REVIEW

Molecular mechanism of ferulic acid and its derivatives in tumor progression

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Abstract

Cancer is a signifcant disease that poses a major threat to human health. The main therapeutic methods for cancer include traditional surgery, radiotherapy, chemotherapy, and new therapeutic methods such as targeted therapy and immunotherapy, which have been developed rapidly in recent years. Recently, the tumor antitumor effects of the active ingredients of natural plants have attracted extensive attention. Ferulic acid (FA), (3-methoxy-4-hydroxyl cinnamic), with the molecular formula is $C_{10}H_{10}O_4$, is a phenolic organic compound found in ferulic, angelica, jujube kernel, and other Chinese medicinal plants but is also, abundant in rice bran, wheat bran, and other food raw materials. FA has anti-infammatory, analgesic, anti-radiation, and immune-enhancing efects and also shows anticancer activity, as it can inhibit the occurrence and development of various malignant tumors, such as liver cancer, lung cancer, colon cancer, and breast cancer. FA can cause mitochondrial apoptosis by inducing the generation of intracellular reactive oxygen species (ROS). FA can also interfere with the cell cycle of cancer cells, arrest most cancer cells in G_0/G_1 phase, and exert an antitumor effect by inducing autophagy; inhibiting cell migration, invasion, and angiogenesis; and synergistically improving the efficacy of chemotherapy drugs and reducing adverse reactions. FA acts on a series of intracellular and extracellular targets and is involved in the regulation of tumor cell signaling pathways, including the phosphatidylinositol 3 kinase (PI3K)/protein kinase B (AKT), B-cell lymphoma-2 (Bcl-2), and tumor protein 53 (P53) pathways and other signaling pathways. In addition, FA derivatives and nanoliposomes, as platforms for drug delivery, have an important regulatory efect on tumor resistance. This paper reviews the efects and mechanisms of antitumor therapies to provide new theoretical support and insight for clinical antitumor therapy.

Keywords Ferulic acid · Tumor · Derivative · Nanoliposome · Mechanism of action

Abbreviations

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Introduction

Malignant tumors are a multifactor and multistep disease, the incidence of which has increased in recent years [\[1](#page-12-0)], but the pathogenesis has not been fully elucidated, so there is no efective aetiological treatment. In 2023, 1,958,310 new cancer cases and 609,820 cancer deaths are projected to occur in the United States [\[2](#page-12-1)]. Globally, there will be an estimated 28.4 million new cancer cases by 2040, a 47 percent increase over the number of new cases in 2020 [[3\]](#page-12-2). This trend refects the growing burden that cancer imposes on global health systems and highlights the need for continued efforts to prevent, detect, and treat this disease.

Current cancer treatments include surgical intervention, radiation, and chemotherapy drugs, which often kill healthy cells and cause a host of adverse reactions in patients. With the continuous development of medicine, new therapies such as induction chemotherapy and targeted therapy have been applied to cancer treatment. Compared with traditional chemoradiotherapy, the survival time of patients has been signifcantly extended with these new therapies, but the adverse reactions and quality of life have not been fundamentally improved $[4, 5]$ $[4, 5]$ $[4, 5]$ $[4, 5]$. Therefore, the continuous exploration of safe and efficient antitumor treatments is still the main research direction of contemporary oncology. Natural plant components play a signifcant role in the prevention and treatment of cancer, have significant efficacy in improving the clinical symptoms, quality of life, and prognosis of cancer patients, and have gradually become an important means of cancer prevention and treatment.

Ferulic acid (FA), a phenolic substance widely found in plant cell walls [[6–](#page-12-5)[8\]](#page-12-6), is an important active component of many traditional Chinese medicines. It has stable physical and chemical properties, outstanding pharmacological activity, and few toxic and side effects on the human body. Its pharmacological activities mainly include antioxidant $[9-11]$ $[9-11]$, anti-inflammatory [[12,](#page-12-9) [13\]](#page-12-10), anti-diabetic [\[14](#page-12-11), [15\]](#page-12-12), blood pressure-low-ering [\[16](#page-12-13)], hepatoprotective [\[17,](#page-12-14) [18\]](#page-12-15), and immunoregulatory [\[19,](#page-12-16) [20](#page-12-17)] activities. With the deepening of pharmacological research on FA, it has shown signifcant antitumor biological activity [[21–](#page-12-18)[24\]](#page-12-19) and is expected to become a potential drug for the treatment of malignant tumors. In this paper, progress related to the antitumor mechanism of FA was reviewed to provide new ideas for solving the current problems of poor efficacy, high toxicity, and drug resistance of traditional antitumor drugs.

Induction of tumor cell apoptosis after FA intervention

Tumor cells can grow without limit and resist programmed death caused by genes. This malignant behavior of tumors not only increases the difficulty of treatment but is also an important cause of cancer-related death. Apoptosis is the most important form of programmed cell death [\[25](#page-12-20)]. FA has a signifcant efect on inducing apoptosis.

Tumor protein 53 (P53)

The expression of p53, a key tumor suppressor gene $[26]$ $[26]$, is altered in most cancers. The loss of p53 function is often a prerequisite for the development of cancer [[27\]](#page-12-22). Niu et al. [\[28\]](#page-12-23) showed that FA could induce the apoptosis of gastric cancer SDC-7901 cells, and the mechanism involved FAmediated upregulation of the mRNA and protein expression of p53. Umut et al. [\[29](#page-12-24)] observed that FA could increase the expression level of p53 in MIA PaCa-2 pancreatic cancer cells while reducing the expression levels of cyclin D1 and cyclin-dependent kinase (CDK) 4/6. In addition, FA was found to reduce colony formation and inhibit cell invasion and migration. The results suggested that FA could promote the apoptosis of MIA PaCa-2 cells by increasing the expression of p53, thus showing an antitumor effect. Folatemediated metabolism is crucial to the stability and function of the genome and afects the occurrence and development of tumors $[30]$ $[30]$. Kumar et al. $[31]$ $[31]$ evaluated the targeted efficacy of chitosan-coated FA-loaded solid lipid nanoparticles and folate conjugate (FFA) in colon cancer. Compared with the control cells, FFA-treated HT-29 cells showed increased p53 levels, increased apoptosis, and loss of mitochondrial membrane potential. Further studies showed that FFA could trigger the release of cytochrome C in colon cancer cells, and the expression of cysteinyl aspartate-specifc proteinase (caspase)-9 and -3 increased after FFA treatment. Together, these results indicate that FFA can activate p53-mediated intrinsic apoptosis, suggesting that these targeted biomaterials could be used as an efective drug in cancer therapy.

B‑cell lymphoma‑2 (Bcl‑2) protein family

Bcl-2 plays an important role in controlling cell apoptosis and enhancing cell survival. miR-34 is abnormally expressed in the tumor process and is considered a tumor suppressor microRNA due to its synergistic efect with the tumor suppressor gene p53 [[32](#page-13-1)]. Increasing the expression of miR-34a can inhibit Bcl-2 and increase apoptosis [\[33](#page-13-2)]. In a human cervical cancer xenograft model, FA treatment was found to reduce tumor weight in a dose-dependent manner, increase miR-34a expression, downregulate Bcl-2 protein expression, and upregulate caspase-3 protein expression [[34\]](#page-13-3). Therefore, the inhibitory effect of FA on the growth of transplanted tumors of human cervical cancer in nude mice may be realized by upregulating miR-34a, thus inhibiting the expression of its target gene Bcl-2, initiating the apoptotic pathway and promoting cell apoptosis. Zhang et al. [[35\]](#page-13-4) used FA treatment on gastric cancer MGC-803 cells, and the results showed that FA could upregulate the expression of Bcl-2-associated X (Bax) mRNA and protein and downregulate the expression of Bcl-2 mRNA and protein, thus efectively inducing the apoptosis of MGC-803 cells and inhibiting their proliferation. These results suggest that the mechanism of FA-induced apoptosis may be related to the activation of the endogenous mitochondrial apoptosis pathway. Isoferulic acid, an isomer of FA, also signifcantly inhibited the proliferation of human renal carcinoma A-498 cells, induced cleaved caspase-3 expression, and promoted the apoptosis of A-498 cells; moreover, isoferulic acid dosedependently downregulated the expression of β-catenin and MYC proto-oncogene (c-Myc), inducing apoptosis [[36](#page-13-5)]. Therefore, isoferulic acid is considered a potential candidate for the treatment of renal carcinoma.

Yue et al. [[37](#page-13-6)] evaluated the potential effects of FA's nitrate compound FXS-3 on the proliferation and metastasis of lung cancer A549 cells. The results showed that FXS-3 can inhibit the activity of A549 cells by upregulating the Bax/Bcl-2 ratio mediated by the c-Jun N-terminal kinase (JNK) and extracellular signal-regulated kinase (ERK)/ p38signaling pathways, which provides an important scientifc basis for the development of FA derivative anticancer drugs. Nitric oxide has a wide range of potential applications in tumor therapy [\[38](#page-13-7)]. Zhang et al. [[37](#page-13-6)] designed and synthesized an FA-nitric oxide donor conjugate. After this coupling was applied to A549 lung cancer cells, it was found that it could upregulate the expression levels of Bax and JNK by downregulating the expression of,Bcl-2, P38 and ERK, thus inhibiting the proliferation of A549 cells and inducing their apoptosis.

