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# Unveiling the veil of lactate in tumor-associated macrophages: a successful strategy for immunometabolic therapy

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Lactate, traditionally regarded as a metabolic waste product at the terminal of the glycolysis process, has recently been found to have multifaceted functional roles in metabolism and beyond. A metabolic reprogramming phenomenon commonly seen in tumor cells, known as the “Warburg effect,” sees high levels of aerobic glycolysis result in an excessive production of lactate. This lactate serves as a substrate that sustains not only the survival of cancer cells but also immune cells. However, it also inhibits the function of tumor-associated macrophages (TAMs), a group of innate immune cells ubiquitously present in solid tumors, thereby facilitating the immune evasion of malignant tumor cells. Characterized by their high plasticity, TAMs are generally divided into the pro-inflammatory M1 phenotype and the pro-tumour M2 phenotype. Through a process of ‘education’ by lactate, TAMs tend to adopt an immunosuppressive phenotype and collaborate with tumor cells to promote angiogenesis. Additionally, there is growing evidence linking metabolic reprogramming with epigenetic modifications, suggesting the participation of histone modification in diverse cellular events within the tumor microenvironment (TME). In this review, we delve into recent discoveries concerning lactate metabolism in tumors, with a particular focus on the impact of lactate on the function of TAMs. We aim to consolidate the molecular mechanisms underlying lactate-induced TAM polarization and angiogenesis and explore the lactate-mediated crosstalk between TAMs and tumor cells. Finally, we also touch upon the latest progress in immunometabolic therapies and drug delivery strategies targeting glycolysis and lactate production, offering new perspectives for future therapeutic approaches.

## KEYWORDS

**lactate, TAMs, immune escape, immunometabolism, targeted drug delivery**

## 1 Introduction

Macrophages are typically the most common immune cell type and are ubiquitous in body tissues. Acting as a bridge between natural and acquired immunity, macrophages not only directly engulf and eliminate pathogens or dead cells, but also act as antigen-presenting cells (APCs) to process and deliver antigens to trigger anti-tumour effects (1, 2). Tumour-associated macrophages (TAMs) are abundantly infiltrated in tumor microenvironment (TME), accounting for more than 50% of solid tumours, and are functionally plastic and heterogeneous (3, 4). In normal tissues, most macrophages undergo a coordinated functional transition between M1 and M2 phenotypes, contributing to the promotion of acute inflammation and tissue repair, respectively (5). Although in the early stages of cancer, TAMs exhibit a pro-inflammatory M1 phenotype to perform anti-tumour effects (6). However, they were ultimately transformed to an immunosuppressive and pro-angiogenic M2 phenotype under the education of TME, thereby facilitating tumour growth and evasion of immune surveillance (7).

In solid tumours, hypoxic zones are prevalent where cells use anaerobic glycolysis to convert pyruvate to lactate under low oxygen conditions (8). In 1923, Otto Heinrich Warburg discovered the phenomenon of aerobic glycolysis in tumour cells under conditions of sufficient oxygen, termed the “Warburg effect” (9). Three main advantages of the “Warburg effect” are known, including a rapid increase in adenosine triphosphate (ATP) levels, the provision of biosynthetic intermediates and the prevention of cellular oxidative stress (10, 11). In addition, this effect has also been found to be present in rapidly proliferating immune cells (12). Previous studies have focused on intermediate metabolites of the tricarboxylic acid (TCA) cycle, such as citric, succinic and fumaric acids (13). Lactate was initially considered to be a by-product of anaerobic glycolysis and was misidentified as a waste product (14). However, lactate has received much attention in recent years. Under physiological conditions (pH 7.2), lactate is present as sodium lactate (15). Whereas in TME at low pH, lactate exists as lactic acid (15). In

2014, lactate was first demonstrated to induce TAM polarization (16). The significant role of lactate in tumor immune escape has also been investigated, but the mechanisms underlying the activation of the pro-angiogenic phenotype of TAMs by lactate remain to be explored, and studies for a comprehensive insight into lactate-targeted immunotherapy are limited.

This review concentrates on the unique role of lactate-activated TAMs in aberrant angiogenesis and immune modulation, with an highlight on recent advances in lactate-targeted drugs and delivery strategies. A detailed description of lactate-mediated interactions between immune cells provides a theoretical basis for a better understanding of the non-metabolic functions of lactate, as well as a potentially valuable avenue for immunometabolic therapies.

## 2 Phenotypic and immunomodulatory effect of TAM in tumors

### 2.1 Phenotypic diversity and metabolic patterns of macrophages

Macrophages are highly plastic and can be polarized into different states, generally summarized as the classically activated M1 macrophages (anti-tumoral phenotype) and the alternatively activated M2 macrophages (pro-tumoral phenotype) (17) (Figure 1). Several signaling molecules, such as PI3K/Akt-ERK signaling (18), STAT3 (19), HIF1 $\alpha$  (20), and STAT6 (21) are involved in macrophage M2-polarization. Generally, metabolic signals and the prototypical polarizing signals like IFN- $\gamma$  and LPS leading to TAMs to express the M1 phenotype, or IL-4 and IL-10 can convert TAMs into the M2 state (22). Notably, M2 macrophages could further be classified into M2a, M2b, M2c and M2d according to their functional state (23). TAMs are thought to predominantly belong to M2d, which can be stimulated by lactate (24, 25). In addition to phenotypic diversity, TAMs also exhibit heterogeneity in their metabolic patterns, which contribute to their

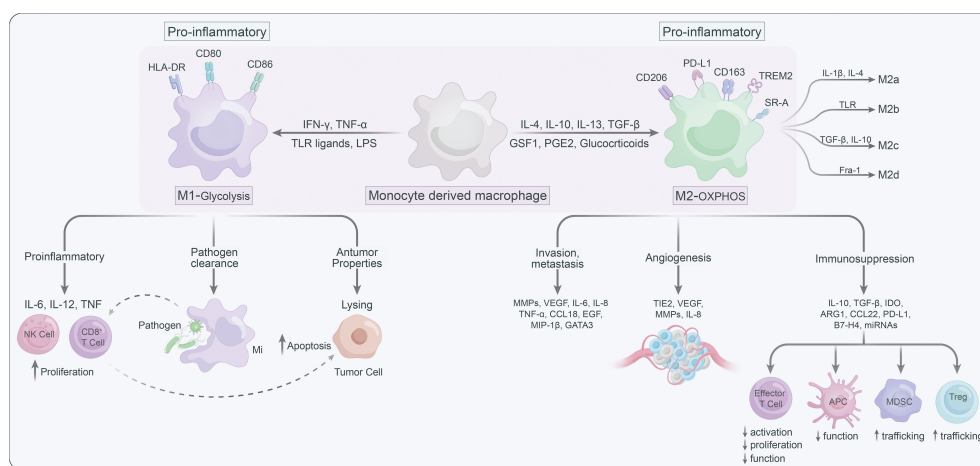


FIGURE 1  
The characteristics of M1 versus M2 macrophages.

