619–36. <https://doi.org/10.1378/chest.100.6.1619>
- 68** Zimmerman JE, Kramer AA, McNair DS *et al.* Acute Physiology and Chronic Health Evaluation (APACHE) IV: hospital mortality assessment for today’s critically ill patients. *Crit Care Med* 2006; **34**: 1297–310. <https://doi.org/10.1097/01.CCM.0000215112.84523.F0>
- 69** Moreno RP, Metnitz PG, Almeida E *et al.* SAPS 3—from evaluation of the patient to evaluation of the intensive care unit. Part 2: development

- of a prognostic model for hospital mortality at ICU admission. *Intensive Care Med* 2005; **31**: 1345–55. <https://doi.org/10.1007/s00134-005-2763-5>
- 70** Metnitz PG, Moreno RP, Almeida E *et al.* SAPS 3—from evaluation of the patient to evaluation of the intensive care unit. Part 1: objectives, methods and cohort description. *Intensive Care Med* 2005; **31**: 1336–44. <https://doi.org/10.1007/s00134-005-2762-6>
- 71** Leroy O, Bailly S, Gangneux JP *et al.* Systemic antifungal therapy for proven or suspected invasive candidiasis: the AmarCAND 2 study. *Ann Intensive Care* 2016; **6**: 2. <https://doi.org/10.1186/s13613-015-0103-7>
- 72** Posteraro B, De Pascale G, Tumbarello M *et al.* Early diagnosis of candidemia in intensive care unit patients with sepsis: a prospective comparison of (1→3)-beta-D-glucan assay, *Candida* score, and colonization index. *Crit Care* 2011; **15**: R249. <https://doi.org/10.1186/cc10507>
- 73** Wojtowicz A, Tissot F, Lamoth F *et al.* Polymorphisms in tumor necrosis factor-alpha increase susceptibility to intra-abdominal *Candida* infection in high-risk surgical ICU patients*. *Crit Care Med* 2014; **42**: e304–8. <https://doi.org/10.1097/CCM.0000000000000208>
- 74** Eggimann P, Pittet D. *Candida* colonization index and subsequent infection in critically ill surgical patients: 20 years later. *Intensive Care Med* 2014; **40**: 1429–48. <https://doi.org/10.1007/s00134-014-3355-z>
- 75** Laine ME, Flannery AH, Moody B *et al.* Need for expanded *Candida* score for empiric antifungal use in medically critically ill patients? *Crit Care* 2019; **23**: 242. <https://doi.org/10.1186/s13054-019-2525-3>
- 76** Cornely OA, Bassetti M, Calandra T *et al.* ESCMID* guideline for the diagnosis and management of *Candida* diseases 2012: non-neutropenic adult patients. *Clin Microbiol Infect* 2012; **18**(Suppl 7):19–37. <https://doi.org/10.1111/1469-0691.12039>
- 77** Mora-Duarte J, Betts R, Rotstein C *et al.* Comparison of caspofungin and amphotericin B for invasive candidiasis. *N Engl J Med* 2002; **347**: 2020–9. <https://doi.org/10.1056/NEJMoa021585>
- 78** Kuse ER, Chetchotaisak P, da Cunha CA *et al.* Micafungin versus liposomal amphotericin B for candidaemia and invasive candidosis: a phase III randomised double-blind trial. *Lancet* 2007; **369**: 1519–27. [https://doi.org/10.1016/S0140-6736\(07\)60605-9](https://doi.org/10.1016/S0140-6736(07)60605-9)
- 79** Reboli AC, Shorr AF, Rotstein C *et al.* Anidulafungin compared with fluconazole for treatment of candidemia and other forms of invasive candidiasis caused by *Candida albicans*: a multivariate analysis of factors associated with improved outcome. *BMC Infect Dis* 2011; **11**: 261. <https://doi.org/10.1186/1471-2334-11-261>
- 80** Pappas PG, Rotstein CM, Betts RF *et al.* Micafungin versus caspofungin for treatment of candidemia and other forms of invasive candidiasis. *Clin Infect Dis* 2007; **45**: 883–93. <https://doi.org/10.1086/520980>
