



Communication

Increase in *TPSB2* and *TPSD1* Expression in Synovium of Hip Osteoarthritis Patients Who Are Overweight

Maho Tsuchiya¹, Kensuke Fukushima¹, Ken Takata¹, Yoshihisa Ohashi¹, Katsufumi Uchiyama², Naonobu Takahira³ , Hiroki Saito¹, Ayumi Tsukada¹, Gen Inoue¹ , Masashi Takaso¹ and Kentaro Uchida^{1,*}

¹ Department of Orthopedic Surgery, Kitasato University School of Medicine, 1-15-1 Kitasato, Minami-ku, Sagami-hara City 252-0374, Japan; 09.ma.10.ho@gmail.com (M.T.); kenfu@r4.dion.ne.jp (K.F.); kentakata41@yahoo.co.jp (K.T.); 44134413oo@gmail.com (Y.O.); kgka.condition-green@xd6.so-net.ne.jp (H.S.); amidesutarere9010@yahoo.co.jp (A.T.); ginoue@kitasato-u.ac.jp (G.I.); mtakaso@kitasato-u.ac.jp (M.T.)

² Department of Patient Safety and Healthcare Administration, Kitasato University School of Medicine, 1-15-1 Kitasato, Minami-ku, Sagami-hara City 252-0374, Japan; katsufu@cf6.so-net.ne.jp

³ Department of Rehabilitation, Kitasato University School of Allied Health Sciences, 1-15-1 Kitasato, Minami-ku, Sagami-hara City 252-0373, Japan; takahira@med.kitasato-u.ac.jp

* Correspondence: kuchida@med.kitasato-u.ac.jp; Tel.: +81-42-778-9217

Abstract: While research suggests that increasing body mass index (BMI) is a risk factor for hip osteoarthritis (HOA), the mechanisms of this effect are not fully understood. Tryptases are among the main proteases found in mast cells (MCs) and contribute to OA pathology. *TPSB2*, which encodes β -tryptase, is increased in the synovium of overweight and obese knee OA patients. However, it remains unclear whether tryptase in the synovium of HOA is increased with increasing BMI. Here, we investigated tryptase genes (*TPSB2* and *TPSD1*) in the synovium of overweight HOA patients. Forty-six patients radiographically diagnosed with HOA were allocated to two groups based on BMI, namely normal ($<25 \text{ kg/m}^2$) and overweight ($25\text{--}29.99 \text{ kg/m}^2$). *TPSB2* and *TPSD1* expression in the synovium of the two groups was compared using real-time polymerase chain reaction. To compare *TPSB2* and *TPSD1* expression in MCs between the groups, we isolated the MC-rich fraction (MC-RF) and MC-poor fraction (MC-PF), extracted using magnetic isolation. *TPSB2* and *TPSD1* expression was increased in the overweight group compared with the normal group. Expression of both genes in the MC-RF was significantly higher than that in MC-PF in both groups. However, *TPSB2* and *TPSD1* expression levels in the MC-RF did not differ between the groups. Tryptase genes were highly expressed in the synovium of overweight HOA patients. Further investigation to reveal the role of tryptase in the relationship between increasing BMI and HOA pathology is required.

Keywords: hip osteoarthritis; mast cell; obesity; tryptase



Citation: Tsuchiya, M.; Fukushima, K.; Takata, K.; Ohashi, Y.; Uchiyama, K.; Takahira, N.; Saito, H.; Tsukada, A.; Inoue, G.; Takaso, M.; et al. Increase in *TPSB2* and *TPSD1* Expression in Synovium of Hip Osteoarthritis Patients Who Are Overweight. *Int. J. Mol. Sci.* **2023**, *24*, 11532. <https://doi.org/10.3390/ijms241411532>

Academic Editor: Giovanna Traina

Received: 20 June 2023

Revised: 14 July 2023

Accepted: 14 July 2023

Published: 16 July 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Osteoarthritis is a degenerative and whole-joint disease that causes pain, stiffness, and reduced mobility. Epidemiological studies suggest that being overweight or obese is a significant risk factor for developing osteoarthritis (OA), particularly in the knee and hip [1–5]. Many studies have investigated the mechanism of knee OA (KOA). Synovial tissue (ST) plays a pivotal role in the inflammatory processes and is involved in OA onset, progression, and pain [6,7]. In contrast, the mechanism of the effect of obesity on ST and the exacerbation of hip osteoarthritis is not completely understood.

