

## Using network control theory to study the dynamics of the structural connectome

Linden Parkes\*,<sup>1,2,3</sup> Jason Z. Kim\*,<sup>4</sup> Jennifer Stiso,<sup>1</sup> Julia K. Brynildsen,<sup>1</sup> Matthew Cieslak,<sup>2,5,6</sup>  
Sydney Covitz,<sup>2,5,6</sup> Raquel E. Gur,<sup>5,6</sup> Ruben C. Gur,<sup>5,6</sup> Fabio Pasqualetti,<sup>7</sup> Russell T.  
Shinohara,<sup>8,9,10</sup> Dale Zhou,<sup>1</sup> Theodore D. Satterthwaite,<sup>2,5,6,9</sup> and Dani S. Bassett<sup>1,6,11,12,13,14</sup>

<sup>1</sup>Department of Bioengineering, University of Pennsylvania, PA 19104, USA

<sup>2</sup>Lifespan Informatics and Neuroimaging Center (PennLINC),

Department of Psychiatry, Perelman School of Medicine,

University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>3</sup>Department of Psychiatry, Rutgers University, Piscataway, NJ 08854, USA

<sup>4</sup>Department of Physics, Cornell University, Ithaca, NY 14853, USA

<sup>5</sup>Penn/CHOP Lifespan Brain Institute, Perelman School of Medicine,

Children's Hospital of Philadelphia Research Institute, Philadelphia, PA 19104, USA

<sup>6</sup>Department of Psychiatry, Perelman School of Medicine,

University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>7</sup>Department of Mechanical Engineering, University of California, Riverside, Riverside, CA 92521, USA

<sup>8</sup>Department of Biostatistics, Epidemiology, and Informatics,

Perelman School of Medicine, Philadelphia, PA 19104, USA

<sup>9</sup>Center for Biomedical Image Computation and Analytics,

University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>10</sup>Penn Statistics in Imaging and Visualization Endeavor (PennSIVE),

Center for Clinical Epidemiology and Biostatistics, Perelman School of Medicine,

University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>11</sup>Department of Neurology, Perelman School of Medicine, Philadelphia, PA 19104, USA

<sup>12</sup>Department of Electrical and Systems Engineering, University of Pennsylvania, PA 19104, USA

<sup>13</sup>Department of Physics and Astronomy, University of Pennsylvania, PA 19104, USA

<sup>14</sup>Santa Fe Institute, Santa Fe, NM 87501, USA

\* These authors contributed equally

Network control theory (NCT) is a simple and powerful tool for studying how network topology informs and constrains dynamics. Compared to other structure-function coupling approaches, the strength of NCT lies in its capacity to predict the patterns of external control signals that may alter dynamics in a desired way. We have extensively developed and validated the application of NCT to the human structural connectome. Through these efforts, we have studied (i) how different aspects of connectome topology affect neural dynamics, (ii) whether NCT outputs cohere with empirical data on brain function and stimulation, and (iii) how NCT outputs vary across development and correlate with behavior and mental health symptoms. In this protocol, we introduce a framework for applying NCT to structural connectomes following two main pathways. Our primary pathway focuses on computing the *control energy* associated with transitioning between specific neural activity states. Our second pathway focuses on computing *average controllability*, which indexes nodes' general capacity to control dynamics. We also provide recommendations for comparing NCT outputs against null network models. Finally, we support this protocol with a Python-based software package called *network control theory for python* (*nctpy*).

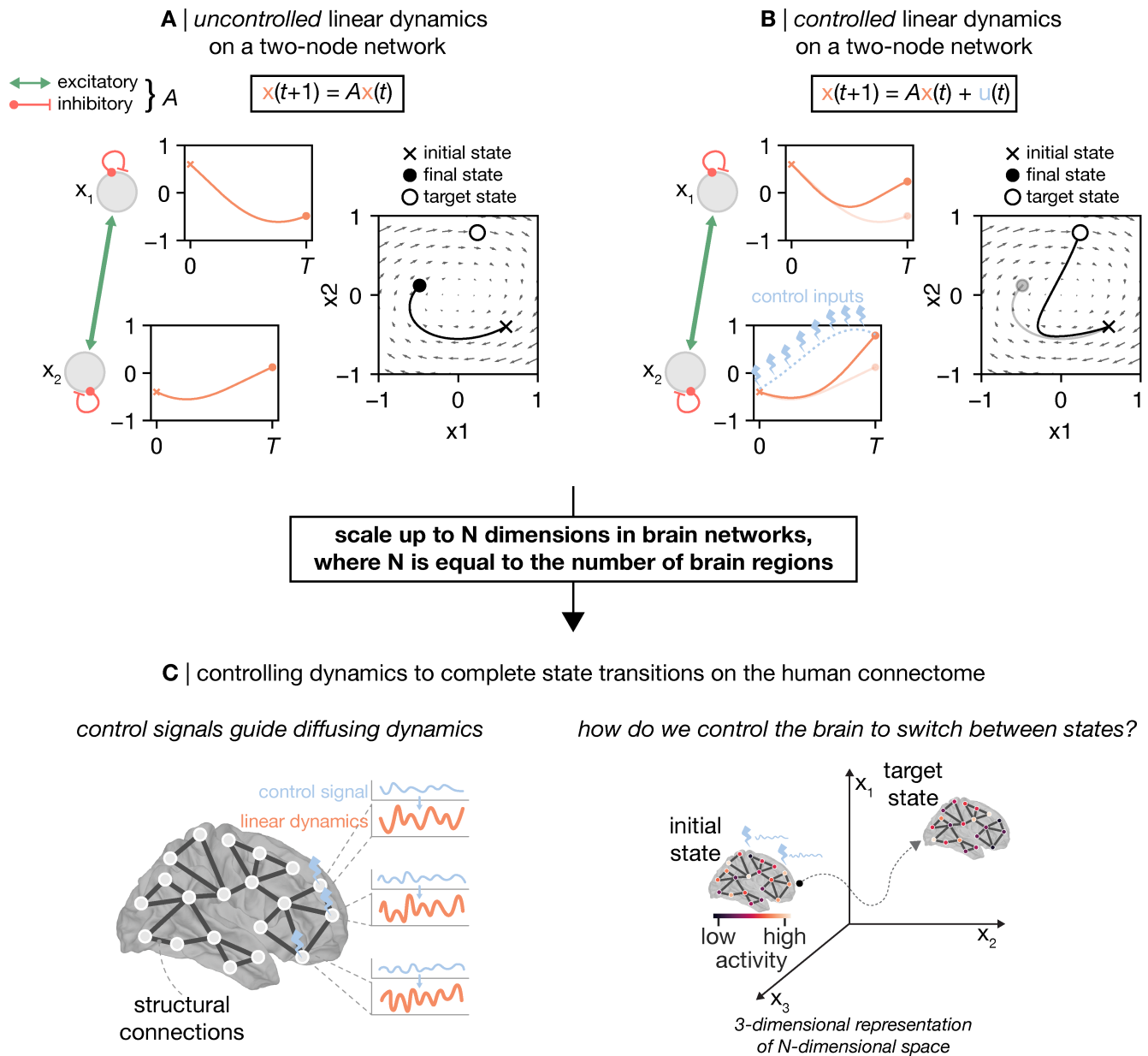
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## I. INTRODUCTION

42 Network neuroscience is principally concerned with studying the connectome [1], the description of whole brain connectivity.  
43 This connectome is often encoded as a graph of nodes interconnected by edges that can be defined across multiple scales, species,  
44 and data modalities [2, 3]. In any case, these descriptions of brain connectivity give rise to complex topology—including hubs,  
45 modules, small-worldness, and core-periphery structure [4]—and understanding how this topology shapes and constrains the  
46 brain’s rich repertoire of dynamics is a central goal of network neuroscience.

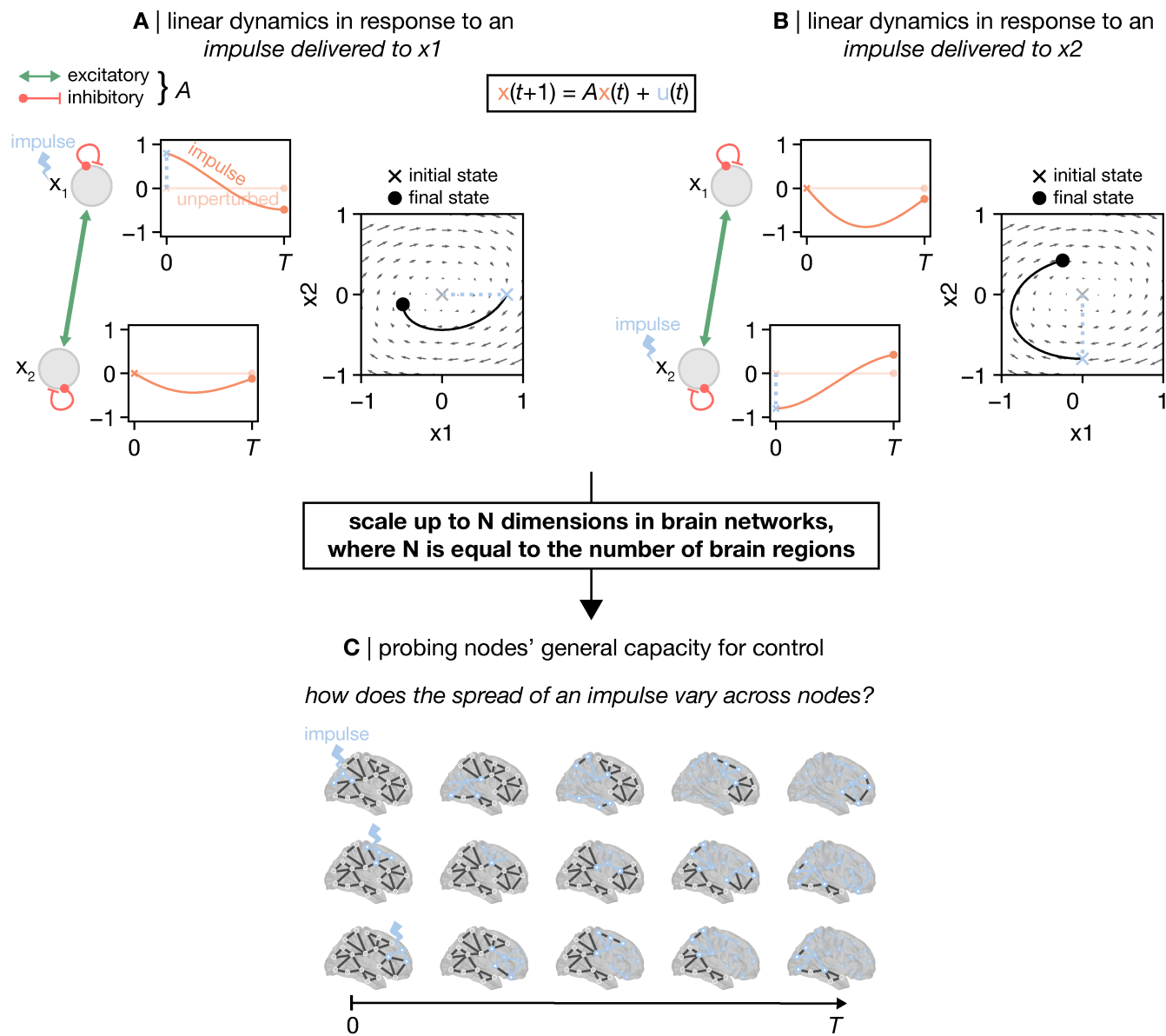
47 Network control theory (NCT) provides an approach to studying these dynamics that yields insights into how they emerge from  
48 the topology of the underlying structural connectome [5–8]. The application of NCT has revolutionized both the understanding  
49 and design of complex networks in contexts as diverse as space and terrestrial exploration, as well as modeling of financial  
50 markets, airline networks, and fire-control systems. Briefly, NCT assumes that inter-nodal communication follows a linear  
51 model of diffusion, where activity from one set of nodes (i.e., an initial *state*) spreads across the network over time along a  
52 series of fronts [4, 9]. Then, upon this dynamical system, NCT models a set of external control signals that are designed to  
53 guide these diffusing activity patterns towards a chosen target *state*. This choice can be informed by a measurement of activity  
54 evoked by behavior, spontaneous activity, or the type of brain system. These control signals are found by minimizing the total  
55 magnitude of their input over a given time horizon; that is, they are designed to achieve a desired state transition with the lowest  
56 amount of *control energy*. Once modeled, these control signals can be examined to determine to what extent, and how, they were  
57 constrained by topology, thus allowing researchers to study how the connectome might be leveraged to control dynamics.

