

Review

Novel strategy for oncogenic alteration-induced lipid metabolism reprogramming in pancreatic cancer

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Abstract

The pathogenesis of pancreatic cancer involves substantial metabolic reprogramming, resulting in abnormal proliferation of tumor cells. This tumorigenic reprogramming is often driven by genetic mutations, such as activating mutations of the *KRAS* oncogene and inactivating or deletions of the tumor suppressor genes *SMAD4*, *CDKN2A*, and *TP53*, which play a critical role in the initiation and development of pancreatic cancer. As a normal cell gradually develops into a cancer cell, a series of signature characteristics are acquired: activation of signaling pathways that sustain proliferation; an ability to resist growth inhibitory signals and evade apoptosis; and an ability to generate new blood vessels and invade and metastasize. In addition to these features, recent research has revealed that metabolic reprogramming and immune escape are two other novel characteristics of tumor cells. The effect of the interactions between tumor and immune cells on metabolic reprogramming is a key factor determining the antitumor immunotherapy response. Lipid metabolism reprogramming, a feature of many malignancies, not only plays a role in maintaining tumor cell proliferation but also alters the tumor microenvironment by inducing the release of metabolites that in turn affect the metabolism of normal immune cells, ultimately leading to the attenuation of the antitumor immune response and resistance to immunotherapy. Pancreatic cancer has been found to have substantial lipid metabolism reprogramming, but the mechanisms remain elusive. Therefore, this review focuses on the mechanisms regulating lipid metabolism reprogramming in pancreatic cancer cells to provide new therapeutic targets and aid the development of new therapeutic strategies for pancreatic cancer.

Key words pancreatic cancer, lipid metabolism reprogramming, tumor microenvironment, immune escape, drug resistance

Introduction

Pancreatic cancer is a malignant tumor of the digestive tract with high mortality. The most common type of pancreatic cancer is pancreatic ductal adenocarcinoma (PDAC), accounting for 90% of cases [1]. Because the early symptoms of pancreatic cancer are not obvious and it has a rapid onset, the 5-year survival rate of patients with pancreatic cancer is only 11%, making it the third deadliest cancer in the United States; pancreatic cancer is predicted to become the second deadliest cancer within 5 years [2,3]. Currently, aggressive treatment, including surgical resection (usually the first

choice), neoadjuvant chemotherapy, and radiotherapy, is the main treatment used in the clinic. However, only 20% of patients are eligible for surgical resection at diagnosis, and even in those who are eligible for and undergo surgery, 80% of patients eventually relapse, and relapsed PDAC is almost always fatal [4,5]. Chemotherapy is the recommended primary treatment for patients who cannot undergo surgery, but its efficacy is suboptimal due to the rapid emergence of drug resistance and severe side effects [6,7]. Hence, it is essential to explore the molecular mechanisms of pancreatic cancer evolution and develop new strategies for

pancreatic cancer diagnosis and treatment.

The most common hallmark of pancreatic cancer is mutation of an oncogene. Molecular analyses have revealed four major PDAC driver genes that are commonly mutated: *KRAS* (~85%), *TP53* (60%–70%), *CDKN2A* (>50%), and *SMAD4* (~50%) [8]. Genetic analysis of clinical specimens revealed that mutations in the *KRAS* oncogene are found in more than 90% of pancreatic cancers and are an early event of stage 1 pancreatic intraepithelial neoplasia (PanIN). Inactivation and deletion of the tumor suppressor genes *SMAD4*, *CDKN2A*, and *TP53* are associated with PanIN progression and invasiveness [9,10]. Somatic gene mutations increase the malignant potential of tumor cells and activate multiple signaling pathways, such as the NOTCH, Hedgehog (Hh), β -catenin, chromosomal reorganization and DNA repair pathways, promoting the proliferation of tumor cells (Figure 1). This suggests that the *KRAS* signaling pathway, a key driver of PDAC initiation, is a prime target for the development of inhibitors; however, it is difficult to develop therapeutic *KRAS* inhibitors [11,12]. Small molecule inhibitors of the *KRAS* G12C mutation sotorasib and adagrasib are under clinical investigation [13,14]. In addition, novel drugs need to target pathways that indirectly affect *KRAS* to treat pancreatic cancer.

The occurrence and development of tumors are complex biological processes with multiple mechanisms and factors. In recent years, reprogramming of energy metabolism has attracted increasing attention in cancer research. It may be an important factor for tumor cell proliferation and is beginning to be recognized as a hallmark of cancer [15,16]. Tumor metabolic reprogramming is not only limited to the Warburg effect, which is related to glycolysis and the tricarboxylic acid cycle but also involves more complex metabolic processes, such as fatty acid and glutamate metabolism [17]. Previous studies have confirmed that abnormal lipid metabolism is one of the hallmarks of malignant tumors and an important potential target for tumor therapy [18]. Energy storage, organelle and membrane maintenance, and the generation of signaling molecules in tumor cells all require fatty acid metabolism to generate energy [18]. Normal cells mainly use fatty acids derived from food, while 90% of the fatty acids used by tumor cells are

derived from *de novo* synthesis, and fatty acid oxidation is enhanced to meet the needs of tumor cells in an unfavorable living environment [19]. Recent studies have shown that lipid and cholesterol accumulation increase drug resistance and promote epithelial-mesenchymal transition (EMT) in basal-type PDAC, resulting in poor outcomes of patients [20,21].

The success of pancreatic cancer immunotherapy shows the important role of the immune system in preventing the progression of tumors, and immunotherapy has become an important treatment method in addition to traditional therapy; however, an immunosuppressive tumor microenvironment is still an obstacle limiting the efficacy of tumor immunotherapy [22,23]. Reprogramming of tumor metabolism affects both immune cells and tumor growth by inducing the release of metabolites such as lactate and PGE. These effects cause metabolic competition and generate an acidic and hypoxic tumor immune microenvironment with high levels of reactive oxygen species, ultimately weakening antitumor immune responses and enabling immune escape [24–26]. Therefore, understanding how metabolic reprogramming modulates antitumor immune responses will help reveal new ideas for targeting metabolic pathways for antitumor immunotherapy.

Gene Mutations Related to Pancreatic Cancer Development and Maintenance

The occurrence and development of pancreatic cancer are generally attributed to the accumulation of genetic alterations, which leads to the activation of oncogenes and the inactivation of tumor suppressor genes. PDAC caused by malignant transformation of the ductal epithelium accounts for approximately 90% of pancreatic cancer cases. The frequency of *KRAS* mutations in PDAC is up to 85%, and *KRAS* is considered a major oncogene that regulates cell proliferation and survival pathways. The high incidence of *KRAS* mutations suggests that therapies targeting the *KRAS* signaling network may be an effective treatment for PDAC. In this section, we summarize the role of *KRAS* in pancreatic cancer, treatment options developed to target *KRAS* mutations, and related challenges. We also discuss other common genetic mutations in PDAC and their effects.

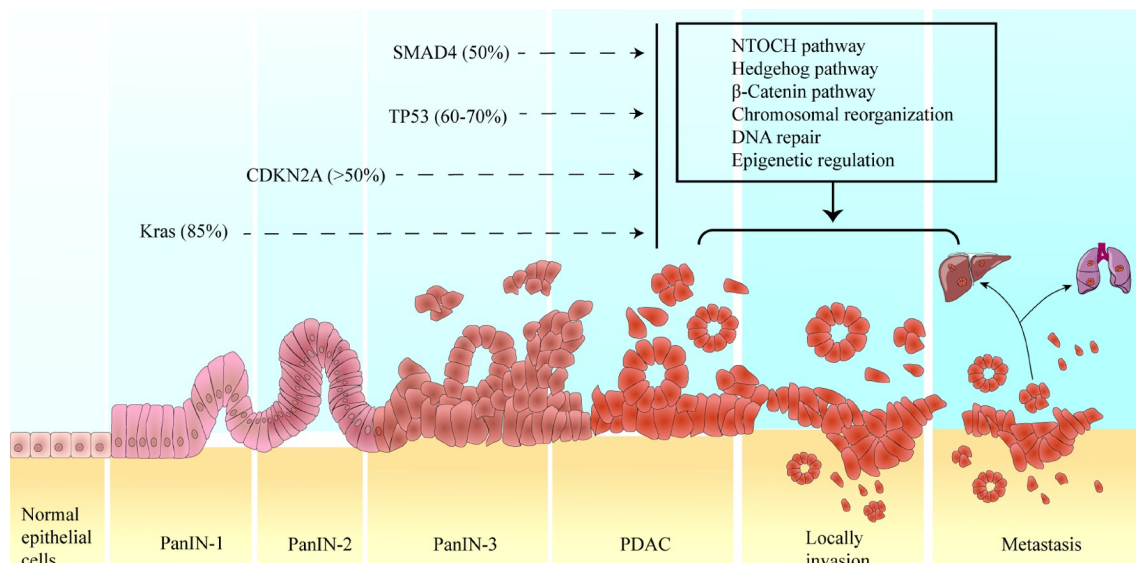


Figure 1. Oncogenic mutations and regulatory mechanisms in the development of pancreatic cancer

KRAS mutations in pancreatic cancer

KRAS is a Kirsten rat sarcoma viral oncogene homolog belonging to the mammalian *RAS* gene family. It encodes the *KRAS* protein, a member of the small GTPase superfamily, which is activated upon binding to GTP and is inactivated upon binding to GDP [27]. The activated GTP-bound state and inactivated GDP-bound state of *KRAS* are strictly regulated by guanine nucleotide exchange factors (GEFs) and GTPase-activating proteins (GAPs) [28,29]. The former catalyzes the formation of the active GTP-bound state, while the latter induces GTP hydrolysis to terminate signaling. In quiescent cells, *KRAS* is mainly present in the inactive GDP-binding state. When cellular transmembrane receptors, such as epidermal growth factor receptor (EGFR), are activated by stimulation signals, GDP is converted into GTP. Once the *KRAS* protein binds to GTP, the signaling switch is triggered, and *KRAS* can interact with numerous downstream effector proteins to activate various intracellular signaling pathways related to cell proliferation, migration, transformation, and survival. The ability of mutant *KRAS* to hydrolyse GTP is impaired, leading to an abnormal hyperactive state. In PDAC, point mutations in codon 12, in which glycine is substituted with another amino acid, are the most common *KRAS* mutations: glycine to aspartic acid mutation (G12D, 45%), glycine to valine mutation (G12V, 35%), and glycine to arginine mutation (G12R, 17%) [30–33]. Uncommon mutations, such as G13 and K117 mutations, also occur. These missense mutations cause *KRAS* to be in an abnormally active state, with consequent activation of some key downstream effector pathways, including the RAF-MEK-ERK pathway and the PI3K-AKT-mTOR pathway [34]. Many recent studies have shown that mutant *KRAS* signaling contributes to the development and maintenance of pancreatic cancer and participates in regulating the survival and metastasis of pancreatic cancer cells, tumor microenvironment remodelling, and the occurrence of metabolic disorders.