Mast cells (MCs) are responsible for the production of inflammatory cytokines and their release in conditions such as asthma and allergies [8,9]. Accumulated evidence suggests that MCs contribute to synovial inflammation and OA progression [10–12]. We previously reported increased expression of MC markers in the ST of overweight and obese patients with KOA [13–15]. Nevertheless, it remains unclear whether MC are increased in the ST of hip osteoarthritis (HOA) with increased body mass index (BMI).

It is well known that tryptases are among the most abundant proteases found in MC [9,16,17]. An increase in tryptase enzyme activity in the synovial extract of KOA patients compared with healthy subjects has been documented [18]. In human MCs, at least four tryptase genes are expressed, *TPSAB1*, *TPSB2*, *TPSD1*, and *TPSG1*, which encode soluble (*TPSAB1*, *TPSB2*) and membrane-anchored (*TPSD1* and *TPSG1*) forms of tryptase [19]. *TPSB2* encodes β -tryptase, the main type released during degranulation in MCs. β -Tryptase is also involved in the production of inflammatory cytokines in inflammatory conditions and inflammatory diseases, as well as being the most abundant protein in human MCs [16,17]. In contrast to *TPSB2*, *TPSD1* encodes δ -tryptase and contains a premature stop codon that results in the loss of the C-terminal regions essential for normal catalytic process. *TPSD1* has been implicated in the pathogenesis of autoimmune diseases [19,20]. Even though *TPSD1* is largely inactive, there is a possibility that increased expression of this gene might be a sign of increased MC activity [21].

Here, we investigated the relationship between *TPSB2* and *TPSD1* expression and BMI in the ST of HOA patients.

2. Results

2.1. Synovial Levels of *TPSB2* and *TPSD1* by BMI

The study enrolled 46 patients radiographically diagnosed with HOA. The distribution of patients according to age and the proportion of patients according to Tönnis grade (2/3) were similar between the two groups ($p = 0.732$ and $p = 0.167$, respectively; Table 1). BMI in the overweight (OW) group was significantly higher than that in the normal-weight (NW) group ($p < 0.001$, Table 1). There was a significant increase in *TPSB2* expression in the OW group compared to the NW group (Figure 1A; $p = 0.038$). In addition, the expression of *TPSD1* in the OW group was higher than that in the NW group (Figure 1B; $p = 0.040$).

Table 1. Clinical characteristics of normal and overweight patients with hip osteoarthritis.

	qPCR			Magnetic Isolation		
	NW (n = 25)	OW (n = 21)	p	NW (n = 7)	OW (n = 7)	p
Age (years)	62.6 ± 9.8	63.0 ± 12.9	0.732	60.6 ± 13.1	58.4 ± 7.8	0.456
Tönnis (2/3), n	8/17	11/10	0.167	4/3	2/5	0.383
BMI (kg/m ²)	21.9 ± 2.2	27.1 ± 1.4	<0.001	22.6 ± 1.34	26.5 ± 1.3	<0.001

NW, normal weight, OW, overweight, BMI, body mass index.

2.2. Correlation between *TPSB2* and *TPSD1* Expression and Proinflammatory Cytokines

Proinflammatory cytokines such as TNF- α , IL-1 β , and IL6 play a pivotal role in synovial inflammation [22,23]. To estimate the possible association of *TPSB2* and *TPSD1* in synovial inflammation, correlation between their expression and inflammatory cytokine expression was investigated. *TPSB2* expression levels positively correlated with *TNFA* ($r = 0.692$, $p < 0.001$) and *IL6* ($r = 0.302$, $p = 0.049$) expression levels (Figure 2A,B). There was no difference between *TPSB2* and *IL1B* expression levels ($r = 0.228$, $p = 0.142$; Figure 2C). Similarly, *TPSD1* expression levels positively correlated with *TNFA* ($r = 0.721$, $p < 0.001$) and *IL6* ($r = 0.333$, $p = 0.029$) expression levels (Figure 2D,E). No correlation was observed between *TPSD1* and *IL1B* ($r = 0.109$, $p = 0.486$, Figure 2F).

