ORIGINAL RESEARCH



In-silico mining to glean SNPs of pharmaco-clinical importance: an investigation with reference to the Indian populated SNPs

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Abstract

Drugs pharmacology is defined by pharmacokinetics and pharmacodynamics and both of them are affected by genetic variability. Genetic variability varies from population to population, and sometimes even within the population, it exists. Single nucleotide polymorphisms (SNPs) are one of the major genetic variability factors which are found to be associated with the pharmacokinetics and pharmacodynamics process of a drug and are responsible for variable drug response and clinical phenotypes. Studies of SNPs can help to perform genome-wide association studies for their association with pharmacological and clinical events, at the same time; their information can direct genome-wide association studies for their association studies for their use as biomarkers. With the aim to mine and characterize Indian populated SNPs of pharmacological and clinical importance. Two hundred six candidate SNPs belonging to 43 genes were retrieved from Indian Genome Variation Database. The distribution pattern of considered SNPs was observed against all five world super-populations (AFR, AMR, EAS, EUR, and SAS). Further, their annotation was done through SNP-nexus by considering Human genome reference builds - hg38, pharmacological and clinical information was supplemented by PharmGKB and ClinVar database. At last, to find out the association between SNPs linkage disequilibrium was observed in terms of r². Overall, the study reported 53 pharmaco-clinical active SNPs and found 24 SNP-pairs as potential markers, and recommended their clinical and experimental validation.

Keywords Single nucleotide polymorphisms · Drug response · Linkage disequilibrium · Molecular marker · Haplotype

Introduction

Drug pharmacology defines in terms of pharmacokinetics (PK) and pharmacodynamics (PD) is a very important aspect of any drug; it decides the fate of the drug as well as its effect on patients (Glassman and Muzykantov 2019). It has been observed that genes that are involved in pharmacokinetics and pharmacodynamics, known as pharmacogenes (PGx-genes), also harbor a range of SNPs (Katara and Yadav 2019). Few of these SNPs are very critical, and their presence alters the structure and activity of their protein's product. The presence of these SNP variants brings

Pramod Katara pmkatara@gmail.com; pkatara@allduniv.ac.in variability in the functionality of pharmacogenes which mainly affects drug's PK and results in variable drug response (VDR) among patients. The more critical situation arises in some circumstances where this VDR resulted adversely and causes adverse drug response (ADR), which is lethal in some cases (Roden et al. 2011; Arbitrio et al. 2021).

The Genome of every person is unique and differs from other individuals even among the members of the same population, pharmacogenomics comes with the possible solution and proposed practice of genetic information in pharmaceutical and clinical care practice (Roden et al. 2019; Katara 2014). Pharmacogenetic testing (pharmacogenotyping) and genome-based medicine (personalized medicine) are the two possible ways to reduce the probability of ADR and enhance the possibilities of precise medication. Both, pharmacogenotyping and personalized medicine needs the use of genomic variant information, especially the variants of pharmacological and clinical (pharmaco-clinical) importance (González-Covarrubias et al. 2020). Contemporary

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studies are mainly relying on SNPs for these purposes (Ji et al. 2021). Numbers of SNPs are already in practice for various pharmacological and clinical screenings such as rs4917639 (CYP2C9*2) and rs1057910 (CYP2C9*3) for Cardiovascular Drugs (Crews et al. 2012; Pandey et al. 2020; Yadav et al. 2022; Yadav and Katara 2018). As studies reveal that most of these SNPs have a specific distribution pattern in the global population and show inter-population genetic variations, thus drugs, as well as markers used for pharmacogenomics purposes in one population, may not be appropriate for another population (Lauschke et al. 2017). Considering the facts, it is essential to include ethnicity in pharmacogenomics to ensure proper dosing, drug efficacy, and to develop pharmacogenomics tests (Sosa-Macías et al. 2016; Katara 2013).

