

Physical Activity as an Adjunct Treatment for People Living with HIV?

Abstract: *This review evaluates physical activity as a candidate for an adjunct treatment, in conjunction with antiretroviral therapy (ART), for people living with HIV (PLWH). Evidence is summarized that chronic, non-resolving inflammation (a principal feature of immune system dysfunction) and a dysfunctional state of the gut environment are key factors in HIV infection that persist despite treatment with ART. In addition, evidence is summarized that regular physical activity may restore normal function of both the immune system and the gut environment and may thereby ameliorate symptoms and non-resolving inflammation-associated comorbidities that burden PLWH. Physicians who care for PLWH could thus consider incorporating physical activity into treatment plans to complement ART. It is also discussed that different types of physical activity can have different effects on the gut environment and immune function, and that future research should establish more specific criteria for the design of exercise regimens tailored to PLWH.*



Keywords: immunodeficiency; exercise intensity; microbiome; gut barrier; pro-inflammatory; recovery

Introduction

Current treatment of infection with human immunodeficiency virus (HIV) focuses on countering the destruction of human immune cells and associated weakening of immune defenses.¹ Such intervention aims to prevent

constitutes a failure to terminate pro-inflammatory signaling.^{6,7} Such non-resolving inflammation is a root cause of multiple diseases and disorders that can occur as comorbidities in PLWH.⁸⁻¹³

This review presents an overview of HIV effects that are successfully

 “Incorporating physical activity into treatment plans may ameliorate lingering effects of HIV” 

progression to stage 3 HIV, or acquired immunodeficiency syndrome (AIDS).²⁻⁴ At the same time, people living with HIV (PLWH) exhibit systemic, low-grade immune system activation, a hallmark of immune system dysfunction, that persists even when viral replication is effectively suppressed⁵ by antiretroviral therapy (ART). This prolonged immune system activation is commonly referred to as *chronic inflammation*, but the term *non-resolving inflammation* has recently been introduced to emphasize that this persistent immune response primarily

addressed by ART, as well as effects that persist in ART-treated PLWH. These lingering effects are aspects of immune system dysfunction, and can be linked to a persistent disruption of the gut environment. Since ART-treated PLWH experience such dysfunction, it is necessary and relevant in modern HIV treatment to understand how to restore a functional immune system and gut environment. Because dysfunction of these systems is implicated in chronic illnesses that disproportionately burden PLWH, ameliorating non-resolving inflammation should be a priority for

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physicians aiming to improve the quality of life of their patients with HIV.

Here, we review the effect of physical activity on immune system function and the gut environment. Attention is given to various types of physical activity and their apparent different impacts on the gut environment. Based on review of the available evidence, a proposal is made that certain types of physical activity may be candidates for adjunct treatment, complementary to ART, that may reconstitute the gut environment and restore normal immune system function (eliminate non-resolving inflammation) in PLWH. We propose that regular, voluntary (interest-based) physical activity with adequate periods of recovery is most likely to have a restorative effect on the gut environment and immune system function and is therefore an attractive candidate for an additional treatment complementary to ART in PLWH. Incorporating physical activity into treatment plans may ameliorate lingering effects of HIV that are not addressed by ART and enhance the quality of life of PLWH.

HIV Effects and Antiretroviral Therapy

Viruses like HIV interact with the immune system in complex ways; they use components of the immune system for their own replication, and at the same time precipitate a dysfunctional state of the immune system as a whole. The retrovirus HIV enters immune cells (especially CD4⁺ T cells) and employs their molecular vehicles for its own replication.¹⁴ To do this, HIV activates gene regulators that trigger replication, such as the transcription factor NF-κB.^{15,16} The latter is a key regulator of the immune response and orchestrates inflammatory signaling.^{17,18} Constitutive, low-grade activation of NF-κB is associated with non-resolving

inflammation in HIV-infected individuals.^{15,19} Furthermore, the take-over of CD4⁺ T cells by HIV eventually results in destruction of immune cells via programmed cell death—mainly as triggered by the human host's immune system in an attempt to eliminate the virus.⁵ Doitsh and Greene state that, during HIV infection, *“most cells are not dying because of a toxic action of products encoded by HIV. Rather, death occurs as a consequence of a powerful defensive innate immune response launched by the host against the virus leading to a cellular form of suicide rather than virological murder.”*⁵ This attack by the immune system reverberates system-wide as non-resolving inflammation. The attack on immune cells involves reactive oxygen species (ROS) that function in the programmed cell death of CD4⁺ T cells²⁰ and also serve as signals that trigger a system-wide, snow-balling, and continuous mobilization of the immune system,²¹ that is, non-resolving inflammation. This non-resolving inflammation involves systemic disruption of redox homeostasis (balance between oxidants and antioxidants) and the gut environment (see next section), and plays a key role in cardiovascular disease, obesity, GI distress, cancer, mental illness and many other conditions.²²⁻²⁷ The recognition of the role of non-resolving inflammation as a root cause for disease has been called *“one of the most important scientific discoveries in health research in recent years.”*^{28,29} Notably, PLWH exhibit a higher incidence of many of these and other comorbidities, which emphasizes the critical importance of addressing non-resolving inflammation in this population.

Another key component of the human immune system that interacts with HIV is the interferon system that detects pathogens and triggers defenses. The interferon system

exhibits low activity in CD4⁺ cells³⁰ and is further suppressed by HIV infection.³¹ Such impaired interferon activation in CD4⁺ cells allows sustained viral replication in these cells. Subsequent activation of interferon in other cells—once HIV infection is established—fails to eradicate the virus and may even promote non-resolving inflammation and further deterioration of host health.³⁰ As stated above, all aspects of immune-system activation involve ROS production that has the potential to snow-ball into non-resolving inflammation under exacerbating conditions, such as a dysfunctional gut environment (see section below). However, additional research is needed to elucidate the complex interactions between HIV and the interferon system.³²

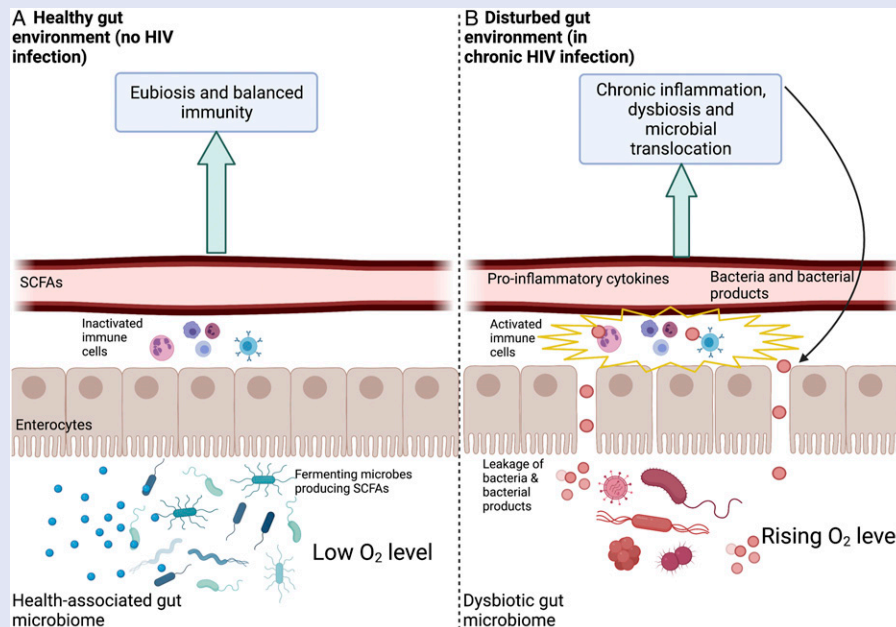
Current HIV treatment based on ART arrests the replication cycle of HIV and has been shown to increase immune cell numbers in most PLWH.³³⁻³⁵ However, some PLWH are “immunological nonresponders” who do respond to ART with cessation of viral replication, but fail to exhibit restoration of CD4⁺ cell counts.³⁶⁻³⁸ More commonly yet, PLWH who receive regular ART treatment that halts viral replication typically show varying levels of persistent non-resolving inflammation.^{8,39-41} Addressing immune-system dysfunction in the context of HIV infection is thus critical even in the era of ART.

HIV and the Gut-Immune Link

Additional mechanistic insight into the repercussions of non-resolving inflammation induced by HIV focuses on the gastrointestinal tract and the gut microbiome.⁴²⁻⁴⁶ Figure 1 depicts the gut environment in its functional state without HIV infection (Figure 1A) as well as in its dysfunctional state during chronic HIV infection (Figure 1B).

Figure 1.

Schematic depiction of a healthy gut environment **(A)** and a gut environment **(B)** that is disturbed as the result of chronic HIV infection. Depicted are the gut content (bottom; fermenting microbes producing short-chain saturated fatty acids, SCFAs, especially in (A)), the cells forming the gut lining (enterocytes), immune cells (between enterocytes and blood vessels) that are either inactivated or activated, and a blood vessel that receives either SCFAs or pro-inflammatory cytokines, bacteria and bacterial products from the gut. Created with [BioRender.com](https://www.biorender.com/).



