Check for updates

OPEN ACCESS

EDITED BY Arvind Bagga, All India Institute of Medical Sciences, India

REVIEWED BY Divya Bhatia, NewYork-Presbyterian, United States William Smoyer, Nationwide Children's Hospital, United States Cynthia Nast, Cedars-Sinai Medical Center, United States

*CORRESPONDENCE Gian Marco Ghiggeri gmarcoghiggeri@gaslini.org

RECEIVED 16 February 2023 ACCEPTED 21 June 2023 PUBLISHED 12 July 2023

CITATION

Vincenti F, Angeletti A and Ghiggeri GM (2023) State of the art in childhood nephrotic syndrome: concrete discoveries and unmet needs. Front. Immunol. 14:1167741. [doi: 10.3389/fimmu.2023.1167741](https://doi.org/10.3389/fimmu.2023.1167741)

COPYRIGHT

© 2023 Vincenti, Angeletti and Ghiggeri. This is an open-access article distributed under the terms of the [Creative Commons](http://creativecommons.org/licenses/by/4.0/) [Attribution License \(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

[State of the art in childhood](https://www.frontiersin.org/articles/10.3389/fimmu.2023.1167741/full) [nephrotic syndrome: concrete](https://www.frontiersin.org/articles/10.3389/fimmu.2023.1167741/full) [discoveries and unmet needs](https://www.frontiersin.org/articles/10.3389/fimmu.2023.1167741/full)

Flavio Vincenti¹, Andrea Angeletti² and Gian Marco Ghiggeri^{2*}

1 Division of Nephrology, Department of Medicine and Department of Surgery, University of California San Francisco, San Francisco, CA, United States, 2Nephrology Dialysis and Transplantation, Istituto di Ricerca e Cura a Carattere Scientifico (IRCCS) Istituto Giannina Gaslini, Genoa, Italy

Nephrotic syndrome (NS) is a clinical entity characterized by proteinuria, hypoalbuminemia, and peripheral edema. NS affects about 2–7 per 100,000 children aged below 18 years old yearly and is classified, based on the response to drugs, into steroid sensitive (SSNS), steroid dependent, (SDNS), multidrug dependent (MDNS), and multidrug resistant (MRNS). Forms of NS that are more difficult to treat are associated with a worse outcome with respect to renal function. In particular, MRNS commonly progresses to end stage renal failure requiring renal transplantation, with recurrence of the original disease in half of the cases. Histological presentations of NS may vary from minimal glomerular lesions (MCD) to focal segmental glomerulosclerosis (FSGS) and, of relevance, the histological patterns do not correlate with the response to treatments. Moreover, around half of MRNS cases are secondary to causative pathogenic variants in genes involved in maintaining the glomerular structure. The pathogenesis of NS is still poorly understood and therapeutic approaches are mostly based on clinical experience. Understanding of pathogenetic mechanisms of NS is one of the 'unmet needs' in nephrology and represents a significant challenge for the scientific community. The scope of the present review includes exploring relevant findings, identifying unmet needs, and reviewing therapeutic developments that characterize NS in the last decades. The main aim is to provide a basis for new perspectives and mechanistic studies in NS.

KEYWORDS

nephrotic syndrome, minimal change disease, focal-segmental glomerulosclerosis, post-transplant recurrence, proteinuria, monoclonal antibodies, rituximab

Introduction

Nephrotic syndrome (NS) is a clinical condition that occurs frequently in children and manifests with the classical clinical triad of severe proteinuria, hypoalbuminemia, and diffuse edema ([1](#page-10-0)). Despite homogeneity of the clinical pattern at presentation, NS may evolve with different outcomes, characterized by unpredictable response to drugs and development of renal failure, that probably reflect different pathological entities [\(1,](#page-10-0) [2\)](#page-10-0).

Histological patterns of childhood NS vary from minimal glomerular lesions (MCD) to focal areas of segmental sclerosis (FSGS). These two histological opposites share the effacement of the slit diaphragm of podocytes, which effectively sustains the proteinuria [\(3](#page-10-0)). If FSGS represents a subsequent stage of MCD or if MCD and FSGS are two distinguished histological entities is still debated [\(4\)](#page-10-0).

Pathogenesis of childhood NS is largely unknown and, therefore, it is defined as idiopathic NS (iNS) in cases without a definite origin, differentiated from secondary NS characterized by causative mechanisms. An important group of secondary NS has a genetic origin and is characterized by the association with a pathogenic variant in genes transcribing for proteins of podocytes and/or of glomerular structures [\(5\)](#page-10-0). Causative genetic variants of NS are identified in 66% of congenital and infantile cases, 30% of children and, in approximately 10-15% of young adults presenting with NS [\(2](#page-10-0), [5\)](#page-10-0). A second group of secondary NS occurring in children (but not limited to young ages) is characterized by the temporal associations with either virus infections or drug administration [\(6,](#page-10-0) [7\)](#page-10-0).

Therefore, a clear differentiation between iNS and secondary NS has key clinical importance for prognosis and for the choice of therapies since genetic NS has, in general, limited response to drugs (see dedicated part). On the other hand, defying the pathological mechanisms that may sustain iNS and, in particular, iNS resistant to drugs, represents one of the major enigmas in nephrology. Several theories have been developed over the years and the analysis of the most relevant findings in this field will be the focus of the present review.

How to define idiopathic nephrotic syndrome

As previously reported, childhood NS is a clinical condition characterized by generic signs (i.e. severe proteinuria, hypoalbuminemia, and edema ([1\)](#page-10-0)) that may occur in secondary NS and/or in association with several glomerulonephritis common in adults, such as primary and secondary autoimmune forms (ie. membranous and lupus nephropathy), metabolic and genetic conditions (ie. diabetic and hypertensive nephropathy and Alport syndrome), and many others. iNS may be defined on the basis of the above typical symptoms combined with the lack of any evidence of a genetic, infective, inflammatory, or autoimmune cause. Further key factors contributing to the classification of iNS are age at onset, response to treatment, and histological patterns.

Age

iNS is a disease that typically affects children and young adults ([1](#page-10-0)). Genetic NS, resulting from molecular modifications of podocyte components, usually manifests in the first 12-24 months of life (30% of cases) and has the peak of onset between 2 and 18 years (50-60% of cases); the remaining 10-20% of genetic NS cases present at older ages ([2](#page-10-0), [5\)](#page-10-0). NS secondary to viral infections and/or associated with drugs may occur any age. Among others, secondary conditions causing NS, such as membranous nephropathy, lupus nephritis, IgA glomerulonephritis, Alport syndrome, metabolic disorders including diabetes mellitus and hypertension, and neoplasms such as myeloma, occur more frequently in adults ([8](#page-10-0)). Therefore, age represents a key classification element of NS.

Response to drugs

The pharmacology approach to iNS (Figure 1) has been consolidated over the years and now represents a crucial element for classification. Corticosteroids represent the first step in the treatment of iNS ([9](#page-10-0), [10](#page-10-0)). Corticosteroid sensitivity occurs in about 80-90% of new-onset iNS, however, half of them develop corticosteroid dependance (SDNS) and about 10-20% result in

corticosteroid resistance (SRNS). Both SDNS and SRNS require the administration of the so=called 'steroid sparing' drugs, to reduce the adverse effects correlated with long courses of corticosteroids [\(11\)](#page-10-0). Classical 'steroid sparing' agents are levamisole, mycophenolate mofetyl (MMF), cyclophosphamide, and calcineurin inhibitors (CNI, cyclosporin and tacrolimus) ([9](#page-10-0), [12](#page-10-0)). Anti-CD20 monoclonal antibodies have been more recently introduced in therapeutic schemes [\(13,](#page-11-0) [14](#page-11-0)). In SDNS, the administration of a single 'steroid sparing' agent is usually effective in maintaining remission, while SRNS may require more than one drug, in which case NS is defined as 'multidrug dependent' (MDNS) ([15](#page-11-0)). Limited SRNS that are resistant to the association of any agent and are defined as multidrug resistant (MRNS). These forms account for around 5-10% of overall cases, and no therapeutic approaches are effective in limiting proteinuria. Moreover, in MRNS the progression to renal failure usually occurs in a few years [\(16,](#page-11-0) [17](#page-11-0)).

Genetic backgrounds

Monogenic NS accounts for a significant number of NS ([18\)](#page-11-0). Causative pathogenic gene variants are identified in 66% of congenital and infantile cases, 30% of children and, in approximately 10-15% of young adults presenting with SR/ MRNS. So far, more than 70 single gene causes of SRNS/MRNS have been reported [\(6\)](#page-10-0). NPHS1 (19q13.12) and NPHS2 (1q25.2) are, by far, the two main autosomal recessive genes of genetic NS, while INF2 (14q32.33) and WT1 (11p13) are the leading cause of autosomal dominant NS. See below for the dedicated section [\(2\)](#page-10-0). Genetic NS must, therefore, be taken into account as a classificatory element in very young patients with NS and in SRNS (see below, the dedicated section).

Renal pathology

Renal pathology is fundamental to exclude secondary causes of NS, such as autoimmune and inflammatory forms, that occur more frequently in adults. There is consensus in considering a renal pathology approach necessary in NS occurring in adults and in SRNS. In childhood, kidney biopsy is usually not performed at the onset of NS and in subjects with SSNS. KDIGO guidelines suggest limiting kidney biopsy in children to those with a late response to corticosteroids or to those suspicious of an underlying secondary renal pathology [\(3,](#page-10-0) [11,](#page-10-0) [19\)](#page-11-0). Recent findings suggested a possible pathogenetic role of anti-nephrin antibodies in some cases of iNS ([20](#page-11-0)). Based on these findings, the diagnostic approach to iNS consolidated over years may be modified and the indication to kidney biopsy may be revised.

Historically, three major histological patterns have been associated with iNS and still represent useful tools for classification of the disease: a) the absence of any glomerular alteration at the optical microscopy is defined as MCD and transient electron microscopy reveals effacement of podocytes with alteration of the slit diaphragm that is considered the reason for proteinuria; b) the second pattern is more rare and is characterized by mesangial expansion with IgM deposit; and c) the third group presents segmental sclerotic lesions that focally interest glomeruli (FSGS); proliferation of Bowman epithelia and adhesion of glomeruli with the capsule, or tip lesions, are variants of FSGS.

For many years, pathology represented the unique classificatory element of NS until it was proven that FSGS may be the common histological final step of different renal diseases ([21\)](#page-11-0). However, the pathological mechanisms explaining the sclerotic degeneration of glomerular tuft common to several glomerulonephritis are still unknown.

The attempt to correlate the different pathology patterns with the response to drugs in children with NS (MCD with SDNS and FSGS with MDNS or MRNS) failed to provide clinical elements for important overlaps between the different categories [\(5\)](#page-10-0). A still open point is, if FSGS is an evolutionary phase of MCD, this seems in contrast with the reported concept on FSGS as the main feature in many autoimmune and degenerative glomerulonephritis [\(4](#page-10-0)).

Causes of secondary nephrotic syndrome (1): monogenic NS susceptibility

Monogenic NS is the first and most unique case in which either the causes and mechanisms of the disease have been elucidated ([Table 1\)](#page-3-0). Moreover, genetic susceptibility offers the opportunity to consider mechanisms that predispose to NS where genetics represents a first hit.

Monogenic NS

The discovery of genetic backgrounds responsible for mendellian NS has represented a real breakthrough in terms of evolution on pathogenesis and clinical impact [\(18\)](#page-11-0). To date, almost 70 genes have been described in association with autosomal recessive (prevalent) and dominant (rare) traits of NS including syndromes [\(22](#page-11-0)–[34](#page-11-0)) (a detailed review on genes involved in iNS is outside the scope of this review). Most mutations are associated with SRNS; six genes involved in GTPase activity have been recently identified as a cause of SDNS ([27](#page-11-0)). Many genes involved in SRNS code for proteins of the the slitdiaphragm, such as nephrin (NPHS1) [\(35](#page-11-0)), or are directly linked to this structure, such as podocin (NPHS2) [\(29](#page-11-0)). Others are components of the cytoskeleton of podocytes such as actinin 4 (ACTN4), myosin, and inverted formin 2 (INF2) ([31](#page-11-0), [32,](#page-11-0) [36](#page-11-0)). All these molecules are directly involved in maintaining the sieving properties of the glomerulus in terms of selectivity towards the charge and the dimension of circulating proteins. Besides confirming that the podocyte is the basic cell of the glomerulus involved in proteinuria, genetic studies have produced progress in the clinical management since, with the important exceptions of mutations of two mitochondrial genes coding for Coenzymes Q2 (COQ2) and Q6 (COQ6) [\(37](#page-11-0)) and of the transient receptor potential cation channel subfamily C member 6 (TRCP6) [\(34](#page-11-0)), genetic NS are only partially responsive to drugs [\(38\)](#page-11-0) and require only symptomatic and antproteinuric generic interventions ([Figure 1\)](#page-1-0). In the former case, ie. mitochondrial COQ2 and COQ6 mutations, oral coenzyme Q10 may revert the clinical phenotypes ([37](#page-11-0)). In the case of the mutation gain of function mutations of $TRCP6$, a $Ca⁺$ channel important for muscle cells functions, Sildenafil down-regulates the channel expression by reducing the cGMP through a peroxisome proliferator–activated receptor needed for its synthesis ([39](#page-11-0), [40\)](#page-11-0). Therefore, it was suggested that Sildenafil, reducing the activity of TRCP6, could revert this genetic form of NS.

