CASE REPORT Open Access

Brazilian XP-E siblings carrying a novel *DDB2* variant developed early-onset melanoma: a case report

Ana Rafaela de Souza Timoteo^{1†}, Isabel Cristina Pinheiro de Almeida^{2†}, Andrey A Yurchenko³, Sheila Ramos de Miranda Henriques², Paulo de Souza Segundo⁴, Fatemeh Rajabi³, Sergey Nikolaev^{3†} and Tirzah Braz Petta^{1,5*†}

Abstract

Background Xeroderma pigmentosum group E (XP-E) is one of the least common forms of XP, a rare syndrome where patients are prone to develop skin cancer in exposed sunlight areas. XP-E patients are generally not diagnosed until they are adults due to the mild phenotype. Case presentation: two XP-E siblings, female, 23 years, and male, 25 years, from a Brazilian consanguineous family carrying the novel missense pathogenic variant in *DDB2* gene, NM_000107.3:c.1027G > C, associated with skin cancer early-onset and severe phenotype, as nodular melanoma in the cornea and in the ear. Conclusion: The assessment of genomic variant pathogenicity was a challenge since this family belongs to an underrepresented population in genomic databases. Given the scarcity of literature documenting XP-E cases and the challenges encountered in achieving an early diagnosis, this report emphasizes the imperative of sun protection measures in XP-E patients. Additionally, it highlights the detrimental impact of the COVID-19 pandemic on cancer diagnosis, leading to the manifestation of a severe phenotype in affected individuals.

Keywords Case Report, Xeroderma Pigmentosum, XP-E, Melanoma

Tirzah Braz Petta

tirzah.petta@usc.edu

Background

Xeroderma pigmentosum (XP) is characterized by increased sensitivity to ultraviolet radiation-induced sunburn, skin cancers, ocular disease, and neurological degeneration [1]. Patients with XP have a 2,000-fold increased risk of melanoma and a 10,000-fold increased risk of nonmelanoma skin cancer XP [2]. XP is classified into eight genetic groups, according to mutated gene: groups XP-A to XP-G presenting alteration in XPA, ERCC3, XPC, ERCC2, DDB2, ERCC4, and ERCC5 genes, respectively, and the group XP-V is characterized by mutations in POLH [3]. The diagnosis is primarily clinic-based and confirmed by genetic tests or by functional cell-based methodologies.

XP-E is a rare form of the disorder, with a few reported cases in which patients with pathogenic variants at the *DDB2* gene result in absent or dysfunctional *DDB2*



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and the use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativeccommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

[†]Ana Rafaela de Souza Timoteo and Isabel Cristina Pinheiro de Almeida contributed equally to the study.

[†]Sergey Nikolaev and Tirzah Braz Petta are both senior author.

^{*}Correspondence:

¹ Departamento de Biologia Celular e Genética, Universidade Federal do Rio Grande do Norte, Av. Senador Salgado Filho, s/n, Natal 59078-970, RN. Brazil

² Hospital Liga contra o Câncer, Av. Miguel Castro, Natal 1355, 59062-000, RN. Brazil

³ Cancer Genomics Lab, B2M, Gustave Roussy Cancer Campus, 114 rue Edouard Vaillant, Villejuif 94805, France

⁴ Hospital Universitário Onofre Lopes, Universidade Federal do Rio Grande do Norte, Av. Nilo Peçanha, 620, 59012-300, RN, Natal, Brazil

⁵ Keck School of Medicine, Department of Pathology, University of Southern California, HMR 315, 2011, Zonal Avenue, Los Angeles, CA 90089-9092, USA

(damage-specific DNA-binding protein 2, also known as p48 subunit) [4, 5]. The human *DDB2* gene, located on chromosome 11 (11p11.2), consists of 10 exons encoding a 48 kDa protein of 427 amino acids exclusively localized in the nucleus and forms a heterodimer with DDB1 forming UV-DDB complex. DDB2 plays a central role in Global Genomic Nucleotide Excision Repair (GG-NER) in recognizing DNA damage [4, 6]. DDB2 binds to pyrimidine dimers including isomers of CPD (cyclobutane pyrimidine dimers) and 6-4PP (6-4 pyrimidine-pyrimidone photoproducts) with a high affinity and specificity.