Role of *KRAS* in the development of pancreatic cancer

KRAS mutations have been identified early in PDAC development. For example, genetic analysis of clinical specimens revealed that *KRAS* mutations were already present in precancerous lesions, such as PanINs and intraductal papillary mucinous neoplasms (IPMNs), suggesting a role for those mutations in the initiation of pancreatic cancer [35]. Furthermore, additional mutations in other genes, such as inactivating mutations and deletions of the tumor suppressor genes *SMAD4*, *CDKN2A*, and *TP53*, are also required for tumor progression [36]. On one hand, *KRAS* mutations have been shown to alter many biological processes in pancreatic cancer cells, resulting in, for example, increased cell proliferation, survival, migration, and invasion and contributing to cell chemoresistance and inflammation. Furthermore, the MAPK signaling pathway was proven to be required for the formation of PanINs in mice because it promotes the dedifferentiation of acinar cells into duct-like cells, which are easily transformed [37]. In a high-fat diet-fed mouse model, oncogenic *KRAS* caused decreased expression of fibroblast growth factor 21, a metabolic regulator that prevents obesity, in acinar cells, and the mice developed extensive inflammation, pancreatic cysts, PanINs and PDAC [38]. The ERK1-MAPK pathway is associated with the resistance of pancreatic cancer cells to chemotherapeutic drugs, such as gemcitabine [39]. The nuclear factor- κ B (NF- κ B) pathway is related to the invasion and metastasis of pancreatic cancer cells and promotes tumor initiation, tumor cell

self-renewal and erlotinib resistance [40]. On the other hand, *KRAS* mutations have been found to alter aspects of the pancreatic cancer microenvironment, such as immune cell infiltration and extracellular matrix structure. Cancer-associated fibroblasts (CAFs), various immune cells, extracellular matrix components, blood vessels and lymph-vessel networks constitute the stromal component of pancreatic tumors and play a dual role in tumor development and metastasis. In general, the mutual crosstalk between tumor cells and CAFs promotes the development of pancreatic cancer, and the Hedgehog signaling pathway is a key regulator. *KRAS*-mutant pancreatic cancer cells secrete Hedgehog ligands that act on Hedgehog receptor-expressing CAFs, and CAFs in turn promote the growth of pancreatic cancer cells by regulating the extracellular matrix, hyaluronic acid, collagen fibres and other factors [41,42]. Furthermore, *KRAS* mutations are also associated with immunosuppressive cell infiltration in the pancreatic cancer microenvironment. The *KRAS* G12D mutation induces the conversion of CD4⁺CD25⁻ T cells into regulatory T cells (Tregs) by upregulating the expressions of IL-10 and transforming growth factor- β [43,44]. This effect is achieved via the activation of the MEK/ERK pathway. Upon the induction of oxidative stress, the *KRAS* G12D protein in cancer cells can also be released from cancer cells into the surrounding microenvironment and then taken up by macrophages. Through STAT3-dependent fatty acid oxidation, *KRAS* G12D causes the transformation of macrophages into an M2-like protumor phenotype [45]. Pancreatic cancer cells bearing oncogenic activated *KRAS* release IL-4, IL-6, IL-13, MCP-1, and CSF-1, which also promotes the recruitment, aggregation, and accumulation of tumor-associated macrophages (TAMs) at tumor sites and induces their polarization into an immunosuppressive phenotype [46]. A high-fat diet combined with *KRAS* mutation can also induce CCL2 secretion from pancreatic epithelial cells and promote the recruitment of myeloid-derived inhibitory cells and protumor macrophages in a peroxisome proliferator-activated receptor-delta-dependent manner [47]. Through the mechanisms described above, *KRAS* mutations significantly increase the infiltration of protumor cells in pancreatic cancer, favoring immune escape and tumor progression. Moreover, *KRAS* mutations also contribute to metabolic disorders. It is generally believed that cancer cells take up more glucose than normal cells and prefer aerobic glycolysis to produce lactate, even in the presence of oxygen, which is called the Warburg effect [48]. The *KRAS* G12D mutation alters multiple metabolic pathways. On one hand, *KRAS* mutations increase glucose uptake and lactate production by increasing the expression of glucose transporters (such as Glut1/SLC2A1), key glycolytic enzymes (such as Hk1 and Hk2), and enzymes of the hexosamine pathway and nonoxidized pentose phosphate pathway via activation of the MAPK and Myc signaling pathways, thus facilitating the survival of pancreatic cancer cells in a hypoxic environment [34,49,50]. In addition to its importance for central carbon metabolism, glutamine is also a valuable and important fuel for tumors. *KRAS* mutations can stimulate glutamine metabolism by inhibiting glutamate dehydrogenase and by activating aspartate aminotransferase, which is critical for pancreatic cancer growth, development and maintenance [51]. Proliferating cells require fatty acid synthesis to produce lipids, which are used in processes such as membrane synthesis and energy generation [18]. In conclusion, the above findings demonstrate that oncogenic *KRAS* mutations promote the initiation, development and maintenance of pancrea-

tic cancer in multiple ways, and therefore, targeting KRAS signaling for the treatment of pancreatic cancer is reasonable (Figure 2).

KRAS targeting strategies and challenges

Strategies targeting KRAS as a treatment for pancreatic cancer have been widely studied, but there are substantial challenges. Current strategies mainly include directly targeting KRAS and targeting proteins upstream and/or downstream of KRAS signaling pathways. KRAS was once considered untargetable because of its high affinity for GTP, which prevented the development of competitive inhibitors of GTP binding. However, in recent years, a switch-II pocket in KRAS G12C was found to bind to a series of covalent small molecule inhibitors, causing the destruction of the switch-I and switch-II regions of KRAS, thereby locking KRAS in a GDP-bound inactive state and preventing downstream signaling [52]. Since the initial success of these selective small-molecule inhibitors targeting the KRAS G12C mutation, more potent covalent inhibitors, such as sotorasib [53], MRTX849 [54], JNJ-74699157 and LY3499446, have been developed. MRTX849 as a treatment for pancreatic cancer is currently being tested in clinical trials, and one patient in the phase I/Ib cohort had a partial response (NCT03785249) [55]. Epidermal

growth factor (EGF) signaling stimulates KRAS activation. The binding of EGF to the EGF receptor (EGFR) on the cell membrane stimulates the phosphorylation of SHC, which complexes with the GEF son of sevenless (SOS) and growth factor receptor-bound protein 2 (GRB2) to promote the binding of GEFs to KRAS, causing KRAS to exist in the activated GTP-bound state [56]. However, first-generation EGFR inhibitors, represented by gefitinib and erlotinib, have shown very limited efficacy [57,58]. This may be due to other resistance mechanisms in pancreatic cancer that allow bypass of EGFR inhibition or resistance resulted from the non-EGFR members of the ERBB family which includes four receptor tyrosine kinases [59]. Irreversible tyrosine kinase inhibitors that can inhibit the activation of all members of the ERBB family, such as afatinib and neratinib, have also been developed and are currently being tested in clinical trials [60–62]. The RAF-MEK-ERK MAPK pathway and the PI3K-AKT-mTOR pathway are the most characteristic downstream signaling pathways of KRAS and are the focus of work to develop drugs targeting various KRAS mutations. However, currently available RAF inhibitors, such as vemurafenib and dabrafenib, have not shown therapeutic efficacy in KRAS-mutant cancers and are even thought to promote tumor development [63,64]. Clinical trials of MEK inhibitors such as selumetinib and

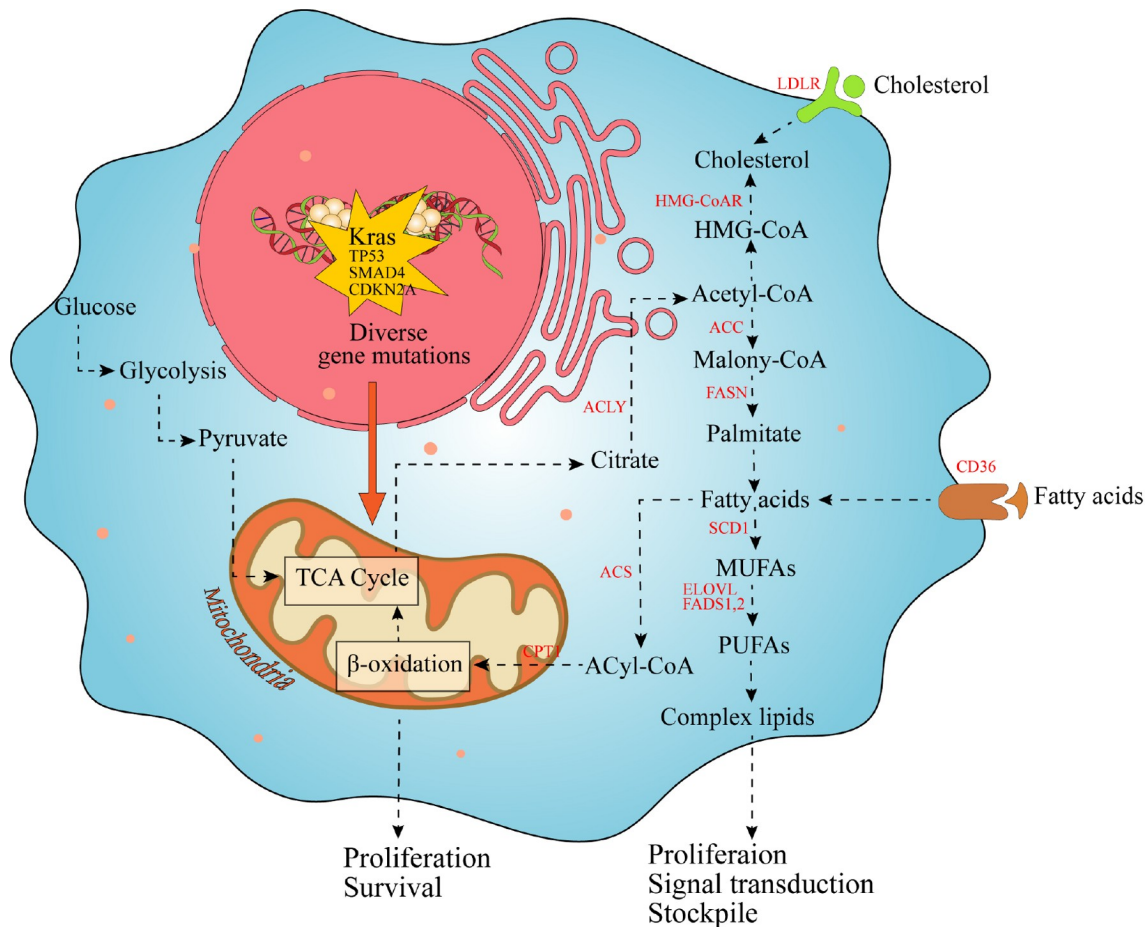


Figure 2. KRAS mutations in pancreatic cancer are involved in many biological processes On one hand, through the downstream RAF-MEK-ERK and PI3K-AKT-mTOR signaling pathways, KRAS mutations increase the proliferation and survival ability of pancreatic cancer cells, contributing to immune escape and drug resistance and leading to metabolic reprogramming. On the other hand, various substances secreted by pancreatic cancer cells caused by KRAS mutations change the tumor immune microenvironment (substances such as IL-4, IL-6, IL-10, IL-13, KrasG12D protein, TGF- β , MCP-1, CSF-1, and CCL2) and extracellular matrix structure (such as Hedgehog ligand and TGF- β) and contribute to tumor angiogenesis (such as VEGF).

trametinib in patients with advanced pancreatic cancer have also failed [65,66]. In addition, although everolimus, an mTOR inhibitor, significantly prolonged progression-free survival and decreased the severe adverse event rate in patients with progressive pancreatic neuroendocrine tumors in a phase III clinical trial [67], PI3K inhibitors are not used in the treatment of pancreatic cancer [68,69]. Despite decades of intensive efforts, no breakthroughs have been achieved in treating pancreatic cancer by targeting KRAS. This is due in part to the multiple alternative signaling pathways linked to KRAS, which may be activated when another is inhibited.

Other common gene mutations in pancreatic cancer

TP53 is a tumor suppressor gene whose encoded product (the P53 protein) plays an important role in the response to cellular stress and in the modulation of the cell cycle, cell apoptosis and cell metabolism [70]. Mutations in *TP53* occur in 60%–70% of PDAC cases; these mutations lead to activation of the cell cycle, loss of cell apoptosis regulation and enhanced metabolism and reshape the tumor microenvironment to promote cancer development. *SMAD4*, also known as *DPC4*, is also a tumor suppressor gene, and *SMAD4* mutation occurs in approximately 50% of pancreatic cancers. The SMAD family plays an important role in mediating transforming growth factor (TGF)- β signal transduction [71,72]. When phosphorylated by activated TGF- β family receptors, *SMAD4* migrates to the nucleus in the form of heterodimeric *SMAD2/SMAD3-SMAD4* complexes and interacts with downstream proteins, leading to cell growth inhibition [73]. *SMAD4* mutations lead to the loss of activated proteins, attenuating the tumor suppressor function of the TGF- β pathway, and are associated with metastasis and related epithelial-mesenchymal transformation of tumor cells. *CDKN2A* encodes a protein that controls the G1/S checkpoint [74]. By inhibiting cyclin-CDK4 and cyclin-CDK6 complexes, which are associated with the G1/S phase transition, *CDKN2A* regulates cell cycle progression. Alterations in the *CDKN2A* gene have been found in >50% of pancreatic cancer cases. *CDKN2A* inactivation acts synergistically with *KRAS* mutations to promote malignant progression of pancreatic cancer. Generally, mutations in *KRAS* and *CDKN2A* occur during carcinogenesis and before invasion into the pancreatic parenchyma, whereas *TP53* and *Smad4* inactivation are relatively later events [75,76]. In addition, some low-frequency mutations have been reported in the development of pancreatic cancer, such as mutations in genes involved in the DNA damage response (*ATM* and *BRCA2*) and epigenetic regulation (*ARID1A*, *ARID1B*, *SMARCA1*, *MLL2*, *MLL3*, and *KDM6A*). The occurrence of pancreatic cancer may involve the accumulation of mutations in multiple genes [77,78].

