



# The role of the atopy patch test in the diagnostic work-up of non-IgE gastrointestinal food allergy in children: a systematic review

Barbara Cuomo<sup>1</sup> · Caterina Anania<sup>2</sup> · Enza D'Auria<sup>3</sup> · Fabio Decimo<sup>4</sup> · Giovanni Cosimo Indirli<sup>5</sup> · Enrica Manca<sup>6</sup> · Gian Luigi Marseglia<sup>7</sup> · Violetta Mastroianni<sup>8</sup> · Valentina Panetta<sup>9</sup> · Angelica Santoro<sup>10</sup> · Marco Ugo Andrea Sartorio<sup>11</sup> · Elisabetta Veronelli<sup>12</sup> · Mauro Calvani<sup>13</sup>

Received: 7 December 2022 / Revised: 16 April 2023 / Accepted: 19 April 2023 / Published online: 30 May 2023  
© The Author(s) 2023

## Abstract

The “Atopy Patch Test” (APT) has been proposed as a diagnostic tool for food allergies (FA), especially in children with FA-related gastrointestinal symptoms. However, its diagnostic accuracy is debated, and its usefulness is controversial. The aim of this systematic review was to evaluate the APT diagnostic accuracy compared with the diagnostic gold standard, i.e., the oral food challenge (OFC), in children affected by non-IgE mediated gastrointestinal food allergies, including the evaluation in milk allergic subgroup. Both classical non-IgE mediated clinical pictures and food induced motility disorders (FPIMD) were considered. The search was conducted in PubMed and Scopus from January 2000 to June 2022 by two independent researchers. The patient, intervention, comparators, outcome, and study design approach (PICOS) format was used for developing key questions, to address the APT diagnostic accuracy compared with the oral food challenge (OFC). The quality of the studies was assessed by the QUADAS-2 system. The meta-analysis was performed to calculate the pooled sensitivity, specificity, DOR (diagnostic odds ratio), PLR (positive likelihood ratio), and NLR (negative likelihood ratio) with their 95% confidence intervals (CI). Out of the 457 citations initially identified via the search (196 on PubMed and 261 on Scopus), 37 advanced to full-text screening, and 16 studies were identified to be included in the systematic review. Reference lists from relevant retrievals were searched, and one additional article was added. Finally, 17 studies were included in the systematic review. The analysis showed that APT has a high specificity of 94% (95%CI: 0.88–0.97) in the group of patients affected by FPIMD. Data showed a high pooled specificity of 96% (95% CI: 0.89–0.98) and the highest accuracy of APT in patients affected by cow’s milk allergy (AUC = 0.93).

**Conclusion:** APT is effective in identifying causative food in children with food-induced motility disorders.

## What is Known:

- Atopy patch test could be a useful diagnostic test for diagnosing food allergy, especially in children with food allergy-related gastrointestinal symptoms.

## What is New:

- Atopy patch test may be a useful tool in diagnosing non IgE food allergy, especially in children with food-induced gastrointestinal motility disorders and cow's milk allergy.

**Keywords** Atopy patch test · Food allergy · Non-IgE mediated food-allergy · Motility disorders · Meta-analysis

## Abbreviations

AEG Allergic eosinophilic gastroenteritis  
APT Atopy patch test

AUC Area under the curve  
CI Confidence intervals  
DOR Diagnostic odds ratio  
EoE Eosinophilic esophagitis  
EC Eosinophilic colitis  
EGIDs Eosinophilic gastrointestinal disorders  
FA Food allergies

Communicated by Tobias Tenenbaum.

✉ Enza D'Auria  
enza.dauria@unimi.it

Extended author information available on the last page of the article

FPIAP	Protein-induced allergic proctocolitis
FPE	Food protein-induced enteropathy
FPIMD	Food induced motility disorders
FPIES	Food protein-induced enterocolitis syndrome
LR	Likelihood ratio
NLR	Negative likelihood ratio
non-IgE-GI food allergy	Non-IgE mediated gastrointestinal food allergy
NPV	Negative predictive value
OFC	Oral food challenge
PICOS	Patient intervention comparators outcome and study design
PLR	Positive likelihood ratio
PPV	Positive predictive value
QUADAS-2	Quality assessment of diagnostic accuracy studies 2
sIgE	Specific IgE
SPT	Skin prick tests
SROC	Summary receiver operating characteristic curve

## Introduction

Non-IgE mediated gastrointestinal (non-IgE-GI) food-induced allergic disorders encompass numerous and different clinical pictures. Some of these are well-characterized, such as Food Protein-Induced Allergic Proctocolitis (FPIAP), Food Protein-Induced Enterocolitis Syndrome (FPIES), Food Protein-Induced Enteropathy Syndrome (FPE), and Eosinophilic Gastrointestinal Disorders (EGIDs) (including Eosinophilic Esophagitis (EoE), Allergic Eosinophilic Gastroenteritis (AEG), and Eosinophilic Colitis (EC)) [1–3]. Others, especially in the first age of life, may present with less specific symptoms such as acute abdominal discomfort, persistent crying and unsettled behavior, frequent regurgitation or vomiting, and persistent watery diarrhea, often in combination with poor growth or constipation [4]. These latter were recently defined as Food Induced Motility Disorders (FPIMD) [4, 5], meaning all entities not included in the above-mentioned classical non-IgE mediated allergy, which improve after dietary elimination of specific food proteins and which motility alteration has been hypothesized, although the exact pathogenetic mechanisms remain largely unknown [6, 7]. FPIMD are included in the group of non-IgE-GI food allergy, as sIgE for foods are not detected in most cases [8].

Diagnosis for non-IgE-GI food allergy is usually based on clinical features by recovery after dietetic therapy and subsequent positivity challenge test. The process is not supported by classical diagnostic tests like skin prick tests (SPT) and serum-specific IgE (sIgE), which are often negative. For these reasons, the APT has

been proposed as a tool in the diagnostic work-up. A positive reaction correlates with infiltrating allergen-specific Th2 cells which secrete interleukin 4 and 13 already 24 h after application of the allergen [9], after 48 h a shift towards a Th1 pattern with the secretion of interferon gamma [10] underlining the role of APT in delayed reaction type IV, rather than immediate type I reaction.

APT are detected as positive and are mainly useful in delayed/mixed reactions (non-IgE gastrointestinal FA, atopic dermatitis, EoE) rather than IgE mediated FA. However, its diagnostic accuracy remains controversial, and it is not routinely recommended because of the lack of a standardized process and the wide variability in sensitivity and specificity of results in previous studies [11, 12].

Although most studies analyzed APT in groups of patients affected by both immediate and delayed allergic reactions [13–15], recent evidence [16] suggests an increased APT diagnostic efficiency if employed in better-selected cohorts.

Two systematic reviews [17, 18] analyzed the accuracy of APT in patients affected by FA. However, the metaanalysis by Luo et al. considered studies including children with different types of food allergies and, in some cases, with atopic dermatitis [17]. The second one [18] provides few informations and does not allow to analyse data from included studies.

The aim of this systematic review was to evaluate the diagnostic accuracy of the APT compared with the diagnostic gold standard, i.e., the oral food challenge (OFC), in children living with non-IgE-GI food allergy, including the evaluation in the milk allergic subgroup.

## Methods

### Search strategy

A comprehensive search was conducted in Medline via PubMed and Scopus (from January 1, 2000, through June 30, 2022), using the keywords “food allergy” and (“patch test” or “atopic patch test”) and (“Food protein-induced enterocolitis syndrome” or “FPIES” or “enterocolitis”), (“Eosinophilic Esophagitis” or “Eosinophilic Colitis” or “Eosinophilic Gastroenteritis”), (“enteropathy” or FPE), (“proctocolitis” or FPIAP), “haematochezia,” “colitis,” “gastritis,” “rectal bleeding,” “failure to thrive,” (“stypsis” or “constipation”).

Two independent researchers (M.U.A.S. and E.M.) screened the databases. The references were imported into a citation manager software (EndNote 20.2.1<sup>®</sup>) for initial duplicate removal.

