



Published in final edited form as:

Gastroenterology. 2023 August ; 165(2): 502–504.e2. doi:10.1053/j.gastro.2023.04.017.

Intratumoral *Malassezia globosa* levels predict survival and therapeutic response to adjuvant chemotherapy in patients with pancreatic ductal adenocarcinoma

Keisuke Okuno, MD, Ph.D.^{1,2}, Masanori Tokunaga, MD, Ph.D.², Daniel Von Hoff, MD³, Yusuke Kinugasa, MD, Ph.D.², Ajay Goel, Ph.D.^{1,4}

¹Department of Molecular Diagnostics and Experimental Therapeutics, Beckman Research Institute of City of Hope, Biomedical Research Center, Monrovia, California, USA

²Department of Gastrointestinal Surgery, Tokyo Medical and Dental University, Tokyo, Japan

³Molecular Medicine Division, Translational Genomics Research Institute, Phoenix, Arizona, USA

⁴City of Hope Comprehensive Cancer Center, Duarte, California, USA

Fungi are microeukaryotes, estimated to encompass between 2.2 to 3.9 million species that inhabit various anatomic sites within the human body with relatively stable colonization – collectively forming the mycobiome^{1, 2}. Despite their low abundance in the human microbiome^{1, 2}, emerging evidence highlights the significant influence of mycobiome on the host's health, including nutrition, metabolism, immunity, and the physiological functions of various body organs^{3, 4}. More specifically, the associations between mycobiome and human cancer have garnered increasing attention in recent years due to the significant enrichment of specific fungi in patients with malignant tumors. However, the biological underpinnings of human mycobiome in cancer pathogenesis and disease progression continue to evolve and remain unclear.

A recent pre-clinical and clinical study revealed that pancreatic ductal adenocarcinomas (PDACs) harbor significant enrichment of one such specific fungi in mice models and human specimens relative to normal pancreatic tissues⁵. Furthermore, this study revealed that the commensal gut fungi – *Malassezia globosa* (*M. globosa*) – migrates from the gut to

Corresponding author: Prof. Ajay Goel, Department of Molecular Diagnostics and Experimental Therapeutics, Beckman Research Institute of City of Hope, 1218 S. Fifth Avenue, Suite 2226, Biomedical Research Center, Monrovia, California 91016; ajgoel@coh.org.

Author Contributions:

Keisuke Okuno, MD, Ph.D. (Conceptualization: Equal; Data curation: Lead; Formal analysis: Lead; Investigation: Lead; Methodology: Lead; Writing – original draft: Lead)

Masanori Tokunaga, MD, Ph.D. (Formal analysis: Equal; Methodology: Equal; Supervision: Equal; Writing – original draft: Equal)

Yusuke Kinugasa, MD, Ph.D. (Formal analysis: Equal; Methodology: Equal; Supervision: Equal; Writing – original draft: Equal)

Daniel Von Hoff, MD (Funding acquisition: Equal; Project administration: Equal; Supervision: Equal; Writing – original draft: Equal)

Ajay Goel, Ph.D. (Conceptualization: Equal; Funding acquisition: Lead; Project administration: Lead; Supervision: Lead; Writing – review & editing: Equal)

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Conflicts of Interest: The authors disclosed no potential conflicts of interest.

the pancreas and promotes pancreatic carcinogenesis via the activation of the complement cascade^{5, 6}. More interestingly, this study elegantly demonstrated that antifungal drugs inhibited the oncogenic progression of PDAC, and the combination of an antifungal drug and chemotherapy exhibited a synergistic anti-cancer effect against PDAC in animal models⁵. Likewise, another commensal fungus, *Candida tropicalis*, was implicated in the pathogenesis of colorectal cancer (CRC) through the accumulation of myeloid-derived suppressor cells⁷. In light of such groundbreaking studies, alterations in the mycobiome have gained increasing attention as functional determinants of disease and potential diagnostic and prognostic biomarkers in various cancers. For instance, as fecal biomarkers, a higher abundance of multiple fungi, including *M. globosa*, has been shown to distinguish CRC patients from healthy individuals accurately⁸. Furthermore, more recent analytical studies demonstrated that tumor-specific mycobiome could predict prognosis in multiple gastrointestinal malignancies⁹. Given this evidence, we hypothesized that high *M. globosa* levels in tissue specimens from patients with PDAC might serve as critical prognostic biomarkers.

