Gene expressions associated with longer lifespan and aging exhibit similarity in mammals

Masaki Takasugi [*](https://orcid.org/0000-0001-9880-1225),*†***, Yuya Yoshida***†***, Yoshiki Nonaka***†* **and Naoko Ohtani [*](https://orcid.org/0000-0001-8934-0797)**

Department of Pathophysiology, Osaka Metropolitan University, Graduate School of Medicine, Osaka, Japan

Received March 09, 2023; Revised May 08, 2023; Editorial Decision June 07, 2023; Accepted June 13, 2023

ABSTRACT

Although molecular features underlying aging and species maximum lifespan (MLS) have been comprehensively studied by transcriptome analyses, the actual impact of transcriptome on aging and MLS remains elusive. Here, we found that transcriptional signatures that are associated with mammalian MLS exhibited significant similarity to those of aging. Moreover, transcriptional signatures of longer MLS and aging both exhibited significant similarity to that of longer-lived mouse strains, suggesting that gene expression patterns associated with species MLS contribute to extended lifespan even within a species and that aging-related gene expression changes overall represent adaptations that extend lifespan rather than deterioration. Finally, we found evidence of co-evolution of MLS and promoter sequences of MLS-associated genes, highlighting the evolutionary contribution of specific transcription factor binding motifs such as that of E2F1 in shaping MLS-associated gene expression signature. Our results highlight the importance of focusing on adaptive aspects of aging transcriptome and demonstrate that cross-species genomics can be a powerful approach for understanding adaptive aging transcriptome.

GRAPHICAL ABSTRACT

INTRODUCTION

Owing to the recent advances in the understanding of the molecular mechanisms of aging, it is becoming clear that aging itself is a treatable condition [\(1\)](#page-12-0). Since aging is the largest risk factor for physical functional decline and numerous diseases, development of its treatment has a potential to vastly improve the quality of life of elderly people [\(2\)](#page-12-0). Therefore, further understanding of the mechanisms of aging is of crucial importance. In order to gain a systematic understanding of fundamental molecular processes of aging, much effort has been devoted to clarify agingrelated gene expression changes [\(3\)](#page-12-0). Microarray and sequencing technologies enabled comprehensive evaluation of aging-related changes in transcriptome [\(4–8\)](#page-12-0), which basically represents the overall state of cells/tissues. To date, *Tabula Muris Senis* [\(7,8\)](#page-12-0) and the Genotype-Tissue Expression (GTEx) consortium [\(6\)](#page-12-0) provided the most comprehensive information of aging-related transcriptomic changes in mouse and human, respectively. These up-to-date largescale analyses, together with numerous preceding studies, identified some common transcriptional features of aging, such as upregulation of inflammatory genes $(3,9)$, which

-^C The Author(s) 2023. Published by Oxford University Press on behalf of Nucleic Acids Research.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*}To whom correspondence should be addressed. Tel: +81 6 6645 3711; Fax: +81 6 6645 3712; Email: masaki.takasugi@omu.ac.jp Correspondence may also be addressed to Naoko Ohtani. Tel: +81 6 6645 3711; Fax: +81 6 6645 3712; Email: naoko.ohtani@omu.ac.jp ^{\dagger}The authors wish it to be known that, in their opinion, the first three authors should be regarded as Joint First Authors.

are believed to contribute to chronic inflammatory state called inflammaging. In addition to the overrepresentation of some gene ontologies, it has been discovered that the majority of aging-related gene expression changes represent reversals or extensions of developmental gene expression changes [\(10\)](#page-12-0), suggesting that aging-related gene expression changes are not just the result of damage accumulation but are associated with the developmental program [\(11,12\)](#page-12-0). A link between development and aging has been also suggested from a physiological point of view (12– 14). Gene expression changes driven by such [developmental](#page-12-0) drift and damage accumulation may contribute to agingrelated deterioration [\(11](#page-12-0)[,15\)](#page-13-0). However, although agingrelated transcriptomic changes have been described in great detail, their functional consequences still remain largely unproven.

Cross-species analysis is another approach to study genetic programs involved in lifespan regulation. For example, it has been reported that genes whose manipulation affects organismal lifespan are more conserved during evolution, and studies of such so-called longevity-associated genes (LAGs) have illuminated species-dependent and -independent longevity-associated pathways [\(16\)](#page-13-0), novel protein-protein interaction networks that may contribute to longevity, and genetic links between aging and age-related diseases [\(17\)](#page-13-0). In addition, investigation of transcriptional features of long-lived species would help to reveal prolongevity genetic programs. Transcriptomic analysis of exceptionally long-lived species, such as the naked mole-rat [\(18,19\)](#page-13-0), blind mole-rat [\(20,21\)](#page-13-0), bat [\(22\)](#page-13-0), gray whale [\(23\)](#page-13-0) and bow-head whale [\(24,25\)](#page-13-0) provided insights into the pathways that might potentially support their long lifespan. In a milestone study published in 2015, a group led by Vadim N Gladyshev reported a transcriptome of liver, kidney, and brain of 33 mammalian species (4 Carnovora, 2 Chiropetra, 1 Didelphimorphia, 1 Diprotodoncia, 1 Erinaceomorpha, 4 Euungulata, 1 Lagomorpha, 1 Monotremata, 8 Primates, 9 Rodentia and 1 Soricomorpha) and uncovered parallel evolution of gene expression and lifespan [\(26\)](#page-13-0). They found that the expressions of DNA repair- and detoxificationrelated genes positively correlate with maximum lifespan (MLS), whereas the expressions of genes involved in central metabolism and growth hormone signaling negatively correlate with MLS. One year later the same group reported transcriptomic study of cultured skin fibroblasts of 16 mammalian species (2 Chiroptera, 1 Eulipotyphla and 13 Rodentia) and confirmed the positive correlation between MLS and the expressions of genes involved in DNA repair [\(27\)](#page-13-0). Recently in 2022, a group led by Vera Gorbunova and Andrei Seluanov reported transcriptomic study of liver, kidney, brain, heart, lung and skin of 26 mammals (3 Eulipotyphla and 23 Rodentia) [\(28\)](#page-13-0). While these mammals are highly diverse in their MLS, they are mostly rodents and are closely related phylogenetically. Analyses using this dataset would thus have high power to detect MLS-associated signatures, although it may be unclear whether the findings apply only to Rodentia and Eulipotyphla or also to other orders of mammals. They found that genes related to DNA repair, microtubule organization, and RNA transport are more expressed in long-lived Rodentia/Eulipotyphla while genes related to energy metabolism and inflammation are less expressed in long-lived Rodentia/ Eulipotyphla.

To date, although cross-species transcriptomic studies revealed many features of MLS-associated gene expression patterns, how expression of these genes relates to aging still remains largely unexplored. Here, we showed that gene expression patterns associated with species MLS and aging exhibited significant similarity. Moreover, transcriptional signatures of longer MLS and aging were all associated with longer lifespan in BXD [\(29,30\)](#page-13-0) mouse strains. Our results suggest that gene expression patterns associated with species MLS contribute to extended lifespan even within a species (i.e. BXD mouse strains), and that aging-related gene expression changes overall represent adaptations that extend lifespan rather than deterioration.

