



## Complete Genome Sequence of *Legionella cardiaca* Strain H63<sup>T</sup>, Isolated from a Case of Native Valve Endocarditis

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**ABSTRACT** We report the complete genome sequence of *Legionella cardiaca* strain  $H63^{T}$ , which had been isolated from aortic valve tissue from a patient with native endocarditis. The genome assembly contains a single 3,477,232-bp contig, with a G+C content of 38.59%, and is predicted to encode 2,948 proteins.

Mong the extrapulmonary manifestations of *Legionella* infection is endocarditis (1, 2). Our laboratory previously described a novel isolate that had been obtained by plating material from resected aortic valve tissue on buffered charcoal yeast extract (BCYE) agar at 37°C and was named *Legionella cardiaca* strain H63<sup>T</sup> (ATCC BAA-2315) (3, 4). Because prior genotypic analysis of *L. cardiaca* involved only DNA-DNA hybridization and phylogenetic analyses of three loci (4) and the mechanisms of *Legionella* endocarditis are unknown, we determined the complete genome of *L. cardiaca* H63<sup>T</sup>.

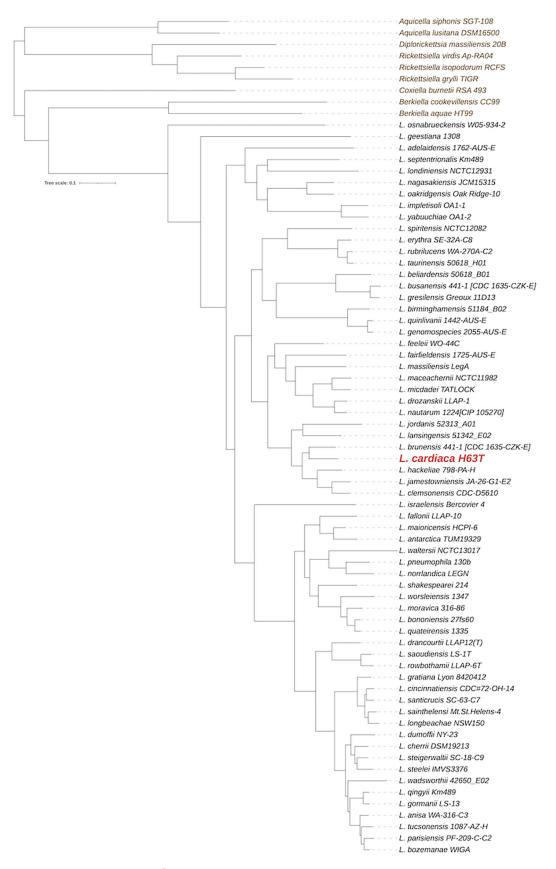
Using the Promega Maxwell 16 system, DNA was isolated from H63<sup>T</sup>, which had been grown from a single colony to confluence on BCYE agar at 37°C for 3 days. DNA was sequenced using Illumina and Pacific Biosciences (PacBio) platforms. For Illumina sequencing, short-read libraries were generated with a KAPA HyperPrep kit (Roche) and sequenced using 150-bp paired-end reads on a NovaSeq 6000 system. For PacBio sequencing, genomic DNA (gDNA) was fragmented to an average size of  $\sim$ 11 kb with a Covaris g-TUBE. DNA was cleaned with SPRIselect beads, followed by library construction using the SMRTbell Express template preparation kit v2.0 (PacBio), which includes single-strand DNA overhang removal, DNA damage repair, end repair/A-tailing, and barcoded overhang adaptor ligation. The library was pooled with other libraries on an equimolar basis and subsequently size selected on a BluePippin instrument with an 8-kb cutoff value. The library pool was purified with SPRIselect beads, quantified with a Qubit 4.0 fluorometer, and assessed with an Agilent fragment analyzer. The final library pool was sequenced with PacBio Sequel II v2.0 chemistry and a single-molecule real-time (SMRT) Cell 8M on a Sequel II instrument at an on-plate concentration of 85 pM. Illumina reads were quality filtered using a combination of Illumina RTA v1.8.70.0 and Trimmomatic v0.38.0 (5). PacBio reads were quality filtered using FastQC v0.72 (6). For Illumina sequencing, 5,802,382 reads were generated, with  $\sim$ 250× coverage; for PacBio sequencing, 1,609,389 reads ( $N_{50}$ , 11,074 bp) were generated. The resulting raw sequencing reads were processed using PacBio SMRTLink v9.0, including demultiplexing by Lima v1.11.0 (7). Genome assembly was performed using the PacBio HGAP4 assembler, which includes overlap determination, followed by consensus polishing with Pilon v1.24 (8) using Illumina 150-bp paired-end reads generated from the same gDNA. Rotation of the chromosome was performed using the IGS automated prokaryotic annotation pipeline (9). The assembly yielded a single closed circular chromosome. Gene annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.4 (10–12).

