

CLINICAL and MOLECULAR HEPATOLOGY

The forum for latest knowledge of hepatobiliary diseases



ChatGPT performance on cirrhosis and HCC Questions

TACE for HCC: 2023 KLCA Practical Recommendations

TARE vs TKI in HCC with Vp1–3 PVT

Core indicators for viral hepatitis elimination in Korea

Fatty liver on chronic hepatitis B outcome

Review

Acute hepatitis C virus infection: clinical update and remaining challenges

Chen-Hua Liu^{1,2,3} and Jia-Horng Kao^{1,2,4,5}

¹Department of Internal Medicine, National Taiwan University Hospital, Taipei; ²Hepatitis Research Center, National Taiwan University Hospital, Taipei; ³Department of Internal Medicine, National Taiwan University Hospital, Yun-Lin Branch, Yunlin; ⁴Graduate Institute of Clinical Medicine, National Taiwan University College of Medicine, Taipei; ⁵Department of Medical Research, National Taiwan University Hospital, Taipei, Taiwan

Acute hepatitis C virus (HCV) infection is a global health concern with substantial geographical variation in the incidence rate. People who have received unsafe medical procedures, used injection drugs, and lived with human immunodeficiency virus are reported to be most susceptible to acute HCV infection. The diagnosis of acute HCV infection is particularly challenging in immunocompromised, reinfected, and superinfected patients due to difficulty in detecting anti-HCV antibody seroconversion and HCV ribonucleic acid from a previously negative antibody response. With an excellent treatment effect on chronic HCV infection, recently, clinical trials investigating the benefit of direct-acting antivirals (DAAs) treatment for acute HCV infection have been conducted. Based on the results of cost-effectiveness analysis, DAAs should be initiated early in acute HCV infection prior to spontaneous viral clearance. Compared to the standard 8–12 week-course of DAAs for chronic HCV infection, DAAs treatment duration may be shortened to 6–8 weeks in acute HCV infection without compromising the efficacy. Standard DAA regimens provide comparable efficacy in treating HCV-reinfected patients and DAA-naïve ones. For cases contracting acute HCV infection from HCV-viremic liver transplant, a 12-week course of pangenotypic DAAs is suggested. While for cases contracting acute HCV infection from HCV-viremic non-liver solid organ transplants, a short course of prophylactic or pre-emptive DAAs is suggested. Currently, prophylactic HCV vaccines are unavailable. In addition to treatment scale-up for acute HCV infection, practice of universal precaution, harm reduction, safe sex, and vigilant surveillance after viral clearance remain critical in reducing HCV transmission. (*Clin Mol Hepatol* 2023;29:623-642)

Keywords: Acute infection; Hepatitis C virus; Direct-acting antiviral

INTRODUCTION

Hepatitis C virus (HCV) infection is a global health concern and a major risk factor for cirrhosis, hepatocellular carcinoma (HCC), hepatic decompensation, and liver transplantation. Notably, the elimination of HCV by 2030 has been proposed

a public health target by the World Health Organization (WHO).¹ Although the global prevalence of HCV has decreased from 0.9% to 0.7% from 2015 to 2020,²⁻⁴ there remains substantial geographical variation in its incidence rate.

While the gold-standard management for chronic HCV infection has been well established, there is yet a standardized

Corresponding author : Jia-Horng Kao

Graduate Institute of Clinical Medicine, National Taiwan University College of Medicine and Hospital, 7 Chung-Shan South Road, Taipei 10002, Taiwan
Tel: +886-2-23123456 ext. 67307, Fax: +886-2-23825962, E-mail: kaojh@ntu.edu.tw
<https://orcid.org/0000-0002-2442-7952>

Editor: Grace Wong, Chinese University of Hong Kong, Hong Kong

Received: Oct. 30, 2022 / **Revised:** Jan. 27, 2023 / **Accepted:** Feb. 16, 2023

management strategy for acute HCV infection. Considering the patients with acute HCV infection are at risk of progressive liver disease and may lead to further viral transmission, it is clinically crucial to develop an effective management strategy for this condition. In this review, we will summarize and discuss the epidemiology, clinical presentation, diagnosis, therapeutic advances, and challenges for acute HCV infection.

EPIDEMIOLOGY

Precise real-world data on the incidence of HCV infection are only available in a limited number of countries, while some studies have attempted to estimate the rates of new HCV infection through modeling based on published data.⁵ The incidence of HCV infection worldwide has reached its peak recently (Fig. 1).⁶ In 2019, the number of newly acquired HCV infection was reported to be 1.5 million per estimation by the WHO.⁷ Although the global incidence of HCV infection has declined since the second-half of the twentieth century, evidenced by an annual decrease of about 250,000 cases during 2015–2019, there remains substantial geographical variations.⁸ The East Mediterranean and European regions are noted with the highest rates of infection, accounting for 470,000 (62.5 per 100,000) and 300,000 (61.8 per 100,000) cases annually (Fig. 2).^{7,8} Table 1 shows the common parenteral routes of HCV transmission. In the East Mediterranean region, the most common routes of HCV transmission are unsafe invasive medical procedures, such as blood transfusion and injections. In the European region, particularly, Eastern Europe, injection drug use (IDU) accounts for a substantial proportion of HCV viral transmission. Furthermore, males account for around 54.6% of new HCV infections in 2019.⁸

People receiving unsafe medical procedures

Unsafe blood transfusion is not considered a major route of HCV transmission at a population level, since blood transfusion does not occur as common as other medical procedures, such as dental treatment. However, concerns for HCV infection via blood transfusion remain present in underdeveloped or developing regions or countries, where the prevalence of HCV is high and the quality of blood screening is suboptimal. The Global Disease Burden (GDB) estimated that the case number of new HCV infections resulted from unsafe medical injections ranged from 952,111 to 1,867,904 in 2000 and 157,592 to 315,120 in 2010.⁹ Additionally the decrease in the number of new HCV infections seemed correlated with the drop in event number of unsafe medical injections, which was 1.35 and 0.36 per year in 2000 and 2010, respectively.¹⁰ However, the rate of injection device re-use is as high as 14%



Figure 1. Peak incidence in hepatitis C virus infection by calendar years in sample countries. Data adapted from that presented in Thrift et al.⁶

Abbreviations:

HCV, hepatitis C virus; PWID, people with inject drugs; PLWH, people living with human immunodeficiency virus; DAA, direct-acting antiviral; SVR, sustained virologic response; HCC, hepatocellular carcinoma; WHO, World Health Organization; IDU, injection drug use; GDB, global disease burden; DOPPS, Dialysis Outcomes Practice Patterns Study; PY, person-year; IFN, interferon; HBV, hepatitis B virus; HIV, human immunodeficiency virus; CDC, Center for Disease Control; MSM, men have sex with men; PrEP, pre-exposure prophylaxis; ALT, alanine aminotransferase; HLA, human leukocyte antigen; EIA, enzyme immunoassay; PEG-IFN, pegylated interferon; RBV, ribavirin; AE, adverse event; SOF, sofosbuvir; FDA, food and drug administration; ITT, intention-to-treat; PP, per-protocol; LDV, ledipasvir; GT, genotype; PrOD, paritaprevir/ritonavir/ombitasvir plus dasabuvir; EBR, elbasvir; GZR, grazoprevir; GLE, glecaprevir; PIB, pibrentasvir; VEL, velpatasvir; OAT, opioid agonist therapy; DCV, daclatasvir; VOX, voxilaprevir; QALY, quality-adjusted life year; AASLD, American Association for the Study of Liver Diseases; IDSA, Infectious Disease Society of America; EASL, European Association for the Study of the Liver; PEP, post-exposure prophylaxis; ChAd, chimpanzee adenovirus; MVA, modified vaccinia Ankara; NS, nonstructural protein; HVR, hypervariable region

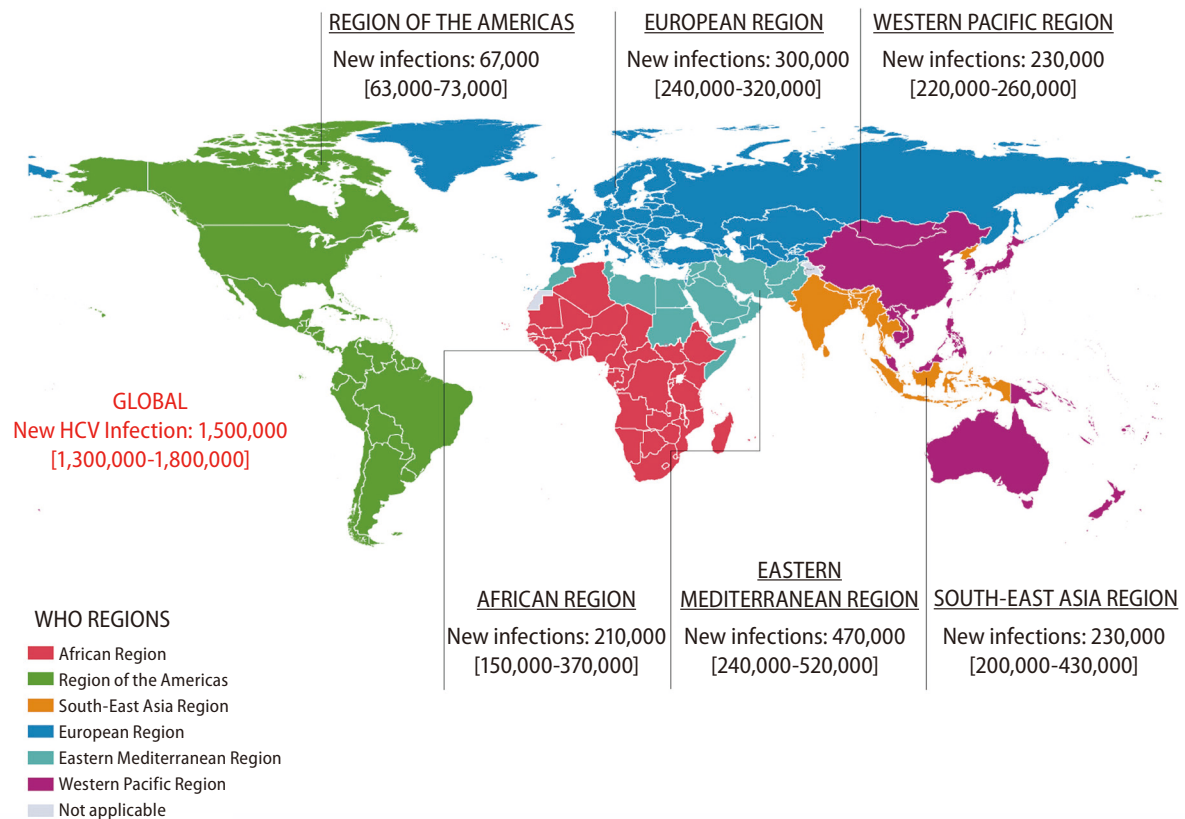


Figure 2. New hepatitis C virus (HCV) infections by World Health Organization region, 2019.