- 81** Kullberg BJ, Viscoli C, Pappas PG *et al.* Isavuconazole versus caspofungin in the treatment of candidemia and other invasive *Candida* infections: the ACTIVE trial. *Clin Infect Dis* 2019; **68**: 1981–9. <https://doi.org/10.1093/cid/ciy827>
- 82** Thompson GR, Soriano A, Skoutelis A *et al.* Rezafungin versus caspofungin in a phase 2, randomized, double-blind study for the treatment of candidemia and invasive candidiasis: the STRIVE trial. *Clin Infect Dis* 2021; **73**: e3647–55. <https://doi.org/10.1093/cid/ciaa1380>
- 83** Andes DR, Safdar N, Baddley JW *et al.* Impact of treatment strategy on outcomes in patients with candidemia and other forms of invasive candidiasis: a patient-level quantitative review of randomized trials. *Clin Infect Dis* 2012; **54**: 1110–22. <https://doi.org/10.1093/cid/cis021>
- 84** Reboli AC, Rotstein C, Pappas PG *et al.* Anidulafungin versus fluconazole for invasive candidiasis. *N Engl J Med* 2007; **356**: 2472–82. <https://doi.org/10.1056/NEJMoa066906>
- 85** Kullberg BJ, Sobel JD, Ruhnke M *et al.* Voriconazole versus a regimen of amphotericin B followed by fluconazole for candidaemia in non-neutropenic patients: a randomised non-inferiority trial. *Lancet* 2005; **366**: 1435–42. [https://doi.org/10.1016/S0140-6736\(05\)67490-9](https://doi.org/10.1016/S0140-6736(05)67490-9)
- 86** Pfaller MA, Andes D, Arendrup MC *et al.* Clinical breakpoints for voriconazole and *Candida* spp. revisited: review of microbiologic, molecular, pharmacodynamic, and clinical data as they pertain to the development of species-specific interpretive criteria. *Diagn Microbiol Infect Dis* 2011; **70**: 330–43. <https://doi.org/10.1016/j.diagmicrobio.2011.03.002>
- 87** Safdar A, Ma J, Saliba F *et al.* Drug-induced nephrotoxicity caused by amphotericin B lipid complex and liposomal amphotericin B: a review and meta-analysis. *Medicine (Baltimore)* 2010; **89**: 236–44. <https://doi.org/10.1097/MD.0b013e3181e9441b>
- 88** Colombo AL, Guimaraes T, Sukienik T *et al.* Prognostic factors and historical trends in the epidemiology of candidemia in critically ill patients: an analysis of five multicenter studies sequentially conducted over a 9-year period. *Intensive Care Med* 2014; **40**: 1489–98. <https://doi.org/10.1007/s00134-014-3400-y>
- 89** Mellinghoff SC, Hartmann P, Cornely FB *et al.* Analyzing candidemia guideline adherence identifies opportunities for antifungal stewardship. *Eur J Clin Microbiol Infect Dis* 2018; **37**: 1563–71. <https://doi.org/10.1007/s10096-018-3285-8>
- 90** Valerio M, Muñoz P, Rodriguez CG *et al.* Antifungal stewardship in a tertiary-care institution: a bedside intervention. *Clin Microbiol Infect* 2015; **21**: 492.e1–e9. <https://doi.org/10.1016/j.cmi.2015.01.013>
- 91** Muñoz P, Bouza E, COMIC (Collaboration Group on Mycosis) Study Group. The current treatment landscape: the need for antifungal stewardship programmes. *J Antimicrob Chemother* 2016; **71**: ii5–ii12. <https://doi.org/10.1093/jac/dkw391>
- 92** Ostrosky-Zeichner L, Kullberg BJ, Bow EJ *et al.* Early treatment of candidemia in adults: a review. *Med Mycol* 2011; **49**: 113–20. <https://doi.org/10.3109/13693786.2010.512300>
- 93** Evans L, Rhodes A, Alhazzani W *et al.* Surviving sepsis campaign: international guidelines for management of sepsis and septic shock 2021. *Intensive Care Med* 2021; **47**: 1181–247. <https://doi.org/10.1007/s00134-021-06506-y>
- 94** Cortegiani A, Russotto V, Maggiore A *et al.* Antifungal agents for preventing fungal infections in non-neutropenic critically ill patients. *Cochrane Database Syst Rev* 2016; **2016**: CD004920.