Lipid Metabolism Reprogramming in Pancreatic Cancer and its Effect

Developing therapeutic *KRAS* inhibitors is a challenging process, so existing drugs and strategies that indirectly target *KRAS* should be explored for pancreatic cancer treatment. Metabolic reprogramming is one of the hallmarks of tumors and an important potential target for tumor therapy. During tumorigenesis and tumor progression, cancer cells need to reprogram their catabolic and anabolic processes to survive and grow; among these processes, lipid metabolism reprogramming is prominent. Research has identified that abnormal lipid metabolism contributes to pancreatic cancer, but the precise mechanism has not been explained in detail. This

section reviews the research progress in understanding the role of lipid metabolism reprogramming and other regulatory mechanisms in the development of pancreatic cancer.

Lipid metabolism reprogramming in pancreatic cancer

Lipids are one of the three major nutrients and metabolites in the human body and play significant physiological roles in cells, for example, constituting the basic structure of cell membranes, storing energy, acting as signal molecules, and synthesizing hormones. There are generally two types of lipids that can be taken up by normal mammalian cells: one type is derived from food (free fatty acids or complexes formed with low-density lipoprotein, among other things), and the other type is derived from synthesis in the body (a small proportion). However, the lipids in tumor cells are mainly derived from tumor cell synthesis [79]. The process includes the following main characteristics: the *de novo* synthesis of fatty acids is increased, while the oxidation of fatty acids is decreased to meet the needs of tumor cell proliferation. The upregulation of lipid metabolism-related proteins and enzymes promotes malignant tumor progression (Figure 3) [80–82]. *De novo* fatty acid synthesis is first catalyzed by ATP citrate lyase (*ACLY*) to generate acetyl coenzyme A (acetyl-CoA); then, acetyl-CoA is carboxylated by acetyl-CoA carboxylase (*ACC*) to generate malonyl coenzyme A (malonyl-CoA). Subsequently, acetyl-CoA and malonyl-CoA are coupled to the acyl carrier protein (*ACP*) domain to generate fatty acid synthase (*FAS*). Finally, *FAS* catalyzes malonyl-CoA to generate palmitate in a process that involves changes in the levels of a variety of rate-limiting enzymes, mainly including increased expression of *ACLY*, *ACC* and *FAS* [83]. Among these enzymes, *ACLY* is the first enzyme in the *de novo* synthesis of fatty acids and a key enzyme linking the glycolysis and lipid metabolism pathways. *ACLY* is highly expressed in PDAC, and *ACLY* expression is negatively correlated with the prognosis of patients. *In vitro* cell experiments have shown that downregulating the expression of endogenous *ACLY* by siRNA can reduce the activity of pancreatic cancer cells and induce apoptosis [84]. In addition, *ACLY* expression in lung cancer tissue is higher than that in normal lung tissue and is associated with tumor stage, degree of differentiation and prognosis [85,86]. A previous study has also found that *ACLY* can regulate colon cancer invasion and metastasis through *CTNNB1* and play an important role in colon cancer progression [87], which indicates that *ACLY* is associated with tumor progression. High expression of *ACC* can be detected in early-stage breast cancer, prostate cancer and hepatocellular carcinoma, and the phosphorylation level of *ACC* is closely related to tumor metastasis. *ACC* inhibitors are effective in cancer treatments [88,89]. Another study showed that inhibition of *ACC* can attenuate WNT and Hedgehog signaling pathways, suppress pancreatic cancer tumor growth, and induce apoptosis of the pancreatic cancer cell lines AsPC-1, BxPC-3, and PANC-1 [90,91]. Another key enzyme in *de novo* fatty acid synthesis is *FAS*. Upregulation of *FAS* expression is a very common feature of human cancer and precancerous lesions and is closely related to chemotherapy resistance, tumor metastasis and poor patient prognosis. Previous studies have shown that the upregulation of *FAS* expression can be used as a marker for the clinical diagnosis of pancreatic cancer [92–95]. These findings suggest that controlling rate-limiting enzymes in fatty acid synthesis can effectively inhibit tumor growth and may be a strategy for pancreatic cancer therapy. In addition to supplying phospholipids

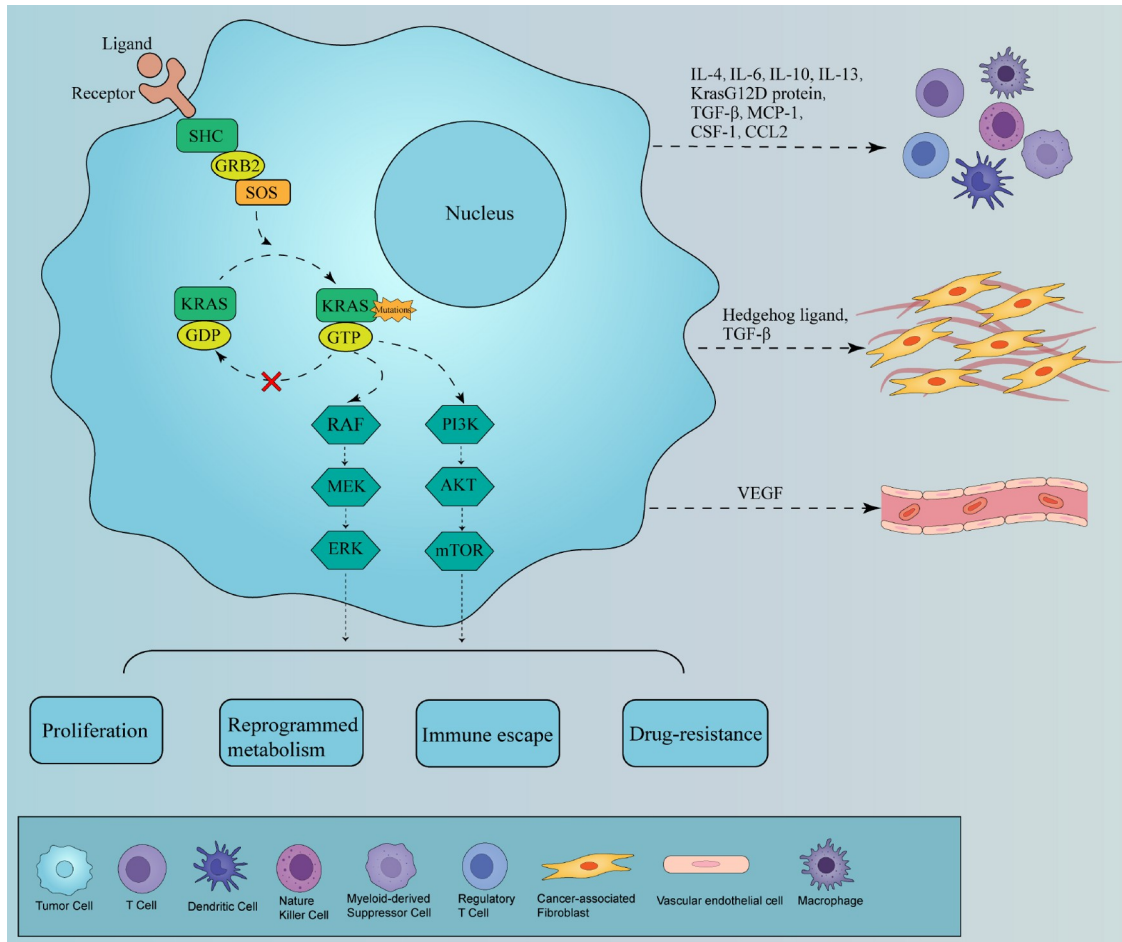


Figure 3. Gene mutations cause reprogramming of lipid metabolism in pancreatic cancer cells to meet their proliferation needs This is reflected in the increased *de novo* synthesis of fatty acids, enhanced activities of various rate-limiting enzymes, and upregulated expression of receptors for transporting exogenous cholesterol and lipid droplets. The synthesized lipids satisfy the proliferation and survival of pancreatic cancer and signal transduction.

and signaling molecules necessary for cell proliferation, fatty acids synthesized by tumor cells enter mitochondria to undergo β -oxidation, which is an important process that provides energy for tumor cell proliferation. Although fatty acid oxidation (FAO) is typically enhanced in other tumors, it is decreased in PDAC due to the unique tumor microenvironment of pancreatic cancer, and the reduction in fatty acid consumption is conducive to tumor cell proliferation and migration [96,97]. Carnitine palmitoyl transferase (CPT) is a rate-limiting enzyme in FAO, and there are two types: CPT1 and CPT2. CPT1 transports fatty acids into mitochondria for β -oxidation. Overexpression of CPTs in the pancreatic cancer cell lines CFPAC-1, BxPC-3, and PANC-1 inhibits tumor growth and increases sensitivity to the chemotherapeutic drug gemcitabine, whereas inhibition of CPT expression promotes pancreatic cancer progression [98]. In tumor cells, an increase in FAO occurs simultaneously with an increase in *de novo* synthesis of fatty acids. CPT1 is a key enzyme in the FAO process. Fatty acids are first activated to generate fatty acyl-CoA and then transported by CPT1 to mitochondria for FAO. After dehydrogenation, water addition, rehydrogenation and thiohydrolysis, acetyl-CoA is generated and enters the tricarboxylic acid cycle. The above process not only generates ATP to supply energy to cells but also prevents lipid toxicity caused by

excessive accumulation of lipids. The generated acetyl-CoA enters the cytoplasm and participates in the metabolic reaction to produce NADPH, which generates a large amount of NADPH to support cell redox homeostasis, thus preventing oxidative damage to tumor cells [99]. FAO plays a key role in tumor cell proliferation and chemotherapy resistance. Inhibition of FAO in mitochondria affects the production of NADPH, increases the production of reactive oxygen species, promotes the consumption of ATP in tumor cells, and results in cell death [100]. Targeting CPT1, a key enzyme in the FAO pathway, can enhance the radiotherapy effect in nasopharyngeal carcinoma patients [101]. Studies have shown enhanced reprogramming of mitochondrial FAO in breast cancer, and the expression of CPT1A/CPT2 is increased in recurrent breast cancer, which is associated with poor prognosis of breast cancer patients [102]. These findings indicate that fatty acid metabolism in cancer cells can be reprogrammed according to cellular energy and nutrient needs to ensure tumor survival.

Gene mutations mediate reprogramming of lipid metabolism in pancreatic cancer

Data from a mouse model of *KRAS*-driven pancreatic cancer show that mutationally activated *KRAS* can not only drive pancreatic

carcinogenesis but also synergize with genetic alterations to reprogram lipid metabolism to promote tumor cell proliferation. The study found that *KRAS* mutation cooperates with signal transduction *GNAS* gene mutation in PDAC, and *GNAS* gene mutation can support PDAC growth by inducing salt-inducible kinases (SIKs). Proteomic studies have shown that this pathway is associated with lipid metabolism and increased content of peroxisomes, organelles required for long-chain fatty acid processing and ether lipid production [103]. Normal cell growth requires the coordinated synthesis of biological macromolecules, and a kinase complex, mammalian target of rapamycin complex 1 (mTORC1), was recently reported to sense nutrient availability and energy supply and regulate the activity of sterol regulatory element binding proteins (SREBPs) to control the synthesis of fatty acids and cholesterol [104–106]. There are three subtypes of SREBPs: SREBP1a, SREBP1c, and SREBP2 [107]. Ferroptosis is another form of cell death that depends on the accumulation of iron and reactive oxygen species in the cell, which causes lipid peroxidation [108,109]. The FBW7-NRA41-SCD1 axis synchronously regulates apoptosis and ferroptosis in pancreatic cancer cells. FBW7 functions as a tumor suppressor by targeting oncoproteins for degradation. Zeng *et al.* [7] found that FBW7-downregulated genes are widely involved in the redox reaction and lipid metabolism. FBW7 regulates lipid peroxidation and promotes cell apoptosis. Further mechanistic studies showed that FBW7 inhibits the expression of stearoyl-CoA desaturase (SCD1) by inhibiting Group 4A member 1 (NR4A1) of the nuclear receptor subfamily. Overexpression of stearoyl-CoA desaturase (SCD) in pancreatic cancer can induce resistance to apoptosis and ferroptosis triggered by hypoxia and nutrient deprivation, resulting in poor prognosis of pancreatic cancer [110,111]. Moreover, increased ferroptosis induced by lipid metabolism inhibits gemcitabine resistance in pancreatic cancer [112]. SREBP1 can affect the expression of SCD, inhibit tumor growth, and affect the occurrence and prognosis of pancreatic cancer [113,114]. The *KRAS* gene controls the levels of cholesterol in cancer cells, and cholesterol is the major component of synthetic hormones and biofilms. Previous studies have found that *KRAS* promotes the growth and metastasis of pancreatic cancer by upregulating the expressions of the transcription factors TFCP2 and SREBP2. SREBP2 can also upregulate hydroxy-methyl glutaryl coenzyme A reductase (HMGR), resulting in an increase in cholesterol synthesis and further upregulation of low-density lipoprotein receptor (LDLR) expression to increase endocytic cholesterol uptake [115,116]. The total serum cholesterol (TSC) level is an important predictor of pancreatic cancer prognosis and affects the Hedgehog and STAT3 signaling pathways to promote the proliferation of pancreatic cancer cells [117–119]. These findings suggest that inhibition of cholesterol metabolism can regulate pancreatic cancer progression. Statins can inhibit the biosynthesis of cholesterol and are not only widely used in cardiovascular disorders to treat hyperlipidemia and atherosclerosis but also show good effects in inhibiting tumor growth [120–122]. In cell experiments, statins inhibited the Akt/PKB signaling pathway through the P27X receptor, inhibited the proliferation of the pancreatic cancer cell lines PANC-1 and MIA PaCa-2, and increased the sensitivity to chemotherapy. *In vivo* experiments have shown that statins can induce the differentiation of pancreatic cancer cells of the basal phenotype in *KRAS* mutant mice by disrupting cholesterol synthesis [21,123]. *KRAS* can also upregulate lipid

