Synthesis and Conformational Analysis of Hydantoin-Based Universal Peptidomimetics

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(NMR, circular dichroism (CD), Fourier transform infrared (FTIR)), and in solid state (X-ray) showed that the presented hydantoin-based peptidomimetics are able to project their substituents in positions superimposable to the side chains of common protein secondary structures such as *α*-helix and *β*-turn, being the open *α*-helix conformation slightly favorable according to molecular modeling, while the closed *β*-turn conformation preferred in solution and in solid state.

■ **INTRODUCTION**

"The most fruitful basis for the discovery of a new drug is to start with an old drug". This is a sentence stated in 1988 by Nobel Laureate Sir James Whyte Black.^{[1](#page-18-0)} Although almost 35 years have passed, the statement is still as relevant today. Indeed, there are highly favorable scaffolds, referred to as privileged scaffold, that are found in many synthetic drugs and can serve as templates to generate structurally diverse bioactive molecules for targeting more than one receptor type, including traditionally "undruggable" targets.² The judicious decoration of these scaffolds, mostly heterocycles, has provided, and still provides, a useful strategy in medicinal chemistry. Indeed, from early "lock and key" theory and combinatorial chemistry to the modern drug repositioning, 3 3 privileged substructure-based diversity-oriented synthesis $(p\bar{D}OS)_+^4$ biology-oriented synthesis (BIOS), 5 5 and complexity-to-diversity (CtD) strategy, 6 6 privileged scaffolds serve as "chemical navigators" to design targeted libraries first, and then to cover unexplored chemical space.

Privileged scaffolds have been exploited also for the design of structural peptidomimetics, encompassing minimalist and universal mimetics, which are a particular class of peptidomimetics where a privileged scaffold that does not possess a peptide character is used as starting point to build structures able to mimic the secondary structures of the proteins, such as *α*-helix and *β*-turn.⁸ Since the pioneering works by Smith and Hirschmann, who used sugar, steroid, and catechol scaffolds for the design of β -turn mimetic,^{[9](#page-19-0)} and Hamilton, who developed terphenyl and related scaffold α -helix mimetics,^{[10](#page-19-0)} many other templates have been used to mimic one preferred peptide secondary conformation, such as pyrrolidine, 11 $\frac{1}{2}$ pyrrolopyrimidines, $\frac{13}{2}$ $\frac{13}{2}$ $\frac{13}{2}$ oligooxopiperazine, $\frac{14}{2}$ $\frac{14}{2}$ $\frac{14}{2}$ bicyclic lactam, 15 benzodiazepine, 16 among others. These mimetics, *i.e.*, minimalist mimetics, possess conformations with energies similar to the global minima where their substituents overlap with some of the key $i + n$ positions of α -helices or β turn. More recently, Burgess and co-workers have introduced the concept of universal peptidomimetics, which are a particular class of structural mimetics designed on scaffolds that being not too rigid are able to mimic different, if not all, secondary structures through rotation around a few of significant degrees of freedom.^{[17](#page-19-0)} Being easy to be functionalized with substituents corresponding to many of the side chains of the protein-derived amino acids, universal peptidomimetics could be very useful for the design of libraries for high-throughput screening programs against diverse targets, including conventionally "undruggable" targets, *i.e.*, those

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proteins that are not yet being targeted.¹⁸ Heterocycles as $\overline{\text{omega}}$ omegatides,^{[19](#page-19-0)} piperidine-piperidinones,^{[20](#page-19-0)} and pyrrolinonepyrrolidine oligomers 21 have been used as core skeletons for the construction of universal peptidomimetics able to target local pairs of amino acids in ideally any secondary structure.

Thanks to their relative flexibility, universal peptidomimetics are very intriguing because, when the binding is mostly governed by the side chains (for example, in protein−protein interactions), they can interact selectively their targets displaying their side chains in the appropriate conformations,

Table 1. Regioselective Synthesis of Hydantoin Intermediates 5

Table 1. continued

overall when the exact binding conformations of the target is unknown. For this reason, there is a great interest in (1) finding new scaffolds for universal peptidomimetics and (2) developing convenient synthetic strategies for the preparation of libraries to supply high-throughput screening programs.

Hydantoin, being a small heterocycle having four derivatizable positions to generate diversity and a maximum of four hydrogen-bond donors/acceptors, are a class of very intriguing, privileged scaffolds exhibiting a wide spectrum of biological activities as evidenced by the number of marketed drugs as anticonvulsants, postsynaptic muscle relaxants, and androgen receptor agonists, and clinical candidates in the treatment of psoriasis, LFA-1 antagonist, and in the treatment of Duchenne muscular dystrophy[.22](#page-19-0) Indeed, even if hydantoin heterocycle could be considered an "old" privileged scaffold, over the past years, the synthetic and pharmaceutical interest in these heterocycle-based compounds, encompassing more elaborated fused- and spiro-hydantoins, has not experienced a decrease in interest as evidenced by the number of publications and patents appeared in the literature dealing with both methodological and medicinal chemistry. The hydantoin core has been also used also as privileged skeleton for the design of minimalist and universal peptidomimetics. 23 In this context, we have described the design and the conformational analysis of racemic, universal peptidomimetic 3-*cyclo*-butylcarbamoyl hydantoins which were synthesized through a regioselective multicomponent (MC) domino process starting from *α*-azido- α *cyclo*-butyl carboxylic ester and isocyanates ([Scheme](#page-1-0) 1).²⁴ Modeling experiments have shown that these compounds can exist at room temperature (rt) in kinetically accessible *α*-helix and *β*-turn conformations which can exchange with no significant entropic penalties, presenting the substituents R^1 , R^2 , and R^3 to the classical $i + n$ side-chain positions of secondary structures. To clarify the importance of the exoquaternary carbon on the stabilization of the conformations

and to widen the synthetic scope of this class of universal peptidomimetics, we present herein the synthesis and conformation analysis of (1) analogues having different quaternary substituents, such a *gem*-dimethyl, *cyclo*-hexyl, and *cyclo*-propyl, and (2) analogues lacking the substituent in that position (glycine derivatives) ([Scheme](#page-1-0) 1). Although the straightforward MC process reported earlier has been the first choice for the synthesis of these derivatives, it failed to give a regioselective process in some cases and high yields in others (see below). For this reason, we exploited a chemoselective process involving only liquid−liquid acid/base extractions for the isolation and purification of all intermediates, thus suitable for combinatorial synthesis purposes. In addition, this synthetic pathway allowed us to prepare a selected collection of enantiomerically pure derivatives instead of racemic mixtures that we would have obtained with the MC process.

■ **RESULTS AND DISCUSSION**

Synthesis. The MC process exploited for the synthesis of racemic 3-*cyclo*-butylcarbamoyl hydantoin universal peptidomimetics F relies on a regioselective intermolecular aza-Michael addition of the *O*-acyl isourea intermediate E formed by a reaction between *in situ* generated carbodiimides C and *α*,*β*-unsaturated fumaric acid mono esters **D** ([Scheme](#page-1-0) 2).^{[25](#page-19-0)} The regiocontrol of the reaction is dictated by the difference between the steric hindrance of the two nucleophilic amino moieties, *i.e.*, one bearing the bulky *cyclo*-butyl $(R^2, R^3 =$ −(CH2)3−) *versus* the other bearing a methylene group. Due to the efficiency, we attempted to exploit the same process for the synthesis of derivatives having different substituents on the azide A. In particular, we sought that other azides linked to a tertiary carbon could have given the same regiochemical control leading to the formation of only one isomer. Indeed, the process was regioselective with *gem*-dimethyl glycine azide

Scheme 3. Synthesis of Hydantoin Intermediate 5h

Scheme 4. Synthesis of Hydantoin Universal Peptidomimetics 8

 $(R² = R³ = Me$, eq 1, [Scheme](#page-1-0) 2) and 1-azidocyclohexane carboxylic acid benzyl ester $(R^2, R^3 = -(CH_2)_5$ –, eq 2, [Scheme](#page-1-0) [2](#page-1-0)) leading to the formation of hydantoins F1 and F2, respectively, but in low yields. This is probably due to the steric hindrance in the reaction between the starting azide and Ph₃P during the first step of the process. Starting with 1azidocyclopropane carboxylic acid benzyl ester $(\mathbb{R}^2, \mathbb{R}^3 =$ $-(CH₂)₂$ -, eq 3, [Scheme](#page-1-0) 2) the process occurred with high yield but low regiocontrol (3:1 mixture of regioisomers F3), probably due to the constrain in the cycle that renders the azido group more accessible in the Staudinger reaction but, at the same time, the resulting amino moiety not much more sterically congested than the amino group bearing a primary carbon in the *O*-acyl isourea intermediate E. Finally, we tried the reaction with azido glycine benzyl ester $(R^2, R^3 = H, eq$ 4, [Scheme](#page-1-0) 2) hoping that the higher nucleophilicity of an alkyl amine compared to the amino group belonging to the glycine moiety would lead certain stereocontrol. However, although the yield was high, the process occurred with no regiocontrol, and an equimolecular mixture of hydantoins F4 was obtained.

With these results in hand, we sought an alternative synthetic pathway that could be exploited for the combinatorial synthesis of libraries of hydantoin-based universal peptidomimetics adopting a simple, technically not demanding multistep protocol that would allow us to avoid tedious chromatographic purifications of the intermediates by employing simple solution-phase protocols based on acid/base liquid/liquid extractions.^{[26](#page-19-0)} Actually, a convenient method to synthesize *N*,*N*′-disubstituted hydantoins under mild conditions consists in the reaction between isocyanates and *N*-alkyl-*α*-amino esters which occurs through a condensation/cyclization domino process [\(Scheme](#page-1-0) 1).^{[27](#page-19-0)} However, to the best of our knowledge, this protocol has never been used starting with *N*-alkyl aspartic acid esters and *α*-aminoester isocyanates, which could give a non-regiospecific process yielding a mixture of the desired hydantoin scaffold 5 along with the six-membered ring 6

arising from the attack of the urea-NH to the ester moiety on the aspartic acid side chain during the cyclization step (see scheme in [Table](#page-2-0) 1). 28 28 28 It is worth noting that if successful, this procedure would allow us to prepare enantiomerically pure hydantoin universal peptidomimetics in contrast to the reported MC domino process through which we synthesized 3-*cyclo*-butylcarbamoyl hydantoins in racemic form ([Scheme](#page-1-0) [1](#page-1-0)).

To check the regioselectivity of the cyclization, we synthesized the urea derivatives 4a and 4b by treating isocyanate 2a and 2b with *N*-*iso*-pentyl and *N*-benzyl aspartic esters 3a and 3b, respectively, in dichloromethane (DCM) at 0 °C for 1 h (entries 1 and 2, [Table](#page-2-0) 1). To our delight, either cyclization triggered by the treatment of 4a and 4b with a strong base (typically 1.0 M NaOH in Schotten−Baumann conditions) in a very short time (less than 5 min) or by leaving overnight 4a and 4b in the presence of a weaker organic base (typically triethylamine (TEA) in DCM solution) yielded the formation of the corresponding hydantoin 5a and 5c with total regiocontrol and very good yields (data not shown).^{[29](#page-20-0)}

With this result in hand, we decided to synthesize a collection of seven systematically modified hydantoin derivatives 5a−h by performing the addition/cyclization process one-pot, *i.e.*, by leaving the reaction with an excess of TEA overnight or, when the cyclization process is not complete in the former conditions, by adding a 1 M aqueous NaOH solution at the end of the condensation step ([Table](#page-2-0) 1). To our delight, the process worked efficiently also with glycine ester isocyanate (entries 8 and 9, [Table](#page-2-0) 1) demonstrating that the regioselectivity achieved in the cyclization step is not driven by the presence of a quaternary carbon which could stabilize a particular reactive conformation. In this way, we obtained hydantoin scaffolds having (1) different exo-quaternary carbon on the N-3 substituent, such as *cyclo*-propyl (entries 3 and 4, [Table](#page-2-0) 1), *cyclo*-hexyl (entry 5, [Table](#page-2-0) 1), *gem*-dimethyl (entries 6 and 7, [Table](#page-2-0) 1), or unsubstituted glycine derivative (entries 8

Chart 1. Structures of Hydantoin Universal Peptidomimetics 8

Ph

 $8p$

HŃ

8q

and 9, [Table](#page-2-0) 1); (2) different substituents on the N-1 position, such as *iso*-pentyl (entry 3, [Table](#page-2-0) 1), benzyl (entries 4, 5, and 8, [Table](#page-2-0) 1), ethyl (entry 6, [Table](#page-2-0) 1), naphthyl (entries 7 and 9, [Table](#page-2-0) 1); and (3) two carboxyl groups orthogonally protected

as benzyl and *tert*-butyl esters (entries 3−5 and 8−9, [Table](#page-2-0) 1) or benzyl and methyl esters (entries 6 and 7, [Table](#page-2-0) 1) ready to be functionalized after selective hydrolysis. It is worth noting that all of these intermediates have been recovered in high

Figure 1. *β*-Turn- and *β*-helix-like conformations of universal peptidomimetics 8.

yields and purities after simple acid/base extraction procedures and used in the following reactions without any further purification. We applied the process also starting from enantiomerically pure *α*-aminoester isocyanates such as leucine benzyl ester isocyanate 2e to have another point of diversity in the final hydantoin-based peptidomimetics ([Scheme](#page-4-0) 3). However, while the addition step worked nicely producing enantiomerically pure intermediate 4c in good yields, all of the attempts made for the cyclization step yielded an almost equimolar mixture of two diastereoisomeric hydantoins 5h, due to epimerization of the stereocenter of the reacting isocyanate 2e. Moreover, we obtained the same epimerization also during the following coupling steps (data not shown), demonstrating the extreme stereochemical instability of this stereocenter.

Starting from intermediates 5a−g the following steps were performed using standard hydrolysis/condensation protocols ([Scheme](#page-4-0) 4), *i.e.*, trifluoroacetic acid (TFA)-promoted hydrolysis of the *tert*-butyl ester of intermediate 5b and coupling with 2,2-diphenylethylamine leading to hydantoin monoamide 7a followed by hydrogenolysis of the benzyl ester and coupling with hexylamine yielding 8a, or hydrogenolysis of the benzyl ester of 5a, 5c−g catalyzed by Pd/C and coupling promoted by 2-(1*H*-benzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU) of the resulting free carboxylic acid with a first amine yielding compounds 7b−i, followed by acid hydrolysis of the *tert*-butyl ester or NaOH promoted hydrolysis of the methyl ester and coupling in the same conditions with a second amine producing final peptidomimetics 8b−r.