functional differences. M1 macrophages, which mostly by relying on glycolysis and pentose phosphate pathway (PPP), are known for their pro-inflammatory roles (26). Furthermore, the pro-inflammatory TAMs can promote proliferation and recruitment of cytotoxic immune cells such as CD8<sup>+</sup> T and NK cells to support the tumor-suppressive functions of the pro inflammatory (M1-type) TAMs (27). While M2 macrophages are generally featured with enhanced oxidative phosphorylation (OXPHOS) and fatty acid oxidation (FAO), can facilitate tumor progression by immune suppression and promotion of cancer cell migration/invasion, angiogenesis, and metastasis (28). In light of this, targeting metabolic pathways has emerged as an attractive strategy for modulating the phenotypes of TAMs and their subsequent immune responses. By altering the metabolic preferences of TAMs, one could potentially shift their functions, providing a novel approach for the treatment of various diseases including cancer (29–31).

## 2.2 TAM-mediated tumor immune escape in the TME

Blocking inhibitory immune checkpoint proteins like PD-1, PD-L1, and CTLA-4 with monoclonal antibodies has revolutionized cancer treatment by counteracting cancer cells' evasion of immune responses and restoring T cell function (32). Unfortunately, a significant challenge in current immunological-checkpoint blockade therapies is that over 60% of patients do not experience any benefits due to resistance observed in certain tumors (33). The concept of “immune editing” elucidates the mechanisms behind the continued growth of tumors despite the presence of a fully operational host immune system (34). During this process, tumor cell division can give rise to reduced immunogenicity, allowing the tumor to evade immune detection by employing immunosuppressive effects or by losing the expression of target antigens (35). This immune evasion is likely to be influenced by factors such as the existing infiltration of immune cells within the tumor during treatment, the composition of the tumor stroma, and the mechanisms of immune resistance (30).

Macrophages as essential tissue-resident antigen-presenting cells, modulating immune responses by transitioning from a pro-inflammatory phenotype to an immunosuppressive phenotype that facilitates tumor growth (36, 37). This non-inflammatory phenotype is distinguished by transcriptional downregulation of iNOS, TNF, IL-12, and Toll-like receptor (TLR) signaling pathway components, alongside transcriptional upregulation of arginase 1, C-type lectin CLEC10A, and IL-10 (38, 39). In the polyoma middle T antigen model of breast cancer, CD4<sup>+</sup> T cell-derived IL-4 is thought to be responsible for the functional shift towards a non-inflammatory phenotype (40). The non-inflammatory polarization of macrophages promotes the expression of PD-L1 in monocytes, leading to the suppression of cytotoxic T cell responses (41). Consequently, tumors manipulate macrophages to establish a microenvironment that facilitates tumor growth by inducing angiogenesis, enhancing tumor cell migration, and promoting invasion (42). This non-inflammatory macrophage polarization as

a key aspect of tumor immunoevasion highlights its central role in shaping the TME.

## 3 Glycolysis, lactate, and tumor microenvironment

### 3.1 Generation of lactate in the tumour microenvironment

The Warburg effect, a hallmark of tumor cells, manifests as aerobic glycolysis, wherein the primary metabolic fate of glucose is its conversion into lactate, despite the presence of oxygen (43, 44). Both cancer cells and stromal cells release significant quantities of lactate into the TME due to the presence of the Warburg effect and reverse Warburg effect (45). Prominently found within the TME, alongside immune system elements like macrophages and lymphocytes, are cells comprising blood vessels, fibroblasts, myofibroblasts, mesenchymal stem cells, adipocytes, and the extracellular matrix (ECM), with TAMs stand out as a prominent constituent (25). In addition to tumor-derived lactate, tumor cells employ miR-375 to reprogram TAMs into lactate producers, thereby fulfilling the energy demands of tumor cells (46).

While tumor cells were once thought to be the primary generators of lactate and protons in the TME for several decades, emerging evidence now reveals that immune and stromal populations within tumors play a crucial role in substantial “lactification” and acidification contributions (47–49). TGF- $\beta$  signaling from cancer cells induces the Warburg effect in cancer-associated fibroblasts, leading to lactate secretion (50, 51). This lactate can be absorbed by neighboring tumor cells through the “reverse Warburg effect” (50). Immune cells also undergo a metabolic transition, resembling Warburg metabolism, to support their functions (52). The PI3K/Akt/mTORC1 pathway plays a critical role in promoting aerobic glycolysis in both immune cells and tumor cells (29). Studies using [18F]FDG-PET have shown increased glucose metabolism in immune cells, with activated T-lymphocytes exhibiting higher glucose uptake in immunocompetent mice (53–55). TAMs, monocytes, and neutrophils exhibit greater glycolytic dependency compared to dendritic cells (56). Tumor-infiltrating T cells have limited glycolytic activity due to glucose competition from highly glycolytic tumor cells (49, 57, 58). The lactate levels in the TME are closely linked to immune cell infiltration and metabolism. Furthermore, co-culture experiments have revealed that tumor cells can induce lactate production in TAMs via the miR-375-LDH-B axis (46).

### 3.2 Impact of lactate on macrophages in normal and non-cancer pathologies

Lactate, often perceived merely as a byproduct of cellular metabolism, actually serves as a significant signaling molecule that plays a crucial role in regulating normal physiology and pathology beyond the scope of cancer (59, 60). Under normal circumstances, systemic lactate concentrations are stringently

maintained at approximately 1–2 mM. However, these concentrations can escalate to an exceptional 40 mM within the TME, potentially impacting cellular function (61). Studies have indicated that tumor cells from both humans and mice excrete lactate *in vitro*, with substantial amounts of lactate being associated with tumor metastasis and unfavorable clinical prognosis (16, 62, 63).