Susceptibility

The existence of rare familial aggregates of patients with corticosteroid sensitive iNS ([41\)](#page-11-0) has long been reported, highlighting the possibility of a common risk linked with susceptibility. The lack of clear inheritance traits supports the existence of a two-hit mechanism, where a susceptibility gene or locus is shared by different individuals in the same family and NS is triggered by a second hit, such as an infectious agent. Familial susceptibility is supported by findings on the genetic architecture of pediatric and adult patients. Genome-wide association studies (GWAS), including a large spectrum of populations in the USA, Europe, and Asia, identified a risk locus in the HLA Class II consisting of HLA-DQA1 and HLA-DQB1 and three additional signals, the Calcium Homeostasis Modulator Family Member 6 (CALHM6), TNF Superfamily Member 15 (TNFSF15), and nephrin ([42](#page-11-0)–[44\)](#page-11-0). The non-HLA loci are linked with expression quantitative trait loci (eQTLs) of monocytes and T cells supporting the concept that an immunomodulatory disfunction has a prominent role in conferring susceptibility to iNS. Large collaborative studies on many thousands of patients is surely the way to obtain further solid information that would rebound on pathogenesis.

APOL1 is a susceptibility gene for FSGS ([45](#page-11-0)) linked with race that has been documented in subjects of West African ancestry. Adults with this ancestry have, in fact, a 4-5 fold higher risk of developing FSGS than Europeans. Genetic variants of APOL1 [\(62,](#page-12-0) [63](#page-12-0)) have likely developed in response to (and to protect from) Trypanosoma brucei rhodesiense and Trypanosoma brucei gambiense, which are very frequent in all African contexts.

Causes of secondary nephrotic syndrome (2): viruses and drugs

Viruses and Drugs have been proposed as potential causes of secondary NS for patients younger than 18 (Table 1). They would be better defined as associated triggers because the real mechanism causing NS is unknown or only partially characterized.

Viruses

Despite the numeric impact of viral infections associated with NS, the clinical importance is limited since they are potentially reversible ([6](#page-10-0), [64\)](#page-12-0). Actually, NS has been demonstrated in TABLE 1 Causes of secondary NS.

mTOR, mammalian target of rapamycin; VEGF, vascular endothelial growth factor.

association with SARS-CoV-5 (in children) and with HIV infections (prevalent in adults).

New episodes of NS were described in two children with acute SARS-CoV-2 infection, whereas the virus has been reported more frequently in association with relapses of NS in patients who had already presented the disease ([46](#page-11-0)). The clinical course has been good in all cases in terms of response to corticosteroids.

HIV infections have been associated with a severe form of collapsing FSGS (HIVAN) complicated by extensive tubular lesions and microcystis [\(47\)](#page-11-0). In the pre-antiretroviral therapy era, HIVAN represented an important cause of ESRD in many countries even though its incidence has declined with the adoption of specific therapies ([48](#page-11-0)). Studies on mechanisms responsible for HIVAN ([49\)](#page-11-0) have shown a direct infection of HIV of podocytes and tubular epithelia leads to the dysregulation of cell cycle with inflammation, alteration of the cytoskeleton, and cell death ([50,](#page-11-0) [51\)](#page-11-0). The high frequency of two variants of Apolipoprotein A1 (G1 and G2) in persons of African ancestry predispose to a very high risk of HIVAN in African Americans and in South Africans with HIV infection [\(52](#page-11-0), [62](#page-12-0)).

Parvovirus B19 DNA has been isolated in renal tissues of a high percentage of patients with glomerular diseases, including MCD, collapsing FSGS, and membranous nephropathy, suggesting a possible role of the virus in the pathogenesis of the disease ([53\)](#page-11-0). The very high frequency of kidney tissues with Parvovirus, absence of specificity, and lack of association with clinical signs of Parvovirus infections speak against a direct involvement in NS.

Drugs

The administration of drugs has been associated with the development of NS; Pamidronate and Lithium were the first to be reported ([54](#page-11-0)–[56\)](#page-11-0). Pamidronate belongs to the class of bisphosphonates that are utilized as first-class agents in osteoporosis and in Paget syndrome. Lithium is a major antipsychotic drug widely utilized in psychiatric patients. More recently, NS has been reported following treatments with three other categories of drugs, ie. mTOR inhibitors, vascular endothelial growth factor blocking agents, and irreversible Bruton's tyrosine kinase inhibitors. The mTOR inhibitor sirolimus is an anti-rejection therapy that may substitute other immudepressors in patients with solid organ transplanationis ([57](#page-11-0)); a mechanism for renal toxicity linked with mTOR has been proposed based on the block of podocytes' compensatory hypertrophy, activated after podocyte loss ([58\)](#page-11-0). Also, the intravitreal vascular endothelial growth factor blocking agents bevacizumab and aflibercet are widely utilized in clinical practice, being drugs of choice for macular degeneration and diabetic retinopathy ([59](#page-11-0)). The irreversible Bruton's tyrosine kinase inhibitor Ibrutinib is essential for the treatment of chronic lymphocytic leukemia ([65](#page-12-0)) and mantle-cell lymphoma ([66](#page-12-0)). Several

reports describe the occurrence of NS following Ibrutinib, but the mechanisms are not understood [\(60](#page-12-0), [61](#page-12-0)).

Mechanisms for idiopathic nephrotic syndrome (1): membrane selectivity, oxidative stress, and circulating factors

The sections below are dedicated to those forms of NS that usually occur in children without a recognized cause and/or of pathogenetic mechanisms. Therefore, as previously mentioned, these forms are defined as iNS. We will summarize the most relevant pathological theories proposed over the years. Despite the absence of any definitive demonstration, the proposed theories may still represent potential future lines of research to be developed and explored (Figure 2; [Table 2\)](#page-5-0).

Glomerular barrier sieving property modification

The glomerular basement membrane (GBM) is a selective filter that allows the passage of proteins according to size and electrical charge: molecules between 10 and 36 Armstrong (A) are filtered in relation to size (maximal for 10 A) whereas molecules > 36 A are filtered on the basis of the charge with privilege for those with a cationic isoelectric point (pI>7) [\(134](#page-13-0), [135\)](#page-13-0). This function is mostly due to the presence of sialic aid in the external membrane of podocytes that confers a negative charge to the structure. In

Primary and secondary childhood NS. Inherited forms of NS, caused by pathological variants in genes transcribing for the main protein of glomerular tuft, represent a unique group of NS for which a clear mechanism has been defined. NS associated with either virus infections or drugs are reported as secondary NS on the basis of both the temporal association and reversibility of proteinuria. All other mechanisms are supposed and require further confirmatory results..

TABLE 2 Idiopathic Nephrotic syndrome (iNS).

This table covers a wide range of potential factors implicated in the pathogenesis of iNS that should be further confirmed.

accordance with the function of sialic acid, podocyte-specific ablation of sialylation in mice caused a phenotype resembling human FSGS ([113,](#page-13-0) [136\)](#page-13-0). Also, sphingolipids play a crucial role in maintaining renal permeability. In line, previous reports suggest that their accumulation in podocytes characterize several metabolic and non-metabolic diseases, including FSGS, and may contribute to altering glomerular permeability [\(114](#page-13-0), [137\)](#page-13-0).

In normal conditions, urine contains minimal amounts of proteins since cationic proteins are in trace amounts in serum and small proteins that cross the glomerular filter are reabsorbed by proximal tubular cells. Filtered albumin is also almost completely reabsorbed by proximal tubular cells via endocytosis ([138,](#page-13-0) [139](#page-13-0)).

iNS is caused by an alteration of the podocyte filtering unit wherein the capacity to repulse proteins on the basis of size and charge is only partially maintained and anionic proteins of any size can pass the filter and are lost into urine. Studies carried out between 1970 and 1980 showed that the selectivity index calculated from urinary excretion and tubular reabsorption (determined by the direct staining in renal biopsies) of albumin and IgG correlated with the intensity of pathological lesions ([140\)](#page-13-0), supporting the concomitant reduction in both pore size and density of GBM in MCD [\(97\)](#page-12-0).

However, modifications of the charge of circulating proteins may partially sustain proteinuria and be a further pathogenetic factor in NS. Studies on charge selectivity were performed considering albumin as a model of sieving modifications, since it accounts for 90% of all urinary proteins in MCD. Albumin is a 46A molecule with anionic charge (pI 4.6). In normal conditions, it is completely repulsed by the podocyte filtering unit for either size and charge characteristics. In MCD, about 50% of urinary albumin isoforms are more cationic compared to serum albumin (4.8-5.2 vs. 4.6) and are also more cationic compared to urinary albumin after remission of the disease. In FSGS, urinary and serum albumin have the same charge, suggesting that charge selectivity of albumin is partially maintained in MCD whereas it is lost in FSGS ([98\)](#page-12-0).

Other studies reported that higher cationic isoforms of albumin are characterized by lower amounts of fatty acids compared to the anionic compounds ([99](#page-12-0)). This finding makes sense, since fatty acids that are transported by albumin in serum confer the characteristic anionic charge to this protein. However, other anionic residues may be implicated in regulating charge properties of circulating proteins. Among others, sialic acid is a strong acidic molecule (pK 2.6-2.9) that is transported in serum by a few proteins such as immunoglobulins, fibrinogen, and alpha-2-macrogobulin. Sialidase enzymes regulate the sialylation of transporter proteins and, in parallel, their charge is anionic for sialylated proteins and cationic for de-sialylated isoforms. Recent findings demonstrate that sialylation of immunoglobulins modulate their cell binding and play an important role in determining a pathological function ([141,](#page-13-0) [142](#page-13-0)). One example is rheumatoid arthritis, where the ratio between de-sialylated and sialylated immunoglobulins directly correlated with the activity of the disease [\(143](#page-13-0)–[145\)](#page-13-0). Moreover, variations in sialic acid of immunoglobulins, and in particular of IgM, modulate the immunoinflammatory functions of T cells in MCD ([110\)](#page-13-0). Therefore, more extensive analysis to investigate the possible pathological role of IgM sialydation in iNS is necessary.

Oxidative stress

iNS is commonly associated with a systemic oxidative milieu. Extensive proteolysis of albumin and α 1-antitrypsin has been documented in the plasma and urine of patients with FSGS [\(111,](#page-13-0) [112\)](#page-13-0). α 1-antitrypsin is an enzyme with anti-proteolytic activity whose fragmentation implies increased proteolysis. Albumin is the principal anti-oxidant in blood: the SH residue of Cys34 is sulphonated in the presence of an oxidative stress with the addition of ±48Da molecular weight and changes of the net charge for the addition of negative residues ([146](#page-13-0), [147\)](#page-13-0). The increased amounts of oxidized albumin in FSGS demonstrates the oxidative stress. Other indirect evidence supports the implication of oxidants in FSGS. Patients carrying a mutation in the two mitochondrial genes COQ2 and COQ6 develop FSGS and revert the phenotype after supplementation with Coenzyme Q10 ([37\)](#page-11-0). All experimental models of FSGS utilize oxidative substances such as adriamycin and puromicin ([148,](#page-13-0) [149\)](#page-13-0). Overall, there is a direct correlation between oxidation and proteolysis that is able to damage renal structures.

Circulating permeability plasma factors

iNS and, in particular, FSGS have been considered the results of circulating plasma factors that bind and permeabilize the filtering glomerular unit ([67,](#page-12-0) [68](#page-12-0), [150](#page-13-0)). Recurrence of NS in the transplanted graft is often used as evidence of the existence of a circulating permeability factor that mediates glomerular injury in around 50% of patients ([69](#page-12-0)–[71](#page-12-0)). The experience described by Gallon et al. ([72\)](#page-12-0) represents definitive support of this concept. They removed a renal graft after a few days of transplantation from a subject with FSGS presenting recurrence of the disease immediately after the kidney transplant. The graft was then transplanted in a second subject with ESRF due to a malformative renal disease, observing the recovery of podocyte effacement previously demonstrated by the renal biopsy during the first transplant and absence of proteinuria in the second patient.

The story of pathogenesis of FSGS coincided for many years with the research of permeability circulating plasma factors. Initial studies tested the efficacy of several potential molecules in inducing permeability in an in vitro model with isolated glomeruli ([73](#page-12-0)). A few molecules emerged as potential plasma factors, however, we are still far from any definitive conclusion and the nature of the permeability factor is still pending ([67\)](#page-12-0). We here describe some molecules that have been reported as valuable candidates for permeability factors.

CD40L

The interest on CD40L and its natural target CD40 derives from the experimental observation that CD40L promotes redistribution of nephrin in both podocytes and glomeruli and increases permeability to albumin [\(74](#page-12-0), [75\)](#page-12-0). CD40L exists as a soluble circulating factor and CD40 is a costimulatory molecule present on the surface of cells involved in immunologic response such as B cells and monocytes/macrophages. CD40 is also constitutively expressed in podocytes ([75\)](#page-12-0). CD40L/CD40 axis promotes inflammatory events [\(76,](#page-12-0) [77\)](#page-12-0) with activation of metalloproteases, chemokines, urokinase, and the soluble urokinase plasminogen activator receptor (suPAR). Serum levels of sCD40L are increased in children with both SSNS and SRNS and in adult patients with biopsy-proven FSGS compared to healthy subjects ([75\)](#page-12-0). On the other hand, serum levels of CD40L are comparable in children with congenital NS and in patients with membranous nephropathy.