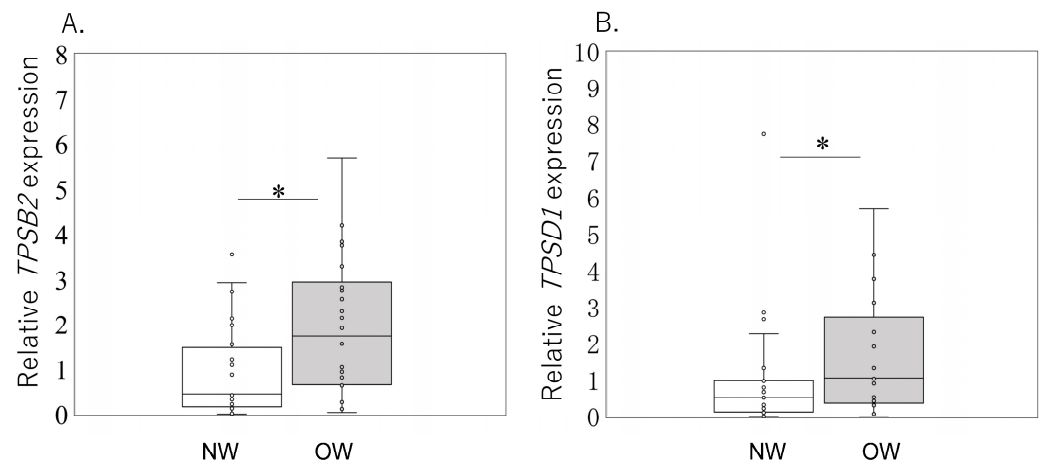


Figure 1. Expression of *TPSB2* and *TPSD1* in the synovial tissue of normal-weight (NW) and overweight (OW) patients. The expression of *TPSB2* and *TPSD1* mRNA in NW and OW groups was estimated by qPCR (A,B). *TPSB2* (A) and *TPSD1* (B) mRNA expression in the synovial tissue of normal-weight ($n = 25$) and overweight ($n = 21$) patients with hip osteoarthritis. Gene expression is presented in box and whisker plots, showing the median, 25th, and 75th percentiles and range. * $p < 0.05$.

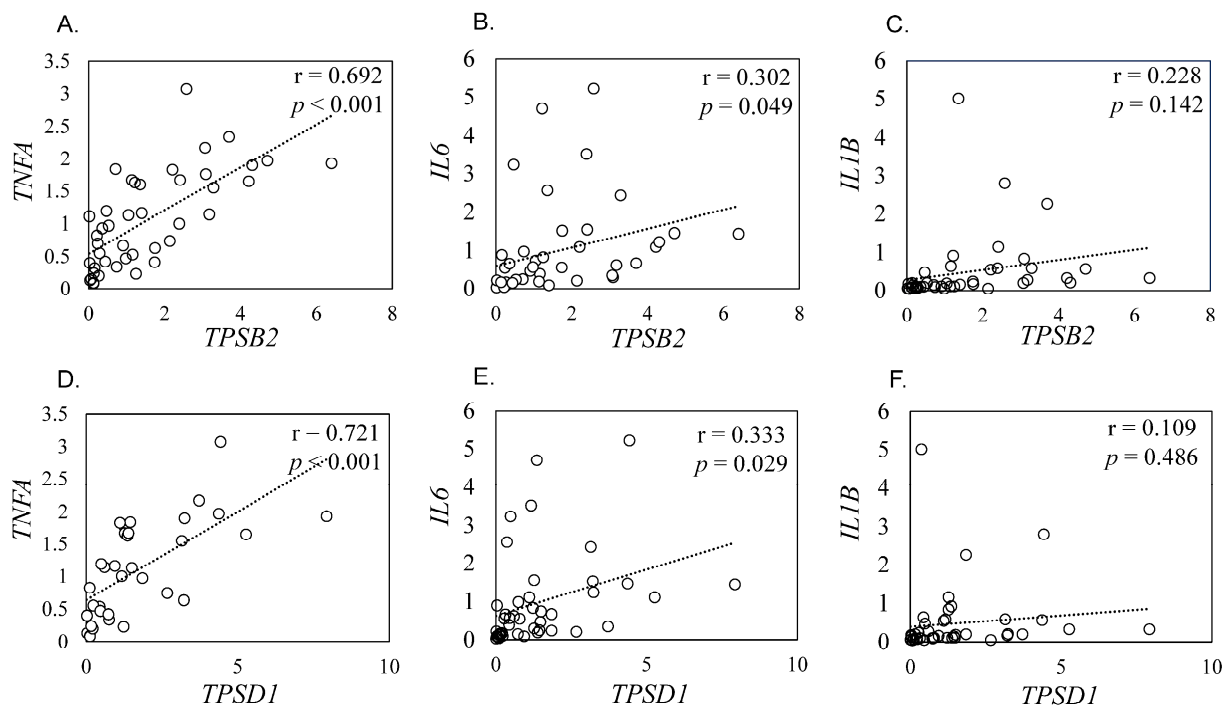


Figure 2. Correlation between *TPSB2* and *TPSD1* expression and proinflammatory cytokines expression. (A–C) Correlation between *TPSB2* and *TNFA* (A), *IL6* (B), and *IL1B* (C). (D–F) Correlation between *TPSD1* and *TNFA* (D), *IL6* (E), and *IL1B* (F).

