

Estrogen Receptor Alpha Phosphorylated at Tyrosine 537 is Associated with Poor Clinical Outcome in Breast Cancer Patients Treated with Tamoxifen

Georgios P. Skliris · Zoann Nugent · Peter H. Watson · Leigh C. Murphy

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Abstract A phospho-tyrosine 537-ER α (p-Y537-ER α) antibody was validated for immunohistochemistry of formalin-fixed paraffin-embedded human breast cancer biopsies and used to interrogate multiple ER positive breast cancer cases present on tissue microarrays already constructed by the Manitoba Breast Tumor Bank. Nuclear p-Y537-ER α protein expression was positively associated with positive nodes (Spearman $r=0.20$, $P=0.0002$) and large tumor size ($r=0.13$, $P=0.02$). On univariate analysis, high levels of p-Y537 were associated with poor overall survival (HR=1.65, 95% CI 1.08–2.52, $P=0.02$) but not relapse free survival from breast cancer. The association with overall survival was not significant on multivariate analysis. These data support the relevance of phosphorylation at p-Y537-ER α in human breast cancers and add further support to the presence of a phosphorylation code for ER α in human breast cancer in vivo.

Keywords Phosphorylation · Tyrosine 537 · Estrogen receptor · Breast cancer · Tissue microarrays · Immunohistochemistry · Endocrine therapy

G. P. Skliris (✉) · L. C. Murphy
Department of Biochemistry and Medical Genetics,
Faculty of Medicine, University of Manitoba,
675 McDermot Avenue,
Winnipeg R3E 0V9 MB, Canada
e-mail: lcmurph@cc.umanitoba.ca

G. P. Skliris · Z. Nugent · P. H. Watson · L. C. Murphy
Manitoba Institute of Cell Biology, University of Manitoba,
675 McDermot Avenue,
Winnipeg, MB, Canada

P. H. Watson
Tumour Tissue Repository and Deeley Research Center,
BC Cancer Agency,
Victoria, BC, Canada

Introduction

Although phosphorylation of tyrosine 537 in the human estrogen receptor has been of interest and explored experimentally for several years, its detection in breast cancer cell lines has been inconsistent [1–3]. The detection of a mutation at this site in ER α in a biopsy sample from a human breast cancer metastasis [4], however, supports its potential relevance in human breast cancer in vivo. However, detection of phosphorylated Y537-ER α in human breast tumor biopsies has not been reported. We have optimized and validated an antibody specific for phosphorylation at Y537 on ER α for immunohistochemistry (IHC) of formalin-fixed paraffin-embedded (FFPE) breast tumor tissues and investigated p-Y537-ER α protein expression in multiple human breast cancer biopsies in order to establish relevance in human breast cancer in vivo.

Materials and Methods

Tissue Microarrays

The tissue microarrays (TMAs) used in this study were constructed prior to this study by the Manitoba Breast Tumor Bank (MBTB, CancerCare Manitoba and University of Manitoba) from over 5,000 cases that have been accrued over the period 1988 to present [5, 6]. Detailed descriptions of the MBTB, case selection, and TMA construction have been published previously [7]. There were over 450 cases represented on the original TMAs, but due to exhaustion of tumor cores from previous use of the TMAs or incomplete data for some cases, the tumor numbers (n) analyzed for some markers were less than

450. This is shown in the results section. MBTB is part of the Canadian Tumor Repository Network (<http://www.CTRNet.ca>) and embraces their policies and operating protocols. The MBTB operates with approval from the Research Ethics Board of the Faculty of Medicine, University of Manitoba.

Antibodies

The phospho-Y537-ER α antibody was a rabbit polyclonal affinity-purified antibody (1 mg/ml; Abcam Inc Cambridge MA) and was used at a 1/150 final dilution. Other phospho-ER α specific antibodies used for IHC were validated previously [8]. HER-2 (mouse monoclonal, clone CB11) and ER α (6F11, mouse monoclonal, Abcam, Cambridge, MA) were used as previously described [9].