58 Recently, we have developed and tested the application of NCT to brain network data across multiple contexts, scales, and  
59 definitions of connectivity [10–20]. Here, we present a protocol for applying NCT to two different structural connectomes: one  
60 defined using undirected connectivity estimated in the human brain [21, 22] and the other using directed connectivity estimated in  
61 the mouse brain [23–25]. Briefly, we detail two common applications of NCT that we—as well as other groups [26–32]—have  
62 deployed that focus on (i) quantifying the amount of energy that is required to complete transitions between specific brain  
63 states (Figure 1) and (ii) examining regions’ general capacity to control dynamics (Figure 2). The former approach is useful for  
64 researchers interested in examining how dynamics can be controlled to move from one place on the network to another, while the  
65 latter is useful for researchers interested in analyzing topographic maps of control. Additionally, we provide recommendations  
66 for visualization of model outputs, and discuss the use of null network models to examine which topological features affect  
67 model outputs.



**FIG. 1**

**Modeling the control energy required to complete a state transition.** Network control theory (NCT) finds the control signals that, when injected into a networked system, will guide simulated neural activity from an *initial state* to a *target state*. Here, we show a two-node toy network ( $x_1, x_2$ ) that illustrates the difference between neural activity (solid orange lines) in the absence (**A**) and presence (**B**) of a control signal (dashed blue line). **A**, Uncontrolled linear dynamics on a two-node network. Given an initial state where  $x_1 = 0.3$  and  $x_2 = -0.2$ , as well as coupling between nodes encoded by  $A$ , uncontrolled neural activity unfolds as shown on the left. These dynamic trajectories can also be represented in two dimensions as a vector field as shown on the right. Under this uncontrolled regime, the state of the system culminates in  $x_1 = -0.24$  and  $x_2 = 0.06$  at time  $T$ . **B**, Controlled linear dynamics on a two-node network. By contrast, when we introduce a control signal to  $x_2$ , the trajectory changes to now culminate in  $x_1 = 0.12$  and  $x_2 = 0.39$  at time  $T$ . Thus, NCT has found the control signal that drove our system from our *initial state* [ $x_1 = 0.3, x_2 = -0.2$ ] to our desired *target state* [ $x_1 = 0.12, x_2 = 0.39$ ]. **C**, NCT applied to the human connectome. The above model can be extended to the scale of  $N$  brain regions that constitute a whole-brain connectome (left). In doing so, we can model and examine the control signals required to transition the brain between various states of interest (right).



**FIG. 2**

**Average controllability: modeling the impulse response of the system from each node.** Network control theory (NCT) can be used to probe regions' general capacity to control dynamics. This is achieved by studying how the system responds to an impulse delivered to each node. Here, we show a two-node toy network ( $x_1, x_2$ ) coupled by  $A$ . Upon this network, we demonstrate how neural activity (solid orange lines) unfolds when an impulse (dashed blue line) is delivered to  $x_1$  (**A**) and  $x_2$  (**B**). **A**, An impulse is delivered to  $x_1$  that sets the initial state of the system to  $[x_1 = 0.4, x_2 = 0]$ . **B**, An impulse is delivered to  $x_2$  that sets the initial state of the system to  $[x_1 = 0, x_2 = -0.4]$ . In each case, the impulse response of the system is quantified as the area under the squared curves of the two orange traces. Intuitively, this measurement corresponds to the amount of activity propagated throughout the system over time. We refer to this measure as the *average controllability*. Thus, for a given time horizon ( $T$ ), a region with higher *average controllability* is better able to broadcast an impulse. **C**, Impulse response modeled for the human connectome. The above model can be extended to the scale of  $N$  brain regions that constitute a whole-brain connectome. In doing so, we can compare each region's capacity to broadcast an impulse across the whole brain.

68

## II. DEVELOPMENT OF THE PROTOCOL

69 The methods that underlie NCT are based on the established fields of control theory and dynamical systems theory. Dating  
 70 back to at least the 19th century [33], control theory is primarily concerned with engineering perturbations to achieve desired  
 71 behaviors in the states of a system, and specifically the evolution of such states over *time*. Hence, one of the most natural ways  
 72 to formulate theories of control is through *differential* and *difference* equations that mathematically define the next state of a

73 system given its current state. A common example of a control system is an inverted pendulum on a cart: the system states are  
74 the positions and velocities of the cart and pendulum, the differential equations are determined through the governing Newtonian  
75 physics, and the control task is to perturb the cart so that the cart and pendulum end up in a desired state. For example, one might  
76 want to push the cart back and forth in such a way as to stabilize the pendulum so that it remains upright [34].

77 From one perspective, the inverted pendulum is not unlike the brain, where the system states are the activities of neural units  
78 (e.g., brain regions), the differential equations are determined through the diffusion of activity through structural connections  
79 between those units, and the control task is to perturb the brain to steer it to a desired state. There is a rich history of such  
80 modeling of the brain as a dynamical system using differential equations, ranging from biophysical models of single neurons  
81 [35], to phenomenological [36] and coarse-grained [37] models of neural populations. In tandem, there is a very practical  
82 translational need to understand how to control brain dynamics [38], for example to compensate for abnormal dynamics that  
83 may be present in neurological and neuropsychiatric disorders.

84 However, there are also many ways in which the brain is not like an inverted pendulum. First is the dimensionality and  
85 complexity of the brain. Understanding how the topology of the structural connectome gives rise to brain function is a difficult  
86 task that has motivated a large body of work in the last two decades. This research has revealed that structure-function coupling  
87 is not one-to-one, varies spatially across the cortex [39–43], and is stronger when indirect structural pathways are accounted for  
88 under multiple models of network communication [44, 45]. Second is the distributed nature of brain states for human function.  
89 While some brain regions may be thought of as supporting specific functions (e.g., the fusiform face area), carrying out complex  
90 human functions typically requires the recruitment of a network of brain regions to a distributed brain state [46]. Finally, biology  
91 imposes relatively tight operating constraints. To support complex human function, the brain needs to optimize for efficient  
92 signaling while balancing the need to minimize wiring cost within the spatial constraints of the cranial cavity. Hence, there  
93 is a need to express the unique complexities and constraints of controlling brain structure-function coupling in the quantitative  
94 formalism of dynamics and control.

95 NCT emerges as a flexible framework that is methodologically based in optimal control theory [47], and can accommodate  
96 a wide range of theoretical and experimental hypotheses and constraints about structure-function coupling through a consistent  
97 mathematical framework [20, 48, 49]. Because NCT posits a model of neural dynamics at the level of individual neural interac-  
98 tions, it allows us to probe the role of the complex structural connectome on brain function at the level of those interactions [14,  
99 50, 51]. Additionally, because NCT parameterizes which regions to control and how, as well as the precise patterns of initial and  
100 target neural activity, it can answer questions ranging from the importance of a single region for propagating activity [10] to the  
101 cost of transitioning between specific brain states [11]. Hence, the development of NCT has largely served to provide a simple,  
102 first-order biophysical model with the flexibility and power to study more advanced hypotheses of function.

The modeling framework of NCT comprises  $N$  nodes (e.g., neurons, brain regions) and  $m$  inputs, and stipulates that the state  
of each node,  $x_i(t)$ , evolves in time as a weighted sum of the state of all upstream nodes,  $x_j(t)$ , and any inputs,  $u_m(t)$ . If the  
evolution of the states can be framed in terms of *states*—where the activity of upstream nodes determines the state of downstream  
nodes at the next *discrete* point in time—then the model takes the form of a difference equation,

$$\mathbf{x}(t+1) = \mathbf{A}\mathbf{x}(t) + \mathbf{B}\mathbf{u}(t), \quad (1)$$

where  $\mathbf{x}(t) = [x_1(t), x_2(t), \dots, x_n(t)]^\top$  is the vector of neural states,  $\mathbf{A}$  is the  $N \times N$  connectome,  $\mathbf{u}(t) = [u_1(t), u_2(t), \dots, u_m(t)]^\top$   
is the vector of independent control signals, and  $\mathbf{B}$  is the  $N \times m$  matrix that quantifies how each input affects the nodes. If instead  
the evolution of the states can be framed in terms of *rates*—where the activity of upstream nodes affects the *continuous* rate at  
which the state of downstream nodes change—then the model takes the form of the differential equation

$$\frac{d}{dt}\mathbf{x}(t) = \mathbf{A}\mathbf{x}(t) + \mathbf{B}\mathbf{u}(t). \quad (2)$$

103 While these two models appear similar at first glance, their definition, properties, and behavior differ substantially. In turn,  
104 the interpretation of the model parameters and outputs can vary dramatically between them.

105 This protocol will discuss two common operationalizations of NCT that can be derived from either of these models. The first  
106 employs a time-varying perturbation,  $\mathbf{u}(t)$ , to drive the neural activity,  $\mathbf{x}(t)$ , from an initial state,  $x_0$ , to a target state,  $x_f$ , given  
107 a balance of constraints on the magnitude of both the neural states and the perturbations. The magnitude of these perturbations  
108 are summarized as the *control energy*, which we interpret as the amount of effort that the model system must exert to complete  
109 a given state transition. The second is *average controllability*, which is the magnitude of the neural activity,  $\mathbf{x}(t)$ , in response  
110 to an impulse stimulus delivered to a single node; a node with higher *average controllability* is better able to leverage graph  
111 topology to spread an impulse throughout the system. Note, *average controllability* is only one example of a node-level NCT  
112 metric that falls within the broader category of *controllability statistics*. This category encompasses NCT outputs that describe  
113 different ways in which the nodes of the system may control its dynamics. While we have used other controllability statistics in  
114 our previous work (e.g., *modal controllability*, see section III), we focus on *average controllability* in this protocol owing to its  
115 simple intuition and broad appeal.

116 We focus on these two operationalizations—*control energy* and *average controllability*—because they encompass two com-  
117 mon sets of questions about the brain. The first set of questions stems from advances in neuroimaging that allow us to empirically



118 measure neural states in many forms such as functional Magnetic Resonance Imaging (fMRI), electrophysiology, and calcium  
119 imaging [52]. Given these state-level empirical data, a natural question is “how does the brain reach or switch between these  
120 states using regimes of internal or external control?” Optimal control theory provides a powerful and flexible set of tools to  
121 explore these questions under various constraints and at different spatio-temporal scales. The second set of questions stems from  
122 empirical evidence demonstrating that individual and groups of brain regions may be important for (i) enabling specific func-  
123 tions, such as visual processing [53], motor processing [54], and cognition [55, 56]; (ii) supporting critical functional processes  
124 in the brain, such as segregation and integration [57, 58]; and (iii) may be disproportionately impacted by diseases processes  
125 [59, 60]. Given these data, a natural question to ask is “what is the contribution of these sets of regions to the control of brain  
126 activity?” Average controllability measures the magnitude of propagation of stimulation along neural tracts, thereby providing a  
127 coarse-grained understanding of this contribution.