They independently screened the search string, reviewed all abstracts, and agreed on which full-text articles to retrieve to assess for potentially eligible studies. Disagreements were resolved through discussion, and, if required, in cases of incongruence, a third reviewer (B.C.) was responsible for mediating a discussion and consequent decisions. The systematic review was based on the PRISMA (preferred reported items for systematic

reviews and meta-analyses) guidelines, and its protocol was registered in the PROSPERO database. The authors had no conflicts of interest, and the study did not receive any funding.

## Eligibility criteria

We developed a PICOS (patient, intervention, comparators, outcome, and study design) approach to formulate the eligibility criteria for the studies. The following question was set: “Are APT as accurate as OFC for non-IgE-GI food allergy in children?” We consider any kind of OFC, both in open and single- or double-blind form. The studies were not restricted to English-language publications, publication type, or study design; however, they were limited to children (0 to 18 years).

We included only studies that allow us to evaluate the diagnostic performance of APT compared to the gold standard for diagnosing food allergy, i.e., OFC in children affected by FPIAP, FPIES, FPE, EGIDs, or FPIMD. Studies were also included if it could be possible to extract data, and if necessary, additional explanations by contacting the authors were requested.

Studies were excluded if the information was not specific to the topic of this review or if the clinical diagnosis was made without the confirmation by OFC, or if data about specificity and/or sensitivity was not provided or impossible to extract.

## Data collection and analysis

Two authors independently retrieved and reviewed the following data (if available) from all included studies: year of publication, first author, design, population size, mean age, type of symptoms, number of allergic subjects, number of patients with positive sIgE or SPT, APT methods used, and data of its accuracy.

Therefore, eligible studies were classified into two categories: (a) studies considering patients living with classic well-defined clinical pictures like FPIAP, FPIES, FPE, and EDGs; and (b) studies including patients living with FPIMD.

Studies were widely discussed in detail and evaluated by all authors in a standardized and independent manner; the methodological quality was evaluated according to criteria proposed by the Quality Assessment of Diagnostic Accuracy Studies 2 (QUADAS-2) tool [19]. At the same time, any divergence was resolved by discussion and agreement among all reviewers. This instrument judges the risk of bias and accessibility from diagnostic accuracy studies. QUADAS-2 contains four key domains (patient selection, index test, reference standard, and flow and timing), and each domain is rated as low, high, and unclear.

## Statistical analysis

Meta-analysis was performed with the midas command in Stata 16.1. Pooled sensitivity, specificity, positive likelihood ratio (PLR), negative likelihood ratio (NLR), and diagnostic

odds ratio (DOR) with their 95% confidence (95%CI) were calculated by a bivariate mixed-effect regression model. The area under the curve (AUC) and relative 95% CI was also calculated. The AUC of summary receiver operating characteristic curve (SROC) results were considered low ( $0.5 > = AUC < = 0.7$ ), moderate ( $0.7 > AUC < = 0.9$ ), or high ( $0.9 > AUC < = 1$ ). The test could be considered highly informative with PLR exceeding 10.0 and NLR below 0.1; moderately informative with PLR values of 5–10 and NLR of 0.1–0.2; or very lowly informative with LR of 2–5 and 0.2–0.5. The  $I^2$  statistic was used to evaluate the heterogeneity between studies; a value of 0% indicates no observed heterogeneity, while values greater than 50% indicate substantial heterogeneity [20].

## Results

The selection and inclusion process of the studies is reported in the PRISMA Statement Flowchart (Fig. 1). The electronic database search identified 196 citations on PubMed and 261 on Scopus. After analyzing titles and abstracts, respectively, 39 and 16 articles remain. After removing duplicates (3 papers), 52 articles were identified, and the full text was assessed for eligibility. A total of 16 articles were then selected, and the other ones were refused for showing data that did not meet the planned inclusion criteria (Fig. 1). Evaluating the most relevant studies' references allowed us to detect one additional article. In total, we included 17 studies in this review.

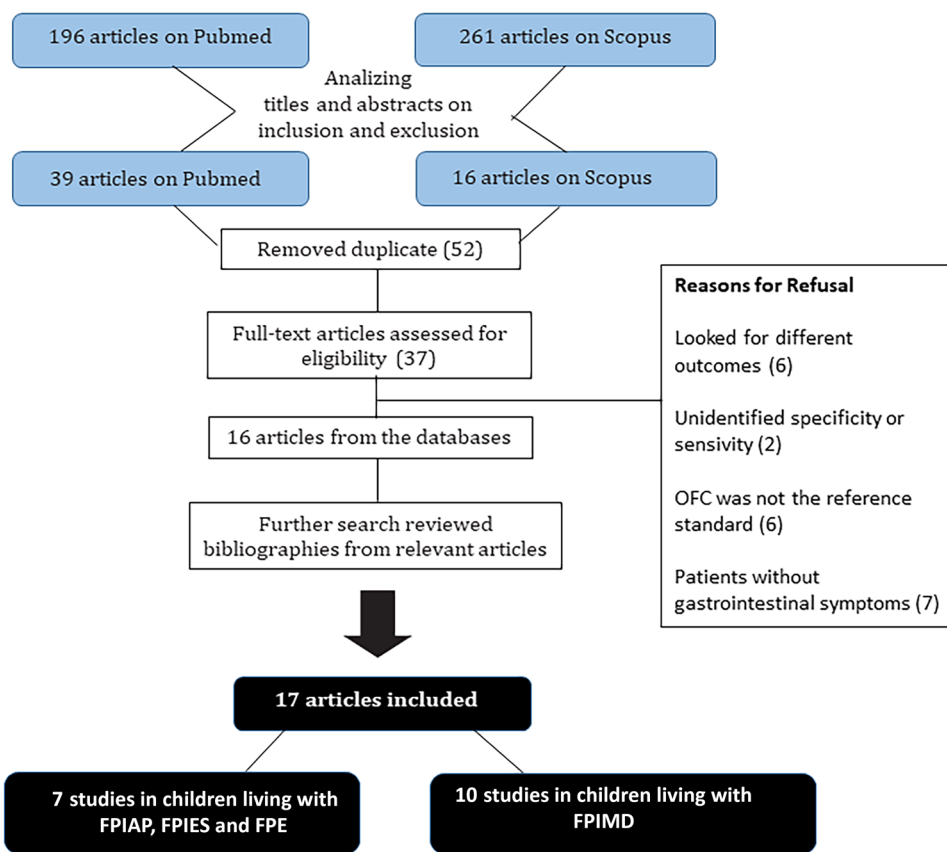
We found three studies evaluating children affected by FPIES [21–23] (Table 1); three studies included patients with FPIAP [24–26] (Table 2), and one included patients affected by FPIAP, FPIES, and FPE [27] (Table 3). Ten studies included [16, 28–36] patients affected by FPIMD (Tables 3 and 4).

Regarding children living with FPIES, reports were conducted on small patient populations (overall 52 patients) and showed very different results: sensitivity ranged from 11.8 to 89%, while specificity ranged from 85.7 to 100%.

One retrospective study allowed to extract data on only eight patients [21]. Two studies [21, 23] received a lower evaluation on Quadas-2, mainly on those domains concerning patients' selection. Specifically, the increased risk of bias was based on the exclusive enrollment of patients not representative of the general population as they were already known to be affected by FA when performing OFC and APT. The only prospective study [22] is qualitatively better than the previous and described high values of specificity and PPV (100%) with lower sensitivity and NPV (respectively 76.2% and 70.6%).