Table 1. Potential routes of HCV transmission

Transmission route
Healthcare-associated transmission
Blood transfusion
Medical injection
Dental treatment
Surgical treatment, including organ transplantation
Hemodialysis
Needlestick injury
IDU
Sexual transmission
Others
Tattoo
Body piercing
Shaving
Nail trimming
Vertical transmission

HCV, hepatitis C virus; IDU, injection drug use.

and 5% in East Mediterranean and South-East Asia, respectively, which raises the concerns about potential HCV transmission due to insufficient device sterilizations in these regions.^{11,12}

Dental visits and hemodialysis are highly associated with new HCV infections. Surveys conducted in Italy and Egypt have shown an odds ratio of acute HCV infection ranging from 1.5 to 3.7 in people who had undergone dental treatment.¹³⁻¹⁵ The Dialysis Outcomes Practice Patterns Study (DOPPS) has reported that the incidence of HCV in hemodialysis patients was 2.9 per 100 person-years (PYs) in 1996 and 1.2 per 100 PYs in 2015.¹⁶ Hemodialysis patients who are coinfecting with hepatitis B virus (HBV) or human immunodeficiency virus (HIV), have been on dialysis for more than ten years, and stay in facilities with an HCV prevalence >20% are at a higher risk of acute HCV infection. Moreover, breaches in sanitary and disinfection practices, such as inadequate hand washing or gloves changing by medical staff before and after patient care, contamination of hemodialysis machine, and the use of shared medication requiring injection, may also

contribute to viral transmission.^{17,18} Although the incidence of HCV infection due to invasive medical procedures has decreased drastically in developed countries, the high incidence remaining in underdeveloped or developing countries suggest that there is still room for improvement in the universal practice of precaution against HCV.

People who inject drugs (PWID)

Of the estimated cases of newly contracted HCV infection in 2015, 23% were attributed to IDU. PWID may account for as high as 60% of newly contracted HCV infections in developed countries.^{19,20} Specifically, young adults aged <30 (12.8–25.1 per 100 PYs) and those who have been incarcerated (5.5–31.6 per 100 PYs), show the highest incidence of HCV.²¹ In young PWID, the risk of HCV contraction is highest (133 per 100 PYs) during the first year of IDU.^{22,23} Furthermore, a meta-analysis enrolling 9,235 PWID from 28 studies yielded a sex difference in the risk of HCV contraction, with the pooled incidence rates being 20.36 per 100 PYs in females and 15.20 per 100 PYs in males (risk ratio: 1.36:1).²⁴ Although such difference was suggested to be mainly associated with different behavioral and social risks among female and male PWID, the actual mechanisms remain elusive, with other potential mediators left largely unexamined.

Although the incidence of HCV in western Europe, Canada, and Australia has been relatively stable or declining, the number remains high or even increasing in the United States (US) and developing countries.^{25–29} Data from the US Center for Disease Control (CDC) HCV surveillance indicated that HCV incidence has increased by 124% from 2013 to 2020. People who reported IDU comprised 66% of new HCV cases in 2020. Similar to the earlier annual reports, population aged from 20 to 39 had the highest incidence of acute HCV infection.²⁹ Shared use of syringes and drug preparation equipment, more frequent injections, having numerous injecting partners, particular injection drug types, having HIV coinfection, and low socioeconomic state are the key risk factors of HCV transmission among PWID.^{22,25,28,30}

People living with HIV (PLWH)

From 1990 to 2014, when interferon (IFN)-free direct-acting antivirals (DAAs) were unavailable, the CASCADE collaboration study, which recruited 16 cohorts across Europe, Canada,

and Australia, found a lack of decline in HCV incidence among HIV-positive men who have sex with men (MSM).³¹ The GDB showed a trend of decreasing acute HCV infection in HIV-positive MSM after 2014, when patient education, mass screening, and scaled-up DAA treatment, were first implemented, particularly in Europe, Canada, and Australia. A meta-analysis summarizing reports between 2000 and 2019 on HIV-positive MSM revealed that the pooled incidence of HCV infection was 8.46 per 1,000 PYs. Although the pooled incidence of HCV infection was 0.12 per 1,000 PYs in HIV-negative MSM not on HIV pre-exposure prophylaxis (PrEP), the incidence increased to 14.80 per 1,000 PYs in those on PrEP.³² Different from the aforementioned risk factors for acute HCV infection in PWIDs, more frequent high-risk sexual behaviors, including unprotected anal intercourse, group sex, having multiple sex partners, sharing sex toys, and recreational drug use, were found to be the major risk factor for HCV infection in HIV-positive MSM.^{33–36}

CLINICAL PRESENTATION

After exposure to HCV, there is a window of one to two weeks before serum HCV ribonucleic acid (RNA) becomes detectable. In patients who develop symptoms, the incubation period between exposure and symptom onset ranges from 2 to 12 weeks. Approximately 80% of people with acute HCV infection do not exhibit symptoms. The remaining 20% may exhibit fever, fatigue, nausea, vomiting, diarrhea, abdominal pain, dark urine, and jaundice.^{37,38} Because the symptoms are usually mild and non-specific, a diagnose of HCV infection is difficult during the acute phase. The elevation of serum alanine transaminase (ALT), an indicator of hepatic injury, typically occurs later than four weeks after viral exposure. Fulminant hepatitis and acute liver failure rarely occur in people with acute HCV infection.³⁹ However, the JFH1, a cell-culture-grown HCV genotype (GT) 2a virus, has been found in a Japanese patient with fulminant HCV infection.^{40–42}

During the acute phase of HCV infection, serum HCV RNA and ALT levels may have significant fluctuations. About 80% of people with acute HCV infection will develop chronic infection, generally defined as the persistence of viremia for more than six months.² Various host and viral factors, including gender, race, HIV or HBV coinfection, presentation of

jaundice, interleukin-28B polymorphism, human leukocyte antigen (HLA) class II alleles, HCV-specific T cell response, viral genotype, peak HCV RNA level, and diversity of viral quasispecies, may affect the course and outcome of acute HCV infection (Table 2).⁴³⁻⁶³ The period between viral exposure and seroconversion consists of three phases: (1) the pre-ramp-up phase (2–14 days after exposure), with intermittent low or below-detection level of HCV RNA; (2) the ramp-up phase (the next 8–10 days following the first phase), with an exponential increase of serum HCV RNA levels; (3) the plateau phase (45 to 68 days following the second phase), with stable serum HCV RNA levels.^{55,64} Although none of the aforementioned host or viral factors can accurately predict spontaneous resolution, assessing the dynamics of HCV RNA levels during the early phase of infection may help discriminate the development of acute resolving hepatitis or chronic infection.⁶⁵ Cases who experience viral rebounds or persistent viral plateau (<1 log₁₀ IU/mL decline in HCV RNA levels following peak) after prior partial viral control (≥1 log₁₀ IU/mL decline in HCV RNA levels following peak) are more likely to develop chronic infection. In contrast, people with significant viral decline after the plateau phase confer spontaneous HCV clearance.⁵⁵ Studies have suggested that the cumulative frequencies of acute resolving HCV infection were 80%, 90%, and 100% in patients who reached HCV RNA clearance at months

3, 4, and 5 of disease onset, respectively.^{66,67} In PLWH with acute HCV infection, those whose serum HCV RNA levels did not reduce for ≥2 log₁₀ at week 4 of diagnosis or remained detectable at week 12 of diagnosis were at a high risk of chronicity.⁶⁸

Superinfection refers to a primary infection of HCV followed by a secondary infection with a distinct HCV strain at a later time point.⁶⁹ Case reports have demonstrated that HCV superinfection may occur in chimpanzees and humans, including PWID, PLWH, organ transplant recipients, and people who receive multiple transfusion or endoscopic examinations.⁷⁰⁻⁷⁵ Fluctuating or persistently elevated ALT levels and/or hepatitis flares may be observed during HCV superinfection.^{69,76} As compared to the relatively benign clinical course in individuals with acute HCV mono-infection, HCV superinfection in patients with chronic HBV infection may result in more aggressive liver diseases.⁷⁷ This is particularly relevant in HBV endemic areas, such as Asia, sub-Saharan Africa, and South America. Studies have indicated that acute HCV infection in HBV carriers may carry a significant risk of developing fulminant or subfulminant hepatitis. In a series of 93 HBV carriers who were superinfected with HCV, the proportions of patients with hepatic decompensation, hepatic failure and mortality were 34%, 11% and 10%, respectively.⁷⁸

Table 2. Host and viral factors that favor spontaneous HCV clearance following acute infection

Host factors
Female ⁴³⁻⁴⁵
White ethnicity ⁴⁶
Lack of HIV coinfection ^{46,47}
HBV coinfection ⁴⁸
Symptomatic (icteric) acute hepatitis ^{49,50}
Interleukin-28B rs12979860 CC and rs8099917 TT alleles ^{44,44,51-55}
HLA class II DQB1*02, DQB1*03, DRB1*04, and DRB1*11 alleles ⁵⁶
Vigorous HCV-specific T-cell response ^{50,57-59}
Viral factors
HCV genotype 1 ^{44,60}
High peak HCV RNA level ^{54,55}
Limited diversity of HCV quasispecies ^{50,61-63}

HCV, hepatitis C virus; HIV, human immunodeficiency virus; HBV, hepatitis B virus; HLA, human leukocyte antigen; RNA, ribonucleic acid.

DIAGNOSIS

Diagnosis of acute HCV infection remains challenging since the gold-standard criteria have not been well-established. Generally, acute HCV infection is defined as the 6-month time period after the initial exposure to HCV. Anti-HCV antibody seroconversion provides direct evidence for HCV contraction, but the window period varies significantly (Table 3). Typically, it takes about 7 to 8 weeks for third-generation enzyme immunoassay (EIA-3) and 9 to 10 weeks for EIA-2 to detect anti-HCV antibodies from the initial HCV exposure.⁷⁹ However, the time to anti-HCV antibody seroconversion may be as long as 12 months or even absent, in PLWH, hemodialysis patients, or patients undergoing organ transplantation.⁸⁰⁻⁸² In these cases, HCV RNA testing become necessary. Moreover, since the anti-HCV antibody assay may have shown positive results at the first visit for most patients, it would be difficult to detect anti-HCV antibody seroconversion if past data about the patients' anti-HCV antibody status

Table 3. Proposed diagnostic criteria for acute HCV infection

Major criteria	Seroconversion of anti-HCV antibody from negative to positive reaction
	Presence of serum HCV RNA from a previously negative reaction of anti-HCV antibody
Ancillary criteria	Markedly-elevated serum ALT levels (>5–20 times of the upper limit of normal)
	Known or suspected exposure to HCV during the past 6 months
	Clinical symptoms of acute illness, including fever, nausea, vomiting, diarrhea, abdominal pain, fatigue, dark urine, and jaundice
	Sudden onset of liver disease
	Excluding other causes of acute liver injuries

HCV, hepatitis C virus; RNA, ribonucleic acid; ALT, alanine aminotransferase.

are not available.