In this article, we share the compelling clinical journey of two siblings, and we have obtained the necessary consent from both the patients and their parents to showcase their images. The rationale behind this decision stems from the fact that the lesions affect their face and eye areas, making the inclusion of pictures immensely valuable in accurately describing the tumors.

The two siblings with a clinical diagnosis of XP, a 20-year-old female (ID_4.19) and a 22 years-old male (ID_4.20) fourth and third child of consanguineous parents (Fig. 1A). They live in a highly consanguineous region with a high prevalence of rare genetic diseases, named Seridó, in the Rio Grande do Norte state, Brazil [7]. They both had the first diagnosis of skin carcinoma at an early age when she was 13 years old and he was 15 years old, not common for XP-E. Both were submitted to multiple surgeries to remove skin lesions in sun-exposed areas. The patient ID_4.19 had removed 19 BCC and 20 SCC and patient ID_4.20 had removed 10 BCC and 25 SCC. Both have early diagnosis of metastatic melanoma at the age of 23 years.

Case presentation

1. The Patient ID_4.19 is female, 23 years (Born in 1999). No neurological abnormalities detected. Her skin presents punctate, flat, pigmented lesions on the eyelids, face, and all UV-exposed areas of the body. She is being followed at the Head and Neck Department since 2012 and had multiple skin lesions removed (Table 1). In May 2018, she underwent a rhinotomy, without reconstruction (Fig. 1C1). Both eyes showed an extensive pigmented lesion on the entire corneal surface, presenting visual acuity of the right eye (RE) 20/100 and left eye (LE) with vision impairment. She underwent an excisional biopsy for a corneal lesion in the LE, diagnosed as SCC and this pigmented lesion was elevated, extending throughout the cornea, making it impossible to assess the internal structures of the eye (Fig. 1C2 and 1C3). She reported a progressive increase of the lesion for the last 5 years when she was in use of topical Mito-

- mycin therapy, however, she lost follow-up and only came back in 2020, after the evolution of the lesion and vision loss of the LE (Fig. 1C4). She underwent evisceration surgery of the LE and the result of corneal biopsy was a nodular melanoma, Breslow thickness 5,5 mm, mitotic index 4/1,0mm2 and ulceration present. The pathology report showed a free/ negative tumor resection margin. Her RE had a SCC lesion and she was treated with topical Mitomycin, with a follow-up with the ophthalmologist. During the COVID-19 pandemic she lost follow-up again and she came back in August 2021 with an expansive and dark lesion on the left eyeball with extraorbital extension, measuring $4.9 \times 2.8 \times 3.4$ cm. The diagnosis was recurrent or residual melanoma, she underwent an additional evisceration surgery. The nodular melanoma showed Breslow thickness 30,0 mm, Clark 5, TIL (tumor infiltrating lymphocytes) and ulceration present, mitotic index 11/1,0 mm2. She was treated with radiotherapy and 6 cycles of cisplatin 100 mg, currently under observation for new lesions and followed by the oncologist.
- 2. The Patient ID_4.20 is male, 25 years (Born in 1997). No neurological abnormalities detected. He presents multiple lesions in areas exposed to UV, such as his back (Fig. 1C5), legs, arms, and face (ears, nose and lips). He is being followed at the Head and Neck Department since 2012 and had multiple skin lesions removed (Table 1). In February 2020, he was submitted to total parotidectomy on the right side to remove a neuroendocrine carcinoma, carcinoid type (Fig. 1C6 and 1C7). He did not present with vision impairment. He lost follow-up during the COVID-19 pandemic and came back to follow-up in February 2021, at 25 y, with a suspicious lesion in the left ear, measuring 1,7×0,8 cm. The diagnosis was nodular melanoma, with 8,0 mm, mitotic index 5/1,00 m2, TIL and ulceration present. The immunohistochemistry result showed stage is pT4bpNx, Breslow thickness 8,0 mm, Clark 5, IHC profile, S100+, MART-1+, p63-, cytokeratin- and EpCAM-. At the same time, he also had two extensive lesions at the parathyroid $(3,2\times2,4)$ cm and $2,2 \times 1,8$ cm), that was extended to the cervical region with nerve compression and neuropathic pain in the right side. Stellate ganglion block was made using analgesic treatment with buprenorphine (5 mg) and pregabalin (75 mg). In September 2022, the CT image analysis showed lung and bone metastasis. He had treatment with cisplatin for the malignant melanoma and due to toxicity, his performance status was low he was admitted to the ICU for a gastrostomy e tracheotomy, and now is under palliative care. Unfortunately, he passed away on 12/06/2022.