128

### III. APPLICATIONS OF THE METHOD

129 Analyzing brain data using a network representation is increasingly popular in neuroscience, and researchers have used a wide  
130 range of connectomic data to perform NCT analysis. For example, the availability of multimodal neuroimaging data in large  
131 cohorts accompanied by clinical and cognitive data [21, 22, 61–63]—as well as indices of neurobiology not measurable *in vivo*  
132 (e.g., high-resolution histology [64], gene expression [65], etc.)—enable researchers to validate NCT against brain function  
133 and biology as well as examine individual differences. Indeed, we have applied NCT in our research with a view towards  
134 achieving these goals. Here, we briefly review selections of this work to show how our protocol may be applied to study the  
135 brain. Specifically, we discuss how model outputs from NCT (A) link to network topology; (B) explain individual differences in  
136 mental health symptoms, cognition, and age; (C) predict effects of neurostimulation; (D) explain switching between functional  
137 task states; and (E) link to neuroanatomy.

138

#### A. Understanding the Influence of Topology

139 In our early work, we began by contextualizing nodal controllability statistics against what we know about connectome topology  
140 from graph theory. Specifically, Gu *et al.* [10] examined how nodal control properties—specifically *average controllability*  
141 (see Pathway B, Section IX B) and *modal controllability*—correlated with nodes’ strength (the sum of a node’s edge weights).  
142 Gu *et al.* [10] found that nodes’ strength correlated strongly positively and negatively with average and modal controllability,  
143 respectively. These relations were conserved across both humans and macaques. Collectively, these results indicate that a node’s  
144 local topological importance predicts its capacity to control the dynamics of a system.

145 We have also examined how connectome topology influences the *control energy* associated with specific state transitions. Bet-  
146 zel *et al.* [49] found that nodes’ topological importance predicted their capacity to facilitate transitions between eight canonical  
147 brain states (seven resting-state cortical networks as well as a subcortical network [66]). Specifically, Betzel *et al.* found that  
148 target states that intersected with the brain’s rich club [58]—a set of highly interconnected nodes that form the connectome’s  
149 core—exhibited low transition energy. This result demonstrates that the rich club is well positioned in the network to act as  
150 an efficient target state to which a diverse set of initial states can transition with low *control energy*. Thus, the topology of  
151 the human connectome may be optimized to guide dynamics toward the rich club, bolstering the idea that these nodes support  
152 functional integration [57, 67, 68].

153 Given these advances in understanding how connectome topology contributes to control, we subsequently analyzed what the  
154 underlying control equations could tell us about network topology. Starting from the NCT equations, Kim *et al.* [50] derived  
155 the features of network architecture that were the most important to determining *control energy*. Kim *et al.* [50] discovered that  
156 a strong and diverse set of connections from stimulated nodes to unstimulated nodes were the leading-order contributors to the  
157 control cost. Using this discovery, the authors reduced the cost of controlling connectomes in the *Drosophila*, mouse, and human  
158 by virtually resecting edges, and developed a method to meaningfully compare the control cost between different species and  
159 connectomes. These results provide simple and quantitative knowledge about the most important features of topology according  
160 to NCT.

161

#### B. Individual Differences

162 While the strong correlation between *average controllability* and strength reported by Gu *et al.* [10] may seem to imply redun-  
163 dancy between nodal controllability statistics and measures from graph theory incorporating weighted degree, we note that this  
164 correlation was spatial (i.e., across the brain) rather than between subjects. In subsequent work examining individual differences,  
165 Parkes *et al.* [69] compared the capacity of *average controllability* and strength to predict psychosis spectrum symptoms using  
166 out-of-sample testing. Parkes *et al.* [69] found that *average controllability* significantly outperformed strength in this predictive

167 task, and demonstrated that this improved performance was concentrated in higher-order default mode cortex [70]. These re-  
168 sults show that while high *average controllability* may depend upon high strength, there exists unique inter-individual variation  
169 between the metrics, and that this variance in *average controllability* couples more tightly to mental health symptoms.

170 We have also shown that *average controllability* exhibits robust developmental and sex effects. *Average controllability* in-  
171 creases between the ages of 8 and 22 years [12] and is higher in females in the cortex while higher in males in subcortex [19].  
172 Furthermore, Tang *et al.* [12] showed that age effects were strongest in nodes with higher controllability, underscoring the de-  
173 velopmental importance of nodes that are well positioned in the network to control dynamics. When examining *control energy*,  
174 Cui *et al.* [71] demonstrated that the amount of energy required to activate the fronto-parietal system—a brain network thought  
175 to support executive function [72]—was negatively correlated with both age and executive function in the same sample. This  
176 result suggests that the developmental emergence of executive function is associated with increased efficiency of neural signaling  
177 within the human connectome.

178

### C. Predicting Stimulation Effects

179 An application of NCT that has clear translational impact is modeling the relationship between brain structure and function.  
180 To this end, we have examined whether NCT can predict the brain's functional response to neurostimulation from its structural  
181 connectome. For example, in patients with epilepsy, Stiso *et al.* [14] found that NCT was able to predict electrophysiological  
182 neuronal responses (measured with electrocorticography) following direct electrical stimulation. This result shows that our  
183 model, wherein neural activity is simulated upon the structural connectome, explains variance in experimentally-manipulated  
184 empirical changes in brain state.

185 We have also examined NCT in the context of non-invasive neurostimulation techniques. In a pair of studies, Medaglia *et al.*  
186 [16, 17] delivered transcranial magnetic stimulation (TMS) to the left inferior frontal gyrus (LIFG) in between repeated sessions  
187 of a set of language tasks. Across both studies, the authors found that NCT metrics extracted from the LIFG explained variance  
188 in changes to task performance before and after TMS. These results demonstrate that NCT can be used to probe the network  
189 mechanisms that underpin how neurostimulation elicits changes in behavior.

190

### D. Modeling Switches Between Functional Brain States

191 In addition to predicting the effects of neurostimulation, NCT can be used to investigate how the topology of the structural  
192 connectome supports transitions between empirically-observed functional brain states. Our group has studied this process using  
193 brain states derived from fMRI. Cornblath *et al.* [20] clustered resting-state fMRI (rs-fMRI) into brain states representing  
194 instantaneous co-activations among canonical brain networks, and used NCT to model the energy required to transition between  
195 those states. Using a series of null network models (see section X), Cornblath *et al.* found that the topology of the structural  
196 connectome was wired to support efficient switching between brain states. This result demonstrates that the topology of the  
197 connectome is optimized to support dynamic fluctuations in resting-state activity.

198 Subsequent work by Braun *et al.* [73] examined transitions between brain states elicited by a working memory task. Braun  
199 *et al.* [73] found that transitioning from a 0-back brain state to the more cognitively demanding 2-back brain state required  
200 more energy than the reverse transition, demonstrating an asymmetry in *control energy*. Braun *et al.* also found that this  
201 energy asymmetry was more pronounced in patients with schizophrenia compared to healthy controls. Thus, while connectome  
202 topology may be setup to enable low-cost fluctuations in resting-state [20], activating cognitively demanding brain states may  
203 require more control effort. Furthermore, this increased control effort appears to scale with within-task differences in cognitive  
204 demand and is further elevated in psychopathology.

205

### E. Biologically informed NCT

206 Neuroscience is increasingly moving toward a multi-scale approach that seeks to understand how features of the brain observed  
207 at one scale link to properties observed at another, and *vice versa* [3, 74–82]. Recently, we have applied this multi-scale approach  
208 to NCT by examining how dynamics within the model are influenced by variations to regions' cellular composition. Specifically,  
209 we [51] examined how regions' profiles of cytoarchitecture impacted the energy associated with state transitions that spanned  
210 the cortical hierarchy (i.e., the sensory-fugal axis [83]). We found that state transitions traversing bottom-up along the cortical  
211 hierarchy of cytoarchitecture required lower *control energy* to complete compared to their top-down counterparts, and observed  
212 that nodes' position along this hierarchy predicted their importance in facilitating these transitions. This result shows that spatial  
213 variations in cortical microstructure constrains macroscopic connectome topology; this effect is consistent with work from  
214 neuroanatomy that describes a precise relationship between regions' profiles of cytoarchitecture and their extrinsic connectivity  
215 [84].

216 In recent work from outside our group, Luppi *et al.* [32] characterized the *control energy* associated with a large set of  
217 activity maps derived from NeuroSynth [85] related to cognition. Additionally, the authors examined how these transition  
218 energies varied when they utilized a broad range of neurotransmitter density maps to modify the control weights. This work ties  
219 together switching between functional brain states and biologically informed connectome analysis to provide the field with a  
220 comprehensive “look-up table” of how regions’ diverse biology impacts *control energy*.

221

#### IV. COMPARISON WITH OTHER METHODS

222 We consider NCT with respect to other models that also seek to understand how communication unfolds within a structurally  
223 interconnected complex system. Specifically, for practicing neuroscientists, we view NCT as complementing both more com-  
224 plex biophysical models of dynamics as well as graph-theoretic measures of inter-nodal communication. While both of these  
225 approaches model communication, they differ in their biological plausibility and complexity.

226 Biophysical models aim to capture neuronal communication by distilling the various biophysical processes necessary for  
227 functional activity into separate model parameters. These parameters are tuned to simulate biologically plausible non-linear  
228 dynamics within and between neurons at multiple scales. For example, at the scale of single neurons, the Hodgkin-Huxley model  
229 is concerned with modeling neuronal spiking activity [86], and is based on parameterizing the flow of sodium and potassium  
230 ions across the cell membrane. At the next scale up, mean-field models focus on the collective activity patterns of co-located  
231 populations of neurons [87, 88]. Coupling multiple mean-field models together—where each model represents distinct neuronal  
232 populations—enables researchers to study how non-linear dynamics emerge from brain structure at the macroscale [87]. In turn,  
233 this approach gives rise to a wide range of complex dynamical behaviors, including synchronized oscillators [89], learning  
234 [90, 91], large-scale traveling brain waves [92], and structure-function coupling [93–95]. Broadly, NCT trades biophysical  
235 accuracy and the complexity of specific model behaviors for more power in designing and studying stimuli. For example, in  
236 lieu of studying state transitions that emerge from different models of associative memory [96] and context integration [97],  
237 NCT allows us to design specific stimuli to transition the model system to states that are known to be important for memory and  
238 cognitive control under specific constraints [49].

239 By contrast, graph-theoretic approaches instantiate relatively simple models of inter-nodal communication that rely on as-  
240 sumptions such as shortest-path routing, spatial proximity, random walks, and diffusion processes [4, 9]. While these assump-  
241 tions are an oversimplification of brain dynamics—and are thus less biologically plausible—their simplicity confers greater  
242 analytic tractability and scalability, which are both desirable features when studying the human brain. This benefit compounds  
243 when the goal of a given study is to examine inter-individual differences, wherein dynamical models may be fit to thousands of  
244 participants. As such, despite their relative simplicity, graph-theoretic approaches have deepened our insights into large-scale  
245 brain organization [55, 56, 98–101], improved our understanding of the link between the brain and mental disorders [59, 102–  
246 104], and helped elucidate the link between structure and function [40, 44, 45, 105–107].

247 We consider NCT as situated between these two modeling approaches. As discussed in Section II, NCT is essentially a model  
248 of two parts, *dynamics* and *control*. For the former component, NCT models dynamics according to a diffusion process; thus,  
249 like graph theory, NCT makes simplifying assumptions of inter-nodal communication, which confers the advantages of analytic  
250 tractability and scalability. However, the second component, *control*, adds an additional layer of model parameterization that  
251 allows researchers to probe how the system might behave under different contexts (e.g., in response to task manipulations,  
252 cognitive control, or neurostimulation protocols). This added flexibility brings NCT closer to biophysical modeling, insofar  
253 as they both seek to understand how the dynamics of a system respond to external perturbation. Indeed, we have shown that  
254 NCT can be used to predict changes in the dynamics of coupled Wilson-Cowan oscillators following simulated stimulation [18],  
255 suggesting that NCT can explain some of the behaviors engendered by non-linear biophysical models.