Other clinical features suggesting acute HCV infection include marked elevation of serum ALT levels (>5–20 times of the upper limit of normal), presentation of symptoms mentioned above, and known or possible exposure to HCV during the past six months (Table 3). However, the ALT elevation is not specific, and only 20% of patients are symptomatic. Furthermore, self-reported information of potential source/route of exposure is largely unreliable.³⁷

For patients with HCV reinfection after spontaneous viral clearance or a course of successful antiviral treatment, documentation of anti-HCV seroconversion would be impossible because the anti-HCV antibodies may persist for years following eradication of HCV.⁸³ Although testing for serum HCV RNA level may help assess viral resurgence, it is not common to monitor serum HCV RNA level after the patients have achieved sustained virologic response at off-therapy week 12 (SVR₁₂), unless an abrupt increase in serum ALT levels is present during the follow-up. In addition, discriminating reinfection from late relapse requires identification of different viral strains through phylogenetic mapping, which cannot be performed without serum samples acquired during the prior episodes of infection and sufficient laboratory techniques.

Diagnosing HCV superinfection in individuals with chronic HCV infection is even more challenging than HCV reinfection due to the need of extensive longitudinal sequence analysis to show the dynamic evolution of viral strains over time. As it is difficult to confirm HCV superinfection, the true incidence and prevalence of HCV superinfection in chronic HCV infection, as well as the clinical consequences, remain largely unknown.

TREATMENT WITH IFN-FREE DAAs

The SVR₂₄ rates of pegylated IFN (PEG-INF) with or without ribavirin (RBV) for 8 to 24 weeks in acute HCV infection range from 60% to 93%.^{84–90} Although the antiviral response rates are fair with IFN-based therapies in patients with acute HCV infection, IFN is obsolete after the introduction of DAAs owing to the concerns about treatment-emergent adverse events (AEs) and the longer treatment duration.

Clinical trials

Sofosbuvir (SOF) plus RBV is the first US food and drug administration (FDA)-approved IFN-free DAA regimen for chronic HCV infection. The DARE-C II study treated 19 patients with acute HCV infection, among which 84% and 74% were PLWH and PWID, with SOF plus RBV for six weeks. However, the SVR₁₂ rates by intention-to-treat (ITT) and per-protocol (PP) analyses were 32% and 40%.⁹¹ The SWIFT-C Cohort-1 study tried to extend the treatment duration of SOF plus RBV to 12 weeks in 17 PLWH with acute HCV infection. Although the SVR₁₂ rate increased from 32% to 59%, SOF plus RBV was not recommended for acute HCV infection because the response rate was far from ideal (Table 4).⁹²

The HepNet Acute HCV IV trial recruited 20 low-risk general patients with acute HCV GT1 or GT4 infection who received sofosbuvir/ledipasvir (SOF/LDV) for six weeks. All patients tolerated the treatment well, and the SVR₁₂ rate was 100%.⁹³ Based on the promising result, Rockstroh et al.⁹⁴ tested this regimen on PLWH with the same condition as mentioned above. However, the SVR₁₂ rates by ITT and PP analyses were 77% and 88%. Patients with a baseline viral load >6.96 log₁₀ IU/mL tended to have a higher risk of relapse with six weeks

Table 4. Summary of IFN-free direct-acting antivirals for acute HCV infection

Study (year)	Regimen	Duration (week)	Patient, n	PLWH (%)	PWID (%)	Hemodialysis (%)	HCV GT	SVR ₁₂ (%) (ITT)	SVR ₁₂ (%) (PP)
Clinical trial									
DARE-C II (2016) ⁹¹	SOF+RBV	6	19	84	74	0	1a (68%) 2b (5%) 3a (21%) 3 (5%)	32	40
SWIFT-C Cohort-1 (2017) ⁹²	SOF+RBV	12	17	100	24	0	1a (65%) 1b (12%) 1 (12%) 2b (6%)	59	59
HepNet Acute HCV IV (2017) ⁹³	SOF/LDV	6	20	0	0	0	1a (55%) 1b (45%)	100	100
Rockstroh et al. (2017) ⁹⁴	SOF/LDV	6	26	100	NA	0	1a (73%) 4a/c/d (27%)	77	88
SWIFT-C Cohort-2 (2017) ⁹⁵	SOF/LDV	8	27	100	19	0	1a (85%) 1b (14%) 4 (4%)	100	100
TARGET3D (2018) ⁹⁶	PrOD±RBV*	8	30	77	53	0	1a (93%) 1b (3%) 1 (3%)	97	100
DAH52 (2019) ⁹⁷	EBR/GZR	8	80	91	NA	0	1a (64%) 4 (36%)	94	99
Martinello et al. (2020) ¹⁰⁰	GLE/PIB	6	30	77	47	0	1a (73%) 1b (3%) 1 (7%) 3a (7%) 4d (10%)	90	96
REACT (2021) ¹⁰¹	SOF/VEL	6	93	70	19	0	1a (62.4%) 1b (4.3%) 1 (1.1%) 3 (16.1%) 4 (16.1%) 1a (60.0%) 1b (2.1%) 2 (4.0%) 3 (17.9%) 4 (15.8%)	82	89
		12	95	68	22	0		91	98

Table 4. Continued

Study (year)	Regimen	Duration (week)	Patient, n	PLWH (%)	PWID (%)	Hemodialysis (%)	HCV GT	SVR ₁₂ (%) (ITT)	SVR ₁₂ (%) (PP)
HepNet Acute HCV V (2022) ¹⁰²	SOF/VEL	8	20	NA	NA	0	1a (60%) 1b (5%) 2 (5%) 3 (15%) 4 (15%)	90	100
Real-world study									
Chromy et al. (2019) ¹⁰⁹	SOF/LDV SOF/VEL PrOD	12	38	100	0	0	1a (66%) 1b (11%) 2 (3%) 3 (13%) 4 (8%)	100	100
Hussein et al. (2019) ¹¹⁰	SOF+DCV [†] SOF/LDV	12	17 2	0 0	0 0	100 100	1a (100%) 1a (100%)	100 100	100 100
He et al. (2018) ¹¹¹	SOF+DCV [†]	24	33	0	0	100	1b (21%) 2a (72%) 1b+2a (6%)	100	100
Goel et al. (2021) ¹¹²	SOF+DCV [†]	8	27	0	0	100	1 (22%) 3 (37%) 4 (7%) Not tested (33%)	93	96
Ji et al. (2022) ¹¹³	EBR/GZR	12	68	0	0	100	1b (100%)	100	100

IFN, interferon; HCV, hepatitis C virus; PLWH, people living with human immunodeficiency virus; PWID, people who inject drugs; GT, genotype; SVR₁₂, sustained virologic response at off-therapy week 12; ITT, intention-to-treat; PP, per-protocol; SOF, sofosbuvir; RBV, ribavirin; LDV, ledipasvir; PrOD, paritaprevir/ritonavir/ombitasvir plus dasabuvir; EBR, elbasvir; GZR, grazoprevir; GLE, glecaprevir; PIB, pibrentasvir; VEL, velpatasvir; DCV, daclatasvir.

[†]PrOD for HCV GT1, PrOD plus RBV for HCV GT1a, or unsubtypable GT1.

¹Daily dose of SOF was 200 mg.

of SOF/LDV.⁹⁴ The SWIFT-C Cohort-2 study extended the treatment duration of SOF/LDV to eight weeks in 27 PLWH, and they all achieved SVR₁₂.⁹⁵ Based on these trial results, SOF/LDV for six weeks is sufficient for non-HIV-positive patients with acute HCV GT1 or GT4 infection, while the treatment duration should be extended to 8 weeks for HIV-positive patients to secure a satisfactory SVR₁₂ rate (Table 4).

In the TARGET3D trial, 30 patients with acute HCV GT1 infection received paritaprevir/ritonavir/ombitasvir plus dasabuvir (PrOD) for eight weeks. RBV was added to PrOD in patients with HCV GT1a or unsubtypeable GT1 infection. Seventy-seven percent and 53% of them were PLWH and PWID. All patients, except for one who discontinued treatment early, achieved SVR₁₂.⁹⁶ Eighty patients with acute HCV GT1 or GT4 infection who received elbasvir/grazoprevir (EBR/GZR) for eight weeks were recruited in the DAHHS2 trial. Most patients (91%) were PLWH. The SVR₁₂ by ITT and PP analyses were 94% and 99%, respectively (Table 4).⁹⁷

Current international guidelines recommend pangenotypic DAA regimens, including glecaprevir/pibrentasvir (GLE/PIB) and sofosbuvir/velpatasvir (SOF/VEL), as the first-line treatment for chronic HCV infection.^{98,99} Treating patients with acute HCV infection with pangenotypic DAAs may be more beneficial than using genotype-specific DAAs because (1) HCV genotyping may not be reliable or possible when the HCV viremia is low during the acute phase of infection; (2) HCV genotyping may not be accessible or available in resource-limited regions; (3) By skipping HCV genotyping, patients may start to receive treatment earlier with the use of pangenotypic DAAs. Martinello et al.¹⁰⁰ conducted a multicenter, international trial to treat 30 patients with recent HCV infection with GLE/PIB for six weeks. Among these patients, 77% and 47% were PLWH and PWID. The SVR₁₂ rate was 90% by ITT analysis. Among the three patients who failed to achieve SVR₁₂, only one had confirmed virologic failure, and the SVR₁₂ by PP analysis was 96%.¹⁰⁰ The REACT trial is a multicenter, international, non-inferior, randomized trial to compare the efficacy of SOF/VEL for 6 or 12 weeks in patients with recent HCV infection.¹⁰¹ About 68% to 79% of patients were PLWH, and 19% to 22% were PWID. By ITT analysis, the SVR₁₂ rates in the 6-week and 12-week arms were 82% and 91% (95% confidence interval: -18.6% to 1.0%), demonstrating a low response rate in the truncated arm by failure to meet the -12% non-inferior confidence bound. The SVR₁₂ rates in the 6-week and 12-week arms were 89% and 98% by

PP analysis. The HepNet Acute HCV V trial, which recruited 20 patients on HIV PrEP or opioid agonist therapy (OAT), extended the treatment duration of SOF/VEL to 8 weeks for acute HCV infection. Eighteen of 20 (90%) patients achieved SVR₁₂, and the remaining two were lost to follow-up (Table 4).¹⁰²

Real-world studies

Limited real-world studies have demonstrated the effectiveness of DAAs for low-risk general patients with acute HCV infection. Furthermore, most of those studies were case reports or case series, although the SVR₁₂ was achieved in all patients.¹⁰³⁻¹⁰⁸ Chromy et al.¹⁰⁹ retrospectively assessed 38 PLWH with acute HCV infection treated with various DAA regimens. All cases received 12 weeks of treatment and achieved SVR₁₂. Four cohort studies have reported that the SVR₁₂ rates in hemodialysis patients with acute HCV infection receiving SOF plus daclatasvir (DCV) for 8–24 weeks, SOF/LDV for 12 weeks, or EBR/GZR for 12 weeks ranged from 93% to 100% (Table 4).¹¹⁰⁻¹¹³

