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Fenton-like Chemistry by a Copper(I) Complex and $_{H_2O_2}$ Relevant to Enzyme Peroxygenase $_{C-H}$ Hydroxylation

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Supporting Information

Experimental details, characterization data, and X-ray crystallographic data for $[3]B(C_6F_5)_4$ (PDF)

Accession Codes

CCDC 2245700 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +441,223 336,033.

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Abstract

Lytic polysaccharide monooxygenases have received significant attention as catalytic convertors of biomass to biofuel. Recent studies suggest that its peroxygenase activity (i.e., using H₂O₂ as an oxidant) is more important than its monooxygenase functionality. Here, we describe new insights into peroxygenase activity, with a copper(I) complex reacting with H₂O₂ leading to site-specific ligand–substrate C–H hydroxylation. $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ (TMG₃tren = 1, 1, 1 – Tris $\left\{2 - [N^{2} - (1, 1, 3, 3 - tetramethylguanidino)]ethyl<math>\right\}$ amine) and a dry source of hydrogen peroxide, ($o - Tol_{3}P = O \cdot H_{2}O_{2}$) react in the stoichiometry, $\left[Cu^{I}(TMG_{3}tren)\right]^{+} + H_{2}O_{2} \rightarrow \left[Cu^{I}(TMG_{3}tren-OH)\right]^{+} + H_{2}O$, wherein a ligand *N*-methyl group undergoes hydroxylation giving TMG₃tren-OH. Furthermore, Fenton-type chemistry (Cu^I + H₂O₂ \rightarrow Cu^{II} – OH + \cdot OH) is displayed, in which (i) a Cu(II) – OH, (\cdot OH) complex could be detected during the reaction and it could be separately isolated and characterized crystallographically and (ii) hydroxyl radical (\cdot OH) scavengers either quenched the ligand hydroxylation reaction and/or (iii) captured the \cdot OH produced.

Graphical Abstract



INTRODUCTION

Oxidative degradation of biomass such as chitin and cellulose is known¹ to be carried out by bacterial and fungal lytic polysaccharide monooxygenases (LPMOs) which comprise a component of the carbohydrate active enzymes (CAZy) family.^{2,3} These mononuclear copper enzymes enable active-site chemistry in the oxidation of recalcitrant polysaccharide C1 and/or C4 C–H bonds (see the diagram in the SI⁴) possessing bond dissociation energies of ~101–104 kcal/mol.⁵ Thus, there is considerable potential to generate biofuels in a sustainable manner, by utilizing LPMOs to break down plentiful biomass materials.^{1c,2c}

Earlier studies revealed a classical monooxygenase activity for LPMOs (Scheme 1a)^{1c,2a,6} which possess a mononuclear Cu active site with a tridentate T-shaped coordination, having protein-derived ligation from 2 His residue imidazole N's plus a primary amine

 $(-NH_2)$ derived from the N-terminal His; the latter comprises a chelate, referred to as the "His brace".^{1b,3b,7} In a monooxygenase reaction cycle,⁸ a cupric-superoxide $(Cu^{II}(O_2^{-}))$ species could form via initial O₂-interaction with a copper(I) center.^{2b,8b,9} This could directly do HAA or, following electron and/or proton transfers would lead to a Cu^{II}- (hydro)peroxide entity that is further transformed into the key species which would affect the difficult hydrogen-atom abstraction (HAA) reaction (e.g., a copper(II)-oxyl $(Cu^{II} - O \cdot)$ species).^{1d,3a,8b,10}

However, in fact, recent biochemical-biophysical studies^{10i,11} detail that LPMOs also are widely functional as peroxygenases and that H₂O₂ is faster reacting with reduced copper(I) LPMOs than is molecular oxygen. The peroxygenase biochemistry (Scheme 1b) is found to lead to observable protein damage resulting in lower product yields and loss of reaction selectivity in comparison to the O2-mediated monooxygenase reactivity. Also, computational studies support the viability of LPMO peroxygenase activity.^{10h,12} Scheme 1c provides mechanistic pathways which have been proposed or can be considered, for the enzyme ligand–copper(I) ion/ H_2O_2 chemistry leading to substrate hydroxylation. The likely reactive species capable of HAA for these difficult substrates are (i) a hydroxyl radical (·OH) produced by copper Fenton chemistry, $(Cu^{I} + H_2O_2 \rightarrow Cu^{II} - OH + \cdot OH)$ (and see below).¹³ (ii) a Cu^{II} – O · · species which may be directly generated from a Cu^I + H₂O₂ reaction with release of H₂O₂; a related route, that has been suggested, could be if the ·OH moiety produced in the Fenton-like reaction, abstracts an H-atom from the Cu^{II}-hydroxide moiety, $Cu^{II} - OH + \cdot OH \rightarrow Cu^{II} - O \cdot + H_2O$ (Scheme 1c)^{8b,12,14} and (iii) a high-valent Cu(III) species,¹⁵ possibly a Cu^{III}(OH)₂ complex (not shown in Scheme 1) derived from direct homolytic cleavage of H_2O_2 in its interaction with Cu^{I} . The reactive species $Cu^{III}(OH)_2$ would affect substrate C-H bond HAA, with one hydroxide bound to copper accepting the proton and then producing H₂O, leaving behind a Cu^{II}-hydroxide species and the substrate carbon radical (R·); rebound, ¹⁶ Cu^{II} – OH + R · \rightarrow Cu^I + R – OH, would complete a catalytic cycle.