The Gut-Immune Link in Health

Figure 1A depicts a functional, health-promoting (eubiotic) gut microbiome characterized by prominent presence of anaerobic, fermenting bacteria that thrive in low-oxygen environments.⁴⁷ These microbes produce short-chain fatty acids (SCFAs) that serve as an energy source for the cells (enterocytes) lining the gut⁴⁸⁻⁵⁰ (for details, see below) and can also diffuse into the bloodstream.^{51,52} Locally, these SCFAs have essential roles in supporting gut-barrier integrity.^{53,54} For example, they act as gene activators for the production of proteins that maintain gut-barrier integrity.⁵⁴⁻⁵⁶ An example of a key SCFA is butyrate (produced by fermenting gut microbes) that increases expression of the tight-junction protein Claudin-1 involved

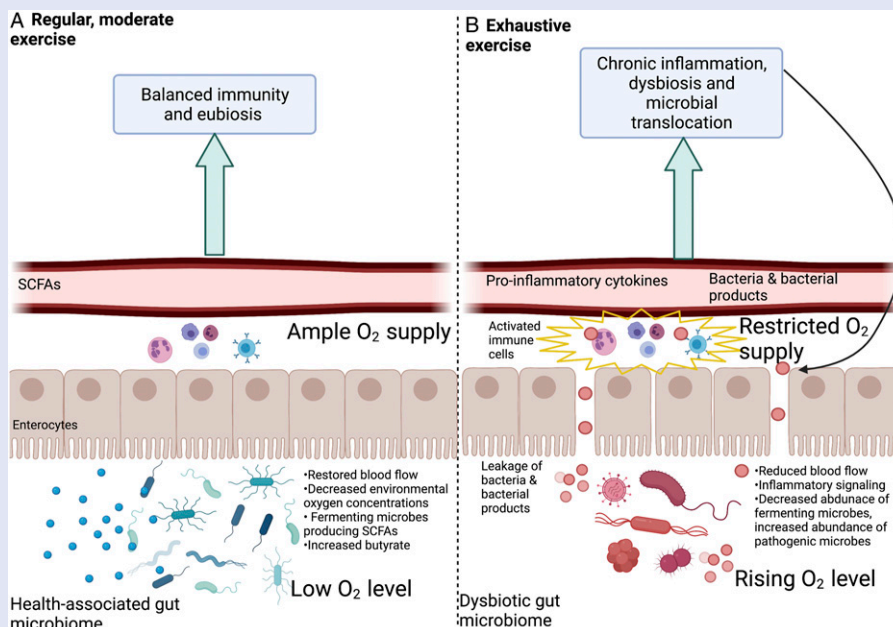
in supporting gut-barrier integrity.⁵⁷ In addition, butyrate also represses a protein (Claudin-2)⁵⁸ that has been suggested to increase barrier permeability.⁵⁹ Gut-barrier integrity supports beneficial gut microbes and these gut microbes, in turn, maintain gut-barrier integrity.⁴⁷

A healthy gut is characterized by a steep oxygen gradient between the (hypoxic) gut lumen and the distal zone of the intestinal epithelium.^{60,61} The enterocytes of the intestinal epithelium require oxygen for their oxidative metabolism that supports their roles in barrier formation,⁵⁵ nutrient absorption⁶² and immunity.^{61,63,64} Enterocytes use butyrate as the fuel they burn with oxygen to produce ATP for these various functions.⁶¹ Therefore, butyrate produced by fermenting gut microbes stimulates oxygen uptake by enterocytes and helps maintain

a low oxygen environment within the gut.^{55,65} By requiring oxygen for their own metabolism, enterocytes thus contribute to maintaining hypoxic concentrations within the gut lumen,⁶¹ which supports anaerobic, SCFA-producing fermenting microbes.^{60,66} In addition to directly supporting gut-barrier integrity, SCFAs play a role in dampening the activity of NF- κ B and the inflammatory response.⁶⁷⁻⁷¹ In the state where gut microbial eubiosis and gut-barrier integrity are maintained, the immune cells associated with the gut lining are not activated to induce systemic effects^{72,73} (Figure 1A). Any condition that interferes with enterocyte function (eg, as seen during HIV infection) triggers production of messengers that initiate a pro-inflammatory cascade involving pro-inflammatory

Figure 2.

The effects of a regimen of regular moderate physical activity (**A**) compared with the effects of exhaustive exercise (**B**). Acclimation to regular moderate physical activity (A) restores (splanchnic) blood flow to the gut during physical activity, which supports maintenance of low oxygen concentrations within the gut lumen and the activity of fermenting, SCFA-producing microbes, strengthens the gut barrier and decreases inflammation. Continuous exhaustive exercise (B) continuously reduces blood flow to the gut, leading to enterocyte dysfunction, gut microbiome dysbiosis, gut-barrier impairment, microbial translocation out of the gut, and non-resolving inflammation. Created with [BioRender.com](https://www.biorender.com).



cytokines.^{74,75} Such adverse conditions include an insufficient supply of butyrate (as fuel) for enterocytes or of oxygen (to burn butyrate) as can be the case during exhaustive exercise (see [Figure 2B](#) in the next major section below).

The Gut-Immune Link as Affected by HIV

[Figure 1B](#) illustrates a feed-forward loop of loss of gut-barrier integrity and non-resolving inflammatory responses in PLWH. Early HIV infection is characterized by widespread destruction of CD4⁺ T cells present in gut mucosa tissue. While ART causes some recovery of these cells, this recovery is not always complete and mucosa impairment may persist after individuals receive treatment.⁷⁶⁻⁷⁸ Disruption of T cells in the gut

mucosa results in impaired gut-barrier integrity, which allows leakage of gut microbes and microbial products into the bloodstream ([Figure 1B](#)). Specifically, lipopolysaccharide (LPS), a cell surface component of gram-negative bacteria, is often identified as a marker of such microbial translocation and increased levels of LPS are observed during chronic HIV infection.^{79,80} Translocation of LPS activates the immune cells associated with the gut lining ([Figure 2B](#)) in a process mediated, for example, by NF-κB.^{81,82} Continuous activation of NF-κB contributes to a cascade of non-resolving immune system activation and may explain the persistent inflammatory state observed in PLWH.⁸³⁻⁸⁵

Furthermore, the disrupted gut barrier in chronic HIV infection ([Figure 1B](#)) results in increased oxygen concentrations within the gut, which shifts the gut microbiome to a disrupted state (dysbiosis) with specific losses in anaerobic fermenters and SCFA production. Increased concentrations of oxygen in the gut as a result of gut-barrier impairment also favor increases in gut bacteria that are facultative anaerobes capable of switching to aerobic (oxygen-dependent) cellular respiration and fast growth.⁸⁶⁻⁸⁸ These fast-growing facultative anaerobes can become pathogenic when undergoing such rapid growth.^{26,88,89} In addition to accelerating the cycle of leaky gut and non-resolving inflammation in PLWH shown in [Figure 1B](#), this shift in microbiome composition is

associated with GI distress and other symptoms.^{86,90}

Since non-resolving inflammation and a dysfunctional gut microbiome are implicated in several conditions that disproportionately burden both untreated and treated PLWH, including cardiovascular disease, chronic kidney disease, cancer, and depression, restoration of the gut environment and normal immune function must be a priority for improving the quality of life of PLWH. The next section examines the potential of regular, moderate physical activity, used as a therapy complementary to ART, to reduce non-resolving inflammation and restore gut barrier integrity.

Physical Activity and the Gut-Immune Link

Figure 2 summarizes effects of physical activity on the gut microbiome, gut-barrier integrity, and inflammation that have emerged from recent studies in various human populations (Figures 2A and 2B).

Intensity, duration, and frequency of exercise, as well as age and training status of individual participants, can all influence the effect of exercise on health.⁹¹⁻⁹⁵ More research is needed to elucidate the threshold of the transition between beneficial and detrimental exercise. A key consideration may be the effect of physical activity on blood flow to the gut, and the fact that acclimation (habituation) to a regular exercise regimen can restore this blood supply.

Effect of Regular Physical Activity With Replete Blood Flow to the Gut

It is well established that regular physical activity can reduce inflammation in various populations.⁹⁶⁻⁹⁸ In principle, physical activity has the potential to divert blood flow from the gut to

the working muscles.⁹⁹ However, during the establishment of a regular regimen of physical activity, a process of acclimation (habituation) takes place that restores blood supply to the gut during physical activity.⁷⁵ Such regular physical activity is associated with fully functional enterocytes and a high abundance of fermenting, butyrate-producing microbes^{75,100-102} (Figure 2A). While some gut microbes produce butyrate from sugars derived from fiber or complex sugars,^{103,104} other microbes convert lactate produced by working muscles to SCFAs like butyrate.^{105,106} The butyrate produced serves to strengthen the gut barrier through the above-described support for enterocytes and, for example, stimulated expression of proteins involved in barrier tightening and downregulation of proteins involved in barrier loosening. A strengthened barrier and associated microbiome eubiosis decrease leakage of microbial products into the bloodstream, and decrease activation of immune system regulators such as NF-κB and the resulting non-resolving inflammation¹⁰⁷⁻¹¹³ (Figure 2A). Suppression of NF-κB by physical activity was associated with reduced muscle loss in a mouse model¹¹⁴ and this effect should also be assessed in humans. Exercise-associated decreases in muscle loss could offer important relief for PLWH since PLWH are often burdened by loss of muscle mass and function.¹¹⁵

Effect of Physical Activity Associated With Continuous Low Blood Flow to the Gut

The type of exercise that increases the risk for non-resolving inflammation may be identified by its effect on the gut—as exercise that continuously exceeds the capacity for acclimatory restoration of blood flow to the gut at a level sufficient to

prevent hypoxia in enterocytes. As far as impacts on the gut environment are concerned, a regimen of continuously exhaustive exercise has similar effects to those seen during the initial transition phase from inactivity to an established regimen of regular, moderate physical activity.^{74,75} Both activities draw blood away from the gut—either continuously or only during a transition phase of acclimation. Whereas acclimation to regular physical activity can restore blood flow to GI cells,⁷⁵ this may not be the case for a regimen of continuously exhaustive exercise.¹¹⁶ Insufficient oxygen supply to the enterocytes is as detrimental as hypoxia within the gut lumen is beneficial. The ability to maintain sufficient blood flow to the gut during exercise is critical to supporting gut barrier integrity and gut microbiome eubiosis^{74,75,117,118} (Figure 2A). As detailed above, insufficient blood flow to enterocytes interferes with their functions and triggers production of messengers such as pro-inflammatory cytokines^{74,75} (Figure 2B).

In summary, exercise that continuously exceeds the capacity to maintain sufficient blood flow to the gut results in enterocyte hypoxia, which impairs gut-barrier integrity,¹¹⁹ increases oxygen concentrations in the gut lumen, and decreases the abundance of anaerobic fermenting microbes^{120,121} (Figure 2B). Such microbiome dysbiosis and impaired barrier parallel changes observed during chronic HIV infection and promote microbial translocation, chronic activation of gut mucosa-associated immune cells, upward-spiraling immune system activation and further impairment of the gut barrier (Figure 2B).