In vivo studies have shown that CD40 is strongly expressed in podocytes of patients with both primary and post-transplant recurrent FSGS and the presence of circulating anti-CD40 antibodies are present in the serum of the same patients ([78](#page-12-0)). It has been proposed that anti-CD40 antibodies purified from the serum of patients with recurrent FSGS disrupt the podocyte

structure by acting on F-actin filaments with the involvement of the suPAR– β 3 integrin signaling pathway ([78](#page-12-0)).

c-Mip

c-Mip (C-maf inducing protein) is an 86 KDa protein with unclear functions that is upregulated in the podocytes of patients with NS in association with conditions such as cancer that predispose the development of NS. c-Mip levels also increase in concomitance with drugs, such as the angiogenesis inhibitors sorafenib and sunitinib [\(79](#page-12-0)-[81](#page-12-0)) that may associate with NS. In vivo studies showed that transgenic mice over-expressing c-Mip develop a disease resembling human MCD ([82](#page-12-0)). However, data on c-Mip serum levels in patients with NS are not available and no pathological mechanisms to correlate c-Mip and proteinuria have been proposed. In the case of inhibitors of angiogenesis that act on vascular endothelial growth factor-receptor tyrosine kinase (VEGF-TKIs), it has been hypothesized that the reduction of RelA, a member of the NFk-B family, is an inhibitor of c-Mip [\(80\)](#page-12-0). More generally, one explanation for proteinuria occurring in association with c-Mip stimulation is that it interacts with factors such as Fyn, a Src kinase that is involved in nephrin phosphorylation ([83](#page-12-0)) and, therefore, c-Mip may directly affect the podocyte slit diaphragm [\(84](#page-12-0)).

uPA/suPAR

The possible implication of suPAR in FSGS derives from in vivo animal models of the disease and does not reflect observation in humans ([85](#page-12-0), [86](#page-12-0)). In fact, serum levels of suPAR are not increased in NS and simply correlate with renal function [\(87\)](#page-12-0). This molecule is a determinant of proteinuria in experimental models of FSGS; mice lacking uPAR (PLAUR-/-) are protected from developing proteinuria ([85](#page-12-0)). The plasminogen activator receptor (uPAR) is expressed by podocytes and it functions to maintain podocyte shape and sieve properties by modifying the $\alpha v\beta3$ -integrin assembly and adhesion to extracellular matrix.

Angiopoietin-like 4

ANGPTL4 is a glycoprotein expressed by several tissues and organs, including glomeruli. It is a known inhibitor of lipoprotein lipase (an enzyme that catalyzes conversion of triglycerides to monoglycerides and free fatty acids) and plays a main role in reducing triglyceride levels in circulation. ANGPTL4 has two isoforms with elevated (pI >8) and neutral (pI 7) isoelectric point ([88\)](#page-12-0) that are characterized by different sialic acid contents: sialylated ANGPTL4 (the neutral isoform found in circulation) is secreted in peripheral organs (mostly skeletal muscle, heart, and adipose tissue) while the hypo-sialylated isoform is produced by podocyte and remains restricted to the kidney. Several observations indicate a potential involvement of ANGPTL4 in NS, but the interpretation of data is not univocal. In fact, circulating sislylated ANGPTL4 is high in NS and plays an anti-proteinuric effect by binding $\alpha v\beta5$ integrin in glomerular endhotelim ([89](#page-12-0)) whereas the renal hypo-syalilated isoform determines proteinuria ([90\)](#page-12-0). In experimental models of NS, levels of the two isoforms of ANGPTL4 are under the control, with an opposite effect, of corticosteroids that up-regulate the expression of sialylated isoforms in adipose tissue and increase their circulating levels while reducing ANGPTL4 expression in podocytes ([91\)](#page-12-0). Therefore, the efficacy of corticosteroids in human NS is likely linked with the combined ability to reduce a potential mediator of proteinuria (renal ANGPTL4) and increase circulating levels of ANGPTL4 that plays an anti-proteinuric effect.

Hemopexin

Plasma hemopexin is a glycoprotein with serine protease activity ([92\)](#page-12-0) involved in iron homeostasis as a binder of free heme. It is considered an antioxidant protein, mostly expressed during acute phases of inflammation ([93](#page-12-0), [94](#page-12-0)). Rats treated with a tail infusion of hemopexin developed reversible proteinuria ([95\)](#page-12-0). In humans, plasma and urine levels of hemopexin have been reported to be decreased in subjects with acute relapse of MCD, compared to the high levels described in proteinuric subjects with FSGS, membranous proliferative glomerulonephritis, or IgA nephropathy, suggesting specificity of changes for MCD ([96](#page-12-0)).

Second-hit mechanism (B7-1)

B7-1 (CD80) is a costimulatory ligand expressed on the surface of antigen-presenting cells (APCs), the binding of which to the Tcell receptors CD28 and CTLA-4 is essential for activating and regulating T-cell immunity. Previous findings described the B7 expression on podocytes' surface of subjects with NS and in posttransplant recurrence of FSGS ([78](#page-12-0), [100](#page-12-0), [101](#page-12-0)). The majority of studies favor the idea that B7-1 may represent a non-specific marker of proteinuric disease, such as MCD, lupus nephritis, membranous nephropathy, and diabetic nephropathy, but not in FSGS [\(102](#page-12-0)– [105\)](#page-12-0). Therefore, de novo expression of glomerular B7-1 in FSGS recurrence may be considered as a local tissue response to nonspecific stimuli (ie. oxidants, inflammatory, infectious) ([106\)](#page-12-0). The expression of B7-1 secondary to these stimuli represent a possible antigen, as part of a second-hit mechanism. Such findings suggest caution in interpretating the significance of glomerular B7-1 expression in post-transplant FSGS recurrence. The reason why B7-1 expression is expressed only in limited patients with posttransplant recurrence of FSGS is unclear. However, such reports raised the hypothesis that B7 blockade may result in a podocyteprotective effect with consequent reduction of proteinuria. Abatacept, a B7-1 inhibitor, was proposed as a specific therapeutic agent in post-transplant FSGS recurrence, with discordant results. In the most recent study, Burke III et al. ([101](#page-12-0)), proposed the administration of abatacept, based on the B7-1 podocytes expression at kidney biopsy, in 12 subjects (median age 12 years old) with NS recurrence after kidney transplant and resistant to conventional treatments with plasmapheresis and rituximab. Nine subjects responded to treatment, of whom seven had a kidney biopsy positive for B7-1, while two were without biopsy. Of note, of the three patients not responding to abatacept, one had a kidney biopsy positive for B7-1. Based on these results, authors suggested that B7-1 podocyte staining may identify subjects who can benefit from abatacept.

Mechanisms for idiopathic nephrotic syndrome (2): complement, interleukines, and immune cells

$C3a/C3aR-IL1\beta$ loop

Recent findings suggested that regulation of C3 convertase in podocytes by CD55, a decay-accelerating factor, is implicated in determining proteinuria and glomerular sclerosis in mice models of glomerulosclerosis induced by adriamycin ([115\)](#page-13-0). C3a/C3aR interaction enhances inflammasome activation in podocytes and the release of active IL-1 β that binds IL-1R1 and leads to actin cytoskeleton rearrangement and podocyte disarrangement. Proteinuria in these mice prevents the uncoupling of IL-1 β /IL-1R1 signaling, providing a causal link (115) (115) (115) . IL-1 β is a member of the interleukin 1 family of cytokines involved in the inflammatory response that is produced by macrophages, monocytes, and dendritic cells.

Anakinra, the receptor antagonist binding IL-1 β to IL-1R1, represents the treatment of choice in several rheumatic diseases. Based on the in vitro and in vivo findings, Anegelli and co ([116\)](#page-13-0). recently administered Anakinra in two patients with multidrugdependent/-resistant NS and in one patient with post-transplant recurrence of FSGS. They induced a complete and two partial remissions respectively ([116\)](#page-13-0). The limited findings supported the hypothesis that, in proteinuric disease, C3a acts as a second-hit in mediating podocyte cytoskeleton rearrangement and that IL-1R1 blockers may limit the damage ([116](#page-13-0)).

T cells/Interleukins

A generic role of T cells was supported by the observation that supernatants of hybridomes from patients with MCD efface podocytes and induce proteinuria in rats [\(117](#page-13-0)). This finding reinforced the concept that MCD is caused by T-cell disorder ([118\)](#page-13-0) and stimulated many studies aimed to characterize the T cell compartment in iNS, without definitive results. Observational studies documented that concomitant measle infections, which stimulate a Tcell response, are associated with proteinuria reduction in iNS patients ([119\)](#page-13-0). Similar associations were reported during type B influenza ([120](#page-13-0)) and ZIKA virus infections [\(121](#page-13-0)).

T helper 2 cells (Th2)

iNS, in particular MCD, is usually characterized by an increased serum level of several cytokines (IL-4, IL-5, IL-9, IL-10, and IL-13) that may be indirectly related to the activation of T helper 2 cells ([122,](#page-13-0) [123](#page-13-0)). IL13 mRNA is up-regulated in the renal tissue of children with MCD and is associated with increased release of IL13 by $CD3^+$ T cells ([122\)](#page-13-0). Moreover, in vivo experiments support a possible role for IL-13 in iNS. Wistar rats over-expressing IL13 spontaneously develop proteinuria and podocyte effacement, while Minnesota-Buffalo, a strain of rats that spontaneously develop proteinuria, have high levels of IL4

and IL13 that precede proteinuria ([124](#page-13-0)). Anti-IL13 drugs are now available for human administration and have already been reported as effective in the treatment of subjects with asthma ([125](#page-13-0)) and atopic dermatitis [\(126\)](#page-13-0). Given the safety of the drug, a small pilot study testing the effectiveness of anti-IL-13 in MCD would be of interest.

T helper 17 cells (Th17)

IL-17 is produced by T helper 17 cells (Th17) that derive from naïve CD4⁺ after stimulation from IL-6 and IL-13. Previous reports associated high serum levels of IL-17 with glomerulosclerosis in patients with FSGS [\(127](#page-13-0)).

Regulatory T cells (Tregs)

Tregs (CD4+ CD25+ Foxp3+) are a subpopulation of T cells, stimulated by IL-2, that suppress immune response. Tregs are able to inhibit T cell proliferation and cytokine production and play a critical role in preventing autoimmunity. Previous papers reported that circulating Tregs are reduced in children affected by MCD and that proteinuria may be reverted by the infusion of Tregs ([124,](#page-13-0) [128\)](#page-13-0) in several murine models of NS (ie. adriamycin, Buffalo-Mna, and LPS). Moreover, patients affected by IPEX, an immunodeficiency hereditary syndrome with polyendocrinopathy and enteropathy characterized by reduced circulating levels of Tregs due to Foxp3 inactivation, may develop MCD ([129](#page-13-0)). Of interest, the reconstitution of the entire T cell compartment after bone marrow transplantation in children with IPEX recover the general symptoms of the syndrome and also normalize the glomerular disease ([130\)](#page-13-0). Tregs are stimulated and increased by IL-2 ([131,](#page-13-0) [132\)](#page-13-0). While the data above support a protective effect of Tregs in NS, the unique report in children with NS treated with IL2 contradicted this concept. In fact, Bonanni et al. [\(151](#page-13-0)) infused IL2 in five children affected by SRNS who presented lower circulating level of Tregs compared to healthy subjects and obtained the normalization of circulating Tregs without any effect on proteinuria.

B cells

For decades, NS has been considered a T cell pathology based on the efficacy of steroids and on the absence of antibody deposition in glomeruli. Results obtained with anti-CD20 monoclonal antibodies have modified this belief. Starting from 2010, several randomized studies proved the non-inferiority of the chimeric anti-CD20 antibody rituximab compared to steroids and CNI in the treatment of steroid-dependent and resistant NS ([13,](#page-11-0) [14](#page-11-0), [16,](#page-11-0) [152](#page-13-0)–[154\)](#page-14-0). Such clinical results strongly support a possible implication of B cells in this pathology. It was initially proved that the positive effect of rituximab is correlated with the depletory effect on B cells but successive studies have widened the panel of cells expressing CD20 to B memory cells, which are now considered the cell effector of rituximab in terms of length of relapse ([155\)](#page-14-0). Through time-of-flight mass cytometry (CyTOF), it was observed that frequencies of class-switched B cells were significantly higher in patients who received rituximab and

relapsed than in non-relapsing individuals ([133](#page-13-0)), a phenomenon already observed in autoimmune conditions such as myasthenia gravis, neuromyelitis optica, and rheumatoid arthritis ([156](#page-14-0), [157\)](#page-14-0). The phenotype frequently associated with recurrence of NS is IgDCD27+CD38+CD95+ Ab-secreting where the presence of CD38 promotes survival in germinal center B cells. The meaning of B cells and, in particular, of switched B cells expressing CD38 in NS can be interpreted as the persistent possibility of secreting antibodies and should be connected with a potential role of antibodies and, more in general, of immunoglobulins. More observational and experimental evidence of an antibody role in iNS is needed since, with the exception of anti-nephrin antibodies described in MCD [\(20\)](#page-11-0), NS patients do not present immune-deposits in glomeruli. This is an evolving story that started only recently and needs to be developed in detail (see the section dedicated to antibodies below).

Proposed mechanisms for idiopathic nephrotic syndrome (3): antibodies and regulatory immunoglobulins

Antibodies

That auto-antibodies may have a pathological role in iNS is not a novel thought at all. However, the possible pathological relevance of auto-antibodies was further considered mostly after the introduction of treatments with anti-CD20 monoclonal antibodies. In previous literature, several papers proposed many possible circulating antibodies as responsible for the disease (see previous section).