- 95** Bailly S, Maubon D, Fournier P *et al.* Impact of antifungal prescription on relative distribution and susceptibility of *Candida* spp.—trends over 10 years. *J Infect* 2016; **72**: 103–11. <https://doi.org/10.1016/j.jinf.2015.09.041>
- 96** Timsit JF, Azoulay E, Schwebel C *et al.* Empirical micafungin treatment and survival without invasive fungal infection in adults with ICU-acquired sepsis, *Candida* colonization, and multiple organ failure: the EMPIRICUS randomized clinical trial. *JAMA* 2016; **316**: 1555–64. <https://doi.org/10.1001/jama.2016.14655>
- 97** Kollef M, Micek S, Hampton N *et al.* Septic shock attributed to *Candida* infection: importance of empiric therapy and source control. *Clin Infect Dis* 2012; **54**: 1739–46. <https://doi.org/10.1093/cid/cis305>
- 98** Vergidis P, Clancy CJ, Shields RK *et al.* Intra-abdominal candidiasis: the importance of early source control and antifungal treatment. *PLoS ONE* 2016; **11**: e0153247. <https://doi.org/10.1371/journal.pone.0153247>
- 99** Desai JV, Mitchell AP, Andes DR. Fungal biofilms, drug resistance, and recurrent infection. *Cold Spring Harb Perspect Med* 2014; **4**: a019729. <https://doi.org/10.1101/cshperspect.a019729>
- 100** Guinea J, Arendrup MC, Canton R *et al.* Genotyping reveals high clonal diversity and widespread genotypes of *Candida* causing candidemia at distant geographical areas. *Front Cell Infect Microbiol* 2020; **10**: 166. <https://doi.org/10.3389/fcimb.2020.00166>

- 101** Ramage G, Rajendran R, Sherry L *et al.* Fungal biofilm resistance. *Int J Microbiol* 2012; **2012**: 528521. <https://doi.org/10.1155/2012/528521>
- 102** Davey ME, O'Toole GA. Microbial biofilms: from ecology to molecular genetics. *Microbiol Mol Biol Rev* 2000; **64**: 847–67. <https://doi.org/10.1128/MMBR.64.4.847-867.2000>
- 103** Toulet D, Debarre C, Imbert C. Could liposomal amphotericin B (L-AMB) lock solutions be useful to inhibit *Candida* spp. biofilms on silicone biomaterials? *J Antimicrob Chemother* 2012; **67**: 430–2. <https://doi.org/10.1093/jac/dkr473>
- 104** Cavalheiro M, Teixeira MC. *Candida* biofilms: threats, challenges, and promising strategies. *Front Med (Lausanne)* 2018; **5**: 28. <https://doi.org/10.3389/fmed.2018.00028>
- 105** Cernakova L, Light C, Salehi B *et al.* Novel therapies for biofilm-based *Candida* spp. Infections. *Adv Exp Med Biol* 2019; **1214**: 93–123. https://doi.org/10.1007/5584_2019_400
- 106** Gill CM, Kenney RM, Hencken L *et al.* T2 *Candida* versus beta-D-glucan to facilitate antifungal discontinuation in the intensive care unit. *Diagn Microbiol Infect Dis* 2019; **95**: 162–5. <https://doi.org/10.1016/j.diagmicrobio.2019.04.016>
- 107** Moreno-García E, Puerta-Alcalde P, Gariup G *et al.* Early stepdown from echinocandin to fluconazole treatment in candidemia: a *post hoc* analysis of three cohort studies. *Open Forum Infect Dis* 2021; **8**: ofab250. <https://doi.org/10.1093/ofid/ofab250>
- 108** Mylonakis E, Zacharioudakis IM, Clancy CJ *et al.* Efficacy of T2 magnetic resonance assay in monitoring candidemia after initiation of antifungal therapy: the serial therapeutic and antifungal monitoring protocol (STAMP) trial. *J Clin Microbiol* 2018; **56**: e01756–17. <https://doi.org/10.1128/JCM.01756-17>
- 109** Gómez-López A. Antifungal therapeutic drug monitoring: focus on drugs without a clear recommendation. *Clin Microbiol Infect* 2020; **26**: 1481–7. <https://doi.org/10.1016/j.cmi.2020.05.037>
