**GENOME SEQUENCES** 





## Complete Genome Sequences and Characteristics of Seven Novel Mycobacteriophages Isolated in East Texas

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**ABSTRACT** Full-genome sequences of seven mycobacteriophages isolated from environmental soil samples are presented. These bacteriophages, with their respective clusters or subclusters are Duplo (A2), Dynamo (P1), Gilberta (A11), MaCh (A11), Nikao (K1), Phloss (N), and Skinny (M1). All had siphovirus-like morphologies, with genome sizes ranging from 43,107 to 82,071 bp.

**B** acteriophages are viruses that exclusively infect bacteria, exhibiting obligate intracellular replication and a limited host range (1, 2). Bacteriophages are the most numerous entities in the biosphere, totaling  $>10^{31}$  particles (3). The bacterium-bacteriophage relationship exhibits constant bidirectional selective pressure, with bacteria evolving to resist viral infection and bacteriophages coevolving to maintain their replicative ability within their hosts (4, 5). The prevalence of antibiotic-resistant bacterial infections has propelled a resurgent interest in phage therapy (6–10). Here, we report seven novel lysogenic bacteriophages.

All bacteriophages were isolated from environmental soil samples collected from various locations in east Texas during 2020 to 2021 (Table 1), using standard methods (5). In short, the soil samples were washed in Middlebrook 7H9 medium prior to centrifugation and supernatant filtration (pore size, 0.22  $\mu$ m). The filtrates were subsequently inoculated with *Mycobacterium smegmatis* mc<sup>2</sup>155 cells and incubated at 25°C for 3.5 days with shaking. Filtered samples of each culture were plated with *M. smegmatis* cells in 7H9 top agar. After purification through three rounds of plating at 37°C for 48 h, the observed plaque morphologies of the various phages ranged from clear to turbid (Table 1). Negative-stain transmission electron microscopy showed all these bacteriophages to exhibit a siphovirus morphotype, with isometric capsids (diameter, ~51.76 to 71.53 nm) and noncontractile tails (length, ~124.23 to 326.19 nm) (Fig. 1 and Table 1), measured using ImageJ (11–13).

Genomic DNA was extracted from lysates of various titers (Table 1) using the Promega Wizard DNA cleanup kit. Preparation for sequencing using the Illumina MiSeq platform (v3 reagents) was conducted with the NEBNext Ultra II library prep kit. Assembly and verification of the untrimmed reads were performed using Newbler v2.9 (16) and Consed v29 (14, 15). Sequencing revealed genomes ranging in length from 43,107 bp (phage Phloss) to 82,071 bp (phage Skinny) (Table 1). All had 3' sticky overhangs (10 to 13 bp long) and an average GC content of 64.7% (range, 61.5% to 67.2%), comparable to the 67.4% GC content of their isolation host, *Mycobacterium smegmatis* mc<sup>2</sup>155 (17). The seven phages were assigned to subclusters A2, A11, K1, M1, and P1 and cluster N (Table 1) based on  $\geq$ 35% gene content similarity (GCS) to other phages, using the GCS tool in PhagesDB (18, 19).

Genome annotation was accomplished using DNA Master v5.23.6; Starterator; Phamerator (20); BLASTp with NCBI GenBank and PhagesDB (21, 22); GeneMark v2.5p (23); HHpred, with the PDB\_mmCIF70\_17\_Apr, Pfam-A\_v35, UniProt-SwissProt-viral70\_3\_Nov\_2021, and NCBI\_Conserved\_Domains\_v3.19 databases (24, 25); Glimmer v3.02 (26); TMHMM v.2.0 (27); SOSUI v1.11 (28); tRNAscan-SE v2.0 (29, 30); and ARAGORN v1.2.41 (31). Default settings were used for all programs (32). An average of 98.0 putative protein-coding genes (range, 70 to 163) and