Accordingly, this study explored the feasibility of tissue-based fungal biomarkers for predicting prognosis in patients with PDAC. Using quantitative nested polymerase chain reaction (PCR) assays in PDAC patient cohorts, we first determined the frequency of *M. globosa*-positivity in PDACs, followed by the quantitative associations between the levels of this fungus and its potential in predicting cancer-related death and tumor recurrence in patients with this malignancy.

To examine the *M. globosa* levels, surgically resected tissue specimens from a cohort of 180 PDAC patients with stages I-III disease were analyzed using quantitative nested PCR assays. Following extraction of DNA, the specific target amplification for the internal transcribed spacer (ITS) regions of *M. globosa* ribosomal DNA was confirmed using the BigDye Terminator Cycle Sequencing (Fig. 1A). Subsequently, in the quantitative nested PCR assays normalized by the *HBB* gene expression, 78 of 180 (43.3%) cases exhibited a Ct value of less than 33, indicating high levels of this fungus, and were defined as *M. globosa*-positive. In contrast, no signal was observed for the remaining 102 cases, even after 50 PCR cycles (Fig. 1B), and they were defined as *M. globosa*-negative. Interestingly, this frequency of *M. globosa*-positivity in PDAC tissues was similar to that in stool samples of healthy individuals (36–53%), which were reported in a previous sequencing-based human gut mycobiome project¹⁰.

First, the correlation between *M. globosa* levels and clinicopathological factors was assessed to determine their prognostic potential. In these analyses, *M. globosa* levels did not significantly correlate with other key clinicopathological factors, including patient demographics, tumor characteristics, and serum tumor markers (Supplementary Table. S1). However, male patients tended to have a higher burden of *M. globosa*-positive cases vs. female patients ($P=0.09$). This higher abundance of *Malassezia* species in male patients was also found in previous studies that quantified *Malassezia* colonization of the skin^{11, 12}. It was unclear whether this higher abundance of *M. globosa* in male patients may affect differential gender burden and prognostic outcomes in PDAC. Therefore, further studies

are warranted on this topic, including investigating larger global populations to confirm the findings further.

Next, to determine whether the intratumoral *M. globosa* levels might serve as predictors of prognosis in patients with PDAC, Kaplan-Meier analyses were performed for overall survival (OS) and recurrence-free survival (RFS). Interestingly, *M. globosa*-positive PDACs exhibited a significantly poor OS (hazard ratio [HR]: 1.69; 95% confidence interval [CI]: 1.21–2.38; log-rank $P < 0.01$; Fig. 1C), as well as markedly poor RFS (HR: 1.51; 95% CI: 1.07–2.11; $P = 0.02$; Fig. 1D). Of note, multivariate Cox regression analyses revealed that the *M. globosa* levels were significant and independent predictors for both OS (HR: 1.66; 95% CI: 1.17–2.35; $P < 0.01$; Fig. 1E) and RFS (HR: 1.51; 95% CI: 1.07–2.13; $P = 0.02$; Fig. 1F), highlighting that intratumoral *M. globosa* levels have the potential to serve as prognostic biomarkers in patients with PDAC.

Finally, we examined the impact of intratumoral *M. globosa* levels on treatment response in PDAC patients treated with adjuvant chemotherapy following curative resection of their tumors, using Kaplan-Meier analyses. In line with our findings for all cases, *M. globosa*-positive PDACs exhibited a significantly poor OS (HR: 2.00; 95% CI: 1.31–3.06; $P < 0.01$; Fig. 1G) and RFS (HR: 1.55; 95% CI: 1.02–2.37; $P = 0.04$; Fig. 1H) in the subset of patients treated with adjuvant chemotherapy. These results illustrate that patients with *M. globosa*-positive PDAC might have greater resistance to adjuvant chemotherapy, rendering an overall poor prognosis in this malignancy.