metabolism enzymes, such as acyl-coenzyme A synthetase long chain family members 3 and 4 (Acsl3 and Acsl4), to increase FAO in a mouse model. Acsl3 can promote fatty acid absorption and retention and β -oxidation and convert fatty acids into acyl-CoA [124]. *KRAS* can also control hormone-sensitive lipase (HSL) to regulate the accumulation and utilization of lipid droplets (LDs) and to regulate the invasive ability of pancreatic cancer [125]. Pancreatic cancer tumors have dense tissue, poor blood supply, hyperoxidation and inflammation. In such a harsh environment, tumor cells must change their metabolism to achieve rapid growth [126]. Glycerophospholipids are an important component of the cell membrane, and lysophospholipids are the metabolic intermediates of glycerophospholipids. The decrease in plasma lysophospholipid levels in patients with pancreatic cancer suggests that lysophospholipids may reduce the risk of pancreatic cancer, which may be due to the increased catabolism of lysophospholipids in pancreatic cancer cells and the abnormal liver function associated with cancer [127–131]. Bile acid is an important component of bile, and there is evidence that bile acid plays an important role in the development of gastrointestinal malignancies. The serum levels of taurocholic acid, bile acid and glycolic acid in patients with pancreatic cancer are significantly increased. Therefore, serum bile acid levels may be an important diagnostic marker in the identification of patients with pancreatic cancer [132,133]. Rozeveld *et al.* [125] reported that the oncogene *KRAS* controls the storage and utilization of lipid droplets in cells by regulating hormone-sensitive lipase. In pancreatic cancer cells, the destruction of the *Kras*-hormone-sensitive lipase axis leads to the storage of excessive lipid droplets in cells, and excess fatty acids enhance the growth and metastasis of cancer cells. In summary, gene mutations can alter lipid metabolism to sustain pancreatic cancer growth. These findings suggest that patients with pancreatic cancer who receive metabolism-modulating treatments may have significantly improved outcomes. Therefore, the study of lipid metabolism is not only beneficial for understanding the progression of pancreatic cancer but also helpful to provide new perspectives for the treatment of pancreatic cancer.

Crosstalk related to lipid metabolism reprogramming within the tumor microenvironment

The tumor microenvironment has been widely accepted to increase the metabolic reprogramming of tumor cells, providing a favorable environment for tumor growth and survival [134]. The tumor microenvironment comprises an immune microenvironment dominated by immune cells and a nonimmune microenvironment dominated by tumor cells and fibroblasts [135]. In recent years, the effect of lipid metabolism on immune cell function has been a focus of research in the field of oncology [136,137]. In normal cells, a stable network formed by factors related to the metabolism of the three major nutrients supports cell growth and function. In the tumor microenvironment, tumor cells are surrounded by layers of different types of cells, including interstitial cells and immune cells, nerve fibres and extracellular matrix. These factors influence tumor cells, generating a hypoxic or nutrient-deprived environment, which forces tumor cells to reprogram their metabolism to absorb enough nutrients from the microenvironment to resist killing by immune cells [138–140]. Lipid metabolism reprogramming in tumor cells is not only driven by the needs of the tumor cells themselves but also regulated by other cells; furthermore, it affects the function

and metabolism of surrounding cells, of which immune cells are the main affected cell population. Previous studies have found that microenvironment immunosuppression mediated by metabolic reprogramming is a key factor limiting immunotherapy efficacy. Abnormal lipid metabolism is closely related to the effector function of immune cells [141–144]. The unique lipid requirements of tumor cells increase the lipid levels in the tumor microenvironment, and a lipid-rich tumor microenvironment further affects the characteristics of immune cells, such as their proliferation, differentiation and execution of cellular functions [145,146]. In immune cells, increased expression of lipid metabolism-related enzymes, such as ACC, the fatty acid transferase CD36, CPT1A, FAS, and SCD1, affects the type of lipids and triggers the accumulation of LDs in cells, thereby affecting the status and function of immune cells [142,147]. The expression of ACC is significantly upregulated in breast and prostate cancers, and inhibition of ACC can reduce the differentiation of T cells expressing interleukin 17 (IL-17) and promote the differentiation of Tregs and tumor cell apoptosis [148]. CD36 can recognize many endogenous ligands, transport fatty acids into cells and activate FAO. Increased CD36 expression in tumors can prevent CD8⁺ T cells from producing cytokines and support Treg survival, thereby decreasing antitumor function. Upregulation of CD36 and FAS expression in T cells is correlated with tumor progression and poor prognosis [149–151]. Increased expression of CPT1A in macrophages promotes fatty acid transport into mitochondria to promote β -oxidation, resulting in a decrease in the levels of proinflammatory cytokines secreted by macrophages. Moreover, inhibitors of CPT1A can suppress Treg differentiation and tumor cell proliferation [152,153]. In addition, while tumor cells have their own metabolic characteristics, their metabolites can also affect the activation of immune cells and induce antitumor immune responses in a variety of ways [24]. Ultimately, the microenvironment is transformed into a place that supports the proliferation and development of tumor cells.

The pancreatic cancer tumor microenvironment contains various types of immune cell subsets, including CD4⁺ T cells, CD8⁺ T cells, B cells, dendritic cells (DCs), macrophages, and natural killer (NK) cells [154]. Among them, DCs, CD4⁺ and CD8⁺ effector T cells, and NK cells are activated to inhibit tumors and prevent immune escape, while other immune cells, such as tolerogenic dendritic cells (tDCs), Tregs, and TAMs, inhibit the antitumor immune response, thereby promoting tumor cell proliferation, invasion, metastasis and angiogenesis [155–160]. Tumor tissue is infiltrated by immune cells (mainly lymphocytes), which are generally thought to attack tumor cells. However, researchers have found that tumor-infiltrating lymphocytes can promote tumor metastasis, resulting in poor prognosis of pancreatic cancer patients [161]. Tregs are an important type of tumor-infiltrating lymphocyte and are a special class of CD4⁺ T cells that promote tumor growth and invasion by suppressing host immune responses and proinflammatory responses. Forkhead transcription factor 3 (FoxP3) belongs to the Forkhead family of transcriptional regulators. FoxP3 can inhibit the expression of target immune genes and is one of the most specific markers of Tregs in tumors [162]. FoxP3⁺ Tregs and CD8⁺ T cells are the two main T-cell populations in the tumor microenvironment. FoxP3⁺ Tregs disrupt the antitumor immune response by inhibiting the function of cytotoxic T lymphocytes, thereby allowing pancreatic cancer cells to escape immune surveillance, and higher levels of FoxP3⁺ Tregs are associated with lower overall survival

and recurrence in pancreatic cancer patients [163]. Abnormal angiogenesis and activation of inhibitory checkpoint pathways in the tumor microenvironment create an immunosuppressive microenvironment, which prevents CD8⁺ T cells from infiltrating the tumor. However, CD8⁺ T cells may directly lead to tumor cell apoptosis by releasing perforin and granzyme, which prevents the development and progression of tumors [164–166]. B lymphocytes mediate antitumor responses by promoting antigen presentation, efficiently priming T cells, and producing antitumor antibodies. However, recent studies have found that interleukin-1 β can induce the expansion of B lymphocytes and promote the proliferation of pancreatic cancer cells [167]. In general, B lymphocytes play a key role in the development of PDAC, but the mechanism of action is still controversial and needs to be further elucidated [168]. Macrophages are important immune cells that maintain tissue and immune system homeostasis. They are also one of the largest leukocyte populations in the tumor stroma and play an important role in tumor progression [169]. Macrophages can adjust their phenotype to respond to microenvironmental stimuli and transduce signals according to their functional requirements [170]. Generally, macrophages can be divided into two types, the classically activated M1 type and the alternatively activated M2 type, according to their polarization state and function [171]. Macrophages infiltrating tumor tissues, known as TAMs, can promote tumor cell growth through a variety of mechanisms, including promoting tumor angiogenesis, enhancing chemotherapy resistance, and suppressing tumor immunity. M1-type macrophages are activated by Th1 cytokines and have proinflammatory and antitumor effects; M2-type macrophages are activated by Th2 cytokines. They have anti-inflammatory effects and induce tissue remodeling and tumor cell proliferation, invasion and metastasis in early-stage tumors [172–175]. Mitchem *et al.* [176] found that inhibition of colony-stimulating factor-1 receptor (CSF1R) or C-motif chemokine ligand 2 (CCR2) on TAMs can reduce the number of tumor-initiating cells and overcome macrophage-induced CD8⁺ cytotoxic T lymphocyte inhibition to improve the effect of chemotherapy in pancreatic cancer. Sanford *et al.* [177] found that pancreatic tumors recruit TAMs through the CCL2/CCR2 chemokine axis to generate an immunosuppressive tumor microenvironment. As such, CCR2 inhibitors may have a strong antitumor effect in PDAC patients and are worthy of further study in clinical trials. Treg infiltration is a distinctive characteristic of PDAC, and Tregs are required for pancreatic tumorigenesis. Tregs can reduce the effect of CD8⁺ T cells and other immune cells and/or suppress T-cell function, resulting in tumor cell evasion of immune surveillance. Jang *et al.* [178] found that the development of pancreatic tumors is accompanied by the gradual accumulation of activated Treg cells, which promote PDAC progression by inhibiting the antitumor response of CD8⁺ T cells. Therefore, a comprehensive understanding of the lipid metabolism patterns of cancer cells and immune cells will aid the development of new therapeutic strategies for pancreatic cancer. Figure 4 summarizes the effect of pancreatic cancer lipid metabolism reprogramming on the immune microenvironment.

Effects of lipid metabolism reprogramming on drug resistance

Abnormal lipid metabolism can not only alter the tumor microenvironment and cause immune escape but is also involved in the resistance to chemotherapy. For the past two decades, gemcitabine has been the cornerstone of pancreatic cancer chemotherapy and

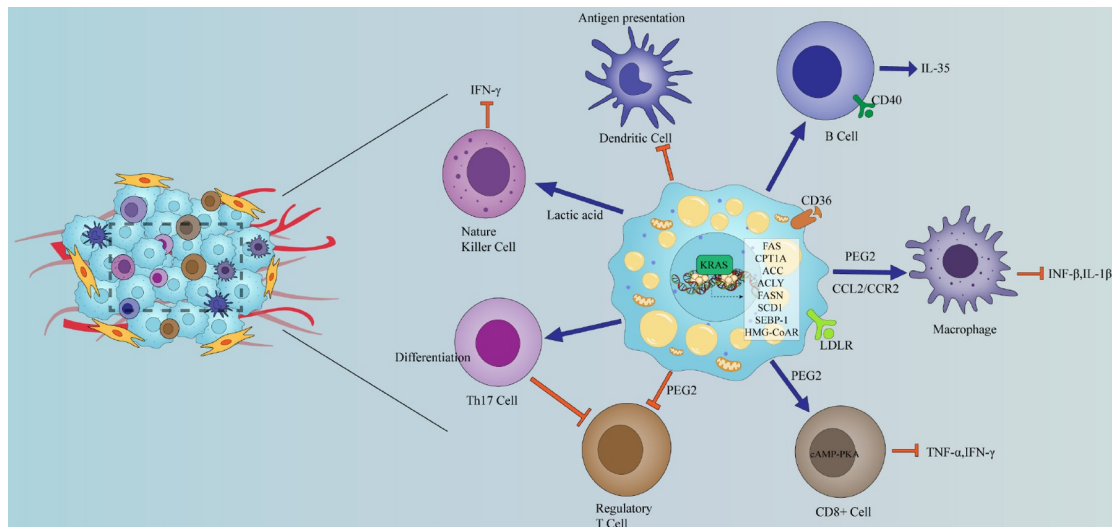


Figure 4. Excessive lipid accumulation in the tumor microenvironment of pancreatic cancer attenuates the antitumor immune response First, the upregulation of various enzymes related to lipid metabolism affects the function of immune cells, including the following aspects: overexpression of ACC and FASN can promote the differentiation of Th17 cells, inhibit the differentiation of Treg cells and promote tumor progression; CD36 and LDLR translocate excessive fatty acids and cholesterol into cells, activate fatty acid β -oxidation, inhibit the secretion of TNF- α and IFN- γ cytokines by CD8 $^{+}$ T cells, and weaken the antitumor response; and the increased expressions of CPT1A, FAS, SCD1 and other enzymes can change the lipid type or trigger the accumulation of intracellular lipid droplets, thereby affecting the status and function of immune cells. Second, tumor cells secrete PEG2 to promote the transformation of M1-type macrophages into M2-type macrophages, and PEG2 activates the cAMP-PKA signaling pathway, leading to the growth arrest of CD8 $^{+}$ T cells and the antitumor activity of Tregs and DCs. Tumor cells can also secrete lactic acid to cause intracellular acidification of NK cells, inhibit the secretion of IFN- γ , and promote NK cell apoptosis. Excessive lipid accumulation leads to antigen presentation dysfunction in DCs, which in turn fails to activate primary T cells and ultimately reduces the antitumor immune response. Pancreatic cancer cells activate the B-cell surface receptor CD40, and B cells secrete IL-35 to promote tumor cell proliferation. Pancreatic cancer can also recruit TAMs through the CCL2/CCR2 chemokine axis, resulting in decreased secretion of proinflammatory factors such as IFN- β and IL-1 β , which in turn causes insufficient recruitment of effector T cells and NK cells and builds an immunosuppressive tumor microenvironment. Increased CPT1A expression in macrophages promotes fatty acid transport into mitochondria, promotes β -oxidation, and inhibits macrophage secretion of the proinflammatory factors IFN- β and IL-1 β .