MATERIALS AND METHODS

Identification of genes associated with MLS in mammals

Genes associated with MLS in mammals were identified as described below using amalgamated cross-species RNA-Seq dataset compiled by Fukushima and Pollock [\(31\)](#page-13-0). This dataset includes transcriptomic data of 52 vertebrates that are normalized by TPM and corrected by surrogate variable analysis (SVA) to remove biases likely arising from heterogeneous sampling/sequencing procedures within the dataset. For the current study, we used transcriptomic data of liver, kidney, brain, heart, ovary, and testis of 29 mammals(3 Artiodactyla, 3 Carnovora, 1 Cingulata, 1 Didelphimorphia, 1 Eulipotyphla, 1 Lagomorpha, 1 Monotremata, 1 Perissodactyla, 10 Primates, 6 Rodentia and 1 Scandentia) of which their MLS is registered in AnAge database [\(32\)](#page-13-0). Detailed information of these species, namely species name, common name, order, maximum lifespan, body weight, and age of sexual maturity are given in Supplementary Table S1. Expression data of different species were joined based on human Ensembl gene name. Expression data of non-human species that do not have 1:1 human orthologs were not joined with those of other species. For the analysis, we only used genes whose expression data were available in 10 or more species in a given tissue. A total of 16 616, 16 590, 16 175, 15 934, 14 677 and 16 001 genes passed these criteria in liver, kidney, brain, heart, ovary and testis, respectively (hereafter referred to as 'all investigated genes'). Association between gene expression and longevity was analyzed by Spearman test or by PGLS analysis using SVA-corrected log₂-transformed TPM values and log₂transformed MLS values. PGLS analysis was conducted using R libraries'ape" and 'nlme'. To take phylogenetic uncertainty into account, we performed PGLS analyses on 100 phylogenetic trees downloaded from vertlife.org [\(33\)](#page-13-0) that were generated using randomized parameters and the maximum *P*-value was used to test the association between MLS and gene expression level. Note that a set of 100 sampled trees has been suggested to capture uncertainty of parameters used for generating phylogenetic trees [\(33\)](#page-13-0). Benjamini, Krieger and Yekutieli method [\(34\)](#page-13-0) was used to control the FDR and a FDR of <10% was considered to be statistically significant.

Enrichment analyses of GO terms and transcription factor targets.

Go enrichment analyses were conducted using GOrilla [\(35\)](#page-13-0) and Enrichr [\(36\)](#page-13-0). Enrichment of transcription factor targets was analyzed by the ChEA3 [\(37\)](#page-13-0) or by Fisher's exact test using the Enrichr gene set library EN-CODE and ChEA Consensus TFs from ChIP-X. All investigated genes as defined above were used as the background except for the analyses using the ChEA3 and Enrichr web tool.

Analysis of evolutionary changes in promoter sequences of MLS-associated genes.

Sequences of 1000 bp promoter regions were retrieved using BioMart [\(38\)](#page-13-0). The genome assembly of Damaraland mole-rat was on the Scaffold level and was not available in BioMart, and thus was excluded from the analysis. Nonredundant vertebrate JASPAR CORE motifs [\(39\)](#page-13-0) were scanned using FIMO [\(40\)](#page-13-0) with default settings.

Retrieval of aging-associated genes.

The lists of genes that are differentially expressed between young and old mice tissues, and the bulk [\(7\)](#page-12-0) and single-cell RNA-Seq data [\(8\)](#page-12-0) of young and old mouse tissues were retrieved from the *Tabula Muris Senis* dataset. The lists of genes that are differentially expressed between young and old human tissues were retrieved from the GTEx dataset [\(6\)](#page-12-0) via Enrichr library GTEx Aging Signatures 2021 [\(36\)](#page-13-0). These signatures were generated by Evangelista et al. using limma-voom R package [\(41\)](#page-13-0) as described in detail in their paper under the section 'Gene expression signatures of aging from GTE' [\(42\)](#page-13-0). Genes differentially expressed between 20–29 years age group and either 30–39, 40–49, 50– 59, 60–69, 70–79 years old group were considered to be associated with aging. Genes upregulated in some age group and downregulated in some other age group were not included in aging-associated genes. The lists of genes that are differentially expressed between 20–59 and 60–99 years old human brain were retrieved from the dataset generated by Carl W. Cotman and colleagues [\(4\)](#page-12-0). The numbers of these aging-associated genes are provided in Supplementary Tables S2. Expression levels shown in the figures are given in TPMs or DESeq2 normalized counts [\(43\)](#page-13-0).

Cross-species comparison of the ratio of T cells in kidney and liver immune cells.

For mouse tissue, processed droplet scRNA-Seq data and the results of clusterization were retrieved from the *Tabula Muris Senis* dataset [\(8\)](#page-12-0). For human tissues, count data were retrieved from the original studies [\(44,45\)](#page-13-0) and processed using Seurat version 2.3.4 and then clustered by shared nearest neighbor method using 29 and 30 principal components for liver and kidney, respectively (resolution $= 0.8$). Cell clusters with high CD45 levels were extracted as immune cells and further classified by shared nearest neighbor method using 30 principal components (resolution $= 0.1$ for mouse kidney, 0.2 for mouse liver, 0.24 for human kidney and 0.2 for human liver). Cell clusters were then classified based on

the expression levels of CD11c, CD11b, CD19 and CD3ε. A cell cluster in human kidney that could not be classified using the above markers was found to express high levels of kidney epithelial markers and was not considered as a part of immune cell. A cell cluster expressing high levels of HBA1 or HBA2 (red blood cell markers) were identified in mouse kidney, human kidney, and human liver, and were not considered as a part of immune cells.

Retrieval of genes associated with longevity of rodentia*/***eulipothyphla and BXD strains.**

The lists of genes that are correlated with MLS of Rodentia/Eulipothyphla (Spearman test, BH-adjusted *P*value < 0.05 and $|\rho| > 0.4$) were retrieved from the study by Lu *et al.* [\(28\)](#page-13-0). The lists of genes whose expression in liver and kidney correlate with the median lifespan of female mice of BXD mouse strains ($|\rho| > 0.4$) were retrieved from the study by Vitiello *et al.* [\(30\)](#page-13-0). Identification of genes associated with the median lifespan of BXD mouse strains were conducted using microarray that contains one or more probes for each gene. Genes identified to be positively associated with lifespan by one probe but also identified to be negatively associated with lifespan by another probe were not included in longevity-associated genes.

Statistical significance test of the overlap between two gene sets.

Statistical significance of the overlap between two gene sets was tested by binominal test under null hypothesis that the two gene sets overlap only by chance. All statistical tests were two-sided.

RESULTS

Aging-like GO enrichment of mammalian MLS-associated genes.

In order to investigate the characteristics of MLSassociated genes, we first extracted expression data of liver, kidney, brain, heart, ovary, and testis of 29 mammals (Figure [1A](#page-3-0), Supplementary Table S1) with known maximum lifespan [\(32\)](#page-13-0) from amalgamated cross-species RNA-Seq dataset compiled by Fukushima and Pollock in 2020 [\(31\)](#page-13-0). We then identified MLS-associated genes using Spearman correlation analysis as described in Figure [1B](#page-3-0) and in Materials and Methods (Supplementary Tables S2, S3). Correlation with FDR less than 10% was considered to be statistically significant. In the liver and kidney, 637 and 470 genes correlated with MLS, respectively (Figure [1C](#page-3-0)), and fewer genes correlated with age of sexual maturity or body weight (Supplementary Table S3). In brain, 68 genes correlated with MLS and about twice the number of genes correlated with age of sexual maturity and body weight. In heart, only eight genes correlated with MLS and even fewer genes correlated with age of sexual maturity or body weight. In ovary, none of the genes correlated with MLS and only one gene correlated with body weight, while as many as 127 genes correlated with age of sexual maturity. In testis, 58, 265 and 10 genes correlated with MLS, age of sexual maturity, and body weight, respectively, suggesting

Figure 1. Identification of mammalian MLS-associated genes. (**A**) The names of 29 mammalian species that were used to extract mammalian MLSassociated genes and a phylogenetic tree drawn by R package 'ape' using phylogenetic data downloaded from VertLife.org. A heatmap showsthe availability of transcriptomic data and each box is colored blue if transcriptomic data was available in the corresponding tissue of corresponding species. Right bar graphs show MLS, age of female sexual maturity, and body weight of each species. (**B**) Procedures for the identification of mammalian MLS-associated genes. (**C**) The bar graph shows the number of MLS-associated genes identified in this study. (**D**) The correlation plot represents the overlap among mammalian MLS-associated genes identified in liver, kidney, brain, heart, ovary and testis by Spearman and PGLS methods. Each pie represents the percentage of overlapping genes in the gene set shown on the left side.

that cross-species differential gene expressions in reproductive organs more reflect age of sexual maturity than MLS. MLS-associated genes found in these tissues had absolute correlation coefficient values greater than 0.53 (Supplementary Table S3) and were largely tissue-specific (Figure [1D](#page-3-0)).