Several recent biochemical studies^{10i,17} on the Cu^{I}/H_2O_2 enzyme reaction reveal generation of protein radicals, via one-electron oxidation of a Tyr and Trp residue near the active site. Solomon and co-workers¹⁰ⁱ could demonstrate direct Cu^{II} hydroxide formation concomitant with protein radical formation, potentially derived from the \cdot OH generated and subsequent reactivity. These observations suggest that scenarios (i) or (ii) (see above) may apply, wherein a copper-mediated Fenton reaction initially occurs in LPMOs. Supporting computational results have been published.^{10h,12}

In the Fenton reaction (with Fe^{II} or Cu^I),^{13a-c,18} the particular situation present (e.g., pH in aqueous media, ligand identity) dictates whether ·OH or a high-valent metal-oxo complex forms (e.g., Fe^{IV} = O);¹⁹ under physiological conditions, carbonate radical anion (CO₃ · -) is present, rather than ·OH.^{13a-c} It is well known that iron- or copper ion-mediated Fenton chemistry effect biological substrate metal-ion/H₂O₂ oxidative damage to peptides or nucleic

acids, where \cdot OH may be generated and react in a site-specific (or localized) manner,²⁰ including possibly in LPMOs.¹² Hydrogen peroxide (or -OOH) can reduce copper(II) complexes,^{10d,21,22} yielding cuprous ions left to react with any excess H₂O₂ present, leading to Fenton chemistry. Furthermore, a recent report indicates conditions where H₂O₂ reduction of copper(II) coordination complexes is observed; this can occur in situations where the ligand which is binding to the metal ion strongly favors copper(I) (e.g., 2,9-dimethyl-1,10-phenanthroline), and \cdot OH is formed if water is present.²³

More broadly, it has been recently suggested that nature may control metal-ion active site oxidative chemistries by utilizing the Fenton reaction in a "constructive manner".²⁴ It should also be noted that the hydroxyl radical may be generated by photolysis of water at metal/ alloy surfaces (or even at the water–gas surface of water microdroplet)²⁵ and in a controlled manner be utilized for organic oxidations including conversion of methane to methanol,²⁶ removal of contaminants in water purification,²⁷ and chemistry applied to bleaching;²⁸ it may even be applied to cancer therapies.²⁹

Here, we illuminate details concerning a chemical system involving a copper-coordination complex, where an LPMO-type peroxygenase reaction is found to occur. Complex $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ reacts with "dry" $H_{2}O_{2}$, ³⁰ according to Scheme 2, where stoichiometric hydroxylation (i.e., formal insertion of an 'O'-atom) of one of the twelve (12) outer ligand methyl groups occurs:

$$\begin{split} & \left[Cu^{I}(TMG_{3}tren) \right]^{+} + H_{2}O_{2} \\ & \rightarrow \left[Cu^{I}(TMG_{3}tren\text{-}OH) \right]^{+} + H_{2}O \end{split}$$

This is a peroxygenase reaction; as Cu^I is left as a final product, a potentially catalytic system is established. As described in this report, our conclusion is that this peroxygenase reaction proceeds via Fenton-type chemistry with copper. Among the experimental observations supporting our supposition, are that a Cu^{II}-hydroxide intermediate could be detected (see below) and that an ·OH reactive species (or an equivalent) could be quenched and/or captured.