Among others, Musante et al. ([107](#page-12-0)) reported that circulating antiactin/ATP synthase beta chain IgM were present in the serum of around 10% of FSGS patients. Moreover, authors described that infusion of such antibodies in Sprague Dawley rats induced proteinuria, which correlated with IgM glomerular deposition. In 2014, Delville et al. ([109\)](#page-13-0) proposed a selection of circulating antibodies with several antigens as targets as biomarkers of posttransplant recurrence FSGS: protein tyrosine phosphatase receptor O, TNF receptor superfamily member 6, Chorionic gonadotropin β , Ribonucleoprotein B, Apoliprotein 2, P2Y purinoceptor 11, Retinoid orphan nuclear receptor a, Chemokine C-C motif-ligand 19, Myosin light kinase, and CD40 [\(109](#page-13-0)). Antibody anti-CD40 was deeply investigated. In vitro studies showed that podocytes express CD40 and that treatment with antibody anti-CD40 resulted in morphologic alterations. Moreover, antibody anti-CD40 induced proteinuria when administered in wild type mice ([75](#page-12-0)). More recently, Jamin et al. [\(158\)](#page-14-0) identified anti-Ubiquitin Carboxyl-Terminal Hydrolase L1 (UCHL1) IgG in patients with relapsing proteinuria and Ye et al. ([108\)](#page-13-0) described anti-Annexin A2 antibodies in patients with NS with IgG deposits along with GBM. The most recent study by Watts et al. [\(20\)](#page-11-0) described the presence of circulating anti-nephrin IgG in patients with MCD and also demonstrated their binding to podocytes' cytoplasm and co-localization with nephrin. Moreover, serum levels of circulating anti-nephrin antibodies correlated with proteinuria and disappeared after remission, suggesting a direct pathogenetic implication.

Overall, available evidence suggests the existence of circulating anti-podocyte antibodies that may characterize different subsets of NS. However, with the exception of anti-nephrin IgG, other studies did not show the concomitance of circulating antibodies with their deposition in glomeruli.

Furthermore, the limited efficacy of anti-CD20 monoclonal antibodies in inducing complete remission, in particular in SRNS, suggests the need for caution and requires more evidence on the pathological role of such antibodies.

Regulatory immunoglobulins

Immunoglobulins may play an important role in NS, with pathological mechanisms different from the classical immune deposits of glomerulonephritis. Previous studies demonstrated that IgM may deposit on the surface of T cells. After deposition, sialylated IgM are internalized differently to non-sialylated IgM, which remain on surface. The internalization of sialylated IgM correlate with T cells proliferation and confer more resistance to steroids [\(141\)](#page-13-0). Moreover, internalization of IgM and T cells' proliferation are both associated with relapse of proteinuria in patients with SDNS ([110\)](#page-13-0). Rituximab seems to have more effect in decreasing the sialylated IgM; therefore we may speculate that the efficacy of rituximab in SSNS may be partially explained by selective depletion of sialylated IgM on T cells.

Therapies for nephrotic syndrome

A detailed analysis and comparison of therapies administered in NS is outside the scope of this review. However, we here present an overview of the therapeutical progresses of the last two decades, with the aim to define how the pathological progresses have been accompanied by therapeutical ones.

Corticosteroids

Steroids still represent the treatment of reference for iNS in case of occurrence and relapses. Based on the KDIGO Guidelines ([159\)](#page-14-0), prednisone at a dose of 60 mg/m ([2](#page-10-0)) for 6 weeks following 6 weeks at 40 mg/m ([2](#page-10-0)) is associated with the best outcome in terms of early relapses and lower incidence of development of SDNS. The administration of different type of corticosteroids, such as methylprednisolone or deflazacort, is not associated with better outcome compared to the scheme reported above. Relapse of iNS is treated with prednisone 40 mg/m ([2](#page-10-0)) every day until normalization of proteinuria and then the same dose given every other day for 40 days. Longer schemes do not improve the outcome in terms of relapse of proteinuria.

Steroid-sparing agents

Long-term complications related to the chronic administration of steroids in SDNS are commonly reported. Therefore, steroidsparing agents (levamisole, cyclophosphamide, alkylating agents, calcinerin inhibitors, mycophenolate mofetil, and rituximab) are administered with the aim to limit the overall steroid dose [\(17,](#page-11-0) [160\)](#page-14-0). Previous randomized clinical studies showed similar efficacy among the different steroid-sparing agents [\(13](#page-11-0), [14,](#page-11-0) [152](#page-13-0), [153](#page-13-0), [161,](#page-14-0) [162](#page-14-0)). Therefore, the choice may depend on the evaluation of possible side effects and safety [\(126](#page-13-0), [127\)](#page-13-0) and on practical aspects. As an example, a single infusion of rituximab allows to avoid the daily administration of therapy. However, several concerns still remain open on rituximab, such as the missing definition of a definitive cumulative dose: recent studies compared the efficacy of different rituximab courses, demonstrating that SDNS patients receiving rituximab for the third or fourth episode of NS relapse had longer remission than patients receiving rituximab for the first release [\(163](#page-14-0)). Moreover, circulating levels of memory B cells better correlate with NS remission [\(155](#page-14-0)) than total B cells and may represent a potential marker for defining the needing of further rituximab infusion.

Also, the development of anti-rituximab antibodies, due to the chimeric nature of these antibodies, was proposed as a factor limiting the efficacy of rituximab [\(164\)](#page-14-0). In our experience, in a significant cohort of NS patients who received at least two rituximab infusions, the development of circulating anti-rituximab antibodies did not affect the response to the treatment [\(165](#page-14-0)).

Multidrug dependence

A subset of patients with NS develop multidrug dependence (MDNS), defined as the need of more than one drug, usually calcinerin inhibitors and micophenolate mofetyl, to keep remission. MDNS is characterized by frequent relapses of NS and may evolve to MRNS that in extreme conditions, however, a specific therapeutical approach for these patients has not yet been proposed.

Multidrug resistance

Currently, the treatment of patients affected by multidrug resistance mostly consist of anti-proteinuric therapies, such as mineral corticoid antagonists (finrenone) and/or drugs that influence the angiotensin and the endothelin renal accomodation ([166,](#page-14-0) [167](#page-14-0)). The main aim of such conservative therapies is to limit the progression to end-stage renal disease. Therefore, MRNS represents one of the most relevant unmet needs in nephrology. A second generation of humanized anti-CD20 monoclonal antibodies, such as ofatumumab ([154\)](#page-14-0) or obinutuzumab [\(168](#page-14-0)), or therapeutical schemes based on the association of different biologics, have been recently proposed [\(169](#page-14-0)).

Conclusions

Concrete discoveries

In the last two decades, in the context of childhood NS, the main findings were represented by the definition of a genetic basis of mendelian forms and by the demonstrated efficacy of anti-CD20 monoclonal antibodies in SDNS.

When monogenic NS is promptly diagnosed, the therapeutical approach requires the administration of antiproteinuric treatments, limiting therefore the immunosuppressive drugs with relative adverse events. Very recently, cyclosporin was demonstrated to be effective in reducing proteinuria in selected patients with monogenic NS [\(38\)](#page-11-0).

Anti-CD20 monoclonal antibodies have modified the therapeutical approach to SDNS with some advantages over previous drugs. An important consequence of this finding is the strong suggestion that B cells may be implicated in iNS. Such findings provided the bases for new studies on the characterization of B cell subsets in iNS.

Urgent unmet needs

The most urgent unmet need is the development of effective therapies in MRNS and in post-transplant NS recurrence. The definition of the pathological mechanisms causing both drug sensitive and resistant iNS remain fundamental and should precede any therapeutic innovation. The efficacy of anti-CD20 has stimulated the research on antibodies and on regulatory immunoglobulins in iNS. Previous data highlighted the existence of circulating IgM with variable charge and isoforms with cationic pI. Whether cationic IgM play a role in iNS and how they are modified by anti-CD20 drugs is a matter of research. Another point of interest is the need to better characterize the amount of immunoglobulins that escape the anti-CD20 effect. New anti-CD20 monoclonal antibodies may have more efficacy in the treatment of MRNS and post-transplant recurrence NS.

The association of anti-CD20 with other monoclonal antibodies targeting long-lived cells producing antibodies, such as plasma cells

Vincenti et al. 10.3389/fi[mmu.2023.1167741](https://doi.org/10.3389/fimmu.2023.1167741)

that expressed CD38, represent the proof-of-concept for an ongoing Phase II study in MDNS, MRNS, and post-transplant recurrence NS (NCT05704400).

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Funding

AA is supported by 'Ricerca Corrente 2023' and '5XMille-5M-2019-23680431'.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/](https://www.frontiersin.org/articles/10.3389/fimmu.2023.1167741/full#supplementary-material)fimmu.2023.1167741/ [full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fimmu.2023.1167741/full#supplementary-material)

References

1. Eddy AA, Symons JM. Nephrotic syndrome in childhood. Lancet (2003) 362 (9384):629–39. doi: [10.1016/S0140-6736\(03\)14184-0](https://doi.org/10.1016/S0140-6736(03)14184-0)

2. Trautmann A, Bodria M, Ozaltin F, Gheisari A, Melk A, Azocar M, et al. Spectrum of steroid-resistant and congenital nephrotic syndrome in children: the PodoNet registry cohort. Clin J Am Soc Nephrol. (2015) 10(4):592–600. doi: [10.2215/](https://doi.org/10.2215/CJN.06260614) CIN.06260614

3. D'Agati V. Pathologic classification of focal segmental glomerulosclerosis. Semin Nephrol. (2003) 23(2):117–34. doi: [10.1053/snep.2003.50012](https://doi.org/10.1053/snep.2003.50012)

4. Maas RJ, Deegens JK, Smeets B, Moeller MJ, Wetzels JF. Minimal change disease and idiopathic FSGS: manifestations of the same disease. Nat Rev Nephrol. (2016) 12 (12):768–76. doi: [10.1038/nrneph.2016.147](https://doi.org/10.1038/nrneph.2016.147)

5. Saleem MA. Molecular stratification of idiopathic nephrotic syndrome. Nat Rev Nephrol. (2019) 15(12):750–65. doi: [10.1038/s41581-019-0217-5](https://doi.org/10.1038/s41581-019-0217-5)

6. Tareyeva IE, Nikolaev AJ, Janushkevitch TN. Nephrotic syndrome induced by insect sting. Lancet (1982) 2(8302):825. doi: [10.1016/S0140-6736\(82\)92718-0](https://doi.org/10.1016/S0140-6736(82)92718-0)

7. Angeletti A, Drovandi S, Sanguineri F, Santaniello M, Ferrando G, Forno R, et al. COVID-19 in children with nephrotic syndrome on anti-CD20 chronic immunosuppression. Clin J Am Soc Nephrol. (2020) 15(10):1494–5. doi: [10.2215/](https://doi.org/10.2215/CJN.06400420) [CJN.06400420](https://doi.org/10.2215/CJN.06400420)

8. Groopman EE, Rasouly HM, Gharavi AG. Genomic medicine for kidney disease. Nat Rev Nephrol. (2018) 14(2):83–104. doi: [10.1038/nrneph.2017.167](https://doi.org/10.1038/nrneph.2017.167)

9. Radhakrishnan J, Cattran DC. The KDIGO practice guideline on glomerulonephritis: reading between the (guide)lines–application to the individual patient. Kidney Int (2012) 82(8):840–56. doi: [10.1038/ki.2012.280](https://doi.org/10.1038/ki.2012.280)

10. Chapter 3: steroid-sensitive nephrotic syndrome in children. Kidney Int Suppl (2011) (2012) 2(2):163–71.

11. Chapter 4: steroid-resistant nephrotic syndrome in children. Kidney Int Suppl (2011) (2012) 2(2):172–6.

12. Trautmann A, Boyer O, Hodson E, Bagga A, Gipson DS, Samuel S, et al. IPNA clinical practice recommendations for the diagnosis and management of children with steroid-sensitive nephrotic syndrome. Pediatr Nephrol. (2023) 38(3):877–919. doi: [10.1007/s00467-022-05739-3](https://doi.org/10.1007/s00467-022-05739-3)

13. Ravani P, Rossi R, Bonanni A, Quinn RR, Sica F, Bodria M, et al. Rituximab in children with steroid-dependent nephrotic syndrome: a multicenter, open-label, noninferiority, randomized controlled trial. J Am Soc Nephrol. (2015) 26(9):2259–66. doi: [10.1681/ASN.2014080799](https://doi.org/10.1681/ASN.2014080799)

14. Iijima K, Sako M, Nozu K, Mori R, Tuchida N, Kamei K, et al. Rituximab for childhood-onset, complicated, frequently relapsing nephrotic syndrome or steroiddependent nephrotic syndrome: a multicentre, double-blind, randomised, placebocontrolled trial. Lancet (2014) 384(9950):1273–81. doi: [10.1016/S0140-6736\(14\)60541-](https://doi.org/10.1016/S0140-6736(14)60541-9) [9](https://doi.org/10.1016/S0140-6736(14)60541-9)

15. Ravani P, Ponticelli A, Siciliano C, Fornoni A, Magnasco A, Sica F, et al. Rituximab is a safe and effective long-term treatment for children with steroid and calcineurin inhibitor-dependent idiopathic nephrotic syndrome. Kidney Int (2013) 84 (5):1025–33. doi: [10.1038/ki.2013.211](https://doi.org/10.1038/ki.2013.211)