Reactive oxygen species (ROS)

ROS are a group of short-lived and highly active oxygencontaining molecules that can induce DNA damage and genotoxic stress, as well as initiate oxidative stress-induced tumor cell death [[39](#page-13-8)]. After treatment with FA, Cao et al. [\[40](#page-13-9)] observed an increase in ROS production and a decrease in superoxide dismutase activity and glutathione content in EC-1 and TE-4 oesophageal cancer cells. In addition, FA could promote the release of lactate dehydrogenase (LDH) and the activation of caspase-3 in oesophageal cancer cells, thus inducing cell apoptosis. Rosaria et al. [[41\]](#page-13-10) found that FA can activate the ERK1/2 pathway through the

participation of ROS and play a proapoptotic role in human glioblastoma U-87 MG cells by reducing the expression levels of Bcl-2, ERK1/2, and c-Myc.

Zinc oxide nanoparticles are efective carriers for the targeted delivery of anticancer drugs to tumor cells [[42](#page-13-11)]. Babu et al. [\[43](#page-13-12)] conjugated zinc oxide nanoparticles with FA (ZnONPs-FA) to act on hepatoma Huh-7 and HepG2 cells. The results showed that ZnONPs-FA could induce oxidative DNA damage and apoptosis by inducing ROS production. Therefore, ZnONPs-FA may be a promising drug for the treatment of liver cancer.

Proliferating cell nuclear antigen (PCNA) pathway

PCNA is the core of many basic cellular processes, such as DNA replication, DNA damage, repair, and chromatin structure maintenance [[44](#page-13-13), [45](#page-13-14)]; it is also an excellent target for cancer therapy [\[46](#page-13-15)]. Arvind et al. [[47\]](#page-13-16) encapsulated FA and aspirin in a new chitosan-coated solid lipid nanoparticle (c-SLN) and observed the potential therapeutic efect of this drug delivery method. The results showed that the viability of MIA PaCa-2 and Panc-1 cells decreased signifcantly after c-SLN treatment. In in vivo studies of oral administration of c-SLN to a pancreatic cancer transplant mouse model, tumor growth was signifcantly inhibited compared with controls. In addition, immunohistochemical analysis showed signifcantly reduced expression of PCNA and MKI67 and increased expression of the apoptotic proteins p-RB, p21, and p-ERK1/2, indicating the proapoptotic efect of this regimen. The mechanisms by which FA induces apoptosis in cancer cells are shown in Fig. [1.](#page-4-0)

Inhibition of tumor cell proliferation in response to FA

Cell proliferation is an important part of cell growth and diferentiation [[48\]](#page-13-17), and active proliferation and metabolism are characteristics of tumor cells [\[49](#page-13-18)]. Unlike normal cells, cancer cells have the characteristics of infnite growth, transformation and metastasis and, therefore, are difficultly eliminated. Thus, inhibiting the proliferation of tumor cells is an important method of tumor therapy. In vitro and in vivo studies have shown that FA can efectively inhibit the proliferation of colorectal cancer, lung cancer and breast cancer cells in a variety of ways.

Cell cycle blockade

The cell cycle is regulated by various factors [[50](#page-13-19)], and impairment of its regulatory mechanism of the cell cycle can lead to uncontrolled growth of normal cells and transformation into tumor cells. Therefore, targeting cell cycle components may be an efective anticancer strategy. Wang et al. [[51](#page-13-20)] showed that the survival rate of osteosarcoma 143B cells after FA treatment was signifcantly reduced. Compared with that in the control group, the proportion of G_0/G_1 phase cells in the FA treatment group was signifcantly increased, the expression levels of CDK2, CDK4, and CDK6 were decreased, and the ratio of Bax/ Bcl-2 was increased. Further studies showed that FA can inhibit phosphatidylinositol 3 kinase (PI3K)/ protein kinase B (AKT) in a dose-dependent manner [\[52\]](#page-13-21). These results suggest that FA may inhibit the proliferation and induce the apoptosis of osteosarcoma cells by inhibiting the PI3K/Akt pathway. Canan et al. [\[53](#page-13-22)] observed that FA can inhibit cell proliferation by increasing the gene expression of TP53 and decreasing the gene expression of CDK2, CDK,4 and CDK6 in prostate cancer PC-3 cells, thus leading to cell cycle arrest in PC-3 cells. Luo et al. [[54](#page-13-23)] found that FA can significantly reduce the number of S phase and G_2/M phase cells in HCT116 and HT-29 colorectal cancer cells and increase the proportion of G1 phase cells in these two cancer cell lines, which refects its antitumor cell proliferation ability.

Gao et al. [[55](#page-13-24)] treated cervical cancer HeLa and CaSki cells with FA, and cell growth was signifcantly inhibited. Further studies showed that FA induced HeLa and CaSki cells to stay in the G_0/G_1 phase of the cell cycle in a dosedependent manner, while FA induced the expression of cycle-related proteins such as p53 and p21 and decreased the expression levels of cyclin D1 and cyclin E. Anwar et al. [[56\]](#page-13-25) reported that FA could significantly reduce the proportion of cells in S phase, thus inhibiting the proliferation of the breast cancer cell line MDA-MB-231. However, in contrast to the above results, the proportion of cells in other phases of the cell cycle did not change signifcantly in this study, and the specifc reasons remain unclear.

Janus kinase/signal transduction and transcriptional activator (JAK/STAT)

The JAK/STAT3 signaling pathway is involved in almost all immune regulatory processes, including tumor cell recognition and tumor-driven immune escape [\[57\]](#page-13-26). Therefore, inhibitors targeting the JAK/STAT3 pathway may inhibit tumor cell growth and stimulate antitumor immunity. Studies have shown [\[58\]](#page-13-27) that FA can effectively reduce the expression levels of the immune factors IL-4, platelet-derived growth factor (PDGF), and granulocyte–macrophage colony-stimulating factor(GM-CSF) as well as the phosphorylation level of JAK2/STAT6 in lung cancer A549 cells, suggesting that FA can inhibit the proliferation and metastasis of lung cancer cells by inhibiting the JAK2/STAT6 immune signaling pathway.

Fig. 1 Mechanism of tumor cell apoptosis induced by FA and its derivatives. FA induces apoptosis by upregulating the expression of P53, reducing the expression of cyclin D1 and cyclin-dependent kinase (CDK) 4/6, upregulating Mir-43a, inhibiting the expression of Bcl-2, and activating the apoptosis pathway to promote cell apoptosis. FA causes DNA oxidative damage and apoptosis by inducing ROS production. *JNK* c-Jun N-terminal kinase, *ERK* extracellular signal-regulated kinase, *C-fox* forkhead box C, *STAT* signal transduc-

tion and transcriptional activator, *JAK* Janus kinase, *BCL-2* B-cell lymphoma-2, *Bax* Bcl-2-associated X, *PI3K* phosphatidylinositol 3 kinase, *AKT* protein kinase B, *mTOR* mammalian target of rapamycin, *P53* tumor protein 53, *Cyt-c* Cytochrome C, *Caspase* cysteinyl aspartate specifc proteinase, *C-Myc* MYC proto-oncogene, *CCND* cyclin D, *CDK* cyclin-dependent kinases, *ROS* reactive oxygen species

Nuclear factor‑kappa B‑gene binding (NF‑κB) pathway

Dysfunctional NF-κB activity is associated with infammatory diseases and cancer [[59\]](#page-13-28). The NF-κB-signaling pathway has long been used as a potential target for disease treatment because it activates antiapoptotic genes and downregulates the expression of proapoptotic factors to induce tumor cell growth. FA can efectively inhibit the activation of the NF-κB signaling pathway [[60](#page-13-29)]. Hiroko et al. [\[61](#page-13-30)] showed that FA can reduce tyrosinase activity by directly binding to enzymes and inhibit the tyrosinase phosphorylation induced by casein kinase 2 (CK2) in B16 melanoma cells in a dose-dependent manner in vitro, thus inhibiting the activation of NF-κB.

Overexpression of the pyrroline‑5‑carboxylate reductase 1 (PYCR1) pathway

Overexpression of PYCR1 is associated with the occurrence and development of cancer [\[62](#page-13-31)]. Yang et al. [\[63](#page-13-32)] found that FA could inhibit the proliferation of breast cancer MCF-7 and 4T1 cells in a dose-dependent manner, and further studies found a direct interaction between FA and PYCR1. PYCR1 can catalyze proline metabolism and synthesis in vivo through enzymatic reactions, which play a role in promoting tumor growth and proliferation. FA can target PYCR1 and inhibit its enzyme activity in a concentrationdependent manner.