Those SNPs which are associated with phenotypes are generally shown to have a non-random association with alleles of different loci. Linkage disequilibrium (LD) is one of the parameters used to measure the non-random association between alleles of different loci, which provides information about the set of SNPs that tend to be inherited together (Crossline et al. 2010). Population-specific LD patterns among genetic variants in different populations can provide information about genetic relatedness between the studied populations (Suarez-Kurtz and Parra 2018). Knowledge about LD patterns among SNPs allows Genome-wide association study (GWAS) to identify and use biomarkers in a population-specific manner, i.e., tagSNPs (Uffelmann et al. 2021). Since allelic distribution and LD of SNPs vary among populations, mining of pharmacogenomics-related SNPs allele frequencies and their LD patterns are crucial and anticipated for association studies (De et al. 2002; Uffelmann et al. 2021).

Population-specific genome projects across the globe that produce a considerable size of sequence data allow scientists to analyze and get the variant patterns of importance (Van Rooij et al. 2017). India is one of the most densely populated South Asian countries whose population shows huge diversity among themselves as well as from the rest of the world (Bongaarts 2009; Jain et al. 2021). Despite this huge diversity, very less is studied on the Indian population, specifically with respect to clinical genomics (GUaRD-IAN Consortium et al. 2019). The current study focused on the analysis of Indian population-specific SNP variants of pharmaco-clinical importance. The study mainly includes a distribution pattern to identify Indian population-specific SNPs and SNP-variants, followed by their pharmaco-clinical annotation. Along with that, pair-wise LD analysis was also performed to find out SNP associations and to observe the possibility to use them as markers.

Methods

Mining of Indian population-specific SNPs belongs to the genes of pharmacokinetics importance

A list of pharmacogenes belonging to pharmacokinetics was collected from PharmGKB (Thorn et al. 2013), https://www.pharmgkb.org/. Indian populated SNP-mining for all the PK-genes was done from the Indian genome variation database (IGVDB, Indian Genome Variation Consortium 2005; http://www.igvdb.res.in). SNP-Variant IDs (rsIDs) and their major and minor allele information were utilized for the whole analysis. A total of 43 genes from 26 gene families comprising 206 SNPs have been investigated (Table 1).

Population-specific distribution analysis of selected SNPs

To infer population-specific worldwide distribution patterns of Indian-SNPs a comparative approach was used against all five super-populations, i.e., African (AFR), American (AMR), East-Asian (EAS), European (EUR), and South-Asian (SAS). Each variant's allele frequency was retrieved from the population genetics database using an ensemble genome browser. These populations' data were obtained from Phase 3 of the 1000 Genome Project https://www.internationalgenome.org/data (The 1000 Genomes Project Consortium, 2015).

Annotation of SNPs

To find out the potential significance of these SNPs or possible functional and physiological consequences of SNPs, structural and functional annotation was performed through SNPnexus, https://www.snp-nexus.org/v4/, the web openaccess, variant annotation tool (Oscanoa et al. 2020). For annotation, Human genome reference builds - hg38 was utilized as a reference. Further clinical and pharmacological annotation for all SNPs was done through ClinVar (Landrum et al. 2018), https://www.ncbi.nlm.nih.gov/clinvar/; and PharmGKB (Thorn et al. 2013), https://www.pharmgkb.org/, database.

Linkage disequilibrium analysis of SNP-Haplotypes (R²)

Investigating patterns of linkage disequilibrium over a wide range of demographic groupings aids in identifying the potential genetic association. R^2 (R squared) is one of the two measures of linkage disequilibrium; that was considered for LD analysis; it quantifies the concordance of alleles for two genetic variants (VanLiere and Rosenberg 2008).