Three studies included children with proctocolitis. Based on QUADAS-2, two of them [24, 26] were low-quality

**Fig. 1** Studies search flow diagram. The electronic databases search identified 196 citations on PubMed and 261 on Scopus. After analyzing titles and abstracts, duplicates were removed and full text were individualized and assessed for eligibility. 16 articles were selected after other ones looked like they had different outcomes or included patients without gastrointestinal symptoms, not comparing to the diagnostic gold standard OFC or not identifying specificity or sensitivity. One additional paper was added from reviewing relevant articles. 17 articles were included in this study



**Table 1** Studies including patients affected by FPIES

	STUDY design	Child's AGE	ALLERGY	Positive subjects IgE/SPT	Allergy prevalence	DIAGNOSYS	APT		PPV	specificity	NPV	sensitivity	QUADAS-2
Zapatero <sup>17</sup> 2005 N:8 <sup>§</sup>	retrospective	9-12 months	FPIES	1	100%	Open challenge	Fish Cooked food and extract	Curates strips removed 48 h reading 48 -96h				37.5%	
Fogg <sup>18</sup> 2006 N: 19 (N:33 OFC)	prospective	5 - 30 months (mean 15.6 m)	FPIES	0	36.8%	Open challenge	Cow's milk, egg, wheat, soy, rice, oat. Not specified if fresh or extract (OFC N 33)	12mm Finn chambers Removed 48 h reading 72h	100%	100%	70.6%	76.2%	
							Cow's milk (OFC N 13)		100%	100%	80%	89%	
Järvinen <sup>19</sup> 2012 N: 25 (N:38 OFC)	unclear	1.5 – 16.8 years (mean 3.3 y)	FPIES	unspecified	100% (8% at the revaluation)	Open challenge	Cow's milk, soy, rice, oat. Not specified if fresh or extract.	unspecified Finn chambers size removed 48 h reading 48 -72h.	40%	85.7%	54.5%	11.8%	

APT atopy patch test; PPV positive predictive value; NPV negative predictive value; OFC oral food challenge

<sup>§</sup> data extract from subjects in which APT were performed

**Table 2** Studies including patients affected by FPIAP

	STUDY design	Child's AGE	ALLERGY	Positive subjects IgE/SPT	Allergy prevalence	DIAGNOSYS	APT		PPV	specificity		NPV	sensitivity	QUADAS-2
<b>Lucarelli<sup>20</sup></b> 2011 N:14	prospective	15 days to 6 months	proctocolitis	0	100%	Open challenge and endoscopy with biopsies.	Cow's milk, soy, rice, wheat, egg, breast milk not specified. Fresh food or dry (not clearly defined)	unspecified Finn chambers size removed 48h reading 48-72h	42.8%	0%			100%	
<b>Alves<sup>21</sup></b> 2015 N: 31	unclear	mean 97.0 days	proctocolitis	unspecified	19.3%	Open challenge	Cow's milk Fresh food	8-mm Finn chambers removed 48 h reading 72h		100%		80.7%	0%	
<b>Arshi<sup>22</sup></b> 2021 N: 28	unclear	mean 5.25 months	proctocolitis	unspecified	42.8%	Open challenge	Fresh food (Cow's milk, egg, peanuts, soybeans, rice, wheat, corn, sesame, red meat)	12-mm Finn chambers removed 48 h reading 48h	52.17%					

studies because the diagnosis were not all confirmed with OFC or apply to a restricted group of selected patients with severe clinical forms not responsive to therapy. The third [25] is qualitatively better than the previous and shows high specificity values (100%) and an NPV of 80.7%.

The only study [27] that enrolled a mixed population affected by FPIAP, FPIES, and FPE is a retrospective one of good quality. It found high values of specificity and PPV for APT performed with fresh foods regardless of which was tested (the best with milk, respectively 100% and 100%, and eggs, 90.9% and 80%). On the contrary, sensitivity values are low (9.1% for milk and 40.4% for egg).

We identified ten studies performed on patients living with FPIMD. These studies have enrolled a large number (n. 770) of children, which were divided into two groups (Tables 4 and 5) to permit separate evaluation of the results of APT diagnostic performance in patients with negative allergy tests and those with mixed positive and negative sIgE-associated forms. Two [28, 30] of these

studies allowed data extraction and were included in each group. Thus, three studies included 320 children affected by FPIMD without specific IgE and a negative SPT (Table 4). Nine studies included 598 children affected by FPIMD with or without specific IgE and positive or negative SPT (Table 5). Nocerino's study [29] was selected for the FPIMD group even if it included a few enterocolitis and enteropathy allergic patients (Table 4).

Studies that evaluated APT diagnostic accuracy in patients with negative sIgE showed good values of specificity and PPV, respectively, of 88.3–100% and 82.8–100% but poor sensitivity (40–80.9%) (Table 4). However, their quality is poor for enrolling patients not representative of the general population as they were already allergic known persons at the beginning of the study and/or were affected by severe clinical forms.

Data analysis from the second group of FPIMD studies showed that APT has high specificity and PPV regardless of sIgE and/or SPT positivity. In this subgroup,

**Table 3** Studies including patients affected by more than one form among FPIAP, FPIES, and FPE

	STUDY design	child's AGE	ALLERGY	Positive subjects IgE/SPT	Allergy prevalence	DIAGNOSYS	APT		PPV		specificity		NPV		sensitivity		QUADAS-2
<b>Sirin Kose<sup>23</sup></b> 2020 N:61&	retrospective	6 - 12.5 months (mean 8 m)	FPIAP FPIES FPE	6 SPT <sup>§</sup> 21 IgE <sup>§</sup>	50.8%	Open challenge	Milk, egg Fresh food	64 mm IQ-UL chambers removed 48 h reading 48-72h	milk 100%	egg 80%	milk 100%	egg 90.9%	milk 48.7%	egg 66.7%	milk 9.1%	egg 44.4%	

APT atopy patch test; PPV positive predictive value; NPV negative predictive value; FPIAP food proteins induced allergic proctocolitis; FPIES food proteins induced enterocolitis syndrome; FPE food proteins enteropathy

& data extract from subjects affected by GI symptoms

§ data from the entire group enrolled, non extracted for those affected by gastrointestinal symptoms

**Table 4** Studies including patients affected by FPIMD without specific IgE and with negative SPT

	STUDY Desygn	Child's AGE	ALLERGY	Positive subjects IgE/SPT	Allergy prevalence	DIAGNO SYS	APT		PPV	specificity	NPV	sensitivity	QUADAS-2
<b>De Boissieu<sup>24</sup> 2003</b> N:32 <sup>£</sup>	retrospective	2-57 months	Gastro - esophageal reflux, colics, diarrhea, constipation failure to thrive	0	65.6%	Open or double blinded challenge	Skimmed cow's milk	unspecified Finn chambers size removed 48 h reading 48-72h	94.40%	90.91%	71.43%	80.95%	
<b>Nocerino<sup>25</sup> 2013</b> N:172	prospective	2-12 months (mean 6.37 m)	Enterocolitis (41.9%), enteropathy (20.9%), gastro-esophageal reflux disease (37.2%)	0	65.7%	DBPCFC	Cow's milk Fresh food	8-mm aluminium cup removed 48 h reading 48-72h	82.81%	88.30%	76.85%	67.95%	
<b>Yukselen<sup>26</sup> 2016</b> N: 116 <sup>£</sup>	unclear	3-60 months	Gastro-esophageal reflux	0	25.8%	Open or blinded challenge and upper endoscopy + scintigraphic evaluation or 24 h esophageal pH	Milk (not specified if fresh or extract)	8 - 12mm Finn chambers removed 48h reading 72h	100%	100 %	82.69%	40%	

APT atopy patch test; PPV positive predictive value; NPV negative predictive value; DBPCFC double blind placebo controlled food challenge  
 £ data extract from subjects without specific IgE and with negative SPT

prospective studies received better QUADAS-2 evaluation and reached specificity and PPV values of 95–100% (range values of 63.6–100% and 66.7–100%, respectively).

Canani [16] showed better data when performing APT with fresh food than with food extracts; Alves [25] and Sirin Kose [27] et al. obtained 100% specificity using fresh milk, while most of the other studies do not clearly declare which type of allergenic material had used.

## Methodological quality

### QUADAS-2

According to the QUADAS-2 criteria, studies enrolling patients with well-defined gastrointestinal clinical pictures (FPIAP, FPIES, and FPE) and those that included patients affected by non-sIgE FPIMD (Tables 1, 2, 3, and 4) resulted in both in lower judgments on QUADAS-2 evaluation, particularly in the domain dedicated to the selection of enrolled patients (Fig. 2). In most cases, children were not representative of the general population as they were already allergic and/or were affected by severe forms. Since inappropriate exclusions may result in over-optimistic estimates or in underestimation of diagnostic

accuracy, the risk of bias was judged as high. The same occurs if APT results were not collected at an appropriate time interval, ideally at the same time as OFC, or if not all the patients receive a diagnosis by positive objective signs at OFC. Studies that enroll subjects with FPIMD both with and without specific IgE (Table 5) appear to be of higher quality.

In general, very few studies have been designed on the specific question of this review [28, 29, 32, 33, 35, 36].