- 110** Van Daele R, Wauters J, Lagrou K *et al.* Pharmacokinetic variability and target attainment of fluconazole in critically ill patients. *Microorganisms* 2021; **9**: 2068. <https://doi.org/10.3390/microorganisms9102068>
- 111** Lewis RE, Andes DR. Managing uncertainty in antifungal dosing: antibiograms, therapeutic drug monitoring and drug-drug interactions. *Curr Opin Infect Dis* 2021; **34**: 288–96. <https://doi.org/10.1097/QCO.0000000000000740>
- 112** Pfaller MA, Diekema DJ, Turnidge JD *et al.* Twenty years of the SENTRY antifungal surveillance program: results for *Candida* Species from 1997–2016. *Open Forum Infect Dis* 2019; **6**: S79–94. <https://doi.org/10.1093/ofid/ofy358>
- 113** Arastehfar A, Gabaldon T, Garcia-Rubio R *et al.* Drug-resistant fungi: an emerging challenge threatening our limited antifungal armamentarium. *Antibiotics (Basel)* 2020; **9**: 877. <https://doi.org/10.3390/antibiotics9120877>
- 114** Hedley KR, Perlin DS. Fungal resistance to echinocandins and the MDR phenomenon in *Candida glabrata*. *J Fungi (Basel)* 2018; **4**: 105. <https://doi.org/10.3390/jof4030105>
- 115** Chow NA, Munoz JF, Gade L *et al.* Tracing the evolutionary history and global expansion of *Candida auris* using population genomic analyses. *mBio* 2020; **11**: e03364–19. <https://doi.org/10.1128/mBio.03364-19>
- 116** Pristov KE, Ghannoum MA. Resistance of *Candida* to azoles and echinocandins worldwide. *Clin Microbiol Infect* 2019; **25**: 792–8. <https://doi.org/10.1016/j.cmi.2019.03.028>
- 117** Lockhart SR, Etienne KA, Vallabhaneni S *et al.* Simultaneous emergence of multidrug-resistant *Candida auris* on 3 continents confirmed by whole-genome sequencing and epidemiological analyses. *Clin Infect Dis* 2017; **64**: 134–40. <https://doi.org/10.1093/cid/ciw691>
- 118** Chowdhary A, Sharma C, Meis JF. *Candida auris*: a rapidly emerging cause of hospital-acquired multidrug-resistant fungal infections globally. *PLoS Pathog* 2017; **13**: e1006290. <https://doi.org/10.1371/journal.ppat.1006290>
- 119** Clancy CJ, Nguyen MH. Emergence of *Candida auris*: an international call to arms. *Clin Infect Dis* 2017; **64**: 141–3. <https://doi.org/10.1093/cid/ciw696>
- 120** CLSI. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts—Fourth Edition: M27*. 2017.
- 121** Arendrup MC, Meletiadis J, Mouton JW *et al.* *EUCAST definitive document E.DEF 7.3.2. Method for the determination of broth dilution minimum inhibitory concentrations of antifungal agents for yeasts*, 2020. https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/AFST/Files/EUCAST_E_Def_7.3.2_Yeast_testing_definitive_revised_2020.pdf
- 122** Knabl L, Lass-Flörl C. Antifungal susceptibility testing in *Candida* species: current methods and promising new tools for shortening the turnaround time. *Expert Rev Anti Infect Ther* 2020; **18**: 779–87. <https://doi.org/10.1080/14787210.2020.1760841>
- 123** Kwon YJ, Won EJ, Jeong SH *et al.* Dynamics and predictors of mortality due to candidemia caused by different *Candida* species: comparison of Intensive Care Unit-Associated Candidemia (ICUAC) and non-ICUAC. *J Fungi (Basel)* 2021; **7**: 597. <https://doi.org/10.3390/jof7080597>
- 124** Pea F. Plasma pharmacokinetics of antimicrobial agents in critically ill patients. *Curr Clin Pharmacol* 2013; **8**: 5–12.