RESULTS AND DISCUSSION

 $\left[Cu^{I}(TMG_{3}tren)\right]^{+}$ (1) (Scheme 2) possesses a tripodal tetradentate N₄ ligand with strong (highly basic) alkylamine donor groups, thus having some similarity to the nitrogenous ligand environment found at the LPMO Cu-active sites. Complex **1** is known to reversibly bind molecular oxygen giving $\left[Cu^{II}(TMG_{3}tren)(O_{2}^{--})\right]^{+}$,³¹ and it was previously observed that under specific oxidizing conditions, an alkoxide–copper(II) complex $\left[Cu^{II}(TMG_{3}trenO^{-})\right]^{+}$ (2) could be isolated and structurally (X-ray) characterized (Figure 1a);³² this observation suggested that a ligand methyl group had undergone hydroxylation.

The experimental observations in that study led to our suggestions that the most likely reactive species which effected the ligand hydroxylation was a Cu^{II} -hydroperoxide, generated (i) directly from $[Cu^{II}(TMG_3tren)]^{2+} + H_2O_2(aq) + base, or$ (ii) by 1-hydroxy-2,2,6,6-tetramethyl-piperidine (TEMPO-H) reductive protonation of the superoxide complex $[Cu^{II}(TMG_3tren)(O_2^{-})]^+$, or (iii) by reduction of $[Cu^{II}(TMG_3tren)]^{2+}$ and/or $[Cu^{II}(TMG_3tren)(O_2^{-})]^+$ effected by phenols which were added. We also speculated that a $Cu^{II}(-OOH)$ could undergo O–O cleavage leading to product, via a $Cu^{II} - O$ species, since the reaction of (1) with PhIO also yielded the hydroxylated ligand alkoxide $[Cu^{II}(TMG_3tren)^{-}]^+$. However, as discussed and referenced above (Introduction), ligand-copper(II) complexes can be reduced with hydrogen peroxide, and we have ourselves observed such reactivity which appeared to lead to Cu^{I}/H_2O_2 Fenton chemistry.^{10d} Could reduction of copper(II) to copper(I) in the presence of hydrogen peroxide be involved in that 2008 study?

Thus, we thought to take advantage of this chemical system and explore new chemistry with **1** where we employ Fenton chemistry conditions that might relate to the peroxygenase chemistry in LPMOs, as described in the Introduction. Would addition of hydrogen peroxide to the cuprous complex lead to ligand methyl group hydroxylation and if so, could mechanistic aspects be investigated?

Here, in testing Cu^I/H₂O₂ reactivity, the alkoxide-copper(II) complex **2** was indeed formed in the reaction of **1** with three equiv dry H₂O₂ (via use of 1.5 equiv ($o - Tol_3P = O \cdot H_2O_2)_2$)³³ in 2-methyltetrahydrofuran (MeTHF) at -70 °C (Scheme 3). A dry solid material source of H₂O₂ allows for careful stoichometric additions as well as use of organic solvents and cryogenic reaction conditions. Observed in the **1**/H₂O₂ reaction was a change from colorless to the green compound **2** { $\lambda_{max}(\epsilon, M^{-1}cm^{-1})$:420(500), 875(270)nm} (Figure 1b). A frozen solution EPR spectrum of the reaction solution (Figure 1c) showed, as previously observed, for [Cu^{II}(TMG₃trenO⁻)]⁺(**2**),³⁴ a reverse axial signal typical of Cu(II) in a trigonal bipyramidal environment. As was determined previously using ESI-MS,³² we here also confirmed the formation of alkoxide complex [Cu^{II}(TMG₃trenO⁻)]⁺(**2**) employing cold spray ionization mass spectrometry (CSI-MS); **2** is characterized by a peak at *m*/*z* 5 18.3 (calcd *m*/*z* 5 18.3; Figure 1d).

Time resolution of the reaction was achieved by quickly injecting $(o-Tol_3PO \cdot H_2O_2)_2$ into a -70 °C solution of **1** into the prechilled CSI-MS instrument.⁴ The mass spectra clearly show peaks due to $[Cu^{II}(TMG_3tren)(OH)]^+$ (m/z = 520.3) at 10 and 54 s. This diminishes as an increasing amount of $[Cu^{II}(TMG_3trenO^-)]^+$ (m/z = 518.3) forms; the final product alkoxide builds up as the Cu^{II}-hydroxide intermediate disappears. Since we have not quantitatively determined instrument response factors for the hydroxide vs alkoxide complexes, strictly speaking we can only say that the hydroxide complex (**3**) forms first. At 120 s, the CSI-MS

signal is essentially pure Cu^{II}-alkoxide **2**; the *m*/*z* 520 peak is exactly the intensity expected and observed for authentic **2**,⁴ possessing a normal isotope distribution pattern (the effect of 63 Cu/ 65 Cu isotope abundance). In fact, we show stronger and clearer evidence for initial formation of $\left[Cu^{II}(TMG_{3}tren) - (OH)\right]^{+}$ (**3**) in other experiments, see below.