Treatment of HCV reinfection

In low-risk general patients with HCV who achieve SVR₁₂ with antiviral treatment, the risk of HCV reinfection ranges from 0.14 to 0.185 per 100 PYs.¹¹⁴⁻¹¹⁶ In PLWH, the risk of HCV reinfection was much higher (3.76 per 100 PYs) according to a recent meta-analysis. PLWH MSM and those with recent drug use had an incidence of reinfection of 6.01 and 5.49 per 100 PYs, respectively.¹¹⁷ In PWID, the risk of HCV reinfection in those with recent drug use, IDU, and those receiving OAT were 5.9, 6.2, and 3.8 per 100 PYs, respectively.¹¹⁸ While the global incidence of new HCV infection in hemodialysis patients was 1.2% in 2015, a recent survey in Taiwan indicated that the risk of HCV reinfection in hemodialysis patients was 0.23 per 100 PYs following the implementation of mass screening, treatment scale-up, and universal precaution.¹¹⁵ A national survey in Australia has demonstrated an increasing number of HCV reinfection from 2016 to 2020 in patients who had achieved SVR₁₂ with DAAs, which entails the importance of early diagnosis and timely treatment for HCV reinfection to reduce viral transmission.¹¹⁹

Data regarding the effectiveness of DAAs in treating HCV reinfection in patients who have achieved SVR₁₂ after a prior course of DAAs are limited. The REACH-C cohort assessed the

effectiveness of treatment with GLE/PIB, SOF/VEL, SOF/LDV, SOF/DCV, EBR/GZR, or sofosbuvir/velpatasvir/voxilaprevir (SOF/VEL/VOX) in 88 Australians with HCV reinfection who achieved SVR₁₂ with a previous course of DAAs. All patients were PLWH, PWID, incarcerated, or on OAT. The SVR₁₂ rate in the 56 patients with available outcomes was 95%, which was comparable to the 95% response rate observed in DAA-naïve patients. Eight patients were treated using the same regimen as their prior treatment, and they all achieved SVR₁₂.¹²⁰ Liu et al.¹²¹ recruited 22 Taiwanese patients diagnosed with HCV reinfection after DAA-induced SVR₁₂. Eighteen (81.8%) of them were PLWH. Twenty patients started retreatment with pangenotypic GLE/PIB or SOF/VEL, and all achieved SVR₁₂. Similar to the REACH-C report, all eleven patients, who were retreated with the same DAA regimen as the prior treatment, achieved SVR₁₂.¹²¹

Treatment of HCV superinfection

Data regarding the use of DAAs in managing chronic HCV-infected patients with HCV superinfection are lacking. Based on the potential genotype switch and the presence of mixed viral strain infection in patients with HCV superinfection, pangenotypic DAA regimens are the preferred treatment choices to secure satisfactory viral eradication.

Treatment of acute HCV infection in patients undergoing organ transplantation

While the rates of solid organ donation and transplantation have increased over the last decade, the demand is still far greater than the supply, necessitating more efficient organ procurement and allocation. HCV-aviremic recipients are conventionally not recommended to receive HCV-viremic organs since HCV transmission occurs immediately after transplantation, and the recipients may develop lethal fibrosing cholestatic hepatitis if the acute HCV infection is left untreated.¹²²⁻¹²⁴ In contrast to IFN, which has poor efficacy and patient tolerance, the excellent effects of DAAs on solid organ recipients with chronic HCV infection has garnered great interest in the application of prophylactic or pre-emptive DAAs to prevent or eradicate HCV transmission in this clinical condition.

Numerous clinical trials and real-world studies have reported the experiences of using DAA treatment, either prophy-

lactically, pre-emptively, or deferred, to manage post-transplant acute HCV infection in HCV-negative patients who have received solid organ transplant from HCV-viremic donors. Although the regimens and treatment durations vary widely across studies, a meta-analysis, which included 852 solid organ recipients from 35 studies, revealed that nearly all patients achieved SVR₁₂. All seven recipients who relapsed after initial DAA treatment achieved SVR₁₂ with a second course of rescue regimens. As HCV was eradicated during the early phase of acute HCV infection, there were no recipient deaths or graft loss.¹²⁵ Table 5 summarizes the representative trials that used DAA treatment to manage acute HCV infection in HCV-negative patients receiving HCV-viremic solid organs.¹²⁶⁻¹⁴² In general, the SVR₁₂ rates were 100% when DAAs were administered for the same duration as indicated for chronic HCV infection. In addition, the SVR₁₂ rate, excluding non-virologic failures, can reach >94% if the solid organ recipients received prophylactic or pre-emptive DAAs for more than one week.

Transplanting HCV-viremic organs to HCV-negative recipients has complex ethical and legal implications because of the intentional transmission of an infectious agent to the recipients. Although this approach provides benefits such as reduced wait time for organ transplant and reduced mortality while on the organ waiting list, these should be weighed against other risks such as HCV transmission from donors to recipients and from recipients to partners, progressive hepatic and extrahepatic diseases in case of DAA failures, and graft loss before making clinical decisions. Most importantly, the local legal regulations must be considered.

Cost-effectiveness of early DAA treatment

Using a willingness-to-pay threshold of USD 100,000 per quality-adjusted life year (QALY), timely DAA treatment for acute HCV infection was cost-effective with an incremental cost-effectiveness ratio (ICER) of USD 19,991 per QALY for patients not at risk of HCV transmission. Furthermore, DAA treatment for acute HCV infection in patients at risk of HCV transmission was cost-saving, with an increase of QALYs by 0.03 and a decrease of costs by USD 3,655.¹⁴³ The cost-effectiveness analysis of prophylactic or pre-emptive DAAs in HCV-negative patients who receive HCV-positive organs remains to be examined.

Table 5. Summary of IFN-free direct-acting antivirals for acute HCV infection after solid organ transplantation from HCV-viremic donors to HCV-aviremic recipients

Study (year)	Regimen	Duration (week)	Strategy	Patient, n	HCV GT	SVR ₁₂ (%) (ITT)	SVR ₁₂ (%) (PP)
Heart							
USHER (2019) ¹²⁶	EBR/GZR	12	Pre-emptive	10	1	90*	100
Bethea et al. (2019) ¹²⁷	GLE/PIB	8	Pre-emptive	25	1-3, indeterminate, NA	100	100
Reyentovich et al. (2020) ²⁸	GLE/PIB	8	Pre-emptive	22	NA	100	100
Smith et al. (2021) ²⁹	GLE/PIB	8	Pre-emptive	22	1-3, indeterminate	100	100
DONATE-HCV (2019) ³⁰	SOF/VEL	4	Pre-emptive	8	1-3, indeterminate	100 [†]	100 [†]
Lung							
Smith et al. (2021) ²⁹	GLE/PIB	8	Prophylactic/pre-emptive	16	1-3, indeterminate	100	100
DONATE-HCV (2019) ³⁰	SOF/VEL	4	Pre-emptive	36	1-3, indeterminate	100 [†]	100 [†]
Cypel et al. (2020) ³¹	SOF/VEL	12	Deferred	20	1-3	90	90
Liver							
Bethea et al. (2020) ³²	GLE/PIB	12	Pre-emptive	9	1-3	100	100
Terrault et al. (2021) ³³	SOF/VEL	12	Pre-emptive	13	NA	100	100
Kwong et al. (2018) ³⁴	SOF/LDV, SOF/DCV, or SOF/VEL±RBV	12-24	Deferred	10	1-3	100	100
Kidney							
THINKER-1 (2017) ³⁵	EBR/GZR	12	Pre-emptive	10	1	100	100
	EBR/GZR+RBV	16			1 (N55A RAS)		
THINKER-2 (2018) ³⁶	EBR/GZR	12	Pre-emptive	10	1	100	100
	EBR/GZR+RBV	16			1 (NA5A RAS)		
EXPANDER (2018) ³⁷	EBR/GZR	12			1a, 1b, 4, indeterminate GT due to low viral load		
	EBR/GZR+RBV	16	Prophylactic	10	1a (NA5A RAS)	100	100
	EBR/GZR+SOF	12			2, 3		
REHANNA (2021) ³⁸	GLE/PIB	4	Prophylactic	10	1, 3	100	100
MYTHIC (2020) ³⁹	GLE/PIB	8	Pre-emptive	30	1, 2, 4	100	100
DAPeR (2021) ⁴⁰	SOF/VEL	2-4 days	Prophylactic	50	1-3	88	88
REFORM HEPC (2021) ⁴¹	SOF/VEL	8 days	Prophylactic	32	NA	97	97
	SOF/VEL+ezetimibe	8 days	Prophylactic	18		94	94

Table 5. Continued

Study (year)	Regimen	Duration (week)	Strategy	Patient, n	HCV GT	SVR ₁₂ (%) (ITT)	SVR ₁₂ (%) (PP)
Terrault et al. (2021) ¹³³	SOF/VEL	12	Pre-emptive	11	NA	100	100
Miscellaneous (heart, lung, kidney, and kidney-pancreas)							
Feld et al. (2020) ¹⁴²	GLE/PIB+ezetimibe	1	Prophylactic	18	1–3	100	100

IFN, interferon; HCV, hepatitis C virus; GT, genotype; SVR₁₂, sustained virologic response at off-therapy week 12; ITT, intention-to-treat; PP, per-protocol; EBR, elbasvir; GZR, grazoprevir; GLE, glecaprevir; PIB, pibrentasvir; NA, not assessed; SOF, sofosbuvir; VEL, velpatasvir; LDV, ledipasvir; DCV, daclatasvir; RBV, ribavirin; NS5A, non-structural protein 5A; RAS, resistance-associated substitution.

*Died on post-transplantation Day 79 due to antibody-mediated rejection and multi-organ failure.

¹One of the 8 patients was pending SVR₁₂ result at the time of publication.

[†]Eight of the 36 patients were pending SVR₁₂ results at the time of publication.

Timing of treatment initiation

There has not been a consensus on the timing of therapeutic intervention for people with acute HCV infection. The possibility of spontaneous viral clearance, the benefits of early treatment initiation (based on the cost-effectiveness of universal DAA treatment), and the risk of HCV transmission as well as potential morbidity/mortality if no treatment is provided, should be considered before treatment initiation for patients with acute HCV infection.

According to the American Association for the Study of Liver Diseases/Infectious Disease Society of America (AASLD/IDSA) and European Association for the Study of the Liver (EASL) guidelines, DAA treatment should be given once acute viremic HCV infection is confirmed.^{98,99,144} While the AASLD guidelines suggest treating acute HCV infection with the same DAA regimens as recommended for chronic HCV infection, the EASL guidelines suggest using GLE/PIB or SOF/VEL for eight weeks to manage acute HCV infection. Although PrEP can significantly reduce HIV transmission, PrEP or post-exposure prophylaxis (PEP) with DAAs is not recommended in this clinical condition due to the lack of evidence.^{99,144,145}

The timely diagnosis and early intervention for acute HCV infection are relevant to HCV-negative patients receiving HCV-viremic organs. Pangenotypic DAAs, such as GLE/PIB or SOF/VEL for 12 weeks, should be initiated within two weeks of transplantation if the recipients are clinically stable. Prophylactic or pre-emptive treatment with pangenotypic DAAs is also recommended in this patient group.^{99,144} However, the ideal pangenotypic regimens remain debatable, as AASLD/IDSA and EASL guidelines have proposed different regimens.