Accordingly, we recovered in good yield and excellent purities a collection of 18 hydantoin-based universal peptidomimetics 8a−r ([Chart](#page-5-0) 1) having different types of substituents, from highly hydrophobic to polar, with a synthetic protocol based in operationally simple and timesaving liquid−liquid acid/base extractions which did not require further chromatographic purification, thus suitable for combinatorial synthesis/high-throughput screening programs. It is worth noting that the key substituents $\mathsf{R}^1,\,\mathsf{R}^2,$ and R^3 have been introduced using simple, commercially available aldehydes (\mathbf{R}^1) and amines $(\mathbf{R}^2,\ \mathbf{R}^3)$ and easy conventional synthetic protocols (reductive amination and coupling reactions, respectively), which render the new synthetic pathway designed herein hypothetically conform to introduce all of the natural (and unnatural) amino acid side chains in the key *i* + *n* positions of *α*-helices or *β*-turn.

Conformational Analysis *In Silico***: Computation.** The propensity of molecules 8a−r to adopt a defined secondary structure was investigated by computational tools. The structures were first submitted to a conformational search by a combined Monte Carlo−Molecular Mechanics (MM)

approach. For each structure, the conformers within 10 kcal/ mol from the minimum were considered. Different parameters were measured to establish whether an *α*-helix or *β*-turn conformation was present, according to the literature (Figure 1).^{[17](#page-19-0)} The three atoms C*i*, C*i* + 4 and C*i* + 7 were related to the *i*, $i + 4$ and $i + 7$ residues of an ideal α -helix, and the ideal interatomic distances $i - i + 4 = 6.2$ Å, $i - i + 7 = 10.3$ Å and i $+ 4 - i + 7 = 5.8$ Å were used as reference. For the *β*-turn conformation, the interatomic distance $d\alpha$ < 7 Å and the absolute value of the dihedral angle C1−C2−C3−N4 *β* < 60° were considered as a condition. The presence of the intramolecular 10-membered ring H-bond was also evaluated. We also considered the less common 3_{10} helix motif, and we found that the compounds 8a−r can efficiently mimic this structure by placing the substituents in coincidence with the *i*, *i* + 2 and *i* + 4 residues of the helix. Results are reported as percentage of conformers meeting the requirements (Table 2 and [Figure](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S1).

β-Turn is described by the presence of the distinctive intramolecular hydrogen bond involving the hydrogen NH_B and forming a 10-membered ring, whereas the *α*-helix structure is established by the presence of a two consecutive *γ*-turn type hydrogen bonding (both NH_A and NH_B are involved) around the hydantoin central core (Figure 1). In general, the *α*-helix

Table 2. Results from Monte Carlo/MM Conformational Analysis*^a*

compound	β -turn (%)	α -helix (%)	3_{10} -helix	global minimum
8a	23	52	11	α -helix
8b	7	66	19	α -helix
8с	9	63	11	α -helix
8d	18	50	19	α -helix
8e	44	54	35	β -turn
8f	10	51	10	α -helix
8g	23	$\mathbf{0}$	$\mathbf{0}$	n.d.
8h	14	52	5	α -helix
8i	15	48	7	n.d.
8j	14	61	10	α -helix
8k	$\mathbf{1}$	60	1	α -helix
81	46	30	$\overline{4}$	β -turn
8m	23	46	10	α -helix
8n	11	43	15	α -helix
80	18	34	9	β -turn
8p	39	23	5	β -turn
8q	21	34	13	β -turn
8r	12	44	11	α -helix

a Results are reported as a percentage of conformers meeting the geometrical requirements for $β$ -turn, $α$ -helix, and 3₁₀-helix.

conformation is preferred because of the higher percentage of conformers adopting this structure for each compound. Looking at the global minimum conformers, the *β*-turn geometry is the most favored in particular for compounds 8o−q having two hydrogen atoms on C3 carbon ($R_4 = R_5 = H$, [Figure](#page-6-0) 1), while for the *gem*-dimethyl 8k−m and cycloalkyl derivatives 8a−j, the *α*-helix is preferred. This last result is in line with that obtained in the previous work for 3-*cyclo*-butylcarbamoyl hydantoins.^{[23](#page-19-0)} For selected molecules 8a, 8m, 8o−r, a density functional theory (DFT) study was performed at the B3LYP6-311G (d,p) level. For each structure, the first lower-energy *α*-helix and *β*-turn conformers were optimized in vacuo. All of the energies were corrected for the zero-point energy (ZPE). Unlike that obtained from MM force field, results from DFT showed that the most stable secondary structure is the α -helix. This difference can be ascribed to the more accurate calculation of the energy from DFT with respect to molecular mechanics. The energy gaps between the *α*-helix and the β -turn conformers varied from 0.17 kcal/mol (8r) to 5.05 kcal/mol (8m). It can be then assumed that in most cases the herein proposed scaffolds can access both the *α*-helix and the *β*-turn secondary structures according to the environment. For compound 8r the energy of the *β*-turn structure found by X-ray single-crystal analysis (*vide infra*) was also calculated without further optimization, resulting to be 2.85 kcal/mol higher than the *β*-turn conformer as obtained from conformational analysis and DFT optimization, indicating a decisive role of the crystal packing in determining the conformation of the residues around the *β*-turn core. Superimposition of the *β*-turn conformation with theoretical *β*-turn models revealed the ability to mimic preferentially the type II β -turn (see rootmean-square deviation (rmsd) values of the backbone in Table 3 and Figure 2). Acceptable rmsd values have been measured

Table 3. Results from DFT Studies

Single point energy of the structure obtained from single-crystal λ ray analysis.

also for the type I′ *β*-turn. The type I and II turns mostly differ in the orientation of the central amide bond. The presence of the hydantoin core allows for the good superimposition of the two orientations thanks to the presence of the two carbonyls of the imide moiety (Figure 2). A comparison within the investigated molecules seems to indicate that the presence of the two hydrogen atoms on C3 carbon is favorable in stabilizing the type II *β*-turn, with compounds 8o−r having the lowest rmsds.

Conformational Analysis in Solution: NMR, Circular Dichroism (CD), Fourier Transform Infrared (FTIR). To investigate the forces that stabilize the secondary structure, we relied on the behavior of the intramolecular hydrogen-bond network of our molecules in solution. Two patterns were

Figure 2. (a) *α*-Helix (left)- and *β*-turn (right)-like conformations for representative compound 8r; (b) superimposition of 8r conformers with an α -helix model in orange (left), 3_{10} -helix model in orange (center), and *β*-turn in green (right). For the *α*-helix, the relevant *i*, *i* + 4, and *i* + 7 positions are highlighted.

identified in our previous work: (1) the α -helix is stabilized by two H-bonds involving both carbonyls on the hydantoin ring and both H_A and H_B , while (2) the β -turn conformation is stabilized by another H-bond not involving any hydantoin-ring carbonyl and only amide NH_B^{24} NH_B^{24} NH_B^{24} An important parameter to assess the presence of hydrogens involved in hydrogen bonds is the chemical shift in a relatively nonpolar solvent such as $CDCI₃$, according to which higher values, typically around 8 ppm, are more related to an internal hydrogen bond. The value of the chemical shifts of the amidic protons H_A and H_B of those compounds for which it was possible to record the ¹H NMR spectrum in deuterated chloroform (2.5 mM solutions) 30 are reported in [Table](#page-8-0) 4.

 H_B protons resonate at lower field than H_A protons in all cases, most of them at ppm higher than 8, indicating their involvement in the formation of hydrogen bonds. Moreover, the rates of H_B/D exchange in ¹H NMR spectra recorded in CD₃OD were very slow for compound 8k, 8l, 8o. Indeed, the spectra of 8k and 8l showed the presence of the amide proton H_B at 8.01 and 8.10 ppm, respectively, integrating for one hydrogen, thus indicating that no exchange occurred, while for 8o there is a peak at 8.16 ppm integrating for 0.25 meaning that only a 75% of hydrogens H_B underwent H/D exchange (see spectra in the Supporting [Information\)](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf). In all spectra recorded in $CD₃OD$, we did not detect the presence of a peak for hydrogen H_A which exchanges quickly with deuterium.

To study more in-depth the presence of these interactions for the more flexible peptidomimetics, namely, the glycyl derivatives having two hydrogen atoms on C3 carbon, we performed variable-temperature $(VT)^{-1}H$ NMR analysis on compounds 80, 8p, 8r in $CDCl₃$ and compounds 8p, 8r in dimethyl sulfoxide (DMSO)-*d*⁶ 2.0 mM solutions. In the relatively nonpolar solvent CDCl₃, low-temperature coefficient values, typically lower than 2.4 ppb/K, are not always very indicative since they can be attributed either to shielded protons or to accessible ones. On the contrary, values significantly larger than 2.4 ppb/K can be assigned to NH protons involved in intramolecular hydrogen bonds which

Table 4. ¹ ^H NMR Data for Hydantoin-Based Universal Mimetics ⁸*^a*

Figure 3. DMSO titration experiments on substrates 8p, 8r.

become accessible to the solvent upon increasing temper-ature.^{[31](#page-20-0)} The $\Delta\delta/\Delta T$ values obtained for H_B of peptidomimetics 8o, 8p, 8r are in all cases very high (22.2, 40.1, and 20.6 ppb/K, respectively, Table 4 and [Figures](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S2−S4) suggesting the involvement of the amide proton in intramolecular hydrogen bonding that is disrupted with the temperature. Interestingly, also the values for H_A are high, although to a lesser extent (Table 4 and [Figures](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S2−S4). This could be explained by the possible involvement of the latter amide proton in less strong hydrogen bond as expected in an *α*-helix conformation. These results corroborate the possibility for the more flexible glycine peptidomimetics to exist in solution as an equilibrium between *β*-turn conformation triggered by an intramolecular hydrogen bond involving H_B and α -helix conformation stabilized by two intramolecular hydrogen bonds involving both H_B and H_A , being the first the preferred one. In very polar solvents, such as DMSO, typically, solventaccessible protons exhibit a $\Delta \delta / \Delta T > 5$ ppb/K, whereas $\Delta \delta /$ $\Delta T < 5$ ppb/K denotes protons involved in H-bonds.^{[32](#page-20-0)} The value found in $8r$ for NH_B is smaller than 5 ppb/K and smaller than that found for NH_A (Table 4 and [Figure](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S6), indicating a preference of NH_B to be involved in intramolecular Hbonding. Interestingly, for molecule 8p, both values are below the 5 ppb/K $\Delta\delta/\Delta T$ threshold (Table 4 and [Figure](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S5), the value for H_A being slightly lower than that of H_B , suggesting that both amide hydrogens are involved in H-bonds. These results suggest a stronger preference to adopt a folded

conformation (β -turn, only NH_B H-bond is present) for 8r and a slight preference for 8p to an open conformation (*α*helix, both NH_A and NH_B are hydrogen-bonded).^{[33](#page-20-0)} The extent of H-bonding can be evaluated also by performing DMSO titration experiments. DMSO is added in small aliquots $(5 \mu L)$ gradually to diluted $CDCl₃$ solutions (2.0 mM) of the compounds of interest. A 1H NMR spectrum is then recorded after each addition, and the chemical shift of the H-bonded protons is plotted against the DMSO aliquots. H-bonding generally results in deshielding, and an increasingly downfieldshifted proton resonance indicates increased H-bond strength.^{[34](#page-20-0)} As the concentration of DMSO increases, the resonance line shifts downfield as a result of increasing Hbonding interactions with DMSO. In contrast, only small changes are seen for the protons involved in intramolecular Hbonding, thus indicating that DMSO cannot compete with the proposed intramolecular H-bond.

We performed DMSO titration experiments on compounds 8p, 8r, and the results are shown in Figure 3 (see also [Figures](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S7 [and](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S8, respectively). As can be seen in Figure 3, the chemical shifts of H_B in compounds 8p, 8r essentially did not change upon dilution with DMSO, whereas we observed a clear increase of the chemical shifts of amidic hydrogens H_A , indicating the presence of a strong intramolecular hydrogen bond involving only the amide proton H_B in all compounds. All of the observations obtained with the NMR experiments seem to underline a preference of the *β*-turn conformation in

Figure 4. CD spectra for peptidomimetics 8p, 8r.

Figure 5. ATR-FTIR spectrum of 8r.

solution. However, two-dimensional (2D) nuclear Overhauser enhancement spectroscopy (NOESY) experiment on glycine derivatives 8o−r did not show evident long-range contacts supporting the possibility that no preferential conformation is adopted.

The ability of universal peptidomimetic 8p, 8r to adopt secondary structure in solution was assessed also by CD spectroscopy. Figure 4 shows the spectra recorded in methanol $(10^{-5}$ M solutions)

The CD profile of compound 8r has a negative absorption band at 194.9 nm and a second positive band centered at 225.4. This profile can be addressed to a right *β*-turn in accordance with those reported in the literature. 35 The results for molecule 8p are even more interesting. In addition to having the typical characteristics of a right *β*-turn as described above (negative band at 194.4 nm, positive band at 225.7 nm) the spectrum has two additional absorption bands, a positive one at 203.4 nm and a negative one at 232.8 nm. This phenomenon reflects a possible contribution from aromatic r esidues π -stacking interactions,^{[36](#page-20-0)} further stabilizing our proposed *β*-turn conformation.

Finally, we registered the attenuated total reflection (ATR)- FTIR spectrum of peptidomimetic 8r, which was the most soluble in chloroform, as model compound for the flexible glycine hydantoin-based universal peptidomimetics (Figure 5). The spectrum was run by depositing a thin layer of 8r in diluted chloroform solution (20 mg/mL) between two quartz plates. Based on literature data, the band at 3431 cm⁻¹ was assigned to the free NH groups, and a second one, more intense, appearing at 3332 cm^{-1} to NH groups involved in an intramolecular hydrogen bonding.³⁷ These findings show that, in solvents of low polarity such as chloroform, secondary structures promoted by intramolecular hydrogen bonds significantly populate the conformational equilibrium of the molecule in solution.

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Conformational Analysis in Solid State: X-ray. Single crystals of 8r were obtained by the slow evaporation of a 1:1 water/acetone solution, after 1 week. Crystallographic data and refinement details are given in Table 5.

Table 5. Crystal Data and Structural Parameters of Compound 8r

no. of measured, independent, and observed $[I > 2\sigma(I)]$ 6841, 3544, 3043 reflections

Compound 8r crystallized in the monoclinic space group *C*2; its structure is shown in Figure 6 as an Oak Ridge thermal ellipsoid plot $(ORTEP)$ diagram,³⁸ indicating the arbitrary atom-numbering scheme used in the following discussion.

Notably, in solid state, the molecule adopts a *β*-turn conformation stabilized by an intramolecular hydrogen bond that persisted during the crystal formation even under the presence of a highly competitive protic solvent $(H₂O)$ in the crystallization solution.