2.3. Tryptase Expression Levels in MCs Derived from the NW and OW Groups

Clinical characteristics of patients determined by magnetic isolation are shown in Table 1. Comparison of the MC-rich fractions (MC-RFs) and MC-poor fractions (MC-PFs) using magnetic beads showed higher expression levels of *TPSB2* and *TPSD1* in the MC-RF than the MC-PF in both NW (*TPSB2*, $p = 0.014$; *TPSD1*, $p = 0.021$) and OW (*TPSB2*, $p = 0.017$; *TPSD1*, $p = 0.138$) (Figure 3A,B, respectively). There were no significant differences in the expression levels of *TPSB2* or *TPSD1* in the MC-RF and MC-PF between the NW and OW groups (*TPSB2*, $p = 1.000$; *TPSD1*, $p = 1.000$)

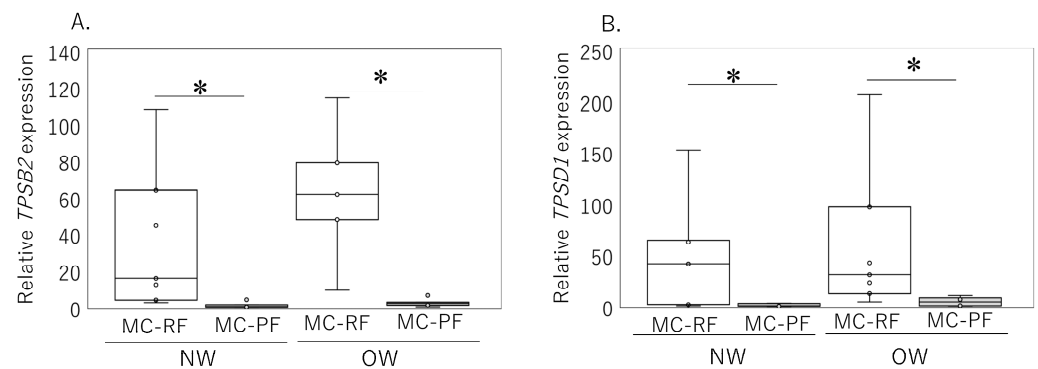


Figure 3. Expression of *TPSB2* and *TPSD1* in mast-cell-rich fraction (MC-RF) and mast-cell-poor fraction (MC-PF) obtained from normal-weight (NW) and overweight (OW) patients. The expression of *TPSB2* (A) and *TPSD1* (B) mRNA in MC-RF and MC-PF (NW, $n = 7$; OW, $n = 7$) was estimated by qPCR. Gene expressions are presented in box and whisker plots, showing the median, 25th, and 75th percentiles and range. * $p < 0.05$.

3. Discussion

In this study, we showed that *TPSB2* and *TPSD1* were significantly increased in HOA patients with OW. This increase in overweight patients may partly explain the link between BMI and HOA.

β -tryptase plays a pivotal role in the inflammatory process through the stimulation of inflammatory cytokine production. β -tryptase stimulates IL-1 β production in synovial macrophages and fibroblasts derived from KOA patients [13]. β -tryptase increased TNF- α and *IL6* production in microglia and peripheral mononuclear cells [24,25]. A recent study also reported that β -tryptase modulated joint lubrication in OA through the cleavage of lubricin [10]. In contrast, recent studies have suggested that increased *TPSD1* gene expression levels were observed in several diseases [26,27]. For example, *TPSD1* expression was increased in patients with aspirin-exacerbated respiratory disease [26]. Increased *TPSD1* expression was also observed in patients with regenerative rotator cuff tear [27]. In our study, *TPSB2* and *TPSD1* positively correlated with *TNFA* and *IL6*. Increased *TPSB2* and *TPSD1* expression may be associated with inflammation in HOA in patients who are overweight.