 Table 1 Details of considered pharmacogenes and their pharmacological role

S. No	Gene	SNPs	Description
1	ABCB1(5)	rs7790722, rs2235040, rs2188531, rs12704363, rs10267099	Transporter
2	ABCC1(17)	rs129116, rs246221, rs903880, rs4781712, rs35597, rs246220, rs2239995, rs2014800, rs215106, rs152022, rs11864374, rs212090, rs246217, rs35587, rs3765129, rs3851713, rs3887893	Transporter
3	ABCC2(10)	rs2002042, rs12260112, rs6584327, rs3740074, rs4148385, rs717620, rs3740065, rs2756104, rs2073337, rs3740063	
4	SLCO1B1(3)	rs999278, rs1292912, rs4149026	
5	ADH1B (6)	rs1229983, rs3840297, rs17033, rs1042026, rs1789882, rs4147536	Metabolizer
6	ALDH2(3)	rs2238151, rs10849970, rs2158029	
7	CES1(3)	rs2002577, rs2307240, rs12443580	
8	CYP1A1(5)	rs4646421, i00243, rs4646903, rs1048943, rs1799814	
9	CYP1A2(3)	rs762551, rs3743484, rs2069526	
10	CYP2A6(2)	rs28399466, rs28399467	
11	CYP2B6(5)	rs8192709, rs4803419, rs4803418, rs2279343, rs1042389	
12	CYP2C19(7)	rs4986893, rs17882787, rs12247175, rs12257497, rs17879393, rs4917623, rs12768009	
13	CYP2C8(2)	rs11572080, rs10509681	
14	CYP2C9(4)	rs9332120, rs9332220, rs2153628, rs9332092	
15	CYP2D6(5)	rs3915951, rs16947, rs1081003, rs1065852, rs17002852	
16	CYP3A4(6)	rs6956344, rs3735451, rs2242480, rs2740574, rs28988574, rs4646437	
17	CYP3A5(1)	rs28365094	
18	CYP3A7(1)	rs2687077	
19	MTHFR (9)	rs1537516, rs1801133, rs4846052, rs2066470, rs1801131, rsI000156, rs2274976, rs3766747, rs3753582	
20	TPMT (4)	rs2518463, i000259, rs1800460, rs2518462	
21	UGT1A8(5)	rs3821242, rs10929301, rs6431628, rs10929303, rs6742078	
22	NAT1(4)	rs7017402, rs4921880, rs13278990, rs7003890	
23	NAT2(5)	rs1041983, rs1801280, rs4646246, rs1208, rs1799930	
24	PPARA (8)	rs7364220, rs4823613, rs4253617, rs9626814, rs9306498, rs9626736, rs5767700, rs4253765	
25	HSD11B1(4)	rs932335, rs2298930, rs2236902, rs1000283	
26	GSTM3(2)	rs7483, rs10735234	
27	GSTA1(3)	rs2290758, rs10948722, rs1051566	
28	AGTR1(10)	rs6801836, rs385338, rs12721275, rs2131127, rs2638363, rs275652, rs2933249, rs4524238, rs5186, rs4681443, rs5182	Recep- tor and
29	AHR (2)	rs2066853, i00241	signaling
30	BDKRB2(2)	rs5225, rs1046248	
31	CTLA4(5)	rs4553808, rs231777, rs11571315, rs11571317, rs3087243	
32	DRD3(6)	rs7638876, rs167770, rs2251177, rs226082, rs4646996, rs324035	
33	IL10(11)	rs1518110, rs3024495, rs3024498, rs1800871, rs10494879, rs2222202, rs11119474, rs3024496, rs1800872, rs1878672, rs3024500	
34	IL18(3)	rs5744247, rs1946519	
35	IL3(1)	rs31481	
36	NR1I2(5)	rs6438546, rs12721607, rs12721612, rs1357459, rs13059232	
37	NR1I3(5)	rs6686001, rs2501873, rs2502805, rs2502815, rs3003596	
38	OPRM1(1)	rs1799971	
39	VDR (8)	rs12717991, rs1544410, rs739837, rs10735810, rs2238136, rs2189480, rs4516035, rs7963776	
40	ANKK1 (1)	rs1800497	
41	HNF4A (6)	rs1028583, rs2071197, rs6093978, rs11086926, rs3212199, rs1800961	Regulator
42	EPHX1(6)	rs2671272, rs2292566, rs2260863, rs1051741, rs1051740, rs2234922	
43	GRIK1(5)	rs363504, rs363430, rs2178865, rs363538, rs2832419	Ion channel

This pair-wise linkage disequilibrium is calculated using the module LDmatrix of the tool LDlink (https://ldlink.nci. nih.gov). Considering the LD, the association of pharmacoclinical SNPs has been observed and screening for potential SNP-markers was performed.

Results and discussion

Change in a single nucleotide can have major impacts, with non-synonymous SNPs playing their role and affecting genes belonging to almost all kinds of physiological processes. As drug pharmacology has clinical significance, genes belonging to PK were explored for their SNP variants. As the distribution of these SNPs-variant plays a significant role. Distribution patterns of all considered SNPs were analyzed for the world population, and further analysis was done to enrich their pharmaco-clinical significance and check for the possibilities of using them as markers.