PI3K/AKT/mammalian target of rapamycin signaling pathway

The PI3K/AKT/mTOR signaling pathway plays an important role in the regulation of cell proliferation, apoptosis, metabolism, and angiogenesis [\[64](#page-13-33)]. FA inhibits the activation of the PI3K/AKT pathway [[65](#page-14-0)]. Luo et al. [\[66\]](#page-14-1) found that in CaSki cells, phosphorylation of Akt and PI3K was reduced by FA in a concentration-dependent manner, leading to cytotoxicity and apoptosis of CaSki cells. Wu et al. [[67\]](#page-14-2) found that FA could signifcantly reduce the expression level of mTOR mRNA and Ki-67 protein in A549 lung cancer graft tissue, increase the expression of caspase-3 protein, and signifcantly inhibit the growth of tumors. The mechanisms by which FA inhibits tumor cell proliferation are shown in Fig. [2](#page-6-0).

Inhibition of tumor cell metastasis and invasion in response to FA

The process of tumor cells invading and metastasizing from the primary lesion to the surrounding tissues and then invading the surrounding tissues or even spreading throughout the whole body is called tumor invasion and metastasis. Cancer cell metastasis is an important cause of cancer death [\[68](#page-14-3)]. The migration and invasion abilities of tumor cells play an important role in the metastasis and recurrence of tumors. Inhibiting the invasion and metastasis of tumor cells can efectively interfere with the cancer process.

Vascular endothelial growth factor (VEGF)

Antiangiogenic therapy is an efective means for the treatment of solid cancer [[69](#page-14-4)]. VEGF is the main medium of angiogenesis, and the application of antiangiogenic agents targeting VEGF is an important strategy for the treatment of many cancers [\[70](#page-14-5), [71\]](#page-14-6). Zhang et al. [[72\]](#page-14-7) found that FA could reduce the expression level of VEGF mRNA and protein in osteosarcoma SaOS-2 cells, and the expression level of CD34 protein in the metastatic tissue of nude mice treated with FA was significantly decreased. The results suggest that FA can reduce osteosarcoma's blood supply capacity by inhibiting the vascularization process to achieve anti-tumor effects. Riham et al. [\[73\]](#page-14-8) prepared FA nanoparticle liposomes. In vitro*,* studies revealed that FA liposomes showed antiangiogenic potential by regulating the expression of cyclin D1 and VEGF after acting on colorectal cancer HCT116 and Caco2 cells. The results show that FA lipid nanoliposomes are an ideal system for the treatment of colorectal cancer and can be used as an efective measure to prevent metastasis.

Fibroblast growth factor 1 (FGF1)

FGF1 plays an important role in tumor progression as a mediator leading to increased invasion and migration [[74,](#page-14-9) [75](#page-14-10)]. Yang et al. [[76\]](#page-14-11) found that FA could inhibit endothelial cell proliferation, migration, and response to FGF1. In vivo*,* angiogenesis measurements showed that FA inhibited FGF1 induced angiogenesis of the rat aortic ring microvasculature. Further studies showed that FA inhibited FGF1-triggered fibroblast growth factor receptor 1 (FGFR1) and PI3K/ Akt signaling [[76\]](#page-14-11). These results suggest that FA is a novel FGFR1 inhibitor with potential antiangiogenic and anticancer activities.

Epithelial–mesenchymal transition (EMT)

EMT is a complex biological transdiferentiation process that confers mesenchymal characteristics to epithelial cells [[77\]](#page-14-12). EMT is considered an important factor in cancer invasion and metastasis. In the course of cancer, EMT is closely related to tumor occurrence, invasion, metastasis and treatment resistance [\[78](#page-14-13)]. Zhang et al. [[79\]](#page-14-14) found in a Transwell experiment that FA had a dose-dependent inhibitory efect on the migration of MDA-MB-231 breast cancer cells, and

Fig. 2 Mechanism of FA and its derivatives inhibiting tumor cell proliferation. FA can inhibit cell proliferation by increasing the gene expression of TP53 and decreasing the gene expression of CDK2, CDK4, and CDK6, thereby leading to cell cycle arrest. It also reduces the expression levels of IL-4, PDGF, and GM-CSF and the phosphorylation level of JAK2/STAT6 and inhibits the JAK2/STAT6 signaling pathway to suppress cell proliferation and metastasis. FA inhibits casein kinase 2 (CK2)-induced phosphorylation of tyrosinase, thereby inhibiting the activation of NF-κB. FA reduces the phosphorylation

of Akt and PI3K and inhibits the activation of the PI3K/AKT pathway. *PI3K* phosphatidylinositol 3 kinase, *AKT* protein kinase B, *mTOR* mammalian target of rapamycin, *TP53* tumor protein 53, *CDK* cyclin-dependent kinases, *JAK* Janus kinase, *STAT* signal transduction and transcriptional activator, *TNF-α* tumor necrosis factor-α, *Bax* Bcl-2-associated X, *PCNA* proliferating cell nuclear antigen, *PDGF* platelet-derived growth factor, *GM-CSF* granulocyte–macrophage colony-stimulating factor, *NF-κB* nuclear factor-k-gene binding, *IL* interleukin

even low concentrations of FA could inhibit the migration of breast cancer cells. Further studies on protein and mRNA levels showed that E-cadherin signifcantly increased and vimentin, Snail, and Slug proteins decreased after FA treatment. These results confrm that FA can inhibit EMT and can inhibit tumor metastasis. Matrix metalloproteinases (MMPs) are closely related to angiogenesis, invasion, metastasis, and the avoidance of immune surveillance in the course of cancer, among which MMP1 and MMP9 are universally upregulated in almost all cancers [\[80\]](#page-14-15). Park et al. [\[81\]](#page-14-16) showed that FA could inhibit the expression of MMP1 and MMP9 in melanoma B16F10 cells and had no cytotoxicity when the concentration was as high as $20 \mu M$, showing its potential as a functional food. Yavuz et al. [[82\]](#page-14-17) found that the gene expression of MMP-2 and MMP-9, which are responsible for extracellular matrix degradation, was signifcantly reduced, TIMP1 expression was increased and invasion and migration were decreased in FA-treated TT human thyroid cancer cells compared to control TT human thyroid cancer cells.

Carbonic anhydrase IX (CAIX) pathway

CAIX is a kind of transmembrane protease induced by a hypoxic microenvironment that belongs to the zinc metalloproteinase family; its high expression in the tumor cell membrane is caused by hypoxia. CAIX, which is induced and activated by hypoxia in the tumor process, is involved in the molecular mechanism of cancer cell invasion and metastasis and has been widely studied as a therapeutic target for cancer [\[83](#page-14-18), [84](#page-14-19)]. Babita et al. [[85\]](#page-14-20) synthesized an FA-triazole compound, and the results showed that this compound could inhibit the activity of HT-29 and HepG2 cells in a concentration-dependent manner. CAIX expression in treated HT-29 and HepG2 cells was signifcantly reduced, and the toxicity to HEK293 cells of the embryonic kidney was small within the same drug concentration range. The results suggest that this compound has good anticancer properties and can selectively inhibit CAIX, indicating that the use of FA as a CAIX inhibitor has good potential as a targeted cancer therapy. The mechanism by which FA inhibits the invasion and metastasis of cancer cells is shown in Fig. [3](#page-8-0).

Enhanced sensitivity to radiotherapy/ chemotherapy

The tolerance of tumor cells to chemoradiotherapy after a period of treatment is a serious problem. Commonly used chemotherapy drugs easily cause patient intolerance or tumor drug resistance, so it is urgent to develop new clinical drugs that have increased efectiveness and reduced toxicity.

FA is poor in water solubility, and its sodium salt, sodium ferulate, has high solubility and stable properties. It can be synthesized artifcially, and after years of research and development, it has been widely used in the feld of medicine. Resveratrol, a natural medicine, has a strong antitumor ability [[86\]](#page-14-21). Yuuga et al. [\[87\]](#page-14-22) found that novel derivatives of FA coupled with resveratrol could inhibit the proliferation of colon cancer HCT116 cells. It was further found that FA derivatives increased the mRNA level of the tumor suppressor p15, a CDK inhibitor. These results suggest that FA derivatives inhibit tumor cell proliferation by increasing p15 expression in HCT116 cells.

Subburayan et al. [[88\]](#page-14-23) reported that FA can produce free radicals, so FA treatment before radiotherapy could enhance the radiation efect of cancer cells. It was also confrmed by experiments that FA can increase ROS, lipid peroxidation markers, and DNA damage in HeLa and ME-80 cervical cancer cells, thereby reducing cell viability, survival rate, and antioxidant status and enhancing radiotherapy sensitivity. Das et al. [\[89](#page-14-24)] observed that the ROS levels in the treatment group were signifcantly higher than those in the control group after FA was used to pretreat lung cancer A549 and liver cancer HepG2 cells before radiotherapy. In this case, exposure to γ rays would further increase ROS levels and improve the radiosensitivity of cancer cells, indicating that the combination of FA and radiotherapy could activate the mitochondrial apoptotic pathway. In addition, compared with radiotherapy alone, combination therapy showed tumor regression, caspase-3 activation, and nuclear fragmentation in tumor tissues. In addition, FA pretreatment protected peripheral blood mononuclear cells (PBMCs) and normal lung fbroblasts (WI38) from radiation damage. In conclusion, FA preconditioning before radiotherapy provides an efective strategy to kill cancer cells and demonstrates the potential to improve radiotherapy efficacy.