The obese condition changes cell phenotypes and response to inflammatory stimulation [11,28,29]. Synovial fibroblasts from obese KOA patients had a high capacity to produce *IL6* protein [11]. Synovial fibroblasts in obese patients highly expressed transcriptional regulators MYC and FOS [29]. Synovial fibroblasts from obese KOA patients exhibited greater aerobic glycolysis, basal lactate secretion, and mitochondrial respiration when stimulated with TNF- α , compared to synovial fibroblasts from subjects with normal weight [28]. Bulk analysis of synovial tissue revealed that synovial *TPSB2* and *TPSD1* expression levels were increased in overweight HOA patients compared to normal-weight patients. We hypothesized that this result reflects the fact that MC-derived OW patients highly expressed *TPSB2* and *TPSD1*, and although *TPSB2* and *TPSD1* expression levels in MCs were similar between normal and overweight patients, an increased ratio/number of MCs results in an increase in *TPSB2* and *TPSD1* in ST of overweight patients. We isolated MCs from normal and overweight patients but found no difference between the groups in *TPSB2* and *TPSD1* expression. These results may suggest that elevated *TPSB2* and *TPSD1* expression in ST of overweight HOA patients may reflect an increased MC number or ratio in ST of overweight patients. In contrast, basophils also express *TPSB2* and *TPSD1* [30]. Therefore, elevated *TPSB2* and *TPSD1* expression in overweight HOA patients may reflect an increase in non-MC populations expressing these genes. Further investigation regarding MC number or MC ratio using flow cytometric analysis or immunohistochemistry is needed to clarify this hypothesis.

Several limitations of the present study warrant mention. First, sample size was small. Second, we did not examine a control population. Inclusion of normal and overweight HOA populations in the study is essential to confirm whether *TPSB2* and *TPSD1* expression is increased in HOA with overweight. Third, we analyzed mRNA expression in the synovium. Further investigation to complement these findings is necessary, such as a protein-profiling study using Western blot as a complementary measure to our gene expression results. Finally, the relationship between *TPSB2* and *TPSD1* expression and synovial inflammation or OA pathology remains to be determined.

4. Material and Methods

4.1. Patients

The study was conducted with a retrospective design in patients radiographically diagnosed with HOA at our institution between 2020 and 2022. Inclusion criteria were age ≥ 40 years and symptomatic primary HOA (Tönnis classification system II–III).

We excluded patients with previous hip surgery, current or previous use of immunosuppressive medication, and HOA caused by rapidly destructive coxarthrosis, idiopathic osteonecrosis of the femoral head, pigmented villonodular synovitis, trauma, or rheumatoid arthritis. Samples were extracted from ST lining the anterior joint capsule in the inferior part of the femoral neck adjacent to the femoral head during the total hip arthroplasty procedure via the anterolateral supine approach in each subject. The subjects were grouped according to the World Health Organization's BMI classifications, namely normal weight (NW; $<25 \text{ kg/m}^2$) and overweight (OW; $25\text{--}29.99 \text{ kg/m}^2$). Expression of synovial tryptase (*TPSB2*, *TPSD1*) mRNA was investigated by real time PCR. Tissues from the remaining two sets of 14 patients (normal, $n = 7$ and overweight, $n = 7$) were used to compare tryptase gene expression between mast cells derived from normal and overweight patients.

4.2. Clinical Assessment

From the radiographic assessment of the progression of HOA, the following grades were assigned based on the Tönnis classification system [31]. Patient background factors and clinical evaluations were compared between patients with normal weight and overweight following quantitative PCR (qPCR) analysis and MC isolation, as described in Table 2.

Table 2. Sequences of the primers used in this study.

Gene	Direction	Primer Sequence (5'–3')	Product Size (bp)
<i>TPSB2</i>	F	CGCAAATACCACCTTGGCG	138
	R	GTGCCATTACCTTGCACAC	
<i>TPSD1</i>	F	CGGAATATCACACCGGCCTC	135
	R	TGCCATTACCTTGCAGACC	
<i>TNFA</i>	F	CTTCTGCCTGCTGCACTTTG	118
	R	GTCACCTCGGGGTTTCGAGAAG	
<i>IL1B</i>	F	GTACCTGTCCCTGCGTGTTGA	153
	R	GGGAACTGGGCAGACTCAA	
<i>IL6</i>	F	GAGGAGACTTGCCTGGTGAA	199
	R	TGGCATTGTGGTTGGGTCA	
<i>GAPDH</i>	F	TGTTGCCATCAATGACCCCTT	202
	R	CTCCACGACGTACTCAGCG	

4.3. qPCR

RNA extraction, cDNA synthesis, and qPCR were conducted using methods previously described in literature [14]. Briefly, primers used for qPCR are listed in Table 2. Gene expression was normalized to that of GAPDH using the delta–delta Ct method. Relative expression was calculated using the mean of all samples (synovial samples from the NW group or MC-poor fraction from the NW group).