plays an important role in the treatment of borderline resectable, locally advanced or advanced metastatic pancreatic cancer. Furthermore, gemcitabine can also be used for postoperative chemotherapy, neoadjuvant chemotherapy, and palliative chemotherapy in patients with distant metastases or locally advanced unresectable disease [179,180]. Gemcitabine enters pancreatic cancer cells as a nucleotide analog and undergoes a series of complex phosphorylation steps to produce derivatives that interfere with DNA synthesis and arrest the pancreatic cancer cell cycle [181]. However, within a few weeks of starting chemotherapy, the response rate of pancreatic cancer patients to gemcitabine is less than 20%, and the remaining 80% of patients survive for less than a year. Resistance to gemcitabine is the main reason for the limited effect of chemotherapy and poor prognosis [182,183]. Fujimura *et al.* [184] used mass spectrometry to analyze the metabolomes of gemcitabine-sensitive and gemcitabine-resistant pancreatic cancer CAPAN-1 and SUIT-2 cell lines and found differences in the amino acid, nucleotide, glucose, lipid, and energy profiles, suggesting that nutrient metabolism may be a reason for chemoresistance. In fact, pancreatic cancer cell lines with high expression of FAS have upregulated PKM2 expression and p53 signaling pathway activity, increased glycolysis to generate energy for tumor cell proliferation, and reduced endoplasmic reticulum stress, and can avoid cell death. These effects maintain the stemness of pancreatic cancer stem cells and result in drug resistance. In another *in vitro* cell experiment, the use of the FAS inhibitor orlistat increased sensitivity to gemcitabine [93,185–187]. Alzoubi *et al.* [188] also found that overexpression of

tumor necrosis factor- α (TNF- α) can downregulate the protein expression of FAS/ACC, thereby inhibiting tumor growth. Pre-clinical studies have found that the antibiotic cerulenin inhibits the proliferation of lung cancer cells by inhibiting FAS. Although agents targeting FAS have not yet been approved for clinical use, FAS holds the promise as a potential therapeutic target [189]. In addition, activation of LDLR increases cholesterol uptake, and downregulation of LDLR induced by oridonin via the ERK/JNK signaling pathway can also increase the sensitivity of PANC-1 cells to gemcitabine and promote tumor cell apoptosis [190]. Currently, an increasing number of preclinical studies and clinical trials are focusing on the feasibility of targeting lipid metabolism-related processes in the treatment of pancreatic cancer. A phase II clinical trial that combines simvastatin with gemcitabine as treatment for patients with locally advanced and metastatic pancreatic cancer failed to achieve the expected clinical benefit (NCT00944463) [191]. Nevertheless, several clinical trials for pancreatic cancer treatment are still underway. For example, a phase I clinical trial was designed to explore whether lowering cholesterol levels with FOLFIRINOX chemotherapy as a treatment for advanced pancreatic cancer could yield beneficial outcomes (NCT04862260). Another phase III trial is testing the curative effect of the combination of simvastatin with digoxin and metformin in patients with advanced pancreatic cancer (NCT02201381).

Conclusions

Genetic alterations drive more than 95% of pancreatic cancers.

Despite decades of research, pancreatic cancer remains a highly lethal malignancy that derives minimal benefit from conventional cytotoxic therapies. Newly approved inhibitors of KRAS G12C can only benefit a subset of patients, and there are currently no drugs targeting KRAS G12D or KRAS G12V. As such, scientists should shift their focus and work to develop indirect targeting strategies. Metabolic reprogramming plays an important role in the occurrence and progression of pancreatic cancer, providing the energy and materials needed for pancreatic cancer cells to survive and further evolve in harsh environments. Immune cells also show abnormal lipid metabolism in the tumor microenvironment. These changes affect the function and status of immune cells and can result in weakened immune responses and immune escape, further promoting invasion and metastasis. Research on strategies targeting genes and enzymes related to tumor and immune cell lipid metabolism has provided powerful evidence for tumor prevention and treatment. Here, research progress in understanding the relationship between lipid metabolism reprogramming and drug resistance in pancreatic cancer was systematically reviewed to provide a theoretical basis for the development of new pancreatic cancer chemotherapy drugs and solutions to overcome the problem of drug resistance. In conclusion, since the initiation and development of pancreatic cancer are driven by various mutations and the dysregulation of signaling factors and not by disruption of a single pathway, the application of multitarget drugs is a promising direction for pancreatic cancer treatment in the future.

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Conflict of Interest

The authors declare that they have no conflict of interest.

References

- Rawla P, Sunkara T, Gaduputi V. Epidemiology of pancreatic cancer: global trends, etiology and risk factors. *World J Oncol* 2019, 10: 10–27
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. *CA Cancer J Clin* 2022, 72: 7–33
- Rahib L, Wehner MR, Matrisian LM, Nead KT. Estimated projection of US cancer incidence and death to 2040. *JAMA Network Open* 2021, 4: e214708
- Huang L, Jansen L, Balavarca Y, Molina-Montes E, Babaei M, van der Geest L, Lemmens V, *et al.* Resection of pancreatic cancer in Europe and USA: an international large-scale study highlighting large variations. *Gut* 2019, 68: 130–139
- Kleeff J, Reiser C, Hinz U, Bachmann J, Debus J, Jaeger D, Friess H, *et al.* Surgery for recurrent pancreatic ductal adenocarcinoma. *Ann Surg* 2007, 245: 566–572
- Springfield C, Jager D, Buchler MW, Strobel O, Hackert T, Palmer DH, Neoptolemos JP. Chemotherapy for pancreatic cancer. *Presse Med* 2019, 48: E159–E74
- Zeng, Pöttler, Lan, Grützmann, Pilarsky, Yang. Chemoresistance in pancreatic cancer. *Int J Mol Sci* 2019, 20: 4504
- Collisson EA, Bailey P, Chang DK, Biankin AV. Molecular subtypes of pancreatic cancer. *Nat Rev Gastroenterol Hepatol* 2019, 16: 207–220
- Collins MA, Bednar F, Zhang Y, Brisset JC, Galbán S, Galbán CJ, Rakshit S, *et al.* Oncogenic Kras is required for both the initiation and maintenance of pancreatic cancer in mice. *J Clin Invest* 2012, 122: 639–653
- Luo J. KRAS mutation in pancreatic cancer. *Semin Oncol* 2021, 48: 10–18
- Nagasaka M, Li YW, Sukari A, Ou SHI, Al-Hallak MN, Azmi AS. KRAS G12C game of thrones, which direct KRAS inhibitor will claim the iron throne? *Cancer Treat Rev* 2020, 84: 101974
- Takashima A, Faller DV. Targeting the RAS oncogene. *Expert Opin Therapeutic Targets* 2013, 17: 507–531
- Canon J, Rex K, Saiki AY, Mohr C, Cooke K, Bagal D, Gaida K, *et al.* The clinical KRAS(G12C) inhibitor AMG 510 drives anti-tumour immunity. *Nature* 2019, 575: 217–223
- Koga T, Suda K, Fujino T, Ohara S, Hamada A, Nishino M, Chiba M, *et al.* KRAS Secondary Mutations that confer acquired resistance to KRAS G12C inhibitors, sotorasib and adagrasib, and overcoming strategies: insights from *in vitro* experiments. *J Thoracic Oncol* 2021, 16: 1321–1332
- Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metab* 2016, 23: 27–47
- Ward PS, Thompson CB. Metabolic reprogramming: a cancer hallmark even warburg did not anticipate. *Cancer Cell* 2012, 21: 297–308
- Altman BJ, Stine ZE, Dang CV. From Krebs to clinic: glutamine metabolism to cancer therapy. *Nat Rev Cancer* 2016, 16: 619–634
- Currie E, Schulze A, Zechner E, Walther TC, Farese Jr. RV. Cellular fatty acid metabolism and cancer. *Cell Metab* 2013, 18: 153–161
- Hapala I, Marza E, Ferreira T. Is fat so bad? Modulation of endoplasmic reticulum stress by lipid droplet formation. *Biol Cell* 2011, 103: 271–285
- Yu S, Wang L, Che D, Zhang M, Li M, Naito M, Xin W, *et al.* Targeting CRABP-II overcomes pancreatic cancer drug resistance by reversing lipid raft cholesterol accumulation and AKT survival signaling. *J Exp Clin Cancer Res* 2022, 41: 88
- Gabitova-Cornell L, Surumbayeva A, Peri S, Franco-Barraza J, Restifo D, Weitz N, Ogier C, *et al.* Cholesterol pathway inhibition induces TGF- β signaling to promote basal differentiation in pancreatic cancer. *Cancer Cell* 2020, 38: 567–583
- Vanpouille-Box C, Lhuillier C, Bezu L, Aranda F, Yamazaki T, Kepp O, Fucikova J, *et al.* Trial watch: immune checkpoint blockers for cancer therapy. *Oncotarget* 2017, 6: e1373237
- Lequeux A, Noman MZ, Xiao M, Van Moer K, Hasim M, Benoit A, Bosseler M, *et al.* Targeting HIF-1 alpha transcriptional activity drives cytotoxic immune effector cells into melanoma and improves combination immunotherapy. *Oncogene* 2021, 40: 4725–4735
- Xia L, Oyang L, Lin J, Tan S, Han Y, Wu N, Yi P, *et al.* The cancer metabolic reprogramming and immune response. *Mol Cancer* 2021, 20: 28
- Vuononvirta J, Marelli-Berg FM, Poobalasingam T. Metabolic regulation of T lymphocyte motility and migration. *Mol Aspects Med* 2021, 77: 100888
- Saka D, Gokalp M, Piyade B, Cevik NC, Arik Sever E, Unutmaz D, Ceyhan G, *et al.* Mechanisms of T-cell exhaustion in pancreatic cancer. *Cancers (Basel)* 2020, 12: 2274
- Prior IA, Lewis PD, Mattos C. A comprehensive survey of Ras mutations