The molecular structure of 8r is characterized by a hydantoin nucleus, substituted at the two nitrogen atoms and at the C3 carbon. The hydantoin ring is planar, with a maximum deviation of 0.032(4) Å (C2) from the best mean plane. The substituents at N1 and C3 extend toward the same direction, with torsion angles of $-61.2(9)$ and 97.2(9)° for C2−C3−C13−C14 and C2−N1−C4−C5, respectively. The divergence between the two values is motivated by the different position of the methylene group bridging the hydantoin nucleus and the amide. While C13 is linked to an asymmetrical carbon (in the *S* configuration), C4 is directly attached to N1 and, thus, remains coplanar with the central ring. The same can be said for C29, which connects N2 to the naphthalene; in this case, the aromatic moiety is oriented in the opposite direction with respect to the other two substituents, with a torsion angle of $-131.6(7)$ ° for C1-N2−C29−C30. Both the naphthalene ring and the *p*chlorophenyl moiety are almost perpendicular to the hydantoin nucleus; in detail, the former forms an angle of 80.3° with the central heterocycle, while the latter is inclined at 86.6°.

The crystal packing ([Figure](#page-11-0) 7) is mainly ensured by two strong H-bonds, one of them, being intramolecular and involving amidic NH_B (N3−H3 in the ORTEP diagram), responsible for the adopted *β*-turn conformation. In detail, the former intramolecular contact (N3−H3N···O4) links the two elongated hydantoin substituents (D···A: 2.727(9) Å; D−H··· A: 1.965(6) Å; D−H···A: 147.2(5)°), while the other, which is

Figure 6. ORTEP diagram of 8r, with the arbitrary atom-numbering scheme. Thermal ellipsoids are drawn at the 40% probability level.

Figure 7. (A) Stick model of 8r in an arbitrary orientation, evidencing the main H-bonds. (B) Stick model showing the crystal packing along the *c* axis. Hydrogen atoms are omitted for the sake of clarity.

intermolecular (N4−H4N···O3^I , ^I at *x*, *y* − 1, *z*), connects the 8r molecules (D···A: 2.843(9) Å; D−H···A: 2.001(6) Å; D− H···A: $166.1(5)°$) forming a chain. Weak-to-very-weak intramolecular and intermolecular C−H···O contacts and a C−H··· Cl interaction contribute to stabilize the network. A complete account of the H-bonds is provided in the [Supporting](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) [Information](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) [\(Table](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf) S1). Despite the considerable number of aromatic moieties, the packing is not significantly influenced by stacking interactions; the only exception is a very weak, almost parallel *π*−*π* contact between two phenyl groups of the benzhydryl moieties of two adjacent molecules (centroid− centroid distance: 4.079 Å; angle between planes: 8.0°). However, numerous C−H···*π* interactions are established between the various aromatic rings.

The Hirshfeld surface (HS) of the 8r was mapped over the normalized contact distance (d_{norm}), according to the following equation

$$
d_{\text{norm}} = \frac{d_i - r_i^{\text{vdW}}}{r_i^{\text{vdW}}} + \frac{d_e - r_e^{\text{vdW}}}{r_e^{\text{vdW}}}
$$

where d_i is the distance between the HS and the nearest nucleus inside the surface, d_e is the distance between the HS and the nearest nucleus outside the surface, and $r^{\rm vdW}$ represents the van der Waals radius of the atom. Details of the HS are provided in Table 6.

Table 6. Characteristics of the HS Generated for 8r

8r	$V(\AA^3)$	$A(\AA^2)$	G	
HS	845.05	641.95	0.673	0.191

The d_{norm} property was visualized with a red-blue-white color scheme, based on the length of the intermolecular contact with respect to the sum of the van der Waals radii ([Figure](#page-12-0) 8A). The inspection revealed a rather unperturbed surface, with only two major red spots corresponding to the short-range H-bond connecting the amide groups of adjacent molecules. Additional small and very feeble spots indicate the presence of minor C−H···O interactions and a weak C−H···Cl contact. The surface mapped over the shape index ([Figure](#page-12-0) 8B) confirmed the absence of strong *π*−*π* stacking interactions; the only notable feature is the presence of deep hollow regions, indicated by the red color, especially in the vicinity of the benzhydryl substituent. Finally, the curvedness plot [\(Figure](#page-12-0) [8](#page-12-0)C) showed the absence of large flat areas, despite the abundance of aromatic moieties, further confirming the marginal contribution of stacking interactions to the overall packing.

The two-dimensional (2D) fingerprint of the HS ([Figure](#page-12-0) 9), providing a visual summary of the contribution of each contact type and the relative area of the surface corresponding to it, revealed a prominence of nonspecific van der Waals H···H contacts (49.1%) . The wings of the plot are occupied by C \cdots $H/H...C$ interactions (24.7%), which include the numerous, weak C−H···*π* H-bonds. O···H/H···O contacts (13.3%) also offer a significant contribution to the surface, representing the strong H-bonds that interconnect the molecules. Cl···H/H···Cl interactions (7.9%) occupy an appreciable portion of the surface, without being particularly relevant for the crystal network. Finally, C···C contacts (1.6%), forming the characteristic arrow-shaped region at the center of the plot, only marginally contribute to the surface. The remaining interactions are negligible and are indicated in [Figure](#page-12-0) 8. These observations were supported by the analysis of the contact enrichments [\(Table](#page-13-0) 7).^{[38](#page-20-0)} The calculations showed that Cl···H/H···Cl, O···H/H···O, and C···H/H···C interactions are enriched $(E_{XY} \geq 1)$ with respect to the corresponding random contacts.

■ **CONCLUSIONS**

In conclusion, we have conceived a new scaffold for universal peptidomimetics based on hydantoin ring. The synthetic strategy for these compounds is based on a chemoselective domino condensation/cyclization process between *α*-aminoester isocyanates and *N*-alkyl aspartic acid diesters which occurs in mild condition (room temperature) followed by conventional deprotection/coupling reactions. All of the intermediates were recovered through time-saving liquid− liquid acid/base extraction procedures in pure form, thus not needing any further chromatographic purification. We synthesized in this way a collection of 18 enantiomerically pure, systematically substituted hydantoins having either different quaternary carbons at the $exo-C_3$ position (*cyclo*hexyl, *cyclo*-propyl, *gem*-dimethyl) or a more flexible methylene group. It is worth noting that a wide range of natural or unnatural amino acid side chains could be incorporated since they come from easily accessible aldehydes (R^1) and amines $(R²$ and $R³$). All of these characteristics render the synthetic

Figure 8. (A) HS mapped over d_{norm} with a fixed color scale in the range −0.5808 au (red) to 2.9223 au (blue), based on the length of the intermolecular contacts with respect to the sum of the van der Waals radii (red: shorter; blue: longer; white: same). (B) HS mapped over the shape index (color scale: −0.9973 to 0.9977 au). Blue areas represent bumps, and red regions indicate hollows. (C) HS mapped over the curvedness (color scale: −4.4169 to 0.8187 au). Green represents flat regions, and blue indicates edges.

Figure 9. Two-dimensional Fingerprint plots of HS, providing a visual summary of the frequency of each combination of d_e and d_i across the HS. Points with a contribution to the surface are colored blue for a small contribution to green for a great contribution.

strategy presented particularly suitable for combinatorial synthesis/high-throughput screening programs. The conformational behavior of the peptidomimetics was studied *in silico*, by molecular modeling, in solution, by NMR, CD, and IR experiments, and in solid state through X-ray analysis. Molecular modeling showed that these scaffolds can adopt kinetically and thermodynamically accessible, intramolecular hydrogen-bond-driven conformations where the key substituents are projected in positions superimposable to some of the key i , $i + n$ side chains of protein secondary structures such as *α*-helix and *β*-turn, the first one being the favored for most of the scaffolds, either for those having a quaternary exo-carbon

or for the more flexible glycine derivatives. On the contrary, VT -NMR experiments recorded in nonpolar $CDCl₃$ and polar DMSO-*d*⁶ and DMSO titration NMR experiments, both performed on the more flexible molecules having two hydrogens bonded to the C-3 carbon (glycine derivatives 8o−r) evidenced the presence in solution of a stronger intramolecular hydrogen bond engaging amide NH_{B} , which is responsible for both *α*-helix and *β*-turn conformations, compared to that involving NH_A (responsible for the only α helix conformation), suggesting the presence of a more populated *β*-turn conformation. Analogously, X-ray analysis showed that glycine derivative 8r adopts in solid state a well-

Table 7. Analysis of the Intermolecular Contacts on the HS of 8r, According to Jelsch et al.^{[39](#page-20-0),*a*}

atoms	H	C	$\mathbf N$	\circ	C ₁
surface $(\%)$	72.2	15.2	0.5	7.8	4.4
contacts (%)					
Н	49.1				
C	24.7	1.6			
N	0.3	0.6	0.0		
\circ	13.3	1.0	0.0	0.6	
Cl	7.9	0.9	0.0	0.0	0.0
enrichments					
Н	0.9				
C	1.1	0.7			
N					
\circ	1.2	0.4			
Cl	1.2	0.7			

^{*a*}The first part of the table gives the surface contribution S_X of each chemical type *X* to the Hirshfeld surface. The second part shows the proportions of the actual contacts (C_{XY}) , and the third part indicates the enrichment ratios (*E_{XY}*) of the various contact types. Reciprocal contacts X···Y and Y···X are merged. E_{XY} were not computed when the random contacts (R_{XY}) were lower than 0.9%. E_{XY} ratios larger than unity indicate enriched contacts (in bold), while those lower than unity are impoverished. The percentages of actual contacts were calculated using CrystalExplorer21.5.

defined *β*-turn conformation with the presence of the characteristic intramolecular hydrogen bond between NH_B and the carbonyl not belonging to the hydantoin ring. All of these features demonstrate that hydantoin scaffolds 8 can be considered a novel class of universal peptidomimetics able to adopt common protein secondary structures with favorable enthalpic and entropic profiles.

■ **MATERIALS AND METHODS**

Materials. Commercially available reagent-grade solvents were employed without purification. Thin-layer chromatography (TLC) was run on silica gel 60 F254 Merck. Visualization of the developed chromatogram was achieved with UV light and ceric ammonium molybdate (CAM) or ninhydrin stains. Flash chromatography (FC) was performed with silica gel 60 (60−200 µm, Merck). ¹H, and ¹³C NMR spectra were run at 400 or 500 MHz. Chemical shifts are expressed in ppm (*δ*), using tetramethylsilane (TMS) as internal standard for ¹H and ¹³C nuclei (δ H and δ C = 0.00). Electrospray ionization (ESI) mass spectra were performed by a Bruker Esquire 3000+ instrument equipped with an MS detector composed of an ESI ionization source and a single-quadrupole mass selective detector or by an Agilent Technologies 1200 series high-performance liquid chromatography (HPLC) system equipped with a DAD and a 6120 MS detector composed by an ESI ionization source and a singlequadrupole mass selective detector. Optical rotations were measured on a Propol Digital Polarimeter with a sodium lamp. Isocyanates 2a− d were synthesized according to the procedure described in ref [40](#page-20-0) and suddenly used in the next step, while *N*-alkyl *α*-aminoester 3 was synthesized according to ref [41.](#page-20-0) CD spectra were recorded on a Jasco J-1000 CD spectrometer at 25 °C, $V_{scan} = 10$ nm/min, DIT = 4 ms, three repetitions per sample, within a range of wavelength between 190 and 250 nm. The blank was preliminarily recorded and subtracted from the sample at each repetition. ATR-FTIR spectrum was run on a Jasco FT/IR-610. X-ray diffraction data for 8r were collected on a Bruker Apex II CCD three-circle diffractometer working at room temperature with graphite-monochromatized Mo K*α* radiation (*λ* = 0.71073 Å). X-ray data were acquired in the *θ* range of 2−21° recording four sets of 360 bidimensional CCD frames with the following operative conditions: omega rotation axis, scan width 0.5°, acquisition time 50 s, sample-to-detector distance 50 mm, *ϕ* angle

fixed at four different values (0, 90, 180, and 270°) for the four different sets. Omega rotation frames were processed using the SAINT software 42 for data reduction (intensity integration, background, Lorentz, and polarization corrections) and for the determination of accurate unit-cell dimensions, obtained by leastsquares refinement of the positions of 3623 independent reflections with $I > 10\sigma(I)$. Absorption effects were empirically evaluated by the SADABS software, 43 and an absorption correction was applied to the data. The structure was solved by direct methods with SIR-2019^{[44](#page-20-0)} and completed by iterative cycles of full-matrix least-squares refinement on F_o^2 and ΔF synthesis using the SHELXL-2018/3 program in the WinGX v2021.3 suite.^{[45](#page-20-0)} The hydrogen atoms were included at geometrically calculated positions and refined using a riding model. Uiso(H) was defined as 1.2Ueq of the parent atoms for phenyl, methylene, and methine residues. The structure was analyzed using
PARST^{[46](#page-20-0)} and Mercury 2021.3.^{[47](#page-20-0)} Graphical representations were generated with ORTEP-3 2020.1, 38 38 38 Mercury, 47 47 47 and CrystalExplorer $21.⁴⁸$ $21.⁴⁸$ $21.⁴⁸$

Computational Details. Conformational analysis was performed with the software Spartan'08 49 by means of the "conformer" distribution" function, using the Monte Carlo search method. For each compound, a variable number of 900−400 conformers were generated, according to the structure. The MMFF force field in vacuo was used for the energy minimization of the found structures. The structures were then clustered according to the default setting of the software (which consists in pruning out higher energy conformers and keeping a diverse set of the low-energy conformers using the RMStorsion definition of nearness). Full geometry optimization of selected lowest-energy conformations was then performed with DFT at the B3LYP 6-311G (d,p) level in vacuo with the software Gaussian'09.^{[50](#page-20-0)} All energies were corrected by adding the ZPE as obtained by frequency calculation at the same level.

General Procedure for the Synthesis of Urea Intermediates 4. To a solution of isocyanate 2 (0.5 mmol, 1 equiv) in DCM (0.3 M solution), a solution of TEA (2 equiv) and *N*-alkyl *α*-aminoester 3 (1.2 equiv) in DCM (0.3 M solution) was added at 0 °C. After 1 h, the reaction was quenched with a 1 N HCl aqueous solution, the temperature raised to rt, and the mixture was extracted with DCM. The collected organic phases were washed with a 1 N HCl aqueous solution (once), brine (once), a saturated aqueous solution of $NAHCO₃$ (twice), and brine (once). The organic phase was dried over $Na₂SO₄$, filtered, and the solvent evaporated. The urea intermediates 4 were used without any further purification.