Lactate is involved in various processes such as inflammation, wound healing, and immune response modulation, with macrophages being key players in these processes (64). During inflammation, lactate act as an energy source for cells, ensuring their function and survival (65). Moreover, lactate is not just a fuel but can also regulate immune cell behavior, specifically macrophages (66). For instance, lactate can influence macrophage polarization, which is an important aspect of the innate immune response (67). Beyond the inflammatory response, lactate has a critical role in wound healing. High concentrations of lactate can promote the migration of macrophages which subsequently transition from a pro-inflammatory M1 phenotype to a pro-healing M2 phenotype (66). As such, lactate serves as a mediator for wound healing by orchestrating the dynamic function of macrophages in the process (66). Further, lactate also plays a significant role in sepsis, a severe systemic inflammatory response syndrome (68). Studies have indicated that lactate produced during sepsis can impair the function of macrophages and other immune cells, contributing to immune dysfunction, a hallmark of sepsis (69, 70). Lactate also influences the process of antigen presentation by macrophages, affecting the host's adaptive immune response (68). Thus, targeting lactate metabolism in macrophages could be a potential therapeutic strategy for sepsis management.

In addition to the aforementioned pathologies, lactate has also been implicated in various other conditions such as neurodegenerative diseases, diabetes, and myocardial infarction (71–73). The common thread in all these pathological conditions is the role of lactate in the modulation of macrophage function and the associated inflammatory responses. It is noteworthy that the lactate concentration in the microenvironment dictates the impact on macrophage function. Under physiological conditions, lactate concentration is relatively low, and thus its impact on macrophages is minimal (74). However, under pathological conditions, the dramatic increase in lactate concentrations can significantly impair macrophage function (75). Understanding the exact mechanisms through which lactate impacts macrophage function in non-cancer pathologies could yield essential insights into the design of novel therapeutics for these diseases.

## 4 Function of lactate in TAM polarization and angiogenesis

Lactate operates as an active signaling molecule that exerts control over the polarization and function of TAMs via receptor-mediated signaling pathways (Figure 2). Exposure of TAMs to lactate elicits a pro-angiogenic phenotype, implicating their crucial involvement in the dysregulation of angiogenic therapies.

Hence, targeting lactate production, shuttling, and/or signaling may offer a promising approach to counteract resistance against angiogenic therapies (Figure 3).

### 4.1 Lactate-monocarboxylate transporters signaling

Lactate build-up can lead to “self-poisoning” and impair the glycolytic process (76). Excessive accumulation of lactate lowers cytoplasmic pH, leading to inhibition of alkaline pH-dependent phosphofructokinase (PFK) and LDH activity, which results in attenuated glycolysis (77). In TME, lactate can induce anti-inflammatory and pro-angiogenic ATMs phenotypes via multiple pathways. For instance, lactate can be transported bidirectionally via MCTs belonging to the SLC16A family, as MCTs are precise regulators in modulating lactate export and import (78, 79). MCT1 and MCT2 can promote lactate efflux in a substrate-dependent concentration gradient, whereas MCT3 and MCT4 are efficient lactate exporters that promote lactate efflux from highly glycolytic cells (79). However, MCT1 and MCT4 are highly expressed in numerous tumors and are linked to adverse prognosis (80). The expression of MCT1 and MCT4 positively correlated with CD163<sup>+</sup> TAMs in human breast cancer and oral squamous cell carcinoma, respectively (81, 82). MCT1 enables lactate influx that promotes glycolysis and M2 phenotypic polarization of TAMs, while knockdown of MCT4 blocks the glycolytic process by decreasing LDH-A expression (83). LDH-A-mediated lactate production increases the NAD/NADH ratio, which is critical for CD40 signaling, and CD40-induced lactate production by fine-tuning the NAD/NADH ratio for M1 macrophage polarization rather than M2 polarization (84).

#### 4.1.1 HIF- $\alpha$

Contrary to LDH-A, LDH-B preferentially oxidizes lactate to pyruvate while reducing NAD to NADH, and pyruvate competes with  $\alpha$ -ketoglutarate to suppress prolyl hydroxylase 2 (PHD2) activity and further inhibit HIF-1 $\alpha$  (85). Further, HIF-1 $\alpha$  can enhance glycolytic activity via upregulating PDK1 expression and blocking the import of lactate-derived pyruvate into mitochondria for the TCA cycle (86). By stabilizing HIF-1 $\alpha$ , lactate could enhance VEGF and Arg1 expression and induces M2-like polarization of TAMs, and VEGF and Arg1 support tumor growth via inducing neovascularization and providing substrates for cancer cell proliferation, respectively (16).

In tumor cells and activated ATMs, intracellular citrate and succinate inhibit HIF- $\alpha$  inhibitory factors (87, 88). Citrate, which accumulates due to the downregulation of isocitrate dehydrogenase (IDH), directly inhibits HIF- $\alpha$  asparagine hydroxylase (FIH) (87). As for succinate, which gets as a result of SDH inhibition, inhibits HIF prolyl hydroxylases (HPHs) (87, 88). Both result in the stabilization and activation of HIF- $\alpha$  and hence the transcription of pro-angiogenic VEGF (87, 88). In breast cancer, sodium/glucose cotransporter 1 (SGLT1) overexpression drives high glycolysis in tumor cells, and secreted lactate promotes polarization of M2 TAMs