- in cancer. *Cancer Res* 2012, 72: 2457–2467
28. Bos JL, Rehmann H, Wittinghofer A. GEFs and GAPs: critical elements in the control of small G proteins. *Cell* 2007, 129: 865–877
 29. Hennig A, Markwart R, Esparza-Franco MA, Ladds G, Rubio I. Ras activation revisited: role of GEF and GAP systems. *Biol Chem* 2015, 396: 831–848
 30. Delpu Y, Hanoun N, Lulka H, Sicard F, Selves J, Buscail L, Torrisani J, *et al.* Genetic and epigenetic alterations in pancreatic carcinogenesis. *Curr Genomics* 2011, 12: 15–24
 31. Haigis KM. KRAS alleles: the devil is in the detail. *Trends Cancer* 2017, 3: 686–697
 32. di Magliano MP, Logsdon CD. Roles for KRAS in pancreatic tumor development and progression. *Gastroenterology* 2013, 144: 1220–1229
 33. Buscail L, Bournet B, Cordelier P. Role of oncogenic KRAS in the diagnosis, prognosis and treatment of pancreatic cancer. *Nat Rev Gastroenterol Hepatol* 2020, 17: 153–168
 34. Jonckheere N, Vasseur R, Van Seuning I. The cornerstone K-RAS mutation in pancreatic adenocarcinoma: from cell signaling network, target genes, biological processes to therapeutic targeting. *Crit Rev Oncol Hematol* 2017, 111: 7–19
 35. Kanda M, Matthaei H, Wu J, Hong SM, Yu J, Borges M, Hruban RH, *et al.* Presence of somatic mutations in most early-stage pancreatic intraepithelial neoplasia. *Gastroenterology* 2012, 142: 730–733
 36. Hingorani SR, Wang L, Multani AS, Combs C, Deramandt TB, Hruban RH, Rustgi AK, *et al.* Trp53R172H and KrasG12D cooperate to promote chromosomal instability and widely metastatic pancreatic ductal adenocarcinoma in mice. *Cancer Cell* 2005, 7: 469–483
 37. Collins MA, Yan W, Sebolt-Leopold JS, Pasca di Magliano M. MAPK signaling is required for dedifferentiation of acinar cells and development of pancreatic intraepithelial neoplasia in mice. *Gastroenterology* 2014, 146: 822–834
 38. Luo Y, Yang Y, Liu M, Wang D, Wang F, Bi Y, Ji J, *et al.* Oncogenic KRAS reduces expression of FGF21 in acinar cells to promote pancreatic tumorigenesis in mice on a high-fat diet. *Gastroenterology* 2019, 157: 1413–1428
 39. Zheng C, Jiao X, Jiang Y, Sun S. ERK1/2 activity contributes to gemcitabine resistance in pancreatic cancer cells. *J Int Med Res* 2013, 41: 300–306
 40. Seguin L, Kato S, Franovic A, Camargo MF, Lesperance J, Elliott KC, Yebra M, *et al.* An integrin β 3-KRAS-RalB complex drives tumour stemness and resistance to EGFR inhibition. *Nat Cell Biol* 2014, 16: 457–468
 41. Walter K, Omura N, Hong SM, Griffith M, Vincent A, Borges M, Goggins M. Overexpression of smoothed activates the sonic hedgehog signaling pathway in pancreatic cancer-associated fibroblasts. *Clin Cancer Res* 2010, 16: 1781–1789
 42. Steele NG, Biffi G, Kemp SB, Zhang Y, Drouillard D, Syu LJ, Hao Y, *et al.* Inhibition of hedgehog signaling alters fibroblast composition in pancreatic cancer. *Clin Cancer Res* 2021, 27: 2023–2037
 43. Cheng H, Fan K, Luo G, Fan Z, Yang C, Huang Q, Jin K, *et al.* KrasG12D mutation contributes to regulatory T cell conversion through activation of the MEK/ERK pathway in pancreatic cancer. *Cancer Lett* 2019, 446: 103–111
 44. Cheng H, Luo G, Jin K, Fan Z, Huang Q, Gong Y, Xu J, *et al.* Kras mutation correlating with circulating regulatory T cells predicts the prognosis of advanced pancreatic cancer patients. *Cancer Med* 2020, 9: 2153–2159
 45. Dai E, Han L, Liu J, Xie Y, Kroemer G, Klionsky DJ, Zeh HJ, *et al.* Autophagy-dependent ferroptosis drives tumor-associated macrophage polarization via release and uptake of oncogenic KRAS protein. *Autophagy* 2020, 16: 2069–2083
 46. Yang W, Yang S, Zhang F, Cheng F, Wang X, Rao J. Influence of the Hippo-YAP signalling pathway on tumor associated macrophages (TAMs) and its implications on cancer immunosuppressive microenvironment. *Ann Transl Med* 2020, 8: 399
 47. Liu Y, Deguchi Y, Wei D, Liu F, Moussalli MJ, Deguchi E, Li D, *et al.* Rapid acceleration of KRAS-mutant pancreatic carcinogenesis via remodeling of tumor immune microenvironment by PPAR δ . *Nat Commun* 2022, 13: 2665
 48. Liberti MV, Locasale JW. The Warburg effect: how does it benefit cancer cells? *Trends Biochem Sci* 2016, 41: 211–218
 49. Gaglio D, Metallo CM, Gameiro PA, Hiller K, Danna LS, Balestrieri C, Alberghina L, *et al.* Oncogenic K-Ras decouples glucose and glutamine metabolism to support cancer cell growth. *Mol Syst Biol* 2011, 7: 523
 50. Perera RM, Bardeesy N. Pancreatic cancer metabolism: breaking it down to build it back up. *Cancer Discovery* 2015, 5: 1247–1261
 51. Son J, Lyssiotis CA, Ying H, Wang X, Hua S, Ligorio M, Perera RM, *et al.* Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature* 2013, 496: 101–105
 52. Ostrem JM, Peters U, Sos ML, Wells JA, Shokat KM. K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interactions. *Nature* 2013, 503: 548–551
 53. FDA approves first KRAS inhibitor: sotorasib. *Cancer Discovery* 2021, 11: OF4
 54. Hallin J, Engstrom LD, Hargis L, Calinisan A, Aranda R, Briere DM, Sudhakar N, *et al.* The KRASG12C inhibitor MRTX849 provides insight toward therapeutic susceptibility of KRAS-Mutant cancers in mouse models and patients. *Cancer Discovery* 2020, 10: 54–71
 55. Christensen JG, Olson P, Briere T, Wiel C, Bergo MO. Targeting Kras^{G12C}-mutant cancer with a mutation-specific inhibitor. *J Intern Med* 2020, 288: 183–191
 56. Fitzgerald TL, Lertpiriyapong K, Cocco L, Martelli AM, Libra M, Candido S, Montalto G, *et al.* Roles of EGFR and KRAS and their downstream signaling pathways in pancreatic cancer and pancreatic cancer stem cells. *Adv Biol Regulation* 2015, 59: 65–81
 57. Sinn M, Bahra M, Liersch T, Gellert K, Messmann H, Bechstein W, Waldschmidt D, *et al.* CONKO-005: adjuvant chemotherapy with gemcitabine plus erlotinib versus gemcitabine alone in patients after R0 resection of pancreatic cancer: a multicenter randomized phase III trial. *J Clin Oncol* 2017, 35: 3330–3337
 58. Moore MJ, Goldstein D, Hamm J, Figer A, Hecht JR, Gallinger S, Au HJ, *et al.* Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J Clin Oncol* 2007, 25: 1960–1966
 59. Moll HP, Pranz K, Musteanu M, Grabner B, Hruschka N, Mohrher J, Aigner P, *et al.* Afatinib restrains K-RAS-driven lung tumorigenesis. *Sci Transl Med* 2018, 10: eao2301
 60. Haas M, Waldschmidt DT, Stahl M, Reinacher-Schick A, Freiberg-Richter J, Fischer von Weikersthal L, Kaiser F, *et al.* Afatinib plus gemcitabine versus gemcitabine alone as first-line treatment of metastatic pancreatic cancer: the randomized, open-label phase II ACCEPT study of the Arbeitsgemeinschaft Internistische Onkologie with an integrated analysis of the 'burden of therapy' method. *Eur J Cancer* 2021, 146: 95–106
 61. van Brummelen EMJ, Huijberts S, van Herpen C, Desar I, Opdam F, van Geel R, *et al.* Phase I study of afatinib and selumetinib in patients with KRAS-mutated colorectal, non-small cell lung, and pancreatic cancer. *Oncologist* 2021, 26: 290–e545

62. Booth L, Poklepovic A, Dent P. Neratinib decreases pro-survival responses of [sorafenib + vorinostat] in pancreatic cancer. *Biochem Pharmacol* 2020, 178: 114067
63. Hatzivassiliou G, Song K, Yen I, Brandhuber BJ, Anderson DJ, Alvarado R, Ludlam MJC, *et al.* RAF inhibitors prime wild-type RAF to activate the MAPK pathway and enhance growth. *Nature* 2010, 464: 431–435
64. Sanchez-Laorden B, Viros A, Girotti MR, Pedersen M, Saturno G, Zambon A, Niculescu-Duvaz D, *et al.* BRAF inhibitors induce metastasis in RAS mutant or inhibitor-resistant melanoma cells by reactivating MEK and ERK signaling. *Sci Signal* 2014, 7: ra30
65. Bodoky G, Timcheva C, Spigel DR, La Stella PJ, Ciuleanu TE, Pover G, Tebbutt NC. A phase II open-label randomized study to assess the efficacy and safety of selumetinib (AZD6244 [ARRY-142886]) versus capecitabine in patients with advanced or metastatic pancreatic cancer who have failed first-line gemcitabine therapy. *Invest New Drugs* 2012, 30: 1216–1223
66. Infante JR, Somer BG, Park JO, Li C-P, Scheulen ME, Kasubhai SM, Oh DY, *et al.* A randomized, double-blind, placebo-controlled trial of trametinib, an oral MEK inhibitor, in combination with gemcitabine for patients with untreated metastatic adenocarcinoma of the pancreas. *Eur J Cancer* 2014, 50: 2072–2081
67. Yao JC, Shah MH, Ito T, Bohas CL, Wolin EM, Van Cutsem E, Hobday TJ, *et al.* Everolimus for advanced pancreatic neuroendocrine tumors. *N Engl J Med* 2011, 364: 514–523
68. Wolpin BM, Hezel AF, Abrams T, Blaszkowsky LS, Meyerhardt JA, Chan JA, Enzinger PC, *et al.* Oral mTOR inhibitor everolimus in patients with gemcitabine-refractory metastatic pancreatic cancer. *J Clin Oncol* 2009, 27: 193–198
69. Javle MM, Shroff RT, Xiong H, Varadhachary GA, Fogelman D, Reddy SA, Davis D, *et al.* Inhibition of the mammalian target of rapamycin (mTOR) in advanced pancreatic cancer: results of two phase II studies. *BMC Cancer* 2010, 10: 368
70. Vogelstein B, Lane D, Levine AJ. Surfing the p53 network. *Nature* 2000, 408: 307–310
71. David CJ, Massagué J. Contextual determinants of TGF β action in development, immunity and cancer. *Nat Rev Mol Cell Biol* 2018, 19: 419–435
72. Cao D, Ashfaq R, Goggins MG, Hruban RH, Kern SE, Iacobuzio-Donahue CA. Differential expression of multiple genes in association with MADH4/DPC4/SMAD4 inactivation in pancreatic cancer. *Int J Clin Exp Pathol* 2008, 1: 510–517
73. Dupont S, Zacchigna L, Cordenonsi M, Soligo S, Adorno M, Rugge M, Piccolo S. Germ-layer specification and control of cell growth by Ectodermin, a Smad4 ubiquitin ligase. *Cell* 2005, 121: 87–99
74. Schutte M, Hruban RH, Geradts J, Maynard R, Hilgers W, Rabindran SK, Moskaluk CA, *et al.* Abrogation of the Rb/p16 tumor-suppressive pathway in virtually all pancreatic carcinomas. *Cancer Res* 1997, 57: 3126–3130
75. Hosoda W, Chianchiano P, Griffin JF, Pittman ME, Brosens LA, Noë M, Yu J, *et al.* Genetic analyses of isolated high-grade pancreatic intraepithelial neoplasia (HG-PanIN) reveal paucity of alterations in TP53 and SMAD4. *J Pathol* 2017, 242: 16–23
76. Makohon-Moore AP, Matsukuma K, Zhang M, Reiter JG, Gerold JM, Jiao Y, Sikkema L, *et al.* Precancerous neoplastic cells can move through the pancreatic ductal system. *Nature* 2018, 561: 201–205
77. Ryan DP, Hong TS, Bardeesy N. Pancreatic adenocarcinoma. *N Engl J Med* 2014, 371: 1039–1049
78. Collisson EA, Bailey P, Chang DK, Biankin AV. Molecular subtypes of pancreatic cancer. *Nat Rev Gastroenterol Hepatol* 2019, 16: 207–220
79. Beloribi-Djefafli S, Vasseur S, Guillaumond F. Lipid metabolic reprogramming in cancer cells. *Oncogenesis* 2016, 5: e189
80. Koundouros N, Poulgiannis G. Reprogramming of fatty acid metabolism in cancer. *Br J Cancer* 2020, 122: 4–22
81. Cheng C, Geng F, Cheng X, Guo D. Lipid metabolism reprogramming and its potential targets in cancer. *Cancer Commun* 2018, 38: 27
82. Zaidi N, Swinnen JV, Smans K. ATP-citrate lyase: a key player in cancer metabolism. *Cancer Res* 2012, 72: 3709–3714
83. Batchuluun B, Pinkosky SL, Steinberg GR. Lipogenesis inhibitors: therapeutic opportunities and challenges. *Nat Rev Drug Discov* 2022, 21: 283–305
84. Zong H, Zhang Y, You Y, Cai T, Wang Y. RETRACTED ARTICLE: Decreased Warburg effect induced by ATP citrate lyase suppression inhibits tumor growth in pancreatic cancer. *Med Oncol* 2015, 32: 85
85. Xin M, Qiao Z, Li J, Liu J, Song S, Zhao X, Miao P, *et al.* miR-22 inhibits tumor growth and metastasis by targeting ATP citrate lyase: evidence in osteosarcoma, prostate cancer, cervical cancer and lung cancer. *Oncotarget* 2016, 7: 44252–44265
86. Csanadi A, Kayser C, Donauer M, Gump V, Aumann K, Rawluk J, Prasse A, *et al.* Prognostic value of malic enzyme and ATP-Citrate lyase in Non-Small cell lung cancer of the young and the elderly. *PLoS ONE* 2015, 10: e0126357
87. Wen J, Min X, Shen M, Hua Q, Han Y, Zhao L, Liu L, *et al.* ACLY facilitates colon cancer cell metastasis by CTNNB1. *J Exp Clin Cancer Res* 2019, 38: 401
88. Rios Garcia M, Steinbauer B, Srivastava K, Singhal M, Mattijssen F, Maida A, Christian S, *et al.* Acetyl-CoA carboxylase 1-dependent protein acetylation controls breast cancer metastasis and recurrence. *Cell Metab* 2017, 26: 842–855.e5
89. Wu X, Huang T. Recent development in acetyl-CoA carboxylase inhibitors and their potential as novel drugs. *Future Medicinal Chem* 2020, 12: 533–561
90. Petrova E, Scholz A, Paul J, Sturz A, Haike K, Siegel F, Mumberg D, *et al.* Acetyl-CoA carboxylase inhibitors attenuate WNT and Hedgehog signaling and suppress pancreatic tumor growth. *Oncotarget* 2017, 8: 48660–48670
91. Nishi K, Suzuki M, Yamamoto N, Matsumoto A, Iwase Y, Yamasaki K, Otagiri M, *et al.* Glutamine deprivation enhances Acetyl-CoA carboxylase inhibitor-induced death of human pancreatic cancer cells. *Anticancer Res* 2018, 38: 6683–6689
92. Yang Y, Liu H, Li Z, Zhao Z, Yip-Schneider M, Fan Q, Schmidt CM, *et al.* Role of fatty acid synthase in gemcitabine and radiation resistance of pancreatic cancers. *Int J Biochem Mol Biol* 2011, 2: 89–98
93. Tian S, Li P, Sheng S, Jin X. Upregulation of pyruvate kinase M2 expression by fatty acid synthase contributes to gemcitabine resistance in pancreatic cancer. *Oncol Lett* 2018, 15: 2211–2217
94. Walter K, Hong SM, Nyhan S, Canto M, Fedarko N, Klein A, Griffith M, *et al.* Serum fatty acid synthase as a marker of pancreatic neoplasia. *Cancer Epidemiol Biomarkers Prevention* 2009, 18: 2380–2385
95. Fazli HR, Moradzadeh M, Mehrbakhsh Z, Sharafkhan M, Masoudi S, Pourshams A, Mohamadkhani A. Diagnostic significance of serum fatty acid synthase in patients with pancreatic cancer. *Middle East J Dig Dis* 2021, 13: 115–120
96. Xiong Y, Liu Z, Zhao X, Ruan S, Zhang X, Wang S, Huang T. CPT1A regulates breast cancer-associated lymphangiogenesis via VEGF signaling. *Biomed Pharmacother* 2018, 106: 1–7
97. De Oliveira MP, Liesa M. The role of mitochondrial fat oxidation in cancer cell proliferation and survival. *Cells* 2020, 9: 2600
98. Luo J, Hong Y, Tao X, Wei X, Zhang L, Li Q. An indispensable role of