Immunohistochemistry, Quantification, and Cut-Off Selection

Immunohistochemistry was performed as described previously [8], where 5 μ m sections were submitted to heat-induced antigen retrieval in the presence of a tris/borate/EDTA buffer (CC1, Ventana Medical Systems, AZ, USA) using an automated tissue immunostainer (Discovery Staining Module, Ventana Medical Systems, AZ, USA). Specificity of the phospho-specific antibodies generally was determined in parallel using antibodies that had been immunoabsorbed (immunoneutralized) with \sim 30 \times excess phospho-specific peptide or non-phosphorylated peptide. Nuclear staining was assessed and *H* scores (0–300) were generated as previously reported [7, 8]. TMAs were evaluated by two investigators (GPS, PHW) and when discordance was found, cases were re-evaluated to reach consensus.

Statistical Methodology

Relapse-free survival was defined as the time from initial surgery to the date of clinically documented local or distant disease recurrence or death attributed to breast cancer (censors were other death and end of follow-up). Overall survival (OS) was defined as the time from initial surgery to the date of death attributable to breast cancer (censors were other deaths and end of follow-up). Survival was analyzed by Cox regression to examine hazard ratios (HR). Each model was tested and complied with the assumption of proportional hazard. Statistical analyses were performed using SASTM version 9.1. Analyses were performed with single predictors and in a multivariate model.

Results

Validation and Optimization of a Phospho-Tyrosine 537 ER Alpha Antibody for Immunohistochemistry

The antibody was screened for its ability to detect nuclear staining in ER α positive but not ER α negative breast tumors (Fig. 1a versus d–f). A breast cancer case showing strong nuclear staining was then chosen for further analysis. One section was stained with the p-Y537-ER α antibody (Fig. 1a) and loss of nuclear staining was observed in a serial section stained with antibody that had been immunoabsorbed with \sim 30 \times excess of the phospho-peptide used to generate the antibody (Fig. 1b), but not with excess non-phospho-peptide (Fig. 1c). It should be noted that while the excess phospho-peptide immunoneutralization lead to loss of nuclear staining (compare Fig. 1a to b), there appeared to be no loss of cytoplasmic staining (Fig. 1b), and indeed, an increase in cytoplasmic staining was sometimes seen (not shown).

Relationship of Phospho-Tyrosine 537 ER α to Known Prognostic Markers in Human Breast Tumor Samples

The validated p-Y537-ER α antibody was then used to stain TMA sections of FFPE ER α positive breast cancer cases. These TMAs have been previously used [7], and the clinical characteristics of these patients have been previously reported [7].

Statistically significant positive correlations were found with size (Spearman $r=0.13$, $P=0.02$, $n=348$) and nodes involved (Spearman $r=0.20$, $P=0.0002$, $n=341$). Furthermore, the median IHC score for p-Y537-ER α in tumors ≤ 2.5 cm was 85 and the median for tumors >2.5 cm was 180, and this difference was statistically significant (Mann–Whitney two-tailed, $P=0.02$). The median IHC score for p-Y537-ER α in node negative patients was 70, and the median for node positive patients was 180. The difference in level of expression of p-Y537-ER α between the node negative and positive groups was statistically significant (Mann–Whitney two-tailed, $P=0.0002$). Large size and positive nodal status are markers of poor prognosis; therefore, these data suggest that detection of high levels of p-Y537-ER α in primary breast tumors is associated with poor prognosis in this cohort of breast cancer patients.

Relationship of Phospho-Tyrosine 537 ER α to Other Phosphorylation Sites on ER Alpha in Human Breast Tumors

Previously, using the same TMAs, we had determined the expression of other known and novel phosphorylated sites