Zheng et al. [[90\]](#page-14-25) synthesized stable PFA&DOX nanoparticles (NPs) with doxorubicin (DOX) from poly ferulic acid (PFA) chemically modifed by FA and found that NPs could sustainably release active components under slightly acidic conditions in vitro, ensuring efective drug administration in the acidic tumor microenvironment. In vivo*,* antitumor studies showed that PFA nanocarriers can promote the accumulation of drugs at the tumor site to obtain better antitumor efects. In addition, PFA&DOX NPs reduced the toxicity of free DOX, thus improving the safety, suggesting that these drug-loaded NPs can provide a reference for future clinical antitumor drug development.

FA also inhibited P-glycoprotein (P-gp) transport in drug-resistant $KB-Ch(R)-8-5$ cells and significantly downregulated ABCB1 gene expression in a concentration-dependent manner. Cytotoxicity analysis showed that FA reduced paclitaxel resistance in KB-Ch(R)-8-5 cells and upregulated apoptosis signaling in paclitaxel-induced

Fig. 3 Mechanism of FA and its derivatives inhibiting tumor cell invasion and migration. FA inhibits cell invasion and migration by reducing the expression levels of VEGF mRNA and protein and CD34 protein. FA inhibits FGF1-induced angiogenesis and inhibits invasion and migration. FA can inhibit the expression of MMP1 and

MMP9 in cells, inhibit EMT, and further suppress cell invasion and metastasis. *VEGF* vascular endothelial growth factor, *FGF* fbroblast growth factor, *FGFR* fbroblast growth factor receptor, *EMT* epithelial-mesenchymal transition, *MMP* matrix metalloproteinase

resistant cells, demonstrating its potential for chemical sensitization [[91](#page-14-26)]. These results suggest that the downregulation of ABCB1 and subsequent apoptosis signal transduction may be responsible for the chemical sensitization potential of FA in P-gp-overexpressing cell lines. However, another study [\[92\]](#page-14-27) pointed out that although FA can offset the ototoxicity of cisplatin to a certain extent, FA shows unnecessary tumor-protective effects, which may limit its clinical use. Therefore, how to ensure the safety of drug use while taking into account the antitumor efficacy is still a problem worth exploring.

Reversing the drug resistance of tumor cells

Tumor cells can mutate under prolonged drug use, showing adaptability. Once tumor resistance occurs, the drug cannot exert its normal anticancer efect. Low intracellular drug concentrations are an important cause of drug resistance in tumor cells.

Hyaluronic acid nano gels can carry drugs well, promote their release in acidic environments, and increase intracellular drug concentrations [[93](#page-14-28)]. CD44, a receptor of hyaluronic acid, has higher expression in many tumor cells than in normal tissues and is associated with the tumorigenicity, invasiveness, and lymphatic metastasis of tumor cells. Zhao et al. [[94](#page-14-29)] found that hyaluronic acid-FA could effectively inhibit the growth of Lewis lung cancer tissue, and its inhibitory effect was comparable to that of cyclophosphamide, indicating that hyaluronic acid-FA targeted drugs could deliver drugs to cells with high CD44 expression through a receptor-mediated mechanism, thus achieving the purpose of enhancing drug efficacy.

Ganesan et al. [[95\]](#page-14-30) studied the regulatory effect of FA on P-gp-mediated multidrug resistance (MDR). The results showed that FA could enhance the cytotoxicity of alamycin and vincristine in KB-Ch(R)-8-5 cells overexpressing P-gp. In addition, FA combined with adriamycin significantly reduced the tumor volume of KB-Ch(R)-8-5 xenogeneic tumors compared with adriamycin alone. Previous studies showed that activation of NF-κB by the PI3K/AKT pathway may lead to increased transcription of MDR1 [[96\]](#page-14-31). It is suggested that FA can reverse MDR by inhibiting the expression of P-gp through inhibition of the PI3K/Akt/NF-κB signaling pathway. Rajeshkumar et al. [[97](#page-14-32)] prepared a nanopolymer of FA and paclitaxel (PTX) supported polyamidoamine (PAMAM) dendrimer coupled with arginine-glycine aspartic acid (RGD). The levels of proapoptotic factors such as caspase 3, caspase 9, p53, and Bax were increased, and the antiapoptotic factors were downregulated in the drug-resistant KB-CH(R)-8-5 cells treated with RGD-PAMAM-FP. The results showed that the polymer could overcome P-gp-mediated MDR. Trans FA could reduce the side effects of doxorubicin and cyclophosphamide, protect the normal function of the heart, liver, and bone, and restore frontal chemotherapy sensitivity in breast cancer model mice by reducing the level of P-gp [[98\]](#page-14-33).

Oxaliplatin is a commonly used chemotherapy drug for the treatment of colorectal cancer. Zhang et al. [[99\]](#page-15-0) found that FA can significantly improve the sensitivity of drug-resistant HCT116 colorectal cancer cells to oxaliplatin (OXA). The mechanism is that FA interferes with the expression of the MDR1 gene by blocking the catabolism of ganglioside acid (GM3). Neuraminidase 3 (NEU3) can catalyze the conversion of GM3 into ceramide trihexosides (Gb3). FA inhibits the activity of NEU3 and then reduces the levels of Gb3 and P-gp, thus improving the sensitivity of drug-resistant cells to OXA chemotherapy. These results suggest that FA can increase chemotherapy sensitivity by remodeling Neu3-mediated GM3 ganglioside metabolism and nay become a new drug to promote the efficacy of chemotherapy in colorectal cancer.

Other antitumor efects

Autophagy

Autophagy is the basis of cell component degradation and recycling. Autophagy is closely related to malignant transformation, tumor progression, and treatment [[100\]](#page-15-1). FA treatment can enhance autophagy and increase the expression of autophagy-related proteins, resulting in the inhibition of mTOR signal transduction [\[101](#page-15-2)].

Wang et al. [[102](#page-15-3)] found that FA could increase the levels of apoptosis and the autophagy biomarkers PINK-1 and Parkin in hepatocellular carcinoma HepG2 cells, suggesting that FA could inhibit the proliferation of HepG2 cells and increase their apoptosis and autophagy. Autophagy is the core target of FA in the treatment of osteosarcoma. Pang et al. [\[103](#page-15-4)] identifed activators of signal transducer and activator of transcription 3 (STAT3), mitogen-activated protein kinase 1 (MAPK1), and phosphoinositin-3 kinase regulatory subunit 1 (PIK3R1) as pharmacological targets of FA in the treatment of osteosarcoma through molecular docking analysis. These fndings from bioinformatics analysis demonstrate the potential therapeutic role of FA in the treatment of osteosarcoma. Claudia et al. [\[104\]](#page-15-5) synthesized a novel compound, FA tributyltin, based on FA and evaluated its efects on colon cancer cells. The results showed that the compound could reduce the activity of colon cancer HCT116, HT-29, and Caco-2 cells. Further studies showed that the decrease in cell viability induced by this compound was associated with G_2/M cell cycle arrest, a process that hardly involved changes in apoptosis or necrotizing apoptotic markers. Autophagy vacuoles and increased LC3-II and p62 autophagy proteins were observed after treatment with this compound, suggesting that the process of cell death is triggered by autophagy. These results suggest that FA derivatives are promising antitumor agents that can trigger autophagy, which may be very important in the case of resistance to the classic apoptosis process.

However, autophagy can be a double-edged sword in different stages of cancer development. In early tumorigenesis, autophagy, as a survival pathway and quality control mechanism, can prevent tumorigenesis and inhibit cancer progression. Once the tumor progresses to an advanced stage, autophagy will instead contribute to the survival and growth of the tumor and promote the aggressiveness of cancer by promoting metastasis [\[105](#page-15-6)]. Autophagy plays roles in both tumor inhibition and promotion, so the exact mechanism of autophagy in cancer remains to be further studied.

Chemical prophylaxis

Cancer chemoprophylaxis refers to the strategy of using natural or synthetic chemicals to prevent, reverse or slow

down the development of cancer, which is a hot research feld in cancer prevention. FA can prevent buccal pouch carcinogenesis induced by 7,12-dimethylbenzene [a] anthracene (DMBA) in hamsters. Oral FA can signifcantly reduce the incidence, volume, and weight of tumors in hamsters, suggesting that FA has an efective chemopreventive efect on DMBA-induced buccal pouch carcinoma in hamsters [\[106](#page-15-7)].