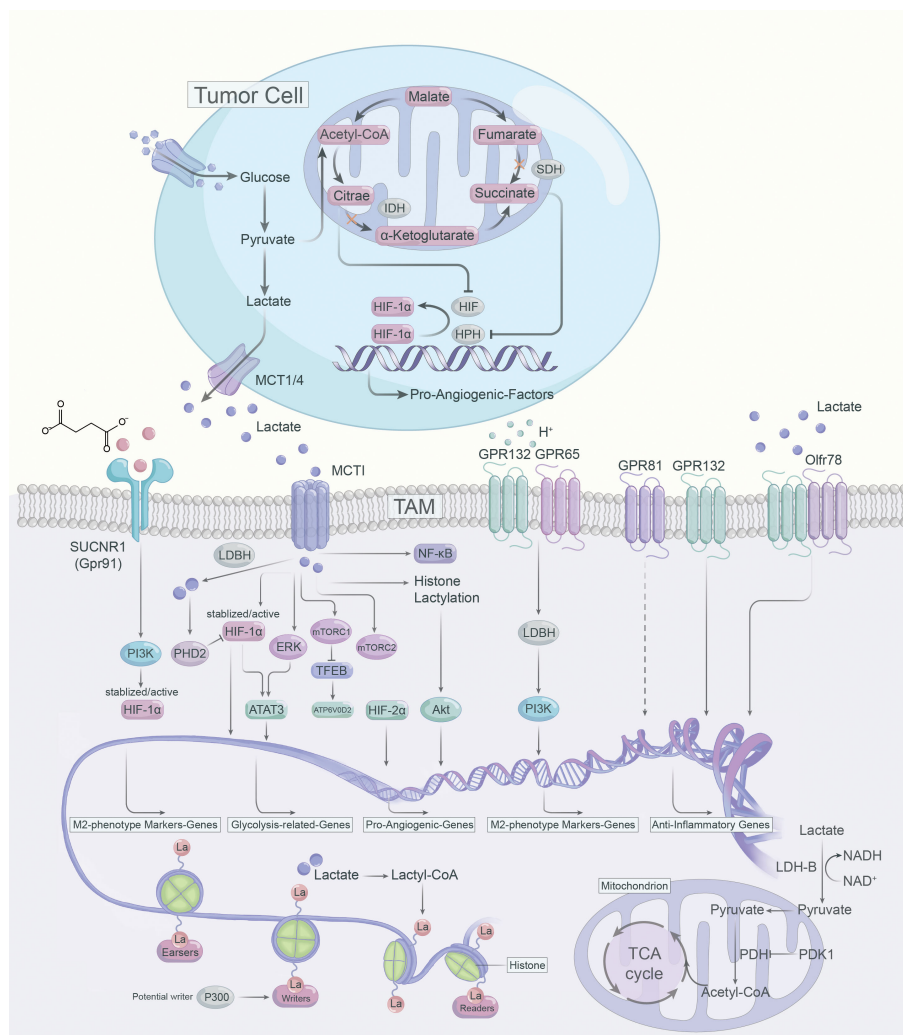


FIGURE 2 Lactate-mediated signaling pathways in the polarization of TAMs.

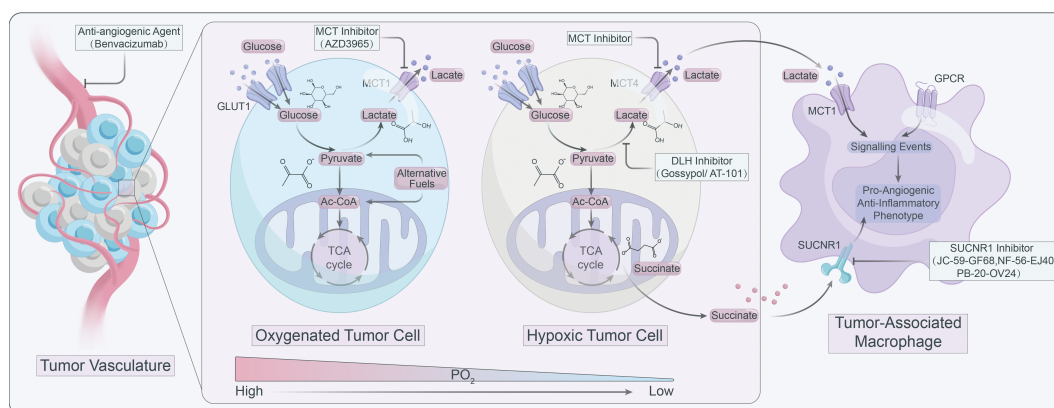


FIGURE 3 Targeting lactate signaling is a promising strategy to counteract resistance against angiogenic therapy.

via activating the HIF-1 $\alpha$ /STAT3 signaling, promoting the polarization of M2 phenotype TAMs, thus creating a vicious cycle between breast cancer cells and TAMs (89). In addition, lactate-mediated activation of the ERK/STAT3 signaling pathway was determined to perform a crucial role in the pro-angiogenic M2-like polarization of ATMs and subsequent tumor neovascularization (25). Similarly, in cervical cancer, lactate contributes to the M2 phenotype of TAMs by promoting HIF-1 $\alpha$  expression and downregulating nuclear factor kappa-B (NF- $\kappa$ B) phosphorylation in TAMs (89).

#### 4.1.2 mTOR

There are insufficient studies on mTOR in ATM. The TSC2-mTOR pathway has been identified as a critical regulator of monocyte differentiation to M2-like TAM (90). TSC2 knockdown activates mTOR, converting macrophages to an angiogenic-promoting M2-like phenotype and increasing the release of IL-10 (90). Conversely, rapamycin inhibits mTOR, leading to the differentiation of monocytes into M1-like TAM (90). Consistently, knockdown of REDD1, an inhibitor of mTOR, impedes glycolysis in TAM and inhibits excessive tumor angiogenesis and metastasis (91). Lactate activates mTORC1 via MCTs and represses TFEB, while suppressing the expression of ATP6V0d2 (a subunit of V-ATPase), a target gene of TFEB (92). However, ATP6V0d2 promotes HIF-2 $\alpha$  lysosomal degradation, blocking ATP6V0d2 induces HIF-2 $\alpha$  activation, which promotes VEGF expression and enhances the pro-angiogenic effect of TAMs (92). Furthermore, in pituitary adenomas, lactate can contribute to tumor invasion via activating mTORC2/Akt signaling (93).

#### 4.1.3 Histone lactylation

Lactate has been considered as a byproduct of sugar metabolism, especially in glycolysis, but in 2019, Zhao et al. first identified and demonstrated that lactate can promote lactonization modifications on histone lysine residues, which in turn directly regulates transcriptional expression of genes, terming it histone lysine lactylation (Kla) (94). TME is rich in lactate and the accumulation of lactate could induce histone Kla in TAMs. The authors used LPS+IFN- $\gamma$  to induce polarization of Bone marrow-derived macrophages (BMDMs) toward M1 phenotype macrophages and found not only a progressive increase in intracellular lactate levels over 24 hours, but also an increasing level of histone Kla (94). It was also found that the expression of the M2-like gene (Arg1) increased continuously with the increase of the Kla level in M1 macrophages. The experimental results suggest that as histone Kla increases, macrophages polarize toward the M2 phenotype, and even macrophages that are already M1 phenotype repolarize toward the M2 phenotype (94). Notably, lactate, an epigenetic regulatory molecule, can induce the expression of Arg1 and other homologous genes involved in wound healing via histone lysine Kla, promoting the function of M2 phenotype TAMs (94). In addition, the acetyltransferase enzyme p300 is involved in the process of histone Kla (95). Lactyl-CoA produced from lactate contributes a lactyl (La) group via p300 to the lysine tail of histones, activating pro-wound healing genes and leading to an

M2-like phenotype (95). However, which enzymes generate the intermediate molecule lactyl-CoA, from which La is derived, and which enzymes deposit (writers), remove (erasers) or recognize and interpret (readers) histone lactylation remain to be identified (96).