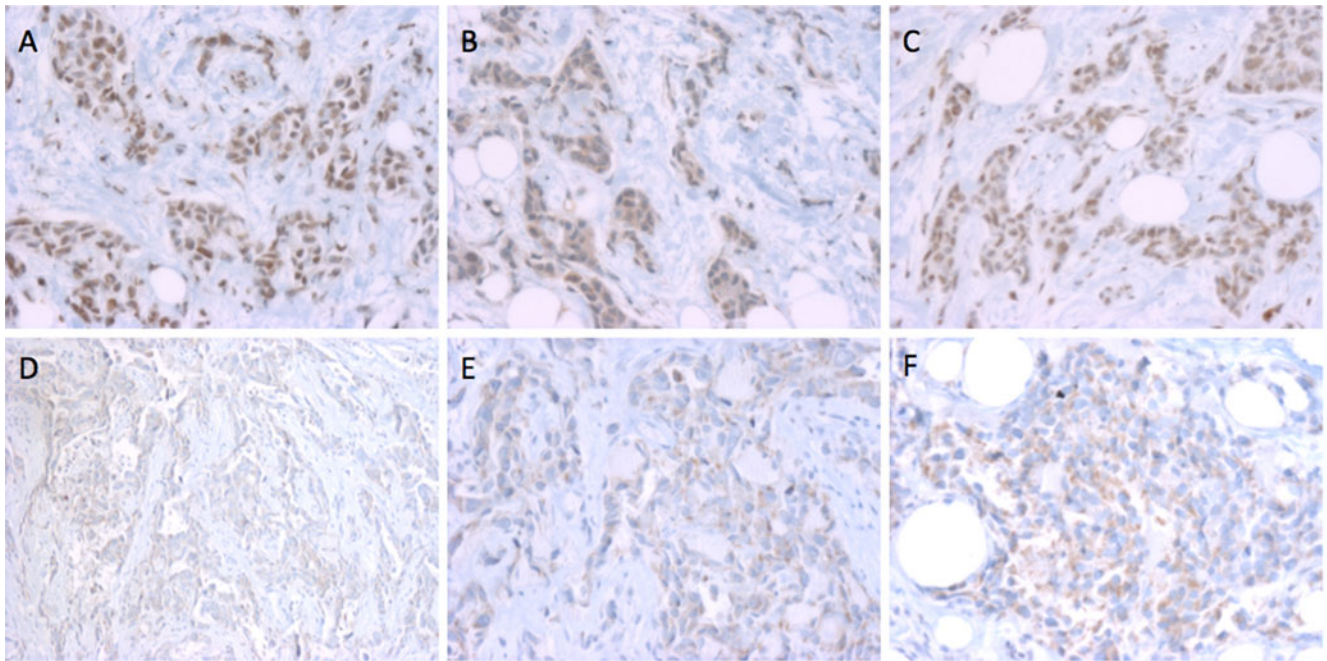


Fig. 1 Immunohistochemical validation of p-Y537-ER α phosphoantibodies in human breast cancer biopsy samples. **a** An ER+ (95 fmol/mg protein by ligand binding assay, LBA) breast tumor section stained with the p-Y537-ER α antibody showing strong, nuclear staining. **b** A serial section of the same tumor using p-Y537-ER α antibody pre-absorbed

with excess of the phosphorylated peptide. **c** A serial section of the same tumor using p-Y537-ER α antibody pre-absorbed with excess of the non-phosphorylated ER α peptide. **d–f** Three different ER α negative (0 fmol/mg protein by LBA) breast tumors stained with the p-Y537-ER α antibody. All magnifications, $\times 500$

in ER α [7, 8]. By comparing p-Y537-ER α expression with other phosphorylated sites in this cohort, p-Y537-ER α was observed to be positively correlated with p-T311-ER α (Spearman $r=0.41$, $P<0.0001$, $n=343$) and p-S559-ER α (Spearman $r=0.24$, $P<0.0001$, $n=340$). When the cases were dichotomized into high or low expression of p-T311-ER α using the median IHC score as a cut-off, the expression of p-Y537-ER α was significantly higher in the high p-T311-ER α versus the low p-T311-ER α (180 versus 40 median IHC score, Mann–Whitney two-tailed, $P<0.0001$). Similarly, the expression of p-Y537 was significantly higher in the high p-S559-ER α versus the low p-S559-ER α expressing tumors (90 versus 25 median IHC score, Mann–Whitney two-tailed, $P<0.0001$). These phosphorylation sites have been found to be previously associated with a poor prognosis in women treated with tamoxifen [7].

Relationship of Phospho-Tyrosine 537 ER α Detection in ER Positive Breast Cancers Patients Treated with Tamoxifen

The relationship of p-Y537-ER α to known and novel markers of poor prognosis in breast cancer suggested that p-Y537-ER α may also be associated with poor prognosis and potentially poor outcome to tamoxifen therapy. Since

patients represented on these TMAs had surgery, plus or minus radiation, followed by adjuvant tamoxifen therapy with a median clinical follow-up was 99 months (range 9–217 months), the relationship of high versus low expression of p-Y537-ER α to clinical outcome was determined.