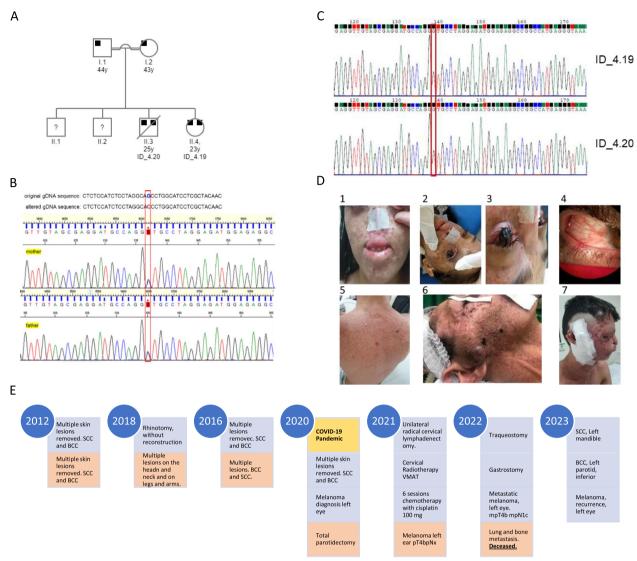


Fig. 1 A Pedigree of the family showing consanguineous marriage. Both parents (I.1 and I.2) are heterozygous for NM_000107.3:c.1027G > C and patient ID_4.19 (II.4) and patient ID_4.20 (II.3) are homozygous for this variant. The two older siblings are asymptomatic and were not tested.

B Reverse sequence electropherogram from mother and father of siblings. The NM_000107.3:c.1027G > C variant is highlighted in red. C Reverse sequence electropherogram from ID_4.19 and ID_4.20. D Photos to evidence severe phenotype. Patient ID_4.19 1. Photo from exeresis to remove lesions in lips, face and nose. 2 and 3. Photo of the nodular, pigmented and elevated lesion of the LE, extending throughout the cornea. 4. Photo showing the melanoma of the RE. Patient ID_4.20 5. Photo showing lesions that were removed and pigmentation on the back, exposed to sunlight. 6. Photo from exeresis of the lesion located in the right parotid gland, before surgery and 7. After the total parotidectomy. E Milestones of patient's clinical history, Patient ID_4.19 is highlighted in blue and for Patient ID_4.20, in orange, unfortunately, he passed away on 12/06/2022

Table 1 Number of lesions according to carcinoma type Basal Cell Carcinoma (BCC) and Squamous cell carcinoma (SCC).

Patient ID	Current age (Years-old)	Age of 1st symptoms		SCC (N)	Melanoma age at onset
ID_4.19	23	13	18	19	20
ID_4.20	25	15	13	22	22

Diagnostic assessment: genetic analysis

The surgical lesion tissue of each sibling was stored in Allprotect[®] Tissue Reagent (Qiagen) and DNA was extracted with DNeasy Blood & Tissue Kit (Qiagen). WES was performed using the Agilent SureSelect v6 and BGISEQ500 sequence platform to the coverage 50X, the mutations were identified using GATK4 according to the GATK Best Practices Protocols [8].