We next examined the enrichment of gene ontology (GO) terms relative to all investigated genes, and found that genes whose expression in the kidney was positively and negatively correlated with mammalian MLS were enriched with inflammatory and mitochondrial genes, respectively (Supplementary Figure S1). Considering that high expression of inflammatory genes and low expression of mitochondrial genes are primary features of aging $(3,9)$, these results may suggest some similarity between MLS-associated genes and aging-associate genes.

MLS- and mouse aging-associated genes overlap significantly.

Although our MLS-associated genes exhibited enrichment of GO terms that is reminiscent of aging transcriptome, it is not clear at this point whether it is just that different genes within the same GO term is associated with mammalian MLS and aging, or whether MLS-associated genes significantly overlap with aging-associated genes (i.e. genes whose expression was correlated with aging). To address this question, we compared MLS-associated genes with the genes whose expression changes during aging in mouse liver, kidney, brain and heart, which were identified by Wyss-Coray and Quake labs and *Tabula Muris* consortium [\(7\)](#page-12-0) (Supplementary Table S2, S4).

Surprisingly, we found a significant overlap between MLS- and aging-associated genes in liver, kidney, and brain (Figure [2A](#page-5-0)). That is, genes positively correlated with both MLS and aging, and genes negatively correlated with both MLS and aging, which we hereafter collectively refer to as 'concordant genes', appeared significantly more often than expected under the assumption of independence of MLS- and aging-associated genes (Figure [2B](#page-5-0)). In addition, in the kidney, genes positively correlated with MLS and simultaneously negatively correlated with aging, and genes negatively correlated with MLS and simultaneously positively correlated with aging, which we hereafter collectively refer to as 'discordant genes', appeared significantly less often than expected under the assumption of independence of MLS- and aging-associated genes (Figure [2B](#page-5-0)). This indicates that the discordance between MLS- and aging-associated gene expression patterns were smaller than expected by chance. These results collectively support the similarity between the transcriptional signatures of aging and longer MLS. This similarity was maintained in the kidney even after removing genes correlated with age of sexual maturity or body weight (Supplementary Figure S2). Thus, although these factors are known to be associated with species MLS, they are not the primary determinant of the similarity between the transcriptional signatures of aging and longer MLS.

Genes positively correlated with both mammalian MLS and mouse aging in the kidney were enriched with inflammation-related genes (Supplementary Figure S1). These inflammatory genes included several T cell markers, such as CD2, CD4 and CD6, suggesting that the enrichment of inflammation-related genes at least partially reflects an increase in T cells in longer-lived mammals (Figure [2C](#page-5-0)). Indeed, re-analysis of *Tabula Muris Senis* [\(8\)](#page-12-0) and public human scRNA-Seq datasets [\(44,45\)](#page-13-0) showed that the percentages of T cells in immune cells in kidney and liver were higher in human than in mouse (Figure [2D](#page-5-0)). Genes negatively correlated with both mammalian MLS and mouse aging in the kidney were enriched with mitochondrial genes (Figure [2C](#page-5-0), Supplementary Figure S1). Re-analysis of *Tabula Muris Senis* scRNA-Seq dataset [\(8\)](#page-12-0) showed that these mitochondrial genes tend to be downregulated during aging in kidney parenchymal cells, such as proximal convoluted tubule epithelial cells (Supplementary Figure S3).

Next, in order to corroborate the similarity between MLS- and aging-associated genes, we extracted MLSassociated genes using another method, namely phylogenetic generalized least squares (PGLS) regression analysis [\(46\)](#page-13-0) (Supplementary Table S2, S3). Unlike Spearman test, PGLS is a parametric test but does take the phylogenetic non-independence of species into consideration. As a result, we found 201, 832, and 114 MLS-associated genes in liver, kidney, and brain, respectively (FDR \lt 0.1). Ovary and testis had only small numbers of MLS-associated genes (23 and 22 genes, respectively). No MLS-associated gene was identified in the heart (Supplementary Table S2, S3). MLS-associated genes identified by PGLS method were largely non-overlapping with those identified by Spearman method (Figure [1D](#page-3-0)). The discrepancy is likely due at least partially to the limited sensitivity of statistical analyses but it should be noted that the overlap between the lists of MLSassociated genes identified by these methods was still statistically significant (Supplementary Figure S4A). Nonetheless, MLS-associated gene expression patterns identified by PGLS method also exhibited significant similarity with gene expression patterns associated with mouse aging (Supplementary Figure S4B and C, Tables S2 and S4). The fact that Spearman and PGLS methods pointed to the same conclusion indicates its robustness. In order to enhance the comprehensiveness of the following analyses, we hereafter combined the lists of MLS-associated genes obtained by these two methods.

In order to further verify our finding, we next investigated the similarity between MLS- and aging-associated gene expression signatures using MLS-associated genes of liver, kidney, brain, heart, lung, and skin of Rodentia and Eulipotyphla that have been identified by Lu *et al.* [\(28\)](#page-13-0). Since 23 out of 26 mammals used in their study were rodents, species-dependent gene expression would be less affected by mammalian order-dependent traits irrelevant to MLS. Rodentia/Eulipotyphla MLS-associated genes identified by Lu *et al*. were largely non-overlapping with mammalian MLS-associated genes identified by us (Figure [3A](#page-6-0)). Nonetheless, Rodentia/Eulipotyphla MLS-associated gene expression patterns exhibited significant similarity with gene expression patterns associated with mouse aging in liver, kidney, brain, heart, and lung (Figure [3B](#page-6-0) and C, Supplementary Table S5, S6). Genes correlated with mouse skin aging was the exception and their overlap with MLSassociated genes was significantly smaller than expected by

Figure 2. A significant similarity between gene expression patterns associated with mammalian MLS and mouse aging. (**A**) The heatmaps show the overlap between mouse aging-associated genes and mammalian MLS-associated genes identified by Spearman test (FDR < 0.1). Each column represents a single gene and the total number of genes shown in the heatmap is indicated in the parenthesis. The top rows show whether the gene was positively (red) or negatively (blue) associated with mammalian MLS in the tissue shown on the top of the heatmap. Other rows indicate whether the gene was upregulated (red), downregulated (blue), or unchanged (white) during aging in mouse in the tissue shown on the left of the heatmap. Only MLS-associated genes whose expression was affected by aging in either of the tissues are shown in the heatmaps. (**B**) The bar graphs represent the odds ratios of concordant (white bars) and discordant genes (black bars) (see main text for the definition). The odds ratios of concordant and discordant genes represent the concordance and discordance, respectively, between transcriptional signatures of mouse aging and longer MLS identified by Spearman test. (C) The plots show the expression levels of indicated genes in the kidney of different species or in male mice of different ages. The plots surrounded by red lines show the expression levels of 'immune system process' (GO:0002376) genes that were positively correlated with both mammalian MLS and mouse aging. The plots surrounded by blue lines show the expression levels of 'mitochondrial part' (GO:0044429) genes that were negatively correlated with both mammalian MLS and mouse aging. (**D**) The bar graphs show the percentages of T cells in the total immune cells in kidney and liver. Cell types were determined based on scRNA-seq data. The white and black bars show the percentages of T cells in mouse and human, respectively. $* FDR < 0.05$ (binominal test).

Genes whose expression positively correlated with Rodentia/Eulipotyphla MLS and mouse aging (Spearman correlation) Genes whose expression negatively correlated with Rodentia/Eulipotyphla MLS and mouse aging (Spearman correlation)

Figure 3. A significant similarity between gene expression patterns associated with Rodentia/Eulipotyphla MLS and mouse aging. (**A**) The correlation plot represents the overlap among mammalian and Rodentia/Eulipotyphla MLS-associated genes identified in liver, kidney, brain, and heart. Each pie represents the percentage of overlapping genes in the gene set shown on the left side. (**B**) The heatmaps show the overlap between genes associated with Rodentia/Eulipotyphla MLS and mouse aging. Each column represents a single gene and the total number of genes shown in the heatmap is indicated in the parenthesis. The top rows show whether the gene was positively (red) or negatively (blue) associated with Rodentia/Eulipotyphla MLS in the tissue shown on the top of the heatmap. The bottom rows indicate whether the gene was upregulated (red) or downregulated (blue) during aging in mouse in the tissue shown on the top of the heatmap. Only MLS-associated genes whose expression was also affected by aging are shown in the heatmaps. (**C**) Bar graphs represent the odds ratios of concordant (white bars) and discordant genes (black bars). * FDR < 0.05 (binominal test).

chance. This could be explained by our observation that aging-related gene expression changes were highly fluctuating in mouse skin (Supplementary Figure S5). Thus, unlike other tissues, aging-related genes in mouse skin are less likely to represent cumulative effects of aging, which could be due to the fact that external factors play major roles in skin aging.