## 4.2 Lactate-G protein-coupled receptors signaling

### 4.2.1 Gpr132

Except MCTs, the G protein-coupled receptor 132 (Gpr132, or G2A) and Gpr65 (or TDAG8) are functional lactate receptor both highly expressed on the surface of macrophages (30, 97, 98). Gpr132 serve as a stress-inducible, seven-pass transmembrane receptor whose activation blocks the cell cycle and modulates proliferation and immunity (99, 100). It was found that cancer cell-derived lactate activated Gpr132 on ATMs to promote the M2 phenotype, and that lactate-Gpr132 axis-activated ATMs enhanced tumor cell metastasis, thus forming a positive feedback loop (97). Gpr132 activation produces inositol triphosphate (IP3) via Gq activation, and increases intracellular calcium concentration (99). Notably, although Gpr132 is also a proton receptor, GPR132 deletion specifically affects calcium mobilization triggered by lactate, rather than by hydrochloric acid (HCl) (97). This suggests that Gpr132 not only acts as a functional receptor for lactate, but that lactate is a key signal for activating Gpr132 (97). Lower Gpr132 expression is associated with better survival in breast cancer patients, so targeting the lactate-Gpr132 axis may help to abrogate the M2 phenotypic polarization of TAMs and lung metastasis of breast cancer (97). Further, activation of the lactate-G2A-PPAR $\gamma$ -axis can mediate the M2 polarization of ATMs and promote breast cancer metastasis (97, 101).

### 4.2.2 Gpr65

Bohn et al. found that in malignant melanoma, lactate promoted anti-inflammatory M2-like TAM polarization through activation of GPR65 (30). Moreover, Lailier reported that in glioblastoma (GBM), the expression of the lactate receptor GPR65 was highest at the mRNA level (102). Interestingly, lactate may be dispensable for the pro-angiogenic TAMs phenotype and anti-inflammatory function mediated by GPCRs (30, 103). Acidic TME was demonstrated to directly activate proton-sensing Gpr65 and Gpr132 on TAMs, independent of lactate, leading to cyclic AMP (cAMP) accumulation, which induces the expression of cAMP early inhibitory factor (ICER) and M2 phenotypic markers (104, 105). In addition, ICER acts as a potent inhibitor of gene transcription, blocking the expression of pro-inflammatory factors such as TNF- $\alpha$  (30, 105).

### 4.2.3 Olfr78

Odorant receptors (ORs) are the largest subfamily of GPCRs (106). 2021, Vadevoo et al. explored the role of an OR termed Olfr78 in TAM (107). Olfr78 recognizes lactate and mediates lactate-induced M2 polarization of macrophages (107). Furthermore, Olfr78 forms a heterodimer with Gpr132, which

promotes M2 macrophage polarization, and subsequently promotes tumor progression and metastasis (107). The combined effect of Olfr78 and Gpr132 enhanced Olfr78 expression and lactate-responsive activity, and conversely, Olfr78 or Gpr132 deficiency reduced lactate-mediated M2 polarization of ATMs (107). However, the potential molecular mechanisms downstream of lactate-Olfr78/Gpr132 signaling in TAMs remain to be explored.

#### 4.2.4 Gpr81

Gpr81 is the most intensively studied lactate receptor in mediating intracellular signaling (108). The expression of Gpr81 is diverse depending on the cell type and tissue microenvironment. For instance, Gpr81 is highly expressed on adipocytes and has been reported to flourish on macrophages in the intestine and lung (108, 109). In addition, DCs in the TME express high levels of Gpr81 (110). Highly expressed Gpr81 is emerging as a critical regulator of tumor growth and metastasis, lactate can modulate Gpr81 expression in lung tumor cells (111). In the TME, lactate-Gpr81 signaling plays an essential role in facilitating PD-L1 expression and chemotherapy resistance (112, 113). Additionally, the expression of MCT1 and MCT4 in tumor cells is regulated by lactate-GPR81 signaling (112). Further, lactate-Gpr81 signaling has been reported to be regulated by the PPAR $\gamma$  in adipocytes and the snail 3/STAT3 pathways in tumor cells pathways (111, 114, 115). In a preclinical model of mouse breast cancer, lactate activates Gpr81 in plasmacytoid dendritic cells (pDCs), inducing intracellular calcium mobilization and inhibiting IFN $\alpha$  production, thus augmenting immunosuppression in TME (116). In monocytes and macrophages, lactate-Gpr81 signaling activation suppresses inflammatory responses by limiting the activation of the  $\beta$ -arrestin/inflammasome pathway, mediates the histoprotective effect (117). Besides, lactate-Gpr81 signaling exerts a role in the suppression of macrophage proinflammatory cytokine production in response to LPS stimulation via inhibition of yes-associated protein (YAP) and NF- $\kappa$ B activation (118). In contrast to its anti-inflammatory role, lactate boosts LPS-stimulated TLR4 activation and NF- $\kappa$ B-dependent inflammatory gene expression in macrophages via monocarboxylate transporters and MD-2 up-regulation (119). Taken together, the above studies suggest that the lactate-Gpr81 signaling axis has a regulatory role in macrophage immune function, and it may also play an essential role in the pro-tumor function of TAMs, but remains to be determined.

## 5 Lactate mediates the communication between TAMs and tumor cells

### 5.1 Lactate as a metabolic fuel for tumour cells

Constitutive glucose uptake, a characteristic of cancer cells, supplies the energy and biosynthetic components necessary for their rapid proliferation (120). Glucose carbon in cultured cancer