256

#### V. EXPERIMENTAL DESIGN

257 The goal of an NCT analysis, as it is conceptualized in this protocol, is to understand how the topology of the structural  
258 connectome supports and constrains spreading dynamics, and to what extent those dynamics can be controlled. Thus, core  
259 to this analysis is the acquisition of one or more structural connectomes from a model organism. These connectomes can be  
260 obtained in numerous ways that each depend on the model organism under study and the available imaging modality. In humans,  
261 for example, structural connectomes are typically extracted from diffusion-weighted imaging (DWI) sequences obtained using  
262 MRI. Tractography algorithms are applied to DWI scans to model the white matter pathways intersecting pairs of brain regions,  
263 which are then used to populate connectome edges with the number of those pathways (e.g., the streamline count) [4]. This  
264 example constitutes a weighted undirected connectome upon which NCT can be conducted. Note that while the application of  
265 NCT is not restricted to weighted undirected connectomes (edges can be weighted or unweighted, directed or undirected), the  
266 edge weight definition determines what types of questions can be addressed using this protocol (for example, see Ref. [48] for  
267 analysis of an unweighted directed *Caenorhabditis elegans* connectome).



268 Given that connectomes are central to the application of NCT, any artifacts present in the connectomes will be reflected in  
269 model outputs. For example, connectomes populated by DWI estimates of connectivity are known to contain false positives  
270 and false negatives, which may be partly mitigated by the use of thresholding techniques [108, 109]. In-scanner head motion  
271 is well known to spuriously impact these estimates of connectivity as well [110, 111]. Thus, the accurate generation and  
272 rigorous quality control of connectomes are both crucial considerations for experimental design. For human connectomes, we  
273 recommend researchers consult the extant literature on the processing and quality control of DWI scans [108, 112, 113] (see also  
274 <https://qsiprep.readthedocs.io/>).

275 Another consideration for connectome estimation is the brain parcellation used to define system nodes. If, as mentioned  
276 above, structural connectivity is determined by streamline count, then variations in the size of regions across the parcellation  
277 will bias connectome edge weights; larger brain regions will intersect with more white matter pathways and thus show higher  
278 connectivity to the remaining regions. As with any analysis of graph topology, this bias will effect the outputs of NCT; for  
279 example, larger regions may show higher *average controllability* just by virtue of being more directly connected to the system.  
280 It is for this reason that we recommend researchers reproduce their results using several different parcellation definitions and  
281 resolutions. Doing so ensures that their results are not driven by a specific parcellation choice.

282 Beyond the core requirement of a connectome, the flexibility of NCT makes it applicable to a broad range of experimental  
283 designs (see Section III); the most critical component is that researchers have hypotheses that pertain to studying the control  
284 of brain dynamics. However, in the case of *control energy*, where researchers will study the control signals,  $\mathbf{u}(t)$ , there are  
285 some additional considerations. First, differences in brain states' magnitude will impact *control energy*, potentially necessitating  
286 the normalization of state magnitude. For example, if researchers are examining transitions between patterns of brain activity  
287 (e.g., using functional data as in [20, 73]), then differences between states' mean activity will impact *control energy*; assuming  
288 a common initial state, target states with higher activity will require more energy to transition to compared to target states with  
289 lower activity. This effect generalizes to binary brain states—where a brain state is encoded as a set of nodes being “on” while  
290 the rest of the nodes are “off”—as well. In this case, differences in state size (i.e., the number of “on” regions) constitute  
291 differences in state magnitude; transitioning to larger target states will require more energy. If there are differences in state  
292 magnitude, we recommend normalizing states before computing *control energy* (see Section IX A, step 3). Note, the need for  
293 this normalization will depend upon researchers' analyses. For example, if researchers are studying individual differences in  
294 the energy associated with a single transition, then normalization may not be necessary so long as state definition is consistent  
295 across subjects. What is critical is that researchers consider what comparisons they want to make and whether variations in state  
296 definition would confound those comparisons.

297 A second consideration is how researchers define their control set,  $B$ . As discussed in Section II, the  $N \times m$  control set defines  
298 the extent to which the nodes of the system can affect changes in its dynamics. In turn, the definition of  $B$  determines the  
299 dimensions of  $\mathbf{u}(t)$ ; the greater the number of control nodes, the more independent control signals will be generated. In our  
300 work, we have often deployed a *uniform full control set*, which means that all of the nodes of the system are designated as  
301 controllers (*full*) and all are given equivalent control over dynamics (*uniform*). In this case,  $m = N$ . Intuitively, this approach  
302 assumes that the entire brain is being controlled—either internally or externally—when completing a state transition. However,  
303 depending on a researcher's hypotheses, this assumption may not be appropriate. Instead, researchers may want to define only  
304 a subset of nodes as controllers (e.g., [49, 50]), or assign variable weights to control nodes (e.g., [30–32, 51]), or both. Note  
305 that assigning variable control weights serves to give some nodes more control over system dynamics than others. In any case,  
306 it is critical that researchers check whether their designated control set was able to complete the associated state transition (see  
307 Section IX A, step 5); successful completion of a state transition is not guaranteed in the model, and completion is less likely  
308 when transitions are driven by a small control set.

309

## VI. EXPERTISE NEEDED TO IMPLEMENT THE PROTOCOL

310 We provide open-source and broadly accessible tools that implement *optimal control* and *average controllability* in a Python-  
311 based software package called *network control theory for python (nctpy)*. In *nctpy*, we provide a flexible implementation that  
312 enables researchers to make model assumptions that best fit their research question. As a result, while a full understanding of  
313 linear systems and optimal control theory are not required, the researcher must have enough expertise to make key modeling  
314 decisions that best represent the data, which we explicitly mark in the protocol.

315 The first piece of expertise needed is to understand the differences between (and implications of) discrete-time systems and  
316 continuous-time systems (see Section II). This difference is not merely a conceptual one, because discrete-time systems display a  
317 fundamentally different set of behaviors than continuous-time systems. That is, a discrete-time system is not simply a temporally  
318 coarse-grained version of a continuous-time system. Instead, each system exhibits different dynamics. As a simple example,  
319 consider a 1-dimensional, discrete-time system that evolves according to  $x(t+1) = -x(t)$ . Starting at 1, this system will alternate  
320 between -1 and 1 over time. There does not exist an equivalent continuous-time system that evolves according to  $\frac{d}{dt}x(t) = ax(t)$   
321 (where  $a$  is a real number) that oscillates in this way.

322 The second piece of expertise needed is to understand the nature of Pathway A (*control energy*, Section IX A) and Pathway

323 B (average controllability, Section IX B) in order to interpret the outputs. Pathway A is solving an optimization problem.  
324 Specifically, first we provide a model of the dynamics (i.e.  $A, B$ ), the initial and target states, and some optimization parameters  
325 (see Section IX A). Then, we solve for the control signals,  $\mathbf{u}(t)$ , that minimize the cost. Hence, all interpretations of  $\mathbf{u}(t)$  should  
326 be made with the understanding that they were determined by the user-defined optimization parameters. Pathway B is not solving  
327 an optimization problem, and thus does not receive any optimization parameters. Rather, it measures the magnitude of the neural  
328 states over time as a result of an impulse stimulation. Because Pathways A and B use the same dynamics but output different  
329 quantities through different means, more expertise in linear systems and optimal control is needed to meaningfully compare and  
330 contrast the two pathways.

331

## VII. LIMITATIONS AND ONGOING DEVELOPMENT

332 Network control theory has the ability to flexibly accommodate many scientific questions and to generate concise knowledge  
333 from a simple model. However, NCT also possesses several limitations that are faced by many in the study of high-dimensional  
334 complex systems, such as numerical stability of algorithms, validation at the scale of microscopic states, approximations of  
335 complex interactions, and interpretation of model parameters.

336

### A. Numerical Stability of Optimization

337 One limitation is the numerical stability of Pathway A under certain parameter conditions, which arises from ill-conditioned  
338 matrices that are built while solving for the control signals. This issue occurs most frequently when using a relatively small  
339 control set—a small  $m$  in the  $N \times m$  matrix  $B$ —to control a network with large  $N$ . It is intuitive that precisely controlling the  
340 initial and target states of the whole brain from only a few nodes is difficult. In light of this limitation, it is crucial that the  
341 researcher carefully study the generated trajectories of the neural activity to ensure that the desired initial and target states are  
342 reached, and that the numerical integrator does not generate a warning of numerically ill-conditioned matrices. In the event that  
343 the control set must be small for the purposes of the research question, one solution may be to extend the control set by heavily  
344 weighting the desired control nodes and lightly weighting the remaining nodes [14]. Another option is to use Pathway B to study  
345 the average controllability of the control set.

346

### B. Validation at the Scale of Microscopic States

347 A second limitation is the validation of the model at the level of individual neural states. Phrased another way: given a  
348 connectome,  $A$ , and stimulations  $\mathbf{u}(t)$  delivered to brain regions  $B$  starting at neural activations  $\mathbf{x}(0)$ , does experimentally mea-  
349 sured neural activity agree with the simulated trajectory  $\mathbf{x}(t)$ ? The challenges associated with addressing this sort of question  
350 extend far beyond NCT and to a significant portion of systems and network neuroscience. Microstate validation between neu-  
351 ral structure and activity is most evident in small systems of neural circuits [114], but how to perform similar validations for  
352 large-scale systems such as the human brain remains an open area of research. Challenges include (i) the multiple possible  
353 scales of constructing brain networks [2, 115]; (ii) differing measures of inter-areal connectivity [108, 116]; (iii) multiple defi-  
354 nitions of simulated neural activity [87, 117–119]; and (iv) the diverse spatial and temporal resolutions at which we can record  
355 whole-brain activity [52]. Along this active area of research, recent work has demonstrated that linear models outperform non-  
356 linear and kernel-based models in both 1-step prediction and model complexity for both fMRI and EEG data [120], as well as  
357 correspondence between control energy and local metabolism [121].

358 Closely linked to ideas of validation is ongoing work on the implementation of the connectome,  $A$ . NCT stipulates a simple but  
359 mechanistic model for the evolution of neural states,  $\mathbf{x}(t)$ , given a stimulus  $\mathbf{u}(t)$ . Hence, the interpretation of the model outputs  
360 is bounded by the interpretability of the model inputs. In the majority of applications, the matrix  $A$  is taken to be the structural  
361 connectome, with the justification that regions must be able to transmit information along structural connections. However, the  
362 distribution of edge weights can vary significantly across different pre-processing pipelines, which implies equally significant  
363 variations in the strength of interactions between regions. Hence, it is important for the interpretation of the results to critically  
364 assess whether the edge from node  $i$  to node  $j$  of the implemented adjacency matrix is a reasonable measure of the strength of  
365 activity diffusion from node  $i$  to node  $j$ .

366

### C. Linear Dynamics

367 A third limitation is the assumption of linear dynamics, which enables the calculation of powerful measures such as optimal  
368 control trajectories, but hinders the biophysical realism of the framework. More sophisticated non-linear models capture complex

dynamics from individual neurons [86] to neural populations [37], thereby enabling the study of fine-grained experimental behavior [122] and complex non-linear phenomena [123]. These models make fewer simplifying assumptions to capture non-linear behaviors of biological systems such as complex memory landscapes [124]. While prior work has shown that linear models outperform many classes of non-linear models in describing and predicting brain-wide neural activity [120], extensions of NCT to non-linear systems will enable greater flexibility to accommodate and explore the impact of non-linear biophysical constraints. While the theory of non-linear control is an active area of research [125], there are immediate applications of NCT to non-linear systems, and many exciting potential extensions of NCT to capture more biophysical realism.