In summary, we, for the first time, demonstrate that intratumoral *M. globosa* levels are associated with cancer-related survival and tumor recurrence in patients with PDAC. In addition, our study provides novel evidence that *M. globosa* levels are a significant predictor of therapeutic response to adjuvant chemotherapy in PDAC patients. This study highlights that *M. globosa* levels are a significant predictor of prognosis in PDAC patients and might be a potential target of antifungal intervention in patients with PDAC.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments:

The PDAC Biomarker Working Group: Naoki Takahashi (Department of Gastroenterology, Saitama Cancer Center, Saitama, Japan), Yasuhide Yamada (Comprehensive Cancer Center, National Center for Global Health and Medicine, Tokyo, Japan), Mitsuro Kanda (Department of Gastroenterological Surgery, Nagoya University Graduate School of Medicine, Nagoya, Japan), Yasuhiro Kodera (Department of Gastroenterological Surgery, Nagoya University Graduate School of Medicine, Nagoya, Japan), Hideo Baba (Department of Gastroenterological Surgery, Graduate School of Medical Sciences, Kumamoto University, Kumamoto, Japan).

Funding:

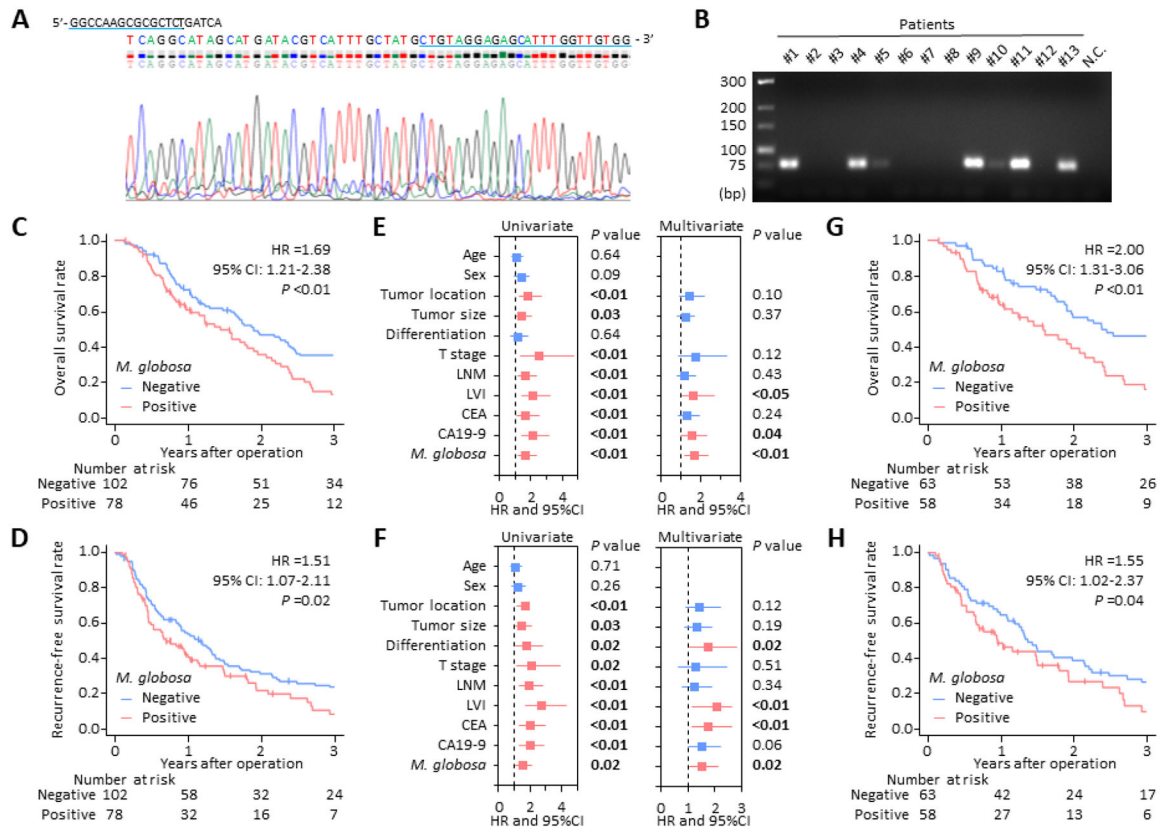
This work was supported by the CA072851, CA184792, CA187956, CA202797, CA227602, and CA214254 grants from the National Cancer Institute and National Institutes of Health.