## Meta-analysis results

Due to the small sample size or lack of necessary data in the studies enrolling children affected by FPIAP, FPIES, and FPE, meta-analysis was conducted only in the group of patients with FPIMD. More analysis was also made in the subgroup of children with suspected milk allergy, the most frequent involved food allergen.

For FPIMD, a total of 8 studies analyzed 491 patients. Overall results of the meta-analysis show that APT has high specificity 94% (95% CI: 0.88–0.97) moderate positive likelihood ratio (PLR 8.3 95% CI: 4.1–16.6) and a low negative likelihood ratio (NLR 0.57 95% CI: 0.40–0.82), while sensitivity 46% (95% CI: 0.27–0.66) appears variable between studies. Two out of three studies (Canani et al. and

**Table 5** Studies patients affected by FPIMD with or without specific IgE and positive or negative SPT

	STUDY Desygn	Child's AGE	ALLERGY	Positive subjects IgE/SPT	Allergy prevalence	DIAGN OSYS	APT		PPV	specificity	NPV	sensitivity	QUADA S-2
<b>Kokkonen<sup>27</sup> 2001 N:84</b>	prospective	1.6–15 years (mean 7.9 y)	Recurrent abdominal pain, some with concomitant diarrhoea, constipation heart burn	13%	33%	Open challenge	Milk, egg, soy, gliadin, wheat, barley, rye, oats (not specified if fresh or extract)	unspecified Finn chambers size removed 48 h reading 48-72h	66.7%	87.5%	77.8%	50%	
<b>De Boissieu<sup>24</sup> 2003 N:35</b>	retrospective	2-57 months	Gastro-esophageal reflux, colics, diarrhea, constipation failure to thrive	8.6%	68.6%	Open or double blinded challenge	Skimmed cow's milk	unspecified Finn chambers size removed 48 h reading 48-72h	95.0%	90.9%	66.7%	79.2%	
<b>Kalach<sup>28</sup> 2005 N: 44<sup>8</sup> (OFC N 36)</b>	prospective	5-78 months (mean 34.3 +/- 7 m)	loose stools, colic, vomiting, gastroesophageal reflux, failure to thrive, all these with or without atopic dermatitis	67% data from OFC positive patients	58.9%	Open challenge	Formula Processed milk product mixed with milk	unspecified Finn chambers size removed 48 h reading 48-72h 25 mm Diallertest	100%	100%	46.2%	41.7%	
<b>Canani<sup>12</sup> 2007 N:60</b>	unclear	3-48 months (mean 23)	Vomiting, Chronic diarrhea, Hematochezia	53% SPT 40% IgE	58.3%	Open challenge	Milk (ml), egg (eg), wheat Fresh food (Fr) and extracts (Ex)	12 mm Finn chambers removed 48 h reading 48-72h (milk N 55 egg N28)	ml Fr 95.2% ml Ex 50% ml Fr 95.8% ml Ex 95.8% ml Fr 67.6% ml Ex 43.4% eg Fr 100% eg Ex 100% eg Fr 10% eg Ex 10% eg Fr 75% eg Ex 33.3% eg Fr 84.2% eg Ex 5.3%				
<b>Cudowska<sup>29</sup> 2010 N: 28</b>	prospective	6 to 144 months	abdominal pain, vomiting, diarrhoea, constipation perianal features, bleeding, loss of appetite, gastro-esophageal reflux, gastritis, ileitis	17.8% SPT 21.4% IgE	60.7%	Open challenge pH-metry monitoring Sigmoid oscopy	Cow's milk (not specified if fresh food or extract)	8 - 12 mm Finn chambers removed 48 h reading 48-72h	92.9%	90.9%	71.4%	76.5%	
<b>Syrgiou<sup>30</sup> 2011 N:48<sup>8</sup></b>	prospective	6 months to 14 years (mean 3.5Y)	Chronic constipation	27%	60.4%	Open challenge	Cow's milk, egg, wheat, rice, corn, potato, chicken, beef, soy, Fresh food	12 mm Finn chambers removed 48 h reading 48-72h	87.5%				
<b>Mowszet<sup>31</sup> 2014 N:61</b>	unclear	3-36 months (mean 13.5 m)	regurgitatio, vomiting, diarrhea, abdominal pain,bleedin, constipation failure to thrive, malnutrition	Milk 9.8% Wheat 6.5%	Milk 45% Wheat 4.7%	Open challenge (61 with milk, 19 with wheat)	Milk, wheat Fresh food	8 mm Finn chambers removed 48 h reading 48-72h	Milk 80.0% (N61) Wheat 100% (N19)	90.9%	39.2%	20.5%	
<b>Boonyaviwat<sup>32</sup> 2015 N:39 (N:41 OFC)</b>	prospective	2 months to 10 years (mean 2.42 Y)	Chronic diarrhoea, haematochezia, vomiting, abdominal pain	11/76 events (14.5%)	73.2%	Open challenge	milk, egg, wheat, soy, shrimp Lyophilized or commercial	12 mm Finn chambers removed 48 h reading 48-72h	Lyophilized 85.7% 63.6% 53.8% 80.0% Commercial 90.0% 90.9% 32.3% 30.0%				
<b>Yukselen<sup>26</sup> 2016 N: 151</b>	unclear	3-60 months	Gastro-esophageal reflux	27 (17.8%) SPT 32 (21%) IgE	43%	Open or blinded challenge and upper endoscopy + scintigraphy or 24 h esophageal pH	Milk (not specified if fresh or extract)	8 - 12mm Finn chambers removed 48h reading 72h	100%	100%	63.70%	24.62%	

APT atopy patch test; PPV positive predictive value; NPV negative predictive value

**Table 5** (continued)

§ data extract from children affected by GI-FA  
 % data extract from children in whom APT were performed  
 OFC Oral Food Challenge  
 esophageal pH esophageal pH-metry

Yukselen et al. [16, 30]) showing lower values for APT were performed with commercial extracts. The heterogeneity is high, with  $I^2$  always greater than 50%. The AUC value was moderate/high (0.90) with high corresponding DOR of 14 (95% CI: 6–34).

The subgroup analysis for milk-allergic patients included eight studies from all clinical groups, including 551 subjects. Seven out of ten studies included FPIMD. Data are like those seen for FPIMD and show even higher pooled specificity of 96% (95% CI: 0.89–0.98) and slightly better accuracy of ATP (AUC = 0.93). The other values are also good: sensitivity 52% (95% CI: 0.31–0.73), PLR 9.7 (95% CI: 4.8–19.6), NLR 0.50 (95% CI: 0.32–0.79), and DOR 19 (95% CI: 8–48). Figure 3 illustrates the results of each meta-analysis.

**Discussion**

APTs are scarcely used in the diagnosis of FA because their diagnostic accuracy and clinical utility in clinical practice are still debated [11, 12]. The present systematic review aimed to investigate the diagnostic accuracy of the APT compared with the diagnostic gold standard, i.e., the OFC, in children living with non-IgE-GI food allergy. To date, the available data on APT have been obtained in mixed populations of patients suffering from both immediate and non-IgE-mediated FA. A recent systematic review by Luo et al. [17] including 41 studies, aimed to evaluate the diagnostic accuracy of APT in children affected by different clinical pictures of FA with or without atopic dermatitis. In a group sub analysis, they found that APT is specific in children with FA-related gastrointestinal symptoms. In contrast to the metaanalysis by Luo, we aimed to evaluate the diagnostic

accuracy of APT exclusively in children with non-IgE mediated food allergy with gastrointestinal symptoms, confirmed by the OFC.