The H63<sup>T</sup> genome assembly contains a single 3,477,232-bp contig ( $\sim$ 4,411× coverage), with a G+C content of 38.593%, and is predicted to encode 2,948 proteins. A rooted species tree based on the concatenated amino acid alignment of 219 single-copy orthologous proteins was generated using OrthoFinder v2.5.4 (13–16), and strain H63<sup>T</sup> was most closely

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**FIG 1** Relationships of *L. cardiaca*  $H63^{T}$  with 63 other sequenced species of *Legionella*. In the rooted species tree, *L. cardiaca* is highlighted in red and the other *Legionella* species and their corresponding strain names appear in black. Appearing at the top of the tree are non-*Legionella* species (in brown) that belong to other genera within the order *Legionellales*. Bar, 0.1 amino acid substitutions per site.

Subject strain for query	GenBank assembly accession no.	ANI (%)
Legionella brunensis	GCF_001467025.1	76.56
Legionella hackeliae	GCF_001467705.1	74.96
Legionella clemsonensis	GCF_002240035.1	74.93
Legionella jamestowniensis	GCF_900640205.1	74.39
Legionella lansingensis	GCF_000622185.1	74.22
Legionella jordanis	GCF_001467765.1	71.95
Legionella feeleii	GCF_001467625.1	72.46
Legionella fairfieldensis	GCF_000621525.1	72.05
Legionella drozanskii	GCF_001467585.1	71.81
Legionella nautarum	GCF_001467895.1	71.77
Legionella maceachernii	GCF_001467845.1	71.55
Legionella micdadei	GCF_001467875.1	71.36
Legionella massiliensis	GCF_000756815.1	71.09

**TABLE 1** ANI values from pairwise comparisons between the genome of *L. cardiaca* strain  $H63^{T}$  and the genomes of the *Legionella* species most closely related to strain  $H63^{T}$ 

related to *Legionella brunensis* (Fig. 1). Pairwise average nucleotide identity (ANI) comparisons (17–21) confirmed that *L. cardiaca* is a distinct species within the *L. brunensis*-containing clade (Table 1), which is linked to disease (4, 22–29). Consistent with H63<sup>T</sup> being virulent in infection models (4), its genome has genes encoding a type IVB secretion system and a type II secretion system and genes linked to iron assimilation (30–38).

**Data availability.** The assembly of the genome is available under GenBank accession number CP119078, and raw reads have been submitted to the NCBI SRA under accession numbers SRR23636844 and SRR23636845.

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## REFERENCES

- Teira A, Sanchez J, Santiago I, Zarauza J, Nan D, Teira R. 2022. *Legionella* endocarditis: a case report and review. Enferm Infecc Microbiol Clin (Engl Ed) 40:190–194. https://doi.org/10.1016/j.eimc.2020.10.022.
- Fraz MSA, Dahle G, Skaug KM, Jarraud S, Frye S, Bjørnholt JV, Nordøy I. 2022. Case report: a prosthetic valve endocarditis caused by *Legionella bozemanae* in an immunocompetent patient. Front Med (Lausanne) 9:1055465. https:// doi.org/10.3389/fmed.2022.1055465.
- Pearce MM, Theodoropoulos N, Noskin GA, Flaherty JP, Stemper ME, Aspeslet T, Cianciotto NP, Reed KD. 2011. Native valve endocarditis due to a novel strain of *Legionella*. J Clin Microbiol 49:3340–3342. https://doi.org/10.1128/ JCM.01066-11.
- Pearce MM, Theodoropoulos N, Mandel MJ, Brown E, Reed KD, Cianciotto NP. 2012. *Legionella cardiaca* sp. nov., isolated from a case of native valve endocarditis in a human heart. Int J Syst Evol Microbiol 62:2946–2954. https://doi .org/10.1099/ijs.0.039248-0.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. https://www.bioinformatics.babraham.ac.uk/projects/fastqc.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, errorprone reads using repeat graphs. Nat Biotechnol 37:540–546. https://doi .org/10.1038/s41587-019-0072-8.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal.pone.0112963.
- Galens K, Orvis J, Daugherty S, Creasy HH, Angiuoli S, White O, Wortman J, Mahurkar A, Giglio MG. 2011. The IGS standard operating procedure for automated prokaryotic annotation. Stand Genomic Sci 4:244–251. https://doi .org/10.4056/sigs.1223234.

- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/ nar/gkw569.
- Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. Nucleic Acids Res 46:D851–D860. https://doi.org/10.1093/nar/gkx1068.
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. Nucleic Acids Res 49: D1020–D1028. https://doi.org/10.1093/nar/gkaa1105.
- Emms DM, Kelly S. 2015. OrthoFinder: solving fundamental biases in whole genome comparisons dramatically improves orthogroup inference accuracy. Genome Biol 16:157. https://doi.org/10.1186/s13059-015-0721-2.
- Emms DM, Kelly S. 2019. OrthoFinder: phylogenetic orthology inference for comparative genomics. Genome Biol 20:238. https://doi.org/10.1186/ s13059-019-1832-y.
- Lubna Asaf S, Jan R, Khan AL, Bilal S, Asif S, Al-Harrasi A, Kim KM. 2022. Unraveling the genome sequence of plant growth promoting *Aspergillus niger* (CSR3) provides insight into the synthesis of secondary metabolites and its comparative genomics. J Fungi (Basel) 8:107. https://doi.org/10.3390/jof8020107.
- Webster J, Dadd-Daigle P, Chapman TA, Kirkby K. 2023. Investigating the defoliating-like VCG2A pathotype of *Verticillium dahliae* through identification and prediction of secreted proteins from genomes of Australian isolates. Plant Pathol 72:334–341. https://doi.org/10.1111/ppa.13660.
- Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM. 2007. DNA-DNA hybridization values and their relationship to whole-genome

sequence similarities. Int J Syst Evol Microbiol 57:81–91. https://doi.org/10 .1099/ijs.0.64483-0.

- Richter M, Rossello-Mora R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci U S A 106:19126–19131. https://doi.org/10.1073/pnas.0906412106.
- Richter M, Rossello-Mora R, Oliver Glockner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 32:929–931. https://doi.org/10.1093/ bioinformatics/btv681.
- Girolamini L, Pascale MR, Mazzotta M, Spiteri S, Marino F, Salaris S, Grottola A, Orsini M, Cristino S. 2022. Combining traditional and molecular techniques supports the discovery of a novel *Legionella* species during environmental surveillance in a healthcare facility. Front Microbiol 13:900936. https://doi.org/10 .3389/fmicb.2022.900936.
- Crespi S, Drasar V, Salva-Serra F, Jaen-Luchoro D, Pineiro-Iglesias B, Lindemann PC, Aliaga-Lozano F, Fernández-Juárez V, Coll-García G, Moore ERB, Bennasar-Figueras A. 2023. *Legionella maioricensis* sp. nov., a new species isolated from the hot water distribution systems of a hospital and a shopping center during routine culturing. Int J Syst Evol Microbiol 73:e005686. https://doi.org/10 .1099/ijsem.0.005686.
- Wilkinson HW, Drasar V, Thacker WL, Benson RF, Schindler J, Potuznikova B, Mayberry WR, Brenner DJ. 1988. *Legionella moravica* sp. nov. and *Legionella brunensis* sp. nov. isolated from cooling-tower water. Ann Inst Pasteur Microbiol 139:393–402. https://doi.org/10.1016/0769-2609(88)90102-0.
- 23. Wilkinson HW, Thacker WL, Steigerwalt AG, Brenner DJ, Ampel NM, Wing EJ. 1985. Second serogroup of *Legionella hackeliae* isolated from a patient with pneumonia. J Clin Microbiol 22:488–489. https://doi.org/10.1128/jcm.22 .4.488-489.1985.
- Palmer A, Painter J, Hassler H, Richards VP, Bruce T, Morrison S, Brown E, Kozak-Muiznieks NA, Lucas C, McNealy TL. 2016. *Legionella clemsonensis* sp. nov.: a green fluorescing *Legionella* strain from a patient with pneumonia. Microbiol Immunol 60:694–701. https://doi.org/10.1111/1348-0421.12439.
- Prochazka B, Indra A, Hasenberger P, Blaschitz M, Wagner L, Wewalka G, Sorschag S, Schmid D, Ruppitsch W. 2016. Draft genome sequence of *Legionella jamestowniensis* isolated from a patient with chronic respiratory disease. Genome Announc 4:e01007-16. https://doi.org/10.1128/genomeA.01007-16.
- Edelstein PH. 2017. Legionella jamestowniensis fatal pneumonia in an immunosuppressed man. J Infect Chemother 23:59–61. https://doi.org/ 10.1016/j.jiac.2016.07.015.
- Thacker WL, Dyke JW, Benson RF, Havlichek DH, Robinson-Dunn B, Stiefel H, Schneider W, Moss CW, Mayberry WR, Brenner DJ. 1992. *Legionella lansingensis* sp. nov. isolated from a patient with pneumonia and underlying