cells undergoes glycolysis to generate pyruvate, which is then converted to lactate by lactate dehydrogenase (LDH) and secreted (121). This disposal of carbon as lactate fulfills multiple roles, including the NADH-dependent recycling of NAD $^{+}$  during glycolysis by LDH and the elimination of protons through monocarboxylate transporters, maintaining intracellular pH homeostasis and acidifying the extracellular space (29). Furthermore, tumors utilize lactate as a fuel source, thereby expanding its metabolic functions within the context of cancer (122). Brandon et al. demonstrated the preference of human non-small cell lung cancer (NSCLC) for lactate over glucose as a primary fuel source to sustain tumor metabolism *in vivo* by fueling the tricarboxylic acid (TCA) cycle (29). Moreover, the application of sodium oxalate in hepatocellular carcinoma (HCC) effectively suppressed lactate dehydrogenase and aerobic glycolysis, resulting in a notable reduction in lactate concentrations, ultimately compromising the viability and proliferation of tumorigenic HCC cells (123). In line with this, a study by Sheng et al. revealed that lactate as the primary contributor to circulating TCA intermediates in fasted mice with genetically engineered lung and pancreatic cancer tumors, even in the presence of glucose, highlighting its significant role in the metabolic turnover, particularly in the context of various tissues and tumors (124). As evidenced by *in vivo* isotope tracing, melanoma xenografts with a higher propensity for metastasis showed greater incorporation of lactate into TCA metabolites via MCT1-facilitated absorption compared to their counterparts with lower metastatic potential (125). In essence, the primary use of lactate in the TCA cycle to produce ATP may separate energy production from aerobic glycolysis, possibly enabling glucose to fuel other cell proliferation and metastasis processes (124, 125). Taken together, lactate is a key metabolic fuel for mouse and human tumor cells *in vivo*, hinting at a link between lactate uptake and metastasis, though more research is needed.

### 5.2 Lactate supports immunosuppression of TAMs

Previously deemed a glycolysis waste product, lactate is now recognized as a critical mediator in tumor-immune cell interactions, facilitating immune evasion (126–128). Elevated lactic acid levels can create an acidic cellular environment, suppressing immune cells like macrophages, T cells and NK cells, thereby fostering tumor proliferation and metastasis (16, 97, 129, 130). TAMs constitute over half of the immune cells in the tumor microenvironment, M1-TAMs enable antigen presentation and immune factor activation, promoting anti-tumor responses (131). In contrast, M2-TAMs dampen inflammation, evade tumor immune surveillance, and stimulate tumor growth and metastasis (132). Lactate can instigate the MCT-HIF1 $\alpha$  pathway, prompting macrophages to polarize towards M2, thereby augmenting the regulatory mechanism of macrophage polarization (133). This transition results in a decrease in IL-12 and an increase in IL-10 expression in M2-TAMs, fostering tumor development (134). Lactate can also

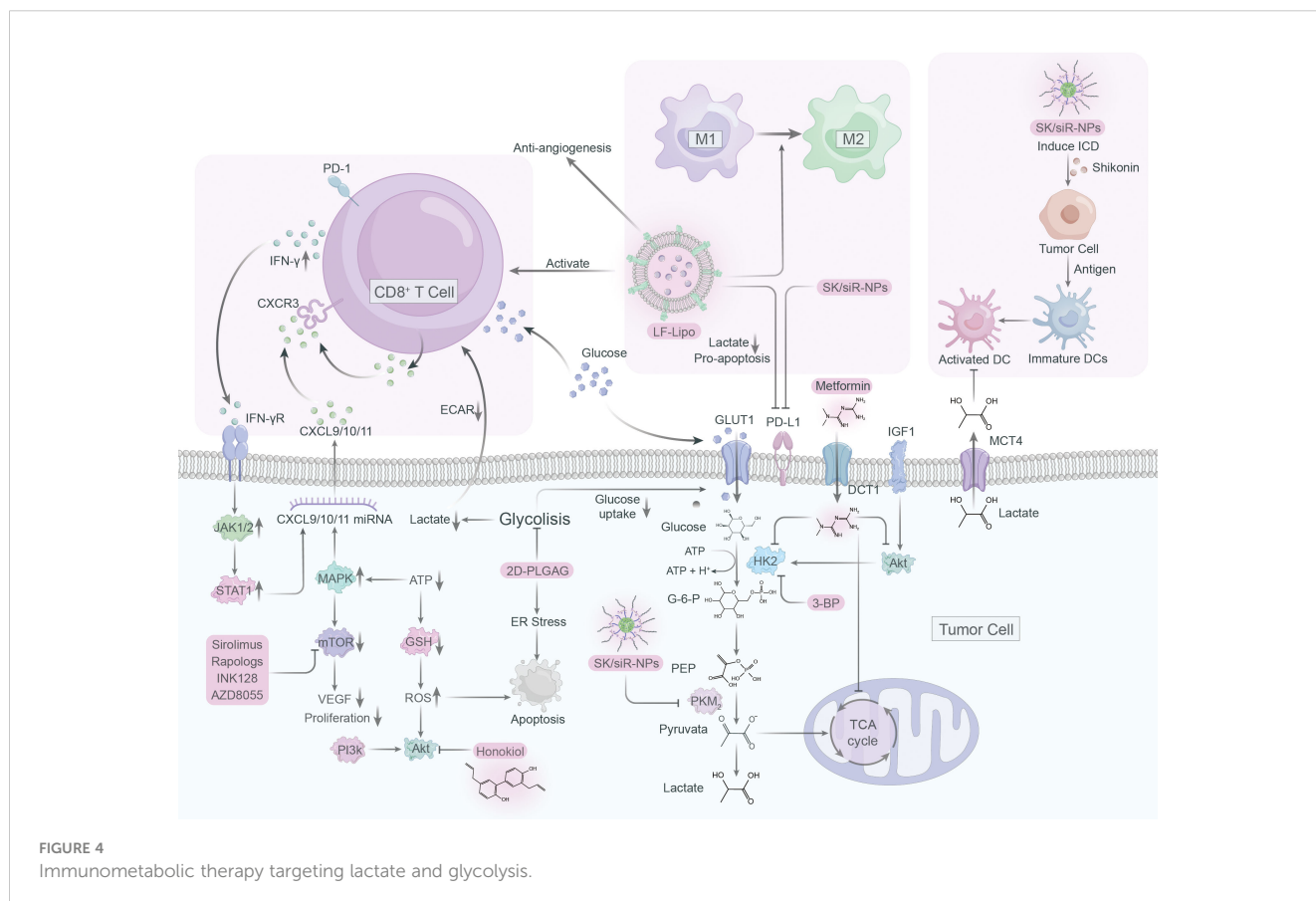
deter HIF2 $\alpha$  degradation by activating mTORC1 in macrophages, further bolstering tumor growth (92). Furthermore, lactate can impede YAP and nuclear factor- $\kappa$ B activation via a GPR81-mediated signal, curtailing pro-inflammatory cytokine production in macrophages, thus inhibiting their pro-inflammatory response to LPS stimulation (118). Modulating lactate levels can facilitate the shift of macrophages from M1 to M2 and enhance PD-L1 expression, aiding in tumor immune evasion (135). Additionally, research suggests that reducing tumor lactate levels can deter M2 macrophage polarization, inhibiting CCL17 secretion and curtailing pituitary adenoma invasion (93). A comprehensive grasp of lactate's effects on TAMs is key, not only for countering the evasion tactics of tumor cells against anti-tumor immune responses but also for forecasting how lactate metabolism could shape targeted therapies in cancer treatment.