Distribution patterns of SNPs

Genomic variation studies are well practices and it is established that most genomic variations are population-specific and follow population-specific distribution patterns. SNPs are also following the same distribution rules; their distribution patterns illustrate plenty about the evolution and population-specific phenotypes (Choudhury et al. 2014; Huang et al. 2015). To detect the Indian population-specific SNPs, their presence was observed in the world super-population, and all 206 SNPs belonging to 43 pharmacogenes were considered for population-specific distribution patterns investigation. The resulting distribution patterns indicate that out of 206 SNPs, 11 SNPs did not shown their presence in any of the world's populations. Such exclusive SNPs were earlier considered population-specific SNPs and also termed "private SNPs" (Baye et al. 2009). From the rest of the 195 SNPs, 84 were found to harbor unique minor and major alleles in Indian populations and were considered "private- allele SNPs" for Indians. The rest of 111 SNPs were observed as global SNPs which were found in all super populations with similar minor and major alleles. On the basis of the obtained results, all SNPs were classified into three classes; (i) Private SNPs (SNPs only present in the Indian population), (ii) Global SNPs with Private Indian alleles, and (iii) Global SNPs with common alleles (Table s1). In total, we got 11 SNPs as Private, 84 SNPs with private Indian alleles, and the rest of 111 SNPs as global SNPs with common alleles (Table s1).



Fig. 1 Column chart shows physical annotation (consequences) of SNPs variants

Information about these distribution patterns is very important and has the potential to answer a lot of questions, especially regarding the causal role of common human DNA variation in complex human traits and for investigating the nature of genetic variation within and between human populations (Hinds et al. 2005; Srivastava and Katara 2020). It also helps in pharmaceutical studies, mainly during clinical trial phases, for drug efficacy and dosage standardization processes in a population-wise manner (Burt and Dhillon 2013; Sim and Ingelman-Sundberg 2011). Application of populations-specific SNPs is also projected for practice to population-specific disease gene mappings (Shastry 2007, 2009).

Annotations

As mentioned, SNPs provide a huge scope in numerous variation-related aspects, but before their use for this purpose, they need to be enriched and annotated. All the considered SNPs belong to genes of pharmacogenomic importance, but not all of them are relevant or affect phenotype. Annotations can provide further details which could help us to deduce more about their potential significance (Miller et al. 2019). Annotation was performed to explore features of the considered SNPs mainly with reference to their physical consequences (Fig. 1), and pharmacological and clinical significance (Table 2).

The impact of all the consequences is not common; it varied from modifier to high impact range. Modifier impacts (158 SNP variants) include variants at downstream, upstream, intron, 3_prime_UTR, and 5_prime_UTR, where predictions are difficult or there is no evidence of impact. Low impacts (18 SNP variants) are created by synonymous_variant, they are assumed to be mostly harmless or unlikely to change protein behavior. Moderate impacts (28 SNP variants) mainly include miss-sense and splice region variants; generally, these are non-disruptive variants that might change protein effectiveness. Consequences like the start_lost and stop_gained variants are assumed to have a high (disruptive) impact (02 SNP variants) on the protein, probably causing protein truncation, loss of function, or triggering nonsense-mediated decay.

Annotation with reference to pharmacological importance and clinical significance suggests that in total 53 SNPs were found to be associated with pharmaco-clinical impact (Table 2). These 53 SNPs are shared by 26 genes, where three genes, i.e., MTHFR, NAT2, and CYP3A4 share four SNPs each, and another three genes, i.e., NR113, VDR, and CYP2D6 share three SNPs each. Twelve genes, i.e., ABCB1, ABCC2, AGTR1, CTLA4, CYP1A2, CYP2B6, CYP2C19, CYP2C8, EPHX1, HNF4A, IL10, and UGT1A8, share two SNPs each, and the rest of 8 SNPs belongs to