Univariate analysis for overall survival from breast cancer death (OS) is shown in Table 1. Detection of high levels of p-Y537-ER α was associated with a poorer OS from breast cancer in women treated with tamoxifen (Fig. 2a). The association of other parameters such as nodal status, size, PR levels, etc., with clinical outcome in this cohort has been previously reported [7]. Size was the only parameter that remained significantly associated with OS from breast cancer death after multivariate analysis. High expression of p-Y537-ER α was not found to be associated with relapse free survival (Table 2, Fig. 2b). These data suggest that high expression of p-Y537-ER α is not an independent factor associated with clinical outcome.

Relationship of Phospho-Tyrosine 537 ER α Expression and HER2 Expression

The expression of HER2 was determined by scoring membrane staining intensity (0, 1, 2, 3) as previously described [9]. There appeared to be an inverse relationship

Table 1 Overall survival from death due to breast cancer

Predictor	<i>N</i>	HR	95% C.I.	<i>P</i>
Age>50 years	420	1.67	0.74–3.81	0.22
Size>2.5 cm	422	2.27	1.56–3.30	<0.0001*
Node +ve	415	2.06	1.42–3.00	0.0002*
Grade	420	1.38	1.02–1.87	0.035*
PR LBA>20	421	0.61	0.42–0.87	0.007*
p-Y537>median	348	1.65	1.08–2.52	0.02*
Multivariate analysis				
Age>50 year	337	1.37	0.42–4.43	0.60
Size>2.5 cm	337	1.90	1.23–2.94	0.0038*
Node +ve	337	1.58	1.02–2.45	0.04*
Grade	337	1.40	0.99–1.97	0.056
PR LBA>20	337	0.67	0.44–1.03	0.065
p-Y537>median	337	1.49	0.96–2.30	0.073

Table 2 Relapse-free survival from recurrence or death due to breast cancer

Predictor	<i>N</i>	HR	95% C.I.	<i>P</i>
Age>50 year	420	1.57	0.80–3.07	0.19
Size>2.5 cm	422	1.85	1.36–2.51	<0.0001*
Node +ve	415	2.02	1.48–2.76	<0.0001*
Grade	420	1.45	1.12–1.88	0.0055*
PR LBA>20	421	0.60	0.44–0.81	0.001*
p-Y537>median	348	1.35	0.96–1.90	0.087
Multivariate analysis				
Age>50 year	337	0.93	0.40–2.15	0.87
Size>2.5 cm	337	1.64	1.16–2.33	0.0055*
Node +ve	337	1.68	1.17–2.39	0.0044*
Grade	337	1.43	1.07–1.91	0.015*
PR LBA>20	337	0.60	0.42–0.86	0.0047*
p-Y537>median	337	1.22	0.86–1.73	0.28