At this stage of experiments, the above results suggest:

$$\begin{bmatrix} Cu^{I}(TMG_{3}tren) \end{bmatrix}^{+} (1) + H_{2}O_{2} \\ \rightarrow \begin{bmatrix} Cu^{II}(TMG_{3}tren)(OH) \end{bmatrix}^{+} (3) + \cdot OH \end{bmatrix}$$
(1)

one of the reaction sequences described above and indicated in Scheme 1, essentially the classical Fenton reaction (with copper(I)). Independently, we could generate copper(II) -hydroxo complex $\left[Cu^{II}(TMG_3tren)(OH)\right]^+(3)^4$ and characterize experimentally its structure via single-crystal X-ray crystallography (Figure 2).⁴ Unlike the alkoxide complex $\left[Cu^{II}(TMG_3trenO^-)\right]^+(2)$, no prominent charge-transfer band is apparent for 3;⁴ however, it does display a reverse-axial EPR spectrum ($g_{\perp} = 2.22$ ($A_{\perp} = 70$ G) and $g_{\parallel} = 1.99$ ($A_{\parallel} = 85$ G)) and a prominent parent ion peak at m/z of 520.3 in CSI-MS.⁴

If the reaction in eq 1 occurs, or even if the products of $\mathbf{1} + H_2O_2$ are $(Cu^{II} - O \cdot + H_2O)$ or $Cu^{III}(OH)_2$ (see above), the $\cdot OH$ (formally) would attack one of the ligand methyl groups in order to proceed to the alkoxide product, **2**. We now present experiments whose results suggest that this is likely the case. When excess $[Cu^{I}(TMG_3tren)]^+(1)$ is reacted with the H_2O_2 reagent, $[Cu^{I}(TMG_3tren)]^+(1)/H_2O_2 = 5:1$ (via use of 0.1 equiv ($o - Tol_3P = O \cdot H_2O_2)_2$), we see from Figure 3a that these reaction conditions do not lead to the observation of the 420 nm UV – vis band associated with alkoxide $[Cu^{II}(TMG_3trenO^-)]^+(2)$ (i.e., that shown in Figure 1b). This reaction of excess **1** with H_2O_2 , conditions such that only one (1) equiv H_2O_2 would react with one molecule of **1**, reveals that essentially no Cu(II) is produced; we observe only ~5% of expected EPR signal intensity which would be due to the presence of a full equivalent of Cu(II) complex (Figure 3b).³⁵

Thus, these results, for reaction conditions where $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)/H_{2}O_{2} = 5:1$, reveal that:

$$\begin{aligned} & \mathbf{5} \Big[\mathrm{Cu}^{\mathrm{I}} (\mathrm{TMG}_{3} \mathrm{tren}) \Big]^{+} + \mathbf{1} \mathrm{H}_{2} \mathrm{O}_{2} \\ & \rightarrow 4 \Big[\mathrm{Cu}^{\mathrm{I}} (\mathrm{TMG}_{3} \mathrm{tren}) \Big]^{+} + \mathbf{1} \Big[\mathrm{Cu}^{\mathrm{I}} (\mathrm{TMG}_{3} \mathrm{tren} - \mathrm{OH}) \Big]^{+} \\ & (+\mathbf{1} \mathrm{H}_{2} \mathrm{O}) \end{aligned}$$

i.e., the oxygenation (by H_2O_2) of the Cu^I-bound TMG₃tren ligand in $[Cu^I(TMG_3tren)]^+(1)$, to give hydroxylated ligand TMG₃tren-OH as a copper(I) complex (following rebound; see also, below), occurs via a peroxygenase stoichiometry, the reaction described in Scheme 2.