Broadly speaking, the linear dynamics assumed by NCT can be thought of as being valid for a non-linear system within small deviations of an operating state [7]. Hence, the most immediate application of NCT to non-linear systems is to linearize the model about an operating point, such as the upright position of an inverted pendulum [126]. Along these lines, the next immediate generalization to NCT is to linear time-varying systems [47], where the model is linearized not about a point, but about a trajectory. While methods to implement control for linearized and time-varying systems are well-established in the control community, a biophysically meaningful implementation and interpretation of the parameters—namely  $A(t)$  and  $B(t)$ —remains an area of active work [127]. Another approximation that is particularly relevant for high-dimensional neural systems is at the limit of weakly coupled oscillators [128, 129], whereby a high-dimensional system of oscillators with weak interactions can be reduced to a low-dimensional phase response curve, allowing for the potential linearization of the system about phase-locked states.

In addition to linearizing dynamics about points and trajectories, NCT can also meaningfully be applied to non-linear dynamical systems that can be made linear through a non-linear change of variables. One such example is by using finite-dimensional *Koopman subspaces*, which allow for the recasting of non-linear systems with single fixed points as higher-dimensional linear systems [130], and closely-related methods in dynamic mode decomposition [131]. Further, advances in non-linear control enable us to probe important coarse-grained questions such as the control set necessary to push non-linear systems between attracting states [132]. Other control strategies take advantage of the ability of non-linear systems to access states that lie outside of their linearization [133].

## VIII. MATERIALS

As discussed above, we split our protocol into two pathways (Figure 3). The primary pathway of our protocol focuses on computing *control energy* (Pathway A, Section IX A). This pathway is illustrated in Figures 3A, 3B, and 3C. Briefly, Figure 3A outlines the inputs required to run our protocol, Figure 3B outlines the model outputs, and Figure 3C outlines some of the variations to model inputs that we have discussed thus far. The second pathway of our protocol focuses on computing nodes' *average controllability* (Pathway B, Section IX B; Figure 3D).

### Equipment

- A computer with *Python* (tested on version 3.9) and `nctpy` installed alongside its dependencies. This protocol has been tested on Mac OS running on Intel Core i5/i7/i9 processors as well as on Apple Silicon. We have also tested this protocol on Linux Ubuntu running on Intel processors. RAM requirements will vary depending on researchers' data and analyses, but we recommend at least 16 GB. Finally, we recommend installing `nctpy` inside a virtual environment managed by Anaconda (<https://www.anaconda.com/>). The following core dependencies are required to run `nctpy`:

- `numpy` (<https://numpy.org/>). Tested on version 1.24.3.
- `scipy` (<https://scipy.org/>). Tested on version 1.10.1.
- `tqdm` (<https://github.com/tqdm/tqdm>). Tested on version 4.65.

Additionally, there are some functions in `nctpy.plotting` and `nctpy.utils` that require the following:

- `statsmodels` (<https://www.statsmodels.org/>). Tested on version 0.13.5.
- `matplotlib` (<https://matplotlib.org/>). Tested on version 3.7.1.
- `seaborn` (<https://seaborn.pydata.org/>). Tested on version 0.12.2.
- `nibabel` (<https://nipy.org/nibabel/>). Tested on version 5.1.
- `nilearn` (<https://nilearn.github.io/>). Tested on version 0.10.1.

Finally, the following optional packages were used to run the analyses illustrated in this protocols paper:

- (optional) `pandas` (<https://pandas.pydata.org/>). Tested on version 1.5.3.
- (optional) `scikit-learn` (<https://scikit-learn.org/>). Tested on version 1.2.2.

417 See <https://github.com/BassettLab/nctpy> for more details. Creating a *Python* environment using Anaconda  
418 and installing the above dependencies should take no longer than 30 minutes.

419

### Input Data

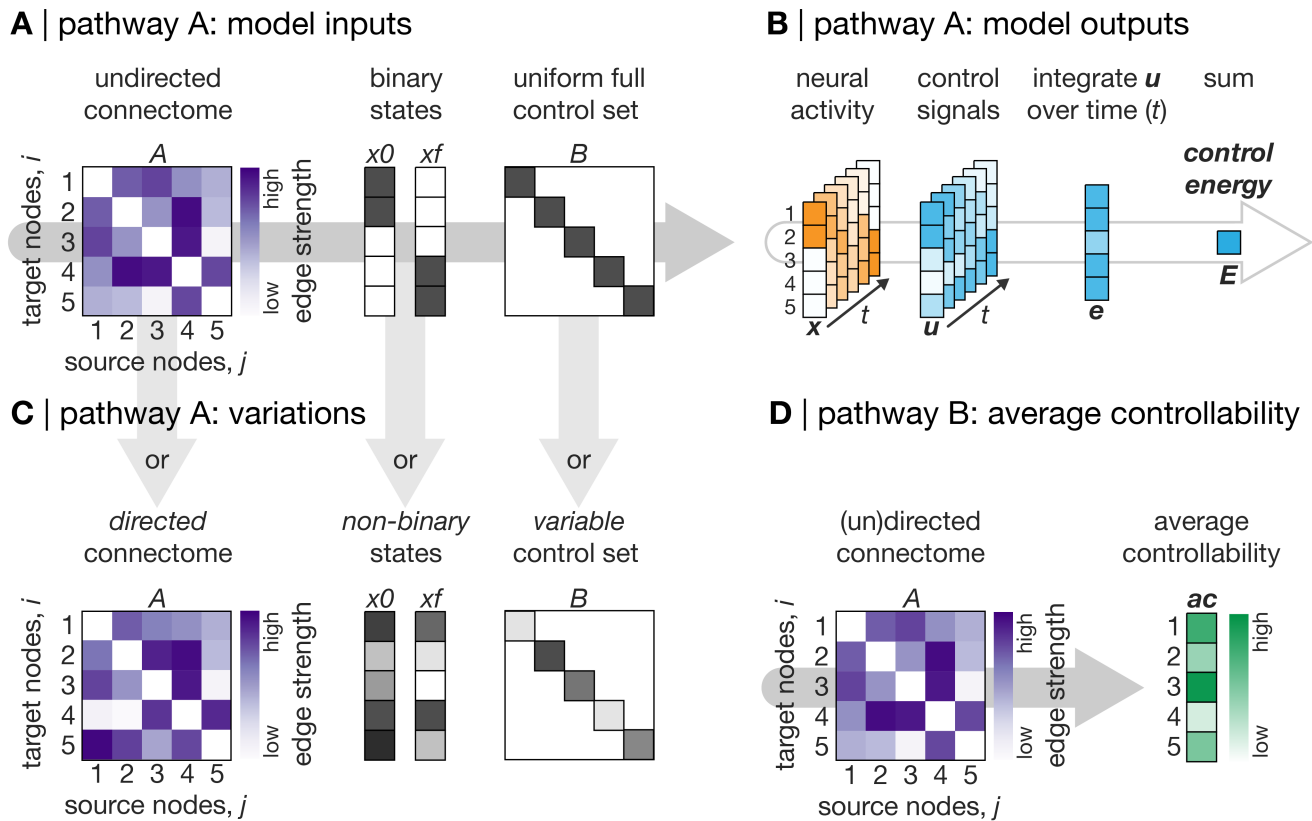
- 420 • **CRITICAL Adjacency matrix,  $A$  (used in all steps):** The adjacency matrix,  $A$  (Figure 3A, left), is the primary input  
421 to our protocol. In  $A$ , the  $N$  nodes of the system are stored on the rows and columns, and the  $N \times N$  edge values are  
422 stored in the entries. As discussed above, both the nodes and the edges of  $A$  can be defined in numerous ways and their  
423 definition depends on the acquired data as well as the research question. For example, the nodes of the system may be  
424 defined as single neurons in organisms such as the *Caenorhabditis elegans* [48, 134] or as brain regions of varying size  
425 and definition in organisms such as the mouse [76], *Drosophila* [50], macaque [135] and human [4]. The edges of  $A$   
426 may be defined as either the directed or undirected structural connectivity between nodes (Figure 3C). Note, effective  
427 functional connectivity between nodes may also be used to define edges [15, 136, 137]. Critically, our model assumes that  
428  $A_{ij}$  encodes the strength of diffusion of activity along the edge connecting *node j to node i*. In other words, our model  
429 assumes that the columns of  $A$  store the source nodes (i.e., projections from node  $j$ ) while the rows store the target nodes  
430 (i.e., projections to node  $i$ ). While this distinction is irrelevant for undirected connectomes where  $A_{ij} = A_{ji}$ , it is crucial  
431 for directed connectomes. Thus, researchers must ensure that their directed  $A$  matrix conforms to the above assumptions.  
432 In any case, NCT can be applied to either participant-level or group-averaged connectomes. Although we focus on a  
433 group-averaged undirected structural connectome in this protocol, we also include an example of our protocol applied to  
434 a directed structural connectome.
- 435 • **Brain states,  $x_0$  and  $x_f$  (used in steps 3-6):** In order to analyze state transitions, researchers also need to provide a pair of  
436 brain states (Figure 3A, middle; initial state,  $x_0$ ; target state,  $x_f$ ) relevant to their hypotheses. Providing these states allows  
437 NCT to find the control signals,  $\mathbf{u}(t)$ , that are required to transition between them and to summarize those control signals  
438 as *control energy* (Figure 3B); here, *control energy* estimates the amount of effort the model has to exert to complete the  
439 transition. Brain states can be defined in a number of ways. The simplest approach is to define each brain state as a binary  
440 vector (Figure 3A, middle), where nodes that are within a given state are assigned an arbitrary constant value (e.g., 1) and  
441 any remaining nodes are assigned a value of 0. In this setup, NCT is tasked with transitioning the brain between actuating  
442 different sets of node to a constant arbitrary level of neural activity. We commonly adopt this approach in our work. An  
443 alternative approach is to allow brain states to represent a variable pattern of activity (Figure 3C, middle). As mentioned  
444 above, Cornblath *et al.* [20] modeled the energy required to transition between brain states derived from clustering of  
445 fMRI data, while Braun *et al.* [73] used task activation maps extracted from an fMRI contrast. These approaches allowed  
446 the authors to generate state vectors that encode non-zero activity across all nodes of the system. In this protocol, we  
447 illustrate examples using both binary and non-binary brain states. Note that brain states are not required for Pathway B  
448 (Figure 3D).
- 449 • **Control set,  $B$  (used in steps 3-6):** In addition to brain states, researchers also need to designate a control set; these are  
450 the nodes that NCT will use to complete state transitions. Intuitively, the control nodes are where time-varying control  
451 signals,  $\mathbf{u}(t)$ , will be injected into the system in order to transition the system between states. Each column  $k$  of  $B$  indicates  
452 the impact of input  $u_k$  on the network nodes. Here, the simplest approach is to define a *uniform full control set* (Figure 3A,  
453 right), which means that all nodes of the system are designated as control nodes and that they are all given the same degree  
454 of control over system dynamics. Alternatively, as discussed above, researchers may also designate partial control sets or  
455 control sets with variable weights (Figure 3C, right). For the latter, variable weights can either be derived *a priori* and  
456 input directly into the model (e.g., [30–32]) or via data-driven optimization approaches (e.g., [51]). We illustrate examples  
457 of using such control sets in this protocol. Note that a control set is not required for Pathway B (Figure 3D).