Additional Effects of Physical Activity on the Gut and Immune System

Additional mechanisms may also contribute to the effects of physical activity on the immune system. For example, adipose tissue around the waist (visceral fat) is metabolically active and releases inflammatory hormones.¹²² Excess visceral fat thereby leads to chronic NF- κ B activation and non-resolving inflammation.^{123,124} Exercise can contribute to preventing excess adipose tissue as one avenue to reduce non-resolving inflammation.^{96,125-127}

Moreover, regular physical activity may also restore immune system balance by supporting redox homeostasis.¹²⁸ All exercise generates oxidants (reactive oxygen species, ROS) in working muscles which, in turn, stimulate production of endogenous antioxidant enzymes that help keep ROS in check.^{129,130} Moderate amounts of ROS are essential to induce these important antioxidant enzymes.¹³¹ Redox homeostasis supports immune function, that is, stimulates immunity against infection without inducing excessive self-attack or non-resolving inflammation.¹³² Regular non-exhaustive physical activity presumably stimulates ROS and antioxidant production in a balanced ratio that maintains redox homeostasis. Antioxidants are needed to keep ROS from triggering excessive NF- κ B activation and programmed cell death.^{133,134} Continuously exhaustive exercise may produce ROS at a level that exceeds the capacity for antioxidant enzyme production and results in chronic redox imbalance. It should also be noted that most of the antioxidant enzymes induced by physical activity require dietary mineral cofactors, such as zinc, selenium, copper, and manganese, and also cooperate with dietary antioxidant vitamins, such as vitamins C and E.¹³⁵

The Transition Between Beneficial and Detrimental Exercise

Intensity and Habituation. There is interest in identifying a quantifiable transition point between beneficial and detrimental physical activity. The authors¹³⁶ of a recent review of the relevant evidence proposed that “*vigorous endurance training with ≥ 60 min at $\geq 70\%$ of VO_{2max} increases the intestinal permeability, with an enhanced effect observed in hot environments, at high altitude, and under dehydration.*” Exercise at 70-80% of VO_{2max} (maximal aerobic capacity, or maximal oxygen consumption) was reported to be the point at which blood flow to the gut decreased by other authors.^{137,138} It should be noted that this recommendation ties the threshold to individual work capacity as a feature of individual fitness level that is, in turn, presumably associated with acclimation/habituation to a particular exercise regimen and varies with individual differences in training status as well as other personal factors.

Evidence for Effects of Physical Activity on Inflammatory Markers in PLWH. There is some evidence that exercise may influence immune markers in PLWH (Table 1). Table 1 shows that exercise can either lower inflammation markers, have no effect, or increase inflammation markers. The intensity at which training programs are completed may affect the response, with moderate and intense exercise reported as having contrasting effects on inflammatory markers in PLWH.¹³⁹ However, some of the studies finding decreases in pro-inflammatory markers in PLWH also involved what can be considered rather intense exercise.

A recent clinical trial consisting of combined aerobic and resistance training over six months, with aerobic exercise performed at 65–80% maximal heart rate, resulted in

statistically significant decreases in percent body fat and decreased levels of the pro-inflammatory cytokines IL-6 and TNF- α .¹⁴⁰ A 12-week resistance training program also significantly decreased subcutaneous body fat, IL-6 and TNF- α in sedentary PLWH¹⁴¹ as well as in PLWH with metabolic syndrome.¹⁴² A recent review¹⁴³ concluded that physical activity is associated with positive changes in the inflammatory environment of PLWH. However, other trials found a mix of decreases in some inflammatory markers and no change in others (eg, Dudgeon et al, 2012).¹⁴⁴ A meta-analysis¹⁴⁵ of a limited number of studies using diverse training regimens concluded that physical activity does not significantly reduce inflammation in PLWH.

Table 1 also includes a summary of recent studies on the effect of exercise on CD4⁺ T cell counts in PLWH. Non-resolving inflammation and dysbiosis are linked to diminished CD4⁺ cell counts¹⁴⁶ and low CD4⁺ counts persist in immunological non-responders (see above). Restoring CD4⁺ cells is a priority of HIV treatment, and elimination of non-resolving inflammation may facilitate recovery of CD4⁺ T cell counts in PLWH. Exercise increased CD4⁺ cell counts in some trials on exercise in PLWH^{141,147-151} but not in others.¹⁵²⁻¹⁵⁴ The potential of physical activity to increase CD4⁺ counts thus warrants further study as a potential avenue to strengthen immunity in PLWH.

Resolving Inflammation and Recovery Time

It should be noted that classification of inflammation regulators into pro- vs anti-inflammatory molecules does not capture the context-dependent roles of these molecules, which applies to both cytokine hormones¹⁵⁵ and another key class of inflammation

Table 1.

Summary of Selected Studies Published Over the Past Ten Years on the Effects of Interventional Physical Activity Programs on Inflammatory Markers and/or CD4⁺ Cell Counts in PWLH. Significant Effects (P<.05) Are Noted in Relation to the Duration and Type of the Physical Activity Completed. Longitudinal Study Assesses Variables Over an Extended Period of Time.

Author(s)	Type of study (original or review)	Duration and type of physical activity	Effects on inflammatory markers	Effects on CD4 ⁺ T-cell count
			Anti-inflammatory effects	
Bonato et al, ¹⁷⁹ 2017	Original study	12 weeks of a) combined aerobic (walking) and strength training or b) aerobic training (walking) alone	Both groups: decrease in pro-inflammatory hsCRP and IL-18; aerobic-only group: decrease in pro-inflammatory IL-6	No longitudinal measure
Bonato et al, ¹⁴³ 2020	Review	Longitudinal, interventional training program	Decrease in pro-inflammatory CRP, IL-8 and IL-6	No longitudinal measure
Dudgeon et al, ¹⁴⁴ 2012	Original study	6 weeks of moderate intensity combined aerobic and strength training	Decrease in salivary cortisol as a stress marker at wake	No longitudinal measure
Ghayomzadeh et al, ¹⁴⁰ 2021	Original study	6-month combined aerobic and resistance training; aerobic exercise completed at 65–80% maximal heart rate	Decrease in pro-inflammatory IL-6 and TNF- α	No longitudinal measure
Pedro et al, ¹⁸⁰ 2017	Original study	16-week heart rate-guided aerobic program	Decrease in pro-inflammatory IL-8	No longitudinal measure
Zanetti et al, ¹⁴¹ 2016a	Original study	12-week resistance training program	Decrease in pro-inflammatory IL-1 β , IL-6, IL-8 and TNF- α	Increase in CD4 ⁺ count
Zanetti et al, ¹⁸¹ 2016b	Original study	12-week resistance training program	Decrease in pro-inflammatory CRP	No longitudinal measure
Zanetti et al, ¹⁴² 2017	Original study	12-week resistance training program	Decrease in pro-inflammatory IL-1 β , IL-6, IL-8, and TNF- α	No longitudinal measure
Zanetti et al, ¹⁸² 2020	Original study	12-week resistance training program; mild to moderate intensity	Decrease in pro-inflammatory IL-1 β and CRP; increase in anti-inflammatory IL-10	No longitudinal measure
			No significant effects	
Cutrono et al, ¹⁸³ 2016	Original study	12 weeks of combined aerobic and resistance training; aerobic sessions completed at 60-80% of age-determined maximum heart rate	No significant changes in inflammatory markers	No longitudinal measure
Ibeneme et al, ¹⁴⁵ 2019a	Review	Aerobic training alone, resistance training alone, and combination of aerobic and resistance training	No significant changes in inflammatory markers	No longitudinal measure
Vingren et al, ¹⁸⁴ 2018	Original study	6-week resistance training program	No significant changes in inflammatory markers	No longitudinal measure

(continued)

Table 1. (continued)

Zanetti et al, ¹⁵¹ 2021	Review	6–24 weeks of resistance training programs	No significant changes in inflammatory markers	Increase in CD4 ⁺ cell count
			Pro-inflammatory effects	
Erlandson et al, ¹³⁹ 2020	Original study	24 weeks of combined aerobic and strength training; first 2 weeks at low intensity; next 10 weeks at moderate intensity (40–50% baseline VO_{2max}); last 12 weeks at moderate or high (60–70% of week 12 VO_{2max}) intensity	Increase in pro-inflammatory IL-6 in high intensity group	No longitudinal measure
Zanetti et al, ¹⁴² 2017	Original study	12-week resistance training program	Decrease in anti-inflammatory IL-10	No longitudinal measure
			No inflammatory markers measured	
Asogwa et al, ¹⁵⁰ 2020	Original study	6 weeks of moderate intensity aerobic training	NA	Increase in CD4 ⁺ count
de Brito-Neto et al, ¹⁴⁹ 2019	Original study	12-week resistance training program	NA	Increase in CD4 ⁺ cell count
Dianatinasab et al, ¹⁸⁵ 2018	Original study	12-week combined aerobic and resistance training	NA	Decrease in CD4 ⁺ cell count
Ezema et al, ¹⁴⁷ 2014	Original study	8 weeks of moderate intensity aerobic training completed at 60–79% of maximum heart rate	NA	Increase in CD4 ⁺ cell count
Ibeneme et al, ¹⁵⁴ 2019b	Review	Aerobic and/or resistance training	NA	No significant changes in CD4 ⁺ count
Maduagwu et al, ¹⁴⁸ 2017	Original study	12-week moderate intensity aerobic training program completed at 50–75% of heart rate reserve	NA	Increase in CD4 ⁺ count
O'Brien et al, ¹⁵³ 2016	Review	Aerobic training alone or a combination of aerobic and resistance training	NA	No significant changes in CD4 ⁺ count
Tiozzo, ¹⁵² 2011	Original study	12-week moderate intensity combined aerobic and resistance training program	NA	No significant changes in CD4 ⁺ count

Abbreviations: CRP, C-reactive protein; hsCRP, high-sensitivity C-reactive protein; IL, interleukin; NA, not applicable (no inflammatory marker measured) TNF, tumor necrosis factor.