The occurrence of skin cancer is associated with ultraviolet-b radiation (UVB) exposure $[107]$ $[107]$. In a study by Amboth [[108](#page-15-9)], FA significantly reduced the incidence, volume, and weight of UVB-induced tumors in the skin of mice compared to control animals. It was also observed that FA therapy could reverse the chronic oxidative damage of UVB-induced skin tumors in mice while regulating the expression of VEGF, inducible nitric oxide synthase (iNOS), tumor necrosis factor- α (TNF- α), and IL-6. FA treatment also modulated the expression of mutated p53, Bcl-2, and Bax in UVB-induced mouse skin tumors. The results showed that FA had a potential inhibitory efect on UVB-induced carcinogenesis in albino mice. In addition, FA protects against the cytotoxicity induced by cyclophosphamide in neuroblastoma SH-SY5Y cells by reducing lipid peroxidation levels [\[109](#page-15-10)].

Aerobic glycolysis

Metabolic changes are the hallmark of cancer, and reprogramming of energy metabolism is a common phenomenon in tumor progression. The change in aerobic glycolysis during the energy metabolism of cancer cells is a signa-ture feature, also known as the Warburg effect [\[110](#page-15-11)]. Long noncoding RNAs (lncRNAs) regulate energy metabolism in cancer [\[111](#page-15-12)]. Cui et al. [[112](#page-15-13)] found that the proliferation of HCT116 and HT-29 cells was inhibited after FA treatment. Further studies showed that FA could inhibit the expression of PKM2 and block aerobic glycolysis. Since PKM2 was positively correlated with lncRNA 495810 expression in this study, it was speculated that FA might inhibit aerobic glycolysis through lncRNA 495810/PKM2 signal transduction. The antitumor effects of FA and its derivatives are shown in Table [1](#page-11-0).

In addition, Priya et al. [[113](#page-15-14)] prepared polyethylene oxide nanofbres by electrospinning technology, and these nanofbres could continuously release efective active substances within 24 h of administration and showed a certain apoptotic efect on breast cancer MCF-7 cells. Tetrahydroisoquinoline plays an important role in the feld of pharmaceutical chemistry due to its extensive pharmacological properties, making it an important scafold for the design of anticancer drugs and a hot topic in current research [[114](#page-15-15)]. Wang et al. [[115](#page-15-16)] synthesized tetrahydroisoquinoline FA derivatives, which were detected by the SRB method to have certain in vitro antitumor activity in colon cancer HT-29 cells and breast cancer MCF-7 cells. Eldin et al. [[116](#page-15-17)] grafted FA onto oligosaccharides, which were then self-assembled into particles. In vitro experiments showed that FA-grafted oligosaccharides have high anticancer activity against human colon cancer HT-29 and LoVo cells, suggesting that FA-grafted oligosaccharides are a good candidate material for relieving colorectal cancer. These results all indicated that FA-related derivatives had excellent antitumor functions but failed to elucidate the specifc antitumor mechanism.

Existing problems and prospects

As a phenolic substance widely present in plant cell walls, FA has been shown to have a broad-spectrum antitumor efect, inhibiting lung cancer, liver cancer, breast cancer, cervical cancer, colorectal cancer, and other related cancers. The anticancer effects of FA are mainly achieved by inhibiting the proliferation of tumor cells, inducing the apoptosis of tumor cells, inhibiting the invasion and migration of tumor cells, and enhancing the efficacy of chemoradiotherapy drugs. In addition, FA can play a role by regulating immune function [\[19,](#page-12-16) [117](#page-15-18)], inducing autophagy, and inhibiting the drug resistance of tumor cells. With the continuous development and improvement of experimental technology, data mining, bioinformatics, and other technologies, the antitumor effect of FA has been fully confirmed in experiments, and its pharmacodynamic mechanism has also been elaborated at multiple levels.

However, there are still some obstacles to the application of FA components as drugs in clinical treatment. On the one hand, FA has poor solubility in water and a poor ability to pass through biological barriers [\[118](#page-15-19)]; therefore, the extent to which it is metabolized in vivo after oral administration is largely unknown, and its absorption status in vivo and concentration in the target site is not easy to determine. Nanocarriers can overcome the restriction of drug action by body barriers (such as the blood–brain barrier and blood–eye barrier) and can reduce drug doses, improve drug availability and reduce side efects after modifcation by targeted groups. However, there is a lack of research on improving the bioavailability of monomer components and targeting them specifcally to tumor tissue through chemical modifcation. On the other hand, we noted that there were signifcant diferences in the doses used in the antitumor studies of FA, which were still in the exploratory stage and failed to form a unifed standard. The above problems may be the reason why FA and its related components have not been developed into clinical drugs. Therefore, in future research, it is necessary to develop a carrier that can selectively deliver drugs to certain parts of the body, identify mutant cells, control the release of drugs, determine the safe and efective dose of FA, and develop a more efective drug regimen to serve

Table 1 The antitumor effects of ferulic acid and its derivatives

cancer patients more efficiently. This still requires further experimental exploration and research.

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Declarations

Conflict of interest The authors declare that they have no competing interests.

Ethical approval Not applicable.

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References

- 1. Qiu H, Cao S, Xu R. Cancer incidence, mortality, and burden in China: a time-trend analysis and comparison with the United States and United Kingdom based on the global epidemiological data released in 2020. Cancer Commun (Lond). 2021;41(10):1037–48.
- 2. Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. CA Cancer J Clin. 2023;73(1):17–48.
- 3. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209–49.
- 4. Zhang Y, Chen L, Hu GQ, Zhang N, Zhu XD, Yang KY, et al. Gemcitabine and cisplatin induction chemotherapy in nasopharyngeal carcinoma. N Engl J Med. 2019;381(12):1124–35.
- 5. Chen R, Manochakian R, James L, Azzouqa AG, Shi H, Zhang Y, et al. Emerging therapeutic agents for advanced non-small cell lung cancer. J Hematol Oncol. 2020;13(1):58.
- 6. Cavalcanti GR, Duarte FIC, Converti A, de Lima ÁAN. Ferulic acid activity in topical formulations: technological and scientifc prospecting. Curr Pharm Des. 2021;27(19):2289–98.
- 7. Bento-Silva A, Vaz Patto MC, do Rosário Bronze M. Relevance, structure and analysis of ferulic acid in maize cell walls. Food Chem. 2018;246:360–78.
- 8. Babbar R, Dhiman S, Grover R, Kaur A, Arora S. A comprehensive review on therapeutic applications of ferulic acid and its novel analogues: a brief literature. Mini Rev Med Chem. 2021;21(12):1578–93.
- 9. Zduńska K, Dana A, Kolodziejczak A, Rotsztejn H. Antioxidant properties of ferulic acid and its possible application. Skin Pharmacol Physiol. 2018;31(6):332–6.
- 10. Chaudhary A, Jaswal VS, Choudhary S, Sharma A, Beniwal V, et al. Ferulic acid: a promising therapeutic phytochemical and recent patents advances. Recent Pat Infamm Allergy Drug Discov. 2019;13(2):115–23.
- 11. Neto-Neves EM, da Silva Maia Bezerra Filho C, Dejani NN, de Sousa DP. ferulic acid and cardiovascular health: therapeutic and preventive potential. Mini Rev Med Chem. 2021;21(13):1625–37.
- 12. Shi Y, Chen X, Qiang S, Su J, Li J. Anti-oxidation and antiinflammatory potency evaluation of ferulic acid derivatives obtained through virtual screening. Int J Mol Sci. 2021;22(21):11305.
- 13. Nouri A, Ghatreh-Samani K, Amini-Khoei H, Mohammadi A, Heidarian E, Najaf M. Ferulic acid prevents cyclosporineinduced nephrotoxicity in rats through exerting anti-oxidant and

anti-infammatory efects via activation of Nrf2/HO-1 signaling and suppression of NF-κB/TNF-α axis. Naunyn Schmiedebergs Arch Pharmacol. 2022;395(4):387–95.