4.4. Magnetic Isolation of MC

MC isolation was performed using magnetic isolation methods reported previously [13]. Briefly, cells were obtained from collagenase-digested synovial samples derived from NW and OW patients. After centrifugation, cells were reacted with biotin-labelled antibody cocktail (anti CD3, CD14, CD19, CD90). All antibodies were purchased from Biolegend (San Diego, CA, USA). Following reaction with streptavidin-conjugated magnetic beads (BD™ IMag Streptavidin Particles Plus—DM, BD Biosciences, Tokyo, Japan), the MC-RF were isolated by negative selection. Positive fractions were also isolated as an MC-PF. *TPSB2* and *TPSD1* in the MC-RF and MC-PF were estimated by qPCR instrument (Bio-Rad CFX Connect, Bio-Rad, Hercules, CA, USA).

4.5. Statistical Analysis

We used non-parametric analyses, as continuous variables in all analyses showed a non-normal distribution by the Shapiro–Wilk test. Continuous and categorical variables between the two groups were compared using the Mann–Whitney U-test and Pearson's chi-squared test, respectively. Gene expression between the groups was compared using the Kruskal–Wallis test adjusted with Bonferroni correction. All statistical analyses were conducted using commercial software (IBM SPSS Statistics for Windows version 28, IBM Corp., Armonk, NY, USA), with p values of less than 0.05 considered to indicate statistical significance.

5. Conclusions

TPSB2 and *TPSD1* expression is increased in synovium of overweight HOA patients. Further investigation is needed to reveal the role of tryptase in the relationship between increasing BMI and HOA pathology.

Author Contributions: Conceptualization, K.F. and K.U. (Kentaro Uchida); Methodology, K.U. (Kentaro Uchida); Validation, K.U. (Kentaro Uchida); Formal analysis, M.T. (Maho Tsuchiya), K.T., Y.O., H.S. and A.T.; Investigation, M.T. (Maho Tsuchiya), K.T., Y.O., H.S. and A.T.; Resources, K.F. and K.U. (Katsufumi Uchiyama); Writing—original draft, M.T. (Maho Tsuchiya) and K.U. (Kentaro Uchida); Writing—review and editing, K.U. (Kentaro Uchida); Visualization, G.I.; Supervision, N.T., G.I. and M.T. (Masashi Takaso); Project administration, M.T. (Masashi Takaso) and K.U. (Kentaro Uchida); Funding acquisition, K.T. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Grant-in-Aid for Young Scientists (Start-up) grant number 22K20965.

Institutional Review Board Statement: This study was approved by the IRB of Kitasato University (reference number: B19-259).

Informed Consent Statement: Written informed consent was obtained from all the patients for harvesting of the synovium for use in this study.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Cheng, K.Y.; Strotmeyer, E.S.; Kado, D.M.; Schousboe, J.T.; Schenk, S.; Nevitt, M.; Lane, N.E.; Hughes-Austin, J.M. The Association of Metabolic Syndrome and Obesity With Clinical Hip Osteoarthritis in the Study of Osteoporotic Fractures and the Osteoporotic Fractures in Men Study Cohorts. *ACR Open Rheumatol.* **2023**, *5*, 115–123. [[CrossRef](#)]
2. Gelber, A.C. Obesity and hip osteoarthritis: The weight of the evidence is increasing. *Am. J. Med.* **2003**, *114*, 158–159. [[CrossRef](#)] [[PubMed](#)]
3. Ji, S.; Liu, L.; Li, J.; Zhao, G.; Cai, Y.; Dong, Y.; Wang, J.; Wu, S. Prevalence and factors associated with knee osteoarthritis among middle-aged and elderly individuals in rural Tianjin: A population-based cross-sectional study. *J. Orthop. Surg. Res.* **2023**, *18*, 266. [[CrossRef](#)] [[PubMed](#)]