16. Magnasco A, Ravani P, Edefonti A, Murer L, Ghio L, Belingheri M, et al. Rituximab in children with resistant idiopathic nephrotic syndrome. J Am Soc Nephrol. (2012) 23(6):1117–24. doi: [10.1681/ASN.2011080775](https://doi.org/10.1681/ASN.2011080775)

17. Tullus K, Webb H, Bagga A. Management of steroid-resistant nephrotic syndrome in children and adolescents. Lancet Child Adolesc Health (2018) 2 (12):880–90. doi: [10.1016/S2352-4642\(18\)30283-9](https://doi.org/10.1016/S2352-4642(18)30283-9)

18. Connaughton DM, Kennedy C, Shril S, Mann N, Murray SL, Williams PA, et al. Monogenic causes of chronic kidney disease in adults. Kidney Int (2019) 95(4):914–28. doi: [10.1016/j.kint.2018.10.031](https://doi.org/10.1016/j.kint.2018.10.031)

19. D'Agati VD, Mengel M. The rise of renal pathology in nephrology: structure illuminates function. Am J Kidney Dis (2013) 61(6):1016–25. doi: [10.1053/](https://doi.org/10.1053/j.ajkd.2012.12.019) [j.ajkd.2012.12.019](https://doi.org/10.1053/j.ajkd.2012.12.019)

20. Watts AJB, Keller KH, Lerner G, Rosales I, Collins AB, Sekulic M, et al. Discovery of autoantibodies targeting nephrin in minimal change disease supports a novel autoimmune etiology. J Am Soc Nephrol. (2022) 33(1):238–52. doi: [10.1681/](https://doi.org/10.1681/ASN.2021060794) [ASN.2021060794](https://doi.org/10.1681/ASN.2021060794)

21. Savige J, Harraka P. Pathogenic variants in the genes affected in alport syndrome (COL4A3-COL4A5) and their association with other kidney conditions: a review. Am J Kidney Dis (2021) 78(6):857–64. doi: [10.1053/j.ajkd.2021.04.017](https://doi.org/10.1053/j.ajkd.2021.04.017)

22. Warejko JK, Tan W, Daga A, Schapiro D, Lawson JA, Shril S, et al. Whole exome sequencing of patients with steroid-resistant nephrotic syndrome. Clin J Am Soc Nephrol. (2018) 13(1):53–62. doi: [10.2215/CJN.04120417](https://doi.org/10.2215/CJN.04120417)

23. Fang Z, Zhang C, Jin Y, Tong J, Liu J, Hao X, et al. Adult-onset focal segmental glomerulosclerosis with steroid-dependent nephrotic syndrome caused by a novel TBC1D8B variant: a case report and literature review. Am J Kidney Dis (2023) 81 (2):240–4. doi: [10.1053/j.ajkd.2022.06.012](https://doi.org/10.1053/j.ajkd.2022.06.012)

24. Weng PL, Majmundar AJ, Khan K, Lim TY, Shril S, Jin G, et al. De novo TRIM8 variants impair its protein localization to nuclear bodies and cause developmental delay, epilepsy, and focal segmental glomerulosclerosis. Am J Hum Genet (2021) 108 (2):357–67. doi: [10.1016/j.ajhg.2021.01.008](https://doi.org/10.1016/j.ajhg.2021.01.008)

25. Drovandi S, Lugani F, Boyer O, La Porta E, Giordano P, Hummel A, et al. Multicentric carpotarsal osteolysis syndrome associated nephropathy: novel variants of MAFB gene and literature review. J Clin Med (2022) 11(15). doi: [10.3390/jcm11154423](https://doi.org/10.3390/jcm11154423)

26. Treimer E, Kalayci T, Schumann S, Suer I, Greco S, Schanze D, et al. Functional characterization of a novel TP53RK mutation identified in a family with Gallowaymowat syndrome. Hum Mutat (2022) 43(12):1866–71. doi: [10.1002/humu.24472](https://doi.org/10.1002/humu.24472)

27. Ashraf S, Kudo H, Rao J, Kikuchi A, Widmeier E, Lawson JA, et al. Mutations in six nephrosis genes delineate a pathogenic pathway amenable to treatment. Nat Commun (2018) 9(1):1960. doi: [10.1038/s41467-018-04193-w](https://doi.org/10.1038/s41467-018-04193-w)

28. Assady S, Wanner N, Skorecki KL, Huber TB. New insights into podocyte biology in glomerular health and disease. J Am Soc Nephrol. (2017) 28(6):1707–15. doi: [10.1681/ASN.2017010027](https://doi.org/10.1681/ASN.2017010027)

29. Caridi G, Bertelli R, Carrea A, Di Duca M, Catarsi P, Artero M, et al. Prevalence, genetics, and clinical features of patients carrying podocin mutations in steroidresistant nonfamilial focal segmental glomerulosclerosis. J Am Soc Nephrol. (2001) 12 (12):2742–6. doi: [10.1681/ASN.V12122742](https://doi.org/10.1681/ASN.V12122742)

30. Caridi G, Bertelli R, Di Duca M, Dagnino M, Emma F, Onetti Muda A, et al. Broadening the spectrum of diseases related to podocin mutations. J Am Soc Nephrol. (2003) 14(5):1278–86. doi: [10.1097/01.ASN.0000060578.79050.E0](https://doi.org/10.1097/01.ASN.0000060578.79050.E0)

31. Kaplan JM, Kim SH, North KN, Rennke H, Correia LA, Tong HQ, et al. Mutations in ACTN4, encoding alpha-actinin-4, cause familial focal segmental glomerulosclerosis. Nat Genet (2000) 24(3):251–6. doi: [10.1038/73456](https://doi.org/10.1038/73456)

32. Liu PJ, Gunther LK, Garone ME, Zhang C, Perez D, Bi-Karchin J, et al. Steroidresistant nephrotic syndrome-associated <i<MYO1E</i< mutations have differential effects on myosin 1e localization, dynamics, and activity. J Am Soc Nephrol. (2022) 33(11):1989–2007. doi: [10.1681/ASN.2021111505](https://doi.org/10.1681/ASN.2021111505)

33. Boyer O, Nevo F, Plaisier E, Funalot B, Gribouval O, Benoit G, et al. INF2 mutations in charcot-Marie-Tooth disease with glomerulopathy. N Engl J Med (2011) 365(25):2377–88. doi: [10.1056/NEJMoa1109122](https://doi.org/10.1056/NEJMoa1109122)

34. Reiser J, Polu KR, Moller CC, Kenlan P, Altintas MM, Wei C, et al. TRPC6 is a glomerular slit diaphragm-associated channel required for normal renal function. *Nat*
Genet (2005) 37(7):739–44. doi: [10.1038/ng1592](https://doi.org/10.1038/ng1592)

35. Pollak MR. Expanding the spectrum of NPHS1-associated disease. Kidney Int (2009) 76(12):1221–3. doi: [10.1038/ki.2009.391](https://doi.org/10.1038/ki.2009.391)

36. Labat-de-Hoz L, Alonso MA. Formins in human disease. Cells (2021) 10 (10):2554. doi: [10.3390/cells10102554](https://doi.org/10.3390/cells10102554)

37. Drovandi S, Lipska-Zietkiewicz BS, Ozaltin F, Emma F, Gulhan B, Boyer O, et al. Oral coenzyme Q10 supplementation leads to better preservation of kidney function in steroid-resistant nephrotic syndrome due to primary coenzyme Q10 deficiency. Kidney Int (2022) 102(3):604–12. doi: [10.1016/j.kint.2022.04.029](https://doi.org/10.1016/j.kint.2022.04.029)

38. Malakasioti G, Iancu D, Milovanova A, Tsygin A, Horinouchi T, Nagano C, et al. A multicenter retrospective study of calcineurin inhibitors in nephrotic syndrome secondary to podocyte gene variants. Kidney Int (2023) 103(5):962–72. doi: [10.1016/](https://doi.org/10.1016/j.kint.2023.02.022) [j.kint.2023.02.022](https://doi.org/10.1016/j.kint.2023.02.022)

39. Tardi NJ, Reiser J. The use of sildenafil for glomerular disease. J Am Soc Nephrol. (2017) 28(5):1329–31. doi: [10.1681/ASN.2017020171](https://doi.org/10.1681/ASN.2017020171)

40. Sonneveld R, Hoenderop JG, Isidori AM, Henique C, Dijkman HB, Berden JH, et al. Sildenafil prevents podocyte injury via PPAR-gamma-Mediated TRPC6 inhibition. J Am Soc Nephrol. (2017) 28(5):1491–505. doi: [10.1681/ASN.2015080885](https://doi.org/10.1681/ASN.2015080885)

41. White RH. The familial nephrotic syndrome. i. a European survey. Clin Nephrol. (1973) 1(4):215–9.

42. Debiec H, Dossier C, Letouze E, Gillies CE, Vivarelli M, Putler RK, et al. Transethnic, genome-wide analysis reveals immune-related risk alleles and phenotypic correlates in pediatric steroid-sensitive nephrotic syndrome. J Am Soc Nephrol. (2018) 29(7):2000–13. doi: [10.1681/ASN.2017111185](https://doi.org/10.1681/ASN.2017111185)

43. Jia X, Horinouchi T, Hitomi Y, Shono A, Khor SS, Omae Y, et al. Strong association of the HLA-DR/DQ locus with childhood steroid-sensitive nephrotic syndrome in the Japanese population. J Am Soc Nephrol. (2018) 29(8):2189-99. doi: [10.1681/ASN.2017080859](https://doi.org/10.1681/ASN.2017080859)

44. Jia X, Yamamura T, Gbadegesin R, McNulty MT, Song K, Nagano C, et al. Common risk variants in NPHS1 and TNFSF15 are associated with childhood steroidsensitive nephrotic syndrome. Kidney Int (2020) 98(5):1308–22. doi: [10.1016/](https://doi.org/10.1016/j.kint.2020.05.029) [j.kint.2020.05.029](https://doi.org/10.1016/j.kint.2020.05.029)

45. Parsa A, Kao WH, Xie D, Astor BC, Li M, Hsu CY, et al. APOL1 risk variants, race, and progression of chronic kidney disease. N Engl J Med (2013) 369(23):2183–96. doi: [10.1056/NEJMoa1310345](https://doi.org/10.1056/NEJMoa1310345)

46. Morello W, Mastrangelo A, Guzzo I, Cusinato L, Petruzzelli Annicchiarico L, Benevenuta C, et al. Prevalence of SARS-CoV-2-IgG antibodies in children with CKD or immunosuppression. Clin J Am Soc Nephrol. (2021) 16(7):1097–9. doi: [10.2215/](https://doi.org/10.2215/CJN.00330121) [CJN.00330121](https://doi.org/10.2215/CJN.00330121)

47. Cohen SD, Kopp JB, Kimmel PL. Kidney diseases associated with human immunodeficiency virus infection. N Engl J Med (2017) 377(24):2363–74. doi: [10.1056/NEJMra1508467](https://doi.org/10.1056/NEJMra1508467)

48. Mallipattu SK, Salem F, Wyatt CM. The changing epidemiology of HIV-related chronic kidney disease in the era of antiretroviral therapy. Kidney Int (2014) 86(2):259-65. doi: [10.1038/ki.2014.44](https://doi.org/10.1038/ki.2014.44)

49. Rednor SJ, Ross MJ. Molecular mechanisms of injury in HIV-associated nephropathy. Front Med (Lausanne). (2018) 5:177. doi: [10.3389/fmed.2018.00177](https://doi.org/10.3389/fmed.2018.00177)

50. Kaufman L, Yang G, Hayashi K, Ashby JR, Huang L, Ross MJ, et al. The homophilic adhesion molecule sidekick-1 contributes to augmented podocyte aggregation in HIV-associated nephropathy. FASEB J (2007) 21(7):1367–75. doi: [10.1096/fj.06-7191com](https://doi.org/10.1096/fj.06-7191com)

51. Kaufman L, Hayashi K, Ross MJ, Ross MD, Klotman PE. Sidekick-1 is upregulated in glomeruli in HIV-associated nephropathy. J Am Soc Nephrol. (2004) 15(7):1721–30. doi: [10.1097/01.ASN.0000128975.28958.C2](https://doi.org/10.1097/01.ASN.0000128975.28958.C2)

52. Thomson R, Finkelstein A. Human trypanolytic factor APOL1 forms pH-gated cation-selective channels in planar lipid bilayers: relevance to trypanosome lysis. Proc Natl Acad Sci USA (2015) 112(9):2894–9. doi: [10.1073/pnas.1421953112](https://doi.org/10.1073/pnas.1421953112)

53. Tanawattanacharoen S, Falk RJ, Jennette JC, Kopp JB. Parvovirus B19 DNA in kidney tissue of patients with focal segmental glomerulosclerosis. Am J Kidney Dis (2000) 35(6):1166–74. doi: [10.1016/S0272-6386\(00\)70055-2](https://doi.org/10.1016/S0272-6386(00)70055-2)

54. Markowitz GS, Appel GB, Fine PL, Fenves AZ, Loon NR, Jagannath S, et al. Collapsing focal segmental glomerulosclerosis following treatment with high-dose pamidronate. J Am Soc Nephrol. (2001) 12(6):1164–72. doi: [10.1681/ASN.V1261164](https://doi.org/10.1681/ASN.V1261164)

55. Petersen CE, Amaral S, Frosch E. Lithium-induced nephrotic syndrome in a prepubertal boy. J Child Adolesc Psychopharmacol (2008) 18(2):210–3. doi: [10.1089/](https://doi.org/10.1089/cap.2007.0118) [cap.2007.0118](https://doi.org/10.1089/cap.2007.0118)