458

### Example Dataset

- 459 • We primarily used undirected structural connectomes derived from DWI performed on the human brain. We obtained  
460 these connectomes from the Philadelphia Neurodevelopmental Cohort (PNC) [21, 22], a community-based study of brain  
461 development in youths aged 8 to 22 years. The neuroimaging sample of the PNC consists of 1,601 participants. From this  
462 original sample, we retained 253 typically developing participants who had no medical co-morbidity or radiological ab-  
463 normalities, and who were not taking psychoactive medications at the time of assessment. Additionally, these participants’  
464 T1-weighted, DWI, and rs-fMRI scans all passed stringent quality control procedures [113, 138, 139].  
465 - Structural connectome reconstruction was performed using QSIprep 0.14.2 [112], which is based on Nipype 1.6.1  
466 [140]. Connectomes were extracted using the 200-node variant of the Schaefer parcellation [115], ordered according to 7





**FIG. 3**

**Schematic representation of the network control theory (NCT) protocol.** Our protocol is split into two pathways. Primarily, our protocol focuses on modeling the *control energy* associated with user-defined control tasks. We refer to this part of our protocol as Pathway A (A, B, and C). Pathway A will be of interest to researchers who seek to study specific state transitions. We also outline a brief protocol for estimating nodes' *average controllability*. We refer to this part of our protocol as Pathway B (D). Pathway B will be of interest to researchers who want to examine nodes' general capacity to control system dynamics. A, Inputs required for Pathway A. To compute *control energy*, researchers must provide a structural connectome ( $A$ ), an initial state ( $x_0$ ), and a target state ( $x_f$ ), and must also define a control set ( $B$ ). B, Model outputs from Pathway A. Given these inputs, our protocol will output the state trajectory (neural activity,  $x(t)$ ) and the control signals ( $u(t)$ ). Once inspected, the control signals can be integrated over time to obtain node-level energy ( $e$ ), which in turn can be summed over nodes to get the *control energy* ( $E$ ). C, Variations to Pathway A. Pathway A can handle a diverse range of inputs, including but not limited to undirected and directed connectomes (left), binary and non-binary brain states (middle), and control sets with uniform or variable weights (right). D, Pathway B: *average controllability*. Pathway B only requires a structural connectome ( $A$ ) as input and will return the *average controllability* of each node. This metric quantifies the impulse response of the system from a given node. Higher *average controllability* indicates that a node is better positioned in the network to propagate dynamics.

467 canonical brain systems [66]. The strength of inter-regional connectivity was summarized using the number of streamlines  
 468 that intersected each pair of parcels. Connectomes were averaged over subjects. This group-averaged connectome was  
 469 thresholded by retaining the edges that were present in at least 60% of participants' connectomes [109]. This process  
 470 resulted in a final connectome with 98% edge density.

471 - rs-fMRI was also obtained from the same 253 PNC participants [21]. These data were used to generate empirical  
 472 brain activity states (see Figure 6). The eXtensible Connectivity Pipeline (XCP-D) [139, 141] was used to post-process  
 473 the outputs of fMRIPrep version 20.2.3 [142]. XCP was built with Nipype 1.7.0 [140]. Processed rs-fMRI time series  
 474 were extracted from the same 200-node parcellation mentioned above [115].

475 • We also studied a directed structural connectome obtained from the Allen Mouse Brain Connectivity Atlas.

476 - Whole-brain structural connectomes were constructed with  $2 \times 10^5$  voxels at a spatial resolution of 100  $\mu\text{m}$ . Voxels  
 477 were assigned to regions (coarse structures) according to a 3-D Allen Mouse Brain Reference Atlas [23]. Isocortex  
 478 was further divided into 6 systems (auditory, lateral, medial, prefrontal, somatomotor, and visual) based on prior work  
 479 that applied community detection to identify stable modules [143]. Connection strengths were modeled for all source



480 and target voxels using data from 428 anterograde tracing experiments in wild type C57BL/6J mice [144]. Normalized  
481 connection strengths were obtained by dividing the connection strengths by the source and target region sizes. Here, we  
482 retained only the 43 isocortical regions. This process resulted in a fully-connected directed structural connectome.

483 In all of the below code, we assume the existence of a *Python* environment with `nctpy` installed alongside its dependencies.  
484 First, we import all the functions we need to run our protocol:

```
485 # import
486 import os
487 import numpy as np
488 import pandas as pd
489 import scipy as sp
490 from scipy import stats
491 from scipy.spatial import distance
492 from sklearn.cluster import KMeans
493 from tqdm import tqdm
494
495 # import plotting libraries
496 import matplotlib.pyplot as plt
497 import seaborn as sns
498 from nilearn import datasets
499 from nilearn import plotting
500
501 # import nctpy functions
502 from nctpy.energies import integrate_u, get_control_inputs
503 from nctpy.pipelines import ComputeControlEnergy, ComputeOptimizedControlEnergy
504 from nctpy.metrics import ave_control
505 from nctpy.utils import (
506     matrix_normalization,
507     convert_states_str2int,
508     normalize_state,
509     normalize_weights,
510     get_null_p,
511     get_fdr_p,
512 )
513 from nctpy.plotting import roi_to_vtx, null_plot, surface_plot, add_module_lines
514 from null_models.geomsurr import geomsurr
```

515 Note that depending on their goals, researchers may only need a subset of this import call. Next, we will load a structural  
516 connectome as our adjacency matrix:

```
517 # directory where data is stored
518 datadir = '/path/to/data'
519 adjacency_file = 'structural_connectome.npy'
520
521 # load adjacency matrix
522 adjacency = np.load(os.path.join(datadir, adjacency_file))
523 n_nodes = adjacency.shape[0]
524 print(adjacency.shape)
525
526 # check for self-connections
527 print(np.any(np.diag(adjacency) > 0))
528
529 # get density including self connections
530 density = np.count_nonzero(np.triu(adjacency, k=0)) / (n_nodes**2 / 2)
531 print(density)
```

```
532 (200, 200)
533 True
534 0.9768
```

535 The above code demonstrates that our connectome comprises 200 nodes, includes self-connections (i.e.,  $A_{ij} > 0$ ), and has an  
536 edge density of 98%. See Figure S1 for *control energy* plotted as a function of edge density.

537

## IX. PROCEDURE

## Core Steps

### Discrete-time versus continuous-time dynamical system

1. **CRITICAL Define a time system.** The first step is to determine whether to model the linear dynamical system in discrete- or continuous-time. In a discrete-time system, the states of the system,  $\mathbf{x}(t)$ , evolve forward in time according to a set of discrete steps ( $\mathbf{x}(t) \rightarrow \mathbf{x}(t+1)$ ). In a continuous-time system, the states of the system are continuously changing in time ( $\mathbf{x}(t) \rightarrow \mathbf{x}(t) + \dot{\mathbf{x}}(t)dt$ ). The choice of time system will depend upon the research question and affects all subsequent analyses owing to differences in the mathematical implementation of NCT under each system. We refer the reader to Karrer *et al.* [8], Kim *et al.* [7], Hespanha [47], and other texts in linear systems theory for extended discussion.
2. **CRITICAL Normalize adjacency matrix (Timing: < 1 second).** Once a time system has been determined, the first practical step in this protocol is to normalize the adjacency matrix,  $A$ . Normalizing  $A$  prior to modeling the dynamical system ensures stability and that the system will not grow to infinity over time. We include a function that normalizes  $A$  appropriately depending on the researcher's chosen time system.

A) Normalizing for discrete-time systems:

```
551 1 # normalize adjacency matrix
552 2 system = "discrete"
553 3 adjacency_norm = matrix_normalization(adjacency, system, c=1)
```

The above call will normalize  $A$  according to the following equation:

$$A_{norm} = \frac{A}{|\lambda(A)|_{max} + c}.$$

Here,  $|\lambda(A)|_{max}$  denotes the largest absolute eigenvalue of the system. Additionally,  $c$  is a user-defined input parameter that determines the rate of decay of system dynamics. We set  $c = 1$  by default, which ensures that all modes of the system decay and thus that activity goes to zero over time (note, this is true of any positive  $c$  value). This normalization ensures that internal dynamics decay in a manner that is necessary for the stabilization of the system. Specifically, the largest absolute value of a matrix's eigenvalues is called the *spectral radius*, and this normalization ensures that the spectral radius is less than 1: a condition known as *Schur stability*. Intuitively, a discrete-time system given by Eq. 1 with no input (i.e.  $\mathbf{u}(t) = 0$ ) will evolve as  $\mathbf{x}(n) = A^n \mathbf{x}(0)$ , and the most unstable eigenmode of the system will evolve as  $\lambda(A)_{max}^n$ . To ensure that this mode does not grow infinitely with  $n$ , it must have a magnitude less than 1.

B) Normalizing for continuous-time systems:

```
564 1 # normalize adjacency matrix
565 2 system = "continuous"
566 3 adjacency_norm = matrix_normalization(adjacency, system, c=1)
```

The above call will normalize  $A$  according to the following equation:

$$A_{norm} = \frac{A}{|\lambda(A)|_{max} + c} - I.$$

Here,  $I$  denotes the identity matrix of size  $N \times N$ . As above, we normalize such that the spectral radius is less than one, but we take the additional step of subtracting the identity. This step exists because a continuous-time system given by Eq. 2 with no input will evolve as  $\mathbf{x}(t) = e^{At} \mathbf{x}(0)$ , and the eigenmodes of the system as  $e^{\lambda_i t}$ . Hence, for the system to decay, all  $\lambda_i$  must have a negative real component, which is achieved through the subtraction of  $I$ .

Irrespective of the chosen time system, the above step outputs  $A_{norm}$ , which contains the structural connectome as a normalized adjacency matrix that is ready for NCT analysis. Importantly, there are several different approaches to normalizing the matrix. Irrespective of approach, the key properties to ensure are the stability of the system as well as the preservation of as much structural information as possible. In all of the code and results shown below,  $A_{norm}$  was produced for a continuous-time system.

## A. Protocol Pathway A: Control Energy

3. **Define a control task: binary brain states (Timing: < 1 second).** To calculate the *control energy* required to complete a state transition, researchers must first define a control task. A control task comprises an initial state,  $x_0$ , a target state,  $x_f$ ,



632 4. **Compute control signals and state trajectory (Timing: < 1 second).** Following the definition of a control task, the  
633 next step is to find the control signals,  $\mathbf{u}(t)$ , that drive the system to transition between  $x_0$  and  $x_f$ .  $\mathbf{u}(t)$  will be an  $m \times T$   
634 matrix of  $m$  time-varying signals injected into the control nodes over a specified time horizon,  $T$ . Here, owing to our use  
635 of a full control set,  $m = N$ . Critically, injecting these control signals into a system whose initial state is encoded by  $x_0$   
636 should result in a system whose final state is encoded by  $x_f$  at time  $T$ . Alongside the control signals, we also extract  
637 the state trajectory,  $\mathbf{x}(t)$ . The state trajectory, which will be an  $N \times T$  matrix, is the time-varying pattern of simulated  
638 neural activity that unfolds as the system traverses between  $x_0$  and  $x_f$ . In this protocol, we find  $\mathbf{u}(t)$  and  $\mathbf{x}(t)$  using the  
639 `get_control_inputs` function:

```
640 1 # set parameters  
641 2 time_horizon = 1 # time horizon (T)  
642 3 rho = 1 # mixing parameter for state trajectory constraint  
643 4 trajectory_constraints = np.eye(n_nodes) # nodes in state trajectory to be constrained  
644 5  
645 6 # get the state trajectory, x(t), and the control signals, u(t)  
646 7 state_trajectory, control_signals, numerical_error = get_control_inputs(  
647 8     A_norm=adjacency_norm,  
648 9     T=time_horizon,  
649 10    B=control_set,  
650 11    x0=initial_state,  
651 12    xf=target_state,  
652 13    system=system,  
653 14    rho=rho,  
654 15    S=trajectory_constraints,  
655 16 )
```

656 By default, we set `time_horizon=1`. Note that this value is arbitrary and does not correspond to any real-world time  
657 units (e.g., seconds). Importantly, `get_control_inputs` utilizes a cost function that includes both the magnitude  
658 of the control signals and the magnitude of the state trajectory. The input parameter `rho` allows researchers to tune the  
659 mixture of these two costs while finding the input  $\mathbf{u}(t)$  that achieves the state transition. Specifically, `rho=1` places equal  
660 cost over the magnitude of the control signals and the state trajectory. Reducing `rho` below 1 increases the extent to  
661 which the state trajectory adds to the cost function alongside the control signals. Conversely, increasing `rho` beyond 1  
662 reduces the state trajectory contribution, thus increasing the relative prioritization of the control signals. Lastly, `S` takes  
663 in an  $N \times N$  matrix whose diagonal elements define which nodes' activity will be constrained in the state trajectory. In  
664 summary, `S` designates which nodes' neural activity will be constrained while `rho` determines by how much it will be  
665 constrained, relative to the control signals. Here, by setting `rho=1` and `S=np.eye(n_nodes)`, we are implementing  
666 what we refer to as *optimal control* [11]. Alternatively, researchers may choose to constrain only a subset of the state  
667 trajectory by defining partial constraint sets. If `S` is set to an  $N \times N$  matrix of zeros, then the state trajectory is completely  
668 unconstrained; we refer to this setup as *minimum control* [20, 51]. In this case, `rho` is ignored. See [here](#) for a notebook  
669 outlining different use cases of `get_control_inputs`.