Data availability:

Data is contained within the article.

REFERENCES

1. Cui L, Morris A, Ghedin E. The human mycobiome in health and disease. *Genome Med* 2013;5:63. [PubMed: 23899327]
2. Huffnagle GB, Noverr MC. The emerging world of the fungal microbiome. *Trends Microbiol* 2013;21:334–41. [PubMed: 23685069]
3. Wu X, Xia Y, He F, et al. Intestinal mycobiota in health and diseases: from a disrupted equilibrium to clinical opportunities. *Microbiome* 2021;9:60. [PubMed: 33715629]
4. Kong HH, Segre JA. Cultivating fungal research. *Science* 2020;368:365–366. [PubMed: 32327584]
5. Aykut B, Pushalkar S, Chen R, et al. The fungal mycobiome promotes pancreatic oncogenesis via activation of MBL. *Nature* 2019;574:264–267. [PubMed: 31578522]
6. Pushalkar S, Hundeyin M, Daley D, et al. The Pancreatic Cancer Microbiome Promotes Oncogenesis by Induction of Innate and Adaptive Immune Suppression. *Cancer Discov* 2018;8:403–416. [PubMed: 29567829]
7. Wang T, Fan C, Yao A, et al. The Adaptor Protein CARD9 Protects against Colon Cancer by Restricting Mycobiota-Mediated Expansion of Myeloid-Derived Suppressor Cells. *Immunity* 2018;49:504–514.e4. [PubMed: 30231984]
8. Coker OO, Nakatsu G, Dai RZ, et al. Enteric fungal microbiota dysbiosis and ecological alterations in colorectal cancer. *Gut* 2019;68:654–662. [PubMed: 30472682]
9. Narunsky-Haziza L, Sepich-Poore GD, Livyatan I, et al. Pan-cancer analyses reveal cancer-type-specific fungal ecologies and bacteriome interactions. *Cell* 2022;185:3789–3806.e17. [PubMed: 36179670]
10. Nash AK, Auchtung TA, Wong MC, et al. The gut mycobiome of the Human Microbiome Project healthy cohort. *Microbiome* 2017;5:153. [PubMed: 29178920]
11. Sugita T, Suzuki M, Goto S, et al. Quantitative analysis of the cutaneous *Malassezia* microbiota in 770 healthy Japanese by age and gender using a real-time PCR assay. *Med Mycol*. 2010;48:229–233. [PubMed: 19462267]
12. Hobi S, Cafarchia C, Romano V, et al. *Malassezia*: Zoonotic Implications, Parallels and Differences in Colonization and Disease in Humans and Animals. *J Fungi (Basel)*. 2022;8:708. [PubMed: 35887463]

**Figure 1.**

Intratumoral *Malassezia globosa* levels and prognostic significance in PDAC. (A) Representative sequences of PCR amplified products in nested PCR assays for detecting *M. globosa* in PDAC specimens. Forward and reverse primer location is indicated by a blue underline. (B) Representative agarose gel electrophoresis (4% agarose) of PCR amplified products in nested PCR assays. Lane 1 is a low-range DNA ladder, lanes 2–14 are PCR amplified products from PDAC tissue specimens, and lane 15 is negative control. (C, D) Kaplan-Meier curves depicting the overall survival (C) and recurrence-free survival (D) for patients with *M. globosa* negative (n = 102) and positive (n = 78) tumors. (E, F) Forest plots with HR for each key clinicopathological factor and *M. globosa* level in univariate and multivariate Cox regression analysis for overall survival (E) and recurrence-free survival (F) in the clinical cohort (n = 180) (G, H) Kaplan-Meier curves depicting the overall survival (G) and recurrence-free survival (H) for PDAC patients who were treated with adjuvant chemotherapy (n = 121). PDAC, pancreatic ductal adenocarcinoma; PCR, polymerase chain reaction; *M. globosa*, *Malassezia globosa*; N.C., negative control; HR, hazard ratio; CI, confidence interval; LNM, lymph node metastases; LVI, lymphovascular invasion; CEA, carcinoembryonic antigen; CA19-9, carbohydrate antigen 19-9.