- CPT-1a to survive cancer cells during energy stress through rewiring cancer metabolism. *Tumour Biol* 2016, Oct 13. doi: 10.1007/s13277-016-5382-6
99. Park JK, Coffey NJ, Limoges A, Le A. The heterogeneity of lipid metabolism in cancer. *Adv Exp Med Biol* 2018, 1063: 33–55
 100. Carracedo A, Cantley LC, Pandolfi PP. Cancer metabolism: fatty acid oxidation in the limelight. *Nat Rev Cancer* 2013, 13: 227–232
 101. Tan Z, Xiao L, Tang M, Bai F, Li J, Li L, Shi F, *et al.* Targeting CPT1A-mediated fatty acid oxidation sensitizes nasopharyngeal carcinoma to radiation therapy. *Theranostics* 2018, 8: 2329–2347
 102. Beloribi-Djefallia S, Vasseur S, Guillaumond F. Lipid metabolic reprogramming in cancer cells. *Oncogenesis* 2016, 5: e189
 103. Patra KC, Kato Y, Mizukami Y, Widholz S, Boukhali M, Revenco I, Grossman EA, *et al.* Mutant GNAS drives pancreatic tumorigenesis by inducing PKA-mediated SIK suppression and reprogramming lipid metabolism. *Nat Cell Biol* 2018, 20: 811–822
 104. Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. *Nat Rev Mol Cell Biol* 2009, 10: 307–318
 105. Laplante M, Sabatini DM. An emerging role of mTOR in lipid biosynthesis. *Curr Biol* 2009, 19: R1046–R1052
 106. Bengoechea-Alonso MT, Ericsson J. SREBP in signal transduction: cholesterol metabolism and beyond. *Curr Opin Cell Biol* 2007, 19: 215–222
 107. Brown MS, Goldstein JL. The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell* 1997, 89: 331–340
 108. Linkermann A, Skouta R, Himmerkus N, Mulay SR, Dewitz C, De Zen F, Prokai A, *et al.* Synchronized renal tubular cell death involves ferroptosis. *Proc Natl Acad Sci USA* 2014, 111: 16836–16841
 109. Yang WS, Stockwell BR. Ferroptosis: death by lipid peroxidation. *Trends Cell Biol* 2016, 26: 165–176
 110. Ye Z, Zhuo Q, Hu Q, Xu X, Mengqi liu X, Zhang Z, Xu W, *et al.* FBW7-NRA41-SCD1 axis synchronously regulates apoptosis and ferroptosis in pancreatic cancer cells. *Redox Biol* 2021, 38: 101807
 111. Gao J, Zhang Z, Liu Y, Zhang Z, Wang M, Gong A, Xia L, *et al.* Stearoyl-CoA desaturase 1 potentiates hypoxic plus nutrient-deprived pancreatic cancer cell ferroptosis resistance. *Oxid Med Cell Longev* 2021, 2021: 6629804
 112. Yang J, Xu J, Zhang B, Tan Z, Meng Q, Hua J, Liu J, *et al.* Ferroptosis: at the crossroad of gemcitabine resistance and tumorigenesis in pancreatic cancer. *Int J Mol Sci* 2021, 22: 10944
 113. Sun Y, He W, Luo M, Zhou Y, Chang G, Ren W, Wu K, *et al.* SREBP1 regulates tumorigenesis and prognosis of pancreatic cancer through targeting lipid metabolism. *Tumor Biol* 2015, 36: 4133–4141
 114. Skrypek K, Balog S, Eriguchi Y, Asahina K. Inhibition of stearyl-CoA desaturase induces the unfolded protein response in pancreatic tumors and suppresses their growth. *Pancreas* 2021, 50: 219–226
 115. Zhang D, Lu P, Zhu K, Wu H, Dai Y. TFPC2 overcomes senescence by cooperating with SREBP2 to activate cholesterol synthesis in pancreatic cancer. *Front Oncol* 2021, 11: 724437
 116. Guillaumond F, Bidaut G, Ouaisi M, Servais S, Gouirand V, Olivares O, Lac S, *et al.* Cholesterol uptake disruption, in association with chemotherapy, is a promising combined metabolic therapy for pancreatic adenocarcinoma. *Proc Natl Acad Sci USA* 2015, 112: 2473–2478
 117. Chen WCY, Boursi B, Mamtani R, Yang YX. Total serum cholesterol and pancreatic cancer: a nested case-control study. *Cancer Epidemiol Biomarkers Prevention* 2019, 28: 363–369
 118. Alexander JI, Martinez E, Vargas A, Zinshteyn D, Sodi V, Connolly DC, Hartman TR, *et al.* Cholesterol and CDON regulate sonic hedgehog release from pancreatic cancer cells. *J Pancreatic Cancer* 2021, 7: 39–47
 119. Jung YY, Ko J, Um J, Chinnathambi A, Alharbi SA, Sethi G, Ahn KS. LDL cholesterol promotes the proliferation of prostate and pancreatic cancer cells by activating the STAT3 pathway. *J Cell Physiol* 2021, 236: 5253–5264
 120. Kazi DS, Penko JM, Bibbins-Domingo K. Statins for primary prevention of cardiovascular disease. *Med Clin N Am* 2017, 101: 689–699
 121. Vallianou N, Kostantinou A, Kougias M, Kazazis C. Statins and cancer. *Anticancer Agents Med Chem* 2014, 14: 706–712
 122. Gong J, Sachdev E, Robbins LA, Lin E, Hendifar AE, Mita MM. Statins and pancreatic cancer. *Oncol Lett* 2017, 13: 1035–1040
 123. Mistafa O, Stenius U. Statins inhibit Akt/PKB signaling via P2X7 receptor in pancreatic cancer cells. *Biochem Pharmacol* 2009, 78: 1115–1126
 124. Padanad MS, Konstantinidou G, Venkateswaran N, Melegari M, Rindhe S, Mitsche M, Yang C, *et al.* Fatty acid oxidation mediated by Acyl-CoA synthetase long chain 3 is required for mutant KRAS lung tumorigenesis. *Cell Rep* 2016, 16: 1614–1628
 125. Rozeveldt CN, Johnson KM, Zhang L, Razidlo GL. KRAS controls pancreatic cancer cell lipid metabolism and invasive potential through the lipase HSL. *Cancer Res* 2020, 80: 4932–4945
 126. Dasgupta A, Shukla SK, Gunda V, King RJ, Singh PK. Evaluating the metabolic alterations in pancreatic cancer. *Methods Mol Biol* 2019, 1882: 221–228
 127. Shu X, Zheng W, Yu D, Li HL, Lan Q, Yang G, Cai H, *et al.* Prospective metabolomics study identifies potential novel blood metabolites associated with pancreatic cancer risk. *Int J Cancer* 2018, 143: 2161–2167
 128. Tao L, Zhou J, Yuan C, Zhang L, Li D, Si D, Xiu D, *et al.* Metabolomics identifies serum and exosomes metabolite markers of pancreatic cancer. *Metabolomics* 2019, 15: 86
 129. Lee JH, Yu SE, Kim KH, Yu MH, Jeong IH, Cho JY, Park SJ, *et al.* Individualized metabolic profiling stratifies pancreatic and biliary tract cancer: a useful tool for innovative screening programs and predictive strategies in healthcare. *EPMA J* 2018, 9: 287–297
 130. Fahrman JF, Bantis LE, Capello M, Scelo G, Dennison JB, Patel N, Murage E, *et al.* A plasma-derived protein-metabolite multiplexed panel for early-stage pancreatic cancer. *J Natl Cancer Institute* 2019, 111: 372–379
 131. Zhang X, Shi X, Lu X, Li Y, Zhan C, Akhtar ML, Yang L, *et al.* Novel metabolomics serum biomarkers for pancreatic ductal adenocarcinoma by the comparison of pre-, postoperative and normal samples. *J Cancer* 2020, 11: 4641–4651
 132. Xiong Y, Shi C, Zhong F, Liu X, Yang P. LC-MS/MS and SWATH based serum metabolomics enables biomarker discovery in pancreatic cancer. *Clinica Chim Acta* 2020, 506: 214–221
 133. Luo X, Liu J, Wang H, Lu H. Metabolomics identified new biomarkers for the precise diagnosis of pancreatic cancer and associated tissue metastasis. *Pharmacol Res* 2020, 156: 104805
 134. Maman S, Witz IP. A history of exploring cancer in context. *Nat Rev Cancer* 2018, 18: 359–376
 135. Anderson NM, Simon MC. The tumor microenvironment. *Curr Biol* 2020, 30: R921–R925
 136. Hao Y, Li D, Xu Y, Ouyang J, Wang Y, Zhang Y, Li B, *et al.* Investigation of lipid metabolism dysregulation and the effects on immune micro-environments in pan-cancer using multiple omics data. *BMC Bioinformatics* 2019, 20: 195
 137. Dougan SK. The pancreatic cancer microenvironment. *Cancer J* 2017, 23: 321–325
 138. Liao Q, Zhou Y, Xia L, Cao D. Lipid metabolism and immune checkpoints. *Adv Exp Med Biol* 2021, 1316: 191–211
 139. Li TJ, Wang WQ, Yu XJ, Liu L. Killing the “BAD”: challenges for