## 6 Targeting lactate and glycolysis for immunometabolic therapy

Although only a few drugs have been studied in clinical trials targeting proteins associated with lactate in macrophages, we present an overview of immunometabolic therapies targeting glycolysis and lactate (Figure 4).

### 6.1 Targeting pyruvate kinase M2

PKM2, an essential glycolytic enzyme, is frequently overexpressed in a variety of solid tumors, playing a critical role in tumorigenesis (136–138). As such, it has become an attractive target for anti-cancer drug development. Shikonin and its analogs alkannin have shown promising results as PKM2-specific inhibitors, effectively reducing the energy supply of tumor cells by interfering with glycolysis (139). Remarkably, these compounds demonstrated a similar effect on drug-sensitive and resistant tumor cells, suggesting their clinical potential (139). However, the limited water solubility of shikonin has hindered its clinical application (140, 141). To solve this issue and improve tumor immunotherapy, a mannosylated lactoferrin nanoparticulate system (Man-LF NPs) has been developed for the co-delivery of shikonin and JQ1, an effective inhibitor of PD-L1 (142). Man-LF NPs could significantly inhibit PD-L1 and reduce lactate production in tumor cells, thus enhancing the therapeutic efficacy of immunotherapy (142). In addition, a versatile nanoparticle codelivering shikonin and PD-L1 knockdown siRNA (SK/siR-NPs) could effectively suppress PD-L1 and reduce lactate production by inhibiting PKM2 (143). SK/siR-NPs has been reported to have significant potential in tumor immunotherapy manifested by induction of immunogenic cell death (ICD), enhanced response of effector T cells, repolarization of TAMs and DC maturation (143).





## 6.2 Targeting mTOR

The mTOR pathway plays a vital role in tumorigenesis by regulating metabolism and promoting tumor growth, making it a promising target for cancer therapy (144). However, specific inhibitors such as rapamycin have limited bioavailability and solubility in water, rendering them less effective (145). To overcome this, a liposome system was developed to co-deliver rapamycin and regorafenib, an anti-angiogenic drug, effectively reducing the lactate production and promoting antitumor immune responses (146). Natural compound honokiol has been shown to inhibit the PI3K/mTOR pathway and reduce immune resistance in tumors, but its limited penetration through the blood-brain barrier has made it difficult to use in treating gliomas (147). A brain-targeted liposomal delivery system was developed by using a peptide that binds to  $\alpha 7$  nicotinic acetylcholine receptors (nAChRs), which are overexpressed on glioma cells and TAMs (148). By employing this system, honokiol and disulfiram/copper were effectively delivered to the tumor site, leading to the inhibition of glucose metabolism, lactate production, M2 to M1 TAM repolarization, triggering of tumor autophagy, and promoting antitumor immunity (148, 149).

## 6.3 Targeting hexokinase

Hexokinase is a vital enzyme in glycolysis that contributes to the development of tumor cells by priming glucose to glucose-6-phosphate, with hexokinase 2 (HK2) being the most active isozyme (150). Though 2-deoxy-D-glucose (2-DG) is a competitive inhibitor of glycolysis that can preferentially kill tumor cells, its toxicity raises concerns about its application in cancer therapy (151, 152). To address this, 2DG-loaded PLGA nanoparticles (2DG-PLGA-NPs) were developed, which effectively promote tumor cell apoptosis, enhance IFN- $\gamma$  production in CD8<sup>+</sup> T cells, and resist anti-PD-1 resistance when combined with sorafenib or anti-PD1 (153). Metformin, a typical diabetic drug, has potential antitumor properties by inhibiting glycolysis and glycogen synthesis (154). Metformin can reduce HK2 activity and impairs glycolysis, as well as indirectly inhibit HK2 by inhibiting IGF1-induced AKT phosphorylation (155). When combined with BPTES nanoparticles, metformin showed an enhanced effect on pancreatic tumor reduction (156).

## 6.4 Targeting epigenetics

Epigenetic readers such as BET proteins, notably BRD4, have been shown to upregulate PD-L1 expression, promoting tumor growth (157, 158). JQ1, a BRD4 inhibitor, can suppress PD-L1

expression and inhibit lactate production (159). Liposomal targeting codelivery of JQ1 and ecognizedt, a histone deacetylase inhibitor, can improve treatment efficacy via epigenetic regulation, with tumor cells and TAM highly expressing the lipoprotein receptor-associated protein 1 (LRP-1) receptor (159). By using lactoferrin-modified targeted liposomal system (LF-Lipo) to co-deliver ecognizedt and JQ1, binding with LRP-1 receptors on tumor cells and TAMs can reduce lactate production, repolarize M2 phenotype TAMs, and enhance CD8<sup>+</sup> T cell antitumor and anti-angiogenesis responses (159).

## 6.5 Targeting glycolysis

Targeting lactate metabolism through blocking glycolysis holds great potential as an efficacious strategy for cancer therapy. 3-BP, a halogenated analog of pyruvate, has emerged as a promising anti-tumor agent due to its selective inhibition of critical glycolysis enzymes including HK2, GAPDH, and 3-PGK, thus reducing ATP production and causing cancer cell death (160, 161). However, clinical applications of 3-BP are limited due to poor pharmacokinetics and systemic toxicity. To overcome these challenges, nanoparticle-based drug delivery technologies, such as liposomes and nanoparticles, have been developed as a targeted drug delivery solution to enhance drug efficacy and reduce toxicity (162). These groundbreaking advancements provide an exceptionally promising avenue for enhancing drug delivery, as they offer finely-tuned specificity and bolstered efficacy.

## 7 Conclusion

Lactate in TME is essential and was previously considered to be a metabolic waste product. We summarize recent evidence that lactate has a broad role in tumour immunity. Kla is a novel post-translational modification (PTM) with lactate as a substrate. The discovery of kla and its effect on macrophages helped to unravel the mystery of the Warburg effect. Although kla was shown to be a consequence of reparative gene expression in macrophages rather than a cause (163), inhibitors of key proteins in lactate metabolism show great promise in preclinical studies. Table 1 summarizes recent advances in drugs targeting lactate metabolism, but these drugs are still worthy of continued exploration in human trials.