The BCFtools/RoH command [9] was used to detect runs of homozygosity (RoH) in sequencing data using Markov model (parameters: minimum length 4 Mb; minimum number of consecutive markers = 10). DNA was extracted from saliva from the parents. For Sanger sequencing, we used 100 ng of DNA and XP-E primers with the PlatinumTM PCR SuperMix High Fidelity from Thermo Fisher, according to the manufacturer. The left primer starts at position 49 and the right primer at position 232, considering the version of the genome GRCh37.p13. The primers sequences are Forward,

GGGAGGCCAGCTAGGGATTA and Reverse, CTC ACCGAACTGATGCCAGA.

A region with 123 Mb in runs of homozygosity (RoH) was identified, which corresponds to 4% of the genome and confirms consanguinity (Fig. 2A). A 25 Mb region was on chr11 36 Mb -61 Mb where DDB2 gene (NM_000107) is located (chr11:47,236,525 -47,260,768). The quality score of the variant is high: 4656.03 and the coverage for ID_4.19 was 127x and for ID_4.20, 38x. In both patients, we found the variant in DDB2 c.1027G > C, at position chr11:47259391 in

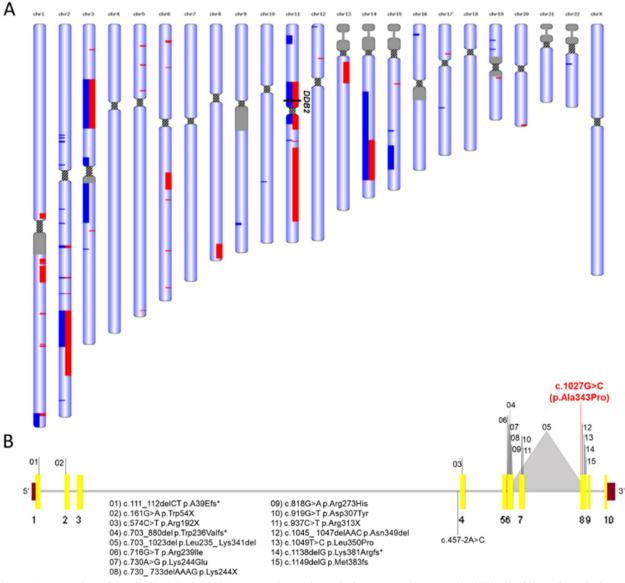


Fig. 2 Genetic analysis of the WES. **A** XP-E patients ROH regions detected at each chromosome. The regions highlighted in blue and in red refer to ROH in Patient ID_4.20 and Patient ID_4.19, respectively. **B** *DDB2* gene schematic representation and pathogenic variants and VUS described in databases. The exons are in yellow, all variants submitted at ClinVar and described in literature are listed (reviewed by Yang and coworkers, 2020 [26] and the variant detected in the two siblings is highlighted in red

exon 8 resulting in p.A343P amino acid change, therefore NM_000107.3:c.1027G>C. DDB2 homozygous missense variants are known to cause Xeroderma Pigmentosum group E (MIM: 278,740). This variant was not found in the general population (1000Genomes [10], dbSNP151 [11], GNOMAD [12]) or in diseaseassociated database CLINVAR [13], LOVD [14]. Both patients carry a homozygous variant and the same variant was detected in heterozygosity in their parents by Sanger sequencing (Fig. 1A and B). The function prediction analysis to identify disease-relevant nonsynonymous Single-Nucleotide Variants was performed using the MCAP [15] and REVEL [16] tools which rendered the scores 0.07 and 0.26 respectively, suggesting that the variant may be pathogenic, but the pathogenicity score is not high. According to a decision tree model that measures and compares the accuracy with four known mutation predictors and seventeen supervised machine-learning algorithms, the variant NM_000107.3:c.1027G > C is pathogenic [17]. Additionally, this G in position c.1027, located in exon 8, is highly conserved among humans and other hominids (Altai Neandertal G/G; Denisova G/G; Vindijia Neandertal G/G). According to the ACMG (American College of Medical Genetics) and to the AMP (Association for Molecular Pathology) guidelines [18], variant segregation with diseases in multiple family members supports evidence of pathogenicity when a gene is known to be associated with the disorder. WES data is available from the corresponding author after reasonable request due to the privacy of patient information data.