However, to further confirm these conclusions, it is required that we show that ligand hydroxylation has occurred, i.e., $\left[Cu^{I}(TMG_{3}tren-OH)\right]^{+}$ is a product. This is, in fact, the case. For the $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)/H_{2}O_{2} = 5:1$ reaction, the product mixture was quenched at -70 °C with 2,6-dimethyl phenyl isocyanide (DIMPI, as a strong copper(I) specific ligand), the solvent was removed, and the reaction mixture was warmed to RT and then extracted with KCN/CD_{3}NO_{2}.⁴ MALDI-TOF MS analysis shows that the most intense peak present is due to unreacted ligand TMG_{3}tren (Figure 3c: m/z 441.3, {(TMG_{3}tren) + H⁺} (calcd m/z 441.3)) which was present in excess. The other major product is one where the methyl group of one ligand has been converted to a $-CH_{2}OH$ (TMG_{3}tren-OH) functionality and in amounts closely correlating with the quantity of $H_{2}O_{2}$ added, m/z 479.2, {(TMG_{3}tren-OH) + Na⁺} (calcd m/z 479.3; Figure 3c). The TMG_{3}tren-OH peak has very close to 1/4 of the intensity as the peak due to unhydroxylated ligand, TMG_{3}tren. Thus the reaction yields are very high, appearing to be nearly quantitative since with the limited amount of $H_{2}O_{2}$ present, only one out of 5 mole-equiv of $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ can undergo conversion to TMG_{3}tren-OH.

Additional CSI-MS based experiments with these reaction conditions where $[Cu^{I}(TMG_{3}tren)]^{+}(1)/H_{2}O_{2} = 5:1$ provide very strong evidence for the Scheme 2 sequence of reactions, i.e., that $[Cu^{II}(TMG_{3}tren)(OH)]^{+}(3)$ is the initially formed species (as an intermediate). By contrast to the reaction conditions with excess hydrogen peroxide, i.e., the data shown in Figure 1, here $[Cu^{II}(TMG_{3}tren)(OH)]^{+}(3)$ (m/z = 520.3) is formed in a highly persistent manner (Figure 3d), lasting for many minutes prior to the start to observing alkoxide $[Cu^{II}(TMG_{3}trenO^{-})]^{+}(2)$ formation (m/z = 518.3; Figure 3d, from 7 min after sample injection, on). It should be emphasized that formation of hydroxide complex **3** implies that the hydroxyl radical must be forming concomitantly (also see Scheme 2).

Experimental observations that further support our characterization of this peroxygenase system (Scheme 2) are:

1. The TMG₃tren ligand has been hydroxylated prior to formation of the final Cu^{II} -alkoxide complex, supporting the reaction as given by eq 1 (vide supra). When the $[Cu^{I}(TMG_{3}tren)]^{+}(1)/H_{2}O_{2} = 1:3$ is quenched prior to alkoxide $[Cu^{II}(TMG_{3}trenO^{-})]^{+}(2)$ formation (based on following UV–vis changes up to where the 420 nm absorption just starts to be observable), a similar workup and analysis of organics reveal that high yields (>95%) of TMG_{3}tren-OH are obtained. The prominent ion peak at m/z of 457.2 is assigned to $\{(TMG_{3}tren-OH) + H^{+}\}$ (calcd m/z 457.3; Figure 4a). Only a trace peak for the starting initial unhydroxylated ligand, TMG_{3}tren (m/z 441.3), is observed in the MALDI-TOF MS spectrum (Figure 4a).

2. With the excess dry H₂O₂ added, we observed additional products of ligand oxygenation, including the overoxidized aldehyde product. Following workup of the reaction mixture containing [Cu^{II}(TMG₃trenO⁻)]⁺(2) and utilizing the DIMPI procedure to remove copper ions (vide supra), mass spectrometric analysis of the organics present reveals that together with a small amount of un-oxidized/oxygenated TMG₃tren, several ligand oxidized types are present (Figure 4). They are (i) the ligand hydroxylated alcohol L-OH {[TMG₃tren - (CH₃)N - CH₂OH] + Na⁺} (*m*/*z* 479 . 2, calcd *m*/*z* 479 . 3; Figure 4b), (ii) the N–H species arising from TMG₃tren-(CH₃)N – CH₂OH *N*-dealkylation, L-NH, {[TMG₃tren - (CH₃)N – H] + K⁺} (*m*/*z* 465 . 2, calcd *m*/*z* 465 . 3; Figure 4b), (released formaldehyde is observed),⁴ and (iii) a small amount of overoxidized aldehyde species L-CHO {[TMG₃tren-(CH₃)N – C(*O*)H] + K⁺} (*m*/*z* 493.2, calcd *m*/*z* 493 . 3; Figure 4b).