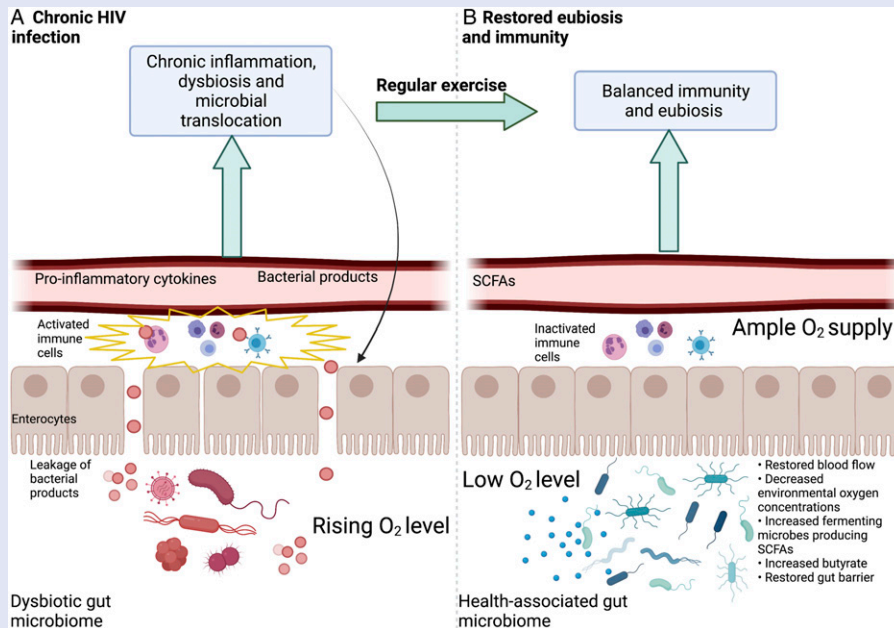
regulators, the fatty-acid-based eicosanoids.^{156,157} Moreover, increased acute inflammation following physical activity is not problematic and instead plays a critical role in triggering the

synthesis of antioxidant enzymes¹³¹ and thus maintaining immune balance.^{129,132,158} In fact, when oxidative signals are eliminated by high-dose antioxidant vitamins taken during athletic training,

synthesis of endogenous antioxidant enzymes as well as muscle building is prevented.¹³¹ Similarly, anti-inflammatory treatments (eg, NSAIDs) targeting exercise-associated injury can actually

Figure 3.

Schematic depiction of proposed changes in the gut of PLWH induced by regular, moderate intensity, interest-driven physical activity. (A) Dysbiosis and impaired barrier present during chronic HIV infection (B) Restored blood flow, eubiosis and reduction of inflammation due to regular physical activity. Created with [BioRender.com](https://www.biorender.com).



prevent healing by blocking the pro-resolution “stop signals” involved in the inflammatory response.¹⁵⁹ Physical activity can thereby acutely increase inflammatory markers while also strengthening pro-resolution pathways.¹⁶⁰ Taken together, these results suggest that short-term increases in markers of inflammation following physical activity may boost, rather than weaken, the anti-inflammatory resolution of inflammation.

The recovery time available for the resolution of acute inflammation may play a role. In other words, exercise performed at an excessive duration,¹⁶¹ without sufficient recovery time between sessions,¹⁶² may be what is most detrimental.¹⁶³ For example, what was described as “vigorous” exercise by the authors^{164,165} increased the levels of the stress hormone cortisol, whereas “moderate” exercise had no such effect. Likewise, “forced” treadmill

running in mice promoted significantly greater increases in cortisol than “voluntary” wheel running.¹⁶⁶ For such exhaustive exercise, constitutive activation of NF- κ B and subsequent tissue degeneration, oxidative stress, gut barrier breakdown, gut microbiome dysbiosis and non-resolving inflammation have been demonstrated.¹⁶⁷⁻¹⁷¹ At this point, it can thus not be excluded that detrimental exercise modalities described as “high-intensity,” “vigorous,” or “forced,” may also be lacking in adequate time for recovery and resolution of inflammation between sessions. A notable study by Schlabe et al (2017)¹⁷² suggests that even marathon training, performed as “moderate endurance training” over a period of 12 months, can be safe for PLWH.

Future research is needed that includes assessment of time allowed

for recovery and resolution of inflammation, and uses consistent standards to quantify work capacity (such as VO_{2max}), what percent of this capacity was reached and for how long during the physical activity, as well as blood flow to the gut across a range of different physical activities. Furthermore, individualized physical activity regimens likely need to be customized for different needs. As stated above, acclimation to a regimen of physical activity can restore blood supply to the gut during exercise. Such acclimation presumably raises the ceiling of work capacity. However, there likely is an upper limit of this acclimation effect that may be exceeded by certain types of exhaustive exercise. The current understanding thus suggests that physical activity recommendations should be expanded to comprehensively emphasize movement that is

interest-based, voluntary, completed at an enjoyable intensity, performed regularly, and allows adequate time for recovery and resolution of inflammation.^{75,173-178}

Proposed Benefits of Physical Activity for PLWH

The evidence reviewed above supports a proposal that regular physical activity—allowing sufficient acclimation and recovery that restore blood flow to the gut—may facilitate the resolution of inflammation by ameliorating the leaky gut present in many treated and untreated PLWH (Figure 3). As stated above (Figure 1B), many PLWH experience a dysbiotic gut microbiome, characterized by gut-barrier disruption and microbial translocation that persist despite ART treatment (Figures 1B and 3A). This microbial translocation activates the immune system and leads to release of pro-inflammatory cytokines, microbes, and microbial products into the bloodstream, further triggering widespread immune activation and gut barrier breakdown (see above; Figures 1B and 3A). The non-resolving inflammation experienced by PLWH (Figure 3B) may thus be reduced by adoption of a regimen of regular physical activity that allows sufficient blood flow to the GI tract, increases abundance of fermenting microbes and SCFAs, and strengthens the gut barrier integrity. This type of physical activity may protect against various comorbidities in both treated and untreated PLWH and serve as a complement to traditional ART (Figure 3B).

At this time, there is some evidence that exercise can reduce markers of inflammation and improve body composition in PLWH (see above; Table 1).^{141,143,179} However, more research is needed to establish this because findings are presently not consistent.¹⁴⁵ It is clear, however,

that physical activity interacts with the same molecular players that are affected by HIV, such as NF-κB. Whereas an acute bout of exercise was associated with acute activation of NF-κB, acclimation to regular exercise suppressed NF-κB in mice models and prevented non-resolving inflammation.¹¹⁴

Physical activity thus offers great promise as a potential lifestyle modification for reducing non-resolving inflammation and inflammation-associated comorbidities in PLWH. Physicians and other healthcare providers should be aware of the specific benefits physical activity may offer to PLWH and consider incorporating physical activity into treatment plans as a complement to ART. Additional research is needed to design individualized training plans customized to match individual fitness levels and interest.

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References

- Phanuphak N, Gulick RM. HIV treatment and prevention 2019: current standards of care. *Curr Opin HIV AIDS*. 2020;15(1):4-12. doi:10.1097/COH.0000000000000588
- Sterne JA, Hernán MA, Ledergerber B, et al. Long-term effectiveness of potent antiretroviral therapy in preventing AIDS and death: a prospective cohort study. *Lancet*.

2005;366(9483):378-384. doi:10.1016/S0140-6736(05)67022-5

- Broder S. The development of antiretroviral therapy and its impact on the HIV-1/AIDS pandemic. *Antiviral Res*. 2010;85(1):1-18. doi:10.1016/j.antiviral.2009.10.002
- CDC. *About HIV/AIDS*. <https://www.cdc.gov/hiv/basics/whatishiv.html>. Published June 1, 2021. Accessed September 25, 2021.
- Doitsh G, Greene WC. Dissecting how CD4 T cells are lost during HIV infection. *Cell Host Microbe*. 2016;19(3):280-291. doi:10.1016/j.chom.2016.02.012
- Schett G, Neurath MF. Resolution of chronic inflammatory disease: universal and tissue-specific concepts. *Nat Commun*. 2018;9(1):3261. doi:10.1038/s41467-018-05800-6
- Buckley CD, Gilroy DW, Serhan CN. Pro-Resolving lipid mediators and Mechanisms in the resolution of acute inflammation. *Immunity*. 2014;40(3):315-327. doi:10.1016/j.immuni.2014.02.009
- Deeks SG, Tracy R, Douek DC. Systemic effects of inflammation on health during chronic HIV infection. *Immunity*. 2013;39(4):633-645. doi:10.1016/j.immuni.2013.10.001
- Maniar A, Ellis C, Asmuth D, Pollard R, Rutledge J. HIV infection and atherosclerosis: evaluating the drivers of inflammation. *Eur J Prev Cardiol*. 2013;20(5):720-728. doi:10.1177/2047487312447843
- Nou E, Lo J, Grinspoon SK. Inflammation, immune activation, and cardiovascular disease in HIV. *AIDS*. 2016;30(10):1495-1509. doi:10.1097/QAD.0000000000001109
- Zicari S, Sessa L, Cotugno N, et al. Immune activation, inflammation, and Non-AIDS Co-Morbidities in HIV-infected patients under long-term ART. *Viruses*. 2019;11(3):200. doi:10.3390/v11030200
- Bloch M, John M, Smith D, Rasmussen T, Wright E. Managing HIV-associated inflammation and ageing in the era of modern ART. *HIV Med*. 2020;21(S3):2-16. doi:10.1111/hiv.12952
- Cribbs SK, Crothers K, Morris A. Pathogenesis of HIV-related lung disease: immunity, infection, and inflammation. *Physiol Rev*. 2020;100(2):603-632. doi:10.1152/physrev.00039.2018