670 In addition to `state_trajectory` and `control_signals`, `get_control_inputs` also outputs `numerical_error`,  
671 which stores two forms of numerical error. The first error is the *inversion* error, which measures the conditioning of the  
672 optimization problem. If this error is small, then solving for the control signals was well-conditioned (see Section VII).  
673 The second error is the *reconstruction* error, which is a measure of the distance between  $x_f$  and  $\mathbf{x}(T)$ . If this error is small,  
674 then the state transition was successfully completed; that is, the neural activity at the end of the simulation was equivalent  
675 to the neural activity encoded by  $x_f$ . We consider errors  $< 1^{-8}$  as adequately small:

```
676 1 # print errors  
677 2 thr = 1e-8  
678 3  
679 4 # the first numerical error corresponds to the inversion error  
680 5 print(  
681 6     "inversion error = {:.2E} (<{:.2E}={:})".format(  
682 7         numerical_error[0], thr, numerical_error[0] < thr  
683 8     )  
684 9 )  
685 10  
686 11 # the second numerical error corresponds to the reconstruction error  
687 12 print(  
688 13     "reconstruction error = {:.2E} (<{:.2E}={:})".format(  
689 14         numerical_error[1], thr, numerical_error[1] < thr  
690 15     )  
691 16 )  
  
692 1 inversion error = 1.36E-15 (<1.00E-08=True)  
693 2 reconstruction error = 5.16E-14 (<1.00E-08=True)
```

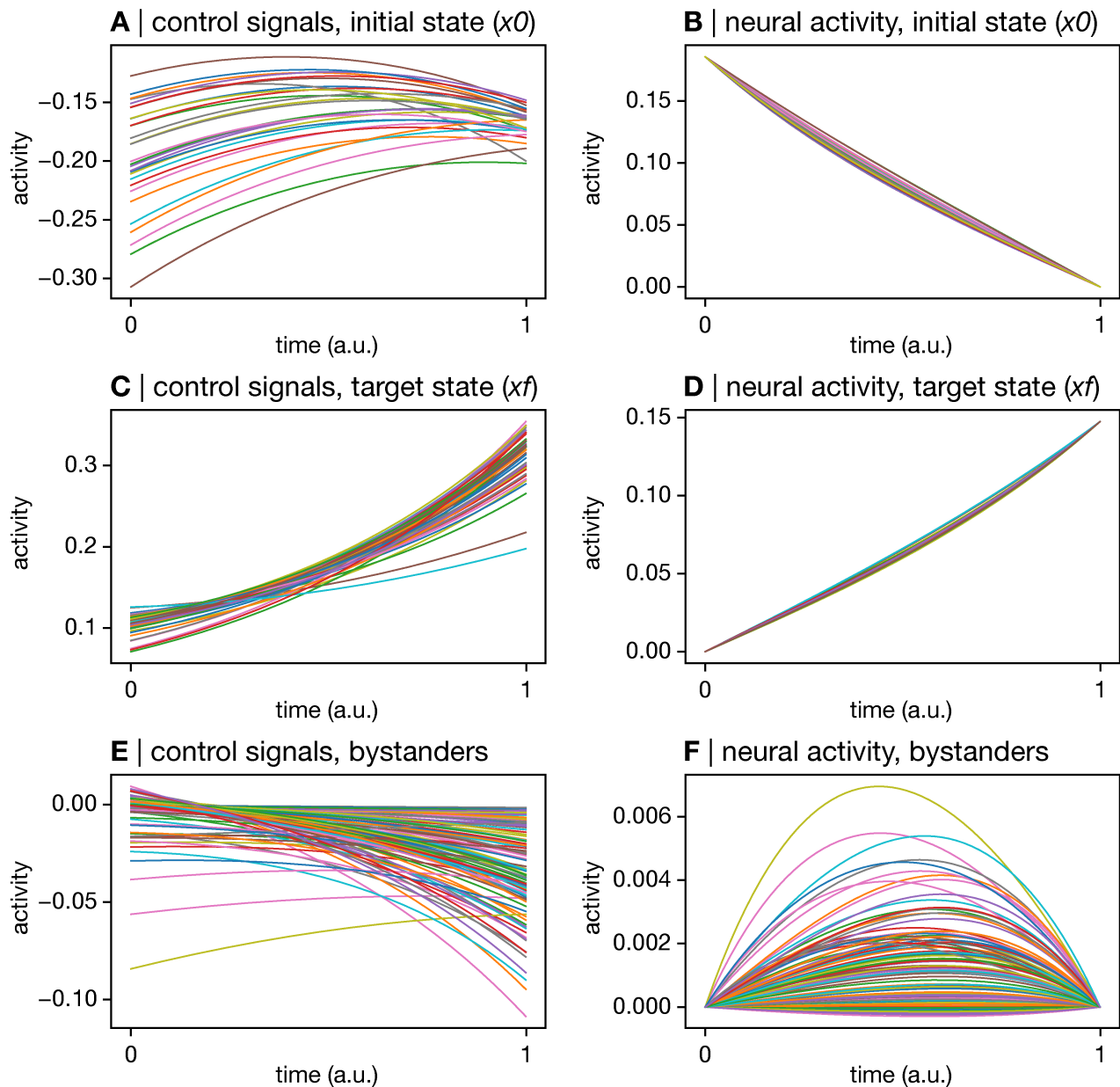
694 **5. Visualize state trajectory and control signals** (Timing: < 1 second). Once  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  have been derived, they should  
695 be visualized before computing *control energy*. Visualization provides intuition regarding how the model is behaving and  
696 is helpful for confirming that the state transition was completed successfully. As noted in Section V, completion of a state  
697 transition is not guaranteed by the model, and an incomplete transition may necessitate revising either the control set (e.g.,  
698 to provide more control over the system if a partial control set was used) or the time horizon (e.g., to provide more time  
699 for the model to complete the transition). We suggest the following simple plot:

```
700 1 f, ax = plt.subplots(3, 2, figsize=(7, 7))
701 2 # plot control signals for initial state
702 3 ax[0, 0].plot(control_signals[:, initial_state != 0], linewidth=0.75)
703 4 ax[0, 0].set_title("A | control signals, x0")
704 5 # plot state trajectory for initial state
705 6 ax[0, 1].plot(state_trajectory[:, initial_state != 0], linewidth=0.75)
706 7 ax[0, 1].set_title("B | neural activity, x0")
707 8
708 9 # plot control signals for target state
709 10 ax[1, 0].plot(control_signals[:, target_state != 0], linewidth=0.75)
710 11 ax[1, 0].set_title("C | control signals, xf")
711 12 # plot state trajectory for target state
712 13 ax[1, 1].plot(state_trajectory[:, target_state != 0], linewidth=0.75)
713 14 ax[1, 1].set_title("D | neural activity, xf")
714 15
715 16 # plot control signals for bystanders
716 17 ax[2, 0].plot(
717 18     control_signals[:, np.logical_and(initial_state == 0, target_state == 0)],
718 19     linewidth=0.75,
719 20 )
720 21 ax[2, 0].set_title("E | control signals, bystanders")
721 22 # plot state trajectory for bystanders
722 23 ax[2, 1].plot(
723 24     state_trajectory[:, np.logical_and(initial_state == 0, target_state == 0)],
724 25     linewidth=0.75,
725 26 )
726 27 ax[2, 1].set_title("F | neural activity, bystanders")
727 28
728 29 for cax in ax.reshape(-1):
729 30     cax.set_ylabel("activity")
730 31     cax.set_xlabel("time (a.u.)")
731 32     cax.set_xticks([0, state_trajectory.shape[0]])
732 33     cax.set_xticklabels([0, time_horizon])
733 34
734 35 f.tight_layout()
735 36 plt.show()
```

736 This plot (Figure 4) shows the control signals (left column) alongside the state trajectory (i.e., neural activity; right column)  
737 separately for nodes within the initial (top row) and the target state (middle row), as well as the bystanders (bottom row).  
738 Note, we define bystanders as nodes that are outside both the initial and target states. We choose this division of nodes  
739 as it provides several simple intuitions about model behavior. First, we can see that the model is driving negative time-  
740 varying control signals into the nodes of the initial state (Figure 4A), which drives their activity to 0 over time (Figure 4B).  
741 Second, we can see that the model is driving positive time-varying control signals into the nodes of the target state (Figure  
742 4C), which drives their activity from 0 to ~0.15 over time (Figure 4D). Note that ~0.15 represents the maximum neural  
743 activity following state normalization for the states presented here; this maximum activity may vary depending on state  
744 definition. Finally, we can see that diverse time-varying control signals are being injected into the bystander nodes (Figure  
745 4E), which are in turn guiding changes in these regions' activity (Figure 4F). In other words, Figure 4 shows that the model  
746 is performing a combination of "turning off" the initial state, "turning on" the target state, as well as guiding diffusing  
747 activity toward the target state via the bystanders. Figure 4 also provides a simple visual way to check whether the state  
748 transition completed successfully; at the end of the simulation, it is apparent that activity in the target state is maximal  
749 while activity in the initial state and bystanders is 0, which accords with our definition of  $xf$ . This behavior explains the  
750 low reconstruction error shown above. Additionally, this plot allows researchers to visualize how model behavior varies  
751 under different control sets (see Section 6 below as well as Figures S2, S3, S4, and S5) and time horizons (see Figures  
752 S6, S7, and S8). Note that while we view Figure 4 as the simplest way to plot initial model outputs, it is only one of  
753 many options. Researchers may choose to plot  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  as heatmaps or on the brain's surface, which would facilitate  
754 visualization of spatial patterning (see step 3a in Section 6 for an example).

755 **6. Compute control energy** (Timing: < 1 second). The final step is to summarize the control signals into *control energy*.  
756 This is done by numerically integrating the control signals over time. In this protocol, we use Simpson's rule—an extension





**FIG. 4**

**Visualize the control signals and the state trajectory.** For a given state transition, the control signals ( $u(t)$ , left column) and the state trajectory ( $x(t)$ , right column) should be visualized. Here, owing to our use of binary brain states, we group this visualization by nodes in the initial state ( $x_0$ , top row), the target state ( $x_f$ , middle row), and the remaining nodes (bystanders, bottom row). This plot provides intuition on model behavior by showing the kinds of control signals that are driving specific changes in neural activity. The top row shows that the model is driving negative time-varying control signals into the nodes of the initial state (A), which drives their activity to 0 over time (B). The middle row shows that the model is driving positive time-varying control signals into the nodes of the target state (C), which drives their activity from 0 to  $\sim 0.15$  over time (D). The bottom row shows that the model is driving diverse time-varying control signals into the bystander nodes (E), which are in turn guiding changes in these regions' activity (F).