MLS- and human aging-associated genes overlap significantly.

Although we established the similarity between gene expression patterns associated with mouse aging and the MLS of Rodentia/Eulipotyphla or a wider range of mammals (hereafter collectively refer to as species MLS), it remains to be tested whether aging-associated gene expression patterns of other species, most importantly human, also exhibit similarity to MLS-associated gene expression patterns. We therefore retrieved genes that are associated with human aging from the GTEx dataset [\(6\)](#page-12-0) via Enrichr library [\(36\)](#page-13-0) and tested their relationship with MLS-associated genes (Supplementary Table S2 and S6). Odds ratios of concordant genes were higher than 1 in mammalian MLS-associated genes in two out of three tissues (liver and brain) and in Rodentia/Eulipotyphla MLS-associated genes in five out of six tissues (liver, kidney, brain, heart, and skin) (Figure [4A](#page-7-0)–D). On the other hand, odds ratios of discordant genes were lower than 1 in mammalian MLS-associated

genes in two out of three tissues (liver and kidney) and in Rodentia/Eulipotyphla MLS-associated genes in all six tissues. Importantly, odds ratios of concordant genes were higher than those of discordant genes in all cases. The similarity between species MLS- and human aging-associated gene expression patterns was statistically significant in kid-ney, brain, and skin (Figure [4B](#page-7-0) and D). In order to corroborate this finding, we retrieved genes associated with human brain aging from another independent dataset generated by Carl W. Cotman and colleagues [\(4\)](#page-12-0). Using this dataset, we confirmed that gene expression pattern associated with MLS in the brain exhibited a significant similarity to gene expression pattern of human brain aging (Figure [4E](#page-7-0) and F). Taken together, our results indicate the similarity between the transcriptional signatures of longer MLS and aging of mouse and human. Genes positively and negatively correlated with species MLS both contributed to the similarity between MLS- and aging-associated gene expression patterns (Supplementary Figure S6).

MLS- and aging-associated expressions are both linked to longer lifespan.

Similarity between gene expression patterns associated with MLS and aging might suggest that a significant portion of aging-related gene expression changes is actually beneficial rather than harmful for longevity. In order to investigate how MLS- and aging-associated genes relate to the

Figure 4. A significant similarity between gene expression patterns associated with species MLS and human aging. (A, C, E) The heatmaps show the overlap between genes associated with species MLS and human aging. Each column represents a single gene and the total number of genes shown in the heatmap is indicated in the parenthesis. The top rows show whether the gene was positively (red) or negatively (blue) associated with species MLS in the tissue shown on the top of the heatmap. The bottom rows indicate whether the gene was upregulated (red), downregulated (blue), or unchanged (white) during aging in human in the tissue shown on the top of the heatmap. Only MLS-associated genes whose expression was also affected by aging are shown in the heatmaps. (**A**) The overlap between genes associated with mammalian MLS and human aging (according to GTEx database). (**C**) The overlap between genes associated with Rodentia/Eulipotyphla MLS and human aging (according to GTEx database). (**E**) The overlap between genes associated with species MLS and human brain aging (according to the study by Berchtold *et al*). (**B**, **D**, **F**) The bar graphs shown in (B), (D) and (F) represent the odds ratios of concordant (white bars) and discordant genes (black bars) in the genes shown in (A), (C) and (E), respectively. * FDR < 0.05 (binominal test).

variation in lifespan within a mammalian species, we next compared MLS- and aging-associated gene expression patterns with those associated with longer median lifespan in genetically heterogeneous BXD mouse strains [\(29,30\)](#page-13-0) that were generated by crossing C57BL/6J and DBA/2J mice. Gene expression patterns associated with longer median lifespan in these strains were identified in liver and kidney (and also in eye, bone and adrenal glands) in a previous study by Ewald *et al.* [\(30\)](#page-13-0). In liver and kidney, genes positively correlated with both species MLS and mouse lifespan, and genes negatively correlated with both species MLS and mouse lifespan, appeared significantly more of-

ten than expected by chance. On the other hand, genes positively correlated with species MLS and simultaneously negatively correlated with mouse lifespan, and genes negatively correlated with species MLS and simultaneously positively correlated with mouse lifespan, appeared significantly less often than expected by chance (Figure [5\)](#page-8-0). These results suggest that gene expression patterns associated with species MLS contribute to extended longevity even within a species. Importantly, the same significant similarities were observed between gene expression patterns associated with mouse aging and mouse lifespan. Gene expressions commonly associated with species MLS and mouse aging also

Figure 5. Gene expression patterns associated with species MLS and/or mouse aging exhibit significant similarities to those associated with the lifespan of genetically heterogeneous mouse strains. The white bars indicated as 'MLS' show the odds ratios of genes positively correlated with both mouse lifespan and species MLS, and genes negatively correlated with both mouse lifespan and species MLS. The white bars indicated as 'Aging' show the odds ratios of genes positively correlated with both mouse lifespan and mouse aging, and genes negatively correlated with both mouse lifespan and mouse aging. The white bars indicated as 'MLS & Aging' show the odds ratios of genes positively correlated with all mouse lifespan, species MLS, and mouse aging, and genes negatively correlated with all mouse lifespan, species MLS, and mouse aging. The black bars indicated as 'MLS' show the odds ratio of genes positively correlated with mouse lifespan and simultaneously negatively correlated with MLS, and genes negatively correlated with mouse lifespan and simultaneously positively correlated with MLS. The black bars indicated as 'Aging' show the odds ratio of genes positively correlated with mouse lifespan and simultaneously negatively correlated with mouse aging, and genes negatively correlated with mouse lifespan and simultaneously positively correlated with mouse aging. The black bars indicated as 'MLS $\&$ Aging' show the odds ratios of genes positively correlated with mouse lifespan and simultaneously negatively correlated with both species MLS and mouse aging, and genes negatively correlated with mouse lifespan and simultaneously positively correlated with both species MLS and mouse aging. $*$ FDR < 0.05 (binominal test).

exhibited similarity to those associated with mouse lifespan (Figure 5, Supplementary Figure S7). These results suggest that aging-related gene expression changes overall represent adaptations that extend lifespan rather than deterioration.

Although both mammalian and Rodentia/Eulipotyphla MLS-associated genes strongly supported our conclusion, it should be noted that these two gene sets have very different relationships with immune regulation. That is, while we found that inflammatory genes were overrepresented in genes that are positively correlated with mammalian MLS, Lu *et al.* [\(28\)](#page-13-0) reported such overrepresentation in genes that are negatively correlated with Rodentia/Eulipotyphla MLS. To better understand this difference, we extracted immune/inflammatory response genes whose Spearman coefficients of correlation between species MLS and expressions in liver, kidney, brain, and heart were significantly different between mammals and Rodentia/Eulipotyphla (Figure [6A](#page-9-0)). We found that genes involved in positive regulation of cytokine production and $NFKB$ signaling tend to be positively and negatively correlated with mammalian and Rodentia/Eulipotyphla MLS, respectively. On the other hand, genes involved in negative regulation of cytokine production and N F κ B signaling tend to be negatively and positively correlated with mammalian and Rodentia/Eulipotyphla MLS, respectively (Figure [6B](#page-9-0)). Importantly, immune/inflammatory response genes whose correlation coefficients with mammalian MLS were significantly higher than with Rodentia/Eulipotyphla MLS showed a weak but significant positive correlation with median lifespan of BXD mouse strains (Figure [6C](#page-9-0)). Immune/inflammatory response genes that were upregulated during aging in mouse liver and kidney were also positively correlated with mouse lifespan. On the other hand, immune/inflammatory response genes whose correlation coefficients with Rodentia/Eulipotyphla MLS were significantly higher than with mammalian MLS did not correlated with mouse lifespan. Thus, adaptation of immune regulation to longer lifespan evolved in different ways in Rodentia/Eulipotyphla and other mammals.