14. Freed EO. HIV-1 and the host cell: an intimate association. *Trends Microbiol.* 2004;12(4):170-177. doi:10.1016/j.tim.2004.02.001
15. Roulston A, Lin R, Beuparlant P, Wainberg MA, Hiscott J. Regulation of human immunodeficiency virus type 1 and cytokine gene expression in myeloid cells by NF-kappa B/Rel transcription factors. *Microbiol Rev.* 1995;59(3):481-505.
16. Bren GD, Trushin SA, Whitman J, Shepard B, Badley AD. HIV gp120 induces, NF-kB Dependent, HIV replication that requires Procaspase 8. *PLoS One.* 2009;4(3):e4875. doi:10.1371/journal.pone.0004875
17. Baldwin AS. THE NF-kB AND IκB PROTEINS: new discoveries and insights. *Annu Rev Immunol.* 1996;14(1):649-681. doi:10.1146/annurev.immunol.14.1.649
18. Oeckinghaus A, Ghosh S. The NF-kappaB family of transcription factors and its regulation. *Cold Spring Harb Perspect Biol.* 2009;1(4):a000034. doi:10.1101/cshperspect.a000034
19. Hayden MS, West AP, Ghosh S. NF-κB and the immune response. *Oncogene.* 2006;25(51):6758-6780. doi:10.1038/sj.onc.1209943
20. Pajusto M, Toivonen TH, Tarkkanen J, Jokitalo E, Mattila PS. Reactive oxygen species induce signals that lead to apoptotic DNA degradation in primary CD4+ T cells. *Apoptosis.* 2005;10(6):1433-1443. doi:10.1007/s10495-005-2050-5
21. Chelombitko MA. Role of reactive oxygen species in inflammation: a minireview. *Mosc Univ Biol Sci Bull.* 2018;73(4):199-202. doi:10.3103/S009639251804003X
22. O'Malley D, Quigley EMM, Dinan TG, Cryan JF. Do interactions between stress and immune responses lead to symptom exacerbations in irritable bowel syndrome? *Brain Behav Immun.* 2011;25(7):1333-1341. doi:10.1016/j.bbi.2011.04.009
23. Myers JL, Allen JC. Nutrition and inflammation: insights on dietary pattern, obesity, and Asthma. *Am J Lifestyle Med.* 2012;6(1):14-17. doi:10.1177/1559827611424259
24. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal.* 2014;20(7):1126-1167. doi:10.1089/ars.2012.5149
25. Lopez-Candales A, Hernández Burgos PM, Hernandez-Suarez DF, Harris D. Linking chronic inflammation with cardiovascular disease: from normal aging to the metabolic syndrome. *J Nat Sci.* 2017;3(4):e341.
26. Zeng MY, Inohara N, Nuñez G. Mechanisms of inflammation-driven bacterial dysbiosis in the gut. *Mucosal Immunol.* 2017;10(1):18-26. doi:10.1038/mi.2016.75
27. Bauer ME, Teixeira AL. Inflammation in psychiatric disorders: what comes first? *Ann N Y Acad Sci.* 2019;1437(1):57-67. doi:10.1111/nyas.13712
28. Slavich GM. Understanding inflammation, its regulation, and relevance for health: a top scientific and public priority. *Brain Behav Immun.* 2015;45:13-14. doi:10.1016/j.bbi.2014.10.012
29. Furman D, Campisi J, Verdin E, et al. Chronic inflammation in the etiology of disease across the life span. *Nat Med.* 2019;25(12):1822-1832. doi:10.1038/s41591-019-0675-0
30. Berg RK, Rahbek SH, Kofod-Olsen E, et al. T cells detect intracellular DNA but fail to induce type I IFN responses: implications for restriction of HIV Replication. *PLoS One.* 2014;9(1):e84513. doi:10.1371/journal.pone.0084513
31. Manel N, Hogstad B, Wang Y, Levy DE, Unutmaz D, Littman DR. A cryptic sensor for HIV-1 activates antiviral innate immunity in dendritic cells. *Nature.* 2010;467(7312):214-217. doi:10.1038/nature09337
32. Utay NS, Douek DC. Interferons and HIV infection: the good, the bad, and the ugly. *Pathog Immun.* 2016;1(1):107-116. doi:10.20411/pai.v1i1.125
33. Autran B, Carcelain G, Li TS, et al. Positive effects of combined antiretroviral therapy on CD4+ T cell homeostasis and function in advanced HIV Disease. *Science.* 1997;277(5322):112-116. doi:10.1126/science.277.5322.112
34. Ledergerber B, Egger M, Opravil M, et al. Clinical progression and virological failure on highly active antiretroviral therapy in HIV-1 patients: a prospective cohort study. *Lancet.* 1999;353(9156):863-868. doi:10.1016/S0140-6736(99)01122-8
35. Silveira MPT, Silveira CPT, Guttier MC, et al. Long-term immune and virological response in HIV-infected patients receiving antiretroviral therapy. *J Clin Pharm Ther.* 2016;41(6):689-694. doi:10.1111/jcpt.12450
36. Gazzola L, Tincati C, Bellistri GM, et al. The absence of CD4+ T cell count recovery despite receipt of virologically suppressive highly active antiretroviral therapy: clinical risk, immunological gaps, and therapeutic options. *Clin Infect Dis Off Publ Infect Dis Soc Am.* 2009;48(3):328-337. doi:10.1086/595851
37. Robbins GK, Spritzler JG, Chan ES, et al. Incomplete reconstitution of T cell subsets on combination antiretroviral therapy in the AIDS Clinical Trials Group protocol 384. *Clin Infect Dis Off Publ Infect Dis Soc Am.* 2009;48(3):350-361. doi:10.1086/595888
38. Massanella M, Gómez-Mora E, Carrillo J, et al. Increased ex vivo cell death of central memory CD4 T cells in treated HIV infected individuals with unsatisfactory immune recovery. *J Transl Med.* 2015;13:230. doi:10.1186/s12967-015-0601-2
39. Lichtfuss GF, Hoy J, Rajasuriar R, et al. Biomarkers of immune dysfunction following combination antiretroviral therapy for HIV infection. *Biomark Med.* 2011;5(2):171-186. doi:10.2217/bmm.11.15
40. Lichtfuss GF, Cheng WJ, Farsakoglu Y, et al. Virologically suppressed HIV patients show activation of NK cells and persistent innate immune activation. *J Immunol Baltim Md* 1950. 2012;189(3):1491-1499. doi:10.4049/jimmunol.1200458
41. Rhoades N, Mendoza N, Jankeel A, et al. Altered immunity and microbial dysbiosis in aged individuals with long-term controlled HIV infection. *Front Immunol.* 2019;10:463. doi:10.3389/fimmu.2019.00463
42. Brenchley J, Douek D. HIV infection and the gastrointestinal immune system. *Mucosal Immunol.* 2008;1(1):23-30. doi:10.1038/mi.2007.1
43. Dinh DM, Volpe GE, Duffalo C, et al. Intestinal microbiota, microbial translocation, and systemic inflammation in chronic HIV Infection. *J Infect Dis.* 2015;211(1):19-27. doi:10.1093/infdis/jiu409
44. Zevin AS, McKinnon L, Burgener A, et al. Microbial translocation and microbiome dysbiosis in HIV-associated immune activation. *Curr Opin HIV AIDS.* 2016;11(2):

- 182-190. doi:10.1097/COH.0000000000000234
45. Alzahrani J, Hussain T, Simar D, et al. Inflammatory and immunometabolic consequences of gut dysfunction in HIV: parallels with IBD and implications for reservoir persistence and non-AIDS comorbidities. *EBioMedicine*. 2019;46:522-531. doi:10.1016/j.ebiom.2019.07.027
 46. Vujkovic-Cvijin I, Sortino O, Verheij E, et al. HIV-associated gut dysbiosis is independent of sexual practice and correlates with noncommunicable diseases. *Nat Commun*. 2020;11(1):2448. doi:10.1038/s41467-020-16222-8
 47. Iebba V, Totino V, Gagliardi A, et al. Eubiosis and dysbiosis: the two sides of the microbiota. *New Microbiol*. 2016;39(1):1-12.
 48. Bergman EN. Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. *Physiol Rev*. 1990;70(2):567-590. doi:10.1152/physrev.1990.70.2.567
 49. Scheppach W. Effects of short chain fatty acids on gut morphology and function. *Gut*. 1994;35(suppl 1):S35-S38. doi:10.1136/gut.35.1_Suppl.S35
 50. den Besten G, van Eunen K, Groen AK, Venema K, Reijnders DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J Lipid Res*. 2013;54(9):2325-2340. doi:10.1194/jlr.R036012
 51. Bugaut M. Occurrence, absorption and metabolism of short chain fatty acids in the digestive tract of mammals. *Comp Biochem Physiol B*. 1987;86(3):439-472. doi:10.1016/0305-0491(87)90433-0
 52. Cummings JH, Pomare EW, Branch WJ, Naylor CP, Macfarlane GT. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut*. 1987;28(10):1221-1227.
 53. Feng Y, Wang Y, Wang P, Huang Y, Wang F. Short-chain fatty acids manifest stimulative and protective effects on intestinal barrier function through the inhibition of NLRP3 inflammasome and autophagy. *Cell Physiol Biochem*. 2018;49(1):190-205. doi:10.1159/000492853
 54. Parada Venegas D, De la Fuente MK, Landskron G, et al. Short chain fatty acids (SCFAs)-mediated gut epithelial and immune regulation and its relevance for inflammatory bowel diseases. *Front Immunol*. 2019;10:277. doi:10.3389/fimmu.2019.00277
 55. Kelly CJ, Zheng L, Campbell EL, et al. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe*. 2015;17(5):662-671. doi:10.1016/j.chom.2015.03.005
 56. Yan H, Ajuwon KM. Butyrate modifies intestinal barrier function in IPEC-J2 cells through a selective upregulation of tight junction proteins and activation of the Akt signaling pathway. *PLoS One*. 2017;12(6):e0179586. doi:10.1371/journal.pone.0179586
 57. Wang HB, Wang PY, Wang X, Wan YL, Liu YC. Butyrate enhances intestinal epithelial barrier function via up-regulation of tight junction protein Claudin-1 transcription. *Dig Dis Sci*. 2012;57(12):3126-3135. doi:10.1007/s10620-012-2259-4
 58. Krishnan M, Singh A, Smith J, et al. HDAC inhibitors regulate claudin-1 expression in colon cancer cells through modulation of mRNA stability. *Oncogene*. 2010;29(2):305-312. doi:10.1038/ncr.2009.324
 59. Luetjig J, Rosenthal R, Barmeyer C, Schulzke JD. Claudin-2 as a mediator of leaky gut barrier during intestinal inflammation. *Tissue Barriers*. 2015;3(1-2):e977176. doi:10.4161/21688370.2014.977176
 60. Albenberg L, Esipova T, Judge C, et al. Correlation between intraluminal oxygen gradient and radial partitioning of intestinal microbiota in humans and mice. *Gastroenterology*. 2014;147(5):1055-1063.e8. doi:10.1053/j.gastro.2014.07.020
 61. Zheng L, Kelly CJ, Colgan SP. Physiologic hypoxia and oxygen homeostasis in the healthy intestine. a review in the theme: cellular responses to hypoxia. *Am J Physiol Cell Physiol*. 2015;309(6):C350-C360. doi:10.1152/ajpcell.00191.2015
 62. Ward JBJ, Keely SJ, Keely SJ. Oxygen in the regulation of intestinal epithelial transport. *J Physiol*. 2014;592(Pt 12):2473-2489. doi:10.1113/jphysiol.2013.270249
 63. Miron N, Cristea V. Enterocytes: active cells in tolerance to food and microbial antigens in the gut. *Clin Exp Immunol*. 2012;167(3):405-412. doi:10.1111/j.1365-2249.2011.04523.x
 64. Zheng L, Kelly CJ, Battista KD, et al. Microbial-derived butyrate promotes epithelial barrier function through IL-10 receptor-dependent repression of Claudin-2. *J Immunol*. 2017;199(8):2976-2984. doi:10.4049/jimmunol.1700105
 65. Colgan SP, Taylor CT. Hypoxia: an alarm signal during intestinal inflammation. *Nat Rev Gastroenterol Hepatol*. 2010;7(5):281-287. doi:10.1038/nrgastro.2010.39
 66. Louis P, Flint HJ. Formation of propionate and butyrate by the human colonic microbiota. *Environ Microbiol*. 2017;19(1):29-41. doi:10.1111/1462-2920.13589
 67. Inan MS, Rasoulpour RJ, Yin L, Hubbard AK, Rosenberg DW, Giardina C. The luminal short-chain fatty acid butyrate modulates NF-κB activity in a human colonic epithelial cell line. *Gastroenterology*. 2000;118(4):724-734. doi:10.1016/S0016-5085(00)70142-9
 68. Segain J, de la Bletiere DR, Bourreille A, et al. Butyrate inhibits inflammatory responses through NFκB inhibition: implications for Crohn's disease. *Gut*. 2000;47(3):397-403. doi:10.1136/gut.47.3.397
 69. Gassull MA. Review article: the intestinal lumen as a therapeutic target in inflammatory bowel disease. *Aliment Pharmacol Ther*. 2006;24(s3):90-95. doi:10.1111/j.1365-2036.2006.03067.x
 70. Liu T, Li J, Liu Y, et al. Short-chain fatty acids suppress lipopolysaccharide-induced production of nitric oxide and proinflammatory cytokines through inhibition of NF-κB pathway in RAW264.7 cells. *Inflammation*. 2012;35(5):1676-1684. doi:10.1007/s10753-012-9484-z
 71. Yang Q, Ouyang J, Sun F, Yang J. Short-chain fatty acids: a soldier fighting against inflammation and protecting from tumorigenesis in people with diabetes. *Front Immunol*. 2020;11:590685. doi:10.3389/fimmu.2020.590685
 72. Macpherson AJ, Geueking MB, McCoy KD. Immune responses that adapt the intestinal mucosa to commensal intestinal bacteria. *Immunology*. 2005;115(2):153-162. doi:10.1111/j.1365-2567.2005.02159.x
 73. Vighi G, Marcucci F, Sensi L, Di Cara G, Frati F. Allergy and the gastrointestinal system. *Clin Exp*