4-(tert-Butyl)-1-methyl N-((1-((Benzyloxy)carbonyl)cyclopropyl) carbamoyl)-N-isopentyl-L-aspartate (4a). Yellow oil. Yield 87% (129 mg). *R_f* (hexane/AcOEt, 70:30) = 0.35; $[\alpha]_D^{20}$ -15.2 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 5.32–5.30 (m, 5H), 5.41 (br s, 1H), 5.14 (d, *J* = 12.4 Hz, 1H), 5.05 (d, *J* = 12.4 Hz, 1H), 4.52 (t, *J* = 6.8 Hz, 1H), 3.64 (s, 3H), 3.27−3.22 (m, 1H), 3.12−3.07 (m, 2H), 2.56 (dd, *J* = 16.8 and 6.8 Hz, 1H), 1.56−1.44 (m, 5H), 1.44 (s, 9H), 1.67–1.14 (m, 2H), 0.90 (d, *J* = 6.0 Hz, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃), *δ* (ppm) = 173.0, 171.2, 170.6, 157.6, 135.9, 128.4, 128.0, 127.8, 81.0, 66.8, 57.4, 52.2, 46.4, 37.7, 36.8, 34.8, 28.0, 26.0, 22.4, 17.8, 17.7; ESI-MS: *m*/*z* (%) calcd 490.3 [M]⁺. Found 491.6 [M + H]⁺ (100); anal. calcd for $C_{26}H_{38}N_2O_7$: C, 63.65; H, 7.81; N, 5.71. Found: C, 63.66; H, 7.81; N, 5.73

4-(tert-Butyl)-1-methyl N-Benzyl-N-((1-((benzyloxy)carbonyl) cyclohexyl)carbamoyl)-L-aspartate (4b). Yellow oil. Yield 83% (112 mg). R_f (hexane/AcOEt, 70:30) = 0.38; $[\alpha]_D^{20}$ -12.7 (*c* = 0.9, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.26–7.18 (m, 10H), 5.01 (s, 2H), 4.89 (t, *J* = 6.8 Hz, 1H), 4.80 (br s, 1H), 4.47 (d, *J* = 17.2 Hz, 1H), 4.33 (d, *J* = 17.2 Hz, 1H), 3.55 (s, 3H), 2.95 (dd, *J* = 16.4 and 6.8 Hz, 1H), 2.56 (dd, *J* = 16.4 and 7.2 Hz, 1H), 1.82− 1.80 (m, 2H), 1.64−1.62 (m, 2H), 1.40−1.33 (m, 4H), 1.33 (s, 9H), 1.04−1.02 (m, 2H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 174.6, 171.4, 170.4, 157.2, 137.3, 136.4, 128.9, 128.4, 128.0, 127.9, 127.7, 126.7, 81.2, 66.5, 58.9, 57.5, 52.2, 36.8, 32.8, 32.3, 28.0, 25.1, 21.2, 21.1; ESI-MS: m/z (%) calcd 552.3 [M]⁺. Found 553.8 [M + H]⁺ (100); anal. calcd for C₃₁H₄₀N₂O₇: C, 67.37; H, 7.30; N, 5.07. Found: C, 67.35; H, 7.31; N, 5.09.

4-(tert-Butyl)-1-methyl N-Benzyl-N-(((S)-1-(benzyloxy)-4-methyl-1-oxopentan-2-yl)carbamoyl)-L-aspartate (4c). Yellow oil. Yield 92% (104 mg). *R_f* (hexane/AcOEt, 70:30) = 0.35; $[\alpha]_D^{\ 20} - 15.8$ ($c =$ 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), *δ* (ppm) = 7.25−7.23 (m, 10H), 5.04 (d, *J* = 12.4 Hz, 1H), 5.01 (d, *J* = 12.4 Hz, 1H), 4.93−4.91 (m, 2H), 4.47 (d, *J* = 17.2 Hz, 1H), 4.39 (br s, 1H), 4.36 (d, *J* = 17.2 Hz, 1H), 3.59 (s, 3H), 2.97 (dd, *J* = 16.8 and 7.2 Hz, 1H), 2.62 (dd, *J* = 16.8 and 6.8 Hz, 1H), 1.35−1.33 (m, 1H), 1.32−1.30 (m, 2H), 1.32 (s, 9H), 0.75 (d, *J* = 6.4 Hz, 3H), 0.73 (d, *J* = 6.4 Hz, 3H); 13C{1 H} NMR (101 MHz, CDCl₃), δ (ppm) = 173.6, 171.7, 170.5, 158.1, 137.4, 136.0, 129.2, 128.8, 128.5, 128.4, 128.0, 127.0, 81.5, 67.0, 57.6, 52.8, 52.6, 51.4, 41.7, 37.0, 28.3, 25.0, 23.0, 22.2; ESI-MS: *m*/*z* (%) calcd 540.3 $[M]^{+}$. Found 563.4 $[M + Na]^{+}$ (100); anal. calcd for $C_{30}H_{40}N_2O_7$: C, 66.65; H, 7.46; N, 5.18. Found: C, 66.66; H, 7.46; N, 5.19.

General Procedure for the Synthesis of Hydantoin Intermediates 5. To a solution of the isocyanate 2 (2.5 mmol, 1 equiv) in DCM (0.3 M solution), a solution of TEA (2 equiv) and *N*-alkyl *α*-aminoester 3 (1.2 equiv) in DCM (0.3 M solution) was added at rt. The solution was left to stir for 18 h. In case the urea derivative (TLC monitoring) is left, a 1 N NaOH aqueous solution (10% in volume) was added and the mixture was vigorously stirred for 5 min. The reaction was quenched with a 1 N HCl aqueous solution and extracted with DCM. The collected organic phases were washed with a 1 N HCl aqueous solution (once), brine (once), a saturated aqueous solution of $NaHCO₃$ (twice), and brine (once). The organic phase was dried over $Na₂SO₄$, filtered, and the solvent evaporated. The hydantoin intermediates 4 were used without any further purification.

Benzyl 1-(4-(2-(tert-Butoxy)-2-oxoethyl)-3-isopentyl-2,5-dioxoimidazolidin-1-yl)cyclopropane-1-carboxylate (5a). Orange gum. Yield 77% (523 mg). R_f (hexane/AcOEt, 70:30) = 0.46; $[\alpha]_D^{-20}$ -31.2 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.31−7.26 (m, 5H), 5.11 (d, 1H, *J* = 12.2 Hz, 1H), 5.07 (d, *J* = 12.2 Hz, 1H), 4.18 (t, *J* = 4.8 Hz, 1H), 3.65−3.55 (m, 1H), 3.08−3.06 (m, 1H,), 2.75 (dd, *J* = 16.8 and 4.3 Hz, 1H), 2.61 (dd, *J* = 16.8 and 5.5 Hz, 1H), 1.78−1.74 (m, 2H), 1.44−1.42 (m, 5H), 1.43 (s, 9H), 0.91 $(d, J = 2.4 \text{ Hz}, 3\text{H})$, 0.89 $(d, J = 2.4 \text{ Hz}, 3\text{H})$; ¹³C{¹H} NMR (101) MHz, CDCl₃), δ (ppm) = 172.6, 170.4, 168.5, 155.6, 135.4, 128.5, 128.2, 127.8, 81.9, 67.4, 55.7, 39.7, 36.6, 32.5, 27.9, 25.8, 22.5, 22.2, 16.5, 16.2; ESI-MS: m/z (%) calcd 458.2 [M]⁺. Found 481.3 [M + Na]⁺ (100); anal. calcd for $C_{25}H_{34}N_2O_6$: C, 65.48; H, 7.47; N, 6.11. Found: C, 65.50; H, 7.48; N, 6.10.

Benzyl 1-(3-Benzyl-4-(2-(tert-butoxy)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)cyclopropane-1-carboxylate (5b). Dark orange gum. Yield 79% (712 mg). R_f (hexane/AcOEt, 70:30) = 0.32; $[\alpha]_D^2$ $^{\prime}$ –20.9 $(c = 1.0, \text{CHCl}_3)$; ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.35– 7.28 (m, 10H), 5.18 (d, *J* = 12.4 Hz, 1H), 5.13 (d, *J* = 12.4 Hz, 1H), 4.88 (d, *J* = 15.6 Hz, 1H), 4.25 (d, *J* = 15.6 Hz, 1H), 4.05 (t, *J* = 4.8 Hz, 1H), 2.65−2.63 (m, 2H), 1.84−1.81 (m, 2H), 1.58−1.56 (m, 2H), 1.39 (s, 9H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 172.3, 170.4, 168.3, 156.3, 136.7, 135.4, 128.9, 128.6, 128.2, 127.9, 127.8, 81.9, 67.5, 55.6, 45.1, 32.6, 27.9, 16.7, 16.3; ESI-MS: *m*/*z* (%) calcd 478.2 [M]⁺. Found 517.3 [M + K]⁺ (100), 501.3 [M + Na]⁺ (87); anal. calcd for $C_{27}H_{30}N_2O_6$: C, 67.77; H, 6.32; N, 5.85. Found: C, 67.78; H, 6.30; N, 5.83.

Benzyl (S)-1-(3-Benzyl-4-(2-(tert-butoxy)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)cyclohexane-1-carboxylate (5c). Orange gum. Yield 72% (578 mg). R_f (hexane/AcOEt, 70:30) = 0.36; $[\alpha]_D^2$ -21.3 ($c = 0.8$, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.25−7.14 (m, 10H), 5.11 (s, 2H), 4.76 (d, *J* = 15.4 Hz, 1H), 4.12 (d, 1H, *J* = 15.4 Hz, 1H), 3.91 (t, 1H, *J* = 5.2 Hz, 1H), 2.79−2.73 (m, 1H), 2.67−2.61 (m, 1H), 2.57 (dd, *J* = 16.6, 4.5 Hz, 1H), 2.50 (dd, *J* $= 16.6, 4.5$ Hz, 1H), 2.12–1.96 (m, 2H), 1.5 –1.40 (m, 6H), 1.31 (s, 9H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 172.5, 171.6, 168.4, 157.0, 135.9, 135.8, 128.9, 128.5, 128.2, 128.1, 127.9, 127.8, 81.8, 67.2, 65.5, 55.7, 45.2, 35.7, 31.7, 31.5, 28.0, 24.9, 22.6, 22.4; ESI-MS: *m/z* (%) calcd 520.3 [M]⁺. Found 521.4 [M + H]⁺; anal. calcd for $C_{30}H_{36}N_{2}O_{6}$: C, 69.21; H, 6.97; N, 5.38. Found: C, 69.21; H, 6.98; N, 5.40.

Benzyl (S)-2-(3-Ethyl-4-(2-methoxy-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)-2-methylpropanoate (5d). Yellow gum. Yield 81% (641 mg). R_f (hexane/AcOEt, 70:30) = 0.28; $[\alpha]_D^{20}$ +11.4 ($c = 1.1$, CHCl₃); ¹H NMR (400 MHz, DMSO- d_6), δ (ppm) = 7.36–7.33 (m 5H), 5.12 (d, *J* = 12.8 Hz, 1H), 5.07 (d, *J* = 12.8 Hz, 1H), 4.35 (t, *J* = 4.4 Hz, 1H), 3.56 (s, 3H), 3.44−3.42 (m, 1H), 3.08−3.06 (m, 1H), 2.95 (dd, *J* = 17.2 and 4.4 Hz, 1H), 2.88 (dd, *J* = 17.2 and 4.4 Hz, 1H), 1.66 (s, 3H), 1.62 (s, 3H), 1.02 (t, *J* = 7.2 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 172.4, 171.7, 169.6, 155.7, 135.7, 128.5, 128.24, 128.22, 67.3, 60.7, 55.4, 52.2, 36.0, 34.2, 24.0, 23.8, 13.1; ESI-MS: m/z (%) calcd 376.2 [M]⁺. Found 399.3 [M + Na]⁺ (100); anal. calcd for C₁₉H₂₄N₂O₆: C, 60.63; H, 6.43; N, 7.44. Found: C, 60.61; H, 6.43; N, 7.45.

Benzyl (S)-2-(4-(2-Methoxy-2-oxoethyl)-3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-1-yl)-2-methylpropanoate (5e). Orange gum. Yield 82% (589 mg). R_f (hexane/AcOEt, 70:30) = 0.36; $[\alpha]_{\text{D}}^2$ ²⁰ −18.2 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.96−7.93 (m, 1H), 7.77−7.72 (m, 2H), 7.41−7.36 (m 2H), 7.25−7.22 (m, 7H), 5.15 (d, *J* = 12.4 Hz, 1H), 5.09 (d, *J* = 12.4 Hz, 1H), 5.07 (d, *J* = 15.6 Hz, 1H), 4.68 (d, *J* = 15.6 Hz, 1H), 3.66 (dd, *J* = 4.8 and 3.6 Hz, 1H), 3.25 (s, 3H), 2.63 (dd, *J* = 16.8 and 3.6 Hz, 1H), 2.47 (dd, *J* = 16.8 and 4.8 Hz, 1H), 1.73 (s, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 172.5, 171.7, 168.9, 156.3, 135.8, 134.9, 131.3, 130.9, 129.4, 128.8, 128.5, 128.2, 128.1, 127.2, 127.0, 126.3, 125.1, 123.4, 67.3, 60.9, 55.8, 51.9, 43.8, 33.5, 24.0, 23.8; ESI-MS: *m/z* (%) calcd 488.2 [M]⁺. Found 511.2 [M + Na]⁺ (100); anal. calcd for $C_{28}H_{28}N_2O_6$: C, 68.84; H, 5.78; N, 5.73. Found: C, 68.84; H, 5.79; N, 5.72.

Benzyl (S)-2-(3-Benzyl-4-(2-(tert-butoxy)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)acetate (5f). Yellowish oil. Yield 74% (612 mg). *Rf* $(\text{hexane/ACOEt}, 70:30) = 0.32; [\alpha]_D^{20} - 10.7 (\text{c} = 1.0, \text{CHCl}_3);$ ¹H NMR (400 MHz, CDCl3), *δ* (ppm) = 7.31−7.28 (m, 10H), 5.18 (d, *J* = 12.4 Hz, 1H), 5.14 (d, *J* = 12.4 Hz, 1H), 4.86 (d, *J* = 15.6 Hz, 1H), 4.32 (s, 2H), 4.22 (d, *J* = 15.6 Hz, 1H), 4.16 (t, *J* = 4.8 Hz, 1H), 4.68−4.65 (m, 2H), 1.34 (s, 9H); ¹³C{¹H} NMR (101 MHz, CDCl₃), *δ* (ppm) = 173.2, 169.6, 168.5, 157.5, 137.1, 136.5, 130.5, 130.1, 130.09, 130.07, 129.9, 129.6, 129.4, 83.6, 69.2, 57.8, 46.8, 41.5, 37.0, 29.5; ESI-MS: m/z (%) calcd 452.2 [M]⁺. Found 475.3 [M + Na]⁺ (100); anal. calcd for $\rm C_{25}H_{28}N_2O_6$: C, 66.36; H, 6.24; N, 6.19. Found: C, 66.37; H, 6.23; N, 6.20.