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	Data for phage: <sup>a</sup>						
Characteristic	Duplo	Dynamo	Gilberta	MaCh	Nikao	Phloss	Skinny
Yr found	2020	2021	2021	2020	2020	2020	2021
Location found	Big Sandy, TX	Longview, TX	Longview, TX	Longview, TX	Longview, TX	Longview, TX	Longview, TX
Soil sampling location	32.588034 N,	32.46402 N,	32.468338 N,	32.465237 N,	32.54954 N,	32.4675 N, 94.725 W	32.467601 N,
	95.063959 W	94.728226 W	94.726844 W	94.727035 W	94.821309 W	:	94.723749 W
Lysate titer (PFU/mL)	$3.0  imes 10^9$	$1.0 \times 10^{10}$	$1.0  imes 10^{9}$	$1.6 \times 10^{10}$	$2.17  imes 10^9$	$8.7 \times 10^{11}$	$1.18  imes 10^9$
Plaque morphology after 48 h	Slightly turbid with	Clear with defined	Clear with defined	Clear with defined	Slightly turbid with	Clear with defined	Clear with defined
at 37°C	defined edges	edges	edges	edges	defined edges	edges	edges
Plaque size (mm)	3.3	0.93	1.5	2.7	1.25	2.0	1.0
Approx coverage ( $ imes$ )	1,516	1,499	5,472	705	414	2,005	499
Genome size (bp)	52,781	46,673	51,470	52,616	59,052	43,107	82,071
GC content (%)	63.5	67.2	63.7	63.6	67.2	66.3	61.5
Overhang sequence	CGGTCGGTTA	CCCGCCCCCCGA	CGGTCGGTTA	CGGTCGGTTA	CTCGGGGGGCAT	CCCGCCGCAATGG	ACCTCCTGCAA
Overhang length (bases)	10	12	10	10	11	13	11
Cluster	A	Ь	A	A	×	Z	Μ
Subcluster	A2	P1	A11	A11	K1		M1
GenBank accession no.	OP297553	OP434454	OP297532	OP297549	OP297530	OP297540	OP297551
SRA accession no.	SRX19690833	SRX19690834	SRX19690836	SRX19690844	SRX19690847	SRX19690850	SRX19690858
Total no. of reads	560,502	485,417	1,968,556	261,625	172,343	608,118	285,773
No. of predicted genes	66	78	66	100	80	70	163
No. of predicted tRNAs	5	0	-	-	0	0	18
tRNA type(s)	Asn, Gln, Glu, Trp, Tyr		Trp	Trp			Trp, Asn, other (Arg/Ala), Tyr, Gln, Pro, Ser, Phe, Met, Arg, His, Leu, Lys, Glv. Val. Thr. Aso. Glu
No. of genes with predicted functions	33	32	36	43	41	28	46
% of genes with predicted functions	33	41	36	43	51	40	28
Key predicted lysogenic life	Integrase, excise,	Integrase, excise,	Integrase, immunity	Immunity repressor	Integrase, excise,	Integrase, excise,	Integrase
cycle genes	immunity repressor	immunity repressor	repressor		immunity repressor	immunity repressor	
No. of orphams	2	0	-	0	0	0	2
Capsid size (nm [ <i>n</i> value])	64.48 (9)	(2) (2) (2) (2) (2) (2) (2) (2) (2) (2)	52.09 (3)	58.67 (21)	67.04 (5)	51.76 (7)	71.53 (3)
Avg tail length (nm [ <i>n</i> value])	124.23 (9)	205.57 (6)	125.54 (3)	130.93 (21)	210.16 (5)	172.04 (7)	326.19 (3)
solated by	Skylar M. Weiss, Jimena H. Segovia	Christina A. Holder, Kaitlyn J. Menard, Brady E. Tyler	Hattie R. Mills, Ashlyn B. Collier, Kalista J. Rivera, Claire P. Martinez	Matthew S. Adams, Camryn L. Hill	Kezia K. Happy	Summer L. Apostalo, Gavin J. Meyer	Jenna F. Curran, Kristen N. Rose
<sup>2</sup> All phages were isolated using the base single-end reads using the NE verified manually using Consed v2	enriched isolation method B Ultra II Library sequencir 9 (14, 15). All phages had a	l (5) and purified through tl ng kit. All had 3' sticky over siphovirus morphotype an	rree sequential 48-h rounds hang genome ends. Genomi d were predicted to be temp	of plating at 37°C. The genc c termini were identified th perate based on the presenc	omes were sequenced using irough buildups of read star ce of predicted lysogeny-rel	J the Illumina shotgun sequ t positions and variations ir ated genes.	iencing method with 150- n genome-wide coverage and

TABLE 1 Properties of seven mycobacteriophages isolated in east Texas, USA, in 2020 and 2021

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**FIG 1** Transmission electron micrographs of the seven bacteriophages, Duplo (A), Dynamo (B), Gilberta (C), MaCh (D), Nikao (E), Phloss (F), and Skinny (G). Capsid sizes and tail lengths are provided in Table 1. Bacteriophage particles were added to 300-mesh carbon–Formvar-coated copper grids (Ted Pella Inc., Redding, CA), stained with 1% (wt/vol) uranyl acetate, and imaged at the University of Arkansas for Medical Sciences Digital Microscopy Laboratory.

3.6 tRNAs (range, 0 to 18) were predicted (Table 1). Functions could only be predicted for 28% to 51% of the putative genes in the phages (Table 1). All phages had at least one of the three key genes associated with a lysogenic life cycle. Duplo, Dynamo, Nikao, and Phloss had the integrase, excise, and immunity repressor genes; Gilberta had both the integrase and immunity repressor genes, while Skinny and MaCh had only the integrase and immunity repressor genes, respectively (Table 1).

**Data availability.** The raw reads of all seven reported mycobacteriophages are available in the Sequence Read Archive (SRA), and their complete genome sequences are available at GenBank. The SRA and GenBank accession numbers are provided in Table 1. High-titer lysates of the phages are archived at the University of Pittsburgh Bacteriophage Institute.

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