Our finding that gene expressions associated with aging and longer lifespan exhibit similarity aligns well with a previous argument by de Magalhaes ˜ *et al.* [\(9\)](#page-12-0) that many of aging-related gene expression changes could represent adaptations to aging. Aging-related increase in the expression of transcriptional repressor REST (RE1-Silencing Transcription Factor) is a proven example of such adaptations. In human brain, age-related upregulation of REST is beneficial for longevity and cognitive performance [\(47,48\)](#page-13-0). Therefore, we tested whether such adaptation also occurred during the evolution of longer lifespan. Enrichment analysis of transcription factor (TF) targets using ChEA3 [\(37\)](#page-13-0) and ENCODE ChIP-seq library showed that REST targets were the most enriched TF targets in genes whose expression in the brain negatively correlated with mammalian MLS (Figure [7A](#page-10-0)). In genes whose expression in the brain negatively correlated with Rodentia/Eulipotyphla MLS, the top 6th enriched TF targets were those of REST. Moreover, consensus REST targets (i.e. genes that were commonly identified to be bound by REST in multiple experiments) were significantly over- and underrepresented in the genes whose expression in the brain was negatively and positively correlated with species MLS, respectively (Figure [7B](#page-10-0)). Thus, suppression of REST targets that is protective against aging-related brain dysfunction seems to occur both during aging and the evolution of longer lifespan.

C

Correlation coefficients with median lifespan of BXD mouse strains (immune/inflammatory response genes)

Figure 6. Immune/inflammatory response genes are differentially associated with MLS of Rodentia/Eulipotyphla and those of a wider range of mammals. (**A**) The heatmapsshow the Spearman coefficients of correlation between species MLS and gene expressions. Each column represents a Spearman coefficient of single gene. Only immune/inflammatory genes (GO:0006945 and GO:0006955) whose Spearman coefficients were significantly different (two-tailed *t* test, FDR < 0.1) between tissues of Rodentia/Eulipotyphla and those of a wider range of mammals are shown in the figure. (**B**) The top 10 enriched GO biological process terms of the genes shown in (A). (**C**) The boxplots show the coefficients of correlation between median lifespan of BXD mouse strains and gene expression. Only immune/inflammatory genes (GO:0006945 and GO:0006955) that were upregulated during aging in mouse liver or kidney, or those whose Spearman coefficients were significantly different (two-tailed *t* test, FDR < 0.1) between tissues of Rodentia/Eulipotyphla and those of a wider range of mammals are shown in the figure. * *P*-value < 0.05 (one-sample Wilcoxon test).

TFs regulating the evolutionary changes in the expression of MLS-associated genes.

It is important to note that the aforementioned results do not necessarily indicate that REST activity itself correlates with MLS in the brain. Rather, REST binding sites might have been increased selectively in the promoter regions of genes associated with MLS in the brain. It is also very

possible that the evolutionary changes in the expression of these REST- and MLS-associated genes were driven by other TFs. Understanding of the regulatory mechanisms that had driven the evolutionary changes in the expression of MLS-associated genes is of great importance, since the effects of MLS-associated gene expression on longevity could depend on the biological context. In order to investigate such regulatory mechanisms, we analyzed the evolu-

Top 10 enriched transcription factor targets in genes whose A B expression negatively correlated with species MLS in the brain **Transcription factor** Data source p-value **Odds ratio REST** U87MG 3.5E-03 3.1 Odds ratio of consensus REST targets MLS CBX2 1.5E-02 1.8 K562 $\frac{1.6}{2.7}$ $q = 2.1 \times 10^{-3}$ **ZNF263 HEK293** 3.3E-02 Δ MEF2A K562 4.3E-02 Mammalian **CTCF** GM12892 5.2E-02 1.6 3 TCF7L2 **HEK293** 8.3E-02 1.5 $\overline{2}$ 1.5
 1.7 **CEBPB** H1HESC 8.8E-02 $q = 3.8 \times 10^{-4}$ ZBTB7A K562 9.4E-02 $\overline{1}$ E_{2F6} HELAS3 1.0E-01 1.6 **CTCFL** 1.4 $1.1E - 01$ K562 Ω Mammalian Rodentia/
Mammalian Rodentia/
MLS Eulipotyphia MLS 1.6 CTCF **MEL** 5.5E-15 ၯ NRF1 **HEPG** 3.3E-13 1.9 를 MAZ **MEL** 6.7E-13 1.9 Rodentia/
Eulipothyphla Genes positively correlated GATA1 G1EER $\overline{1}$ $.5E-11$ 1.6 \Box with species MLS in the brain **MYOG** Myocyte $1.5E-11$ 1.6 1.2E-09 1.7 **REST** Myocyte Genes negatively correlated MYOD₁ $2.0E-09$ 1.6 Myocyte with species MLS in the brain GM12878 2.1E-09 1.6 EGR1 4.5E-09 1.7 MXI1 CH12LX **ESRRA** GM12878 7.0E-09 2.0 Genes positively correlated
with mammalian MLS in the liver C · Artiodactyla PRDM5 (repressor) CREB3L1 (activator) BARHL1 (repressor) \bullet Carnivora 1.2 Cingulata Ŏ $\overline{2}$ · Didelphimorphia 0.9 1 \bullet Eulipotyphla 1.5 · Lagomorpha 0.6 Odds ratio of genes containing TF motif $\overline{1}$ 0.5 ó Monotremata 0.3 0.5 \bullet Perissodactyla $R^2 = 0.55$ $R^2 = 0.48$ $R^2 = 0.42$ \bullet Primates $q = 1.8 \times 10^{-2}$ $q = 5.0 \times 10^{-2}$ $q = 5.4 \times 10^{-3}$ $\overline{0}$ $\overline{0}$ $\overline{0}$ · Rodentia $\overline{2}$ 2 $\overline{\mathbf{A}}$ 6 \mathfrak{D} 6 4 6 Δ Scandentia Genes negatively correlated
with mammalian MLS in the brain Genes negatively correlated with Genes positively correlated with
mammalianMLS in the kidney mammalian MLS in the brain RFX6 (activator) LHX2 (activator) E2F1 (activator) TGIF2LY (repressor) 1.2 1.5 0.2 1.5 0.9 0.15 1 $\overline{1}$ 0.6 0.1 $R^2 = 0.35$ $R^2 = 0.56$ 0.5 0.5 0.3 0.05 $q = 3.9 \times 10^{-2}$ $q = 4.2 \times 10^{-3}$ $R^2 = 0.43$ $R^2 = 0.50$ $= 7.6 \times$ 10^{-3} O $= 3.9 \times 10^{-2}$ α $\boldsymbol{0}$ Ω $\boldsymbol{0}$ \cap $\overline{2}$ $\overline{4}$ $\,6$ $\overline{2}$ $\overline{\mathbf{A}}$ 6 \overline{c} $\overline{4}$ 6 $\overline{2}$ $\overline{4}$ 66

Figure 7. TFsregulating MLS-associated genes expression and their evolutionary changes. (**A**) Top 10 transcription factortargetsthat were overrepresented in the genes whose expression in the brain negatively correlated with species MLS. (**B**) The bar graph shows the odds ratios of consensus REST target genes in the genes whose expression in the brain correlated with species MLS. (**C**) The plots show the odds ratio of genes containing at least one motif of the indicated TFs in their 1000 bp promoter regions. * FDR < 0.05 [Fisher's exact test for (A, B), and Spearman correlation test for (C)].