- Immunol.* 2008;153(s1):3-6. doi:10.1111/j.1365-2249.2008.03713.x
74. Royes LFF. Cross-talk between gut and brain elicited by physical exercise. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866(10):165877. doi:10.1016/j.bbadis.2020.165877
 75. Keirns BH, Koemel NA, Sciarillo CM, Anderson KL, Emerson SR. Exercise and intestinal permeability: another form of exercise-induced hormesis? *Am J Physiol Gastrointest Liver Physiol.* 2020;319(4):G512-G518. doi:10.1152/ajpgi.00232.2020
 76. Mehndru S, Poles MA, Tenner-Racz K, et al. Lack of mucosal immune reconstitution during prolonged treatment of acute and early HIV-1 infection. *PLoS Med.* 2006;3(12):e484. doi:10.1371/journal.pmed.0030484
 77. Somsouk M, Estes JD, Deleage C, et al. Gut epithelial barrier and systemic inflammation during chronic HIV infection. *AIDS Lond Engl.* 2015;29(1):43-51. doi:10.1097/QAD.0000000000000511
 78. Ponte R, Mehraj V, Ghali P, Couëdel-Courteille A, Cheyner R, Routy JP. Reversing gut damage in HIV infection: using non-human primate models to instruct clinical research. *EBioMedicine.* 2016;4:40-49. doi:10.1016/j.ebiom.2016.01.028
 79. Sandler NG, Douek DC. Microbial translocation in HIV infection: causes, consequences and treatment opportunities. *Nat Rev Microbiol.* 2012;10(9):655-666. doi:10.1038/nrmicro2848
 80. Ramendra R, Isnard S, Mehraj V, et al. Circulating LPS and (1→3)-β-D-Glucan: A Folie à Deux Contributing to HIV-associated immune activation. *Front Immunol.* 2019;10:465. doi:10.3389/fimmu.2019.00465
 81. Andreaskos E, Sacre SM, Smith C, et al. Distinct pathways of LPS-induced NF-κB activation and cytokine production in human myeloid and nonmyeloid cells defined by selective utilization of MyD88 and Mal/TIRAP. *Blood.* 2004;103(6):2229-2237. doi:10.1182/blood-2003-04-1356
 82. Candelli M, Franza L, Pignataro G, et al. Interaction between Lipopolysaccharide and Gut Microbiota in Inflammatory Bowel Diseases. *Int J Mol Sci.* 2021;22(12):6242. doi:10.3390/ijms22126242
 83. Hoessel B, Schmid JA. The complexity of NF-κB signaling in inflammation and cancer. *Mol Cancer.* 2013;12(1):86. doi:10.1186/1476-4598-12-86
 84. Liu T, Zhang L, Joo D, Sun SC. NF-κB signaling in inflammation. *Signal Transduct Target Ther.* 2017;2:17023. doi:10.1038/sigtrans.2017.23
 85. Mussbacher M, Salzmann M, Brostjan C, et al. Cell type-specific roles of NF-κB linking inflammation and thrombosis. *Front Immunol.* 2019;10:85. doi:10.3389/fimmu.2019.00085
 86. Rigottier-Gois L. Dysbiosis in inflammatory bowel diseases: the oxygen hypothesis. *ISME J.* 2013;7(7):1256-1261. doi:10.1038/ismej.2013.80
 87. Stecher B. The roles of inflammation, nutrient availability and the commensal microbiota in enteric pathogen infection. *Microbiol Spectr.* 2015;3(3):MBP-0008-2014. doi:10.1128/microbiolspec.MBP-0008-2014
 88. Rivera-Chavez F, Zhang L, Faber F. Depletion of butyrate-producing clostridia from the gut microbiome drives an aerobic luminal expansion of Salmonella. *Cell Host Microbe.* 2016;19(4):443-454.
 89. Winter SE, Bäumlner AJ. Why related bacterial species bloom simultaneously in the gut: principles underlying the "Like will to like" concept. *Cell Microbiol.* 2014;16(2):179-184. doi:10.1111/cmi.12245
 90. Lloyd-Price J, Arze C, Ananthakrishnan AN, et al. Multi-omics of the gut microbial ecosystem in inflammatory bowel diseases. *Nature.* 2019;569(7758):655-662. doi:10.1038/s41586-019-1237-9
 91. Shephard RJ. Absolute versus relative intensity of physical activity in a dose-response context. *Med Sci Sports Exerc.* 2001;33(6):S400.
 92. Woo JS, Derleth C, Stratton JR, Levy WC. The influence of age, gender, and training on exercise efficiency. *J Am Coll Cardiol.* 2006;47(5):1049-1057. doi:10.1016/j.jacc.2005.09.066
 93. Clark J. The impact of duration on effectiveness of exercise, the implication for periodization of training and goal setting for individuals who are overfat, a meta-analysis. *Biol Sport.* 2016;33(4):309-333. doi:10.5604/20831862.1212974
 94. Seifi-skishahr F, Damirchi A, Farjaminezhad M, Babaei P. Physical training status determines oxidative stress and redox changes in response to an acute aerobic exercise. *Biochem Res Int.* 2016;2016:e3757623. doi:10.1155/2016/3757623
 95. Raffin J, Barthélémy JC, Dupré C, et al. Exercise frequency determines heart rate variability gains in older people: a meta-analysis and meta-regression. *Sports Med Auckl NZ.* 2019;49(5):719-729. doi:10.1007/s40279-019-01097-7
 96. Woods JA, Wilund KR, Martin SA, Kistler BM. Exercise, inflammation and aging. *Aging Dis.* 2011;3(1):130-140.
 97. Simpson RJ, Kunz H, Agha N, Graff R. Chapter fifteen - exercise and the regulation of immune functions. In: Bouchard C, ed. *Progress in Molecular Biology and Translational Science. Vol 135. Molecular and Cellular Regulation of Adaptation to Exercise.* Academic Press; 2015:355-380. doi:10.1016/bs.pmbts.2015.08.001
 98. Scheffer DdL, Latini A. Exercise-induced immune system response: anti-inflammatory status on peripheral and central organs. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866(10):165823. doi:10.1016/j.bbadis.2020.165823
 99. Otte JA, Oostveen E, Geelkerken RH, Groeneveld ABJ, Kolkman JJ. Exercise induces gastric ischemia in healthy volunteers: a tonometry study. *J Appl Physiol.* 2001;91(2):866-871. doi:10.1152/jap.2001.91.2.866
 100. Evans CC, LePard KJ, Kwak JW, et al. Exercise prevents weight gain and alters the gut microbiota in a mouse model of high fat diet-induced obesity. *PLoS One.* 2014;9(3):e92193. doi:10.1371/journal.pone.0092193
 101. Kang SS, Jeraldo PR, Kurti A, et al. Diet and exercise orthogonally alter the gut microbiome and reveal independent associations with anxiety and cognition. *Mol Neurodegener.* 2014;9(1):36. doi:10.1186/1750-1326-9-36
 102. Monda V, Villano I, Messina A, et al. Exercise modifies the gut microbiota with positive health effects. *Oxid Med Cell Longev.* 2017;2017:1-8. doi:10.1155/2017/3831972
 103. Perrin P, Pierre F, Patry Y, et al. Only fibres promoting a stable butyrate producing colonic ecosystem decrease the rate of aberrant crypt foci in rats. *Gut.* 2001;48(1):53-61. doi:10.1136/gut.48.1.53