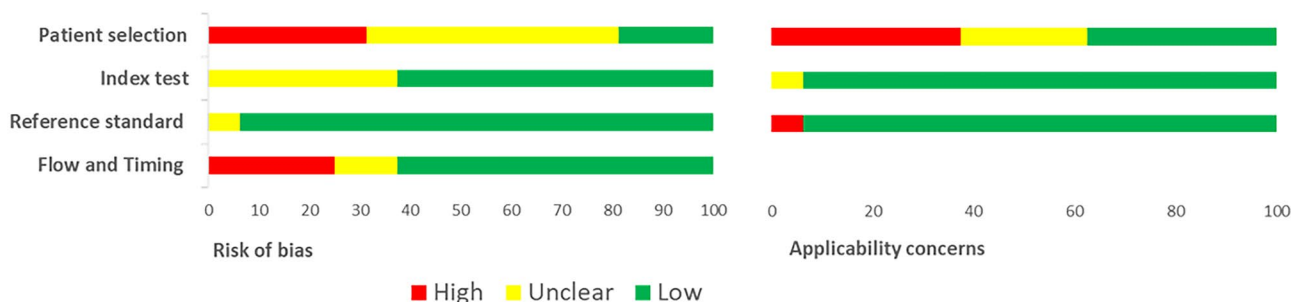
Most available studies focused on FPIMD [8, 37–39] which have not been specifically considered in the previous meta-analysis by Luo et al.

In this group, statistical analysis showed high diagnostic performance of APT, especially in the subgroup of milk-allergic patients (94% with 95% CI: 0.88–0.97, for milk-allergic patients 96% with 95% CI: 0.89–0.98). Thus, APT could help identify offending foods in allergic patients, leading to simplifying the diagnostic process. In fact, in non-IgE-GI food allergy, except for acute FPIES, the delay between food assumption and reaction makes it difficult to suspect the responsible food, while the shortness between food assumption and reaction facilitates the offending food identification in IgE-mediated allergies.

Our search found only a few studies evaluating APT efficacy in the well-known characterized picture of gastrointestinal allergy like in FPIAP, FPIES, and FPE. In detail, studies investigating the APT diagnostic role in FPIES enrolled only a small population and showed different results, probably due to different methodological accuracy. However, the most methodologically correct study by Fogg [22] showed an optimal (100%) specificity and a lousy (76.2%) sensitivity of APT.

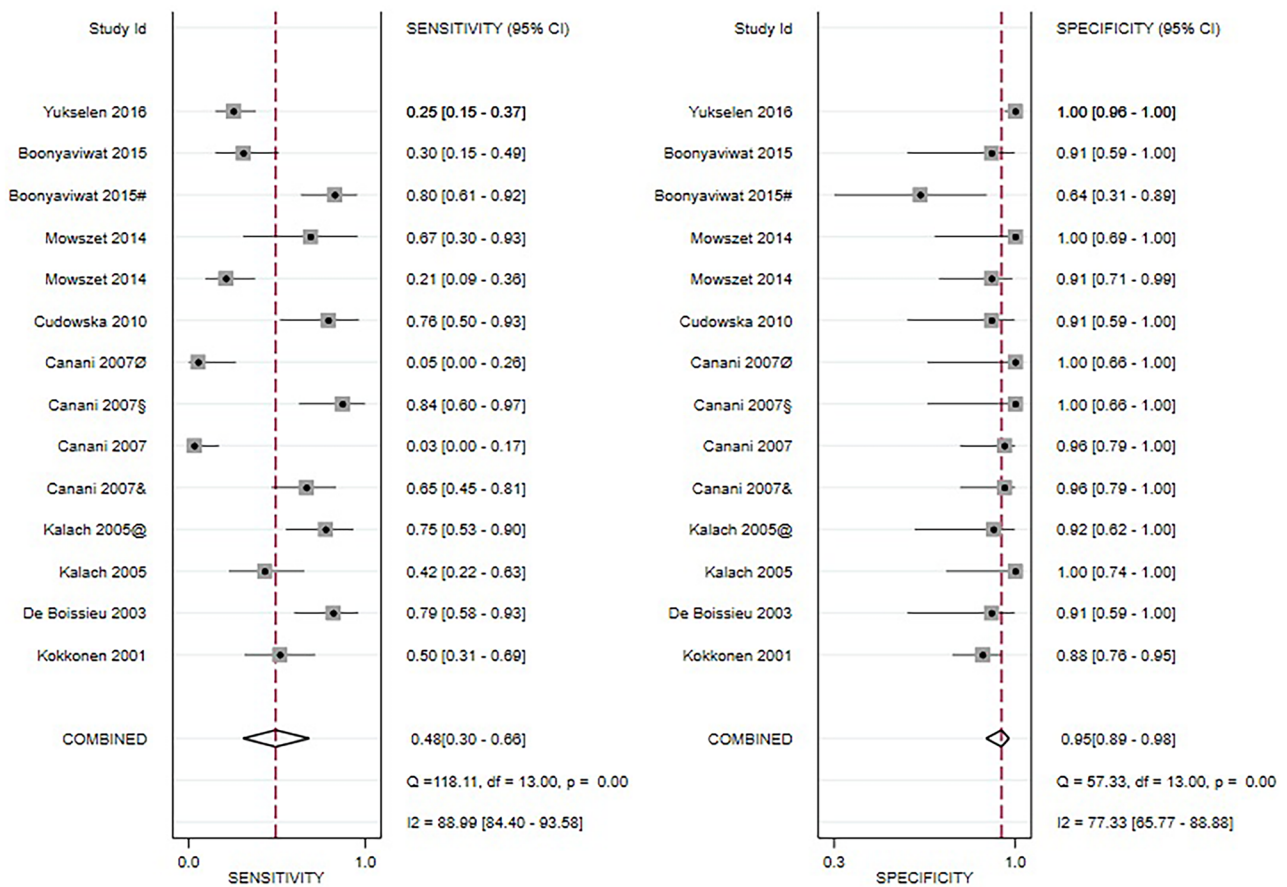
Studies enrolling patients suffering from FPIAP also showed very different results. The prospective one obtained a low QUADAS-2 score, and its results contrast with the retrospective one, which was methodologically accurate: the first showed a 100% sensitivity, the second a 100% specificity, and a third unspecified study provided only the low value of VPP (52.17%).

Sirine Kose [27] study enrolled patients affected by FPIES, FPIAP, and FPE and showed that specificity and



**Fig.2** QUADAS-2 results. Proportion of studies with low, high or unclear RISK of BIAS. Proportion of studies with low, high or unclear. Concerns regarding applicability