 $Log_{2}MLS$

tionary changes in the presence of TF binding sequences in the 1000 bp upstream regions of transcriptional start sites (TSS) of mammalian MLS-associated genes using FIMO [\(40\)](#page-13-0) and vertebrate JASPAR CORE position weight matrices [\(39\)](#page-13-0). We limited our TFBS analysis to 1000 bp upstream regions of TSS due to technical reasons, although it should be kept in mind that other regions also contain many TFBS. Rodentia/Eulipotyphla MLS-associated genes were not investigated because promoter sequences were not available for the majority of species used in the comparative study [\(28\)](#page-13-0). For each motif and each species, we calculated [\(1\)](#page-12-0) the ratio of genes containing at least one motif in their promoters and [\(2\)](#page-12-0) the average number of motifs per promoter for MLS-associated genes and background genes (i.e. all investigated genes). We then calculated the odds ratios of these two values between MLS-associated genes and background genes and tested their correlation with MLS (Spearman test). Motif was considered to be associated with MLS genes if the odds ratios of both of these values were significantly correlated with MLS (FDR \langle 5%). As a result, we found seven TF binding motifs whose presence in the promoter regions of mammalian MLS-associated genes correlated with MLS (Figure 7C, Supplementary Table S7), suggesting that MLS evolved in parallel with promoter

Figure 8. A summary of the study.

sequences of MLS-associated genes. Importantly, evolutionary changes in the presence of these transcriptional activator- and repressor-binding motifs were all consistent with the evolutionary changes in the expression of MLSassociated genes (Figure [7C](#page-10-0)). For example, in genes whose expression in the brain positively correlated with MLS, the presence of the binding motif of transcriptional activator LHX2, which has been reported to play major roles in forebrain development [\(49\)](#page-13-0), was positively correlated with MLS. On the other hand, in genes whose expression in the brain negatively correlated with MLS, the presence of the binding motifs of transcriptional activator E2F1 [\(50\)](#page-13-0) and putative transcriptional repressor TGIF2LY [\(51\)](#page-13-0) was negatively and positively correlated with MLS, respectively. Notably, genes whose expression in the brain negatively correlated with MLS not only lost binding motifs of an activator E2F (E2F1) during the evolution of longevity (Figure [7C](#page-10-0)), but were also enriched with genes targeted by a repressor E2F (E2F6) [\(50\)](#page-13-0) in the longest-lived primate (human) (Figure $7A$). These consistent results suggest that E2F transcription factors rather than REST had shaped MLS-associated gene expression signature in the brain during the evolution of longevity.

DISCUSSION

Our conclusions are based on the analyses of two different types of MLS-associated genes, namely those identified by

Lu *et al.* [\(28\)](#page-13-0) in Rodentia/Eulipotyphla and those identified by us in a wider range of mammals. Identification of MLS-associated genes in Rodentia/Eulipotyphla would be less affected by mammalian order-dependent traits irrelevant to MLS. On the other hand, findings obtained from the investigations of genes associated with longevity in a wider range of mammals including primates and human would more likely to be relevant to human biology. Importantly, MLS-associated genesidentified using Spearman and PGLS methods both lead to the same conclusions. The complementarity of these analyses therefore enhances the robustness and generality of our conclusions (Figure 8). Regarding the identification of MLS-associated genes, two novel sophisticated methods using ranking-based method [\(23\)](#page-13-0) and machine learning analysis [\(52\)](#page-14-0) have been recently developed by groups led by Alexey A. Moskalev, Vadim E. Fraifeld and Robi Tacutu, and their utilization may allow identification of additional overlaps between MLS- and aging-associated genes.

In this study, we clearly showed that genes correlated with mouse/human aging and species MLS overlap significantly. MLS-associated gene expression signature is likely contributing to longer lifespan and thus our results suggest that a significant portion of aging-related gene expression changes could be beneficial for longevity. Although dysregulated inflammatory responses underlie pathogenesis of many diseases, their physiological functions are also crucial for tumor suppression [\(53\)](#page-14-0). High expression of inflammatory genes in long-lived mammals may represent better immunity. Indeed, we found that gene expression patterns associated with species MLS and aging were all associated with longer lifespan in genetically heterogeneous mouse strains. This strongly supports the notion that gene expression patterns associated with species MLS contribute to extended lifespan even within a species, and that agingrelated gene expression changes overall represent adaptations that extend lifespan.

There are some caveats regarding the interpretation of our results. First, considering that loss of proteostasis is a major hallmark of aging [\(54\)](#page-14-0), age-related changes in mRNA levels may tend not to be reflected in protein levels. In the current study, we only analyzed mRNA levels and it is not clear if the same conclusion could be drawn from the same analysis using the levels of functional proteins instead of mRNA levels. Second, expression data used in this study was quantified at the gene level and not at the transcript level. Thus, we cannot exclude the possibility that age- or species-dependent splicing has some influence on our results. Third, changes in mRNA levels during aging and in the evolution of longevity may be the result rather than the cause of aging/longevity. For example, age-related decreases in mRNA levels may reflect DNA damage blocking their transcription, and longer-lived species may downregulate damage-vulnerable gene transcripts. Fourth, molecular changes observed during aging are also often observed during development (10,11[,55–57\)](#page-14-0). Many development-associated genes are subsumed within aging-associated genes, especially when the latter were identified by comparing young adult and old animals. Therefore, the similarity between gene expressions associated with aging and longer lifespan may be at least partially due to a link between genetic programs regulating development and longevity. Finally, it should be noted that although genes associated with aging and longer lifespan exhibited statistically significant similarity, these two gene sets were still largely non-overlapping with each other. One of the major reasons for this is that genes identified to be associated with aging or longer lifespan highly depend on species, strains, sex, conditions of animals, and methods used for the analysis. For example, as described above for Spearman and PGLS methods, each statistical method has strengths and weaknesses. In this regard it should be also noted that DESeq2 [\(43\)](#page-13-0) that was used to identify mouse aging-related genes has been shown to severely understate FDR values when the sample size is large [\(58\)](#page-14-0). Also, it has been reported that genes associated with aging greatly vary among 129sv, BALB/c, CBA, DBA, C57BL/6, C3H and B6C3F mice [\(59\)](#page-14-0). If the majority of genes associated with longer lifespan in a given set of species overlapped with genes associated with aging in a given strain of animal, these MLS-associated genes would not overlap to the same extent with genes associated with aging in other strains or species. Therefore, if genes associated with longer lifespan and aging have similarity in mammals regardless of species, strains, sex, and other conditions, the overlap between these two gene sets, even if statistically significant, theoretically has to be weak.

DATA AVAILABILITY

All the data analyzed in this study were retrieved from freely accessible public databases as described above. Source data underlying the figures are provided in the supplementary tables.

SUPPLEMENTARY DATA

[Supplementary](https://academic.oup.com/nar/article-lookup/doi/10.1093/nar/gkad544#supplementary-data) Data are available at NAR Online.

ACKNOWLEDGEMENTS

We thank Dr Nobuyuki Kutsukake for technical and scientific advice.

FUNDING

Japan Society for the Promotion of Science (JSPS-KAKENHI) [20K15695 to M.T., 22H03540 to N.O., and 23KJ1831 to Y.N.]; Astellas Foundation for Research on Metabolic Disorders [2021A3097 to M.T.]; Takeda Science Foundation [to M.T. and N.O.]; Japan Science and Technology Agency (JST SPRING) [JPMJSP2139 to Y.Y.]. Funding for open access charge: Japan Science and Technology Agency (JST SPRING) [JPMJSP2139 to Y.Y.]. *Conflict of interest statement.* None declared.