Table 2	Reported SNPs v	with pharmaco-cli	nical significance and their phenotypic association
S. No.	Gene name	Variation ID	Affected pharmacology and major phenotypic changes#
Α	Indian Populati	on Specific SNP	Variants
1	ABCB1	rs2235040	- Toxicity
2	ABCC2	rs717620	- Metabolism/PK; Dubin-Johnson syndrome
3	ABCC2	rs3740065	- Toxicity; Metabolism/PK; Leukemia, B-Cell, Acute
4	ANKK1	rs1800497	- Toxicity; Alcoholism; Substance-Related Disorders
5	CES1	rs2307240	- Efficacy; Acute coronary syndrome
6	CYP1A1	rs1048943	- Efficacy; Breast Neoplasms
7	CYP2C8	rs11572080	- Toxicity; CYP2C8 Haplotype Polymorphism
8	CYP2D6	rs17002852	- Tramadol response
9	HNF4A	rs11086926	- Familial hyperinsulinism, Maturity-onset diabetes of the young, type 1
10	HNF4A	rs1800961	- All Highly Penetrant, Familial hyperinsulinism, Maturity-onset diabetes of the young, type 1, Monogenic diabetes
11	MTHFR	rs1801133	- Drug Toxicity; hematotoxicity; Leukopenia; mucositis; Neoplasms; Neutropenia; Osteosarcoma; Precursor Cell Lymphoblastic Leukemia-Lymphoma; primary central nervous system lymphoma; Thrombocytopenia; Toxic liver disease
12	MTHFR	rs2066470	- Homocystinuria due to methylene tetrahydrofolate reductase deficiency, Homocystinuria due to MTHFR deficiency
13	MTHFR	rs1801131	- Efficacy; Neoplasms
14	MTHFR	rs2274976	- Homocystinuria
15	NR1I3	rs2502815	- Toxicity; Anemia; Nasopharyngeal Neoplasms
16	ТРМТ	rs1800460	- Toxicity; Neoplasms
17	VDR	rs1544410	- Efficacy; Osteoporosis
18	VDR	rs4516035	- Metabolism/PK
19	CYP2D6	rs1065852	- Toxicity; Opioid-Related Disorders
20	CYP3A4	rs2740574	- Toxicity; Breast Neoplasms
21	CYP3A4	rs4646437	- Metabolism/PK; Kidney Transplantation; liver transplantation; Proteinuria
22	IL10	rs1800872	- Efficacy; Kidney Transplantation
23	NR1I3	rs2501873	- Dosage; Inflammatory bowel disease
24	VDR	rs739837	- Basal cell carcinoma
25	CYP3A4	rs3735451	- Toxicity; Vitamin D-dependent rickets type II with alopecia
B.	Global SNP Va	riants	
1	AGTR1	rs5182	- Efficacy; Coronary Artery Disease
2	ABCB1	rs10267099	- Toxicity; Breast Neoplasms; Neutropenia
3	AGTR1	rs5186	- Toxicity / Efficacy; Drug Hypersensitivity; Hypertension; Hypertrophy, Left Ventricular; Renal tubular dysgenesis,
4	CTLA4	rs4553808	- Toxicity/PK; Kidney Transplantation
5	CYP1A2	rs762551	- Toxicity/Dosage; Stevens-Johnson Syndrome; Arthritis, Rheumatoid/ Depressive Disorder, Major
6	CYP1A2	rs2069526	- Toxicity; Depressive Disorder, Major
7	CYP2B6	rs8192709	- Toxicity; Carcinoma, Non-Small-Cell Lung, Cystitis; Transplantation
8	CYP2B6	rs4803419	- Metabolism/PK; HIV Infections, efavirenz response - Metabolism/PK
9	CYP2C19	rs4986893	- Toxicity; Cough, Maculopapular Exanthema; Tuberculosis
10	CYP2C19	rs12768009	- Metabolism/PK; HIV Infections
11	CYP2C8	rs10509681	- Toxicity; CYP2C8 HAPLOTYPE POLYMORPHISM, rosiglitazone response - Metabolism/PK
12	EPHX1	rs1051740	- Metabolism/PK; Kidney Transplantation, Epilepsy; Ovarian Neoplasms - carbamazepine response, Cystic fibrosis
13	IL18	rs5744247	- Efficacy; Breast Neoplasms;Colorectal Neoplasms
14	NAT2	rs1041983	- Toxic liver disease; Tuberculosis; Drug interaction with drug; Drug Toxicity; Tuberculosis
15	NAT2	rs1801280	- Other/Toxicity; Tobacco Use Disorder; Ovarian Neoplasms, ACETYLATION
16	NAT2	rs1208	- Toxicity; Drug interaction with drug; Drug Toxicity; Acetylation, Slow
17	NAT2	rs1799930	- Toxicity; Drug interaction with drug; Drug Toxicity; ethambutol, isoniazid, pyrazinamide, and rifampin response
18	OPRM1	rs1799971	- Efficacy/Toxicity; Tobacco Use Disorder; Erectile Dysfunction; Sexual Dysfunctions,
19	PPARA	rs4823613	- Efficacy; Organ Transplantation
20	UGT1A8	rs10929303	- Toxicity; Liver Failure, Acute; HIV Infections; nephrolithiasis; Lucey-Driscoll syndrome, Gilbert syndrome, Crigler-Najjar syndrome
21	UGT1A8	rs6742078	- Biliruhin Serum Level of Quantitative Trait Locus 1 Gilbert syndrome