CHALLENGES AND FUTURE PERSPECTIVES

Since the gold-standard diagnostic criteria for acute HCV infection has not been well-established, to detect *de novo* infection, reinfection, or superinfection of HCV promptly, clinicians should stay alert to this potential diagnosis, particularly when encountering patients with markedly elevated ALT levels and/or risk factors of HCV transmission. While the guidelines recommend initiating DAA treatment over waiting for spontaneous viral clearance in managing acute HCV infection, it is important to note the DAA regimens are not yet licensed to treat acute HCV infection. Furthermore, the dis-

crepant pricing and payment policies across countries may hinder the universal and early access to DAA treatment. In addition to early identification and timely therapeutic intervention, public health strategies, such as universal precaution, harm reduction, safe sex, and post-SVR₁₂ viral surveillance, should also be implemented to efficiently reduce the risk of HCV transmission.¹⁴⁶

With the high HCV infection rates observed in endemic areas and in high-risk PWID and PLWH, development of prophylactic vaccines against HCV should be prioritized to combat HCV transmission. Despite years of vigorous work, only two candidate vaccines have advanced to human studies. The first one consists of the recombinant full-length E1/E2 glycoprotein of HCV GT1a with an oil-in-water adjuvant (MF59C.1).¹⁴⁷ This vaccine did not enter further efficiency studies due to failure in generating antibodies that neutralized heterologous HCV pseudo-particles in study volunteers. The second vaccine consists of a chimpanzee adenovirus vector (ChAd3) as prime and a modified vaccinia Ankara (MVA) vector as boost, both of which encode the HCV GT1b non-structural proteins (NS). This vaccine was tested in phase 1–2 randomized, double-blind, placebo-controlled trial in PWID. Although it generated strong HCV-specific T-cell responses and lowered the peak HCV RNA level, it cannot prevent chronic HCV infection.¹⁴⁸ Recently, a bivalent pangentypic prophylactic vaccine, which consists of a chimpanzee adenovirus vector (ChAdOx1) encoding conserved sequences across HCV GT1-6 and a modified HCV glycoprotein E2 with deletions of hypervariable regions (HVR) 1 and 2, has successfully induced both neutralizing antibody and CD4+ and CD8+ T cell response in mice experiments.¹⁴⁹ Further translational research is needed to confirm this new candidate's clinical effectiveness.

CONCLUSION

Acute HCV infection remains a threat to global health, despite a decrease of incidence over the past decades. Transmitted through parenteral routes, new cases of HCV infection comprise mainly of people receiving unsafe medical procedures, PWID, and PLWH.

Diagnosing acute HCV infection can be challenging, particularly in immunocompromised, reinfected, and superinfected patients, since it is difficult to detect anti-HCV antibody

seroconversion and confirm the presence of HCV RNA from a previously negative antibody response. Therefore, both comprehensive history taking and prudent laboratory assessment are needed in order to more accurately diagnose new cases.

To date, numerous clinical trials and real-world cohort studies have supported the role of DAAs in managing acute HCV infection. Furthermore, early initiation of DAA treatment was found to be cost-saving and cost-effective in patients with acute HCV infection, regardless of the risk of HCV transmission. Although the efficacy of DAAs in treating acute *de novo* HCV infection and HCV reinfection is excellent, there is not yet a consensus on the optimal DAA regimens and the duration of treatment. In HCV-negative recipients who have received HCV-viremic organs, the transmission of HCV from donors to recipients typically happens within a few days of transplantation. With a well-defined timing and route of HCV transmission, the administration of prophylactic or pre-emptive DAAs during the peri-transplant period almost always eradicate HCV in the organ recipients, although the optimized DAA regimens and treatment duration remain to be determined. Overall, treatment of acute HCV infection with DAAs can improve patient outcomes and reduce HCV transmission. Prophylactic vaccines against HCV are not yet available, and a pangentypic HCV vaccine is under active development. Patient education and public health practices, such as universal precaution, harm reduction, safe sex, and vigilant post-SVR₁₂ surveillance, may help achieve the WHO goal of HCV eradication by 2030.

Authors' contributions

Conception and design: CH Liu, JH Kao; Drafting of the article: CH Liu, JH Kao; Critical revision of the article for important intellectual content: CH Liu, JH Kao; Final approval of the article: CH Liu, JH Kao; Administrative, technical, or logistic support: JH Kao; Collection and assembly of data: CH Liu.

Acknowledgements

The authors thank Hui-Ju Lin and Pin-Chin Huang for managing the administrative affairs; the 7th Core Lab of the National Taiwan University Hospital, and the 1st Common Laboratory of the National Taiwan University Hospital, Yun-Lin Branch, for the technical support; Jo-Hsuan Wu for polishing the manuscript writing.

Conflicts of Interest

Chen-Hua Liu: advisory board for Abbvie, Gilead Sciences, Merck Sharp & Dohme; speaker's bureau for Abbott, Abbvie, Gilead Sciences, Merck Sharp & Dohme; research grant from Abbvie, Gilead Science, Merck Sharp & Dohme. Jia- Horng Kao: advisory board for Abbott, Abbvie, Bristol-Myers Squibb, Gilead Sciences, GlaxoSmithKline, Merck Sharp & Dohme, Novartis, Roche; speaker's bureau for Abbott, Abbvie, Bayer, Bristol-Myers Squibb, Gilead Sciences, GlaxoSmithKline, Merck Sharp & Dohme, Novartis, Roche.

REFERENCES

1. Geneva: World Health Organization (WHO). Guidelines for the care and treatment of persons diagnosed with chronic hepatitis C virus infection. 2018. WHO web site, <<https://www.who.int/publications/i/item/9789241550345>>. Accessed 27 Jan 2023.
2. Polaris Observatory HCV Collaborators. Global change in hepatitis C virus prevalence and cascade of care between 2015 and 2020: a modelling study. *Lancet Gastroenterol Hepatol* 2022;7:396-415.
3. Lauer GM, Walker BD. Hepatitis C virus infection. *N Engl J Med* 2001;345:41-52.
4. Liu CH, Kao JH. Nanomedicines in the treatment of hepatitis C virus infection in Asian patients: optimizing use of peginterferon alfa. *Int J Nanomedicine* 2014;9:2051-2067.
5. Razavi H, Waked I, Sarrazin C, Myers RP, Idilman R, Calinas F, et al. The present and future disease burden of hepatitis C virus (HCV) infection with today's treatment paradigm. *J Viral Hepat* 2014;21 Suppl 1:34-59.
6. Thrift AP, El-Serag HB, Kanwal F. Global epidemiology and burden of HCV infection and HCV-related disease. *Nat Rev Gastroenterol Hepatol* 2017;14:122-132.
7. Geneva: World Health Organization (WHO). Global progress report on HIV, viral hepatitis and sexually transmitted infections, 2021. WHO web site, <<https://www.who.int/publications/i/item/9789240027077>>. Accessed 27 Jan 2023.
8. Geneva: World Health Organization (WHO). Global hepatitis report 2017. WHO web site, <<https://www.who.int/publications/i/item/9789241565455>>. Accessed 27 Jan 2023.
9. Pépin J, Abou Chakra CN, Pépin E, Nault V, Valiquette L. Evolution of the global burden of viral infections from unsafe medical injections, 2000-2010. *PLoS One* 2014;9:e99677.
10. Pépin J, Abou Chakra CN, Pépin E, Nault V. Evolution of the global use of unsafe medical injections, 2000-2010. *PLoS One* 2013;8:e80948.
11. Hauri AM, Armstrong GL, Hutin YJ. The global burden of disease attributable to contaminated injections given in health care settings. *Int J STD AIDS* 2004;15:7-16.
12. Gore C, Lazarus JV, Peck RJJ, Sperle I, Safreed-Harmon K. Unnecessary injecting of medicines is still a major public health challenge globally. *Trop Med Int Health* 2013;18:1157-1159.
13. Mele A, Tosti ME, Marzolini A, Moiraghi A, Ragni P, Gallo G, et al. Prevention of hepatitis C in Italy: lessons from surveillance of type-specific acute viral hepatitis. SEIEVA collaborating Group. *J Viral Hepat* 2000;7:30-35.
14. Paez Jimenez A, Sharaf Eldin N, Rimlinger F, El-Daly M, El-Hariri H, El-Hoseiny M, et al. HCV iatrogenic and intrafamilial transmission in Greater Cairo, Egypt. *Gut* 2010;59:1554-1560.
15. El Gaafary MM, Rekecewicz C, Abdel-Rahman AG, Allam MF, El Hosseiny M, Hamid MA, et al. Surveillance of acute hepatitis C in Cairo, Egypt. *J Med Virol* 2005;76:520-525.
16. Jadoul M, Bieber BA, Martin P, Akiba T, Nwankwo C, Arduino JM, et al. Prevalence, incidence, and risk factors for hepatitis C virus infection in hemodialysis patients. *Kidney Int* 2019;95:939-947.
17. Liu CH, Kao JH. Pan-genotypic direct-acting antivirals for patients with hepatitis C virus infection and chronic kidney disease stage 4 or 5. *Hepatol Int* 2022;16:1001-1019.
18. Fabrizi F, Messa P. Transmission of hepatitis C virus in dialysis units: a systematic review of reports on outbreaks. *Int J Artif Organs* 2015;38:471-480.
19. Degenhardt L, Charlson F, Stanaway J, Larney S, Alexander LT, Hickman M, et al. Estimating the burden of disease attributable to injecting drug use as a risk factor for HIV, hepatitis C, and hepatitis B: findings from the Global Burden of Disease Study 2013. *Lancet Infect Dis* 2016;16:1385-1398.
20. Hajarizadeh B, Grebely J, Dore GJ. Epidemiology and natural history of HCV infection. *Nat Rev Gastroenterol Hepatol* 2013;10:553-562.
21. Martinello M, Matthews GV. Enhancing the detection and management of acute hepatitis C virus infection. *Int J Drug Policy* 2015;26:899-910.
22. Maher L, Jalaludin B, Chant KG, Jayasuriya R, Sladden T, Kaldor JM, et al. Incidence and risk factors for hepatitis C seroconversion in injecting drug users in Australia. *Addiction* 2006;101:1499-1508.
23. Hagan H, Pouget ER, Des Jarlais DC, Lelutiu-Weinberger C.