- 125** Bouman CS, van Kan HJ, Koopmans RP *et al.* Discrepancies between observed and predicted continuous venovenous hemofiltration removal of antimicrobial agents in critically ill patients and the effects on dosing. *Intensive Care Med* 2006; **32**: 2013–9. <https://doi.org/10.1007/s00134-006-0397-x>
- 126** Yagasaki K, Gando S, Matsuda N *et al.* Pharmacokinetics and the most suitable dosing regimen of fluconazole in critically ill patients receiving continuous hemodiafiltration. *Intensive Care Med* 2003; **29**: 1844–8. <https://doi.org/10.1007/s00134-003-1980-z>
- 127** Riche CVW, Cassol R, Pasqualotto AC. Is the frequency of candidemia increasing in COVID-19 patients receiving corticosteroids. *J Fungi (Basel)* 2020; **6**: 286. <https://doi.org/10.3390/jof6040286>
- 128** Arastehfar A, Carvalho A, Nguyen MH *et al.* COVID-19-associated candidiasis (CAC): an underestimated complication in the absence of immunological predispositions? *J Fungi (Basel)* 2020; **6**: 211. <https://doi.org/10.3390/jof6040211>
- 129** Leon C, Ruiz-Santana S, Saavedra P *et al.* Value of beta-D-glucan and *Candida albicans* germ tube antibody for discriminating between *Candida* colonization and invasive candidiasis in patients with severe abdominal conditions. *Intensive Care Med* 2012; **38**: 1315–25. <https://doi.org/10.1007/s00134-012-2616-y>
- 130** Delaloye J, Calandra T. Invasive candidiasis as a cause of sepsis in the critically ill patient. *Virulence* 2014; **5**: 161–9. <https://doi.org/10.4161/viru.26187>
- 131** Poissy J, Damonti L, Bignon A *et al.* Risk factors for candidemia: a prospective matched case-control study. *Crit Care* 2020; **24**: 109. <https://doi.org/10.1186/s13054-020-2766-1>
- 132** Zilberberg M, Yu HT, Chaudhari P *et al.* Relationship of fluconazole prophylaxis with fungal microbiology in hospitalized intra-abdominal surgery patients: a descriptive cohort study. *Crit Care* 2014; **18**: 590. <https://doi.org/10.1186/s13054-014-0590-1>
- 133** Lortholary O, Renaudat C, Sitbon K *et al.* The risk and clinical outcome of candidemia depending on underlying malignancy. *Intensive Care Med* 2017; **43**: 652–62. <https://doi.org/10.1007/s00134-017-4743-y>
- 134** Maertens JA, Girmenia C, Brüggemann RJ *et al.* European Guidelines for primary antifungal prophylaxis in adult haematology patients: summary of the updated recommendations from the European Conference on Infections in Leukaemia. *J Antimicrob Chemother* 2018; **73**: 3221–30.

- 135** Freifeld AG, Bow EJ, Sepkowitz KA *et al.* Clinical practice guideline for the use of antimicrobial agents in neutropenic patients with cancer: 2010 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2011; **52**: 427–31. <https://doi.org/10.1093/cid/ciq147>
- 136** Kontoyiannis DP, Reddy BT, Hanna H *et al.* Breakthrough candidemia in patients with cancer differs from *de novo* candidemia in host factors and *Candida* species but not intensity. *Infect Control Hosp Epidemiol* 2002; **23**: 542–5. <https://doi.org/10.1086/502104>
- 137** Girmenia C, Martino P, Cassone A. Breakthrough candidemia during antifungal treatment with fluconazole in patients with hematologic malignancies. *Blood* 1996; **87**: 838–9. <https://doi.org/10.1182/blood.V87.2.838.bloodjournal872838>
- 138** Uzun O, Ascioglu S, Anaissie EJ *et al.* Risk factors and predictors of outcome in patients with cancer and breakthrough candidemia. *Clin Infect Dis* 2001; **32**: 1713–7. <https://doi.org/10.1086/320757>
- 139** Pfeiffer CD, Garcia-Effron G, Zaas AK *et al.* Breakthrough invasive candidiasis in patients on micafungin. *J Clin Microbiol* 2010; **48**: 2373–80. <https://doi.org/10.1128/JCM.02390-09>