A

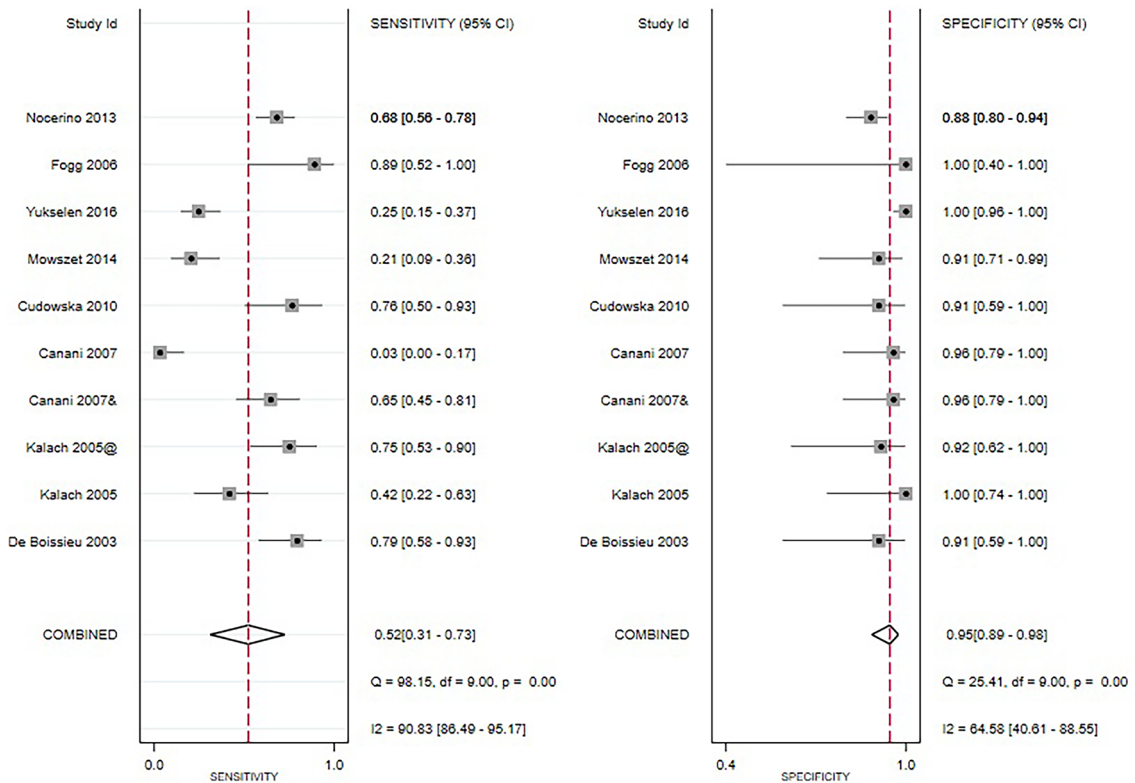
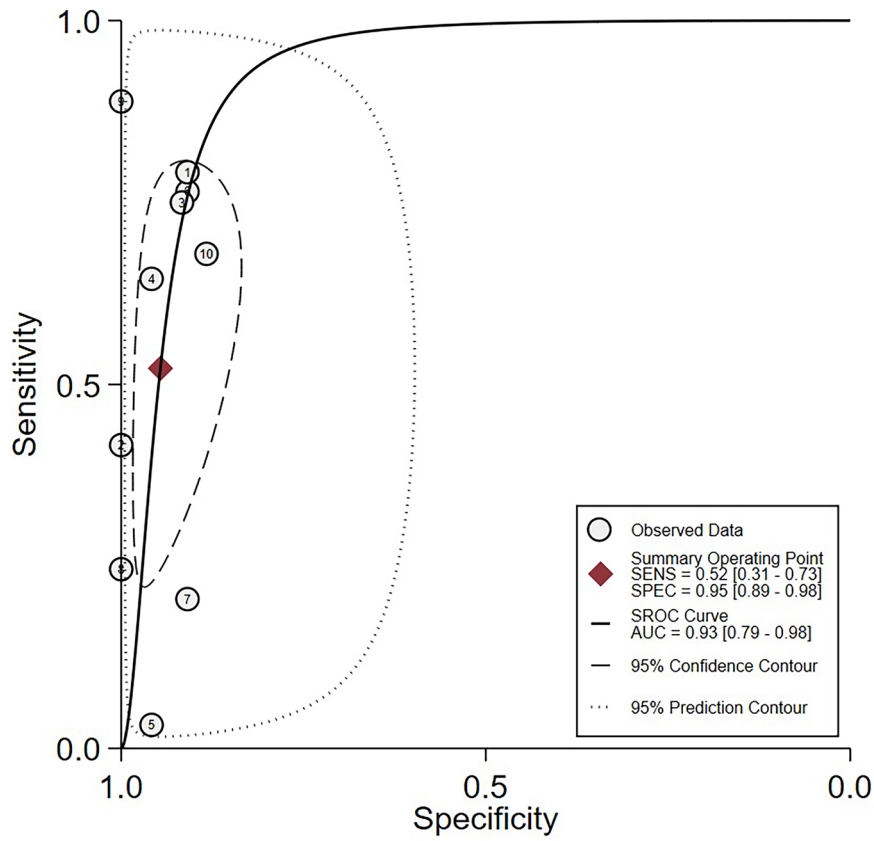
**Fig. 3** A Results of meta-analysis conducted in the group of FPIMD patients. @ dial, & Fresh, Ø eggs, #Lyophilized, + Wheat, § eggs + fresh. B Results of meta-analysis conducted in the group of milk allergic patients. @ dial, & Fresh, Ø eggs, #Lyophilized, + Wheat, § eggs + fresh

PPV values are high and varied according to the offending food, both 100% for cow’s milk and 90.9% and 80%, respectively, for hen’s egg. Different food could also account for the different results in the studies. Thus, no definitive conclusion can be drawn about APT role in patients affected by FPIAP, FPIES, and FPE, and meta-analysis was not possible. Further methodologically adequate studies are needed.

In summary, our systematic review provides further important information compared to the two previous meta-analyses. It has been designed on the specific outcome and the search included 4 more studies. Two of those were released later [26, 27]. The remaining two are studies that have been excluded from the previous meta-analysis. We have decided to include them for different reasons. For Kalach’s study [32], we justified its inclusion by extracting data for GI allergy excluding atopic dermatitis. Shirogoy’s study [34] was included after counting non-responder patients to an elimination diet in the group of non-allergic subjects. Our study provides more in-depth results based on the different clinical pictures of GI-FA.

In the FPIMD group for APT in general and for cows’ milk APT, specificity is higher in our study (0.94 and 0.96) than in Luo’s meta-analysis [17] (0.91 and 0.86), respectively. Thus, our data showing a very high AUC in FPIMD and even more for milk allergy indicate that APT is an accurate tool for diagnosing FA in FPMID, especially in the case of CMA. Instead, because of the low sensitivity value, negative APT results cannot exclude an FA diagnosis. For this reason, to increase the APT diagnostic performance, it has been suggested that it could be useful to perform APT joined to the search for sIgE or SPT [16, 36, 40–42].

We did not include studies about APT efficacy in patients affected by EGDIs. These studies were excluded because they did not report sensitivity and specificity [43, 44] or for the absence of comparison between APT results with OFC [45–47]. Most of the studies enrolling patients living with EoE focus on APT efficacy versus exclusion of the suspected food, and diagnosis is generally confirmed on symptom relief and histologic remission without documentation of clinical and histological relapsing after reintroduction



**B**

Fig. 3 (continued)

of the offending food. Only in Spergel et al. [48] study, OFC was performed but sensitivity and specificity cannot be calculated.

Limits of our systematic review are that our analysis does not allow us to draw any conclusion regarding diagnostic test performance when carried out with fresh, lyophilized, or commercial extracts allergenic foods, as the majority of the studies did not give any data about. Following the analysis, one characteristic that affected our review was the limitation of heterogeneity between studies, which may be explained by non-standardized performing test.

## Conclusions

This systematic review suggests that the APT test may be a useful tool in children living with FPIMD, especially in children affected by CMA.

**Authors' contribution** The first author, Barbara Cuomo, and the last one, Mauro Calvani, designed the work, and all authors gave the same contribution to perform data analysis and interpretation, the process of drafting, and the critical revision of the article. Meta-analysis was performed by Valentina Panetta. All authors gave the final approval of the version to be published.

**Funding** Open access funding provided by Università degli Studi di Milano within the CRUI-CARE Agreement.

**Data availability** PROSPERO2022 CRD42022322897 Available from: [https://www.crd.york.ac.uk/prospero/display\\_record.php?ID=CRD42022322897](https://www.crd.york.ac.uk/prospero/display_record.php?ID=CRD42022322897)