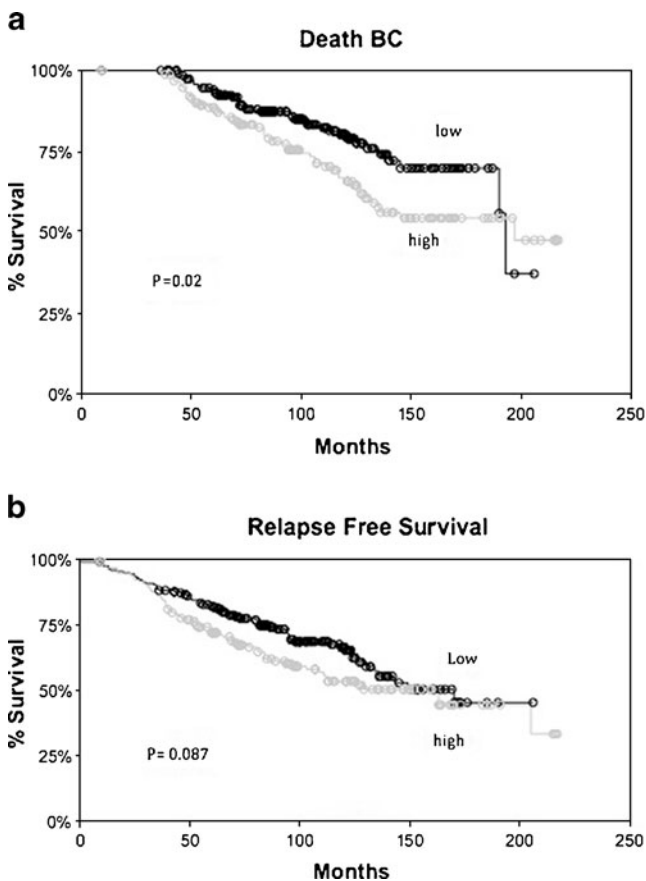


Fig. 2 **a** Kaplan–Meier graphs of overall survival from breast cancer specific death with respect to expression p-Y537-ER α (high>median *H* score). **b** Kaplan–Meier graphs of relapse free survival from breast cancer recurrence or breast cancer specific death with respect to expression of p-Y537-ER α (high>median *H* score). *P* value represents the significance of a simple survival analysis without the proportional hazard assumption that was applied in the analyses presented in Tables 1 and 2

between p-Y537-ER α and HER2 staining (Spearman $r=-0.065$, $P=0.024$, $n=187$). Furthermore, the median *H* score for p-Y537-ER α in tumors where HER2 was not detected was significantly higher than those tumors which were $\geq 2+$ for HER2 staining (90 versus 45, $P=0.027$). In addition, in a small group of tumors ($n=9$) that were matched for ER α , PR, and grade, no significant difference was observed in p-Y537-ER α expression in the HER2 negative versus the HER2 over-expressing group (90 versus 120, $P=0.93$ Mann–Whitney). These data suggest that phosphorylation of Y537 is unlikely to be due to HER2 over-expression in ER positive breast cancer.

Discussion

Post-translational modifications (PTMs) such as phosphorylation are known to be important in regulation of protein function [10]. ER α can be phosphorylated at multiple residues including serines, threonines, and tyrosines. We and others have previously been able to establish the potential relevance in vivo of phosphorylation at several serine and threonine residues by IHC mainly in breast cancer biopsy samples [7, 11–15]. While phosphorylation at tyrosines on ER α has been of interest due to the previously determined interactions of ER α , c-Src, and growth factor receptor kinases, tyrosine phosphorylation on ER α has only been performed to date using cultured cells [1, 3, 16]. However, the finding from Suzanne Fuquas laboratory that at least one breast cancer metastasis was found to express a missense mutant ER α such that an asparagine residue replaced tyrosine 537 [4] suggests that this residue and its phosphorylation may have a role in metastasis in breast

cancer *in vivo*. Furthermore, this mutant displayed potent estrogen independent transcriptional activity that was minimally affected by either tamoxifen or ICI164384 [4], supporting a role of tyrosine phosphorylation in breast cancer progression and possibly antiestrogen resistance. Previous evidence suggests that phosphorylation at Y-537 increases the efficiency of estrogen binding [17], and site-directed mutagenesis experiments to eliminate phosphorylation at this site resulted in increased ligand-independent transcriptional activity that was still however inhibited by antiestrogen [16]. Other experiments using an excess of a 12 amino acid peptide containing p-Y537 of human ER α demonstrated that this disrupted the dimerization of ER α such that the receptor no longer bound to DNA containing an ERE using an electromobility shift assay [18]. Furthermore, Y537 is at the base of helix 12 in ER α . Upon ligand binding, the positioning of helix 12 is critical for coactivator recruitment [19]. Interestingly, mutation of Y537 to a serine was found to impart stability to recombinant ligand binding domain fragments of ER α to facilitate crystallization in the absence of ligand [20]. It appears that the structure of the amino acid at residue 537 as well as its phosphorylation can have significant functional consequences for ER α . The limited data currently available in the literature suggest that in particular, phosphorylation at Y537 can affect stability of the receptor dimer and coactivator recruitment. Therefore, it is tempting to speculate that persistent phosphorylation at Y537, which could be concluded from our ability to detect p-Y537 (often an unstable event) in breast tumor biopsy samples, could significantly alter the cyclical and dynamic interaction of the ligand-bound receptor with target gene promoters, which is known to be critical for regulation of gene transcription [21, 22]. This could lead in turn to deregulated estrogen signaling and increased risk of disease progression. These data together with the current detection of p-Y537 in human breast cancer biopsy samples, now strongly support the relevance of phosphorylation of Y-537 in breast cancer *in vivo*.