- 140** Bizerra FC, Jimenez-Ortigosa C, Souza AC *et al.* Breakthrough candidemia due to multidrug-resistant *Candida glabrata* during prophylaxis with a low dose of micafungin. *Antimicrob Agents Chemother* 2014; **58**: 2438–40. <https://doi.org/10.1128/AAC.02189-13>
- 141** Breda GL, Tuon FF, Meis JF *et al.* Breakthrough candidemia after the introduction of broad spectrum antifungal agents: a 5-year retrospective study. *Med Mycol* 2018; **56**: 406–15. <https://doi.org/10.1093/mmy/myx077>
- 142** Cuervo G, Garcia-Vidal C, Nucci M *et al.* Breakthrough candidaemia in the era of broad-spectrum antifungal therapies. *Clin Microbiol Infect* 2016; **22**: 181–8. <https://doi.org/10.1016/j.cmi.2015.09.029>
- 143** Nucci M, Colombo AL. Risk factors for breakthrough candidemia. *Eur J Clin Microbiol Infect Dis* 2002; **21**: 209–11. <https://doi.org/10.1007/s10096-002-0697-1>
- 144** Orasch C, Mertz D, Garbino J *et al.* Fluconazole non-susceptible breakthrough candidemia after prolonged low-dose prophylaxis: a prospective FUNGINOS study. *J Infect* 2018; **76**: 489–95. <https://doi.org/10.1016/j.jinf.2017.12.018>
- 145** Sanchez-Portocarrero J, Perez-Cecilia E, Corral O *et al.* The central nervous system and infection by *Candida* species. *Diagn Microbiol Infect Dis* 2000; **37**: 169–79. [https://doi.org/10.1016/S0732-8893\(00\)00140-1](https://doi.org/10.1016/S0732-8893(00)00140-1)
- 146** Nguyen MH, Yu VL. Meningitis caused by *Candida* species: an emerging problem in neurosurgical patients. *Clin Infect Dis* 1995; **21**: 323–7. <https://doi.org/10.1093/clinids/21.2.323>
- 147** Casado JL, Quereda C, Oliva J *et al.* Candidal meningitis in HIV-infected patients: analysis of 14 cases. *Clin Infect Dis* 1997; **25**: 673–6. <https://doi.org/10.1086/513746>
- 148** Fennelly AM, Slenker AK, Murphy LC *et al.* *Candida* cerebral abscesses: a case report and review of the literature. *Med Mycol* 2013; **51**: 779–84. <https://doi.org/10.3109/13693786.2013.789566>
- 149** Lefort A, Chartier L, Sendid B *et al.* Diagnosis, management and outcome of *Candida endocarditis*. *Clin Microbiol Infect* 2012; **18**: E99–E109. <https://doi.org/10.1111/j.1469-0691.2012.03764.x>
- 150** Drummond RA, Collar AL, Swamydas M *et al.* CARD9-dependent neutrophil recruitment protects against fungal invasion of the central nervous system. *PLoS Pathog* 2015; **11**: e1005293. <https://doi.org/10.1371/journal.ppat.1005293>
- 151** Baddley JW, Benjamin DK Jr, Patel M *et al.* *Candida* infective endocarditis. *Eur J Clin Microbiol Infect Dis* 2008; **27**: 519–29. <https://doi.org/10.1007/s10096-008-0466-x>
- 152** Rivoisy C, Vena A, Schaeffer L *et al.* Prosthetic valve *Candida* spp. endocarditis: new insights into long-term prognosis—the ESCAPE study. *Clin Infect Dis* 2018; **66**: 825–32. <https://doi.org/10.1093/cid/cix913>
- 153** Schwartz S, Kontoyiannis DP, Harrison T *et al.* Advances in the diagnosis and treatment of fungal infections of the CNS. *Lancet Neurol* 2018; **17**: 362–72. [https://doi.org/10.1016/S1474-4422\(18\)30030-9](https://doi.org/10.1016/S1474-4422(18)30030-9)
- 154** Chaussade H, Cazals X, Desoubeaux G *et al.* Central nervous system candidiasis beyond neonates: lessons from a nationwide study. *Med Mycol* 2021; **59**: 266–77. <https://doi.org/10.1093/mmy/myaa051>
- 155** Chen TL, Chen HP, Fung CP *et al.* Clinical characteristics, treatment and prognostic factors of candidal meningitis in a teaching hospital in Taiwan. *Scand J Infect Dis* 2004; **36**: 124–30. <https://doi.org/10.1080/00365540310017573>