- 14. Li X, Wu J, Xu F, Chu C, Li X, Shi X, et al. Use of ferulic acid in the management of diabetes mellitus and its complications. Molecules. 2022;27(18):6010.
- 15. Zhao J, Gao J, Li H. Ferulic acid confers protection on islet β cells and placental tissues of rats with gestational diabetes mellitus. Cell Mol Biol (Noisy-le-grand). 2020;66(1):37–41.
- 16. Alam MA. Anti-hypertensive efect of cereal antioxidant ferulic acid and its mechanism of action. Front Nutr. 2019;6:121.
- 17. Cheng Q, Li C, Yang CF, Zhong YJ, Wu D, Shi L, et al. Methyl ferulic acid attenuates liver fbrosis and hepatic stellate cell activation through the TGF-β1/Smad and NOX4/ROS pathways. Chem Biol Interact. 2019;299:131–9.
- 18. Cao L, Li Z, Yang Z, Wang M, Zhang W, Ren Y, et al. Ferulic acid positively modulates the infammatory response to septic liver injury through the GSK-3β/NF-κB/CREB pathway. Life Sci. 2021;277: 119584.
- 19. Sin Singer Brugiolo A, Carvalho Gouveia AC, de Souza Alves CC, de Castro ESFM, Esteves de Oliveira É, Ferreira AP. Ferulic acid supresses Th2 immune response and prevents remodeling in ovalbumin-induced pulmonary allergy associated with inhibition of epithelial-derived cytokines. Pulm Pharmacol Ther. 2017;45:202–9.
- 20. Ahmadifar E, Moghadam MS, Dawood MAO, Hoseinifar SH. *Lactobacillus fermentum* and/or ferulic acid improved the immune responses, antioxidative defence and resistance against *Aeromonas hydrophila* in common carp (*Cyprinus carpio*) fngerlings. Fish Shellfsh Immunol. 2019;94:916–23.
- 21. Choi YE, Park E. Ferulic acid in combination with PARP inhibitor sensitizes breast cancer cells as chemotherapeutic strategy. Biochem Biophys Res Commun. 2015;458(3):520–4.
- 22. Serafm TL, Carvalho FS, Marques MP, Calheiros R, Silva T, Garrido J, et al. Lipophilic cafeic and ferulic acid derivatives presenting cytotoxicity against human breast cancer cells. Chem Res Toxicol. 2011;24(5):763–74.
- 23. Wang B, Zhang W, Zhou X, Liu M, Hou X, Cheng Z, et al. Development of dual-targeted nano-dandelion based on an oligomeric hyaluronic acid polymer targeting tumor-associated macrophages for combination therapy of non-small cell lung cancer. Drug Deliv. 2019;26(1):1265–79.
- 24. Nasr Bouzaiene N, Kilani Jaziri S, Kovacic H, Chekir-Ghedira L, Ghedira K, Luis J. The efects of cafeic, coumaric and ferulic acids on proliferation, superoxide production, adhesion and migration of human tumor cells in vitro. Eur J Pharmacol. 2015;766:99–105.
- 25. Morana O, Wood W, Gregory CD. The apoptosis paradox in cancer. Int J Mol Sci. 2022;23(3):1328.
- 26. Hu J, Cao J, Topatana W, Juengpanich S, Li S, Zhang B, et al. Targeting mutant p53 for cancer therapy: direct and indirect strategies. J Hematol Oncol. 2021;14(1):157.
- 27. Zhang C, Liu J, Xu D, Zhang T, Hu W, Feng Z. Gain-of-function mutant p53 in cancer progression and therapy. J Mol Cell Biol. 2020;12(9):674–87.
- 28. Niu YX, Zhang Y, Wei LQ, Xu Y. The inducing efects of ferulic acid on the apoptosis of gastric cancer SGC-7901 cells and lts infuence on COX-2, survivin, XIAP and p53. Western J Tradit Chin Med. 2019;32(01):19–23.
- 29. Fahrioğlu U, Dodurga Y, Elmas L, Seçme M. Ferulic acid decreases cell viability and colony formation while inhibiting migration of MIA PaCa-2 human pancreatic cancer cells in vitro. Gene. 2016;576(1 Pt 3):476–82.
- 30. Myte R, Gylling B, Schneede J, Ueland PM, Häggström J, Hultdin J, et al. Components of one-carbon metabolism

other than folate and colorectal cancer risk. Epidemiology. 2016;27(6):787–96.