## Declarations

**Ethical approval** Ethical approval is not applicable to the content of this meta-analysis.

**Competing interests** The authors declare no competing interests.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

- Calvani M, Anania C, Cuomo B, D'Auria E, Decimo F, Indirli GC, Marseglia G, Mastrorilli V, Sartorio MUA, Santoro A et al (2021) Non-IgE- or Mixed IgE/Non-IgE Mediated gastrointestinal food allergies in the first years of life: old and new tools for diagnosis. *Nutrients* 13(1):226. <https://doi.org/10.3390/nu13010226>
- Sicherer SH, Sampson HA (2018) Food allergy, a review and update on epidemiology, pathogenesis, diagnosis, prevention, and management. *J Allergy Clin Immunol* 141:41–58. <https://doi.org/10.1016/j.jaci.2017.11.003>
- Calvani M, Anania C, Bianchi A, D'Auria E, Cardinale F, Votto M, Duse M, Manti S, Tosca MA, Cardinale F, et al (2021) Update on food protein-induced-entero-colitis syndrome (FPIES). *Acta Biomed* 92: 1–16. <https://doi.org/10.23750/abm.v91i11-S.10316>
- Canani BR, Caffarelli C, Calvani M, Martelli A, Carucci L, Cozzolino T, Alvisi P, Agostoni C, Lionetti P, Marseglia GL (2022) Diagnostic therapeutic care pathway for pediatric food allergies and intolerances in Italy: a joint position paper by the Italian Society for Pediatric Gastroenterology Hepatology and Nutrition (SIGENP) and the Italian Society for Pediatric Allergy and Immunology (SIAIP). *Ital J Pediatrics* 48:87. <https://doi.org/10.1186/s13052-022-01277-8>
- Meyer R, Lozinsky AC, Fleischer DM, Vieira MC, Du Toit G, Vandenplas I, Dupont C, Knibb R, Uysal P, Cavkaytar O et al (2020) Diagnosis and management of Non-IgE gastrointestinal allergies in breastfed infants - an EAACI position paper. *Allergy* 75(1):14–32. <https://doi.org/10.1111/all.13947>
- Pensabene L, Salvatore S, D'Auria E, Parisi F, Concolino D, Borrelli O, Thapar N, Staiano A, Vandenplas Y, Saps M (2018) Cow's milk protein allergy in infancy: a risk factor for functional gastrointestinal disorders in children? *Nutrients* 10–1716. <https://doi.org/10.3390/nu10111716>
- Schäppi MG, Borrelli O, Knafelz D, Williams S, Smith VV, Milla PJ, Lindley KJ (2008) Mast cell–nerve interactions in children with functional dyspepsia. *J Pediatr Gastroenterol Nutr* 47:472–480. <https://doi.org/10.1097/MPG.0b013e318186008e>
- Labrosse R, Graham F, Caubet JC (2020) Non-IgE mediated gastrointestinal food allergies in children: an update. *Nutrients* 12:2086. <https://doi.org/10.3390/nu12072086>
- Walter A, Seegräber M (2019) Wollenberg A (2018) Food-related contact dermatitis, contact urticaria, and atopy patch test with food. *Nature Clin Rev Allergy Immunol* 56(1):19–31. <https://doi.org/10.1007/s12016-018-8687-y>
- Wollenberg A, Vogel S (2013) Patch testing for noncontact dermatitis: the atopy patch test for food and inhalants. *Curr Allergy Asthma Rep* 13(5):539–544. <https://doi.org/10.1007/s11882-013-0368-6>
- Muraro A, Werfel T, Hoffmann-Sommergruber K, Roberts G, Beyer K, Bindslev-Jensen C, Cardona V, Dubois A, du Toit G, Eigenmann P et al (2014) EAACI food allergy and ana-phylaxis guidelines: diagnosis and management of food allergy. *Allergy* 69:1008–1025. <https://doi.org/10.1111/all.12429>
- Nowak-Węgrzyn A, Chehade M, Groetch ME, Spergel JM, Wood RA, Allen K, Atkins D, Bahna S, Barad A, Berin C et al (2017) International consensus guidelines for the diagnosis and management of food protein–induced enterocolitis syndrome: executive summary- Workgroup Report of the Adverse Reactions to Foods Committee, American Academy of Allergy, Asthma & Immunology. *J Allergy Clin Immunol* 139:1111–1126. <https://doi.org/10.1016/j.jaci.2016.12.966>
- Devillers AC, de Waard-van der Spek FB, Mulder PG, Oranje AP, (2009) Delayed- and immediate-type reactions in the atopy patch test with food allergens in young children with atopic dermatitis. *Pediatr Allergy Immunol* 20(1):53–58. <https://doi.org/10.1111/j.1399-3038.2008.00760.x>
- Pustisek N, Jaklin-Kekez A, Frkanec R, Sikanić-Dugić N, Misak Z, Jadresin O, Kolacek S (2010) Our experiences with the use of atopy patch test in the diagnosis of cow's milk hypersensitivity. *Acta Dermatovenerol Croat* 18(1):14–20

15. Mehl A, Rolinck-Werninghaus C, Staden U, Verstege A, Wahn U, Beyer K, Niggemann B (2006) The atopy patch test in the diagnostic workup of suspected food-related symptoms in children. *J Allergy Clin Immunol* 118(4):923–929. <https://doi.org/10.1016/j.jaci.2006.07.003>
16. Canani RB, Ruotolo S, Auricchio L, Caldore M, Porcaro F, Manguso F, Terrin G, Troncone R (2007) Diagnostic accuracy of the atopy patch test in children with food allergy-related gastrointestinal symptoms. *Allergy* 62(7):738–743. <https://doi.org/10.1111/j.1398-9995.2007.01351.x>
17. Luo Y, Zhang G-Q, Li Z-Y (2019) The diagnostic value of APT for food allergy in children: a systematic review and meta-analysis. *Pediatr Allergy Immunol* 30:451–461. <https://doi.org/10.1111/pai.13031>
18. Gayam S, Zinn Z, Chelliah M, Teng J (2018) Patch testing in gastrointestinal diseases—a systematic review of the patch test and atopy patch test. *J Eur Acad Dermatol Venereol* 32:e349–e351. <https://doi.org/10.1111/jdv.14923>
19. Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, Leeflang MM, Sterne JA, Bossuyt PM (2011) QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med*. Oct 18;155(8):529–36. <https://doi.org/10.7326/0003-4819-155-8-201110180-00009>
20. Higgins JP, Thompson SG, Deeks JJ, Altman DG (2003) Measuring inconsistency in meta-analyses. *BMJ* 327(7414):557–560. <https://doi.org/10.1136/bmj.327.7414.557>
21. Zapatero RL, Alonso LE, Martín FE, Martínez MMI (2005) Food-protein-induced enterocolitis syndrome caused by fish. *Allergol Immunopathol (Madr)* 33(6):312–316. [https://doi.org/10.1016/s0301-0546\(05\)73249-3](https://doi.org/10.1016/s0301-0546(05)73249-3)
22. Fogg MI, Brown-Whitehorn TA, Pawlowski NA, Spergel JM (2006) Atopy patch test for the diagnosis of food protein-induced enterocolitis syndrome. *Pediatr Allergy Immunol* 17:351–355. <https://doi.org/10.1111/j.1399-3038.2006.00418.x>
23. Järvinen KM, Caubet JC, Sickles L, Ford LS, Sampson HA, Nowak-Węgrzyn A (2012) Poor utility of atopy patch test in predicting tolerance development in food protein-induced enterocolitis syndrome. *Ann Allergy Asthma Immunol* 109(3):221–222. <https://doi.org/10.1016/j.anai.2012.06.020>
24. Lucarelli S, Di Nardo G, Lastrucci G, D'Alfonso Y, Marcheggiano A, Federici T, Frediani S, Frediani T, Cucchiara S (2011) Allergic proctocolitis refractory to maternal hypoallergenic diet in exclusively breast-fed infants: a clinical observation. *BMC Gastroenterol* 16(11):82. <https://doi.org/10.1186/1471-230x-11-82>
25. Alves FA, Cheik MFA, De Nápolis ACR, Rezende ÉRMDA, Barros CP, Segundo GRS (2015) Poor utility of the atopy patch test in infants with fresh rectal bleeding. *Ann Allergy Asthma Immunol* 115:161–162. <https://doi.org/10.1016/j.anai.2015.06.005>
26. Arshi S, Khoshmirsafa M, Khalife M, Nabavi M, Bermanian MH, Shokri S, Seif F, Yousefi A, Fallahpour M (2021) Atopy patch test in the diagnosis of food allergens in infants with allergic proctocolitis compared with elimination/introduction C. *Iran J Allergy Asthma Immunol* Sep 28;20(5):520–524. <https://doi.org/10.18502/ijaai.v20i5.7402>
27. Sirin Kose S, Asilsoy S, Tezcan D, Atakul G, Al S, Atay O, Kangalli Boyacioglu O, Kangalli Boyacioglu O, Uzuner N, Anal O et al (2020) Atopy patch test in children with cow's milk and hen's egg allergy: do clinical symptoms matter? *Allergol Immunopathol (Madr)* 48:323–331. <https://doi.org/10.1016/j.aller.2020.03.002>
28. De Boissieu D, Waguet JC, Dupont C (2003) The atopy patch tests for detection of cow's milk allergy with digestive symptoms. *J Pediatr* 142(2):203–205. <https://doi.org/10.1067/mpd.2003.92>
29. Nocerino R, Granata V, Di Costanzo M, Pezzella V, Leone L, Passariello A, Troncone TR, Berni Canani R (2013) Atopy patch tests are useful to predict oral tolerance in children with gastrointestinal symptoms related to non-IgE mediated cow's milk allergy. *Allergy* 68:246–248. <https://doi.org/10.1186/2045-7022-3-S3-P41>
30. Yukselen A, Celtik C (2016) Food allergy in children with refractory gastroesophageal reflux disease. *Pediatr Int* 58:254–258. <https://doi.org/10.1111/ped.12779>
31. Kokkonen J, Ruuska T, Karttunen TJ, Niinimäki A (2001) Mucosal pathology of the foregut associated with food allergy and recurrent abdominal pains in children. *Acta Paediatr* 90(1):16–21. <https://doi.org/10.1080/080352501750064824>
32. Kalach N, Soulaïnes P, de Boissieu D, Dupont C (2005) A pilot study of the usefulness and safety of a ready-to-use atopy patch test (Diallertest) versus a comparator (Finn Chamber) during cow's milk allergy in children. *J Allergy Clin Immunol* 116(6):1321–1326. <https://doi.org/10.1016/j.jaci.2005.08.033>
33. Cudowska B, Kaczmarek M (2010) Atopy patch test in the diagnosis of food allergy in children with gastrointestinal symptoms. *Adv Med Sci* 55(2):153–160. <https://doi.org/10.2478/v10039-010-0038-z>
34. Syrigou EI, Pitsios C, Panagiotou I, Chouliaras G, Kitsiou S, Kanariou M, Roma-Giannikou E (2011) Food allergy-related paediatric constipation: the usefulness of atopy patch test. *Eur J Pediatr* 170(9):1173–1178. <https://doi.org/10.1007/s00431-011-1417-6>
35. Mowszet K, Matusiewicz K, Iwańczak B (2014) Value of the atopy patch test in the diagnosis of food allergy in children with gastrointestinal symptoms. *Adv Clin Exp Med* 23(3):403–9. <https://doi.org/10.17219/acem/37136>
36. Boonyaviwat O, Pacharn P, Jirapongsananuruk O, Vichyanond P, Visitsunthorn N (2015) Role of atopy patch test for diagnosis of food allergy-related gastrointestinal symptoms in children. *Pediatr Allergy Immunol* 26(8):737–741. <https://doi.org/10.1111/pai.12382>
37. Ruffner MA, Spergel JM (2016) Non-IgE mediated food allergy syndrome. *Ann Allergy Asthma Immunol* 117(5):452–454. <https://doi.org/10.1016/j.anai.2016.04.014>
38. Cianferoni A (2020) Non-IgE mediated food allergy. *Curr Pediatr Rev* 16:95–105. <https://doi.org/10.2174/1573396315666191031103714>
39. Nowak-Węgrzyn A, Mehr KY, SS, Koletzko S (2015) Non-IgE mediated gastrointestinal food allergy. *J Allergy Clin Immunol* 135:1114–1124. <https://doi.org/10.1016/j.jaci.2015.03.025>
40. Isolauri E, Turjanmaa K (1996) Combined skin prick and patch testing enhances identification of food allergy in infants with atopic dermatitis. *J Allergy Clin Immunol* 97:9–15. [https://doi.org/10.1016/S0091-6749\(96\)70277-4](https://doi.org/10.1016/S0091-6749(96)70277-4)
41. Roehr C, Reibel S, Ziegert M, Sommerfeld C, Wahn U, Niggemann B (2001) Atopy patch test together with level of specific IgE reduces the need for oral food challenges in children with atopic dermatitis. *J Allergy Clin Immunol* 107:548–553. <https://doi.org/10.1067/MAI.2001.112849>
42. Niggemann B, Reibel S, Wahn U (2000) The atopy patch test (APT) a useful tool for the diagnosis of food allergy in children with atopic dermatitis. *Allergy* 55:281–285. <https://doi.org/10.1034/j.1398-9995.2000.00464.x>
43. Erwin EA, James HR, Gutekunst HM, Russo JM, Kelleher KJ, Platts-Mills TA (2010) Serum IgE measurement and detection of food allergy in pediatric patients with eosinophilic esophagitis. *Ann Allergy Asthma Immunol* 104(6):496–502. <https://doi.org/10.1016/j.anai.2010.03.018>
44. Dalby K, Nielsen RG, Kruse-Andersen S, Fenger C, Bindslev-Jensen C, Ljungberg S, Larsen K, Walsted AM, Husby S (2010) Eosinophilic oesophagitis in infants and children in the region of southern Denmark: a prospective study of prevalence and clinical presentation. *Pediatr Gastroenterol Nutr* 51(3):280–282. <https://doi.org/10.1097/MPG.0b013e3181d1b107>
45. Syrigou E, Angelakopoulou A, Zande M, Panagiotou I, Roma E, Pitsios C (2015) Allergy-test-driven elimination diet is useful in children with eosinophilic esophagitis, regardless of the severity