REFERENCES

- [1.](#page-0-0) Gonzalez-Freire,M., Diaz-Ruiz,A., Hauser,D., Martinez-Romero,J., Ferrucci,L., Bernier,M. and de Cabo,R. (2020) The road ahead for health and lifespan interventions. *Ageing Res. Rev.*, **59**, 101037.
- [2.](#page-0-0) Budovsky,A., Muradian,K.K. and Fraifeld,V.E. (2006) From disease-oriented to aging/longevity-oriented studies. *Rejuvenation Res.*, **9**, 207–210.
- [3.](#page-0-0) Frenk,S. and Houseley,J. (2018) Gene expression hallmarks of cellular ageing. *Biogerontology*, **19**, 547–566.
- [4.](#page-0-0) Berchtold,N.C., Cribbs,D.H., Coleman,P.D., Rogers,J., Head,E., Kim,R., Beach,T., Miller,C., Troncoso,J., Trojanowski,J.Q. *et al.* (2008) Gene expression changes in the course of normal brain aging are sexually dimorphic. *Proc. Natl. Acad. Sci. U.S.A.*, **105**, 15605–15610.
- [5.](#page-0-0) Zahn,J.M., Poosala,S., Owen,A.B., Ingram,D.K., Lustig,A., Carter,A., Weeraratna,A.T., Taub,D.D., Gorospe,M., Mazan-Mamczarz,K. *et al.* (2007) AGEMAP: a gene expression database for aging in mice. *PLoS Genet.*, **3**, e201.
- [6.](#page-0-0) GTEx-consortium (2013) The Genotype-Tissue Expression (GTEx) project. *Nat. Genet.*, **45**, 580–585.
- [7.](#page-0-0) Schaum, N., Lehallier, B., Hahn, O., Pálovics, R., Hosseinzadeh, S., Lee,S.E., Sit,R., Lee,D.P., Losada,P.M., Zardeneta,M.E. *et al.* (2020) Ageing hallmarks exhibit organ-specific temporal signatures. *Nature*, **583**, 596–602.
- [8.](#page-0-0) Tabula Muris Consortium (2020) A single-cell transcriptomic atlas characterizes ageing tissues in the mouse. *Nature*, **583**, 590–595.
- [9.](#page-0-0) de Magalhães, J.P., Curado, J. and Church, G.M. (2009) Meta-analysis of age-related gene expression profiles identifies common signatures of aging. *Bioinformatics*, **25**, 875–881.
- [10.](#page-1-0) Somel,M., Guo,S., Fu,N., Yan,Z., Hu,H.Y., Xu,Y., Yuan,Y., Ning,Z., Hu,Y., Menzel,C. *et al.* (2010) MicroRNA, mRNA, and protein expression link development and aging in human and macaque brain. *Genome Res.*, **20**, 1207–1218.
- [11.](#page-1-0) de Magalhães, J.P. (2012) Programmatic features of aging originating in development: aging mechanisms beyond molecular damage? *FASEB J.*, **26**, 4821–4826.
- [12.](#page-1-0) Blagosklonny,M.V. (2013) Aging is not programmed: genetic pseudo-program is a shadow of developmental growth. *Cell Cycle*, **12**, $3736 - 3742$.
- [13.](#page-1-0) Dilman,V.M. (1971) Age-associated elevation of hypothalamic, threshold to feedback control, and its role in development, ageine, and disease. *Lancet*, **1**, 1211–1219.
- [14.](#page-1-0) Dilman,V.M., Revskoy,S.Y. and Golubev,A.G. (1986) Neuroendocrine-ontogenetic mechanism of aging: toward an integrated theory of aging. *Int. Rev. Neurobiol.*, **28**, 89–156.
- [15.](#page-1-0) Horvath,S. and Raj,K. (2018) DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nat. Rev. Genet.*, **19**, 371–384.
- [16.](#page-1-0) Yanai,H., Budovsky,A., Barzilay,T., Tacutu,R. and Fraifeld,V.E. (2017) Wide-scale comparative analysis of longevity genes and interventions. *Aging Cell*, **16**, 1267–1275.
- [17.](#page-1-0) Budovsky,A., Abramovich,A., Cohen,R., Chalifa-Caspi,V. and Fraifeld,V. (2007) Longevity network: construction and implications. *Mech. Ageing Dev.*, **128**, 117–124.
- [18.](#page-1-0) Kim,E.B., Fang,X., Fushan,A.A., Huang,Z., Lobanov,A.V., Han,L., Marino,S.M., Sun,X., Turanov,A.A., Yang,P. *et al.* (2011) Genome sequencing reveals insights into physiology and longevity of the naked mole rat. *Nature*, **479**, 223–227.
- [19.](#page-1-0) Yu,C., Li,Y., Holmes,A., Szafranski,K., Faulkes,C.G., Coen,C.W., Buffenstein, R., Platzer, M., de Magalhães, J.P. and Church, G.M. (2011) RNA sequencing reveals differential expression of mitochondrial and oxidation reduction genes in the long-lived naked mole-rat when compared to mice. *PLoS One*, **6**, e26729.
- [20.](#page-1-0) Li,K., Wang,L., Knisbacher,B.A., Xu,Q., Levanon,E.Y., Wang,H., Frenkel-Morgenstern,M., Tagore,S., Fang,X., Bazak,L. *et al.* (2016) Transcriptome, genetic editing, and microRNA divergence substantiate sympatric speciation of blind mole rat, Spalax. *Proc. Natl. Acad. Sci. U.S.A.*, **113**, 7584–7589.
- [21.](#page-1-0) Malik,A., Domankevich,V., Lijuan,H., Xiaodong,F., Korol,A., Avivi,A. and Shams,I. (2016) Genome maintenance and bioenergetics of the long-lived hypoxia-tolerant and cancer-resistant blind mole rat, Spalax: a cross-species analysis of brain transcriptome. *Sci. Rep.*, **6**, 38624.
- [22.](#page-1-0) Huang,Z., Whelan,C.V., Foley,N.M., Jebb,D., Touzalin,F., Petit,E.J., Puechmaille,S.J. and Teeling,E.C. (2019) Longitudinal comparative transcriptomics reveals unique mechanisms underlying extended healthspan in bats. *Nat. Ecol. Evol.*, **3**, 1110–1120.
- [23.](#page-1-0) Toren,D., Kulaga,A., Jethva,M., Rubin,E., Snezhkina,A.V., Kudryavtseva,A.V., Nowicki,D., Tacutu,R., Moskalev,A.A. and Fraifeld,V.E. (2020) Gray whale transcriptome reveals longevity adaptations associated with DNA repair and ubiquitination. *Aging Cell*, **19**, e13158.
- [24.](#page-1-0) Seim,I., Ma,S., Zhou,X., Gerashchenko,M.V., Lee,S.G., Suydam,R., George,J.C., Bickham,J.W. and Gladyshev,V.N. (2014) The transcriptome of the bowhead whale Balaena mysticetus reveals adaptations of the longest-lived mammal. *Aging (Albany NY)*, **6**, 879–899.
- [25.](#page-1-0) Keane,M., Semeiks,J., Webb,A.E., Li,Y.I., Quesada,V., Craig,T., Madsen,L.B., van Dam,S., Brawand,D., Marques,P.I. *et al.* (2015) Insights into the evolution of longevity from the bowhead whale genome. *Cell Rep.*, **10**, 112–122.
- [26.](#page-1-0) Fushan,A.A., Turanov,A.A., Lee,S.G., Kim,E.B., Lobanov,A.V., Yim,S.H., Buffenstein,R., Lee,S.R., Chang,K.T., Rhee,H. *et al.* (2015) Gene expression defines natural changes in mammalian lifespan. *Aging Cell*, **14**, 352–365.
- [27.](#page-1-0) Ma,S., Upneja,A., Galecki,A., Tsai,Y.M., Burant,C.F., Raskind,S., Zhang,Q., Zhang,Z.D., Seluanov,A., Gorbunova,V. *et al.* (2016) Cell culture-based profiling across mammals reveals DNA repair and metabolism as determinants of species longevity. *Elife*, **5**, e19130.
- [28.](#page-1-0) Lu,J.Y., Simon,M., Zhao,Y., Ablaeva,J., Corson,N., Choi,Y., Yamada,K.Y.H., Schork,N.J., Hood,W.R., Hill,G.E. *et al.* (2022) Comparative transcriptomics reveals circadian and pluripotency networks as two pillars of longevity regulation. *Cell Metab.*, **34**, 836–856.