- 31. Senthil Kumar C, Thangam R, Mary SA, Kannan PR, Arun G, Madhan B. Targeted delivery and apoptosis induction of trans-resveratrol-ferulic acid loaded chitosan coated folic acid conjugate solid lipid nanoparticles in colon cancer cells. Carbohydr Polym. 2020;231: 115682.
- 32. Zhang L, Liao Y, Tang L. MicroRNA-34 family: a potential tumor suppressor and therapeutic candidate in cancer. J Exp Clin Cancer Res. 2019;38(1):53.
- 33. Huang Q, Zheng Y, Ou Y, Xiong H, Yang H, Zhang Z, et al. Retraction notice to "miR-34a/Bcl-2 signaling pathway contributes to age-related hearing loss by modulating hair cell apoptosis" [Neurosci. Lett. 661 (November) (2017) 51-56]. Neurosci Lett. 2019;707: 134290.
- 34. Fu J, Wang H. Inhibitory efect of ferulic acid mediated by miR-34a on transplanted tumor of human cervical cancer in nude mice. Chin J Gerontol. 2021;41(13):2833–5. [https://doi.org/10.](https://doi.org/10.3969/j.issn.1005-9202.2021.13.041) [3969/j.issn.1005-9202.2021.13.041.](https://doi.org/10.3969/j.issn.1005-9202.2021.13.041)
- 35. Zhang Y, Li HL, Wang HP, Gu J, Ma CL, Wu HY. Efects of ferulic acid on gastric cancer cell line MGC-803 proliferation. Chin J Inf Tradit Chin Med. 2016;23(09):70–3. [https://doi.org/](https://doi.org/10.3969/j.issn.1005-5304.2016.09.017) [10.3969/j.issn.1005-5304.2016.09.017](https://doi.org/10.3969/j.issn.1005-5304.2016.09.017).
- 36. Zhang QF, Liu W, Liu JX, Qiu J, Zhang Y, Tian CH, et al. Efect of isoferulic acid on inhibiting cell proliferation and inducing cell apoptosis in human renal carcinoma cell A-498. J Clin Urol. 2021;36(07):549–54. [https://doi.org/10.13201/j.issn.1001-1420.](https://doi.org/10.13201/j.issn.1001-1420.2021.07.009) [2021.07.009](https://doi.org/10.13201/j.issn.1001-1420.2021.07.009).
- 37. Yue SJ, Zhang PX, Zhu Y, Li NG, Chen YY, Li JJ, et al. A ferulic acid derivative FXS-3 inhibits proliferation and metastasis of human lung cancer A549 cells via positive JNK signaling pathway and negative ERK/p38, AKT/mTOR and MEK/ERK signaling pathways. Molecules. 2019;24(11):2165.
- 38. Sun F, Wang Y, Luo X, Ma Z, Xu Y, Zhang X, et al. Anti-CD24 antibody-nitric oxide conjugate selectively and potently suppresses hepatic carcinoma. Cancer Res. 2019;79(13):3395–405.
- 39. Moloney JN, Cotter TG. ROS signalling in the biology of cancer. Semin Cell Dev Biol. 2018;80:50–64.
- 40. Cao Y, Zhang H, Tang J, Wang R. Ferulic acid mitigates growth and invasion of esophageal squamous cell carcinoma through inducing ferroptotic cell death. Dis Markers. 2022;2022:4607966.
- 41. Grasso R, Dell'Albani P, Carbone C, Spatuzza M, Bonfanti R, Sposito G, et al. Synergic pro-apoptotic effects of ferulic acid and nanostructured lipid carrier in glioblastoma cells assessed through molecular and delayed luminescence studies. Sci Rep. 2020;10(1):4680.
- 42. Singh TA, Das J, Sil PC. Zinc oxide nanoparticles: a comprehensive review on its synthesis, anticancer and drug delivery applications as well as health risks. Adv Colloid Interface Sci. 2020;286: 102317.
- 43. Ezhuthupurakkal PB, Ariraman S, Arumugam S, Subramaniyan N, Muthuvel SK, Kumpati P, et al. Anticancer potential of ZnO nanoparticle-ferulic acid conjugate on Huh-7 and HepG2 cells and diethyl nitrosamine induced hepatocellular cancer on Wistar albino rat. Nanomedicine. 2018;14(2):415–28.
- 44. Stoimenov I, Helleday T. PCNA on the crossroad of cancer. Biochem Soc Trans. 2009;37(Pt 3):605–13.
- 45. Cardano M, Tribioli C, Prosperi E. Targeting proliferating cell nuclear antigen (PCNA) as an efective strategy to inhibit tumor cell proliferation. Curr Cancer Drug Targets. 2020;20(4):240–52.
- 46. Horsfall AJ, Abell AD, Bruning JB. Targeting PCNA with peptide mimetics for therapeutic purposes. ChemBioChem. 2020;21(4):442–50.
- 47. Thakkar A, Chenreddy S, Wang J, Prabhu S. Ferulic acid combined with aspirin demonstrates chemopreventive potential towards pancreatic cancer when delivered using chitosan-coated solid-lipid nanoparticles. Cell Biosci. 2015;5:46.
- 48. Díaz-Coránguez M, Liu X, Antonetti DA. Tight junctions in cell proliferation. Int J Mol Sci. 2019;20(23):5972.
- 49. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646–74.
- 50. Matthews HK, Bertoli C, de Bruin RAM. Cell cycle control in cancer. Nat Rev Mol Cell Biol. 2022;23(1):74–88.
- 51. Wang K, Liao GJ, Liu WJ. Efect of ferulic acid on proliferation, invasion and apoptosis of osteosarcoma cells. Chin J Exp Surg. 2019;01:51–3. [https://doi.org/10.3760/cma.j.issn.1001-9030.](https://doi.org/10.3760/cma.j.issn.1001-9030.2019.01.016) [2019.01.016.](https://doi.org/10.3760/cma.j.issn.1001-9030.2019.01.016)
- 52. Wang T, Gong X, Jiang R, Li H, Du W, Kuang G. Ferulic acid inhibits proliferation and promotes apoptosis via blockage of PI3K/Akt pathway in osteosarcoma cell. Am J Transl Res. 2016;8(2):968–80.
- 53. Eroğlu C, Seçme M, Bağcı G, Dodurga Y. Assessment of the anticancer mechanism of ferulic acid via cell cycle and apoptotic pathways in human prostate cancer cell lines. Tumour Biol. 2015;36(12):9437–46.
- 54. Luo Y, Wang CZ, Sawadogo R, Yuan J, Zeng J, Xu M, et al. 4-vinylguaiacol, an active metabolite of ferulic acid by enteric microbiota and probiotics, possesses signifcant activities against drug-resistant human colorectal cancer cells. ACS Omega. 2021;6(7):4551–61.
- 55. Gao J, Yu H, Guo W, Kong Y, Gu L, Li Q, et al. The anticancer efects of ferulic acid is associated with induction of cell cycle arrest and autophagy in cervical cancer cells. Cancer Cell Int. 2018;18:102.
- 56. Al-Mutairi A, Rahman A, Rao MS. Low doses of thymoquinone and ferulic acid in combination efectively inhibit proliferation of cultured MDA-MB 231 breast adenocarcinoma cells. Nutr Cancer. 2021;73(2):282–9.
- 57. Owen KL, Brockwell NK, Parker BS. JAK-STAT signaling: a double-edged sword of immune regulation and cancer progression. Cancers (Basel). 2019;11(12):2002.
- 58. Guo F, Zhao RM, Li JL, Liu YX, Yang SJ. Ferulic acid inhibits lung cancer cell proliferation and metastasis by regulating JAK2/STAT6 immune signaling pathway. Chin J Immunol. 2021;37(04):459–62. [https://doi.org/10.3969/j.issn.1000-484X.](https://doi.org/10.3969/j.issn.1000-484X.2021.04.015) [2021.04.015.](https://doi.org/10.3969/j.issn.1000-484X.2021.04.015)
- 59. Yu H, Lin L, Zhang Z, Zhang H, Hu H. Targeting NF-κB pathway for the therapy of diseases: mechanism and clinical study. Signal Transduct Target Ther. 2020;5(1):209.
- 60. Wu X, Lin L, Wu H. Ferulic acid alleviates lipopolysaccharideinduced acute lung injury through inhibiting TLR4/NF-κB signaling pathway. J Biochem Mol Toxicol. 2021;35(3): e22664.
- 61. Maruyama H, Kawakami F, Lwin TT, Imai M, Shamsa F. Biochemical characterization of ferulic acid and cafeic acid which efectively inhibit melanin synthesis via diferent mechanisms in B16 melanoma cells. Biol Pharm Bull. 2018;41(5):806–10.
- 62. Xu Y, Zuo W, Wang X, Zhang Q, Gan X, Tan N, et al. Deciphering the efects of PYCR1 on cell function and its associated mechanism in hepatocellular carcinoma. Int J Biol Sci. 2021;17(9):2223–39.
- 63. Yang RP, An N, Shan SH, Shi JY, Li HQ, He SL, et al. 569 Ferulic acid binds to pyrroline-5-carboxylate reductase 1 to inhibit proliferation of breast cancer cells. Chin J Biochem Mol Biol. 2023;39(01):79–86. [https://doi.org/10.13865/j.cnki.cjbmb.2022.](https://doi.org/10.13865/j.cnki.cjbmb.2022.10.1407) [10.1407.](https://doi.org/10.13865/j.cnki.cjbmb.2022.10.1407)
- 64. Xu F, Na L, Li Y, Chen L. Roles of the PI3K/AKT/mTOR signalling pathways in neurodegenerative diseases and tumours. Cell Biosci. 2020;10(1):54.
- 65. Li D, Rui YX, Guo SD, Luan F, Liu R, Zeng N. Ferulic acid: a review of its pharmacology, pharmacokinetics and derivatives. Life Sci. 2021;284: 119921.
- 66. Luo L, Zhu S, Tong Y, Peng S. Ferulic acid induces apoptosis of HeLa and Caski cervical carcinoma cells by down-regulating the phosphatidylinositol 3-kinase (PI3K)/Akt signaling pathway. Med Sci Monit. 2020;26: e920095.
- 67. Wu J, Wang YX, Wei N, Wu XZ, Liu XE. Ferulic acid inhibits lung cancer growth and its mechanism. Zhejiang Med J. 2018;40(12):1303–6. [https://doi.org/10.12056/j.issn.1006-2785.](https://doi.org/10.12056/j.issn.1006-2785.2018.40.12.2017-2134) [2018.40.12.2017-2134.](https://doi.org/10.12056/j.issn.1006-2785.2018.40.12.2017-2134)
- 68. Fares J, Fares MY, Khachfe HH, Salhab HA, Fares Y. Molecular principles of metastasis: a hallmark of cancer revisited. Signal Transduct Target Ther. 2020;5(1):28.
- 69. Itatani Y, Kawada K, Yamamoto T, Sakai Y. Resistance to anti-angiogenic therapy in cancer-alterations to anti-VEGF pathway. Int J Mol Sci. 2018;19(4):1232.
- 70. Frezzetti D, Gallo M, Maiello MR, D'Alessio A, Esposito C, Chicchinelli N, et al. VEGF as a potential target in lung cancer. Expert Opin Ther Targets. 2017;21(10):959–66.
- 71. Chen Y, Zhang L, Liu WX, Wang K. VEGF and SEMA4D have synergistic efects on the promotion of angiogenesis in epithelial ovarian cancer. Cell Mol Biol Lett. 2018;23:2.
- 72. Zhang XD, Wu Q, Yang SH. Ferulic acid promoting apoptosis in human osteosarcoma cell lines. Pak J Med Sci. 2017;33(1):127–31.
- 73. El-Gogary RI, Nasr M, Rahsed LA, Hamzawy MA. Ferulic acid nanocapsules as a promising treatment modality for colorectal cancer: preparation and in vitro/in vivo appraisal. Life Sci. 2022;298: 120500.
- 74. Wang D, Wang C, Sun L, Lu X, Shi J, Chen J, et al. MiR-143–3p increases the radiosensitivity of breast cancer cells through FGF1. Cell Mol Biol (Noisy-le-grand). 2022;67(5):256–62.