chronic lymphocytic leukemia. J Clin Microbiol 30:2398–2401. https://doi .org/10.1128/jcm.30.9.2398-2401.1992.

- Vinh DC, Garceau R, Martinez G, Wiebe D, Burdz T, Reimer A, Bernard K. 2007. Legionella jordanis lower respiratory tract infection: case report and review. J Clin Microbiol 45:2321–2323. https://doi.org/10.1128/JCM.00314-07.
- Ricketts KD, Joseph CA, European Working Group for Legionella Infections. 2007. Legionnaires disease in Europe: 2005–2006. Euro Surveill 12: E7–E8. https://doi.org/10.2807/esm.12.12.00753-en.
- Burstein D, Amaro F, Zusman T, Lifshitz Z, Cohen O, Gilbert JA, Pupko T, Shuman HA, Segal G. 2016. Genomic analysis of 38 *Legionella* species identifies large and diverse effector repertoires. Nat Genet 48:167–175. https://doi.org/10.1038/ng.3481.
- Wexler M, Zusman T, Linsky M, Lifshitz Z, Segal G. 2022. The Legionella genus core effectors display functional conservation among orthologs by themselves or combined with an accessory protein. Curr Res Microb Sci 3: 100105. https://doi.org/10.1016/j.crmicr.2022.100105.
- White RC, Cianciotto NP. 2019. Assessing the impact, genomics, and evolution of type II secretion across a large, medically-important genus: the *Legionella* type II secretion paradigm. Microb Genom 5:e000273. https://doi.org/10.1099/mgen.0.000273.
- Hickey EK, Cianciotto NP. 1994. Cloning and sequencing of the Legionella pneumophila fur gene. Gene 143:117–121. https://doi.org/10.1016/0378 -1119(94)90615-7.
- 34. Hickey EK, Cianciotto NP. 1997. An iron- and *fur*-repressed *Legionella* pneumophila gene that promotes intracellular infection and encodes a protein with similarity to the *Escherichia coli* aerobactin synthetases. Infect Immun 65:133–143. https://doi.org/10.1128/iai.65.1.133-143.1997.
- Robey M, Cianciotto NP. 2002. *Legionella pneumophila feoAB* promotes ferrous iron uptake and intracellular infection. Infect Immun 70:5659–5669. https://doi.org/10.1128/IAI.70.10.5659-5669.2002.
- Portier E, Zheng H, Sahr T, Burnside DM, Mallama C, Buchrieser C, Cianciotto NP, Héchard Y. 2015. *IroT/mavN*, a new iron-regulated gene involved in *Legionella pneumophila* virulence against amoebae and macrophages. Environ Microbiol 17:1338–1350. https://doi.org/10.1111/1462-2920.12604.
- O'Connor TJ, Zheng H, VanRheenen SM, Ghosh S, Cianciotto NP, Isberg RR. 2016. Iron limitation triggers early egress by the intracellular bacterial pathogen *Legionella pneumophila*. Infect Immun 84:2185–2197. https:// doi.org/10.1128/IAI.01306-15.
- Zheng H, Chatfield CH, Liles MR, Cianciotto NP. 2013. Secreted pyomelanin of *Legionella pneumophila* promotes bacterial iron uptake and growth under iron-limiting conditions. Infect Immun 81:4182–4191. https://doi.org/10.1128/ IAI.00858-13.