56. Gong R, Wang P, Dworkin L. What we need to know about the effect of lithium on the kidney. Am J Physiol Renal Physiol (2016) 311(6):F1168–71. doi: [10.1152/](https://doi.org/10.1152/ajprenal.00145.2016) [ajprenal.00145.2016](https://doi.org/10.1152/ajprenal.00145.2016)

57. Puelles VG, van der Wolde JW, Wanner N, Scheppach MW, Cullen-McEwen LA, Bork T, et al. mTOR-mediated podocyte hypertrophy regulates glomerular integrity in mice and humans. JCI Insight (2019) 4(18). doi: [10.1172/jci.insight.99271](https://doi.org/10.1172/jci.insight.99271)

58. Letavernier E, Bruneval P, Mandet C, Duong Van Huyen JP, Peraldi MN, Helal I, et al. High sirolimus levels may induce focal segmental glomerulosclerosis de novo. Clin J Am Soc Nephrol. (2007) 2(2):326–33. doi: [10.2215/CJN.03751106](https://doi.org/10.2215/CJN.03751106)

59. Nobakht N, Nguyen HA, Kamgar MK, Abdelnour L, Rastogi A, Hanna RM. Development of collapsing focal and segmental glomerulosclerosis after receiving intravitreal vascular endothelial growth factor blockade. Kidney Int Rep (2019) 4 (10):1508–12. doi: [10.1016/j.ekir.2019.07.019](https://doi.org/10.1016/j.ekir.2019.07.019)

60. Jhaveri KD, Wanchoo R, Sakhiya V, Ross DW, Fishbane S. Adverse renal effects of novel molecular oncologic targeted therapies: a narrative review. Kidney Int Rep (2017) 2(1):108–23. doi: [10.1016/j.ekir.2016.09.055](https://doi.org/10.1016/j.ekir.2016.09.055)

61. Byrd JC, Furman RR, Coutre SE, Flinn IW, Burger JA, Blum KA, et al. Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. N Engl J Med (2013) 369 (1):32–42. doi: [10.1056/NEJMoa1215637](https://doi.org/10.1056/NEJMoa1215637)

62. Genovese G, Friedman DJ, Ross MD, Lecordier L, Uzureau P, Freedman BI, et al. Association of trypanolytic ApoL1 variants with kidney disease in African americans. Science (2010) 329(5993):841–5. doi: [10.1126/science.1193032](https://doi.org/10.1126/science.1193032)

63. Beckerman P, Bi-Karchin J, Park AS, Qiu C, Dummer PD, Soomro I, et al. Transgenic expression of human APOL1 risk variants in podocytes induces kidney disease in mice. Nat Med (2017) 23(4):429–38. doi: [10.1038/nm.4287](https://doi.org/10.1038/nm.4287)

64. Gulati A, Sinha A, Sreenivas V, Math A, Hari P, Bagga A. Daily corticosteroids reduce infection-associated relapses in frequently relapsing nephrotic syndrome: a randomized controlled trial. Clin J Am Soc Nephrol. (2011) 6(1):63–9. doi: [10.2215/](https://doi.org/10.2215/CJN.01850310) CIN.01850310

65. Jain N, Keating M, Thompson P, Ferrajoli A, Burger J, Borthakur G, et al. Ibrutinib and venetoclax for first-line treatment of CLL. N Engl J Med (2019) 380 (22):2095–103. doi: [10.1056/NEJMoa1900574](https://doi.org/10.1056/NEJMoa1900574)

66. Ponader S, Chen SS, Buggy JJ, Balakrishnan K, Gandhi V, Wierda WG, et al. The bruton tyrosine kinase inhibitor PCI-32765 thwarts chronic lymphocytic leukemia cell survival and tissue homing in vitro and in vivo. Blood (2012) 119(5):1182–9. doi: [10.1182/blood-2011-10-386417](https://doi.org/10.1182/blood-2011-10-386417)

67. Musante L, Candiano G, Bruschi M, Zennaro C, Carraro M, Artero M, et al. Characterization of plasma factors that alter the permeability to albumin within isolated glomeruli. Proteomics (2002) 2(2):197–205. doi: [10.1002/1615-9861\(200202\)](https://doi.org/10.1002/1615-9861(200202)2:2%3C197::AID-PROT197%3E3.0.CO;2-6) [2:2<197::AID-PROT197>3.0.CO;2-6](https://doi.org/10.1002/1615-9861(200202)2:2%3C197::AID-PROT197%3E3.0.CO;2-6)

68. Savin VJ, Sharma R, Sharma M, McCarthy ET, Swan SK, Ellis E, et al. Circulating factor associated with increased glomerular permeability to albumin in recurrent focal segmental glomerulosclerosis. N Engl J Med (1996) 334(14):878-83. doi: [10.1056/NEJM199604043341402](https://doi.org/10.1056/NEJM199604043341402)

69. Vincenti F, Ghiggeri GM. New insights into the pathogenesis and the therapy of recurrent focal glomerulosclerosis. Am J Transplant. (2005) 5(6):1179–85. doi: [10.1111/](https://doi.org/10.1111/j.1600-6143.2005.00968.x) [j.1600-6143.2005.00968.x](https://doi.org/10.1111/j.1600-6143.2005.00968.x)

70. Dantal J, Baatard R, Hourmant M, Cantarovich D, Buzelin F, Soulillou JP. Recurrent nephrotic syndrome following renal transplantation in patients with focal glomerulosclerosis. a one-center study of plasma exchange effects. Transplantation (1991) 52(5):827–31. doi: [10.1097/00007890-199111000-00014](https://doi.org/10.1097/00007890-199111000-00014)

71. Dantal J, Bigot E, Bogers W, Testa A, Kriaa F, Jacques Y, et al. Effect of plasma protein adsorption on protein excretion in kidney-transplant recipients with recurrent nephrotic syndrome. N Engl J Med (1994) 330(1):7–14. doi: [10.1056/](https://doi.org/10.1056/NEJM199401063300102) [NEJM199401063300102](https://doi.org/10.1056/NEJM199401063300102)

72. Gallon L, Leventhal J, Skaro A, Kanwar Y, Alvarado A. Resolution of recurrent focal segmental glomerulosclerosis after retransplantation. N Engl J Med (2012) 366 (17):1648–9. doi: [10.1056/NEJMc1202500](https://doi.org/10.1056/NEJMc1202500)

73. McCarthy ET, Sharma M, Savin VJ. Circulating permeability factors in idiopathic nephrotic syndrome and focal segmental glomerulosclerosis. *Clin J Am*
S*oc Nephrol*. (2010) 5(11):2115–21. doi: [10.2215/CJN.03800609](https://doi.org/10.2215/CJN.03800609)

74. Kairaitis L, Wang Y, Zheng L, Tay YC, Wang Y, Harris DC. Blockade of CD40- CD40 ligand protects against renal injury in chronic proteinuric renal disease. *Kidney*
I*nt* (2003) 64(4):1265–72. doi: [10.1046/j.1523-1755.2003.00223.x](https://doi.org/10.1046/j.1523-1755.2003.00223.x)

75. Doublier S, Zennaro C, Musante L, Spatola T, Candiano G, Bruschi M, et al. Soluble CD40 ligand directly alters glomerular permeability and may act as a
circulating permeability factor in FSGS. *PloS One* (2017) 12(11):e0188045. doi: [10.1371/journal.pone.0188045](https://doi.org/10.1371/journal.pone.0188045)

76. Hassan GS, Merhi Y, Mourad W. CD40 ligand: a neo-inflammatory molecule in vascular diseases. Immunobiology (2012) 217(5):521–32. doi: [10.1016/](https://doi.org/10.1016/j.imbio.2011.03.010) [j.imbio.2011.03.010](https://doi.org/10.1016/j.imbio.2011.03.010)

77. Quezada SA, Jarvinen LZ, Lind EF, Noelle RJ. CD40/CD154 interactions at the interface of tolerance and immunity. Annu Rev Immunol (2004) 22:307–28. doi: [10.1146/annurev.immunol.22.012703.104533](https://doi.org/10.1146/annurev.immunol.22.012703.104533)

78. Delville M, Baye E, Durrbach A, Audard V, Kofman T, Braun L, et al. B7-1 blockade does not improve post-transplant nephrotic syndrome caused by recurrent FSGS. J Am Soc Nephrol. (2016) 27(8):2520–7. doi: [10.1681/ASN.2015091002](https://doi.org/10.1681/ASN.2015091002)

79. Ivy SP, Wick JY, Kaufman BM. An overview of small-molecule inhibitors of VEGFR signaling. Nat Rev Clin Oncol (2009) 6(10):569–79. doi: [10.1038/](https://doi.org/10.1038/nrclinonc.2009.130) [nrclinonc.2009.130](https://doi.org/10.1038/nrclinonc.2009.130)

80. Izzedine H, Mangier M, Ory V, Zhang SY, Sendeyo K, Bouachi K, et al. Expression patterns of RelA and c-mip are associated with different glomerular diseases following anti-VEGF therapy. Kidney Int (2014) 85(2):457–70. doi: [10.1038/](https://doi.org/10.1038/ki.2013.344) [ki.2013.344](https://doi.org/10.1038/ki.2013.344)

81. Echeverria V, Burgess S, Gamble-George J, Zeitlin R, Lin X, Cao C, et al. Sorafenib inhibits nuclear factor kappa b, decreases inducible nitric oxide synthase and cyclooxygenase-2 expression, and restores working memory in APPswe mice. Neuroscience (2009) 162(4):1220–31. doi: [10.1016/j.neuroscience.2009.05.019](https://doi.org/10.1016/j.neuroscience.2009.05.019)

82. Zhang SY, Kamal M, Dahan K, Pawlak A, Ory V, Desvaux D, et al. C-mip impairs podocyte proximal signaling and induces heavy proteinuria. Sci Signal (2010) 3 (122):ra39. doi: [10.1126/scisignal.2000678](https://doi.org/10.1126/scisignal.2000678)

83. Patrakka J, Tryggvason K. Nephrin–a unique structural and signaling protein of the kidney filter. Trends Mol Med (2007) 13(9):396–403. doi: [10.1016/](https://doi.org/10.1016/j.molmed.2007.06.006) [j.molmed.2007.06.006](https://doi.org/10.1016/j.molmed.2007.06.006)

84. Sendeyo K, Audard V, Zhang SY, Fan Q, Bouachi K, Ollero M, et al. Upregulation of c-mip is closely related to podocyte dysfunction in membranous nephropathy. Kidney Int (2013) 83(3):414–25. doi: [10.1038/ki.2012.426](https://doi.org/10.1038/ki.2012.426)

85. Wei C, Moller CC, Altintas MM, Li J, Schwarz K, Zacchigna S, et al. Modification of kidney barrier function by the urokinase receptor. Nat Med (2008) 14(1):55–63. doi: [10.1038/nm1696](https://doi.org/10.1038/nm1696)

86. Wei C, Trachtman H, Li J, Dong C, Friedman AL, Gassman JJ, et al. Circulating suPAR in two cohorts of primary FSGS. J Am Soc Nephrol. (2012) 23(12):2051–9. doi: [10.1681/ASN.2012030302](https://doi.org/10.1681/ASN.2012030302)

87. Hayek SS, Sever S, Ko YA, Trachtman H, Awad M, Wadhwani S, et al. Soluble urokinase receptor and chronic kidney disease. N Engl J Med (2015) 373(20):1916–25. doi: [10.1056/NEJMoa1506362](https://doi.org/10.1056/NEJMoa1506362)

88. Zhu P, Goh YY, Chin HF, Kersten S, Tan NS. Angiopoietin-like 4: a decade of research. Biosci Rep (2012) 32(3):211–9. doi: [10.1042/BSR20110102](https://doi.org/10.1042/BSR20110102)

89. Clement LC, Mace C, Del Nogal Avila M, Marshall CB, Chugh SS. The proteinuria-hypertriglyceridemia connection as a basis for novel therapeutics for nephrotic syndrome. Transl Res (2015) 165(4):499–504. doi: [10.1016/j.trsl.2014.06.004](https://doi.org/10.1016/j.trsl.2014.06.004)

90. Clement LC, Avila-Casado C, Mace C, Soria E, Bakker WW, Kersten S, et al. Podocyte-secreted angiopoietin-like-4 mediates proteinuria in glucocorticoid-sensitive nephrotic syndrome. Nat Med (2011) 17(1):117–22. doi: [10.1038/nm.2261](https://doi.org/10.1038/nm.2261)

91. Chugh SS, Clement LC, Mace C. New insights into human minimal change disease: lessons from animal models. Am J Kidney Dis (2012) 59(2):284–92. doi: [10.1053/j.ajkd.2011.07.024](https://doi.org/10.1053/j.ajkd.2011.07.024)

92. Bakker WW, Borghuis T, Harmsen MC, van den Berg A, Kema IP, Niezen KE, et al. Protease activity of plasma hemopexin. Kidney Int (2005) 68(2):603–10. doi: [10.1111/j.1523-1755.2005.00438.x](https://doi.org/10.1111/j.1523-1755.2005.00438.x)

93. Immenschuh S, Song DX, Satoh H, Muller-Eberhard U. The type II hemopexin interleukin-6 response element predominates the transcriptional regulation of the hemopexin acute phase responsiveness. Biochem Biophys Res Commun (1995) 207 (1):202–8. doi: [10.1006/bbrc.1995.1173](https://doi.org/10.1006/bbrc.1995.1173)