lable 2 (continued)						
S. No.	Gene name	Variation ID	Affected pharmacology and major phenotypic changes#			
22	CYP2D6	rs16947	- Toxicity; Bradycardia; Debrisoquine, ultrarapid metabolism, Tramadol response, Deutetrabena- zine response			
23	CYP3A4	rs2242480	- Efficacy/Toxicity; Coronary Artery Disease; Heroin Dependence			
24	DRD3	rs167770	- Efficacy; Anxiety Disorders			
25	EPHX1	rs2234922	- Toxicity/Dosage; Congenital Abnormalities; Craniofacial Abnormalities/ Carcinoma, Non-Small- Cell Lung/ Epilepsy; carbamazepine response			
26	IL10	rs1800871	- Dosage; liver/Kidney transplantation			
27	NR1I3	rs3003596	- Metabolism/PK; HIV Infections			
28	CTLA4	rs3087243	- Toxicity; Psoriasis Autoimmune lymphoproliferative syndrome type V, chronic fatigue syndrome with infection-triggered onset			

#Source-ClinVar database and PharmGKB database

OPRM1, PPARA, TPMT, IL18, DRD3, CYP1A1, ANKK1, and CES1 genes. Though very brief information is available, interestingly, it has been observed that most of these SNPs are reported to be associated with multiple phenotypic outcomes. rs1799971, which belongs to OPRM1, was found to be involved in a huge number of events and shows 56 annotation hits in the PharmGKB (Thorn et al. 2013) database. Though we were concerned to enrich the private Indian SNPs we did not find hits for all 11 SNPs during pharmaco-clinical annotation. As they are only reported in Indian populations, there are possibilities that very less is known and studied about their significance and phenotypic associations.

As all of these SNPs belong to the Absorption/Distribution /Metabolism/Excretion or Toxicity (ADMET) process, the pharmacokinetics of drugs get affected and result in increased severity of related diseases (e.g., Leukemia in the presence of rs3740065). At the same time, a few abnormalities also emerged in terms of disease and syndrome (e.g., Dubin-Johnson syndrome caused by the presence of rs717620). Though these SNPs belong to genes involved in drugs pharmacokinetics, in other ways, they also affect the pharmacodynamics of drugs, mainly the efficacy of the drugs (e.g., rs1048943), and alter the dosage of the drug as well (e.g., rs2501873), which finally results in variable drug response (VDR).

Linkage disequilibrium between SNP-Haplotypes

Haplotypes are ensembles of genetic polymorphisms that co-occur throughout a single chromosome and are critical to fully describing and inferring an individual's genome. Linkage disequilibrium is a sensitive indicator of the population genetic forces that structure a genome. Pair-wise Linkage disequilibrium, in terms of r², was mined between all considered haplotype-SNPs to find SNP-pairs and observe the strength of associations between them (García-Fernández et al. 2018). Linkage disequilibrium was done in two sets of the population (i) among all five super-populations, and (ii) all South Asian sub-populations independently. In both super-populations and South Asian sub-populations, all those pairs were considered significant if they showed $r^2 \ge 0.8$ in at least one population.