104. Cheng C, Wei H, Peng J. 370 Dietary soluble fiber increases the intestinal butyrate-producing bacteria, reduces intestinal permeability, and improves metabolic syndrome in sows during perinatal period. *J Anim Sci*. 2019; 97(suppl 3):133. doi:10.1093/jas/skz258.271
105. Flint HJ, Duncan SH, Scott KP, Louis P. Interactions and competition within the microbial community of the human colon: links between diet and health. *Environ Microbiol*. 2007;9(5): 1101-1111. doi:10.1111/j.1462-2920.2007.01281.x
106. Scheiman J, Lubner JM, Chavkin TA, et al. Meta-omics analysis of elite athletes identifies a performance-enhancing microbe that functions via lactate metabolism. *Nat Med*. 2019; 25(7):1104-1109. doi:10.1038/s41591-019-0485-4
107. Heuvelin E, Lebreton C, Grangette C, Pot B, Cerf-Bensussan N, Heyman M. Mechanisms involved in alleviation of intestinal inflammation by bifidobacterium breve soluble factors. *PLoS One*. 2009;4(4):e5184. doi:10.1371/journal.pone.0005184
108. Yan F, Polk DB. Disruption of NF- κ B signalling by ancient microbial molecules: novel therapies of the future? *Gut*. 2010;59(4):421-426. doi:10.1136/gut.2009.179614
109. Ferreira CM, Vieira AT, Vinolo MAR, Oliveira FA, Curi R, Martins FdS. The central role of the gut microbiota in chronic inflammatory diseases. *J Immunol Res*. 2014;2014:e689492. doi:10.1155/2014/689492
110. Mu Q, Kirby J, Reilly CM, Luo XM. Leaky gut as a danger signal for autoimmune diseases. *Front Immunol*. 2017;8:598. doi:10.3389/fimmu.2017.00598
111. Lazar V, Ditu LM, Pircalabioru GG, et al. Aspects of gut microbiota and immune system interactions in infectious diseases, immunopathology, and cancer. *Front Immunol*. 2018;9:1830. doi:10.3389/fimmu.2018.01830
112. Divella R, Palma GD, Tufaro A, et al. Diet, probiotics and physical activity: the right allies for a healthy microbiota. *Anticancer Res*. 2021; 41(6):2759-2772. doi:10.21873/anticancer.15057
113. Singh R, Zogg H, Wei L, et al. Gut microbial dysbiosis in the pathogenesis of gastrointestinal dysmotility and metabolic disorders. *J Neurogastroenterol Motil*. 2021; 27(1):19-34. doi:10.5056/jnm20149
114. Liu HW, Chang SJ. Moderate exercise suppresses NF- κ B signaling and activates the SIRT1-AMPK-PGC1 α Axis to Attenuate Muscle Loss in Diabetic db/db Mice. *Front Physiol*. 2018;9:636. doi:10.3389/fphys.2018.00636
115. Oliveira VHF, Borsari AL, Weibel AR, Erlandson KM, Deminice R. Sarcopenia in people living with the human immunodeficiency Virus: a systematic review and meta-analysis. *Eur J Clin Nutr*. 2020;74(7): 1009-1021. doi:10.1038/s41430-020-0637-0
116. Clark A, Mach N. The Crosstalk between the gut microbiota and mitochondria during exercise. *Front Physiol*. 2017;8:319. doi:10.3389/fphys.2017.00319
117. van Wijck K, Lenaerts K, van Loon LJC, Peters WHM, Buurman WA, Dejong CHC. Exercise-induced splanchnic hypoperfusion results in gut dysfunction in healthy men. *PLoS One*. 2011;6(7):e22366. doi:10.1371/journal.pone.0022366
118. Codella R, Luzi L, Terruzzi I. Exercise has the guts: how physical activity may positively modulate gut microbiota in chronic and immune-based diseases. *Dig Liver Dis*. 2018; 50(4):331-341. doi:10.1016/j.dld.2017.11.016
119. Lian P, Braber S, Varasteh S, Wichers HJ, Folkerts G. Hypoxia and heat stress affect epithelial integrity in a Caco-2/HT-29 co-culture. *Sci Rep*. 2021;11(1):13186. doi:10.1038/s41598-021-92574-5
120. Litvak Y, Byndloss MX, Bäuml AJ. Colonocyte metabolism shapes the gut microbiota. *Science*. 2018; 362(6418):eaat9076. doi:10.1126/science.aat9076
121. Konjar Š, Pavšič M, Veldhoen M. Regulation of oxygen homeostasis at the intestinal epithelial barrier site. *Int J Mol Sci*. 2021;22(17):9170. doi:10.3390/ijms22179170
122. Adams MS, Adams RB, Wessman CA, Demmig-Adams B. Nutritional cues tie living organisms to their environment and its sustainability. *Front Nutr*. 2016;3:28. doi:10.3389/fnut.2016.00028
123. Fontana L, Eagon JC, Trujillo ME, Scherer PE, Klein S. Visceral fat adipokine secretion is associated with systemic inflammation in obese humans. *Diabetes*. 2007;56(4): 1010-1013. doi:10.2337/db06-1656
124. Baker RG, Hayden MS, Ghosh S. NF- κ B, inflammation and metabolic disease. *Cell Metab*. 2011;13(1):11-22. doi:10.1016/j.cmet.2010.12.008
125. Flynn MG, McFarlin BK, Markofski MM. The anti-inflammatory actions of exercise training. *Am J Lifestyle Med*. 2007;1(3):220-235. doi:10.1177/1559827607300283
126. Mika A, Macaluso F, Barone R, Di Felice V, Sledzinski T. Effect of exercise on fatty acid metabolism and adipokine secretion in adipose tissue. *Front Physiol*. 2019;10:26. doi:10.3389/fphys.2019.00026
127. Niemi GM, Rewane A, Algotar AM. Exercise and fitness effect on obesity. In: *StatPearls*. Treasure Island (FL): StatPearls Publishing; 2021. <http://www.ncbi.nlm.nih.gov/books/NBK539893/>. Accessed October 6, 2021.
128. Margaritelis NV, Paschalis V, Theodorou AA, Kyparos A, Nikolaidis MG. Redox basis of exercise physiology. *Redox Biol*. 2020;35: 101499. doi:10.1016/j.redox.2020.101499
129. Radak Z, Chung HY, Goto S. Systemic adaptation to oxidative challenge induced by regular exercise. *Free Radic Biol Med*. 2008;44(2):153-159. doi:10.1016/j.freeradbiomed.2007.01.029
130. Freiberg MS, Chang C-CH, Kuller LH, et al. HIV infection and the risk of acute myocardial infarction. *JAMA Intern Med*. 2013;173(8):614-622. doi:10.1001/jamainternmed.2013.3728
131. Adams R, Egbo K, Demmig-Adams B. High-dose vitamin C supplements diminish the benefits of exercise in athletic training and disease prevention. *Nutr Food Sci*. 2014;44: 95-101.
132. Amir Aslani B, Ghojdi S. Studies on oxidants and antioxidants with a brief glance at their relevance to the immune system. *Life Sci*. 2016;146: 163-173. doi:10.1016/j.lfs.2016.01.014
133. Baruchel S, Wainberg MA. The role of oxidative stress in disease progression in individuals infected by the human immunodeficiency virus. *J Leukoc Biol*. 1992;52(1):111-114. doi:10.1002/jlb.52.1.111
134. Malorni W, Rivabene R, Teresa Santini M, Donelli G. N-Acetylcysteine inhibits apoptosis and decreases viral particles in HIV-chronically infected

- U937 cells. *FEBS Lett.* 1993;327(1):75-78. doi:10.1016/0014-5793(93)81043-Y
135. Tran E, Demmig-Adams B. Vitamins and minerals: powerful medicine or potent toxins? *Nutr Food Sci.* 2007;37(1):50-60. doi:10.1108/00346650710726959
136. Ribeiro FM, Petriz B, Marques G, Kamilla LH, Franco OL. Is there an exercise-intensity threshold capable of avoiding the leaky gut? *Front Nutr.* 2021;8:75. doi:10.3389/fnut.2021.627289
137. Casey E, Mistry DJ, MacKnight JM. Training room management of medical conditions: sports gastroenterology. *Clin Sports Med.* 2005;24(3):525-540, viii. doi:10.1016/j.csm.2005.05.002
138. van Wijck K, Lenaerts K, Grootjans J, et al. Physiology and pathophysiology of splanchnic hypoperfusion and intestinal injury during exercise: strategies for evaluation and prevention. *Am J Physiol Gastrointest Liver Physiol.* 2012;303(2):G155-G168. doi:10.1152/ajpgi.00066.2012
139. Erlandson KM, Wilson MP, MaWhinney S, et al. The impact of moderate or high-intensity combined exercise on systemic inflammation among older persons with and without HIV. *J Infect Dis.* 2020;223(7):1161-1170. doi:10.1093/infdis/jiaa494
140. Ghayomzadeh M, Earnest CP, Hackett D, et al. Combination of resistance and aerobic exercise for six months improves bone mass and physical function in HIV infected individuals: a randomized controlled trial. *Scand J Med Sci Sports.* 2021;31(3):720-732. doi:10.1111/sms.13871
141. Zanetti HR, da Cruz LG, Lourenço CLM, de F Neves F, Silva-Vergara ML, Mendes EL. Non-linear resistance training reduces inflammatory biomarkers in persons living with HIV: a randomized controlled trial. *Eur J Sport Sci.* 2016a;16(8):1232-1239. doi:10.1080/17461391.2016.1167962
142. Zanetti HR, da Cruz LG, Lourenço CL, Neves FF, Silva-Vergara ML, Mendes EL. Does nonlinear resistance training reduce metabolic syndrome in people living with HIV? A randomized clinical trial. *J Sports Med Phys Fitness.* 2017;57(5):678-684. doi:10.23736/s0022-4707.16.06294-0
143. Bonato M, Turrini F, De Zan V, et al. A mobile application for exercise intervention in people living with HIV. *Med Sci Sports Exerc.* 2020;52(2):425-433. doi:10.1249/MSS.0000000000002125
144. Dudgeon WD, Jagggers JR, Phillips KD, et al. Moderate-intensity exercise improves body composition and improves physiological markers of stress in HIV-infected men. *ISRN AIDS.* 2012;2012:e145127. doi:10.5402/2012/145127
145. Ibeneme SC, Omeje C, Myezwa H, et al. Effects of physical exercises on inflammatory biomarkers and cardiopulmonary function in patients living with HIV: a systematic review with meta-analysis. *BMC Infect Dis.* 2019a;19(1):359. doi:10.1186/s12879-019-3960-0
146. Lu D, Zhang JB, Wang YX, et al. Association between CD4+ T cell counts and gut microbiota and serum cytokines levels in HIV-infected immunological non-responders. *BMC Infect Dis.* 2021;21(1):742. doi:10.1186/s12879-021-06491-z
147. Ezema CI, Onwunali AA, Lamina S, Ezugwu UA, Amaeze AA, Nwankwo MJ. Effect of aerobic exercise training on cardiovascular parameters and CD4 cell count of people living with human immunodeficiency virus/acquired immune deficiency syndrome: a randomized controlled trial. *Niger J Clin Pract.* 2014;17(5):543. doi:10.4103/1119-3077.141414
148. Maduagwu S, Kaidal A, Gashau W, et al. Effect of aerobic exercise on CD4 cell count and lipid profile of HIV infected persons in North Eastern Nigeria. *J AIDS Clin Res.* 2015;06:508. doi:10.4172/2155-6113.1000508
149. de Brito-Neto JG, de Andrade MF, de Almeida VD, et al. Strength training improves body composition, muscle strength and increases CD4+ T lymphocyte levels in people living with HIV/AIDS. *Infect Dis Rep.* 2019;11(1):7925. doi:10.4081/idr.2019.7925
150. Asogwa EI, Obeagu EI, Ekine RS, et al. Effects of 6-weeks moderate intensity aerobic exercise on CD4 count, bone mineral density and weight of people living with HIV/AIDS in Alex-Ekwueme federal university teaching hospital Ebonyi State. *J Pharm Res Int.* 2020;25:13-22. doi:10.9734/jpri/2020/v32i2330784
151. Zanetti HR, Lopes LTP, Gonçalves A, et al. Effects of resistance training on muscle strength, body composition and immune-inflammatory markers in people living with HIV: a systematic review and Meta-analysis of randomized controlled trials. *HIV Res Clin Pract.* 2021;22(5):119-127. doi:10.1080/25787489.2021.1975448
152. Tiozzo E. The Effect of Combined Moderate-Intensity Training on Immune Functioning, Metabolic Variables, and Quality of Life in HIV-infected Individuals Receiving Highly Active Antiretroviral Therapy. Univ Miami. Published Online 2011. Accessed December 15, 2021. <https://www.semanticscholar.org/paper/The-Effect-of-Combined-Moderate-Intensity-Training-Tiozzo/7c556549e41891e4eeab54a7838bc53da93015ed>
153. O'Brien KK, Tynan AM, Nixon SA, Glazier RH. Effectiveness of aerobic exercise for adults living with HIV: systematic review and meta-analysis using the Cochrane Collaboration protocol. *BMC Infect Dis.* 2016;16:182. doi:10.1186/s12879-016-1478-2
154. Ibeneme SC, Irem FO, Iloanusi NI, et al. Impact of physical exercises on immune function, bone mineral density, and quality of life in people living with HIV/AIDS: a systematic review with meta-analysis. *BMC Infect Dis.* 2019;19(1):340. doi:10.1186/s12879-019-3916-4
155. Scheller J, Chalaris A, Schmidt-Arras D, Rose-John S. The pro- and anti-inflammatory properties of the cytokine interleukin-6. *Biochim Biophys Acta BBA - Mol Cell Res* 2011;1813(5):878-888. doi:10.1016/j.bbamcr.2011.01.034
156. Harizi H, Corcuff JB, Gualde N. Arachidonic-acid-derived eicosanoids: roles in biology and immunopathology. *Trends Mol Med.* 2008;14(10):461-469. doi:10.1016/j.molmed.2008.08.005
157. Dennis EA, Norris PC. Eicosanoid storm in infection and inflammation. *Nat Rev Immunol.* 2015;15(8):511-523. doi:10.1038/nri3859
158. Caillaud C, Py G, Eydoux N, Legros P, Prefaut C, Mercier J. Antioxidants and mitochondrial respiration in lung, diaphragm, and locomotor muscles: effect of exercise. *Free Radic Biol Med.* 1999;26(9):1292-1299. doi:10.1016/S0891-5849(98)00342-6
159. Markworth JF, Maddipati KR, Cameron-Smith D. Emerging roles of pro-resolving lipid mediators in