To provide still further evidence for this Fenton-like chemistry, we sought to identify the presence of \cdot OH (or its equivalent) by employing trapping reagents and/or external substrates which have C–H/O – H bonds (Scheme 4). Inclusion of ten (10) equiv 2,4,6-tri*t*-butylphenoxyl radical (T*t*BuArO \cdot) with solutions of $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ prior to addition of H₂O₂(1/H₂O₂ = 5:1) quells the peroxygenase type ligand hydroxylation chemistry (Scheme 2); little or no alkoxide complex **2** is formed (UV–vis criterion). We deduce that \cdot OH produced by the 1/H₂O₂ reaction reacts with excess T*t*BuArO \cdot present, and elimination of isobutylene (formed in 59% yield) as well as additional documented phenolic chemistry³⁶ gives 2,6,-di*t*-butyl-1,4-hydroquinone (as explained in the SI), which is detected in GC–MS as 2,6-di*tert*butyl-1,4-benzoquinone formed in 20% yield based on copper (so effectively ~100%).⁴ This implies capture of " \cdot OH" in near quantitative yields. However, addition of only two equiv. T*t*BuArO \cdot gave only an ~25% yield of the benzoquinone; the efficiency of trapping goes up as the quantity of added trapping agent is increased. Related experiments with excess H₂O₂ and monitored by EPR spectroscopy are also consistent with our conclusions (Figure S5).⁴

For an experiment where 10 equiv trityl radical were added (as Gomberg's dimer) (Scheme 4), again no alkoxide complex **2** formed (UV–vis criterion). Here, the ·OH released from the copper complex (1)/H₂O₂ reaction would be trapped by the trityl radical to directly form triphenylcarbinol; this was generated in 18% yield. As was mentioned above, due to the stoichiometry of this reaction, this 18% yield is very high, as 20% is the theoretical maximum. Again, when only a limited amount of added Gomberg's dimer is used (2 equiv), the trapping efficiency is only 3% based on the amount of copper and the stoichiometry of reaction employed ($1/H_2O_2 = 5:1$). Excess amounts of added 2,6-di-*t*-butyl-4-methoxyphenol or xanthene were also observed to "capture" the ·OH generated in reactions of **1** with H₂O₂ (Scheme 4), through HAA to produce 2,6-di-*t*-butyl-4-methoxy phenoxyl radical and xanthone, respectively.⁴ See Table S3 for details/yields for the trapping/quenching experiments.

It is interesting to survey a number of recently published LPMO biomimetic studies.^{22,37} In those reports involving ligand–copper(II) complex reactions with added hydrogen peroxide and an oxidizable substrate, Simaan and Hitomi,^{37a} Kaizer,²² Itoh,^{37b} Castillo,^{37c,d} Cowan,^{37e} and their co-workers have utilized mono- or poly-nuclear Cu^{II}-complexes, some with a His-Brace like ligand. Added H₂O₂ (aq) likely leads to Cu^{II}-OOH moiety and to oxidation of ligands (e.g., ACC oxidase substrate analogs) or exogenous glucose derivatives (e.g., as polysaccharides or surrogates). However, neither a specific O₂ reduced-derivative (e.g., ·OH) nor a metal-based strong oxidant (e.g., a Cu^{II}-oxyl) has been yet identified. It is notable, however, that Simaan and Hitomi,^{37a} and Kaizer²² provide evidence that excess H₂O₂ at some stage effects cupric ion reduction (via Cu–O heterolytic cleavage of the presumed Cu^{II} – OOH moiety)^{10c,10,21b} and the real oxidant species is something like "Cu^I – OOH." This latter hypothesis points to Fenton-like reactivity.

Based on these experimental results, we can establish plausible reaction pathways for the $[Cu^{I}(TMG_{3}tren)]^{+}(1)$ and dry $H_{2}O_{2}$ reaction which leads to C–H activation in an overall peroxygenase reaction, shown in Scheme 5 for the differing stoichiometries tested experimentally. The most likely initial reaction is formation of a Cu^{II}-hydroxide complex (**3**) plus a ·OH species. The latter reactive entity performs HAA from a ligand methyl group, producing water and a ligand carbon radical; subsequent rebound from the Cu^{II}-hydroxide gives Cu^I(TMG₃tren-OH).³⁸ This reaction mechanism was evaluated and is further supported, by density functional theory (DFT) calculations on the full complex and its reaction with $H_{2}O_{2}$.