Benzyl (S)-2-(4-(2-(tert-Butoxy)-2-oxoethyl)-3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-1-yl)acetate (5g). Yellowish oil. Yield 88% (582 mg). *R_f* (hexane/AcOEt, 70:30) = 0.28; $[\alpha]_D^2$ +8.2 ($c = 1.0$, CHCl₃); ^fH NMR (400 MHz, CDCl₃), δ (ppm) = 7.99−7.96 (m, 1H), 7.80−7.74 (m, 2H), 7.44−7.42 (m, 2H), 7.31− 7.23 (m, 7H), 5.32 (d, *J* = 15.6 Hz, 1H), 5.12 (s, 2H), 4.65 (d, *J* = 15.6 Hz, 1H), 4.30 (s, 2H), 3.89 (dd, *J* = 5.2 and 4.0 Hz, 1H), 2.68 (dd, *J* = 16.8 and 4.0 Hz, 1H), 2.56 (dd, *J* = 16.8 and 5.2 Hz, 1H), 1.20 (s, 9H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 171.6, 167.7, 166.9, 155.8, 135.0, 134.0, 131.2, 130.7, 129.2, 128.8, 128.5, 128.3, 127.1, 126.6, 126.3, 125.2, 123.2, 82.0, 67.6, 56.5, 43.7, 40.0, 35.2, 27.8; ESI-MS: *m*/*z* (%) calcd 502.2 [M]⁺ . Found 525.3 [M + Na]⁺ (100); anal. calcd for C₂₉H₃₀N₂O₆: C, 69.31; H, 6.02; N, 5.57. Found: C, 69.33; H, 6.02; N, 5.58.

Benzyl 2-((S)-3-Benzyl-4-(2-(tert-butoxy)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)-4-methylpentanoate (5h). Yellow gum. Yield 89% (217 mg). R_f (hexane/AcOEt, 70:30) = 0.32; ¹H NMR (400 MHz, CDCl3), *δ* (ppm), mixture of diastereoisomers = 7.34−7.27 (m, 20H), 5.21−5.17 (m, 4H), 4.90−4.83 (m, 4H), 4.29−4.26 (m, 2H), 4.15−4.09 (m, 2H), 2.68−2.63 (m, 4H), 2.34−2.30 (m, 2H), 1.96− 1.93 (m, 2H), 1.66−1.64 (m, 2H), 1.41 (s, 9H), 1.40 (s, 9H), 0.98− 0.95 (m, 12H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm), mixture of diastereoisomers = 172.2, 172.1, 169.76, 169.73, 168.6, 156.7, 156.4, 136.1, 135.67, 135.62, 129.2, 129.1, 128.88, 128.85, 128.6, 128.58, 128.51, 128.3, 128.2, 128.17, 128.12, 128.0, 127.9, 127.3, 82.2, 67.8, 67.7, 56.2, 56.1, 52.4, 52.0, 45.6, 45.5, 37.2, 37.0, 36.0, 35.7, 30.0, 28.2, 25.2, 23.4, 23.3, 21.4, 21.3; ESI-MS: *m*/*z* (%) calcd 508.3 [M]⁺. Found 531.3 [M + Na]⁺ (100); anal. calcd for $C_{29}H_{36}N_2O_6$: C, 68.48; H, 7.13; N, 5.51. Found: C, 68.46; H, 7.14; N, 5.52.

General Procedure for the Synthesis of Hydantoin Monoamides 7. To a solution of hydantoin derivative 5 (1.2 mmol, 1 equiv) in a 1:1 mixture AcOEt/MeOH (0.1 M solution), a catalytic amount of Pd/C was added at rt. The mixture was vigorously stirred under hydrogen atmosphere. When the starting material is totally consumed (around 3 h, TLC monitoring), the mixture is filtered on a Celite pad, washed with MeOH, and the solvent evaporated. The crude was dissolved in DMF (0.1 M solution), and HBTU (1.1 equiv) followed by TEA (1.1 equiv) and the amine (1.1 equiv) were added at rt. The reaction was left to stir for 12 h. The solution was diluted with AcOEt and washed with brine (three times), a 1 N HCl aqueous solution (twice), brine (once), a saturated $NAHCO₃$ aqueous solution (twice), and brine (twice). The organic phase was dried over $Na₂SO₄$, filtered, and the solvent evaporated. Hydantoin monoamides 7 were used without any further purification.

Benzyl 1-(3-Benzyl-4-(2-((2,2-diphenylethyl)amino)-2-oxoethyl)- 2,5-dioxoimidazolidin-1-yl)cyclopropane-1-carboxylate (7a). Yellow gum. Yield 75% (230 mg). *Rf* (hexane/AcOEt, 50:50) = 0.36; $[\alpha]_D^{20}$ –24.2 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.32−7.19 (m, 20H), 5.17 (br s, 1H), 5.14 (d, *J* = 10.0 Hz, 1H), 5.06 (d, *J* = 10.0 Hz, 1H), 4.62 (d, *J* = 12.0 Hz, 1H), 4.26 (d, *J* = 12.0 Hz, 1H), 4.16−4.14 (m, 1H), 4.06 (t, *J* = 6.4 Hz, 1H), 3.86−3.84 (m, 1H), 3.69−3.67 (m, 1H), 3.45 (dd, *J* = 12.8 and 3.2 Hz, 1H), 2.14−2.11 (m, 1H), 1.83−1.81 (m, 4H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 173.0, 170.5, 167.8, 156.3, 141.7, 141.6, 136.4, 135.5, 128.94, 128.92, 128.8, 128.7, 128.6, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.1, 67.7, 56.0, 50.5, 45.5, 44.0, 36.8, 32.8, 16.8, 16.3; ESI-MS: *m/z* (%) calcd 601.3 [M]⁺. Found 624.9 [M + Na]⁺ (100), 602.0 $[M + H]^+$ (12); anal. calcd for $C_{37}H_{35}N_3O_5$: C, 73.86; H, 5.86; N, 6.98. Found: C, 73.86; H, 5.87; N, 7.00

tert-Butyl 2-(3-Benzyl-1-(1-((2,2-diphenylethyl)carbamoyl) cyclopropyl)-2,5-dioxoimidazolidin-4-yl)acetate (7b). Amorphous white solid. Yield 80% (421 mg). *Rf* (hexane/AcOEt, 50:50) = $0.31; [\alpha]_D^{20} - 15.4$ (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), *δ* (ppm) = 7.19−7.12 (m, 16H), 4.53 (d, *J* = 15.2 Hz, 1H), 4.24 (d, *J* = 15.2 Hz, 1H), 4.24 (s, 1H), 3.92−3.90 (m, 1H), 3.65−3.61 (m, 2H), 2.80 (dd, *J* = 18.0 and 3.2 Hz, 1H), 2.52 (dd, *J* = 4.4 Hz, 1H), 1.54− 1.51 (m, 4H), 1.29 (s, 9H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* $(ppm) = 172.0, 169.4, 166.3, 156.0, 142.9, 142.6, 136.1, 128.9, 128.5,$ 128.3, 128.25, 128.23, 128.05, 128.02, 126.2, 126.1, 77.2, 56.1, 49.9, 48.9, 45.9, 45.2, 32.8, 25.4, 16.0, 15.5; ESI-MS: *m*/*z* (%) calcd 567.3 $[M]^{+}$. Found 590.5 $[M + Na]^{+}$ (100); anal. calcd for $C_{34}H_{37}N_{3}O_{5}$: C, 71.94; H, 6.57; N, 7.40. Found: C, 71.95; H, 6.58; N, 7.39.

tert-Butyl 2-(1-(1-((4-Fluorobenzyl)carbamoyl)cyclopropyl)-3 isopentyl-2,5-dioxoimidazolidin-4-yl)acetate (7c). Amorphous white solid. Yield 86% (256 mg). *Rf* (hexane/AcOEt, 50:50) = $0.15; [\alpha]_D^{20}$ +9.4 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), *δ* (ppm) = 7.94 (br s, 1H), 7.13−7.11 (m, 2H), 6.89−6.85 (m, 2H), 4.45 (dd, *J* = 15.6 and 6.4 Hz, 1H), 4.27 (dd, *J* = 15.6 and 5.6 Hz, 1H), 3.84 (dd, *J* = 4.4 and 2.4 Hz, 1H), 3.46−3.42 (m, 1H), 3.09− 3.05 (m, 1H), 3.02 (dd, *J* = 18.0 and 2.4 Hz, 1H), 2.73 (dd, 18.0 and 4.4 Hz, 1H), 1.83−1.81 (m, 1H), 1.65−1.63 (m, 1H), 1.48 (septet, *J* = 6.4 Hz, 1H), 1.38−1.14 (m, 4H), 1.13 (s, 9H), 0.86 (d, *J* = 6.4 Hz, 3H), 0.84 (d, *J* = 6.4 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 172.1, 169.7, 169.2, 161.8 (d, *J* = 244.2 Hz), 155.5, 134.5 (d, *J* = 3.0 Hz), 128.6 (d, *J* = 8.1 Hz), 115.1 (d, *J* = 22.2 Hz), 83.2, 55.6, 42.8, 39.6, 37.0, 33.6, 33.5, 27.7, 25.9, 22.4, 22.3, 16.3, 15.9; ESI-MS: *m/z* (%) calcd 475.2 [M]⁺. Found 498.3 [M + Na]⁺ (100); anal. calcd for $C_{25}H_{34}FN_{3}O_{5}$: C, 63.14; H, 7.21; N, 8.84. Found: C, 63.16; H, 7.21; N, 8.85.

tert-Butyl 2-(3-Benzyl-1-(1-((2,2-diphenylethyl)carbamoyl) cyclohexyl)-2,5-dioxoimidazolidin-4-yl)acetate (7d). Yellow gum. Yield 81% (518 mg). R_f (hexane/AcOEt, 50:50) = 0.41; $[\alpha]_D^2$ $+13.8$ $(c = 1.0, \text{CHCl}_3)$; ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.35– 7.24 (m, 15H), 7.00 (t, *J* = 5.6 Hz, 1H), 4.75 (d, *J* = 15.2 Hz, 1H), 4.38 (t, *J* = 8.0 Hz, 1H), 4.20 (d, *J* = 15.2 Hz, 1H), 4.10−4.07(m, 1H), 8.81–3.73 (m, 2H), 2.85 (dd, *J* = 17.6 and 3.6 Hz, 1H), 2.69–
2.62 (m 1H), 2.09–2.07 (m, 1H), 1.78–1.28 (m, 9H), 1.45 (s, 9H); 2.62 (m 1H), 2.09−2.07 (m, 1H), 1.78−1.28 (m, 9H), 1.45 (s, 9H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 172.5, 172.3, 169.2, 156.7, 142.3, 142.2, 135.8, 129.1, 128.5, 128.4, 128.2, 128.12, 128.10, 127.9, 126.5, 126.4, 82.5, 70.0, 55.3, 50.4, 45.3, 44.3, 34.0, 32.1, 28.1, 25.1, 22.8, 22.2; ESI-MS: m/z (%) calcd 609.3 [M]⁺. Found 632.4 [M + Na]⁺ (100); anal. calcd for C₃₇H₄₃N₃O₅: C, 72.88; H, 7.11; N, 6.89. Found: C, 72.89; H, 7.13; N, 6.90.

Methyl (S)-2-(3-Ethyl-1-(1-(hexylamino)-2-methyl-1-oxopropan-2-yl)-2,5-dioxoimidazolidin-4-yl)acetate (7e). Amorphous white solid. Yield 79% (267 mg). *Rf* (hexane/AcOEt, 50:50) = 0.35; $[\alpha]_D^{20}$ +22.3 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ $(ppm) = 6.70$ (br s, 1H), 3.99 (dd, $J = 4.4$ and 3.2 Hz, 1H), 3.70 (s, 3H), 3.56 (sextet, *J* = 7.2 Hz, 1H), 3.24−3.19 (m, 3H), 3.08 (dd, *J* = 17.6 and 3,2 Hz, 1H), 2.91 (dd, *J* = 17.6 and 4.4 Hz, 1H), 1.80 (s, 3H), 1.73 (s, 3H), 1.49−1.47 (m, 2H), 1.27−1.23 (m, 6H), 1.35 (t, *J* $= 7.2$ Hz, 3H), 0.86 (t, *J* = 6.8 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CDCl3), *δ* (ppm) = 172.9, 171.9, 170.2, 155.8, 62.3, 55.2, 52.3, 39.9, 35.9, 32.9, 31.5, 29.1, 26.5, 25.2, 24.6, 22.5, 14.0, 13.2; ESI-MS: *m*/*z* (%) calcd 369.2 [M]+ . Found 392.4 [M + Na]⁺ (100); anal. calcd for C18H31N3O5: C, 58.52; H, 8.46; N, 11.37. Found: C, 58.51; H, 8.48; N, 11.38.

Methyl (S)-2-(1-(1-(Isobutylamino)-2-methyl-1-oxopropan-2-yl)- 3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-4-yl)acetate (7f). Yellow gum. Yield 91% (208 mg). *Rf* (hexane/AcOEt, 50:50) = 0.37; $[\alpha]_D^{20}$ –10.0 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 8.02−8.00 (m, 1H), 7.80−7.78 (m, 2H), 7.45−7.43 (m, 2H), 7.34−7.32 (m, 2H), 6.92 (br t, *J* = 4.8 Hz, 1H), 4.97 (d, *J* = 14.8 Hz, 1H), 4.85 (d, *J* = 14.8 Hz, 1H), 3.61 (dd, *J* = 4.4 and 2.8 Hz, 1H), 3.14 (s, 3H), 3.10−3.08 (m, 1H), 3.02−3.00 (m, 1H), 2.71 (dd, *J* = 18.0 Hz and 2.8 Hz, 1H), 2.38 (dd, *J* = 18.0 and 4.4 Hz, 1H), 1.76 (s, 3H), 1.75−1.73 (m, 1H), 1.71 (s, 3H), 0.85 (d, *J* = 4.0 Hz, 3H), 0.83 $(d, J = 4.0 \text{ Hz. } 3\text{H})$; ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 173.5, 172.2, 170.4, 156.7, 134.4, 131.7, 131.6, 130.0, 129.1, 128.1, 127.2, 126.6, 125.4, 124.0, 62.9, 56.4, 52.2, 47.8, 45.1, 33.3, 28.7, 25.8, 24.9, 20.6; ESI-MS: m/z (%) calcd 453.2 [M]⁺. Found 476.2 [M + Na]⁺ (100); anal. calcd for C₂₅H₃₁N₃O₅: C, 66.21; H, 6.89; N, 9.27. Found: C, 66.19; H, 6.88; N, 9.28.