Discussion and conclusions

In this paper, we present two XP-E patients from a Brazilian consanguineous family affected with early-onset multiple skin lesions and malignant melanoma. XP-E is conventionally described as a mild XP phenotype, with residual levels of photoproducts DNA repair (between 50 and 100%), which makes the diagnosis difficult and generally, they are not diagnosed until adulthood [19, 20]. The reason for the phenotypic variations in XP-E patients may depend on environmental exposure and lifestyle. During the undiagnosed period, they accumulate large quantities of pre-carcinogenic lesions, and as a consequence, they may develop hundreds of skin cancers [21]. Unfortunately, the COVID-19 pandemic has difficult access to healthcare [22], especially in vulnerable rural areas, resulting in the evolution of the lesion into melanoma.

The parents are heterozygous and asymptomatic and 2 siblings from 4 are homozygous for the variant c.1027G > C (p.A343P), in *DDB2* gene. This variant was not found in the general population from

public databases. Despite the novel variant pathogenicity score is not extremely high, the position of modification seems to be extremely important to DDB2 DNA-binding function. DDB2 is ubiquitously present in human tissues and localizes ahead of XP-C to repair CPD and 6-4PP lesions in DNA [6]. One significant challenge we faced is the limited data of underrepresented populations in genomic databases and this continuous lack of diversity poses unique difficulties when it comes to accurately determining the pathogenicity of genomic variants. Genomic variant pathogenicity is typically evaluated by comparing the observed variants to known variants cataloged in databases. These databases primarily consist of genomic data from individuals of European ancestry, with a disproportionately smaller representation of other ethnic groups. As a result, the majority of our current understanding of variant pathogenicity is biased towards European populations. Without an adequate representation of diverse populations, we may miss crucial insights into variant pathogenicity and fail to provide accurate diagnoses, risk assessments, and tailored treatment plans for individuals from underrepresented backgrounds.

In a study with 89 XP patients, it was observed a specific propensity to ocular problems in XP-C group compared with XP-E and XP-V patients, while these two last groups had more skin cancers [23]. The patient ID_4.19 presented lesions in both eyes, with nodular melanoma in LE cornea, and residual/recurrent disease after one year. The etiology of ocular problems in XP-E patients is not well known since there are few reports.

This report presents the consequence of high UV light incidence in the XP-E outcome in patients living in a rural area in a tropical country. In the region Serido, UVA/UVB incidence is extremely high, therefore increasing the frequency of pyrimidine dimers [24, 25], in addition to this environmental risk, these patients lost follow-up during the COVID-19 pandemic, these two factors can explain, respectively, the anticipation and severity of the clinical manifestation observed in these 2 patients, with melanoma diagnosis at 20 and 22 years.

Abbreviations

ACMG

1000Genomes The 1000 Genomes Project

American College of Medical Genetics and Genomics

AMP Association for Molecular Pathology

BCC Basal Cell Carcinoma

CPD Cyclobutane Pyrimidine Dimers

dbSNP151The Single Nucleotide Polymorphism databaseDDB2Damage-specific DNA-binding protein 2GG-NERGlobal Genomic Nucleotide Excision Repair

GNOMAD Genome Aggregation Database

Mb Megabase

CT Computer Tomography
ICU Intensive Care Unit

.OVD Leiden Open Variation Database

ROH Runs of homozygosity
SCC Squamous Cell Carcinoma
TIL Tumor-infiltrating Lymphocytes

UVA Ultraviolet A UVB Ultraviolet B

XP Xeroderma pigmentosum WES Whole Exome Sequencing

6-4PP 6-4 pyrimidine-pyrimidone photoproducts

Acknowledgements

We want to express our heartfelt gratitude for this family invaluable contribution to science and their willingness to allow us to share and learn from their clinical journey has been instrumental to advance medical knowledge and improve patient care. Their courage, trust, and generosity in sharing their personal experiences have made a significant impact on our research and will undoubtedly benefit countless individuals facing similar challenges. By offering insights into their journey, they have not only helped us gain a deeper understanding of this condition but also provided hope and inspiration to others who may be on a similar path.