- immunotherapy in pancreatic cancer. *Biochim Biophys Acta Rev Cancer* 2020, 1874: 188384
140. Tao J, Yang G, Zhou W, Qiu J, Chen G, Luo W, Zhao F, *et al.* Targeting hypoxic tumor microenvironment in pancreatic cancer. *J Hematol Oncol* 2021, 14: 14
 141. Netea-Maier RT, Smit JWA, Netea MG. Metabolic changes in tumor cells and tumor-associated macrophages: a mutual relationship. *Cancer Lett* 2018, 413: 102–109
 142. Liu X, Hartman CL, Li L, Albert CJ, Si F, Gao A, Huang L, *et al.* Reprogramming lipid metabolism prevents effector T-cell senescence and enhances tumor immunotherapy. *Sci Transl Med* 2021, 13: eaaz6314
 143. Kaymak I, Williams KS, Cantor JR, Jones RG. Immunometabolic interplay in the tumor microenvironment. *Cancer Cell* 2021, 39: 28–37
 144. Corn KC, Windham MKA, Rafat M. Lipids in the tumor microenvironment: From cancer progression to treatment. *Prog Lipid Res* 2020, 80: 101055
 145. Leone RD, Powell JD. Metabolism of immune cells in cancer. *Nat Rev Cancer* 2020, 20: 516–531
 146. Ding X, Zhao T, Lee CC, Yan C, Du H. Lysosomal acid lipase deficiency controls T- and B-Regulatory cell homeostasis in the lymph nodes of mice with human cancer xenotransplants. *Am J Pathol* 2021, 191: 353–367
 147. Zhang J, Song Y, Shi Q, Fu L. Research progress on FASN and MGLL in the regulation of abnormal lipid metabolism and the relationship between tumor invasion and metastasis. *Front Med* 2021, 15: 649–656
 148. Berod L, Friedrich C, Nandan A, Freitag J, Hagemann S, Harmrolfs K, Sandouk A, *et al.* De novo fatty acid synthesis controls the fate between regulatory T and T helper 17 cells. *Nat Med* 2014, 20: 1327–1333
 149. Subramanian M, Marelli-Berg FM. CD36 pumps fat to defang killer T cells in tumors. *Cell Metab* 2021, 33: 1509–1511
 150. Ma X, Xiao L, Liu L, Ye L, Su P, Bi E, Wang Q, *et al.* CD36-mediated ferroptosis dampens intratumoral CD8⁺ T cell effector function and impairs their antitumor ability. *Cell Metab* 2021, 33: 1001–1012.e5
 151. Chen YJ, Liao WX, Huang SZ, Yu YF, Wen JY, Chen J, Lin DG, *et al.* Prognostic and immunological role of CD36: pan-cancer analysis. *J Cancer* 2021, 12: 4762–4773
 152. Malandrino MI, Fucho R, Weber M, Calderon-Dominguez M, Mir JF, Valcarcel L, Escoté X, *et al.* Enhanced fatty acid oxidation in adipocytes and macrophages reduces lipid-induced triglyceride accumulation and inflammation. *Am J Physiol Endocrinol Metab* 2015, 308: E756–E769
 153. Mikalayeva V, Cesleviciene I, Sarapiniene I, Zvikas V, Skeberdis VA, Jakstas V, Bordel S. Fatty acid synthesis and degradation interplay to regulate the oxidative stress in cancer cells. *Int J Mol Sci* 2019, 20: 1384
 154. Zhou J, Jiang Y, Huang Y, Wang Q, Kaifi JT, Kimchi ET, Chabu CY, *et al.* Single-cell RNA sequencing to characterize the response of pancreatic cancer to anti-PD-1 immunotherapy. *Transl Oncol* 2022, 15: 101262
 155. Li W, Song X, Yu H, Zhang M, Li F, Cao C, Jiang Q. Dendritic cell-based cancer immunotherapy for pancreatic cancer. *Arab J Gastroenterol* 2018, 19: 1–6
 156. Ajina R, Weiner LM. T-cell immunity in pancreatic cancer. *Pancreas* 2020, 49: 1014–1023
 157. Fincham REA, Delvecchio FR, Goulart MR, Yeong JPS, Kocher HM. Natural killer cells in pancreatic cancer stroma. *World J Gastroenterol* 2021, 27: 3483–3501
 158. Huang H, Wang Z, Zhang Y, Pradhan RN, Ganguly D, Chandra R, Murimwa G, *et al.* Mesothelial cell-derived antigen-presenting cancer-associated fibroblasts induce expansion of regulatory T cells in pancreatic cancer. *Cancer Cell* 2022, 40: 656–673
 159. Ye H, Zhou Q, Zheng S, Li G, Lin Q, Wei L, Fu Z, *et al.* Tumor-associated macrophages promote progression and the Warburg effect via CCL18/NF- κ B/VCAM-1 pathway in pancreatic ductal adenocarcinoma. *Cell Death Dis* 2018, 9: 453
 160. Bellone G, Carbone A, Smirne C, Scirelli T, Buffolino A, Novarino A, Stacchini A, *et al.* Cooperative induction of a tolerogenic dendritic cell phenotype by cytokines secreted by pancreatic carcinoma cells. *J Immunol* 2006, 177: 3448–3460
 161. Evans A, Costello E. The role of inflammatory cells in fostering pancreatic cancer cell growth and invasion. *Front Physiol* 2012, 3: 270
 162. Kim CH. FOXP3 and its role in the immune system. *Adv Exp Med Biol* 2009, 665: 17–29
 163. Hu L, Zhu M, Shen Y, Zhong Z, Wu B. The prognostic value of intratumoral and peritumoral tumor-infiltrating FoxP3⁺ Treg cells in of pancreatic adenocarcinoma: a meta-analysis. *World J Surg Onc* 2021, 19: 300
 164. Nolz JC. Molecular mechanisms of CD8⁺ T cell trafficking and localization. *Cell Mol Life Sci* 2015, 72: 2461–2473
 165. Maimela NR, Liu S, Zhang Y. Fates of CD8⁺ T cells in Tumor Microenvironment. *Comput Struct Biotechnol J* 2019, 17: 1–13
 166. Hui L, Chen Y. Tumor microenvironment: sanctuary of the devil. *Cancer Lett* 2015, 368: 7–13
 167. Takahashi R, Macchini M, Sunagawa M, Jiang Z, Tanaka T, Valenti G, Renz BW, *et al.* Interleukin-1beta-induced pancreatitis promotes pancreatic ductal adenocarcinoma via B lymphocyte-mediated immune suppression. *Gut* 2021, 70: 330–341
 168. Philip PA. Targeting B cells in pancreatic adenocarcinoma: does RESOLVE resolve the question? *Ann Oncol* 2021, 32: 582–583
 169. Davies LC, Taylor PR. Tissue-resident macrophages: then and now. *Immunology* 2015, 144: 541–548
 170. Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 2012, 122: 787–795
 171. Mills CD, Kincaid K, Alt JM, Heilman MJ, Hill AM. M-1/M-2 macrophages and the Th1/Th2 paradigm. *J Immunol* 2000, 164: 6166–6173
 172. Geeraerts X, Bolli E, Fendt SM, Van Ginderachter JA. Macrophage metabolism as therapeutic target for cancer, atherosclerosis, and obesity. *Front Immunol* 2017, 8: 289
 173. Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaili S, Mardani F, Seifi B, *et al.* Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol* 2018, 233: 6425–6440
 174. Vogel DYS, Glim JE, Stavenuiter AWD, Breur M, Heijnen P, Amor S, Dijkstra CD, *et al.* Human macrophage polarization in vitro: maturation and activation methods compared. *Immunobiology* 2014, 219: 695–703
 175. Van Ginderachter JA, Movahedi K, Hassanzadeh Ghassabeh G, Meerschaut S, Beschin A, Raes G, De Baetselier P. Classical and alternative activation of mononuclear phagocytes: picking the best of both worlds for tumor promotion. *Immunobiology* 2006, 211: 487–501
 176. Mitchem JB, Brennan DJ, Knolhoff BL, Belt BA, Zhu Y, Sanford DE, Belaygorod L, *et al.* Targeting tumor-infiltrating macrophages decreases tumor-initiating cells, relieves immunosuppression, and improves chemotherapeutic responses. *Cancer Res* 2013, 73: 1128–1141
 177. Sanford DE, Belt BA, Panni RZ, Mayer A, Deshpande AD, Carpenter D, Mitchem JB, *et al.* Inflammatory monocyte mobilization decreases patient survival in pancreatic cancer: a role for targeting the CCL2/CCR2 axis. *Clin Cancer Res* 2013, 19: 3404–3415
 178. Jang JE, Hajdu CH, Liot C, Miller G, Dustin ML, Bar-Sagi D. Crosstalk between regulatory T cells and tumor-associated dendritic cells negates anti-tumor immunity in pancreatic cancer. *Cell Rep* 2017, 20: 558–571

179. Health Commission of PRC N. Chinese guidelines for diagnosis and treatment of gastric cancer 2018 (English version). *Chin J Cancer Res* 2019, 31: 707–737
180. Balaban EP, Mangu PB, Khorana AA, Shah MA, Mukherjee S, Crane CH, Javle MM, *et al.* Locally advanced, unresectable pancreatic cancer: american society of clinical oncology clinical practice guideline. *J Clin Oncol* 2016, 34: 2654–2668
181. Springfield C, Jäger D, Büchler MW, Strobel O, Hackert T, Palmer DH, Neoptolemos JP. Chemotherapy for pancreatic cancer. *La Presse Médicale* 2019, 48: e159–e174
182. Catenacci DVT, Junttila MR, Karrison T, Bahary N, Horiba MN, Nattam SR, Marsh R, *et al.* Randomized phase Ib/II study of gemcitabine plus placebo or vismodegib, a hedgehog pathway inhibitor, in patients with metastatic pancreatic cancer. *J Clin Oncol* 2015, 33: 4284–4292
183. Binenbaum Y, Na'ara S, Gil Z. Gemcitabine resistance in pancreatic ductal adenocarcinoma. *Drug Resistance Updates* 2015, 23: 55–68
184. Fujimura Y, Ikenaga N, Ohuchida K, Setoyama D, Irie M, Miura D, Wariishi H, *et al.* Mass spectrometry-based metabolic profiling of gemcitabine-sensitive and gemcitabine-resistant pancreatic cancer cells. *Pancreas* 2014, 43: 311–318
185. Amrutkar M, Vethe NT, Verbeke CS, Aasrum M, Finstadsveen AV, Santha P, Gladhaug IP, *et al.* Differential gemcitabine sensitivity in primary human pancreatic cancer cells and paired stellate cells is driven by heterogeneous drug uptake and processing. *Cancers (Basel)* 2020, 12: 3628
186. Zhao H, Duan Q, Zhang Z, Li H, Wu H, Shen Q, Wang C, *et al.* Up-regulation of glycolysis promotes the stemness and EMT phenotypes in gemcitabine-resistant pancreatic cancer cells. *J Cell Mol Med* 2017, 21: 2055–2067
187. Tadros S, Shukla SK, King RJ, Gunda V, Vernucci E, Abrego J, Chaika NV, *et al.* De Novo lipid synthesis facilitates gemcitabine resistance through endoplasmic reticulum stress in pancreatic cancer. *Cancer Res* 2017, 77: 5503–5517
188. Al-Zoubi M, Chipitsyna G, Saxena S, Sarosiek K, Gandhi A, Kang CY, Relles D, *et al.* Overexpressing TNF-Alpha in pancreatic ductal adenocarcinoma cells and fibroblasts modifies cell survival and reduces fatty acid synthesis via downregulation of sterol regulatory element binding protein-1 and activation of acetyl CoA carboxylase. *J Gastrointest Surg* 2014, 18: 257–268
189. Gouw AM, Eberlin LS, Margulis K, Sullivan DK, Toal GG, Tong L, Zare RN, *et al.* Oncogene KRAS activates fatty acid synthase, resulting in specific ERK and lipid signatures associated with lung adenocarcinoma. *Proc Natl Acad Sci USA* 2017, 114: 4300–4305
190. Wang B, Shen C, Li Y, Zhang T, Huang H, Ren J, Hu Z, *et al.* Oridonin overcomes the gemcitabine resistant PANC-1/Gem cells by regulating GST pi and LRP/1 ERK/JNK signalling. *Onco Targets Ther* 2019, 12: 5751–5765
191. Hong JY, Nam EM, Lee J, Park JO, Lee SC, Song SY, Choi SH, *et al.* Randomized double-blinded, placebo-controlled phase II trial of simvastatin and gemcitabine in advanced pancreatic cancer patients. *Cancer Chemother Pharmacol* 2014, 73: 125–130