Methyl (S)-2-(3-Benzyl-1-(2-((2,2-diphenylethyl)amino)-2-oxoethyl)-2,5-dioxoimidazolidin-4-yl)acetate (7g). Orange gum. Yield 82% (174 mg). *R_f* (hexane/AcOEt, 50:50) = 0.43; $[\alpha]_D^2$ -6.0 (*c* = 1.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.17−7.02 (m, 15H), 7.00 (br s, 1H), 4.49 (d, *J* = 15.2 Hz, 1H), 4.31 (d, *J* = 15.2 Hz, 1H), 4.17 (t, *J* = 8.0 Hz, 1H), 4.12 (s, 2H), 3.86−3.77 (m, 3H), 3.39 (s, 3H), 2.75 (dd, *J* = 17.6 and 3.6 Hz, 1H), 2.58 (dd, *J* $= 17.6$ and 4.4 Hz, 1H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 171.2, 169.9, 166.8, 155.7, 142.1, 142.0, 135.5, 129.0, 128.5, 128.4, 128.3, 128.23, 128.20, 128.1, 126.6, 56.1, 52.3, 50.1, 45.6, 44.6, 41.8, 32.8, 29.7; ESI-MS: m/z (%) calcd 499.2 [M]⁺. Found 522.2 [M + Na]⁺ (100); anal. calcd for C₂₉H₂₉N₃O₅: C, 69.72; H, 5.85; N, 8.41. Found: C, 69.74; H, 5.85; N, 8.40.

tert-Butyl (S)-2-(1-(2-(Isobutylamino)-2-oxoethyl)-3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-4-yl)acetate (7h). Yellow gum. Yield 93% (278 mg). *Rf* (hexane/AcOEt, 50:50) = 0.24; $[\alpha]_D^{20}$ –23.1 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 8.06−8.04 (m, 1H), 7.83−7.80 (m, 2H), 7.49−7.35 (m, 4H), 7.23 (br t, *J* = 5.6 Hz, 1H), 5.18 (d, *J* = 14.8 Hz, 1H), 4.75 (d, *J* = 14.8 Hz, 1H), 4.23 (d, *J* = 16.8 Hz, 1H), 4.18 (d, *J* = 16.8 Hz, 1H), 3.67 (dd, *J* = 4.4 and 2.8 Hz, 1H), 3.10−3.07 (m, 1H), 2.93−2.90 (m, 1H), 2.83 (dd, *J* = 17.6 and 2.8 Hz, 1H), 2.60 (dd, *J* = 17.6 and 4.4 Hz, 1H), 1.77 (septet, *J* = 6.8 Hz, 1H), 1.19 (s, 9H), 0.82 (d, *J* = 6.8 Hz, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 171.5, 169.1, 166.4, 155.8, 134.1, 131.3, 130.7, 129.7, 128.9, 127.6, 127.3, 126.5, 125.1, 123.3, 82.9, 56.4, 47.2, 44.4, 42.1, 33.9, 28.4, 27.7, 20.2, 20.1; ESI-MS: m/z (%) calcd 467.2 [M]⁺. Found 490.1 [M + Na]⁺ (100); anal. calcd for $C_{26}H_{33}N_3O_5$: C, 66.79; H, 7.11; N, 8.99. Found: C, 66.80; H, 7.11; N, 9.00.

tert-Butyl (S)-2-(1-(2-((4-Chlorobenzyl)amino)-2-oxoethyl)-3- (naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-4-yl)acetate (7i). Amorphous white solid. Yield 90% (185 mg). *Rf* (hexane/AcOEt, $50:50$ = 0.28; $[\alpha]_D^{20}$ -12.9 ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CDCl3), *δ* (ppm) = 8.11−8.09 (m, 1H), 7.90−7.88 (m, 2H), 7.78 (br t, *J* = 6.0 Hz, 1H), 7.57−7.54 (m, 2H), 7.46−7.43 (m, 2H), 7.27 (d, *J* = 8.4 Hz, 2H), 7.20 (d, *J* = 8.4 Hz, 2H), 5.29 (d, *J* = 14.8 Hz, 1H),

4.78 (d, *J* = 14.8 Hz, 1H), 4.56 (dd, *J* = 15.6 and 6.4 Hz, 1H), 4.38 (dd, *J* = 15.6 and 5.6 Hz, 1H), 4.37−4.36 (m, 2H), 3.74 (dd, *J* = 4.4 and 2.4 Hz, 1H), 2.89 (dd, *J* = 18.0 and 2.4 Hz, 1H), 2.67 (dd, *J* = 18.0 and 4.4 Hz, 1H), 1.10 (s, 9H); ¹³C{¹H} NMR (101 MHz, CDCl3), *δ* (ppm) = 171.5, 169.1, 166.6, 136.7, 134.1, 132.8, 131.3, 130.6, 129.7, 128.9, 128.65, 128.61, 127.6, 127.3, 126.5, 125.1, 123.2, 83.1, 56.3, 44.2, 42.5, 42.1, 33.9, 27.6; ESI-MS: *m*/*z* (%) calcd 535.2 [M]⁺. Found 558.4 [M + Na]⁺ (68); anal. calcd for $C_{29}H_{30}CIN_3O_5$: C, 64.98; H, 5.64; N, 7.84. Found: C, 64.96; H, 5.65; N, 7.85.

General Procedure for the Hydrolysis of the tert-Butyl Ester of Hydantoin Monoamides 7. To a solution of hydantoin monoamide 7 (1.0 mmol, 1 equiv) in DCM (0.1 M solution), TFA (30% in volume) was added at rt. The mixture was stirred until the starting material is totally consumed (around 12 h, TLC monitoring). The solvents were evaporated and co-evaporated twice with cyclohexane. The obtained acids were used in the coupling reaction without any further purification.

General Procedure for the Hydrolysis of the Methyl Ester of Hydantoin Monoamides 7. To a solution of hydantoin monoamide 7 (1.0 mmol, 1 equiv) in MeOH (0.1 M solution), a 1 M NaOH aqueous solution (10% in volume) was added at rt. The reaction was stirred until complete disappearance of the starting material (around 3 h, TLC monitoring). The organic solvent was evaporated, the mixture was diluted with AcOEt, and a 1 M HCl aqueous solution was added until acidic pH was reached. The phases were separated, and the aqueous phase was washed three times with AcOEt. The collected organic phases were dried over Na₂SO₄, filtered, and the solvent evaporated. The obtained acids were used in the coupling reaction without any further purification.

General Procedure for the Synthesis of Hydantoin Universal Peptidomimetics 8. The acid obtained was dissolved in DMF (1.0 mmol, 0.1 M solution), and HBTU (1.1 equiv) followed by TEA (1.1 equiv) and the amine (1.1 equiv) were added at rt. The reaction was left to stir for 12 h. The solution was diluted with AcOEt and washed with brine (three times), a 1 N HCl aqueous solution (twice), brine (once), a saturated $NAHCO₃$ aqueous solution (twice), and brine (twice). The organic phase was dried over $Na₂SO₄$, filtered, and the solvent evaporated. Hydantoin universal peptidomimetics 8 were purified by flash chromatography (from AcOEt/hexane = 80:20 to AcOEt 100%).

(S)-1-(3-Benzyl-4-(2-((2,2-diphenylethyl)amino)-2-oxoethyl)-2,5 dioxoimidazolidin-1-yl)-N-hexylcyclopropane-1-carboxamide (8a). Yellowish gum. Yield 82% (134 mg). *Rf* (AcOEt/hexane, 80/20) = $0.29; [α]_D$ ²⁰ −31.2 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 8.03 (br s, 1H), 7.24−7.05 (m, 15H), 5.34 (br s, 1H), 4.57 (d, *J* = 14.8 Hz, 1H), 4.04−4.02 (m, 2H), 3.69−3.63 (m, 3H), 3.24 (m, 1H), 3.14 (m, 1H), 2.82 (br s, 1H), 2.30 (br s, 1H), 1.85−1.82 (m, 1H), 1.63−1.60 (m, 2H), 1.50−1.46 (m, 2H), 1.16−1.10 (m, 7H), 0.73 (t, *J* = 6.8 Hz, 3H); ¹³C{¹H} NMR (400 MHz, CDCl₃), δ $(ppm) = 172.1, 169.3, 167.6, 156.1, 141.3, 135.8, 129.0, 128.9, 128.2,$ 128.1, 127.9, 127.8, 127.15, 127.11, 55.5, 50.3, 45.4, 43.9, 40.6, 34.1, 33.7, 31.5, 29.1, 26.6, 22.6, 16.1, 15.7, 14.0; ESI-MS: *m*/*z* (%) calcd 594.3 $[M]^{+}$. Found 617.4 $[M + Na]^{+}$ (100), 595.4 $[M + H]^{+}$ (5); anal. calcd for C₃₆H₄₂N₄O₄: C, 72.70; H, 7.12; N, 9.42. Found: C, 72.69; H, 7.12; N, 9.40

(S)-1-(4-(2-((2-(1H-Indol-3-yl)ethyl)amino)-2-oxoethyl)-3-benzyl-2,5-dioxoimidazolidin-1-yl)-N-(2,2-diphenylethyl)cyclopropane-1 carboxamide (8b). Amorphous white solid. Yield 82% (101 mg). *Rf* $(\text{ACOEt}) = 0.35; [\alpha]_{\text{D}}^{20} - 10.1 (\text{c} = 1.0, \text{CHCl}_3);$ ¹H NMR (400 MHz, DMSO- d_6), δ (ppm) = 10.88 (s, 1H), 8.52 (brs, 1H), 8.25 (s, 1H), 7.56 (d, *J* = 7.8 Hz, 1H), 7.45−7.05 (m, 18H), 7.00 (t, *J* = 7.2 Hz, 1H), 4.57 (d, *J* = 15.9 Hz, 1H), 4.33 (t, *J* = 8.05 Hz, 1H), 4.30 (d, *J* = 15.9 Hz, 1H), 4.02 (t, *J* = 3.4 Hz, 1H), 3.78− 3.75 (m, 1H), 3.63− 3.61 (m, 1H), 3.35−3.27 (m, 3H), 2.82−2.80 (m, 2H), 2.73−2.71 (m, 1H), 1.47−1.45 (m, 1H), 1.37−1.35 (m, 1H), 1.16−1.13 (m, 2H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 171.4, 168.8, 165.7, 155.4, 142.2, 142.0, 135.5, 128.3, 127.9, 127.7, 127.64, 127.62, 127.43, 127.41, 125.6, 125.5, 76.5, 55.4, 49.3, 48.2, 45.3, 44.6, 32.1, 24.8, 15.4, 14.8; ESI-MS: *m/z* (%) calcd 653.3 [M]⁺. Found 676.5 [M

 $+$ Na]⁺ (100); anal. calcd for C₄₀H₃₉N₅O₄: C, 73.49; H, 6.01; N, 10.71. Found: C, 73.50; H, 6.00; N, 10.72.

(S)-1-(3-Benzyl-4-(2-((4-hydroxyphenethyl)amino)-2-oxoethyl)- 2,5-dioxoimidazolidin-1-yl)-N-(2,2-diphenylethyl)cyclopropane-1 carboxamide (8c). Amorphous white solid. Yield 85% (84 mg). *Rf* $(ACOEt) = 0.27; [\alpha]_D^{20} + 29.5 (\textit{c} = 1.1, CHCl_3);$ ¹H NMR (400 MHz, DMSO-*d*6), *δ* (ppm) = 9.17 (s, 1H), 8.47 (s, 1H), 8.16 (s, 1H), 7.24− 7.14 (m, 15H,), 6.97 (d, *J* = 8.4 Hz, 2H), 6.66 (d, *J* = 8.4 Hz, 2H), 4.51 (d, *J* = 15.9 Hz, 1H), 4.31 (t, *J* = 7.0 Hz, 1H), 4.24 (d, *J* = 15.9 Hz, 1H), 3.96 (s, 1H), 3.80−3.78 (m, 1H), 3.65−3.63 (m, 1H), 3.26−3.24 (m, 2H), 3.17−3.13 (m, 2H), 2.70 (br s, 2H), 1.48−1.46 (m, 1H), 1.31−1.29 (m, 1H), 1.15−1.13 (m, 2H); 13C{1 H} NMR $(101 \text{ MHz}, \text{ DMSO-}d_6)$, δ (ppm) = 173.0, 169.8, 168.7, 156.2, 143.3, 143.2, 137.2, 129.8, 129.6, 128.9, 128.7, 128.4, 128.0, 127.8, 126.6, 115.7, 56.3, 49.8, 44.9, 44.8, 41.2, 34.4, 33.8, 33.4, 15.8, 15.2; ESI-MS: *m/z* (%) calcd 630.3 [M]⁺. Found 631.5 [M + H]⁺ (100); anal. calcd for $C_{38}H_{38}N_4O_5$: C, 72.36; H, 6.07; N, 8.88. Found: C, 72.36; H, 6.08; N, 8.90.