Figure 5 shows the calculated reaction coordinate based on the proposed mechanism in Scheme 5, top (see the SI for computational details). In the initial structure (Figure 5, **0**), the H_2O_2 associates with the complex through van der Waals interactions but does not bind directly to the Cu (Cu–O distance: 3.27 Å). The reaction proceeds through homolytic cleavage of the H₂O₂ forming a Cu^{II} – OH and \cdot OH that is 14.9 kcal/mol downhill in ΔG (Figure 5, 2) through a low barrier of $\Delta G^{\ddagger} = 3.0$ kcal/mol (Figure 5, 1). Immediately after homolytic cleavage, the resulting OH (Figure 5, 2a) is not properly oriented to abstract an H atom from the ligand methyl group and must reorient to the proper conformation (Figure 5, 2b) to perform HAA from the C-H bond. This rearrangement involves a small increase in the O···O distance (2.25 Å in 2a to 2.56 Å in 2b) and a rotation of the ·OH fragment; this proceeds through a low barrier of 0.9 kcal/mol (Figure 5, 2a-2b). From 2b, the ·OH performs HAA from the ligand methyl C–H bond with almost no barrier, $\Delta G^{\ddagger} = 0.35$ kcal/mol (Figure 5, 3), producing a water molecule and the ligand methyl radical. This HAA step is further downhill by 23.1 kcal/ mol in ΔG (Figure 5, 4). Finally, the methyl radical rebound occurs with the highest barrier in this process, $\Delta G^{\dagger} = 7.0$ kcal/mol (Figure 5, 5), due to the significant steric reorganization of the complex to reach this transition state. The Cu^I hydroxylated ligand complex product 6 (i.e., Cu^I(TMG₃tren-OH)) is 28.1 kcal/mol downhill from the previous step and 66.5 kcal/mol downhill from the starting structure. Each step in the proposed mechanism is thermodynamically favorable, with very low barriers for

O–O cleavage and HAA, and a reasonable, limiting barrier for the rebound hydroxylation. Furthermore, the \cdot OH reorientation to a conformation conducive to HAA from the ligand would result in a finite lifetime for the Cu^{II} – OH and \cdot OH, consistent with the observation of species **3** by ESI-MS (Figures 2 and 3) and the radical trapping results. Thus, the calculations in Figure 5 show that the proposed mechanism in Scheme 5 is thermodynamically and kinetically feasible, and consistent with the experimental results presented above.

Scheme 2 and the upper part of Scheme 5 represent a first round of a peroxygenase catalytic cycle, as Cu(I) is regenerated and can accept a new substrate (here, a new unhydroxylated ligand). However, when excess H_2O_2 is present (Scheme 5, bottom), $\left[Cu^{I}(TMG_3tren-OH)\right]^+$ is oxidized by hydrogen peroxide to a cupric form, subsequently leading to TMG₃tren-OH deprotonation and formation of alkoxide $\left[Cu^{II}(TMG_3trenO^-)\right]^+$ (2). *N*-Dealkylation can otherwise occur (vide supra), producing formaldehyde plus a $Cu^{II}(TMG_3tren-(CH_3)N - H)$ species, as we observe experimentally.

CONCLUSIONS

In this study, we have provided considerable new insights into site-specific Fentontype peroxygenase chemistry, quite likely relevant to LPMOs³⁹ and perhaps also to copper-dependent *p*-methane monooxygenases (*p*MMOs).^{24,40} Using a synthetic analog $[Cu^{I}(TMG_{3}tren)]^{+}(1), Cu^{I}/H_{2}O_{2}$ reactions occur. Our experimental results indicate that this leads to a cupric-hydroxide plus hydroxyl radical as suggested in the study on a LPMO by Solomon and co-workers;¹⁰ⁱ subsequent *N*-methyl group hydroxylation occurs leaving behind Cu^{I} . The generation of a ·OH intermediate (or possibly a $Cu^{III}(OH)_{2}$ or $Cu^{II} - O \cdot$ species) was demonstrated via capture or quenching with radical scavengers or external substrates. The proposed reaction mechanism is further determined to be thermodynamically and kinetically feasible by DFT reaction coordinate calculations. The overall reaction, $[Cu^{I}(TMG_{3}tren)]^{+} + H_{2}O_{2} \rightarrow [Cu^{I}(TMG_{3}tren-OH)]^{+} (+H_{2}O)$, is consistent with LPMO peroxygenase catalytic behavior. This study provides a fresh perspective on Fentonlike copper chemistry and previously proposed mechanisms and nature of key intermediates in peroxygenase reactivity, including LPMOs.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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(38). Note: we tested directly for the viability of this latter process and the results support our rebound hypothesis. When the hydroxide complex $\left[Cu^{II}(TMG_{3}tren)(OH)\right]^{+}$ (3) is exposed to 10 equiv

trityl radical (as Gomberg's dimer) at -70 °C, $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ was produced in high yields (ESI detection of 1 and depletion of trityl radical occurred (UV-vis monitoring)) and a 67% yield of triphenylmethanol was obtained. See Supporting Information.

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Figure 1.