- 75. Henriksson ML, Edin S, Dahlin AM, Oldenborg PA, Öberg Å, Van Guelpen B, et al. Colorectal cancer cells activate adjacent fbroblasts resulting in FGF1/FGFR3 signaling and increased invasion. Am J Pathol. 2011;178(3):1387–94.
- 76. Yang GW, Jiang JS, Lu WQ. Ferulic acid exerts anti-angiogenic and anti-tumor activity by targeting fbroblast growth factor receptor 1-mediated angiogenesis. Int J Mol Sci. 2015;16(10):24011–31.
- 77. Chen HT, Liu H, Mao MJ, Tan Y, Mo XQ, Meng XJ, et al. Crosstalk between autophagy and epithelial-mesenchymal transition and its application in cancer therapy. Mol Cancer. 2019;18(1):101.
- 78. Pastushenko I, Blanpain C. EMT transition states during tumor progression and metastasis. Trends Cell Biol. 2019;29(3):212–26.
- 79. Zhang X, Lin D, Jiang R, Li H, Wan J, Li H. Ferulic acid exerts antitumor activity and inhibits metastasis in breast cancer cells by regulating epithelial to mesenchymal transition. Oncol Rep. 2016;36(1):271–8.
- 80. Gobin E, Bagwell K, Wagner J, Mysona D, Sandirasegarane S, Smith N, et al. A pan-cancer perspective of matrix metalloproteases (MMP) gene expression profle and their diagnostic/ prognostic potential. BMC Cancer. 2019;19(1):581.
- 81. Park HJ, Cho JH, Hong SH, Kim DH, Jung HY, Kang IK, et al. Whitening and anti-wrinkle activities of ferulic acid isolated from *Tetragonia tetragonioides* in B16F10 melanoma and CCD-986sk fbroblast cells. J Nat Med. 2018;72(1):127–35.
- 82. Dodurga Y, Eroğlu C, Seçme M, Elmas L, Avcı ÇB, Şatıroğlu-Tufan NL. Anti-proliferative and anti-invasive efects of ferulic acid in TT medullary thyroid cancer cells interacting with URG4/ URGCP. Tumour Biol. 2016;37(2):1933–40.
- 83. Ward C, Meehan J, Gray M, Kunkler IH, Langdon SP, Argyle DJ. Carbonic anhydrase IX (CAIX), cancer, and radiation responsiveness. Metabolites. 2018;8(1):13.
- 84. Takacova M, Kajanova I, Kolarcikova M, Lapinova J, Zatovicova M, Pastorekova S. Understanding metabolic alterations and heterogeneity in cancer progression through validated immunodetection of key molecular components: a case of carbonic anhydrase IX. Cancer Metastasis Rev. 2021;40(4):1035–53.
- 85. Aneja B, Queen A, Khan P, Shamsi F, Hussain A, Hasan P, et al. Design, synthesis & biological evaluation of ferulic acid-based small molecule inhibitors against tumor-associated carbonic anhydrase IX. Bioorg Med Chem. 2020;28(9): 115424.
- 86. Mu Q, Najaf M. Resveratrol for targeting the tumor microenvironment and its interactions with cancer cells. Int Immunopharmacol. 2021;98: 107895.
- 87. Sawata Y, Matsukawa T, Doi S, Tsunoda T, Arikawa N, Matsunaga N, et al. A novel compound, ferulic acid-bound resveratrol, induces the tumor suppressor gene p15 and inhibits the threedimensional proliferation of colorectal cancer cells. Mol Cell Biochem. 2019;462(1–2):25–31.
- 88. Karthikeyan S, Kanimozhi G, Prasad NR, Mahalakshmi R. Radiosensitizing efect of ferulic acid on human cervical carcinoma cells in vitro. Toxicol In Vitro. 2011;25(7):1366–75.
- 89. Das U, Manna K, Adhikary A, Mishra S, Saha KD, Sharma RD, et al. Ferulic acid enhances the radiation sensitivity of lung and liver carcinoma cells by collapsing redox homeostasis: mechanistic involvement of Akt/p38 MAPK signalling pathway. Free Radic Res. 2019;53(9–10):944–67.
- 90. Zheng Y, You X, Chen L, Huang J, Wang L, Wu J, et al. Biotherapeutic nanoparticles of poly(ferulic acid) delivering doxorubicin for cancer therapy. J Biomed Nanotechnol. 2019;15(8):1734–43.
- 91. Muthusamy G, Balupillai A, Ramasamy K, Shanmugam M, Gunaseelan S, Mary B, et al. Ferulic acid reverses ABCB1-mediated paclitaxel resistance in MDR cell lines. Eur J Pharmacol. 2016;786:194–203.
- 92. Paciello F, Fetoni AR, Mezzogori D, Rolesi R, Di Pino A, Paludetti G, et al. The dual role of curcumin and ferulic acid in counteracting chemoresistance and cisplatin-induced ototoxicity. Sci Rep. 2020;10(1):1063.
- 93. Cheng X, Hu T, Li C, Shi S, Xu Y, Jia C, et al. Acid-sensitive and L61-crosslinked hyaluronic acid nanogels for overcoming tumor drug-resistance. Int J Biol Macromol. 2021;188:11–23.
- 94. Zhao YX, Wei Y, Chen Z, Zhang SP, Liu F. The anti tumor efect of ferulic acid decoreted hyaluronic acid on spontaneous lung metastasis model mice of lewis lung cancer. Guangdong Chem Ind. 2018;45(13):30–1. [https://doi.org/10.3969/j.issn.1007-1865.](https://doi.org/10.3969/j.issn.1007-1865.2018.13.015) [2018.13.015.](https://doi.org/10.3969/j.issn.1007-1865.2018.13.015)
- 95. Muthusamy G, Gunaseelan S, Prasad NR. Ferulic acid reverses P-glycoprotein-mediated multidrug resistance via inhibition of PI3K/Akt/NF-κB signaling pathway. J Nutr Biochem. 2019;63:62–71.
- 96. Xi G, Hayes E, Lewis R, Ichi S, Mania-Farnell B, Shim K, et al. CD133 and DNA-PK regulate MDR1 via the PI3K- or Akt-NF-κB pathway in multidrug-resistant glioblastoma cells in vitro. Oncogene. 2016;35(42):5576.
- 97. Anbazhagan R, Muthusamy G, Krishnamoorthi R, Kumaresan S, Rajendra Prasad N, Lai JY, et al. PAMAM G4.5 dendrimers for targeted delivery of ferulic acid and paclitaxel to overcome P-glycoprotein-mediated multidrug resistance. Biotechnol Bioeng. 2021;118(3):1213–23.
- 98. Helmy SA, El-Mofty S, El Gayar AM, El-Sherbiny IM, El-Far YM. Novel doxorubicin/folate-targeted trans-ferulic acid-loaded PLGA nanoparticles combination: in-vivo superiority over standard chemotherapeutic regimen for breast cancer treatment. Biomed Pharmacother. 2022;145: 112376.
- 99. Zhang X, Shan S, Shi J, Li H, Li Z. Polyphenol from millet bran increases the sensitivity of colorectal cancer cells to oxaliplatin by blocking the ganglioside GM3 catabolism. Food Funct. 2021;12(1):291–301.
- 100. Yamazaki T, Bravo-San Pedro JM, Galluzzi L, Kroemer G, Pietrocola F. Autophagy in the cancer-immunity dialogue. Adv Drug Deliv Rev. 2021;169:40–50.
- 101. Li C, Chen L, Song M, Fang Z, Zhang L, Coffie JW, et al. Ferulic acid protects cardiomyocytes from TNF-α/cycloheximideinduced apoptosis by regulating autophagy. Arch Pharm Res. 2020;43(8):863–74.
- 102. Wang J, Lai X, Yuan D, Liu Y, Wang J, Liang Y. Efects of ferulic acid, a major component of rice bran, on proliferation, apoptosis, and autophagy of HepG2 cells. Food Res Int. 2022;161: 111816.
- 103. Pang G, Yi T, Luo H, Jiang L. Preclinical fndings: the pharmacological targets and molecular mechanisms of ferulic acid treatment for COVID-19 and osteosarcoma via targeting autophagy. Front Endocrinol (Lausanne). 2022;13: 971687.
- 104. Pellerito C, Emanuele S, Ferrante F, Celesia A, Giuliano M, Fiore T. Tributyltin(IV) ferulate, a novel synthetic ferulic acid derivative, induces autophagic cell death in colon cancer cells: From chemical synthesis to biochemical efects. J Inorg Biochem. 2020;205: 110999.
- 105. Li X, He S, Ma B. Autophagy and autophagy-related proteins in cancer. Mol Cancer. 2020;19(1):12.
- 106. Balakrishnan S, Menon VP, Manoharan S. Ferulic acid inhibits 7,12-dimethylbenz[a]anthracene-induced hamster buccal pouch carcinogenesis. J Med Food. 2008;11(4):693–700.
- 107. Willenbrink TJ, Ruiz ES, Cornejo CM, Schmults CD, Arron ST, Jambusaria-Pahlajani A. Field cancerization: defnition, epidemiology, risk factors, and outcomes. J Am Acad Dermatol. 2020;83(3):709–17.
- 108. Ambothi K, Prasad NR, Balupillai A. Ferulic acid inhibits UVB-radiation induced photocarcinogenesis through modulating infammatory and apoptotic signaling in Swiss albino mice. Food Chem Toxicol. 2015;82:72–8.
- 109. Ayna A, Özbolat SN, Darendelioglu E. Quercetin, chrysin, cafeic acid and ferulic acid ameliorate cyclophosphamide-induced toxicities in SH-SY5Y cells. Mol Biol Rep. 2020;47(11):8535–43.
- 110. Wu Z, Wu J, Zhao Q, Fu S, Jin J. Emerging roles of aerobic glycolysis in breast cancer. Clin Transl Oncol. 2020;22(5):631–46.
- 111. Tan YT, Lin JF, Li T, Li JJ, Xu RH, Ju HQ. LncRNA-mediated posttranslational modifications and reprogramming of energy metabolism in cancer. Cancer Commun (Lond). 2021;41(2):109–20.
- 112. Cui K, Wu H, Fan J, Zhang L, Li H, Guo H, et al. The mixture of ferulic acid and P-coumaric acid suppresses colorectal cancer through lncRNA 495810/PKM2 mediated aerobic glycolysis. Int J Mol Sci. 2022;23(20):12106.
- 113. Vashisth P, Sharma M, Nikhil K, Singh H, Panwar R, Pruthi PA, et al. Antiproliferative activity of ferulic acid-encapsulated electrospun PLGA/PEO nanofbers against MCF-7 human breast carcinoma cells. 3 Biotech. 2015;5(3):303–15.
- 114. Faheem, Karan Kumar B, Venkata Gowri Chandra Sekhar K, Chander S, Kunjiappan S, Murugesan S. 1,2,3,4-Tetrahydroisoquinoline (THIQ) as privileged scafold for anticancer de novo drug design. Expert Opin Drug Discov. 2021;16(10):1119–47.
- 115. Wang J, Zhang H, Li JM, Zhang EL, Tao ZL. Design, synthesis and biological evaluation of tetrahydroisoquinoline ferulic acid derivatives as potential antitumor agents. J Suzhou Univ. 2018;33(03):109–13.
- 116. Johnson EM, Lee H, Jayabalan R, Suh JW. Ferulic acid grafted self-assembled fructo-oligosaccharide micro particle for targeted delivery to colon. Carbohydr Polym. 2020;247: 116550.
- 117. Cui HR, Zhang JY, Cheng XH, Zheng JX, Zhang Q, Zheng R, et al. Immunometabolism at the service of traditional Chinese medicine. Pharmacol Res. 2022;176: 106081.
- 118. Stompor-Gorący M, Machaczka M. Recent advances in biological activity, new formulations and prodrugs of ferulic acid. Int J Mol Sci. 2021;22(23):12889.

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