(S)-1-(3-Benzyl-4-(2-(cyclohexylamino)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)-N-(2,2-diphenylethyl)cyclopropane-1-carboxamide (8d). Amorphous white solid. Yield 92% (163 mg). *Rf* (AcOEt/ hexane, 80:20) = 0.22; $[\alpha]_D^{20} - 37.5$ ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 8.12 (br s, 1H), 7.26–7.20 (m, 15H), 5.03 (d, *J* = 7.6 Hz, 1H), 4.48 (d, *J* = 15.6 Hz, 1H), 4.42 (d, *J* = 15.6 Hz, 1H), 4.37 (t, *J* = 7.6 Hz, 1H), 3.98−3.94 (m, 1H), 3.77−3.70 (m, 2H), 3.54−3.48 (m, 1H), 2.85 (dd, *J* = 16.8 and 2.4 Hz, 1H), 2.33 (dd, *J* = 16.8 and 4.4 Hz, 1H), 1.87−1.56 (m, 10H), 1.19−1.02 (m, 4H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 172.0, 169.5, 166.4, 156.0, 142.9, 142.6, 136.1, 129.0, 128.5, 128.3, 128.26, 128.24, 128.0, 126.2, 126.1, 56.1, 49.9, 48.9, 45.9, 45.2, 34.1, 33.7, 32.8, 25.4, 24.8, 24.7, 16.0, 15.5; ESI-MS: m/z (%) calcd 592.3 [M]⁺. Found 615.5 $[M + Na]^+$ (100); anal. calcd for $C_{36}H_{40}N_4O_4$: C, 72.95; H, 6.80; N, 9.45. Found: C, 72.97; H, 6.81; N, 9.44.

tert-Butyl (S)-(6-(2-(3-Benzyl-1-(1-((2,2-diphenylethyl) carbamoyl)cyclopropyl)-2,5-dioxoimidazolidin-4-yl)acetamido) hexyl)carbamate (8e). Amorphous white solid. Yield 78% (78 mg). R_f (AcOEt/hexane, 90:10) = 0.27; $[\alpha]_D^{20}$ –7.9 (*c* = 1.0, MeOH); ¹H NMR (400 MHz, CD₃OD), δ (ppm) = 8.67 (br s, 0.5 H), 7.16–7.10 (m, 15H), 4.40 (d, *J* = 15.6 Hz, 1H), 4.35 (d, *J* = 15.6 Hz, 1H), 4.30 (t, *J* = 7.6 Hz, 1H), 3.85−3.80 (m, 2H), 3.64−3.62 (m, 1H), 3.00− 2.80 (m, 4H), 2.74 (dd, *J* = 17.2 and 2.8 Hz, 1H), 2.55 (dd, *J* = 17.2 and 4.8 Hz, 1H), 1.57−1.55 (m, 1H), 1.38−1.30 (m, 3H), 1.32 (s, 9H), 1.19–1.15 (m, 8H). ¹³C{¹H} NMR (101 MHz, CD₃OD), δ (ppm) = 173.3, 170.9, 170.8, 168.3, 156.6, 142.7, 142.5, 136.5, 128.4, 128.0, 127.99, 127.92, 127.7, 127.5, 126.0, 78.4, 60.1, 56.6, 49.7, 45.1, 44.9, 39.8, 39.2, 33.2, 29.6, 28.9, 27.4, 26.4, 26.2, 15.4, 14.8; ESI-MS: *m/z* (%) calcd 709.4 [M]⁺. Found 710.5 [M + H]⁺ (100); anal. calcd for $C_{41}H_{51}N_5O_6$: C, 69.37; H, 7.24; N, 9.87. Found: C, 69.38; H, 7.22; N, 9.90.

(S)-N-(4-Fluorobenzyl)-1-(4-(2-(hexylamino)-2-oxoethyl)-3-isopentyl-2,5-dioxoimidazolidin-1-yl)cyclopropane-1-carboxamide (8f). Amorphous white solid. Yield 85% (152 mg). *Rf* (AcOEt/hexane, $80:20$ = 0.22; $\left[\alpha\right]_{D}^{20}$ –17.3 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CD3OD), *δ* (ppm) = 7.23−7.18 (m, 2H), 6.93−6.88 (m, 2H), 4.40 (d, *J* = 15.2 Hz, 1H), 4.25 (d, *J* = 15.2 Hz, 1H), 4.06 (dd, *J* = 4.8 and 2.8 Hz, 1H), 3.46−3.44 (m, 1H), 3.11−3.08 (m, 1H), 2.90−2.87 (m, 2H), 2.83−2.78 (m, 2H), 1.69−1.66 (m, 1H), 1.50−1.11 (m, 14H), 0.88−0.86 (m, 6H), 0.81 (t, *J* = 7.2 Hz, 3H); 13C{1 H} NMR (101 MHz, CD3OD), *δ* (ppm) = 173.7, 171.0, 168.5, 161.9 (d, *J* = 244.4 Hz), 156.3, 134.6 (d, *J* = 4.0 Hz), 128.6 (d, *J* = 8.1 Hz), 114.5 (d, *J* = 21.2 Hz), 56.2, 42.3, 39.3, 38.5, 36.6, 33.3, 33.1, 31.4, 28.9, 26.3, 25.6, 22.3, 22.2, 21.4, 15.4, 15.0, 12.9; ESI-MS: m/z (%) calcd 502.3 [M]⁺. Found 525.3 $[M + Na]^+$ (100); anal. calcd for $C_{27}H_{39}FN_4O_4$: C, 64.52; H, 7.82; N, 11.15. Found: C, 64.52; H, 7.81; N, 11.15.

(S)-1-(4-(2-((4-Chlorobenzyl)amino)-2-oxoethyl)-3-isopentyl-2,5 dioxoimidazolidin-1-yl)-N-(4-fluorobenzyl)cyclopropane-1-carboxamide (8g). Yellow gum. Yield 85% (117 mg). *Rf* (AcOEt/hexane, $80:20$) = 0.28; $[\alpha]_D^{20}$ –22.4 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CD₃OD), δ (ppm) = 7.13–7.08 (m, 4H), 6.97–6.95 (m, 2H), 8.82– 6.80 (m, 2H), 4.33 (d, *J* = 15.2 Hz, 1H), 4.16 (d, *J* = 15.2 Hz, 1H), 4.08−4.05 (m, 2H), 4.91 (d, *J* = 14.8 Hz, 1H), 3.35−3.32 (m, 1H),

3.01−2.99 (m, 1H), 2.95−2.92 (m, 1H), 2.84−2.80 (m, 1H), 1.64− 1.62 (m, 1H), 1.45−1.18 (m, 6H), 0.79 (d, *J* = 4.8 Hz, 3H), 0.77 (d, *J* $= 4.8$ Hz, 3H); ¹³C{¹H} NMR (101 MHz, CD₃OD), δ (ppm) = 172.2, 169.5, 167.2, 160.5 (d, *J* = 244.4 Hz), 154.8, 135.5, 132.9, 131.5, 127.4 (d, *J* = 8.1 Hz), 127.3, 127.0, 113.3 (d, *J* = 22.2 Hz), 54.7, 41.2, 40.8, 38.1, 35.2, 31.9, 24.3, 20.3, 20.2, 14.4, 14.0; ESI-MS: *m/z* (%) calcd 542.2 [M]⁺. Found 565.3 [M + Na]⁺ (100); anal. calcd for C28H32ClFN4O4: C, 61.93; H, 5.94; N, 10.32. Found: C, 61.94; H, 5.96; N, 10.32.

1-(3-Benzyl-4-(2-(cyclohexylamino)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)-N-(2,2-diphenylethyl)cyclohexane-1-carboxamide (8h). Amorphous white solid. Yield 91% (201 mg). *Rf* (AcOEt/ hexane, 80:20) = 0.21; $[\alpha]_D^{\alpha}$ ⁰ -6.7 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl3), *δ* (ppm) = 7.76 (t, *J* = 5.2 Hz, 1H), 7.18−7.14 (m, 15H), 5.78 (d, *J* = 7.6 Hz, 1H), 4.63 (d, *J* = 15.2 Hz, 1H), 4.33 (t, *J* = 8.0 Hz, 1H), 4.10 (d, *J* = 15.2 Hz, 1H), 3.99−3.95 (m, 1H), 3.71− 3.68 (m, 1H), 3.60−3.58 (m, 2H), 2.76 (dd, *J* = 16.8 and 2.8 Hz, 1H), 2.60−2.57 (m, 2H), 2.48 (dd, *J* = 16.8 and 4.0 Hz, 1H), 1.99− 1.02 (m, 18H); 13C{1 H} NMR (101 MHz, CDCl3), *δ* (ppm) = 172.9, 172.8, 156.9, 142.6, 142.5, 136.1, 128.9, 128.4, 128.34, 128.32, 128.2, 128.0, 127.9, 126.4, 126.3, 67.0, 55.5, 50.2, 48.9, 45.1, 44.5, 34.2, 32.8, 32.7, 32.3, 25.4, 25.1, 24.9, 23.0, 22.1; ESI-MS: *m*/*z* (%) calcd 634.4 [M]⁺. Found 657.5 [M + Na]⁺ (100); anal. calcd for $C_{39}H_{46}N_4O_4$: C, 73.79; H, 7.30; N, 8.83. Found: C, 73.81; H, 7.31; N, 8.82.

1-(4-(2-((2-(1H-Indol-3-yl)ethyl)amino)-2-oxoethyl)-3-benzyl-2,5 dioxoimidazolidin-1-yl)-N-(2,2-diphenylethyl)cyclohexane-1-carboxamide (8i). Amorphous white solid. Yield 72% (72 mg). *Rf* $(\text{ACOEt}) = 0.23; [\alpha]_{\text{D}}^{20} + 16.5 (\text{c} = 1.0, \text{CHCl}_3); {}^{1}\text{H NMR}$ (400 MHz, CDCl₃), δ (ppm) = 8.16 (s, 1H), 7.57 (br s, 1H), 7.48 (d, *J* = 7.6 Hz, 1H), 7.26 (d, *J* = 7.6 Hz, 1H), 7.20−7.00 (m, 17H), 6.88 (s, 1H), 5.41 (br s, 1H), 4.51 (d, *J* = 15.2 Hz, 1H), 4.34 (t, *J* = 7.8 Hz, 1H), 3.97−3.95 (m, 1H), 3.95 (d, *J* = 15.2 Hz, 1H), 3.67−3.65 (m, 2H), 2.88−2.86 (m, 2H), 2.64 (d, *J* = 15.4 Hz, 1H), 2.61−2.59 (m, 2H), 2.26 (d, *J* = 15.4 Hz, 1H), 2.04−1.56 (m, 10H); 13C{1 H} NMR (101 MHz, CDCl₃), δ (ppm) = 172.9, 172.7, 167.7, 156.8, 142.6, 142.5, 136.5, 136.0, 128.9, 128.4, 128.36, 128.33, 128.2, 128.0, 127.9, 126.3, 122.4, 112.3, 111.5, 100.0, 67.1, 55.4, 50.2, 45.1, 44.5, 39.8, 34.4, 32.3, 31.3, 29.7, 25.07, 25.00, 23.0, 22.2; ESI-MS: *m*/*z* (%) calcd 695.3 [M]⁺. Found 718.4 [M + Na]⁺ (100); anal. calcd for $C_{43}H_{45}N_5O_4$: C, 74.22; H, 6.52; N, 10.06. Found: C, 74.23; H, 6.51; N, 10.04.

(S)-1-(3-Benzyl-4-(2-(isobutylamino)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl)-N-(2,2-diphenylethyl)cyclohexane-1-carboxamide (8j). Amorphous white solid. Yield 83% (111 mg). *Rf* (AcOEt/hexane, $80:20$ = 0.28; $[\alpha]_D^{20}$ +32.6 ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 7.71 (t, *J* = 5.2 Hz, 1H), 7.19–7.14 (m, 15H), 5.89 (br s, 1H), 4.61 (d, *J* = 15.2 Hz, 1H), 4.33 (t, *J* = 8.0 Hz, 1H), 4.13 (d, *J* = 15.2 Hz, 1H), 3.98−3.95 (m, 1H), 3.71 (s, 1H), 3.68− 3.66 (m, 1H), 2.97−2.95 (m, 1H), 2.85−2.83 (m, 2H), 2.56−2.54 (m, 2H), 1.99−1.97 (m, 1H), 1.67−1.64 (m, 2H), 1.43−1.40 (m, 7H), 0.80 (d, *J* = 6.4 Hz, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 173.0, 172.7, 168.0, 156.8, 142.6, 142.5, 136.2, 128.9, 128.4, 128.3, 128.2, 128.0, 126.4, 126.3, 67.0, 55.6, 50.2, 47.3, 45.1, 44.5, 34.3, 32.3, 31.3, 28.3, 25.1, 23.0, 22.1, 20.24, 20.21; ESI-MS: *m*/*z* (%) calcd 608.3 $[M]^{+}$. Found 609.3 $[M + H]^{+}$ (100); anal. calcd for C37H44N4O4: C, 73.00; H, 7.29; N, 9.20. Found: C, 73.01; H, 7.28; N, 9.20.

(S)-2-(3-Ethyl-4-(2-((4-hydroxyphenethyl)amino)-2-oxoethyl)- 2,5-dioxoimidazolidin-1-yl)-N-hexyl-2-methylpropanamide (8k). Amorphous white solid. Yield 93% (228 mg). *Rf* (AcOEt/hexane, $80:20$) = 0.12; $[\alpha]_{D}^{20}$ –14.6 ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CD₃OD), δ (ppm) = 8.00 (br s, 1H), 7.00 (d, *J* = 6.8 Hz, 2H), 6.70 (d, *J* = 6.8 Hz, 2H), 4.11 (t, *J* = 3.2 Hz, 1H), 3.49−3.47 (m, 1H), 3.36−3.33 (m, 2H), 3.19−3.14 (m, 2H), 3.10−3.06 (m, 1H), 2.87 (dd, *J* = 13.2 and 2.4 Hz, 1H), 2.82 (dd, *J* = 13.2 and 3.6 Hz, 1H), 2.68 (t, *J* = 6.0 Hz, 2H), 1.77 (s, 3H), 1.67 (s, 3H), 1.51−1.49 (m, 2H), 1.27−1.15 (m, 6H), 1.11 (t, *J* = 5.6 Hz, 3H), 0.86 (t, *J* = 4.8 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CD₃OD), *δ* (ppm) = 173.6, 172.2, 167.9, 155.3, 154.5, 128.5, 128.1, 113.8, 60.6, 54.8, 39.6, 38.6, 34.4, 33.0, 32.8, 30.2, 27.6, 25.2, 23.3, 22.5, 21.1, 11.0; ESI-MS: *m*/*z* (%) calcd 474.3 $[M]^{+}$. Found 475.5 $[M + H]^{+}$ (37), 475.5 $[M + Na]^{+}$

 (100) , 513.5 [M + K]⁺ (22); anal. calcd for C₂₅H₃₈N₄O₅: C, 63.27; H, 8.07; N, 11.81. Found: C, 63.26; H, 8.07; N, 11.80.

(S)-N-Isobutyl-2-(4-(2-(isobutylamino)-2-oxoethyl)-3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-1-yl)-2-methylpropanamide (8l). Amorphous white solid. Yield 91% (169 mg). *Rf* (AcOEt/hexane, $80:20$) = 0.30; $[\alpha]_D^{20}$ –21.4 ($c = 0.9$, CHCl₃); ¹H NMR (400 MHz, CD₃OD), δ (ppm) = 8.09 (t, *J* = 5.2 Hz, 1H), 8.04–8.02 (m, 1H), 7.77−7.74 (m, 2H), 7.42−7.35 (m, 4H), 4.94 (d, *J* = 15.2 Hz, 1H), 4.82 (d, *J* = 15.2 Hz, 1H), 3.72 (dd, *J* = 4.4 and 2.8 Hz, 1H), 3.00− 2.98 (m, 1H), 2.90−2.88 (m, 1H), 2.64−2.62 (m, 2H), 2.51 (dd, *J* = 16.4 and 4.4 Hz, 1H), 2.45 (dd, *J* = 13.2 and 6.8 Hz, 1H), 1.77−1.72 (m, 1H), 1.70 (s, 3H), 1.62 (s, 3H), 1.43−1.41 (m, 1H), 0.83 (d, *J* = 4.4 Hz, 3H), 0.81 (d, *J* = 4.4 Hz, 3H), 0.68 (t, *J* = 6.4 Hz, 6H); ¹³C{¹H} NMR (101 MHz, CD₃OD), *δ* (ppm) = 175.2, 173.6, 169.0, 157.2, 134.4, 132.0, 131.7, 129.1, 128.8, 127.3, 126.6, 126.1, 125.3, 123.5, 78.3, 62.3, 57.1, 46.8, 44.0, 34.2, 28.4, 28.2, 25.0, 24.0, 19.7, 19.4; ESI-MS: m/z (%) calcd 494.3 [M]⁺. Found 517.4 [M + Na]⁺ (100); anal. calcd for $C_{28}H_{38}N_4O_4$: C, 67.99; H, 7.74; N, 11.33. Found: C, 68.00; H, 7.73; N, 11.33.