(a) ChemDraw representation of $[Cu^{II}(TMG_3trenO^-)]^+(2)$ based on its crystallographic determination.³² (b) UV–vis spectral changes (over 1 h) when $[Cu^{I}(TMG_3tren)]^+(1)$ reacts with three equiv H₂O₂ in MeTHF at –70 °C. (c) X-band EPR spectrum (red) { $g_{\perp} = 2.27$ ($A \perp = 82$ G) and $g_{\parallel} = 1.99$ ($A_{\parallel} = 82$ G)} and simulation (black) of complex **2** in frozen MeTHF at 20 K. (d) Time-resolution of CSI-MS spectrum for the formation of **2** upon addition of 3 equiv H₂O₂ to a solution of 1 at –70 °C, 10 s (red) after injection, then at 54 s (green), and finally at 120 s (black), which is identified as pure alkoxide complex **2**.



Figure 2.

Displacement ellipsoid plot (30% probability level) of one of the two crystallographically independent $\left[Cu^{II}(TMG_{3}tren)(OH)\right]^{+}$ cations (3) at 110(2) K. Hydrogen atoms and lattice solvent molecules are omitted for clarity except for a hydrogen atom on the hydroxo ligand. The hydroxo O-atom is H-bonded to two partially occupied crystal lattice water molecules (not shown); O1A…O1W (H-bonding) = 2.771 Å (gray, C; white, H; blue, N; red, O; green, Cu).

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Figure 3.

(a) UV-vis spectral changes of **1** with 0.2 equiv H_2O_2 in MeTHF at -70 °C. (b) X-band EPR spectrum of authentic complex **2** (red) and the product solution obtained with 0.2 equiv H_2O_2 added to **1** (blue) in frozen MeTHF at 20 K. (c) Matrix-assisted laser desorption/ ionization-time-of-flight mass spectrometry (MALDI-TOF MS) spectrum; the reaction of excess **1** and H_2O_2 after metal ions were removed by treatment with DIMPI and KCN/CD₃CN. (d) CSI-MS spectra for the reaction of **1** and 0.2 equiv H_2O_2 in MeTHF at -70 °C. The times indicated in the various panels indicate the number of seconds or minutes following sample injection. Also, see the text.



Figure 4.

MALDI-TOF MS spectrum; metal ions removed by treatment with DIMPI and KCN/CD₃CN. (a) Prior to, or (b) after, the formation of Cu^{II} -alkoxide species with excess of H_2O_2 added to **1**. (c) Oxidized products in the reaction of **1** and excess of H_2O_2 .

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Figure 5.

DFT-calculated reaction coordinate for homolytic H_2O_2 cleavage, subsequent HAA and rebound ligand hydroxylation by $\left[Cu^{I}(TMG_3tren)\right]^+$. Optimized structures and singlet energies are shown for each species. Thermodynamics are calculated at -70 °C.





LPMO Reaction Scheme (a) Monooxygenase and (b) Peroxygenase Reaction Pathway; (c) Proposed Mechanisms Relevant to the LPMOs Cu-Site, Processing H₂O₂



Scheme 3.

Reaction of Complex $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ with 3 equiv $H_{2}O_{2}$ Leads to Cu^{II} -Alkoxide Complex (2)



Scheme 4.

Capture/Trapping of a Hydroxyl Radical (·OH) Derived from $\left[Cu^{I}(TMG_{3}tren)\right]^{+}(1)$ Reactivity with Hydrogen Peroxide^{*a*}

^{*a*} Partial or nearly full inhibition of the peroxygenase chemistry where **1** is converted to alkoxide $\left[Cu^{II}(TMG_{3}trenO^{-})\right]^{+}$ (**2**) occurs. Also, see the text.

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Scheme 5.

Proposed Courses of Reaction of $[Cu^{I}(TMG_{3}tren)]^{+}(1)$ with Varying Amounts of $H_{2}O_{2}{}^{a}$

^{*a*} (Upper): $1/H_2O_2 = 5:1$ and the chemistry shown is for that one complex which reacts with H_2O_2 in a stoichiometric manner. (Lower): excess H_2O_2 relative to complex (1) produces the same hydroxylated ligand complex Cu^{I} (TMG₃tren-OH); however the excess oxidant present leads to further chemistry (far right).



Scheme 2.

Complex $[Cu^{I}(TMG_{3}tren)]^{+}(1)$ Reacts with Dry $H_{2}O_{2}$ to Afford Cu^{I} Complex Product Where a Ligand Methyl Group Has Been Hydroxylated, in Accordance with the Peroxygenase Pathway Postulated in LPMOs; See Text for Further Explanation