94. Gutteridge JM. Lipid peroxidation and antioxidants as biomarkers of tissue damage. Clin Chem (1995) 41(12 Pt 2):1819–28. doi: [10.1093/clinchem/41.12.1819](https://doi.org/10.1093/clinchem/41.12.1819)

95. Lennon R, Singh A, Welsh GI, Coward RJ, Satchell S, Ni L, et al. Hemopexin induces nephrin-dependent reorganization of the actin cytoskeleton in podocytes. \overline{JAm} Soc Nephrol. (2008) 19(11):2140–9. doi: [10.1681/ASN.2007080940](https://doi.org/10.1681/ASN.2007080940)

96. Bakker WW, van Dael CM, Pierik LJ, van Wijk JA, Nauta J, Borghuis T, et al. Altered activity of plasma hemopexin in patients with minimal change disease in relapse. Pediatr Nephrol. (2005) 20(10):1410–5. doi: [10.1007/s00467-005-1936-3](https://doi.org/10.1007/s00467-005-1936-3)

97. Bridges CR, Myers BD, Brenner BM, Deen WM. Glomerular charge alterations in human minimal change nephropathy. Kidney Int (1982) 22(6):677–84. doi: [10.1038/](https://doi.org/10.1038/ki.1982.229) [ki.1982.229](https://doi.org/10.1038/ki.1982.229)

98. Ghiggeri GM, Candiano G, Ginevri F, Gusmano R, Ciardi MR, Perfumo F, et al. Renal selectivity properties towards endogenous albumin in minimal change nephropathy. Kidney Int (1987) 32(1):69–77. doi: [10.1038/ki.1987.173](https://doi.org/10.1038/ki.1987.173)

99. Ghiggeri GM, Ginevri F, Candiano G, Oleggini R, Perfumo F, Queirolo C, et al. Characterization of cationic albumin in minimal change nephropathy. Kidney Int (1987) 32(4):547–53. doi: [10.1038/ki.1987.243](https://doi.org/10.1038/ki.1987.243)

100. Yu CC, Fornoni A, Weins A, Hakroush S, Maiguel D, Sageshima J, et al. Abatacept in B7-1-positive proteinuric kidney disease. N Engl J Med (2013) 369 (25):2416–23. doi: [10.1056/NEJMoa1304572](https://doi.org/10.1056/NEJMoa1304572)

101. Burke GW 3rd, Chandar J, Sageshima J, Ortigosa-Goggins M, Amarapurkar P, Mitrofanova A, et al. Benefit of B7-1 staining and abatacept for treatment-resistant post-transplant focal segmental glomerulosclerosis in a predominantly pediatric cohort: time for a reappraisal. Pediatr Nephrol. (2023) 38(1):145–59. doi: [10.1007/](https://doi.org/10.1007/s00467-022-05549-7) [s00467-022-05549-7](https://doi.org/10.1007/s00467-022-05549-7)

102. Garin EH, Diaz LN, Mu W, Wasserfall C, Araya C, Segal M, et al. Urinary CD80 excretion increases in idiopathic minimal-change disease. J Am Soc Nephrol. (2009) 20(2):260–6. doi: [10.1681/ASN.2007080836](https://doi.org/10.1681/ASN.2007080836)

103. Garin EH, Mu W, Arthur JM, Rivard CJ, Araya CE, Shimada M, et al. Urinary CD80 is elevated in minimal change disease but not in focal segmental glomerulosclerosis. Kidney Int (2010) 78(3):296–302. doi: [10.1038/ki.2010.143](https://doi.org/10.1038/ki.2010.143)

104. Fiorina P, Vergani A, Bassi R, Niewczas MA, Altintas MM, Pezzolesi MG, et al. Role of podocyte B7-1 in diabetic nephropathy. J Am Soc Nephrol. (2014) 25(7):1415– 29. doi: [10.1681/ASN.2013050518](https://doi.org/10.1681/ASN.2013050518)

105. Mishra OP, Kumar R, Narayan G, Srivastava P, Abhinay A, Prasad R, et al. Toll-like receptor 3 (TLR-3), TLR-4 and CD80 expression in peripheral blood mononuclear cells and urinary CD80 levels in children with idiopathic nephrotic syndrome. Pediatr Nephrol. (2017) 32(8):1355–61. doi: [10.1007/s00467-017-3613-8](https://doi.org/10.1007/s00467-017-3613-8)

106. Reiser J, von Gersdorff G, Loos M, Oh J, Asanuma K, Giardino L, et al. Induction of B7-1 in podocytes is associated with nephrotic syndrome. J Clin Invest. (2004) 113(10):1390–7. doi: [10.1172/JCI20402](https://doi.org/10.1172/JCI20402)

107. Musante L, Candiano G, Bruschi M, Santucci L, Carnemolla B, Orecchia P, et al. Circulating anti-actin and anti-ATP synthase antibodies identify a sub-set of patients with idiopathic nephrotic syndrome. Clin Exp Immunol (2005) 141(3):491–9. doi: [10.1111/j.1365-2249.2005.02862.x](https://doi.org/10.1111/j.1365-2249.2005.02862.x)

108. Ye Q, Zhang Y, Zhuang J, Bi Y, Xu H, Shen Q, et al. The important roles and molecular mechanisms of annexin A(2) autoantibody in children with nephrotic syndrome. Ann Transl Med (2021) 9(18):1452. doi: [10.21037/atm-21-3988](https://doi.org/10.21037/atm-21-3988)

109. Delville M, Sigdel TK, Wei C, Li J, Hsieh SC, Fornoni A, et al. A circulating antibody panel for pretransplant prediction of FSGS recurrence after kidney transplantation. Sci Transl Med (2014) 6(256):256ra136. doi: [10.1126/](https://doi.org/10.1126/scitranslmed.3008538) [scitranslmed.3008538](https://doi.org/10.1126/scitranslmed.3008538)

110. Colucci M, Carsetti R, Rosado MM, Cascioli S, Bruschi M, Candiano G, et al. Atypical IgM on T cells predict relapse and steroid dependence in idiopathic nephrotic syndrome. Kidney Int (2019) 96(4):971–82. doi: [10.1016/j.kint.2019.04.006](https://doi.org/10.1016/j.kint.2019.04.006)

111. Musante L, Candiano G, Petretto A, Bruschi M, Dimasi N, Caridi G, et al. Active focal segmental glomerulosclerosis is associated with massive oxidation of plasma albumin. J Am Soc Nephrol. (2007) 18(3):799–810. doi: [10.1681/](https://doi.org/10.1681/ASN.2006090965) [ASN.2006090965](https://doi.org/10.1681/ASN.2006090965)

112. Candiano G, Musante L, Bruschi M, Petretto A, Santucci L, Del Boccio P, et al. Repetitive fragmentation products of albumin and alpha1-antitrypsin in glomerular diseases associated with nephrotic syndrome. J Am Soc Nephrol. (2006) 17(11):3139– 48. doi: [10.1681/ASN.2006050486](https://doi.org/10.1681/ASN.2006050486)

113. Niculovic KM, Blume L, Wedekind H, Kats E, Albers I, Groos S, et al. Podocyte-specific sialylation-deficient mice serve as a model for human FSGS. J Am Soc Nephrol (2019) 30(6):1021–35. doi: [10.1681/ASN.2018090951](https://doi.org/10.1681/ASN.2018090951)

114. Merscher S, Fornoni A. Podocyte pathology and nephropathy - sphingolipids in glomerular diseases. Front Endocrinol (Lausanne). (2014) 5:127. doi: [10.3389/](https://doi.org/10.3389/fendo.2014.00127) [fendo.2014.00127](https://doi.org/10.3389/fendo.2014.00127)

115. Angeletti A, Cantarelli C, Petrosyan A, Andrighetto S, Budge K, D'Agati VD, et al. Loss of decay-accelerating factor triggers podocyte injury and glomerulosclerosis. J Exp Med (2020) 217(9). doi: [10.1084/jem.20191699](https://doi.org/10.1084/jem.20191699)

116. Angeletti A, Magnasco A, Trivelli A, Degl'Innocenti LM, Piaggio G, Lugani F, et al. Refractory minimal change disease and focal segmental glomerular sclerosis treated with anakinra. Kidney Int Rep (2022) 7(1):121–4. doi: [10.1016/](https://doi.org/10.1016/j.ekir.2021.10.018) [j.ekir.2021.10.018](https://doi.org/10.1016/j.ekir.2021.10.018)

117. Koyama A, Fujisaki M, Kobayashi M, Igarashi M, Narita M. A glomerular permeability factor produced by human T cell hybridomas. Kidney Int (1991) 40 (3):453–60. doi: [10.1038/ki.1991.232](https://doi.org/10.1038/ki.1991.232)

118. Shalhoub RJ. Pathogenesis of lipoid nephrosis: a disorder of T-cell function. Lancet (1974) 2(7880):556–60. doi: [10.1016/S0140-6736\(74\)91880-7](https://doi.org/10.1016/S0140-6736(74)91880-7)

119. Lin CY, Hsu HC. Histopathological and immunological studies in spontaneous remission of nephrotic syndrome after intercurrent measles infection. Nephron (1986) 42(2):110–5. doi: [10.1159/000183647](https://doi.org/10.1159/000183647)

120. Tamura H, Kuraoka S, Hidaka Y, Nagata H, Furuie K, Nakazato H. A case of nephrotic syndrome that resolved with influenza b infection. Case Rep Nephrol Dial. (2021) 11(1):103–9. doi: [10.1159/000515062](https://doi.org/10.1159/000515062)

121. Peralta-Aros C, Garcia-Nieto V. Does zika virus infection induce prolonged remissions in children with idiopathic nephrotic syndrome? Pediatr Nephrol. (2017) 32 (5):897–900.

122. Yap HK, Cheung W, Murugasu B, Sim SK, Seah CC, Jordan SC. Th1 and Th2 cytokine mRNA profiles in childhood nephrotic syndrome: evidence for increased IL-
13 mRNA expression in relapse. *J Am Soc Nephrol*. (1999) 10(3):529–37. doi: [10.1681/](https://doi.org/10.1681/ASN.V103529) [ASN.V103529](https://doi.org/10.1681/ASN.V103529)

123. Lama G, Luongo I, Tirino G, Borriello A, Carangio C, Salsano ME. T-Lymphocyte populations and cytokines in childhood nephrotic syndrome. Am J Kidney Dis (2002) 39(5):958–65. doi: [10.1053/ajkd.2002.32769](https://doi.org/10.1053/ajkd.2002.32769)

124. Le Berre L, Bruneau S, Naulet J, Renaudin K, Buzelin F, Usal C, et al. Induction of T regulatory cells attenuates idiopathic nephrotic syndrome. J Am Soc Nephrol. (2009) 20(1):57–67. doi: [10.1681/ASN.2007111244](https://doi.org/10.1681/ASN.2007111244)

125. Nettis E, Brussino L, Patella V, Bonzano L, Detoraki A, Leo Di E, et al. Effectiveness and safety of dupilumab in patients with chronic rhinosinusitis with nasal polyps and associated comorbidities: a multicentric prospective study in real life. Clin Mol Allergy (2022) 20(1):6. doi: [10.1186/s12948-022-00171-2](https://doi.org/10.1186/s12948-022-00171-2)

126. Simpson EL, Gooderham M, Wollenberg A, Weidinger S, Armstrong A, Soung J, et al. Efficacy and safety of lebrikizumab in combination with topical corticosteroids in adolescents and adults with moderate-to-Severe atopic dermatitis: a randomized clinical trial (ADhere). JAMA Dermatol (2023) 159(2):182–91. doi: [10.1001/](https://doi.org/10.1001/jamadermatol.2022.5534) [jamadermatol.2022.5534](https://doi.org/10.1001/jamadermatol.2022.5534)

127. Liu LL, Qin Y, Cai JF, Wang HY, Tao JL, Li H, et al. Th17/Treg imbalance in adult patients with minimal change nephrotic syndrome. Clin Immunol (2011) 139 (3):314–20. doi: [10.1016/j.clim.2011.02.018](https://doi.org/10.1016/j.clim.2011.02.018)

128. Couser WG. Basic and translational concepts of immune-mediated glomerular diseases. J Am Soc Nephrol. (2012) 23(3):381–99. doi: [10.1681/ASN.2011030304](https://doi.org/10.1681/ASN.2011030304)

129. Hashimura Y, Nozu K, Kanegane H, Miyawaki T, Hayakawa A, Yoshikawa N, et al. Minimal change nephrotic syndrome associated with immune dysregulation,
polyendocrinopathy, enteropathy, X-linked syndrome. *Pediatr Nephrol*. (2009) 24 (6):1181–6. doi: [10.1007/s00467-009-1119-8](https://doi.org/10.1007/s00467-009-1119-8)

130. Bertelli R, Bodria M, Nobile M, Alloisio S, Barbieri R, Montobbio G, et al. Regulation of innate immunity by the nucleotide pathway in children with idiopathic nephrotic syndrome. Clin Exp Immunol (2011) 166(1):55–63. doi: [10.1111/j.1365-](https://doi.org/10.1111/j.1365-2249.2011.04441.x) [2249.2011.04441.x](https://doi.org/10.1111/j.1365-2249.2011.04441.x)

131. Polhill T, Zhang GY, Hu M, Sawyer A, Zhou JJ, Saito M, et al. IL-2/IL-2Ab complexes induce regulatory T cell expansion and protect against proteinuric CKD. J Am Soc Nephrol. (2012) 23(8):1303–8. doi: [10.1681/ASN.2011111130](https://doi.org/10.1681/ASN.2011111130)