The association of pY537 with large tumor size and positive node status suggested that it might be a poor prognostic marker. This was supported by a significant association of detection of high levels of nuclear p-Y537 with a poorer overall survival. However, this relationship did not remain significant on multivariate analysis suggesting that p-Y537's association with tumor size and/or nodal status was driving the significance associated with univariate analysis. Recently, we reported that some phosphorylated sites on ER α were associated with good outcome, but others were associated with poor outcome in the same patient cohort. P-Y537 was found positively correlated with two other sites, p-T311 and p-S559 also associated with poor outcome [7]; however, if we added the presence of high p-Y537 to the P7-score that we had previously developed [7], to generate a P8-

score, no further value for predicting prognosis was found. Interestingly, as was noted previously, the phospho-sites associated with poor outcome seemed to cluster in the C-terminal region while those associated with good outcome were located in the N-terminal region of ER α [7]. P-Y537 reinforces this pattern, although the functional significance remains to be determined. It is speculated that differential phosphorylation in the N- versus the C-terminal regions may influence how these two regions interact with each other to recruit co-factor complexes [23, 24].

The kinases involved in phosphorylation at Y537 *in vivo* are unknown. Experimental data from cells in culture have suggested that HER2 and/or Src-like kinases are possible candidates. However, the detection of activated c-Src in the nucleus of ER+ breast cancer patients treated with tamoxifen is generally associated with improved patient outcome [25]. In contrast, increased expression of HER2 and/or EGFR in breast tumors is associated with poor patient outcome and when co-expressed with ER α is also associated with tamoxifen resistance [26, 27]. In the current study, no association was found between p-Y537-ER α and increased HER2 expression, and EGFR was not measured in these tumors. Interestingly, p-Y537-ER α is required for the receptor to interact with the SH2 domain of c-Src and to regulate c-Src/EGFR-like signaling [28, 29], but such interactions appear to be associated most often with the extra-nuclear, so-called non-genomic actions of ER α , and in the present study, only nuclear localization of p-Y537-ER α was measured. Recently, the nuclear localization of receptor tyrosine kinases such as EGFR, HER2, and IGF1R has been reported [30, 31], and functional roles of nuclear localized receptor tyrosine kinases in transcription, DNA synthesis and cellular proliferation, DNA repair, chemo- and radio-resistance have been suggested [32]. Importantly, high levels of nuclear localized EGFR have been correlated with poor patient outcome in breast cancer [32] as well as in other cancer sites [31]. Since nuclear localized HER2 or EGFR was not assessed in this current study, it is possible that the nuclear localized HER2 and/or EGFR may have a role in phosphorylation of ER α at p-Y537. This exciting possibility requires further investigation.

Somatic ER mutations in primary breast cancer are relatively rare [33]. Due to the paucity of banked tissue from distant breast cancer metastases, studies in this regard are also rare. However, of interest is the observation that ER mutations were found in two out of eight breast cancer metastases analyzed by Fuqua's group [4]. Fuqua's group have also identified a more frequently occurring mutation in early breast cancers and interestingly this mutation alters a residue, K303, which can undergo post-translational modification [34]. In this case, the mutation (K303R) results in the inability of the site to be acetylated and also regulates the phosphorylation of a down-stream site S305

[35]. Such data underscore the importance of PTMs of ER α and suggest the possibility that altered regulation of PTMs may have important roles in breast cancer progression.

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Conflict of Interest The authors declare that they have no conflict of interest.