Over the past decade, cancer immunotherapy has been recognized as one of the most promising therapeutic strategies. A large amount of research is currently focusing on PD-1/PD-L1-targeted drugs to restore or enhance isolated components of the immune system, but only a small number of individuals have had appreciable results. Metabolic checkpoints have attracted significant attention as promising therapeutic targets to enhance the anti-

TABLE 1 Pharmaceutical drugs targeting lactate metabolism in tumors.

Target	Inhibitors	Clinical trials
LDH-A	GENE-140, NHI, Galloflavin, FX-11, Stiripentol, Nifuroxazide, Honokiol, Oxamate, Sirtinol, Quiflapon	Phase II/III (Pancreatic cancer, Small cell lung cancer, Non-small cell lung cancer, Esophageal cancer, Ocular melanoma, Intracranial malignant tumor, Rhabdomyosarcoma)
LDH-B	Quinoline 3-sulfonamides, NHI	None
GLUT1	WZB117, STF-31 KRH3955, BAY-876 STF-118804, STF 31, Rapamycin, Fucoxanthin, Quercetin	None
Hexokinase	3-BrPA, 2-DG, DMJ, Lonidamine, Sorafenib, WZB-117, STF-31, 7ACC2 STF-31, PFK-158, KGP94, CG-5	Phase II (Ovarian cancer, Pancreatic cancer, Prostate cancer)
MCT1	AZD3965, AR-C155858, GW-1100, SR13800, AZB1109, Chrysin, Quercetin, Pterostilbene, DIDS, SLC-0111, 7ACC2	Phase II (Non-small cell lung cancer, Melanoma) Phase I/II (Head and neck cancer, Melanoma, Colorectal cancer)
MCT4	BAY-8002, Pterostilbene, CHC, Naringenin, Arctigenin, AZD3965, BAY-8002, CB-839, DIDS	Phase III (Renal cell carcinoma)
HIF-1 $\alpha$	Echinomycin, PX-478, 2ME, YC-1, Digoxin, DCA, Resveratrol, LW6, TEI-6720, Curcumin, 2-DG	Phase II (Glioblastoma, Renal carcinoma, Kaposi's sarcoma)
PKM2	Shikonin, DASA-10, TEPP-46, Sulfasalazine, ML265, DASA-58, MEDI3039, GSK-2837808A, Thyroid hormone, EGCG	Preclinical studies
mTOR	Rapamycin, Everolimus, Temsirolimus, AZD8055, NVP-BEZ-235, PP242, Gedatolisib, OSI-027, AZD2014, MLN0128	Phase II (Breast cancer, Renal cell carcinoma)
Histone deacetylases (HDACs)	Sodium butyrate, Trichostatin A, Vorinostat, Valproic acid, Mocetinostat	Preclinical studies
Bromodomain and extra-terminal (BET) proteins	I-BET151, JQ1, OTX015, TEN-010, ABBV-075	Preclinical studies
Glycolysis	3-BP	Preclinical studies

NHI, N-hydroxyindole; FX11, 3-dihydroxy-6-methyl-7-(phenylmethyl)-4-propylnaphthalene-1-carboxylic acid; 3-BrPA, 3-bromopyruvate; 2-DG, 2-deoxy-D-glucose; DMJ, deoxymannojirimycin; DIDS, 4,4'-diisothiocyanatostilbene-2,2'-disulfonic acid disodium salt hydrate; 7ACC2, 7-(N-benzyl-N-methylamino)-2-oxo-2H-chromene-3-carboxylic acid; CHC,  $\alpha$ -cyano-4-hydroxycinnamate; EGCG, epigallocatechin-3-gallate; 3-BP, 3-Bromopyruvate.

cancer immune response, and therefore exploring strategies to target metabolic pathways is a necessary and feasible direction of research.

The metabolism of tumor cells is highly plastic, overlapping with that of normal cells and differing *in vitro* and *in vivo* environments, leading to significant challenges in the development of pharmaceuticals that currently target cancer metabolism. The development of drug delivery technologies and physical strategies for integrated therapy may address these limitations, for example, nanotechnology-based targeted delivery and phototherapy.

In conclusion, the understanding of the role of lactate in tumor metabolic reprogramming and tumor immunity is still only the tip of the iceberg up to now, and further exploration will bring about interesting discoveries.

## Author contributions

LS and AZ were involved in the conception of the study. HT and XZ were involved in writing the article. LS and AZ critically

revised the manuscript. All authors contributed to the article and approved the submitted version.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## Glossary

APCs	antigen-presenting cells
TAMs	tumor-associated macrophages
TME	tumor microenvironment
ATP	adenosine triphosphate
TCA	tricarboxylic acid
PPP	pentose phosphate pathway
OXPPOS	oxidative phosphorylation
FAO	fatty acid oxidation
TLR	Toll-like receptor
ECM	extracellular matrix
MCTs	monocarboxylate transporters
PFK	phosphofructokinase
PHD2	prolyl hydroxylase 2
IDH	isocitrate dehydrogenase
HPHs	HIF prolyl hydroxylases
SGLT1	sodium/glucose cotransporter 1
NF- $\kappa$ B	nuclear factor kappa-B
Kla	histone lysine lactylation
BMDMs	bone marrow-derived macrophages
GPCRs	G protein-coupled receptors
La	lactyl
IP3	inositol triphosphate
HCl	hydrochloric acid
GBM	glioblastoma
cAMP	cyclic AMP
Ors	odorant receptors
pDCs	plasmacytoid dendritic cells
YAP	yes-associated protein
LDH	lactate dehydrogenase
non-small cell lung cancer	NSCLC
TCA	tricarboxylic acid
HCC	hepatocellular carcinoma
PKM2	pyruvate kinase M2
Man-LF NPs	mannosylated lactoferrin nanoparticulate system
SK/siR-NPs	nanoparticle codelivering shikonin and PD-L1 knockdown siRNA
ICD	immunogenic cell death
nAChRs	$\alpha$ 7 nicotinic acetylcholine receptors
HK2	hexokinase 2

(Continued)

## Continued

2-DG	2-deoxy-D-glucose
2DG-PLGA-NPs	2DG-loaded PLGA nanoparticles
LRP-1	lipoprotein receptor-associated protein 1
LF-Lipo	lactoferrin-modified targeted liposomal system