- immunological and adaptive responses to exercise-induced muscle injury. *Exerc Immunol Rev.* 2016;22:110-134.
160. Vella L, Markworth JF, Farnfield MM, Maddipati KR, Russell AP, Cameron-Smith D. Intramuscular inflammatory and resolving lipid profile responses to an acute bout of resistance exercise in men. *Physiol Rep.* 2019;7(13):e14108. doi:10.14814/phy2.14108
161. Stranahan AM, Mattson MP. Exercise-induced hormesis. In: Mattson MP, Calabrese EJ, eds. *Hormesis: A Revolution in Biology, Toxicology and Medicine.* Totowa, NJ: Humana Press; 2010:109-122. doi:10.1007/978-1-60761-495-1_6
162. Mattson MP, Calabrese EJ. Hormesis: what it is and why it matters. In: *Hormesis: A Revolution in Biology, Toxicology and Medicine.* Humana Press Inc; 2010:1-13. doi:10.1007/978-1-60761-495-1_1
163. Degerström J, Østerud B. Increased inflammatory response of blood cells to repeated bout of endurance exercise. *Med Sci Sports Exerc.* 2006;38(7):1297-1303. doi:10.1249/01.mss.0000227315.93351.8d
164. Hill EE, Zack E, Battaglini C, Viru M, Viru A, Hackney AC. Exercise and circulating cortisol levels: the intensity threshold effect. *J Endocrinol Invest.* 2008;31(7):587-591. doi:10.1007/BF03345606
165. Ponce P, Del Arco A, Loprinzi P. Physical activity versus psychological stress: effects on salivary cortisol and working memory performance. *Med Kaunas Lith.* 2019;55(5):E119. doi:10.3390/medicina55050119
166. Sasaki H, Hattori Y, Ikeda Y, et al. Forced rather than voluntary exercise entrains peripheral clocks via a corticosterone/noradrenaline increase in PER2::LUC mice. *Sci Rep.* 2016;6(1):27607. doi:10.1038/srep27607
167. Rosa JC, Lira FS, Eguchi R, et al. Exhaustive exercise increases inflammatory response via Toll like receptor-4 and NF-κBp65 pathway in rat adipose tissue. *J Cell Physiol.* 2011;226(6):1604-1607. doi:10.1002/jcp.22490
168. Clark A, Mach N. Exercise-induced stress behavior, gut-microbiota-brain axis and diet: a systematic review for athletes. *J Int Soc Sports Nutr.* 2016;13:43. doi:10.1186/s12970-016-0155-6
169. Dokladny K, Zuhl MN, Moseley PL. Intestinal epithelial barrier function and tight junction proteins with heat and exercise. *J Appl Physiol.* 2016;120(6):692-701. doi:10.1152/jappphysiol.00536.2015
170. Liao P, He Q, Zhou X, et al. Repetitive bouts of exhaustive exercise induces a systemic inflammatory response and multi-organ damage in rats. *Front Physiol.* 2020;11:685. doi:10.3389/fphys.2020.00685
171. Suzuki K, Tominaga T, Ruhee RT, Ma S. Characterization and modulation of systemic inflammatory response to exhaustive exercise in relation to oxidative stress. *Antioxidants.* 2020;9(5):401. doi:10.3390/antiox9050401
172. Schlabe S, Vogel M, Boesecke C, et al. Moderate endurance training (marathon-training) – effects on immunologic and metabolic parameters in HIV-infected patients: the 42 KM cologne project. *BMC Infect Dis.* 2017;17(1):550. doi:10.1186/s12879-017-2651-y
173. Hillsdon M, Thorogood M, Anstiss T, Morris J. Randomised controlled trials of physical activity promotion in free living populations: a review. *J Epidemiol Community Health.* 1995;49(5):448-453. doi:10.1136/jech.49.5.448
174. Fernandes JL, Serrano CV, Toledo F, et al. Acute and chronic effects of exercise on inflammatory markers and B-type natriuretic peptide in patients with coronary artery disease. *Clin Res Cardiol.* 2011;100(1):77-84. doi:10.1007/s00392-010-0215-x
175. Ke Z, Yip SP, Li L, Zheng XX, Tong KY. The effects of voluntary, involuntary, and forced exercises on brain-derived neurotrophic factor and motor function recovery: a rat brain Ischemia Model. *PLoS One.* 2011;6(2):e16643. doi:10.1371/journal.pone.0016643
176. Cook MD, Martin SA, Williams C, et al. Forced treadmill exercise training exacerbates inflammation and causes mortality while voluntary wheel training is protective in a mouse model of Colitis. *Brain Behav Immun.* 2013;33:46-56. doi:10.1016/j.bbi.2013.05.005
177. Chair SY, Cheng HY, Chew HSJ, Zang YL, Siow EKC, Cao X. Leisure-time physical activity and depressive symptoms among patients with coronary heart disease: the mediating role of physical activity self-efficacy. *Worldviews Evid Based Nurs.* 2020;17(2):144-150. doi:10.1111/wvn.12425
178. Berry A, McCabe CS, Halls S, Muir S, Walsh N. Beliefs, motives and gains associated with physical activity in people with osteoarthritis. *Musculoskeletal Care.* 2021;19(1):52-58. doi:10.1002/msc.1507
179. Bonato M, Galli L, Passeri L, et al. A pilot study of brisk walking in sedentary combination antiretroviral treatment (cART)- treated patients: benefit on soluble and cell inflammatory markers. *BMC Infect Dis.* 2017;17(1):61. doi:10.1186/s12879-016-2095-9
180. Pedro RE, Candido N, Guariglia DA, et al. Exercise improves cytokine profile in HIV-infected people: a randomized clinical trial. *Cytokine.* 2017;99:18-23. doi:10.1016/j.cyto.2017.06.019
181. Zanetti HR, da Cruz LG, Lourenço CL, et al. Nonlinear resistance training enhances the lipid profile and reduces inflammation marker in people living with HIV: a randomized clinical trial. *J Phys Act Health.* 2016;13(7):765-770. doi:10.1123/jpah.2015-0540
182. Zanetti HR, Gonçalves A, Teixeira Paranhos Lopes L, et al. Effects of exercise training and statin use in people living with human immunodeficiency virus with dyslipidemia. *Med Sci Sports Exerc.* 2020;52(1):16-24. doi:10.1249/MSS.0000000000002120
183. Cutrono S, Lewis J, Perry A, Signorile J, Tiozzo E, Jacobs K. The effect of a community-based exercise program on inflammation, metabolic risk, and fitness levels among persons living with HIV/AIDS. *AIDS Behav.* 2016;20:1123-1131. doi:10.1007/s10461-015-1245-1
184. Vingren JL, Curtis JH, Levitt DE, et al. Adding resistance training to the standard of care for inpatient substance abuse treatment in men with human immunodeficiency virus improves skeletal muscle health without altering cytokine concentrations. *J Strength Cond Res.* 2018;32(1):76-82. doi:10.1519/JSC.0000000000002289
185. Dianatinasab M, Fararouei M, Padehban V, et al. The effect of a 12-week combinational exercise program on CD4 count and mental health among HIV infected women: a randomized control trial. *J Exerc Sci fit.* 2018;16(1):21-25. doi:10.1016/j.jesf.2018.02.001