References

1. Arnold S, Obourn J, Jaffe H, Notides A (1995) Phosphorylation of the human estrogen receptor on tyrosine 537 in vivo and by Src family tyrosines in vitro. *Mol Endocrinol* 9:24–33
2. Marquez DC, Lee J, Lin T, Pietras RJ (2001) Epidermal growth factor receptor and tyrosine phosphorylation of estrogen receptor. *Endocr* 16(2):73–81
3. Pietras R, Arboleda J, Reese D, Wongvipat N, Pegram M, Ramos L, Gorman C, Parker M, Sliwkowski M, Slamon D (1995) HER-2 tyrosine kinase pathway targets estrogen receptor and promotes hormone independent growth in human breast cancer cells. *Oncogene* 10(12):2435–2446
4. Zhang QX, Borg A, Wolf DM, Oesterreich S, Fuqua SA (1997) An estrogen receptor mutant with strong hormone-independent activity from a metastatic breast cancer. *Cancer Res* 57(7):1244–1249
5. Watson P, Snell L, Parisien M (1996) The NCIC-Manitoba Breast Tumor Bank: a resource for applied cancer research. *CMAJ* 155:281–283
6. Snell L, Watson P (2006) Breast tissue banking: collection, handling, storage, and release of tissue for breast cancer research. *Meth Mol Med* 120:3–24
7. Skliris GP, Nugent ZJ, Rowan BG, Penner CR, Watson PH, Murphy LC (2010) A phosphorylation code for oestrogen receptor-alpha predicts clinical outcome to endocrine therapy in breast cancer. *Endocr Relat Cancer* 17(3):589–597
8. Skliris GP, Rowan BG, Al-Dhaheer M, Williams C, Troup S, Begic S, Parisien M, Watson PH, Murphy LC (2009) Immunohistochemical validation of multiple phospho-specific epitopes for estrogen receptor alpha (ERalpha) in tissue microarrays of ERalpha positive human breast carcinomas. *Breast Cancer Res Treat* 118(3):443–453
9. Weitsman G, Li L, Skliris G, Davie J, Ung K, Curtis-Snell L, Tomes L, Watson P, Murphy L (2006) Estrogen receptor-alpha phosphorylated at Serine 118 is present at the promoters of estrogen-regulated genes and is not altered due to Her2 over-expression. *Cancer Res* 66:10162–10170
10. Weigel N, Moore N (2007) Steroid receptor phosphorylation: a key modulator of multiple receptor functions. *Mol Endocrinol* 21:2311–2319
11. Murphy LC, Cherlet T, Adeyinka A, Niu Y, Snell L, Watson P (2004) Phospho-serine-118 estrogen receptor-alpha detection in human breast tumors in vivo. *Clin Cancer Res* 10:1354–1359
12. Gee J, Robertson J, Gutteridge E, Ellis I, Pinder S, Rubini M, Nicholson R (2005) Epidermal growth factor receptor/HER2/insulin-like growth factor receptor signalling and oestrogen receptor activity in clinical breast cancer. *Endocr Relat Cancer* 12(Suppl 1):S99–S111
13. Yamashita H, Nishio M, Toyama T, Sugiura H, Kondo N, Kobayashi S, Fuji Y, Iwase H (2008) Low phosphorylation of estrogen receptor alpha (ER{alpha}) serine 118 and high phosphorylation of ER{alpha} serine 167 improve survival in ER-positive breast cancer. *Endocr Relat Cancer* 15:755–763
14. Holm C, Kok M, Michalides R, Fles R, Koomstra RH, Wesseling J, Hauptmann M, Neeffjes J, Peterse JL, Stal O et al (2009) Phosphorylation of the oestrogen receptor alpha at serine 305 and prediction of tamoxifen resistance in breast cancer. *J Pathol* 217(3):372–379
15. Jiang J, Sarwar N, Peston D, Kulinskaya E, Shousha S, Coombes R, Ali S (2007) Phosphorylation of estrogen receptor-alpha at Ser167 is indicative of longer disease-free and overall survival in breast cancer patients. *Clin Cancer Res* 13:5769–5776
16. Weis K, Ekena K, Thomas J, Lazennec G, Katzenellenbogen B (1996) Constitutively active human estrogen receptors containing amino acid substitutions for tyrosine 537 in the receptor protein. *Mol Endocrinol* 10:1388–1398
17. Arnold SF, Melamed M, Vorojeikina DP, Notides AC, Sasson S (1997) Estradiol-binding mechanism and binding capacity of the human estrogen receptor is regulated by tyrosine phosphorylation. *Mol Endocrinol* 11(1):48–53
18. Arnold SF, Notides AC (1995) An antiestrogen: a phosphotyrosyl peptide that blocks dimerization of the human estrogen receptor. *Proc Natl Acad Sci USA* 92(16):7475–7479
19. Shiau A, Barstad D, Loria P, Cheng L, Kushner P, Agard D, Greene G (1998) The structural basis of estrogen receptor/coactivator recognition and the antagonism of this interaction by tamoxifen. *Cell* 95:927–937
20. Nettles KW, Bruning JB, Gil G, Nowak J, Sharma SK, Hahm JB, Kulp K, Hochberg RB, Zhou H, Katzenellenbogen JA et al (2008) NFKappaB selectivity of estrogen receptor ligands revealed by comparative crystallographic analyses. *Nat Chem Biol* 4(4):241–247
21. Reid G, Hubner M, Metivier R, Brand H, Denger S, Manu D, Beaudouin J, Ellenberg J, Gannon F (2003) Cyclic, proteasome-mediated turnover of unliganded and liganded ERalpha on responsive promoters is an integral feature of estrogen signaling. *Mol Cell* 11(1):695–707
22. Metivier R, Penot G, Hubner M, Reid G, Brand H, Kos M, Gannon F (2003) Estrogen receptor-alpha directs ordered, cyclical, and combinatorial recruitment of cofactors on a natural target promoter. *Cell* 115(6):751–763
23. Dutertre M, Smith C (2003) Ligand-independent interactions of p160/steroid receptor coactivators and CREB-binding protein (CBP) with estrogen receptor-alpha: regulation by phosphorylation sites in the A/B region depends on other receptor domains. *Mol Endocrinol* 17:1296–1314
24. Kobayashi Y, Kitamoto T, Masuhiro Y, Watanabe M, Kase T, Metzger D, Yanagisawa J, Kato S (2000) p300 mediates functional synergism between AF-1 and AF-2 of estrogen receptor alpha and beta by interacting directly with the N-terminal A/B domains. *J Biol Chem* 275:15645–15651
25. Campbell EJ, McDuff E, Tatarov O, Tovey S, Brunton V, Cooke TG, Edwards J (2008) Phosphorylated c-Src in the nucleus is associated with improved patient outcome in ER-positive breast cancer. *Br J Cancer* 99(11):1769–1774
26. Newby J, Johnston S, Smith I, Dowsett M (1997) Expression of epidermal growth factor receptor and c-erbB2 during the development of tamoxifen resistance in human breast cancer. *Clin Cancer Res* 3:1643–1651
27. Gutierrez MC, Detre S, Johnston S, Mohsin SK, Shou J, Allred DC, Schiff R, Osborne CK, Dowsett M (2005) Molecular changes in tamoxifen-resistant breast cancer: relationship between estrogen receptor, HER-2, and p38 mitogen-activated protein kinase. *J Clin Oncol* 23(11):2469–2476