4. Murphy, N.J.; Eyles, J.P.; Hunter, D.J. Hip Osteoarthritis: Etiopathogenesis and Implications for Management. *Adv. Ther.* **2016**, *33*, 1921–1946. [[CrossRef](#)]
5. Park, J.M. Association between obesity and osteoarthritis in the South Korean older population: A nationwide population-based study. *Medicine* **2023**, *102*, e33455. [[CrossRef](#)]
6. Jones, G.C.; Riley, G.P.; Buttle, D.J. The role of proteases in pathologies of the synovial joint. *Int. J. Biochem. Cell Biol.* **2008**, *40*, 1199–1218. [[CrossRef](#)] [[PubMed](#)]
7. Sanchez-Lopez, E.; Coras, R.; Torres, A.; Lane, N.E.; Guma, M. Synovial inflammation in osteoarthritis progression. *Nat. Rev. Rheumatol.* **2022**, *18*, 258–275. [[CrossRef](#)]
8. Maun, H.R.; Jackman, J.K.; Choy, D.F.; Loyet, K.M.; Staton, T.L.; Jia, G.; Dressen, A.; Hackney, J.A.; Bremer, M.; Walters, B.T.; et al. An Allosteric Anti-tryptase Antibody for the Treatment of Mast Cell-Mediated Severe Asthma. *Cell* **2019**, *179*, 417–431.e19. [[CrossRef](#)]
9. Sommerhoff, C.P.; Schaschke, N. Mast cell tryptase beta as a target in allergic inflammation: An evolving story. *Curr. Pharm. Des.* **2007**, *13*, 313–332. [[CrossRef](#)]
10. Das, N.; de Almeida, L.G.N.; Derakhshani, A.; Young, D.; Mehdinejadani, K.; Salo, P.; Rezanooff, A.; Jay, G.D.; Sommerhoff, C.P.; Schmidt, T.A.; et al. Tryptase beta regulation of joint lubrication and inflammation via proteoglycan-4 in osteoarthritis. *Nat. Commun.* **2023**, *14*, 1910. [[CrossRef](#)]
11. Nanus, D.E.; Wijesinghe, S.N.; Pearson, M.J.; Hadjicharalambous, M.R.; Rosser, A.; Davis, E.T.; Lindsay, M.A.; Jones, S.W. Regulation of the Inflammatory Synovial Fibroblast Phenotype by Metastasis-Associated Lung Adenocarcinoma Transcript 1 Long Noncoding RNA in Obese Patients With Osteoarthritis. *Arthritis Rheumatol.* **2020**, *72*, 609–619. [[CrossRef](#)]
12. Wang, Q.; Lepus, C.M.; Raghu, H.; Reber, L.L.; Tsai, M.M.; Wong, H.H.; von Kaeppler, E.; Lingampalli, N.; Bloom, M.S.; Hu, N.; et al. IgE-mediated mast cell activation promotes inflammation and cartilage destruction in osteoarthritis. *eLife* **2019**, *8*, e39905. [[CrossRef](#)]
13. Takata, K.; Uchida, K.; Mukai, M.; Takano, S.; Aikawa, J.; Iwase, D.; Sekiguchi, H.; Miyagi, M.; Inoue, G.; Takaso, M. Increase in Tryptase and Its Role in the Synovial Membrane of Overweight and Obese Patients with Osteoarthritis of the Knee. *Diabetes Metab. Syndr. Obes.* **2020**, *13*, 1491–1497. [[CrossRef](#)]
14. Tsukada, A.; Takata, K.; Takano, S.; Ohashi, Y.; Mukai, M.; Aikawa, J.; Iwase, D.; Inoue, G.; Takaso, M.; Uchida, K. Increased NMUR1 Expression in Mast Cells in the Synovial Membrane of Obese Osteoarthritis Patients. *Int. J. Mol. Sci.* **2022**, *23*, 11237. [[CrossRef](#)]
15. Uchida, K.; Takano, S.; Inoue, G.; Iwase, D.; Aikawa, J.; Takata, K.; Tazawa, R.; Kawakubo, A.; Sekiguchi, H.; Takaso, M. Increase in mast cell marker expression in the synovium of obese patients with osteoarthritis of the knee. *Diabetes Metab. Syndr. Obes.* **2019**, *12*, 377–382. [[CrossRef](#)] [[PubMed](#)]
16. Hallgren, J.; Pejler, G. Biology of mast cell tryptase. An inflammatory mediator. *FEBS J.* **2006**, *273*, 1871–1895. [[CrossRef](#)] [[PubMed](#)]
17. Sommerhoff, C.P.; Bode, W.; Pereira, P.J.; Stubbs, M.T.; Sturzebecher, J.; Piechottka, G.P.; Matschiner, G.; Bergner, A. The structure of the human betaII-tryptase tetramer: Fo(u)r better or worse. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 10984–10991. [[CrossRef](#)]
18. Nakano, S.; Mishiro, T.; Takahara, S.; Yokoi, H.; Hamada, D.; Yukata, K.; Takata, Y.; Goto, T.; Egawa, H.; Yasuoka, S.; et al. Distinct expression of mast cell tryptase and protease activated receptor-2 in synovia of rheumatoid arthritis and osteoarthritis. *Clin. Rheumatol.* **2007**, *26*, 1284–1292. [[CrossRef](#)]
19. Caughey, G.H. Mast cell tryptases and chymases in inflammation and host defense. *Immunol. Rev.* **2007**, *217*, 141–154. [[CrossRef](#)] [[PubMed](#)]
20. Pallaoro, M.; Fejzo, M.S.; Shayesteh, L.; Blount, J.L.; Caughey, G.H. Characterization of genes encoding known and novel human mast cell tryptases on chromosome 16p13.3. *J. Biol. Chem.* **1999**, *274*, 3355–3362. [[CrossRef](#)]
21. Kennedy, R.B.; Oberg, A.L.; Ovsyannikova, I.G.; Haralambieva, I.H.; Grill, D.; Poland, G.A. Transcriptomic profiles of high and low antibody responders to smallpox vaccine. *Genes Immun.* **2013**, *14*, 277–285. [[CrossRef](#)]
22. Mathiessen, A.; Conaghan, P.G. Synovitis in osteoarthritis: Current understanding with therapeutic implications. *Arthritis Res. Ther.* **2017**, *19*, 18. [[CrossRef](#)]
23. Rahmati, M.; Mobasheri, A.; Mozafari, M. Inflammatory mediators in osteoarthritis: A critical review of the state-of-the-art, current prospects, and future challenges. *Bone* **2016**, *85*, 81–90. [[CrossRef](#)]
24. Malamud, V.; Vaaknin, A.; Abramsky, O.; Mor, M.; Burgess, L.E.; Ben-Yehudah, A.; Lorberboum-Galski, H. Tryptase activates peripheral blood mononuclear cells causing the synthesis and release of TNF α , IL6 and IL1 β : Possible relevance to multiple sclerosis. *J. Neuroimmunol.* **2003**, *138*, 115–122. [[CrossRef](#)] [[PubMed](#)]
25. Zhang, S.; Zeng, X.; Yang, H.; Hu, G.; He, S. Mast cell tryptase induces microglia activation via protease-activated receptor 2 signaling. *Cell. Physiol. Biochem.* **2012**, *29*, 931–940. [[CrossRef](#)] [[PubMed](#)]
26. Bangert, C.; Villazala-Merino, S.; Fahrenberger, M.; Krausgruber, T.; Bauer, W.M.; Stanek, V.; Campion, N.J.; Bartosik, T.; Quint, T.; Regelsberger, G.; et al. Comprehensive Analysis of Nasal Polyps Reveals a More Pronounced Type 2 Transcriptomic Profile of Epithelial Cells and Mast Cells in Aspirin-Exacerbated Respiratory Disease. *Front. Immunol.* **2022**, *13*, 850494. [[CrossRef](#)]
27. Rai, M.F.; Cai, L.; Tycksen, E.D.; Chamberlain, A.; Keener, J. RNA-Seq analysis reveals sex-dependent transcriptomic profiles of human subacromial bursa stratified by tear etiology. *J. Orthop. Res.* **2022**, *40*, 2713–2727. [[CrossRef](#)]