- [29.](#page-1-0) Ashbrook,D.G., Arends,D., Prins,P., Mulligan,M.K., Roy,S., Williams,E.G., Lutz,C.M., Valenzuela,A., Bohl,C.J., Ingels,J.F. *et al.* (2021) A platform for experimental precision medicine: the extended BXD mouse family. *Cell Syst.*, **12**, 235–247.
- [30.](#page-1-0) Vitiello,D., Dakhovnik,A., Statzer,C. and Ewald,C.Y. (2021) Lifespan-associated gene expression signatures of recombinant BXD mice implicates Coro7 and set in longevity. *Front. Genet.*, **12**, 694033.
- [31.](#page-1-0) Fukushima,K. and Pollock,D.D. (2020) Amalgamated cross-species transcriptomes reveal organ-specific propensity in gene expression evolution. *Nat. Commun.*, **11**, 4459.
- [32.](#page-1-0) Tacutu,R., Thornton,D., Johnson,E., Budovsky,A., Barardo,D., Craig,T., Diana,E., Lehmann,G., Toren,D., Wang,J. *et al.* (2018) Human ageing genomic resources: new and updated databases. *Nucleic Acids Res.*, **46**, D1083–d1090.
- [33.](#page-1-0) Upham,N.S., Esselstyn,J.A. and Jetz,W. (2019) Inferring the mammal tree: species-level sets of phylogenies for questions in ecology, evolution, and conservation. *PLoS Biol.*, **17**, e3000494.
- [34.](#page-1-0) Benjamini,Y., Krieger,A.M. and Yekutieli,D. (2006) Adaptive linear step-up procedures that control the false discovery rate. *Biometrika*, **93**, 491–507.
- [35.](#page-2-0) Eden,E., Navon,R., Steinfeld,I., Lipson,D. and Yakhini,Z. (2009) GOrilla: a tool for discovery and visualization of enriched GO terms in ranked gene lists. *BMC Bioinf.*, **10**, 48.
- [36.](#page-2-0) Chen,E.Y., Tan,C.M., Kou,Y., Duan,Q., Wang,Z., Meirelles,G.V., Clark,N.R. and Ma'ayan,A. (2013) Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. *BMC Bioinf.*, **14**, 128.
- [37.](#page-2-0) Keenan,A.B., Torre,D., Lachmann,A., Leong,A.K., Wojciechowicz,M.L., Utti,V., Jagodnik,K.M., Kropiwnicki,E., Wang,Z. and Ma'ayan,A. (2019) ChEA3: transcription factor enrichment analysis by orthogonal omics integration. *Nucleic Acids Res.*, **47**, W212–W224.
- [38.](#page-2-0) Smedley,D., Haider,S., Ballester,B., Holland,R., London,D., Thorisson,G. and Kasprzyk,A. (2009) BioMart–biological queries made easy. *Bmc Genomics [Electronic Resource]*, **10**, 22.
- [39.](#page-2-0) Castro-Mondragon,J.A., Riudavets-Puig,R., Rauluseviciute,I., Lemma,R.B., Turchi,L., Blanc-Mathieu,R., Lucas,J., Boddie,P., Khan,A., Manosalva Pérez,N. et al. (2022) JASPAR 2022: the 9th release of the open-access database of transcription factor binding profiles. *Nucleic Acids Res.*, **50**, D165–D173.
- [40.](#page-2-0) Grant,C.E., Bailey,T.L. and Noble,W.S. (2011) FIMO: scanning for occurrences of a given motif. *Bioinformatics*, **27**, 1017–1018.
- [41.](#page-2-0) Law,C.W., Chen,Y., Shi,W. and Smyth,G.K. (2014) voom: precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol.*, **15**, R29.
- [42.](#page-2-0) Evangelista,J.E., Clarke,D.J.B., Xie,Z., Lachmann,A., Jeon,M., Chen,K., Jagodnik,K.M., Jenkins,S.L., Kuleshov,M.V., Wojciechowicz,M.L. *et al.* (2022) SigCom LINCS: data and metadata search engine for a million gene expression signatures. *Nucleic Acids Res.*, **50**, W697–W709.
- [43.](#page-2-0) Love,M.I., Huber,W. and Anders,S. (2014) Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.*, **15**, 550.
- [44.](#page-2-0) MacParland,S.A., Liu,J.C., Ma,X.Z., Innes,B.T., Bartczak,A.M., Gage,B.K., Manuel,J., Khuu,N., Echeverri,J., Linares,I. *et al.* (2018) Single cell RNA sequencing of human liver reveals distinct intrahepatic macrophage populations. *Nat. Commun.*, **9**, 4383.
- [45.](#page-2-0) Young,M.D., Mitchell,T.J., Vieira Braga,F.A., Tran,M.G.B., Stewart,B.J., Ferdinand,J.R., Collord,G., Botting,R.A., Popescu,D.M., Loudon,K.W. *et al.* (2018) Single-cell transcriptomes from human kidneys reveal the cellular identity of renal tumors. *Science*, **361**, 594–599.
- [46.](#page-4-0) Grafen,A. (1989) The phylogenetic regression. *Philos. Trans. R. Soc. Lond. B Biol. Sci.*, **326**, 119–157.
- [47.](#page-8-0) Lu,T., Aron,L., Zullo,J., Pan,Y., Kim,H., Chen,Y., Yang,T.H., Kim,H.M., Drake,D., Liu,X.S. *et al.* (2014) REST and stress resistance in ageing and Alzheimer's disease. *Nature*, **507**, 448–454.
- [48.](#page-8-0) Zullo,J.M., Drake,D., Aron,L., O'Hern,P., Dhamne,S.C., Davidsohn,N., Mao,C.A., Klein,W.H., Rotenberg,A., Bennett,D.A. *et al.* (2019) Regulation of lifespan by neural excitation and REST. *Nature*, **574**, 359–364.
- [49.](#page-11-0) Chou,S.J. and Tole,S. (2019) Lhx2, an evolutionarily conserved, multifunctional regulator of forebrain development. *Brain Res.*, **1705**, $1 - 14$
- [50.](#page-11-0) Kent,L.N. and Leone,G. (2019) The broken cycle: E2F dysfunction in cancer. *Nat. Rev. Cancer*, **19**, 326–338.
- [51.](#page-11-0) Blanco-Arias,P., Sargent,C.A. and Affara,N.A. (2002) The human-specific Yp11.2/Xq21.3 homology block encodes a potentially functional testis-specific TGIF-like retroposon. *Mamm. Genome*, **13**, 463–468.
- [52.](#page-11-0) Kulaga,A.Y., Ursu,E., Toren,D., Tyshchenko,V., Guinea,R., Pushkova,M., Fraifeld,V.E. and Tacutu,R. (2021) Machine learninganalysis of longevity-associated gene expression landscapes in mammals. *Int. J. Mol. Sci.*, **22**, 1073.
- [53.](#page-11-0) Egen,J.G., Ouyang,W. and Wu,L.C. (2020) Human Anti-tumor Immunity: insights from Immunotherapy Clinical Trials. *Immunity*, **52**, 36–54.
- [54.](#page-12-0) Hipp,M.S., Kasturi,P. and Hartl,F.U. (2019) The proteostasis network and its decline in ageing. *Nat. Rev. Mol. Cell Biol.*, **20**, 421–435.
- [55.](#page-12-0) Takasugi,M. (2011) Progressive age-dependent DNA methylation changes start before adulthood in mouse tissues. *Mech. Ageing Dev.*, **132**, 65–71.
- [56.](#page-12-0) Horvath,S. (2013) DNA methylation age of human tissues and cell types. *Genome Biol.*, **14**, R115.
- [57.](#page-12-0) de Magalhães, J.P. (2023) Ageing as a software design flaw. *Genome Biol.*, **24**, 51.
- [58.](#page-12-0) Li,Y., Ge,X., Peng,F., Li,W. and Li,J.J. (2022) Exaggerated false positives by popular differential expression methods when analyzing human population samples. *Genome Biol.*, **23**, 79.
- [59.](#page-12-0) Park,S.K., Kim,K., Page,G.P., Allison,D.B., Weindruch,R. and Prolla,T.A. (2009) Gene expression profiling of aging in multiple mouse strains: identification of aging biomarkers and impact of dietary antioxidants. *Aging Cell*, **8**, 484–495.

-^C The Author(s) 2023. Published by Oxford University Press on behalf of Nucleic Acids Research.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.