Authors' contributions

ARST did the acquisition, analysis, and interpretation of data and has worked on the original draft; ICPA is the head and neck surgeon treating patient ID 4.19, she helped with the acquisition, analysis, interpretation of data, article writing and discussion. She shares the first authorship with ARST; AAY is the bioinformatician scientist responsible for the WES analysis and data interpretation, he helped with article writing and discussion; SRMH is the head and neck surgeon treating patient ID_4.20, she helped with the acquisition, analysis, and interpretation of data; PSS is the ophthalmologist responsible for both patient, he helped with the acquisition, analysis, and interpretation of data; FR performed Sanger sequencing for variant confirmation in the siblings; SN performed WES analysis, interpretation and payment, he helped writing the original manuscript and proofs; TBP is the geneticist following the patients and she contributed with conception, design of the work, acquisition, analysis, and interpretation of data, writing original and revised version of the manuscript. SN and TBP share the last authorship. Authors read and approved the manuscript.

Funding

TBP received funding from the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), for fellowships and laboratory consumables. SN received funding from the Fondation Gustave Roussy for the molecular analysis. The funding bodies played no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request. Datasets will be publicly freely available at the time the primary manuscript is accepted for publication.

Declarations

Ethics approval and consent to participate

This study was approved by the Ethics Committee of Liga contra o Cancer Hospital and Universidade de Sao Paulo, and written informed consent was obtained from all participants (CEP/ICB/USP, 48347515.3.0000.5467).

Consent for publication

Written informed consent was obtained from the patient's parents for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

Competing interests

The authors declare that they have no competing interests.