(S)-2-(4-(2-((2-(1H-Indol-3-yl)ethyl)amino)-2-oxoethyl)-3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-1-yl)-N-isobutyl-2-methylpropanamide (8m). Amorphous white solid. Yield 69% (59 mg). *Rf* $(ACOEt) = 0.28; [\alpha]_D^{20} - 36.9 (c = 0.9, CHCl_3);$ ¹H NMR (400 MHz, CD₃OD), δ (ppm) = 7.97–7.94 (m, 1H), 7.73–7.69 (m, 2H), 7.38−7.36 (m, 3H), 7.32−7.30 (m, 2H), 7.23−7.21 (m, 1H), 6.99− 6.97 (m, 1H), 7.91−7.86 (m, 2H), 4.85 (d, *J* = 15.2 Hz, 1H), 4.68 (d, *J* = 15.2 Hz, 1H), 3.69 (dd, *J* = 4.8 and 2.8 Hz, 1H), 3.12−3.10 (m, 1H), 2.99−2.95 (m, 2H), 2.88 (dd, *J* = 8.4 and 6.8 Hz, 1H), 2.63− 2.58 (m, 3H), 2.42 (dd, *J* = 16.4 and 4.8 Hz, 1H), 1.74−1.71 (m, 1H), 1.71 (s, 3H), 1.63 (s, 3H), 0.78 (d, *J* = 2.8 Hz, 3H), 0.76 (d, *J* = 2.8 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CD₃OD), δ (ppm) = 174.8, 173.3, 168.6, 156.9, 136.8, 134.0, 131.5, 131.4, 128.8, 128.5, 127.3, 127.0, 126.3, 125.8, 124.9, 123.2, 121.8, 118.3, 117.9, 111.7, 110.9, 78.0, 61.9, 56.8, 43.6, 39.6, 34.1, 28.2, 24.7, 24.5, 23.7, 19.4; ESI-MS: *m/z* (%) calcd 581.3 [M]⁺. Found 604.3 [M + Na]⁺ (100); anal. calcd for $C_{34}H_{39}N_5O_4$: C, 70.20; H, 6.76; N, 12.04. Found: C, 70.19; H, 6.77; N, 12.05.

(S)-2-(3-Benzyl-1-(2-((2,2-d iphenylethyl)amino)-2-oxoethyl)-2,5 dioxoimidazolidin-4-yl)-N-cyclohexylacetamide (8n). Amorphous white solid. Yield 88% (152 mg). R_f (AcOEt/hexane, 80:20) = 0.18 ; $[α]_D^{20} - 12.9$ (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), *δ* (ppm) = 7.72 (br s, 1H), 7.19−7.11 (m, 15H), 5.42 (br s, 1H), 5.54 (d, *J* = 15.2 Hz, 1H), 4.32 (d, *J* = 15.2 Hz, 1H), 4.27 (t, *J* = 7.6 Hz, 1H), 4.13−4.02 (m, 2H), 3.89−3.86 (m, 1H), 3.80 (br s, 1H), 3.75− 3.72 (m, 1H), 3.50−3.48 (m, 1H), 2.73 (d, *J* = 14.4 Hz, 1H), 2.38 (d, *J* = 14.4 Hz, 1H), 1.80–1.52 (m, 5H), 1.24–0.97 (m, 5H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 171.8, 166.6, 155.9, 142.5, 142.3, 135.9, 129.0, 128.4, 128.3, 128.2, 128.1, 128.0, 126.5, 126.4, 56.5, 49.9, 48.9, 45.7, 44.5, 41.9, 34.6, 32.8, 32.7, 25.4, 24.8, 24.7; ESI-MS: *m/z* (%) calcd 566.3 [M]⁺. Found 589.4 [M + Na]⁺ (100); anal. calcd for $C_{34}H_{38}N_4O_4$: C, 72.06; H, 6.76; N, 9.89. Found: C, 72.04; H, 6.77; N, 9.90.

(S)-2,2′*-(3-(Naphthalen-1-ylmethyl)-2,5-dioxoimidazolidine-1,4 diyl)bis(N-isobutylacetamide) (8o).* Amorphous white solid. Yield 85% (97 mg). R_f (AcOEt/hexane, 80:20) = 0.30; $[\alpha]_D^2$ ⁰ -39.0 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CD₃OD), δ (ppm) = 8.51 (br s, 0.3H), 8.16−8.14 (m, 1H), 7.92−7.87 (m, 2H), 7.55−7.44 (m, 4H), 5.10 (d, *J* = 15.6 Hz, 1H), 5.02 (d, *J* = 15.6 Hz, 1H), 4.26 (d, *J* = 17.2 Hz, 1H), 4.16 (d, *J* = 17.2 Hz, 1H), 4.07 (dd, *J* = 4.8 and 3.2 Hz, 1H), 3.16−3.14 (m, 1H), 3.12−3.10 (m, 1H), 2.81 (dd, *J* = 16.4 and 2.8 Hz, 1H), 2.74−2.66 (m, 2H), 2.57−2.51 (m, 1H), 1.89−1.85 (m, 1H), 1.55−1.50 (m, 1H), 0.93 (d, *J* = 1.6 Hz, 3H), 0.91 (d, *J* = 1.6 Hz, 3H), 0.79 (d, *J* = 6.0 Hz, 3H), 0.77 (d, *J* = 6.0 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CD₃OD), *δ* (ppm) = 173.0, 168.4, 167.9, 167.8, 156.5, 134.1, 131.4, 128.8, 128.4, 126.8, 126.4, 125.8, 124.9, 123.1, 78.1, 57.32, 57.31, 43.7, 41.0, 33.7, 28.0, 27.9, 19.2, 19.1, 19.0; ESI-MS: *m/z* (%) calcd 466.3 [M]⁺. Found 489.2 [M + Na]⁺ (100); anal. calcd for $C_{26}H_{34}N_4O_4$: C, 66.93; H, 7.35; N, 12.01. Found: C, 66.93; H, 7.37; N, 12.00.

(S)-N-(4-Chlorobenzyl)-2-(1-(2-(isobutylamino)-2-oxoethyl)-3- (naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-4-yl)acetamide (8p). Amorphous white solid. Yield 81% (131 mg). *Rf* (AcOEt/ hexane, 80:20) = 0.27; $[\alpha]_D^{20}$ –16.0 (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃), δ (ppm) = 8.06–8.03 (m, 2H), 7.76–7.74 (m, 2H), 7.45−7.33 (m, 4H), 7.15 (d, *J* = 8.4 Hz, 2H), 6.93 (d, *J* = 8.4 Hz, 2H), 6.39 (br t, *J* = 5.6 Hz, 1H), 5.01 (d, *J* = 14.8 Hz, 1H), 4.86 (d, *J* = 14.8 Hz, 1H), 4.15 (d, *J* = 17.2 Hz, 1H), 4.04 (d, *J* = 17.2 Hz, 1H), 4.02 (dd, *J* = 14.8 and 6.4 Hz, 1H), 3.73−3.72 (m, 1H), 3.61 (dd, *J* = 14.8 and 5.2 Hz, 1H), 2.99−2.97 (m, 1H), 2.85−2.72 (m, 2H), 2.57 (dd, *J* = 16.4 and 4.8 Hz, 1H), 1.67 (septet, *J* = 6.8 Hz, 1H), 0.72 (d, *J* $= 6.8$ Hz, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 171.9, 167.9, 167.2, 155.8, 136.0, 133.9, 133.4, 131.5, 131.2, 129.6, 128.8, 128.7, 127.8, 127.2, 126.5, 125.2, 123.6, 56.8, 47.4, 44.4, 42.6, 41.8, 34.5, 28.0, 20.22, 20.20; ESI-MS: *m*/*z* (%) calcd 534.2 [M]+ . Found 557.2 $[M + Na]$ ⁺ (100); anal. calcd for C₂₉H₃₁ClN₄O₄: C, 65.10; H, 5.84; N, 10.47. Found: C, 65.09; H, 5.85; N, 10.46.

(S)-N-Isobutyl-2-(3-(naphthalen-1-ylmethyl)-4-(2-((naphthalen-1-ylmethyl)amino)-2-oxoethyl)-2,5-dioxoimidazolidin-1-yl) acetamide (8q). Amorphous white solid. Yield 81% (153 mg). *Rf* $(\text{ACOEt}) = 0.43; [\alpha]_{\text{D}}^{20} - 28.5 (\text{c} = 1.0, \text{CHCl}_3);$ ¹H NMR (400 MHz, CD₃OD), *δ* (ppm) = 8.01–7.95 (m, 2H), 7.75–7.68 (m, 4H), 7.37−7.33 (m, 8H), 5.00 (d, *J* = 15.2 Hz, 1H), 4.86 (d, *J* = 15.2 Hz, 1H), 4.82 (d, *J* = 15.2 Hz, 1H), 4.73 (d, *J* = 15.2 Hz, 1H), 4.24 (d, *J* = 17.2 Hz, 1H), 4.15 (d, *J* = 17.2 Hz, 1H), 3.95 (dd, *J* = 4.8 and 3.2 Hz, 1H), 2.64 (dd, *J* = 16.8 and 3.2 Hz, 1H), 2.53 (dd, *J* = 16.8 and 4.8 Hz, 1H), 2.34 (dd, *J* = 13.2 and 6.8 Hz, 1H), 2.18 (dd, *J* = 13.2 and 6.4 Hz, 1H), 1.22 (septet, *J* = 6.8 Hz, 1H), 0.50 (d, *J* = 6.8 Hz, 3H), 0.48 (d, $J = 6.8$ Hz, 3H); ¹³C{¹H} NMR (101 MHz, CD₃OD), δ (ppm) = 173.0, 168.3, 167.9, 156.5, 134.0, 133.8, 133.1, 131.3, 131.2, 128.7, 128.4, 128.3, 127.7, 126.6, 126.4, 125.9, 125.4, 125.3, 125.0, 124.9, 123.0, 122.9, 57.2, 46.2, 43.6, 41.1, 40.8, 33.7, 27.8, 18.94, 18.92; ESI-MS: m/z (%) calcd 550.3 [M]⁺. Found 573.3 [M + Na]⁺ (100); anal. calcd for $C_{33}H_{34}N_4O_4$: C, 71.98; H, 6.22; N, 10.17. Found: C, 71.98; H, 6.22; N, 10.15.

(S)-N-(4-Chlorobenzyl)-2-(4-(2-((2,2-diphenylethyl)amino)-2-oxoethyl)-3-(naphthalen-1-ylmethyl)-2,5-dioxoimidazolidin-1-yl) acetamide (8r). Crystalline white solid. Yield 79% (127 mg). *Rf* $(ACOEt) = 0.38$; mp 132–134 °C; $[\alpha]_D^{20}$ +11.5 ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CDCl3), *δ* (ppm) = 8.40 (br t, *J* = 6.0 Hz, 1H), 7.91 (d, *J* = 8.4 Hz, 1H), 7.82−7.80 (m 1H), 7.76−7.73 (m, 1H), 7.46− 7.34 (m, 2H), 7.28−7.20 (m, 5H), 7.17−7.13 (m, 4H), 7.05−6.93 (m, 7H), 4.87 (d, *J* = 15.2 Hz, 1H), 4.84 (br s, 1H), 4.70 (d, *J* = 15.2 Hz, 1H), 4.50 (dd, *J* = 15.2 and 6.4 Hz, 1H), 4.29 (d, *J* = 17.2 Hz, 1H), 4.24 (d, *J* = 17.2 Hz, 1H), 4.20 (dd, *J* = 11.2 and 5.2 Hz, 1H), 3.71 (t, *J* = 8.0 Hz, 1H), 3.65−3.64 (m, 1H), 3.16−3.14 (m, 1H), 2.89−2.87 (m, 1H), 2.65 (dd, *J* = 16.4 and 2.4 Hz, 1H), 2.10 (dd, *J* = 16.4 and 4.8 Hz, 1H); ¹³C{¹H} NMR (101 MHz, CDCl₃), δ (ppm) = 171.7, 167.3, 166.6, 155.8, 141.23, 141.21, 137.1, 133.8, 132.6, 131.5, 131.2, 129.5, 128.9, 128.87, 128.85, 128.7, 128.4, 128.3, 127.8, 127.7, 127.3, 127.1, 127.0, 126.6, 125.0, 123.5, 56.8, 50.0, 44.5, 43.2, 42.5, 42.1, 34.6; ESI-MS: m/z (%) calcd 658.2 [M]⁺. Found 681.5 [M + Na]⁺ (100); anal. calcd for $C_{39}H_{35}CIN_4O_4$: C, 71.06; H, 5.35; N, 8.50. Found: C, 71.04; H, 5.36; N, 8.51.

■ **ASSOCIATED CONTENT**

\bullet Supporting Information

The Supporting Information is available free of charge at [https://pubs.acs.org/doi/10.1021/acs.joc.2c01903.](https://pubs.acs.org/doi/10.1021/acs.joc.2c01903?goto=supporting-info)

Copies of the $^1\mathrm{H}$, $^{13}\mathrm{C}$ NMR, and MS spectra for all new compounds and copies of the VT ¹ H NMR and DMSO titration ¹H NMR experiments [\(PDF](https://pubs.acs.org/doi/suppl/10.1021/acs.joc.2c01903/suppl_file/jo2c01903_si_001.pdf))

Accession Codes

CCDC [2191858](https://summary.ccdc.cam.ac.uk/structure-summary?pid=ccdc:2191858&id=doi:10.1021/acs.joc.2c01903) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

CCDC-2191858 contains the supporting crystallographic data for this paper. These data can be obtained free of charge *via*www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB21EZ, U.K.; fax: ++44 1223 336 033; or [deposit@ccdc.cam.ac.uk\)](mailto:deposit@ccdc.cam.ac.uk).

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Notes

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