- Meta-regression of hepatitis C virus infection in relation to time since onset of illicit drug injection: the influence of time and place. *Am J Epidemiol* 2008;168:1099-1109.
24. Esmaeili A, Mirzazadeh A, Carter GM, Esmaeili A, Hajarizadeh B, Sacks HS, et al. Higher incidence of HCV in females compared to males who inject drugs: A systematic review and meta-analysis. *J Viral Hepat* 2017;24:117-127.
 25. Morris MD, Shiboski S, Bruneau J, Hahn JA, Hellard M, Prins M, et al.; International Collaboration of Incident HIV and HCV in Injecting Cohorts (InC3). Geographic differences in temporal incidence trends of hepatitis C virus infection among people who inject drugs: The InC3 Collaboration. *Clin Infect Dis* 2017;64:860-869. Erratum in: *Clin Infect Dis* 2017;65:706-707.
 26. Palmateer NE, Taylor A, Goldberg DJ, Munro A, Aitken C, Shepherd SJ, et al. Rapid decline in HCV incidence among people who inject drugs associated with national scale-up in coverage of a combination of harm reduction interventions. *PLoS One* 2014;9:e104515.
 27. van den Berg CH, Smit C, Bakker M, Geskus RB, Berkhout B, Jurliaans S, et al. Major decline of hepatitis C virus incidence rate over two decades in a cohort of drug users. *Eur J Epidemiol* 2007;22:183-193.
 28. Grebely J, Lima VD, Marshall BD, Milloy MJ, DeBeck K, Montaner J, et al. Declining incidence of hepatitis C virus infection among people who inject drugs in a Canadian setting, 1996-2012. *PLoS One* 2014;9:e97726.
 29. Centers for Disease Control and Prevention (CDC), the United States. Hepatitis C Surveillance 2020. CDC web site, <<https://www.cdc.gov/hepatitis/statistics/2020surveillance/hepatitis-c.htm>>. Accessed 27 Jan 2023.
 30. Pouget ER, Hagan H, Des Jarlais DC. Meta-analysis of hepatitis C seroconversion in relation to shared syringes and drug preparation equipment. *Addiction* 2012;107:1057-1065.
 31. van Santen DK, van der Helm JJ, Del Amo J, Meyer L, D'Arminio Monforte A, Price M, et al.; CASCADE Collaboration in EuroCoord. Lack of decline in hepatitis C virus incidence among HIV-positive men who have sex with men during 1990-2014. *J Hepatol* 2017;67:255-262.
 32. Jin F, Dore GJ, Matthews G, Luhmann N, Macdonald V, Bajis S, et al. Prevalence and incidence of hepatitis C virus infection in men who have sex with men: a systematic review and meta-analysis. *Lancet Gastroenterol Hepatol* 2021;6:39-56.
 33. Hagan H, Jordan AE, Neurer J, Cleland CM. Incidence of sexually transmitted hepatitis C virus infection in HIV-positive men who have sex with men. *AIDS* 2015;29:2335-2345.
 34. Hasse B, Ledergerber B, Hirschel B, Vernazza P, Glass TR, Jeanin A, et al.; Swiss HIV Cohort Study. Frequency and determinants of unprotected sex among HIV-infected persons: the Swiss HIV cohort study. *Clin Infect Dis* 2010;51:1314-1322.
 35. Apers L, Vanden Berghe W, De Wit S, Kabeya K, Callens S, Buyze J, et al. Risk factors for HCV acquisition among HIV-positive MSM in Belgium. *J Acquir Immune Defic Syndr* 2015;68:585-593.
 36. Compton WM, Jones CM. Substance use among men who have sex with men. *N Engl J Med* 2021;385:352-356.
 37. Santantonio T, Wiegand J, Gerlach JT. Acute hepatitis C: current status and remaining challenges. *J Hepatol* 2008;49:625-633.
 38. Kanda T, Yokosuka O, Imazeki F, Saisho H. Acute hepatitis C virus infection, 1986-2001: a rare cause of fulminant hepatitis in Chiba, Japan. *Hepatogastroenterology* 2004;51:556-558.
 39. Takano S, Omata M, Ohto M, Satomura Y. Prospective assessment of incidence of fulminant hepatitis in post-transfusion hepatitis: a study of 504 cases. *Dig Dis Sci* 1994;39:28-32.
 40. Kato T, Date T, Miyamoto M, Furusaka A, Tokushige K, Mizokami M, et al. Efficient replication of the genotype 2a hepatitis C virus subgenomic replicon. *Gastroenterology* 2003;125:1808-1817.
 41. Wakita T, Pietschmann T, Kato T, Date T, Miyamoto M, Zhao Z, et al. Production of infectious hepatitis C virus in tissue culture from a cloned viral genome. *Nat Med* 2005;11:791-796.
 42. Kanda T, Matsumoto N, Ishii T, Arima S, Shibuya S, Honda M, et al. Chronic hepatitis C: Acute exacerbation and alanine aminotransferase flare. *Viruses* 2023;15:183.
 43. Micallef JM, Kaldor JM, Dore GJ. Spontaneous viral clearance following acute hepatitis C infection: a systematic review of longitudinal studies. *J Viral Hepat* 2006;13:34-41.
 44. Grebely J, Page K, Sacks-Davis R, van der Loeff MS, Rice TM, Bruneau J, et al.; InC3 Study Group. The effects of female sex, viral genotype, and IL28B genotype on spontaneous clearance of acute hepatitis C virus infection. *Hepatology* 2014;59:109-120.
 45. van den Berg CH, Grady BP, Schinkel J, van de Laar T, Molenkamp R, van Houdt R, et al. Female sex and IL28B, a synergism for spontaneous viral clearance in hepatitis C virus (HCV) seroconverters from a community-based cohort. *PLoS One* 2011;6:e27555.
 46. Thomas DL, Astemborski J, Rai RM, Anania FA, Schaeffer M, Galai N, et al. The natural history of hepatitis C virus infection: host, viral, and environmental factors. *JAMA* 2000;284:450-

- 456.
47. Monin MB, Ingiliz P, Lutz T, Scholten S, Cordes C, Martínez-Rebollar M, et al.; PROBE-C study group. Low spontaneous clearance rates of recently acquired hepatitis C virus in human immunodeficiency virus-positive men who have sex with men (PROBE-C study). *Clin Infect Dis* 2023;76:e607-e612.
 48. Soriano V, Mocroft A, Rockstroh J, Ledergerber B, Knysz B, Chaplinskas S, et al.; EuroSIDA Study Group. Spontaneous viral clearance, viral load, and genotype distribution of hepatitis C virus (HCV) in HIV-infected patients with anti-HCV antibodies in Europe. *J Infect Dis* 2008;198:1337-1344.
 49. Tillmann HL, Thompson AJ, Patel K, Wiese M, Tenckhoff H, Nischalke HD, et al.; German Anti-D Study Group. A polymorphism near IL28B is associated with spontaneous clearance of acute hepatitis C virus and jaundice. *Gastroenterology* 2010;139:1586-1592.e1.
 50. Thomson EC, Fleming VM, Main J, Klenerman P, Weber J, Eliahoo J, et al. Predicting spontaneous clearance of acute hepatitis C virus in a large cohort of HIV-1-infected men. *Gut* 2011;60:837-845.
 51. Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, et al. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature* 2009;461:798-801.
 52. Yang M, Rao HY, Feng B, Zhang W, Wei L. Impact of interleukin 28B polymorphisms on spontaneous clearance of hepatitis C virus infection: a meta-analysis. *J Gastroenterol Hepatol* 2013;28:1114-1121.
 53. Miri HH, Fazeli P, Ali-Hassanzadeh M, Bemani P, Kabelitz D, Kalandar K. Correlation between IL-28 polymorphism and spontaneous clearance in HCV patients: systematic review and meta-analysis. *Arch Virol* 2021;166:2469-2478.
 54. Liu L, Fisher BE, Thomas DL, Cox AL, Ray SC. Spontaneous clearance of primary acute hepatitis C virus infection correlated with high initial viral RNA level and rapid HVR1 evolution. *Hepatology* 2012;55:1684-1691.
 55. Hajarizadeh B, Grady B, Page K, Kim AY, McGovern BH, Cox AL, et al.; InC3 Study Group. Patterns of hepatitis C virus RNA levels during acute infection: the InC3 study. *PLoS One* 2015;10:e0122232.
 56. Gauthiez E, Habfast-Robertson I, Rüeger S, Kutalik Z, Aubert V, Berg T, et al.; Swiss Hepatitis C Cohort Study. A systematic review and meta-analysis of HCV clearance. *Liver Int* 2017;37:1431-1445.
 57. Thimme R, Oldach D, Chang KM, Steiger C, Ray SC, Chisari FV. Determinants of viral clearance and persistence during acute hepatitis C virus infection. *J Exp Med* 2001;194:1395-1406.
 58. Kuntzen T, Timm J, Berical A, Lewis-Ximenez LL, Jones A, Nolan B, et al. Viral sequence evolution in acute hepatitis C virus infection. *J Virol* 2007;81:11658-11668.
 59. Schulze Zur Wiesch J, Ciuffreda D, Lewis-Ximenez L, Kasprovicz V, Nolan BE, Streeck H, et al. Broadly directed virus-specific CD4+ T cell responses are primed during acute hepatitis C infection, but rapidly disappear from human blood with viral persistence. *J Exp Med* 2012;209:61-75.
 60. Harris HE, Eldridge KP, Harbour S, Alexander G, Teo CG, Ramsay ME; HCV National Register Steering Group. Does the clinical outcome of hepatitis C infection vary with the infecting hepatitis C virus type? *J Viral Hepat* 2007;14:213-220.
 61. Farci P, Shimoda A, Coiana A, Diaz G, Peddis G, Melpolder JC, et al. The outcome of acute hepatitis C predicted by the evolution of the viral quasispecies. *Science* 2000;288:339-344.
 62. Liu CH, Chen BF, Chen SC, Lai MY, Kao JH, Chen DS. Selective transmission of hepatitis C virus quasi species through a needlestick accident in acute resolving hepatitis. *Clin Infect Dis* 2006;42:1254-1259.
 63. Bull RA, Luciani F, McElroy K, Gaudieri S, Pham ST, Chopra A, et al. Sequential bottlenecks drive viral evolution in early acute hepatitis C virus infection. *PLoS Pathog* 2011;7:e1002243.
 64. Martinello M, Hajarizadeh B, Grebely J, Dore GJ, Matthews GV. Management of acute HCV infection in the era of direct-acting antiviral therapy. *Nat Rev Gastroenterol Hepatol* 2018;15:412-424.
 65. Layden TJ, Mika B, Wiley TE. Hepatitis C kinetics: mathematical modeling of viral response to therapy. *Semin Liver Dis* 2000;20:173-183.
 66. Santantonio T, Medda E, Ferrari C, Fabris P, Cariti G, Massari M, et al. Risk factors and outcome among a large patient cohort with community-acquired acute hepatitis C in Italy. *Clin Infect Dis* 2006;43:1154-1159.
 67. Hajarizadeh B, Grebely J, Applegate T, Matthews GV, Amin J, Petoumenos K, et al.; ATAC study group. Dynamics of HCV RNA levels during acute hepatitis C virus infection. *J Med Virol* 2014;86:1722-1729.
 68. Vogel M, Boesecke C, Rockstroh JK. Acute hepatitis C infection in HIV-positive patients. *Curr Opin Infect Dis* 2011;24:1-6.
 69. Blackard JT, Sherman KE. Hepatitis C virus coinfection and superinfection. *J Infect Dis* 2007;195:519-524.
 70. Okamoto H, Mishiro S, Tokita H, Tsuda F, Miyakawa Y, Mayumi M. Superinfection of chimpanzees carrying hepatitis C virus of genotype II/1b with that of genotype III/2a or I/1a. *Hepatology*