LD in super-populations

LD results for five super-populations indicate that in total 98 SNP-pairs are significantly coupled; the maximum number of SNP pairs with $r^2 \ge 0.8$ belongs to chromosome 10 (32-pairs) followed by chromosome number 1 (27-pairs). A further observation indicates that out of 94 pairs, 18 pairs are associated in all five populations, and 20 pairs show association in only one of the five populations. SNP-pairs indicate that each of rs2756104, rs4148385, and rs6584327 was found significantly associated with 7 other SNPs, and rs12260112, rs2073337, rs3740063, and rs3740074 were associated with 6 other SNPs. Other frequent SNPs are rs1537516, rs222202, rs3024496, rs3024500, rs3753582, and rs3766747 each of which is associated with 4 other SNPs.

Observations for inter-population variability in LD across super-population indicate that AMR and EUR populations almost show a similar association among SNP-pairs which are 71 and 70 in number, respectively. The most frequent number of pairs, i.e., 85, was observed in EAS, which includes 17 unique pairs that were not observed in other populations. SNP-pair in SAS (56) is almost comparable to AMR, EUR, and EAS populations but does not carry any single unique pair. On the other hand, the AFR population only displays 26 SNP-pairs with significant values (Fig. 2), and most of them are common with four other populations.

LD in SAS sub-populations

Mining for SAS sub-populations resulted in 87 SNP pairs in total, out of them 33 SNP-pairs belong to chromosome 1, and 21 belong to chromosome 10, rest of SNP-pairs are shared by chromosome- 5|2, 3|3, 2|4, 2|6, 8|7, 2|8, 3|12, 1|15,



Fig. 2 Range wise comparative representation of LD in SNP-pairs among world super-population (AFR: African, AMR: Ad Mixed American, EAS: East Asian, EUR: European, SAS: South Asian)



Fig. 3 Range wise comparative representation of LD sharing among SAS subpopulations (BEB: Bengali from Bangladesh, GIH: Gujrati Indian from Houston, Texas, ITU: Indian Telugu from the UK, STU: Sri Lankan Tamil from the UK, PJL: Punjabi from Lahore, Pakistan)

4|16, 1|19, 1|20 and 1|22. Out of these 87, 24 SNP-pairs were found common in all five populations, and in fact, most of them show $r^2 \sim 1$ (Fig. 3), which indicates that they are strongly associated with each other in all South Asian populations. Interestingly it has been found that out of all five populations' only ITU shares unique (12) SNP pairs. It has been observed that rs3024500 is associated with 7 other SNPs and each of rs10494879, rs1878672, rs3024496 associated with 6 other SNPs, and rs12260112, rs2066470, rs2073337, rs2222202, rs2756104, rs3740074, rs3753582, rs3766747, rs4148385, and rs6584327 are associated with 5 SNPs. Other frequent SNPs are rs1518110, rs1537516, rs1800871, rs1800872, rs2242480, and rs3735451 each of which is associated with 4 other SNPs.

Results for inter-population LD variations, among the population, indicate that four populations, i.e., STU (69), PJL (70), GIH (68), and BEB (72) share similar kind of association patterns among SNP-pairs. On the other hand, ITU also shows 62 SNP-pairs, but out of them, 12 are found unique and do not share by any other South Asian population.

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Fig. 4 Synopsis of overall analysis of Indian populated SNPs belongs to pharmacokinetics genes

SNP biomarker

SNP-pairs with high LD are very interesting and important for molecular biology purposes; they can be utilized as a potential biomarker for genome-wide association studies (GWAS) and related phenotypic features (Alwi 2005; Uffelmann et al. 2021). On the basis of population-specific LD, SNP-pair can be either considered as a universal marker or a population-specific (private) marker. As annotation indicated that 53 SNPs are associated with pharmaco-clinical phenotypes, out of these 53, 21 SNPs were significantly paired with other SNPs and formed 24 significant pairs in total (Fig. 4). Interestingly, out of 21 clinically significant SNPs, 7 SNPs, i.e., rs1800871 (4), rs1800872 (4), rs2066470 (2), rs2242480 (2), rs2501873 (2), rs3003596 (2), and rs4646437 (2) are involved in multiple associations (Table 3).