- of symptoms. *Pediatr Allergy Immunol* 26(4):323–329. <https://doi.org/10.1111/pai.12389>
46. Spergel JM, Brown-Whitehorn TF, Cianferoni A, Shuker M, Wang ML, Verma R, Liacouras CA (2012) Identification of causative foods in children with eosinophilic esophagitis treated with an elimination diet. *J Allergy Clin Immunol* 130(2):461–7.e5. <https://doi.org/10.1016/j.jaci.2012.05.021>
47. Spergel JM, Andrews T, Brown-Whitehorn TF, Beausoleil JL, Liacouras CA (2005) Treatment of eosinophilic esophagitis with specific food elimination diet directed by a combination of skin prick and patch tests. *Ann Allergy Asthma Immunol* 95(4):336–343. [https://doi.org/10.1016/S1081-1206\(10\)61151-9](https://doi.org/10.1016/S1081-1206(10)61151-9)
48. Spergel JM, Brown-Whitehorn T, Beausoleil JL, Shuker M, Liacouras CA (2007) Predictive values for skin prick test and atopy patch test for eosinophilic esophagitis. *J Allergy Clin Immunol* 119(2):509–511. <https://doi.org/10.1016/j.jaci.2006.11.016>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

## Authors and Affiliations

Barbara Cuomo<sup>1</sup>  · Caterina Anania<sup>2</sup> · Enza D'Auria<sup>3</sup>  · Fabio Decimo<sup>4</sup> · Giovanni Cosimo Indirli<sup>5</sup> · Enrica Manca<sup>6</sup> · Gian Luigi Marseglia<sup>7</sup> · Violetta Mastroianni<sup>8</sup> · Valentina Panetta<sup>9</sup> · Angelica Santoro<sup>10</sup> · Marco Ugo Andrea Sartorio<sup>11</sup> · Elisabetta Veronelli<sup>12</sup> · Mauro Calvani<sup>13</sup>

Barbara Cuomo  
cuomoba@gmail.com

Mauro Calvani  
maurocalvani58@gmail.com

- <sup>1</sup> Operative Complex Unit of Pediatrics, Belcolle Hospital, 00100 Viterbo, Italy
- <sup>2</sup> Department of Maternal Infantile and Urological Sciences, Sapienza University of Rome, 00185 Rome, Italy
- <sup>3</sup> Allergy Unit, Department of Pediatrics, Buzzi Children's Hospital, Milan 20154, Italy
- <sup>4</sup> Department of Woman, Child and General and Specialized Surgery, University of Campania "Luigi Vanvitelli, 80138 Naples, Italy
- <sup>5</sup> Pediatric Allergology and Immunology (SIAIP) for Regions Puglia and Basilicata, 73100 Lecce, Italy
- <sup>6</sup> Pediatrics Department, Policlinico Riuniti, University Hospital of Foggia, 71122 Foggia, Italy

<sup>7</sup> Pediatrics Department, Pediatric Clinic, Policlinico San Matteo, University of Pavia, 27100 Pavia, Italy

<sup>8</sup> Operative Complex Unit of Pediatrics and Emergency, Giovanni XXIII Hospital, 70056 Bari, Italy

<sup>9</sup> L'altrastatistica srl -Consultancy & Training- Biostatistics office, Rome Cap 00174, Italy

<sup>10</sup> Mother-Child Department, Pediatric Clinic, University of Parma, 43121 Parma, Italy

<sup>11</sup> Pediatric Allergology Unit, Department of Childhood and Developmental Medicine, Fatebenefratelli-Sacco Hospital, 20121 Milan, Italy

<sup>12</sup> Pediatric Department, Garbagnate Milanese Hospital, ASST Rhodense, 70056 Garbagnate Milanese, Italy

<sup>13</sup> Operative Unit of Pediatrics, S. Camillo-Forlanini Hospital, 00152 Rome, Italy