Received: 4 October 2022 Accepted: 31 July 2023 Published online: 12 August 2023

References

- Kraemer KH, Digiovanna JJ. Forty years of research on xeroderma pigmentosum at the US National Institutes of Health. Photochem Photobiol. 2015;91(2):452–9.
- Moriwaki S, Kanda F, Hayashi M, Yamashita D, Sakai Y, Nishigori C. Xeroderma pigmentosum clinical practice guidelines. J Dermatol. 2017;44(10):1087–96.
- Masutani C, Kusumoto R, Yamada A, Dohmae N, Yokoi M, Yuasa M, et al. The XPV (xeroderma pigmentosum variant) gene encodes human DNA polymerase eta [see comments]. Nature. 1999;399:700–4.
- Scrima A, Koníčková R, Czyzewski BK, Kawasaki Y, Jeffrey PD, Groisman R, et al. Structural basis of UV DNA-Damage recognition by the DDB1-DDB2 complex. Cell. 2008;135(7):1213–23.
- Oh KS, Emmert S, Tamura D, Digiovanna JJ, Kraemer KH. Multiple skin cancers in adults with mutations in the XP-E (DDB2) DNA repair gene. J Invest Dermatol. 2011;131(3):785–8.
- Wittschieben B, Iwai S, Wood RD. DDB1-DDB2 (xeroderma pigmentosum group E) protein complex recognizes a cyclobutane pyrimidine dimer, mismatches, apurinic/apyrimidinic sites, and compound in DNA. J Biol Chem. 2005;280(48):39982–9.
- 7. Freire-Maia N. Genetic effects in brazilian populations due to consanguineous marriages. Am J Med Genet. 1990;35:115–7.
- 8. Van der Auwera GA, Carneiro MO, Hartl C, Poplin R, Del Angel G, Levy-Moonshine A et al. From FastQ data to high confidence variant calls: the Genome Analysis Toolkit best practices pipeline. Curr Protoc Bioinforma. 2013;43(1110):11.10.1–11.10.33.
- Narasimhan V, Danecek P, Scally A, Xue Y, Tyler-Smith C, Durbin R. BCFtools/RoH: a hidden Markov model approach for detecting autozygosity from next-generation sequencing data. Bioinforma Oxf Engl. 2016;32(11):1749–51.
- The 1000 Genomes Project Consortium, Auton A, Abecasis GR, Steering committee, Altshuler DM, et al. A global reference for human genetic variation. Nature. 2015;526(7571):68–74. PMID: 26826718.
- Sherry ST, Ward MH, Kholodov M, Baker J, Phan L, Smigielski EM, et al. dbSNP: the NCBI database of genetic variation. Nucleic Acids Res. 2001;29(1):308–11.
- 12. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. Nature. 2020;581(7809):434–43.
- Landrum MJ, Lee JM, Benson M, Brown GR, Chao C, Chitipiralla S, et al. ClinVar: improving access to variant interpretations and supporting evidence. Nucleic Acids Res. 2018;46(D1):D1062–7.
- Fokkema IFAC, Taschner PEM, Schaafsma GCP, Celli J, Laros JFJ, den Dunnen JT. LOVD v.2.0: the next generation in gene variant databases. Hum Mutat. 2011;32(5):557–63.
- Jagadeesh KA, Wenger AM, Berger MJ, Guturu H, Stenson PD, Cooper DN, et al. M-CAP eliminates a majority of variants of uncertain significance in clinical exomes at high sensitivity. Nat Genet. 2016;48(12):1581–6.
- Ioannidis NM, Rothstein JH, Pejaver V, Middha S, McDonnell SK, Baheti S, et al. REVEL: an Ensemble Method for Predicting the pathogenicity of rare missense variants. Am J Hum Genet. 2016;99(4):877–85.
- Do Nascimento PMH, Medeiros IG, Falcão RM, Stransky B, De Souza JES. A decision tree to improve identification of pathogenic mutations in clinical practice. BMC Med Inform Decis Mak. 2020. PMID: 32151256.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med Off J Am Coll Med Genet. 2015;17(5):405–24.
- Menck CF, Munford V. DNA repair diseases: what do they tell us about cancer and aging? Genet Mol Biol. 2014;37(1 Suppl):220–33.
- Cleaver J. Common pathways for ultraviolet skin carcinogenesis in the repair and replication defective groups of xeroderma pigmentosum. J Dermatol Sci. 2000;23(1):1–11.
- 21. Yurchenko AA, Rajabi F, Braz-Petta T, Fassihi H, Lehmann A, Nishigori C, et al. Genomic mutation landscape of skin cancers from DNA repair-deficient xeroderma pigmentosum patients. Nat Commun. 2023;14(1):2561.
- 22. Bigoni A, Malik AM, Tasca R, Carrera MBM, Schiesari LMC, Gambardella DD, et al. Brazil's health system functionality amidst of the COVID-19 pandemic: an analysis of resilience. Lancet Reg Health Am. 2022;10:100222.

- Fassihi H, Sethi M, Fawcett H, Wing J, Chandler N, Mohammed S, et al. Deep phenotyping of 89 xeroderma pigmentosum patients reveals unexpected heterogeneity dependent on the precise molecular defect. Proc Natl Acad Sci U S A; 2016. PMID: 26884178.
- Schuch AP, Moreno NC, Schuch NJ, Menck CFM, Garcia CCM. Sunlight damage to cellular DNA: focus on oxidatively generated lesions. Free Radic Biol Med. 2017. PMID: 28109890.
- Santiago KM, Castro LP, Neto JPD, de Nóbrega AF, Pinto CAL, Ashton-Prolla P, et al. Comprehensive germline mutation analysis and clinical profile in a large cohort of brazilian Xeroderma pigmentosum patients. J Eur Acad Dermatol Venereol. 2020. PMID: 32228487.
- 26. Yang R, Kong Q, Duan Y, Li W, Sang H. Identification of a novel DDB2 mutation in a chinese Han family with Xeroderma pigmentosum group E:a case report and literature review. BMC Med Genet. 2020;21(1):67.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