High r² values indicate that both SNPs of SNP-pairs convey similar information, as one allele of the first SNP of the pair is often observed with one allele of the second SNP of the pair, so only one of the two SNPs needs to be genotyped to capture the presence of the allelic polymorphism (Bush and Moore 2012). It is noticeable that all 24 SNP-pairs in Table 3 show presence in both, the world and SAS populations, but out of these, 13 have Indian population-specific variants (minor-major alleles) which are different from the rest of the population. As noted, before, patterns of LD are population-specific, thus tag SNPs selected for one population may not work well for different populations. To avoid false-positive tagging or redundant information, the use of these 13 SNP-pairs for global genotyping needs to take care of allelic variations during LD analysis. The rest of the 11 SNP-pairs, as their minor and major alleles are common in global populations, their use of genotyping will be straightforward with similar alleles in all populations in comparison and thus act as universal SNP-tags (Li et al. 2008).

across populations							
S. No.	rsID of SNP-Pairs		World	SAS			
	Reference*	Associated SNPs	ALL	ALL			
1	rs2066470	rs3766747	0.934	1.0			
2	rs2502815	rs2502805	0.788	0.801			
3	rs1800871	rs1800872	0.998	1.0			
4	rs2501873	rs3003596	0.828	0.992			
5	rs2274976	rs1537516	0.638	0.716			
6	rs4553808	rs231777	0.801	0.846			
7	rs167770	rs226082	0.948	0.991			
8	rs4646437	rs2242480	0.724	0.915			
9	rs1208	rs1801280	0.823	0.914			
10	rs11572080	rs10509681	1.0	1.0			
11	rs739837	rs7963776	0.807	0.927			
12	rs4803419	rs4803418	0.936	1.0			
13	rs4823613	rs5767700	0.589	0.773			
14	rs2242480	rs2687077	0.482	0.723			
15	rs2066470	rs3753582	0.932	0.97			
16	rs1800872	rs1518110	0.962	0.94			
17	rs1800871	rs1518110	0.961	0.94			
18	rs1800872	rs1800871	0.998	1.0			
19	rs3003596	rs2501873	0.828	0.992			
20	rs1800871	rs1518110	0.961	0.94			
21	rs1800872	rs1518110	0.962	0.94			
22	rs2242480	rs3735451	0.768	0.914			
23	rs3087243	rs11571315	0.467	0.709			
24	rs4646437	rs3735451	0.676	0.833			

 Table 3 Association (LD) between SNP-pairs and their distribution across populations

* Found to be associated with pharmaco-clinical features

Though in Table 3, Only, SNP-Pairs with pharmaco-clinical significance were mentioned, various other SNP-pairs, whose phenotypic information is not available, were also mined. The use of all of these SNPs-pair for population-specific GWAS mainly with reference to the pharmacokinetics studies is expected. As they all belong to genes involved in the ADMET process, it is anticipated that as soon as their annotation is available, they will be further explored for their pharmaco-clinical connection.

Conclusion

It is imperative to have knowledge about population-specific SNPs with relevance to pharmacogenomics. In this study, Mining, distribution pattern, and annotation were performed for 206 SNPs of pharmacokinetics importance, reported in the Indian population. Distribution patterns suggest a wide range of distribution of SNPs among the world's super-population and South Asian sub-population, eleven private SNPs for the Indian population were also observed. Annotation suggested that out of 206 considered SNPs, 53 are reported to be associated with pharmaco-clinical phenotypes and affect the pharmacokinetics of the drugs. Linkage disequilibrium through r^2 suggests that the number of SNPs forms considerable SNP-pairs, whose number varies from population to population. Further, it has been observed that 24 of these SNP-pairs have the potential to be used as markers (Tag-SNPs) for pharmaco-clinical phenotypes. It is assumed that the information produced through this study, after experimental validation, may be useful for further GWAS for pharmacogenomics-based pharmacoclinical phenotype purposes among populations in the near future.

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Author contributions AY: Contributed to the implementation of the research, to the analysis of the results, and to the writing of the manuscript SS: Contribute to population-wise data analysis and interpretation ST: Performed SNP annotation for their clinical significance NK: Prepared figures PK: Design and supervised the work, and writing & communicating the manuscript Note: All authors reviewed the manuscript.

Declarations

Competing interests The authors declare no competing interests.

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