- 156** Smego RA J, Perfect JR, Durack DT. Combined therapy with amphotericin B and 5-fluorocytosine for *Candida* meningitis. *Rev Infect Dis* 1984; **6**: 791–801. <https://doi.org/10.1093/clinids/6.6.791>
- 157** Weiler S, Fiegl D, MacFarland R *et al.* Human tissue distribution of voriconazole. *Antimicrob Agents Chemother* 2011; **55**: 925–8. <https://doi.org/10.1128/AAC.00949-10>
- 158** Peman J, Ruiz-Gaitan A. Candidemia from urinary tract source: the challenge of candiduria. *Hosp Pract (1995)* 2018; **46**: 243–5. <https://doi.org/10.1080/21548331.2018.1538623>
- 159** Cuervo G, Garcia-Vidal C, Puig-Asensio M *et al.* Echinocandins compared to fluconazole for candidemia of a urinary tract source: a propensity score analysis. *Clin Infect Dis* 2017; **64**: 1374–9. <https://doi.org/10.1093/cid/cix033>
- 160** Sobel JD, Kauffman CA, McKinsey D *et al.* Candiduria: a randomized, double-blind study of treatment with fluconazole and placebo. The National Institute of Allergy and Infectious Diseases (NIAID) mycoses study group. *Clin Infect Dis* 2000; **30**: 19–24. <https://doi.org/10.1086/313580>
- 161** Grau S, Luque S, Echeverria-Esnal D *et al.* Urinary micafungin levels are sufficient to treat urinary tract infections caused by *Candida* spp. *Int J Antimicrob Agents* 2016; **48**: 212–4. <https://doi.org/10.1016/j.ijantimicag.2016.05.010>
- 162** Chhablani J. Fungal endophthalmitis. *Expert Rev Anti Infect Ther* 2011; **9**: 1191–201. <https://doi.org/10.1586/eri.11.139>
- 163** Savani DV, Perfect JR, Cobo LM *et al.* Penetration of new azole compounds into the eye and efficacy in experimental *Candida* endophthalmitis. *Antimicrob Agents Chemother* 1987; **31**: 6–10. <https://doi.org/10.1128/AAC.31.1.6>
- 164** Barza M. Treatment options for candidal endophthalmitis [editorial comment]. *Clin Infect Dis* 1998; **27**: 1134–6. <https://doi.org/10.1086/514973>
- 165** O'Day DM, Foulds G, Williams TE *et al.* Ocular uptake of fluconazole following oral administration. *Arch Ophthalmol* 1990; **108**: 1006–8. <https://doi.org/10.1001/archophth.1990.01070090108050>
- 166** Lashof AMO, Rothova A, Sobel JD *et al.* Ocular manifestations of candidemia. *Clin Infect Dis* 2011; **53**: 262–8. <https://doi.org/10.1093/cid/cir355>
- 167** Ellis ME, Al-Abdely H, Sandridge A *et al.* Fungal endocarditis: evidence in the world literature, 1965–1995. *Clin Infect Dis* 2001; **32**: 50–62. <https://doi.org/10.1086/317550>
- 168** Boland JM, Chung HH, Robberts FJ *et al.* Fungal prosthetic valve endocarditis: Mayo Clinic experience with a clinicopathological analysis. *Mycoses* 2011; **54**: 354–60. <https://doi.org/10.1111/j.1439-0507.2010.01884.x>
- 169** Meena DS, Kumar D, Agarwal M *et al.* Clinical features, diagnosis and treatment outcome of fungal endocarditis: a systematic review of reported cases. *Mycoses* 2022; **65**:294–302. <https://doi.org/10.1111/myc.13398>
- 170** Giuliano S, Guastalegname M, Russo A *et al.* *Candida* endocarditis: systematic literature review from 1997 to 2014 and analysis of 29 cases

- from the Italian study of endocarditis. *Expert Rev Anti Infect Ther* 2017; **15**: 807–18. <https://doi.org/10.1080/14787210.2017.1372749>
- 171** Foong KS, Sung A, Burnham JP *et al.* Risk factors predicting *Candida* infective endocarditis in patients with candidemia. *Med Mycol* 2020; **58**: 593–9. <https://doi.org/10.1093/mmy/myz104>
- 172** Gamaletsou MN, Kontoyiannis DP, Sipsas NV *et al.* *Candida* osteomyelitis: analysis of 207 pediatric and adult cases (1970–2011). *Clin Infect Dis* 2012; **55**: 1338–51. <https://doi.org/10.1093/cid/cis660>
- 173** Neofytos D, Huprikar S, Reboli A *et al.* Treatment and outcomes of *Candida* osteomyelitis: review of 53 cases from the PATH alliance(R) registry. *Eur J Clin Microbiol Infect Dis* 2014; **33**: 135–41. <https://doi.org/10.1007/s10096-013-1939-0>