28. Migliaccio A, Castoria G, Di Domenico M, de Falco A, Bilancio A, Lombardi M, Barone MV, Ametrano D, Zannini MS, Abbondanza C et al (2000) Steroid-induced androgen receptor-oestradial receptor beta-*Src* complex triggers prostate cancer cell proliferation. *EMBO J* 19(20):5406–5417
29. Varricchio L, Migliaccio A, Castoria G, Yamaguchi H, de Falco A, Di Domenico M, Giovannelli P, Farrar W, Appella E, Auricchio F (2007) Inhibition of estradiol receptor/*Src* association and cell growth by an estradiol receptor alpha tyrosine-phosphorylated peptide. *Mol Cancer Res* 5(11):1213–1221
30. Lin SY, Makino K, Xia W, Matin A, Wen Y, Kwong KY, Bourguignon L, Hung MC (2001) Nuclear localization of EGF receptor and its potential new role as a transcription factor. *Nat Cell Biol* 3(9):802–808
31. Aleksic T, Chitnis MM, Perestenko OV, Gao S, Thomas PH, Turner GD, Protheroe AS, Howarth M, Macaulay VM (2010) Type 1 insulin-like growth factor receptor translocates to the nucleus of human tumor cells. *Cancer Res* 70(16):6412–6419
32. Wang YN, Yamaguchi H, Hsu JM, Hung MC (2010) Nuclear trafficking of the epidermal growth factor receptor family membrane proteins. *Oncogene* 29(28):3997–4006
33. Barone I, Brusco L, Fuqua SA (2010) Estrogen receptor mutations and changes in downstream gene expression and signaling. *Clin Cancer Res* 16(10):2702–2708
34. Fuqua SA, Wiltschke C, Zhang QX, Borg A, Castles CG, Friedrichs WE, Hopp T, Hilsenbeck S, Mohsin S, O'Connell P et al (2000) A hypersensitive estrogen receptor-alpha mutation in premalignant breast lesions. *Cancer Res* 60(15):4026–4029
35. Cui Y, Zhang M, Pestell R, Curran EM, Welshons WV, Fuqua SA (2004) Phosphorylation of estrogen receptor alpha blocks its acetylation and regulates estrogen sensitivity. *Cancer Res* 64(24):9199–9208