- 1994;20:1131-1136.
71. Herring BL, Page-Shafer K, Tobler LH, Delwart EL. Frequent hepatitis C virus superinfection in injection drug users. *J Infect Dis* 2004;190:1396-1403
 72. Widell A, Månsson S, Persson NH, Thysell H, Hermodsson S, Blohme I. Hepatitis C superinfection in hepatitis C virus (HCV)-infected patients transplanted with an HCV-infected kidney. *Transplantation* 1995;60:642-647.
 73. Lai ME, Mazzoleni AP, Argiolo F, De Virgili S, Balestrieri A, Purcell RH, et al. Hepatitis C virus in multiple episodes of acute hepatitis in polytransfused thalassaemic children. *Lancet* 1994;343:388-390.
 74. Kao JH, Chen PJ, Lai MY, Chen DS. Superinfection of heterologous hepatitis C virus in a patient with chronic type C hepatitis. *Gastroenterology* 1993;105:583-587.
 75. Kao JH, Chen PJ, Wang JT, Yang PM, Lai MY, Wang TH, et al. Superinfection by homotypic virus in hepatitis C virus carriers: studies on patients with post-transfusion hepatitis. *J Med Virol* 1996;50:303-308.
 76. Kao JH, Chen PJ, Lai MY, Yang PM, Sheu JC, Wang TH, et al. Mixed infections of hepatitis C virus as a factor in acute exacerbations of chronic type C hepatitis. *J Infect Dis* 1994;170:1128-1133.
 77. Liaw YF. Hepatitis C virus superinfection in patients with chronic hepatitis B virus infection. *J Gastroenterol* 2002;37 Suppl 13:65-68.
 78. Liaw YF, Chen YC, Sheen IS, Chien RN, Yeh CT, Chu CM. Impact of acute hepatitis C virus superinfection in patients with chronic hepatitis B virus infection. *Gastroenterology* 2004;126:1024-1029.
 79. Carithers RL Jr, Marquardt A, Gretch DR. Diagnostic testing for hepatitis C. *Semin Liver Dis* 2000;20:159-171.
 80. Schroeter M, Zoellner B, Polywka S, Laufs R, Feucht HH. Prolonged time until seroconversion among hemodialysis patients: the need for HCV PCR. *Intervirology* 2005;48:213-215.
 81. Thomson EC, Nastouli E, Main J, Karayiannis P, Eliahoo J, Muir D, et al. Delayed anti-HCV antibody response in HIV-positive men acutely infected with HCV. *AIDS* 2009;23:89-93.
 82. Preiksaitis JK, Cockfield SM, Fenton JM, Burton NI, Chui LW. Serologic responses to hepatitis C virus in solid organ transplant recipients. *Transplantation* 1997;64:1775-1780.
 83. Toyoda H, Kumada T, Kiriya S, Sone Y, Tanikawa M, Hisanaga Y, et al. Changes in hepatitis C virus (HCV) antibody status in patients with chronic hepatitis C after eradication of HCV infection by interferon therapy. *Clin Infect Dis* 2005;40:e49-e54.
 84. Santantonio T, Fasano M, Sagnelli E, Tundo P, Babudieri S, Fabris P, et al.; Acute Hepatitis C Study Group. Acute hepatitis C: a 24-week course of pegylated interferon α -2b versus a 12-week course of pegylated interferon α -2b alone or with ribavirin. *Hepatology* 2014;59:2101-2109.
 85. Martinello M, Hellard M, Shaw D, Petoumenos K, Applegate T, Grebely J, et al. Short duration response-guided treatment is effective for most individuals with recent hepatitis C infection: the ATAC II and DARE-C I studies. *Antivir Ther* 2016;21:425-434.
 86. Corey KE, Mendez-Navarro J, Gorospe EC, Zheng H, Chung RT. Early treatment improves outcomes in acute hepatitis C virus infection: a meta-analysis. *J Viral Hepat* 2010;17:201-207.
 87. Liu CH, Liang CC, Liu CJ, Lin JW, Chen SI, Hung PH, et al. Pegylated interferon alfa-2a monotherapy for hemodialysis patients with acute hepatitis C. *Clin Infect Dis* 2010;51:541-549.
 88. Fabrizi F, Dixit V, Messa P, Martin P. Interferon therapy of acute hepatitis C in dialysis patients: meta-analysis. *J Viral Hepat* 2012;19:784-791.
 89. Liu CH, Sheng WH, Sun HY, Hsieh SM, Lo YC, Liu CJ, et al. Peginterferon plus Ribavirin for HIV-infected patients with treatment-naïve acute or chronic HCV infection in Taiwan: A prospective cohort study. *Sci Rep* 2015;5:17410.
 90. Zhang B, Nguyen NH, Yee BE, Yip B, Ayoub WS, Lutchman GA, et al. Treatment of acute hepatitis C infection with pegylated interferon and ribavirin in patients coinfecting with human immunodeficiency virus: A systematic review and meta-analysis. *Intervirology* 2015;58:242-249.
 91. Martinello M, Gane E, Hellard M, Sasadeusz J, Shaw D, Petoumenos K, et al. Sofosbuvir and ribavirin for 6 weeks is not effective among people with recent hepatitis C virus infection: The DARE-C II study. *Hepatology* 2016;64:1911-1921.
 92. Naggie S, Marks KM, Hughes M, Fierer DS, Macbrayne C, Kim A, et al.; AIDS Clinical Trials Group (ACTG) A5327 Study Team. Sofosbuvir plus ribavirin without interferon for treatment of acute hepatitis C virus infection in HIV-1-infected individuals: SWIFT-C. *Clin Infect Dis* 2017;64:1035-1042.
 93. Deterding K, Spinner CD, Schott E, Welzel TM, Gerken G, Klinker H, et al.; HepNet Acute HCV IV Study Group. Ledipasvir plus sofosbuvir fixed-dose combination for 6 weeks in patients with acute hepatitis C virus genotype 1 mono-infection (HepNet Acute HCV IV): an open-label, single-arm, phase 2 study. *Lancet Infect Dis* 2017;17:215-222.
 94. Rockstroh JK, Bhagani S, Hyland RH, Yun C, Dvory-Sobol H, Zheng W, et al. Ledipasvir-sofosbuvir for 6 weeks to treat acute

- hepatitis C virus genotype 1 or 4 infection in patients with HIV coinfection: an open-label, single-arm trial. *Lancet Gastroenterol Hepatol* 2017;2:347-353.
95. Naggie S, Fierer DS, Hughes MD, Kim AY, Luetkemeyer A, Vu V, et al.; Acquired Immunodeficiency Syndrome Clinical Trials Group (ACTG) A5327 Study Team. Ledipasvir/Sofosbuvir for 8 weeks to treat acute hepatitis C virus infections in men with human immunodeficiency virus infections: Sofosbuvir-containing regimens without interferon for treatment of acute HCV in HIV-1 infected individuals. *Clin Infect Dis* 2019;69:514-522.
96. Martinello M, Bhagani S, Gane E, Orkin C, Cooke G, Dore GJ, et al. Shortened therapy of eight weeks with paritaprevir/ritonavir/ombitasvir and dasabuvir is highly effective in people with recent HCV genotype 1 infection. *J Viral Hepat* 2018;25:1180-1188.
97. Boerekamps A, De Weggheleire A, van den Berk GE, Lauw FN, Claassen MAA, Posthouwer D, et al. Treatment of acute hepatitis C genotypes 1 and 4 with 8 weeks of grazoprevir plus elbasvir (DAHHS2): an open-label, multicentre, single-arm, phase 3b trial. *Lancet Gastroenterol Hepatol* 2019;4:269-277.
98. AASLD-IDS A HCV Guidance Panel. Hepatitis C Guidance 2018 Update: AASLD-IDS recommendations for testing, managing, and treating hepatitis C virus infection. *Clin Infect Dis* 2018;67:1477-1492.
99. European Association for the Study of the Liver; Clinical Practice Guidelines Panel: Chair; EASL Governing Board representative; Panel members. EASL recommendations on treatment of hepatitis C: Final update of the series. *J Hepatol* 2020;73:1170-1218. Erratum in: *J Hepatol* 2023;78:452.
100. Martinello M, Orkin C, Cooke G, Bhagani S, Gane E, Kulasegaram R, et al. Short-duration pan-genotypic therapy with Glecaprevir/Pibrentasvir for 6 weeks among people with recent hepatitis C viral infection. *Hepatology* 2020;72:7-18.
101. Matthews GV, Bhagani S, Van der Valk M, Rockstroh J, Feld JJ, Rauch A, et al.; REACT study group; Protocol Steering Committee; Coordinating Centre; Site Principal Investigators. Sofosbuvir/velpatasvir for 12 vs. 6 weeks for the treatment of recently acquired hepatitis C infection. *J Hepatol* 2021;75:829-839.
102. Maasoumy B, Ingiliz P, Spinner C, Cordes C, Stellbrink HJ, zur Wiesch JS, et al. Sofosbuvir plus velpatasvir for 8 weeks in patients with acute hepatitis C: multicenter, single arm, phase 2 study (The HepNet acute HCV-V study). *J Hepatol* 2022;77 Suppl 1:S582.
103. Micallef S, Gauci J, Gerada J. Acute hepatitis C infection with secondary liver injury successfully treated with sofosbuvir/velpatasvir combination. *Br J Hosp Med (Lond)* 2020;81:1-3.
104. Wu SH, Chu CJ, Huang YH, Hou MC. Successful treatment with sofosbuvir and daclatasvir plus ribavirin in acute hepatitis C-infected patient with hepatic decompensation. *J Chin Med Assoc* 2019;82:595-598.
105. Hatanaka T, Naganuma A, Tateyama Y, Yoshinari F, Hoshino T, Sato K, et al. Ledipasvir and sofosbuvir for acute hepatitis C virus mono-infection associated with a high risk of acute liver failure. *Intern Med* 2019;58:2969-2975.
106. Li C, Hu J. A case report of sofosbuvir and daclatasvir to treat a patient with acute hepatitis C virus genotype 2 mono-infection. *Medicine (Baltimore)* 2018;97:e0416.
107. Brancaccio G, Sorbo MC, Frigeri F, Rizzo V, Cantone M, Genderini F, et al. Treatment of acute hepatitis C with ledipasvir and sofosbuvir in patients with hematological malignancies allows early re-start of chemotherapy. *Clin Gastroenterol Hepatol* 2018;16:977-978.
108. Liu H, Zhang T, Yan Y. Sofosbuvir and ribavirin in acute hepatitis C-infected patient with decompensated cirrhosis. *Medicine (Baltimore)* 2016;95:e5555.
109. Chromy D, Mandorfer M, Bucsecs T, Schwabl P, Scheiner B, Schmidbauer C, et al. High efficacy of interferon-free therapy for acute hepatitis C in HIV-positive patients. *United European Gastroenterol J* 2019;7:507-516.
110. Hussein NR, Saleema ZSM, Abd QH. Direct acting antiviral treatment for patients with end-stage kidney disease with acute HCV infection. *Mediterr J Hematol Infect Dis* 2019;11:e2019034.
111. He YL, Yang SJ, Hu CH, Dong J, Gao H, Yan TT, et al. Safety and efficacy of sofosbuvir-based treatment of acute hepatitis C in end-stage renal disease patients undergoing haemodialysis. *Aliment Pharmacol Ther* 2018;47:526-532.
112. Goel A, Bhaduria DS, Kaul A, Verma A, Tiwari P, Rungta S, et al. Acute hepatitis C treatment in advanced renal failure using 8 weeks of pan-genotypic daclatasvir and reduced-dose sofosbuvir. *Nephrol Dial Transplant* 2021;36:1867-1871.
113. Ji Q, Chu X, Zhou Y, Liu X, Zhao W, Ye W. Safety and efficacy of grazoprevir/elbasvir in the treatment of acute hepatitis C in hemodialysis patients. *J Med Virol* 2022;94:675-682.
114. Simmons B, Saleem J, Hill A, Riley RD, Cooke GS. Risk of late relapse or reinfection with hepatitis C Virus after achieving a sustained virological response: A systematic review and meta-analysis. *Clin Infect Dis* 2016;62:683-694.
115. Liu CH, Peng CY, Kao WY, Yang SS, Shih YL, Lin CL, et al. Hepa-