- 174** Slenker AK, Keith SW, Horn DL. Two hundred and eleven cases of *Candida* osteomyelitis: 17 case reports and a review of the literature. *Diagn Microbiol Infect Dis* 2012; **73**: 89–93. <https://doi.org/10.1016/j.diagmicrobio.2012.02.004>
- 175** Saconi ES, de Carvalho VC, de Oliveira PRD *et al.* Prosthetic joint infection due to *Candida* species: case series and review of literature. *Medicine (Baltimore)* 2020; **99**: e19735. <https://doi.org/10.1097/MD.00000000000019735>
- 176** Lee YR, Kim HJ, Lee EJ *et al.* Prosthetic joint infections caused by *Candida* species: a systematic review and a case series. *Mycopathologia* 2019; **184**: 23–33. <https://doi.org/10.1007/s11046-018-0286-1>
- 177** Meena DS, Kumar D. *Candida* pneumonia: an innocent bystander or a silent killer? *Med Princ Pract* 2022; **31**:98–102. <https://doi.org/10.1159/000520111>
- 178** Neoh CF, Slavin M, Chen SC *et al.* Echinocandins in the treatment of candidaemia and invasive candidiasis: clinical and economic perspectives. *Int J Antimicrob Agents* 2014; **43**: 207–14. <https://doi.org/10.1016/j.ijantimicag.2013.08.010>
- 179** Ou HT, Lee TY, Chen YC *et al.* Pharmacoeconomic analysis of antifungal therapy for primary treatment of invasive candidiasis caused by *Candida albicans* and non-*albicans Candida* species. *BMC Infect Dis* 2017; **17**: 481. <https://doi.org/10.1186/s12879-017-2573-8>
- 180** Heimann SM, Cornely OA, Wisplinghoff H *et al.* Candidemia in the intensive care unit: analysis of direct treatment costs and clinical outcome in patients treated with echinocandins or fluconazole. *Eur J Clin Microbiol Infect Dis* 2015; **34**: 331–8. <https://doi.org/10.1007/s10096-014-2230-8>
- 181** Thompson GR, Soriano A, Cornely OA *et al.* Rezafungin versus caspofungin for treatment of candidaemia and invasive candidiasis (ReSTORE): a multicentre, double-blind, double-dummy, randomised phase 3 trial. *Lancet* 2023; **401**:49–59. [https://doi.org/10.1016/S0140-6736\(22\)02324-8](https://doi.org/10.1016/S0140-6736(22)02324-8)
- 182** Spec A, Pullman J, Thompson GR *et al.* MSG-10: a phase 2 study of oral ibrexafungerp (SCY-078) following initial echinocandin therapy in non-neutropenic patients with invasive candidiasis. *J Antimicrob Chemother* 2019; **74**: 3056–62. <https://doi.org/10.1093/jac/dkz277>
- 183** Ghannoum M, Arendrup MC, Chaturvedi VP *et al.* Ibrexafungerp: a novel oral triterpenoid antifungal in development for the treatment of *Candida auris* infections. *Antibiotics (Basel)* 2020; **9**: 539. <https://doi.org/10.3390/antibiotics9090539>
- 184** Lee A, Wang N, Carter CL *et al.* Therapeutic potential of fosmanogepix (APX001) for intra-abdominal candidiasis: from lesion penetration to efficacy in a mouse model. *Antimicrob Agents Chemother* 2021; **65**: e02476-20. <https://doi.org/10.1128/AAC.02476-20>
- 185** Arendrup MC, Chowdhary A, Jorgensen KM *et al.* Manogepix (APX001A) *in vitro* activity against *Candida auris*: head-to-head comparison of EUCAST and CLSI MICs. *Antimicrob Agents Chemother* 2020; **64**: e00656-20. <https://doi.org/10.1128/AAC.00656-20>
- 186** Wiederhold NP, Najvar LK, Jaramillo R *et al.* The novel arylamidine T-2307 demonstrates *in vitro* and *in vivo* activity against *Candida auris*. *Antimicrob Agents Chemother* 2020; **64**: e02198-19. <https://doi.org/10.1128/AAC.02198-19>
- 187** Fioriti S, Brescini L, Pallotta F *et al.* Antifungal combinations against *Candida* species: from bench to bedside. *J Fungi (Basel)* 2022; **8**:1077. <https://doi.org/10.3390/jof8101077>