

Complete Genome Sequence of Legionella cardiaca Strain H63^T, Isolated from a Case of Native Valve Endocarditis

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ABSTRACT We report the complete genome sequence of Legionella cardiaca strain H63^T, which had been isolated from aortic valve tissue from a patient with native endocarditis. The genome assembly contains a single $3,477,232$ -bp contig, with a $G+C$ content of 38.59%, and is predicted to encode 2,948 proteins.

mong the extrapulmonary manifestations of Legionella infection is endocarditis ([1](#page-2-0), [2\)](#page-2-1). Our laboratory previously described a novel isolate that had been obtained by plating material from resected aortic valve tissue on buffered charcoal yeast extract (BCYE) agar at 37°C and was named Legionella cardiaca strain H63^T (ATCC BAA-2315) [\(3](#page-2-2), [4](#page-2-3)). Because prior genotypic analysis of L. cardiaca involved only DNA-DNA hybridization and phylogenetic analyses of three loci ([4](#page-2-3)) and the mechanisms of Legionella endocarditis are unknown, we determined the complete genome of L. cardiaca H63^T.

Using the Promega Maxwell 16 system, DNA was isolated from H63^T, which had been grown from a single colony to confluence on BCYE agar at 37°C for 3 days. DNA was sequenced using Illumina and Pacific Biosciences (PacBio) platforms. For Illumina sequencing, short-read libraries were generated with a KAPA HyperPrep kit (Roche) and sequenced using 150-bp paired-end reads on a NovaSeq 6000 system. For PacBio sequencing, genomic DNA (gDNA) was fragmented to an average size of \sim 11 kb with a Covaris g-TUBE. DNA was cleaned with SPRIselect beads, followed by library construction using the SMRTbell Express template preparation kit v2.0 (PacBio), which includes single-strand DNA overhang removal, DNA damage repair, end repair/A-tailing, and barcoded overhang adaptor ligation. The library was pooled with other libraries on an equimolar basis and subsequently size selected on a BluePippin instrument with an 8-kb cutoff value. The library pool was purified with SPRIselect beads, quantified with a Qubit 4.0 fluorometer, and assessed with an Agilent fragment analyzer. The final library pool was sequenced with PacBio Sequel II v2.0 chemistry and a single-molecule real-time (SMRT) Cell 8M on a Sequel II instrument at an on-plate concentration of 85 pM. Illumina reads were quality filtered using a combination of Illumina RTA v1.8.70.0 and Trimmomatic v0.38.0 ([5\)](#page-2-4). PacBio reads were quality filtered using FastQC v0.72 [\(6](#page-2-5)). For Illumina sequencing, 5,802,382 reads were generated, with \sim 250 \times coverage; for PacBio sequencing, 1,609,389 reads (N_{50} , 11,074 bp) were generated. The resulting raw sequencing reads were processed using PacBio SMRTLink v9.0, including demultiplexing by Lima v1.11.0 [\(7\)](#page-2-6). Genome assembly was performed using the PacBio HGAP4 assembler, which includes overlap determination, followed by consensus polishing with Pilon v1.24 [\(8\)](#page-2-7) using Illumina 150-bp paired-end reads generated from the same gDNA. Rotation of the chromosome was performed using the IGS automated prokaryotic annotation pipeline [\(9\)](#page-2-8). The assembly yielded a single closed circular chromosome. Gene annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.4 ([10](#page-2-9)[–](#page-2-10)[12](#page-2-11)).

The H63^T genome assembly contains a single 3,477,232-bp contig (\sim 4,411 \times coverage), with a $G+C$ content of 38.593%, and is predicted to encode 2,948 proteins. A rooted species tree based on the concatenated amino acid alignment of 219 single-copy orthologous proteins was generated using OrthoFinder v2.5.4 [\(13](#page-2-12)–[16\)](#page-2-13), and strain H63T was most closely Editor Vanja Klepac-Ceraj, Wellesley College Copyright © 2023 Lopez et al. This is an openaccess article distributed under the terms of the [Creative Commons Attribution 4.0](https://creativecommons.org/licenses/by/4.0/) [International license](https://creativecommons.org/licenses/by/4.0/).

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FIG 1 Relationships of L. cardiaca H63^T with 63 other sequenced species of Legionella. In the rooted species tree, L. cardiaca is highlighted in red and the other Legionella species and their corresponding strain names appear in black. Appearing at the top of the tree are non-Legionella species (in brown) that belong to other genera within the order Legionellales. Bar, 0.1 amino acid substitutions per site.

TABLE 1 ANI values from pairwise comparisons between the genome of L. cardiaca strain H63^T and the genomes of the Legionella species most closely related to strain H63^T

related to Legionella brunensis ([Fig. 1](#page-1-0)). Pairwise average nucleotide identity (ANI) comparisons $(17–21)$ $(17–21)$ $(17–21)$ $(17–21)$ confirmed that L. cardiaca is a distinct species within the L. brunensis-containing clade [\(Table 1\)](#page-2-15), which is linked to disease [\(4,](#page-2-3) [22](#page-3-1)-[29\)](#page-3-2). Consistent with $H63^T$ being virulent in infection models [\(4](#page-2-3)), its genome has genes encoding a type IVB secretion system and a type II secretion system and genes linked to iron assimilation ([30](#page-3-3)–[38\)](#page-3-4).

Data availability. The assembly of the genome is available under GenBank accession number [CP119078,](https://www.ncbi.nlm.nih.gov/nuccore/CP119078) and raw reads have been submitted to the NCBI SRA under accession numbers [SRR23636844](https://www.ncbi.nlm.nih.gov/sra/SRR23636844) and [SRR23636845.](https://www.ncbi.nlm.nih.gov/sra/SRR23636845)

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