132. Bertelli R, Di Donato A, Cioni M, Grassi F, Ikehata M, Bonanni A, et al. LPS nephropathy in mice is ameliorated by IL-2 independently of regulatory T cells activity. PloS One (2014) 9(10):e111285. doi: [10.1371/journal.pone.0111285](https://doi.org/10.1371/journal.pone.0111285)

133. Fribourg M, Cioni M, Ghiggeri G, Cantarelli C, Leventhal JS, Budge K, et al. CyTOF-enabled analysis identifies class-switched b cells as the main lymphocyte subset associated with disease relapse in children with idiopathic nephrotic syndrome. Front Immunol (2021) 12:726428. doi: 10.3389/fi[mmu.2021.726428](https://doi.org/10.3389/fimmu.2021.726428)

134. Brenner BM, Hostetter TH, Humes HD. Molecular basis of proteinuria of glomerular origin. N Engl J Med (1978) 298(15):826–33. doi: [10.1056/](https://doi.org/10.1056/NEJM197804132981507) [NEJM197804132981507](https://doi.org/10.1056/NEJM197804132981507)

135. Rennke HG, Patel Y, Venkatachalam MA. Glomerular filtration of proteins: clearance of anionic, neutral, and cationic horseradish peroxidase in the rat. Kidney Int (1978) 13(4):278–88. doi: [10.1038/ki.1978.41](https://doi.org/10.1038/ki.1978.41)

136. Blau EB, Haas JE. Glomerular sialic acid and proteinuria in human renal disease. Lab Invest. (1973) 28(4):477–81.

137. Fornoni A, Merscher S, Kopp JB. Lipid biology of the podocyte–new perspectives offer new opportunities. Nat Rev Nephrol. (2014) 10(7):379–88. doi: [10.1038/nrneph.2014.87](https://doi.org/10.1038/nrneph.2014.87)

138. Cui S, Verroust PJ, Moestrup SK, Christensen EI. Megalin/gp330 mediates uptake of albumin in renal proximal tubule. Am J Physiol (1996) 271(4 Pt 2):F900-907. doi: [10.1152/ajprenal.1996.271.4.F900](https://doi.org/10.1152/ajprenal.1996.271.4.F900)

139. Amsellem S, Gburek J, Hamard G, Nielsen R, Willnow TE, Devuyst O, et al. Cubilin is essential for albumin reabsorption in the renal proximal tubule. J Am Soc Nephrol. (2010) 21(11):1859–67. doi: [10.1681/ASN.2010050492](https://doi.org/10.1681/ASN.2010050492)

140. Bu L, Mirocha J, Haas M. Immunoglobulin g/albumin staining in tubular protein reabsorption droplets in minimal change disease and focal segmental glomerulosclerosis. Nephrol Dial Transplant. (2021) 36(6):1016–22. doi: [10.1093/ndt/](https://doi.org/10.1093/ndt/gfaa039) [gfaa039](https://doi.org/10.1093/ndt/gfaa039)

141. Colucci M, Stockmann H, Butera A, Masotti A, Baldassarre A, Giorda E, et al. Sialylation of n-linked glycans influences the immunomodulatory effects of IgM on T cells. J Immunol (2015) 194(1):151–7. doi: [10.4049/jimmunol.1402025](https://doi.org/10.4049/jimmunol.1402025)

142. Bosseboeuf A, Allain-Maillet S, Mennesson N, Tallet A, Rossi C, Garderet L, et al. Pro-inflammatory state in monoclonal gammopathy of undetermined significance and in multiple myeloma is characterized by low sialylation of pathogen-specific and other monoclonal immunoglobulins. Front Immunol (2017) 8:1347. doi: [10.3389/](https://doi.org/10.3389/fimmu.2017.01347) fi[mmu.2017.01347](https://doi.org/10.3389/fimmu.2017.01347)

143. Dalziel M, Crispin M, Scanlan CN, Zitzmann N, Dwek RA. Emerging principles for the therapeutic exploitation of glycosylation. Science (2014) 343 (6166):1235681. doi: [10.1126/science.1235681](https://doi.org/10.1126/science.1235681)

144. Parekh RB, Roitt IM, Isenberg DA, Dwek RA, Ansell BM, Rademacher TW. Galactosylation of IgG associated oligosaccharides: reduction in patients with adult and juvenile onset rheumatoid arthritis and relation to disease activity. Lancet (1988) 1 (8592):966–9. doi: [10.1016/S0140-6736\(88\)91781-3](https://doi.org/10.1016/S0140-6736(88)91781-3)

145. Sumar N, Colaco CB, Bodman KB, Parekh R, Williams P, Dwek R, et al. Abnormalities in the glycosylation of IgG in spouses of patients with rheumatoid arthritis. A Family study. J Autoimmun (1991) 4(6):907–14. doi: [10.1016/0896-8411](https://doi.org/10.1016/0896-8411(91)90053-f) [\(91\)90053-f](https://doi.org/10.1016/0896-8411(91)90053-f)

146. Candiano G, Petretto A, Bruschi M, Santucci L, Dimuccio V, Prunotto M, et al. The oxido-redox potential of albumin methodological approach and relevance to human diseases. J Proteomics. (2009) 73(2):188–95. doi: [10.1016/j.jprot.2009.06.006](https://doi.org/10.1016/j.jprot.2009.06.006)

147. Bruschi M, Candiano G, Santucci L, Ghiggeri GM. Oxidized albumin. the long way of a protein of uncertain function. Biochim Biophys Acta (2013) 1830(12):5473–9. doi: [10.1016/j.bbagen.2013.04.017](https://doi.org/10.1016/j.bbagen.2013.04.017)

148. Ghiggeri GM, Cercignani G, Ginevri F, Bertelli R, Zetta L, Greco F, et al. Puromycin aminonucleoside metabolism by glomeruli and glomerular epithelial cells. vitro. Kidney Int (1991) 40(1):35–42. doi: [10.1038/ki.1991.176](https://doi.org/10.1038/ki.1991.176)

149. Bertelli R, Bonanni A, Caridi G, Canepa A, Ghiggeri GM. Molecular and cellular mechanisms for proteinuria in minimal change disease. Front Med (Lausanne). (2018) 5:170. doi: [10.3389/fmed.2018.00170](https://doi.org/10.3389/fmed.2018.00170)

150. Candiano G, Musante L, Carraro M, Faccini L, Campanacci L, Zennaro C, et al. Apolipoproteins prevent glomerular albumin permeability induced in vitro by serum from patients with focal segmental glomerulosclerosis. J Am Soc Nephrol. (2001) 12 (1):143–50. doi: [10.1681/ASN.V121143](https://doi.org/10.1681/ASN.V121143)

151. Bonanni A, Bertelli R, Rossi R, Bruschi M, Di Donato A, Ravani P, et al. A pilot study of IL2 in drug-resistant idiopathic nephrotic syndrome. PloS One (2015) 10(9): e0138343. doi: [10.1371/journal.pone.0138343](https://doi.org/10.1371/journal.pone.0138343)

152. Ravani P, Magnasco A, Edefonti A, Murer L, Rossi R, Ghio L, et al. Short-term effects of rituximab in children with steroid- and calcineurin-dependent nephrotic
syndrome: a randomized controlled trial. *Clin J Am Soc Nephrol*. (2011) 6(6):1308–15. doi: [10.2215/CJN.09421010](https://doi.org/10.2215/CJN.09421010)

153. Basu B, Sander A, Roy B, Preussler S, Barua S, Mahapatr TKS, et al. Efficacy of rituximab vs tacrolimus in pediatric corticosteroid-dependent nephrotic syndrome: a randomized clinical trial. JAMA Pediatr (2018) 172(8):757–64. doi: [10.1001/](https://doi.org/10.1001/jamapediatrics.2018.1323) [jamapediatrics.2018.1323](https://doi.org/10.1001/jamapediatrics.2018.1323)

154. Basu B. Ofatumumab for rituximab-resistant nephrotic syndrome. N Engl J Med (2014) 370(13):1268–70. doi: [10.1056/NEJMc1308488](https://doi.org/10.1056/NEJMc1308488)

155. Colucci M, Carsetti R, Cascioli S, Casiraghi F, Perna A, Rava L, et al. B cell reconstitution after rituximab treatment in idiopathic nephrotic syndrome. *J Am Soc*
Nephrol. (2016) 27(6):1811–22. doi: [10.1681/ASN.2015050523](https://doi.org/10.1681/ASN.2015050523)

156. Lebrun C, Cohen M, Rosenthal-Allieri MA, Bresch S, Benzaken S, Marignier R, et al. Only follow-up of memory b cells helps monitor rituximab administration to patients with neuromyelitis optica spectrum disorders. Neurol Ther (2018) 7(2):373–83. doi: [10.1007/s40120-018-0101-4](https://doi.org/10.1007/s40120-018-0101-4)

157. Leandro MJ, Cambridge G, Ehrenstein MR, Edwards JC. Reconstitution of peripheral blood b cells after depletion with rituximab in patients with rheumatoid arthritis. Arthritis Rheumatol (2006) 54(2):613–20. doi: [10.1002/art.21617](https://doi.org/10.1002/art.21617)

158. Jamin A, Berthelot L, Couderc A, Chemouny JM, Boedec E, Dehoux L, et al. Autoantibodies against podocytic UCHL1 are associated with idiopathic nephrotic syndrome relapses and induce proteinuria in mice. J Autoimmun (2018) 89:149–61. doi: [10.1016/j.jaut.2017.12.014](https://doi.org/10.1016/j.jaut.2017.12.014)

159. Kidney disease: improving global outcomes glomerular diseases work g. KDIGO 2021 clinical practice guideline for the management of glomerular diseases. Kidney Int (2021) 100(4S):S1–S276.

160. Trautmann A, Vivarelli M, Samuel S, Gipson D, Sinha A, Schaefer F, et al. IPNA clinical practice recommendations for the diagnosis and management of children with steroid-resistant nephrotic syndrome. Pediatr Nephrol (2020) 35(8):1529–61. doi: [10.1007/s00467-020-04519-1](https://doi.org/10.1007/s00467-020-04519-1)

161. Larkins NG, Liu ID, Willis NS, Craig JC, Hodson EM. Non-corticosteroid immunosuppressive medications for steroid-sensitive nephrotic syndrome in children. Cochrane Database Syst Rev (2020) 4(4):CD002290. doi: [10.1002/](https://doi.org/10.1002/14651858.CD002290.pub5) [14651858.CD002290.pub5](https://doi.org/10.1002/14651858.CD002290.pub5)

162. Muhlig AK, Lee JY, Kemper MJ, Kronbichler A, Yang JW, Lee JM, et al. Levamisole in children with idiopathic nephrotic syndrome: clinical efficacy and pathophysiological aspects. J Clin Med (2019) 8(6). doi: [10.3390/jcm8060860](https://doi.org/10.3390/jcm8060860)

163. Chan EY, Yu ELM, Angeletti A, Arslan Z, Basu B, Boyer O, et al. Long-term efficacy and safety of repeated rituximab to maintain remission in idiopathic childhood nephrotic syndrome: an international study. J Am Soc Nephrol. (2022) 33(6):1193–207. doi: [10.1681/ASN.2021111472](https://doi.org/10.1681/ASN.2021111472)

164. Ravani P, Colucci M, Bruschi M, Vivarelli M, Cioni M, Di Donato A, et al. Human or chimeric monoclonal anti-CD20 antibodies for children with nephrotic syndrome: a superiority randomized trial. J Am Soc Nephrol. (2021) 32(10):2652-63. doi: [10.1681/ASN.2021040561](https://doi.org/10.1681/ASN.2021040561)

165. Angeletti A, Bruschi M, Colucci M, Kajana X, La Porta E, Caridi G, et al. Circulating anti-rituximab antibodies do not affect response to rituximab in steroiddependent nephrotic syndrome. Kidney Int Rep (2022) 7(11):2509–12. doi: [10.1016/](https://doi.org/10.1016/j.ekir.2022.08.006) [j.ekir.2022.08.006](https://doi.org/10.1016/j.ekir.2022.08.006)

166. Bhatt DL, Szarek M, Steg PG, Cannon CP, Leiter LA, McGuire DK, et al. Sotagliflozin in patients with diabetes and recent worsening heart failure. N Engl J Med (2021) 384(2):117–28. doi: [10.1056/NEJMoa2030183](https://doi.org/10.1056/NEJMoa2030183)

167. Bakris GL, Agarwal R, Anker SD, Pitt B, Ruilope LM, Rossing P, et al. Effect of finerenone on chronic kidney disease outcomes in type 2 diabetes. N Engl J Med (2020) 383(23):2219–29. doi: [10.1056/NEJMoa2025845](https://doi.org/10.1056/NEJMoa2025845)

168. Furie RA, Aroca G, Cascino MD, Garg JP, Rovin BH, Alvarez A, et al. B-cell depletion with obinutuzumab for the treatment of proliferative lupus nephritis: a randomised, double-blind, placebo-controlled trial. Ann Rheum Dis (2022) 81(1):100– 7. doi: [10.1136/annrheumdis-2021-220920](https://doi.org/10.1136/annrheumdis-2021-220920)

169. Basu B, Angeletti A, Islam B, Ghiggeri GM. New and old anti-CD20 monoclonal antibodies for nephrotic syndrome. Where We Are? Front Immunol (2022) 13:805697. doi: 10.3389/fi[mmu.2022.805697](https://doi.org/10.3389/fimmu.2022.805697)