- titis C virus reinfection in patients on haemodialysis after achieving sustained virologic response with antiviral treatment. *Aliment Pharmacol Ther* 2022;55:434-445.
116. Liu CH, Sun HY, Peng CY, Hsieh SM, Yang SS, Kao WY, et al. Hepatitis C virus reinfection in people with HIV in Taiwan after achieving sustained virologic response with antiviral treatment: The RECUR study. *Open Forum Infect Dis* 2022;9:ofac348.
 117. Hosseini-Hooshyar S, Hajarizadeh B, Bajis S, Law M, Janjua NZ, Fierer DS, et al. Risk of hepatitis C reinfection following successful therapy among people living with HIV: a global systematic review, meta-analysis, and meta-regression. *Lancet HIV* 2022;9:e414-e427.
 118. Hajarizadeh B, Cunningham EB, Valerio H, Martinello M, Law M, Janjua NZ, et al. Hepatitis C reinfection after successful antiviral treatment among people who inject drugs: A meta-analysis. *J Hepatol* 2020;72:643-657.
 119. Carson JM, Barbieri S, Matthews GV, Dore GJ, Hajarizadeh B. National trends in retreatment of HCV due to reinfection or treatment failure in Australia. *J Hepatol* 2023;78:260-270. Erratum in: *J Hepatol* 2023 Mar 8. doi: 10.1016/j.jhep.2023.02.020.
 120. Carson JM, Hajarizadeh B, Hanson J, O'Beirne J, Iser D, Read P, et al.; REACH-C Study Group. Effectiveness of treatment for hepatitis C virus reinfection following direct acting antiviral therapy in the REACH-C cohort. *Int J Drug Policy* 2021;96:103422.
 121. Liu CH, Liu CJ, Su TH, Tseng TC, Chen PJ, Kao JH. Sofosbuvir/velpatasvir or glecaprevir/pibrentasvir for treating patients with hepatitis C virus reinfection following direct-acting antiviral-induced sustained virologic response. *Adv Dig Med* 2023;10:34-42.
 122. Weinfurter K, Reddy KR. Hepatitis C viraemic organs in solid organ transplantation. *J Hepatol* 2021;74:716-733.
 123. Zahid MN. Transplantation of organs from hepatitis C virus-positive donors under direct-acting antiviral regimens. *J Clin Med* 2022;11:770.
 124. Liu CH, Chen YS, Wang SS, Kao JH. Treatment of de novo hepatitis C virus-related fibrosing cholestatic hepatitis after orthotopic heart transplantation by ledipasvir and sofosbuvir. *J Formos Med Assoc* 2017;116:407-409.
 125. Raasikh T, Jamali T, Flores A, Cotton RT, Ramanathan V, Tan HP, et al. Systematic review: hepatitis C viraemic allografts to hepatitis C-negative recipients in solid organ transplantation. *Aliment Pharmacol Ther* 2021;54:571-582.
 126. McLean RC, Reese PP, Acker M, Atluri P, Bermudez C, Goldberg LR, et al. Transplanting hepatitis C virus-infected hearts into uninfected recipients: A single-arm trial. *Am J Transplant* 2019;19:2533-2542.
 127. Bethea ED, Gaj K, Gustafson JL, Axtell A, Lebeis T, Schoenike M, et al. Pre-emptive pangenotypic direct acting antiviral therapy in donor HCV-positive to recipient HCV-negative heart transplantation: an open-label study. *Lancet Gastroenterol Hepatol* 2019;4:771-780.
 128. Reyentovich A, Gidea CG, Smith D, Lonze B, Kon Z, Fargnoli A, et al. Outcomes of the treatment with Glecaprevir/Pibrentasvir following heart transplantation utilizing hepatitis C viremic donors. *Clin Transplant* 2020;34:e13989.
 129. Smith DE, Chen S, Fargnoli A, Lewis T, Galloway AC, Kon ZN, et al. Impact of early initiation of direct-acting antiviral therapy in thoracic organ transplantation from hepatitis C virus positive donors. *Semin Thorac Cardiovasc Surg* 2021;33:407-415.
 130. Woolley AE, Singh SK, Goldberg HJ, Mallidi HR, Givertz MM, Mehra MR, et al.; DONATE HCV Trial Team. Heart and lung transplants from HCV-infected donors to uninfected recipients. *N Engl J Med* 2019;380:1606-1617.
 131. Cypel M, Feld JJ, Galasso M, Pinto Ribeiro RV, Marks N, Kuczynski M, et al. Prevention of viral transmission during lung transplantation with hepatitis C-viraemic donors: an open-label, single-centre, pilot trial. *Lancet Respir Med* 2020;8:192-201.
 132. Bethea E, Arvind A, Gustafson J, Andersson K, Pratt D, Bhan I, et al. Immediate administration of antiviral therapy after transplantation of hepatitis C-infected livers into uninfected recipients: Implications for therapeutic planning. *Am J Transplant* 2020;20:1619-1628.
 133. Terrault NA, Burton J, Ghobrial M, Verna E, Bayer J, Klein C, et al. Prospective multicenter study of early antiviral therapy in liver and kidney transplant recipients of HCV-viremic donors. *Hepatology* 2021;73:2110-2123.
 134. Kwong AJ, Wall A, Melcher M, Wang U, Ahmed A, Subramanian A, et al. Liver transplantation for hepatitis C virus (HCV) non-viremic recipients with HCV viremic donors. *Am J Transplant* 2019;19:1380-1387.
 135. Goldberg DS, Abt PL, Blumberg EA, Van Deerlin VM, Levine M, et al. Trial of transplantation of HCV-infected kidneys into uninfected recipients. *N Engl J Med* 2017;376:2394-2395.
 136. Reese PP, Abt PL, Blumberg EA, Van Deerlin VM, Bloom RD, Potluri VS, et al. Twelve-month outcomes after transplant of hepatitis C-infected kidneys into uninfected recipients: A single-group trial. *Ann Intern Med* 2018;169:273-281.
 137. Durand CM, Bowring MG, Brown DM, Chattergoon MA, Mas-

- saccesi G, Bair N, et al. Direct-acting antiviral prophylaxis in kidney transplantation from hepatitis C virus-infected donors to noninfected recipients: An open-label nonrandomized trial. *Ann Intern Med* 2018;168:533-540.
138. Durand CM, Barnaba B, Yu S, Brown DM, Chattergoon MA, Bair N, et al. Four-week direct-acting antiviral prophylaxis for kidney transplantation from hepatitis C-viremic donors to hepatitis C-negative recipients: An open-label nonrandomized study. *Ann Intern Med* 2021;174:137-138.
139. Sise ME, Goldberg DS, Kort JJ, Schaubel DE, Alloway RR, Durand CM, et al. Multicenter study to transplant hepatitis C-infected kidneys (MYTHIC): An open-label study of combined glecaprevir and pibrentasvir to treat recipients of transplanted kidneys from deceased donors with hepatitis C virus infection. *J Am Soc Nephrol* 2020;31:2678-2687.
140. Gupta G, Yakubu I, Zhang Y, Kimball P, Kang L, Mitchell K, et al. Outcomes of short-duration antiviral prophylaxis for hepatitis C positive donor kidney transplants. *Am J Transplant* 2021;21:3734-3742.
141. Chen R, Li D, Zhang M, Yuan X. Sofosbuvir/velpatasvir prophylaxis for 12 weeks in hepatitis C virus (HCV)-negative recipients receiving kidney transplantation from HCV-positive donors. *Ann Transplant* 2021;26:e933313.
142. Feld JJ, Cypel M, Kumar D, Dahari H, Pinto Ribeiro RV, Marks N, et al. Short-course, direct-acting antivirals and ezetimibe to prevent HCV infection in recipients of organs from HCV-infected donors: a phase 3, single-centre, open-label study. *Lancet Gastroenterol Hepatol* 2020;5:649-657. Erratum in: *Lancet Gastroenterol Hepatol* 2020;5:e6.
143. Bethea ED, Chen Q, Hur C, Chung RT, Chhatwal J. Should we treat acute hepatitis C? A decision and cost-effectiveness analysis. *Hepatology* 2018;67:837-846.
144. AASLD-IDSA. HCV guidance: recommendations for testing, managing, and treating hepatitis C. AASLD-IDSA web site, <<https://www.hcvguidelines.org/>>. Accessed 27 Jan 2023.
145. Naggie S, Holland DP, Sulkowski MS, Thomas DL. Hepatitis C virus postexposure prophylaxis in the healthcare worker: Why direct-acting antivirals don't change a thing. *Clin Infect Dis* 2017;64:92-99.
146. Liu CH, Kao JH. Last mile to microelimination of hepatitis C virus infection among people living with human immunodeficiency virus. *Clin Infect Dis* 2021;73:e2172-e2174.
147. Frey SE, Houghton M, Coates S, Abrignani S, Chien D, Rosa D, et al. Safety and immunogenicity of HCV E1E2 vaccine adjuvanted with MF59 administered to healthy adults. *Vaccine* 2010;28:6367-6373.
148. Page K, Melia MT, Veenhuis RT, Winter M, Rousseau KE, Mas-saccesi G, et al. Randomized trial of a vaccine regimen to prevent chronic HCV infection. *N Engl J Med* 2021;384:541-549.
149. Donnison T, McGregor J, Chinnakannan S, Hutchings C, Center RJ, Pountourios P, et al. A pan-genotype hepatitis C virus viral vector vaccine generates T cells and neutralizing antibodies in mice. *Hepatology* 2022;76:1190-1202.