28. Farah, H.; Wijesinghe, S.N.; Nicholson, T.; Alnajjar, F.; Certo, M.; Alghamdi, A.; Davis, E.T.; Young, S.P.; Mauro, C.; Jones, S.W. Differential Metabotypes in Synovial Fibroblasts and Synovial Fluid in Hip Osteoarthritis Patients Support Inflammatory Responses. *Int. J. Mol. Sci.* **2022**, *23*, 3266. [[CrossRef](#)] [[PubMed](#)]
29. Wijesinghe, S.N.; Badoume, A.; Nanus, D.E.; Sharma-Oates, A.; Farah, H.; Certo, M.; Alnajjar, F.; Davis, E.T.; Mauro, C.; Lindsay, M.A.; et al. Obesity defined molecular endotypes in the synovium of patients with osteoarthritis provides a rationale for therapeutic targeting of fibroblast subsets. *Clin. Transl. Med.* **2023**, *13*, e1232. [[CrossRef](#)]
30. Jogie-Brahim, S.; Min, H.K.; Fukuoka, Y.; Xia, H.Z.; Schwartz, L.B. Expression of alpha-tryptase and beta-tryptase by human basophils. *J. Allergy Clin. Immunol.* **2004**, *113*, 1086–1092. [[CrossRef](#)]
31. Kovalenko, B.; Bremjit, P.; Fernando, N. Classifications in Brief: Tonnis Classification of Hip Osteoarthritis. *Clin. Orthop. Relat. Res.* **2018**, *476*, 1680–1684. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.