757 of the trapezoidal rule that fits a polynomial through neighboring sets of points—to achieve this integration, yielding a  
 758 vector of node-level energy:

```

759 1 # integrate control signals to get control energy
760 2 node_energy = integrate_u(control_signals)
761 3
762 4 print(node_energy.shape)
763 5 print(np.round(node_energy[:5], 2))
    
```

```
764 1 (200,)
765 2 [21.13 37.65 23.55 21.55 28.34]
```

766 Finally, these node-level energies can be summed to produce a single estimate of *control energy*:

```
767 1 # summarize nodal energy
768 2 energy = np.sum(node_energy)
769 3
770 4 print(np.round(energy, 2))
```

```
771 1 2604.71
```

772 *Wrapping Pathway A for ease of use*

773 Steps 1-6 outline how to extract the *control energy* for a single control task, which we defined as completing a state transition between the visual system and the default mode system using control signals delivered to all system nodes (to view the above steps in a single notebook, see [here](#)). Alternatively, researchers may want to examine many control tasks within the context of a single study. Thus, in `nctpy` we include a *Python* class called `ComputeControlEnergy` that wraps all of the above steps (excluding step 5) and applies them over a list of control tasks. In addition to an adjacency matrix, `ComputeControlEnergy` expects a dictionary as input wherein each entry is a control task that includes: (i) an initial state,  $x_0$ ; (ii) a target stage,  $x_f$ ; (iii) a control set,  $B$ ; (iv) a matrix of state trajectory constraints,  $S$ ; and (v) a constraint parameter,  $\rho$ . For example, using the `states` variable defined above, this dictionary could be created as follows:

```
782 1 # initialize list of control tasks
783 2 control_tasks = []
784 3
785 4 # define control set using a uniform full control set
786 5 # note, here we use the same control set for all control tasks
787 6 control_set = np.eye(n_nodes)
788 7
789 8 # define state trajectory constraints
790 9 # note, here we constrain the full state trajectory equally for all control tasks
791 10 trajectory_constraints = np.eye(n_nodes)
792 11
793 12 # define mixing parameter
794 13 # note, here we use the same rho for all control tasks
795 14 rho = 1
796 15
797 16 # assemble control tasks
798 17 n_states = len(state_labels)
799 18 for initial_idx in np.arange(n_states):
800 19     initial_state = normalize_state(states == initial_idx) # initial state
801 20     for target_idx in np.arange(n_states):
802 21         target_state = normalize_state(states == target_idx) # target state
803 22
804 23         control_task = dict() # initialize dict
805 24         control_task["x0"] = initial_state # store initial state
806 25         control_task["xf"] = target_state # store target state
807 26         control_task["B"] = control_set # store control set
808 27         control_task["S"] = trajectory_constraints # store state trajectory constraints
809 28         control_task["rho"] = rho # store rho
810 29         control_tasks.append(control_task)
```

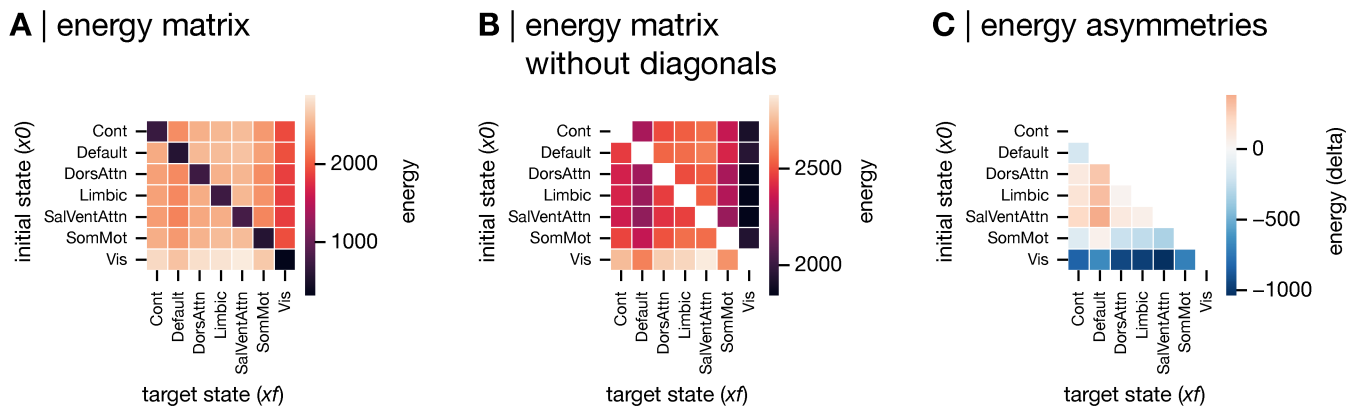
811 Note that for simplicity, the above section of code assumes the same control set ( $B$ ), trajectory constraints ( $S$ ), and  $\rho$  for  
812 all control tasks, but researchers may vary these parameters over transitions according to their needs. Next, we initialize  
813 and run `ComputeControlEnergy`:

```
814 1 # compute control energy across all control tasks
815 2 compute_control_energy = ComputeControlEnergy(
816 3     A=adjacency, control_tasks=control_tasks, system="continuous", c=1, T=1
817 4 )
818 5 compute_control_energy.run()
```

819 ComputeControlEnergy will perform matrix normalization internally, and hence here we input  $A$  rather than  
820  $A_{norm}$ . Apart from the `control_tasks` variable, ComputeControlEnergy also requires that the user specify  
821 the time system (`system='continuous'`), normalization constant (`c=1`), and time horizon (`T=1`) as input argu-  
822 ments. Once instantiated, `ComputeControlEnergy.run()` will run steps 1-6 (excluding step 5). Once com-  
823 pleted, a single estimate of *control energy* per control task will be stored in `ComputeControlEnergy.E`. Note that  
824 ComputeControlEnergy will not output the state trajectory, control signals, or node-level energy. These energy  
825 values can be trivially reshaped into a matrix and visualized (Figure 5):

```
826 1 # reshape energy into matrix
827 2 energy_matrix = np.reshape(compute_control_energy.E, (n_states, n_states))
828 3
829 4 # subtract lower triangle from upper to examine energy asymmetries
830 5 energy_matrix_delta = energy_matrix.transpose() - energy_matrix
831 6
832 7 f, ax = plt.subplots(1, 3, figsize=(7, 4))
833 8
834 9 # plot energy matrix
835 10 sns.heatmap(
836 11     energy_matrix,
837 12     ax=ax[0],
838 13     square=True,
839 14     linewidth=0.5,
840 15     cbar_kws={"label": "energy", "shrink": 0.25},
841 16 )
842 17
843 18 # plot without self-transitions
844 19 # setup mask to exclude persistence energy (i.e., transitions where i==j)
845 20 mask = np.zeros_like(energy_matrix)
846 21 mask[np.eye(n_states) == 1] = True
847 22 sns.heatmap(
848 23     energy_matrix,
849 24     ax=ax[1],
850 25     square=True,
851 26     linewidth=0.5,
852 27     cbar_kws={"label": "energy", "shrink": 0.25},
853 28     mask=mask,
854 29 )
855 30
856 31 # plot energy asymmetries
857 32 mask = np.triu(np.ones_like(energy_matrix, dtype=bool))
858 33 sns.heatmap(
859 34     energy_matrix_delta,
860 35     ax=ax[2],
861 36     square=True,
862 37     linewidth=0.5,
863 38     cbar_kws={"label": "energy (delta)", "shrink": 0.25},
864 39     mask=mask,
865 40     cmap="RdBu_r",
866 41     center=0,
867 42 )
868 43
869 44 for cax in ax:
870 45     cax.set_ylabel("initial state (x0)")
871 46     cax.set_xlabel("target state (xf)")
872 47     cax.set_yticklabels(state_labels, rotation=0, size=6)
873 48     cax.set_xticklabels(state_labels, rotation=90, size=6)
874 49 f.tight_layout()
875 50 plt.show()
```

876 In Figure 5A, the diagonals represent the energy associated with control tasks that have identical initial and target states.  
877 We refer to these energies as persistence energy [20, 73, 145], which we interpret as the amount of *control energy* required  
878 to maintain a given brain state (see Figure S9 for a persistence energy example of Figure 4). Persistence energy is  
879 typically lower than the energy for control tasks wherein the initial and target states differ (i.e., the off-diagonal elements  
880 of `energy_matrix`). Thus, in order to better visualize the variance in energy across state transitions, we recommend  
881 also plotting `energy_matrix` excluding the diagonal elements (Figure 5B). Additionally, as the energy associated with  
882 transitioning from  $x_0$  to  $x_f$  is not necessarily equivalent to that associated with the reverse direction, researchers may  
883 subtract the upper and lower triangles of `energy_matrix` to examine energy asymmetries (Figure 5C). Indeed, we  
884 have done this in our previous work (see [51, 73]).



**FIG. 5**

**Visualize control energy.** In cases where multiple state transitions are considered, we recommend visualizing *control energy* using the following heatmaps. **A**, Full energy matrix. The full energy matrix shows energy for all state transitions, including those where the initial and target state are the same (diagonal elements). This form of energy is referred to as persistence energy and is interpreted as the amount of effort required to maintain neural activity in a given state. Persistence energy is typically lower than the energy associated with transitioning between different states. **B**, Between-state energy matrix. In order to visualize variance in *control energy* for transitions between states, we recommend plotting the energy matrix without the diagonal as well. **C**, Energy asymmetry matrix. Finally, we recommend subtracting the transpose of the energy matrix and visualizing the lower (or upper) triangle of the ensuing asymmetry matrix. Doing so allows researchers to see the asymmetries present in the *control energy*.

### Variations to Pathway A

All of the above constitutes our complete protocol for calculating *control energy*. However, there are multiple variations to the above protocol that researchers may wish to consider depending on their research goals. In this section, we illustrate a selection of these variants that are likely to be of broad interest to the field of neuroscience (Figure 3C). These variations include (A) studying non-binary brain states; (B) implementing partial and non-uniform control sets; and (C) examining directed structural connectomes.

#### Non-binary brain states

In Section IX A, we illustrated a state transition between the visual system and the default mode system using a binary definition of brain states extracted from a canonical system-level grouping of brain regions [66]. Below, we provide an example of using non-binary brain states instead. This example draws on aforementioned work from Cornblath *et al.* [20], who modeled the energy required to transition between clusters of rs-fMRI activity.

3A) *Define a control task: non-binary brain states.* To define non-binary brain activity states, we cluster rs-fMRI data along the time dimension to extract co-activation states [20]. To achieve this goal, we first load the rs-fMRI data for 253 participants extracted from the same parcellation that defined our structural connectome. Then, we concatenate these time series end-to-end across subjects:

```

1 # load resting-state time series
2 rsfmri_file = 'pnc_schaefer200_rsts.npy'
3 rsfmri = np.load(os.path.join(datadir, rsfmri_file))
4
5 n_trs = rsfmri.shape[0]
6 n_nodes_rsfmri = rsfmri.shape[1]
7 n_subs = rsfmri.shape[2]
8 print('n_trs, {0}; n_nodes, {1}; n_subs, {2}'.format(n_trs, n_nodes_rsfmri, n_subs))
9
10 rsfmri_concat = np.zeros((n_trs * n_subs, n_nodes_rsfmri))
11 print(rsfmri_concat.shape)
12
13 for i in np.arange(n_subs):
14     # z score and concatenate subject i's time series
15     start_idx = i * n_trs

```

```
915     16     end_idx = start_idx + n_trs
916     17     rsfmri_concat[start_idx:end_idx, :] = sp.stats.zscore(rsfmri[:, :, i], axis=0)
```

```
917     1 n_trs, 120; n_nodes, 200; n_subs, 253
918     2 (30360, 200)
```

919 This process assigns empirical time series to each of the system nodes comprising 30,360 time points (120 TRs by  
920 253 participants) of rs-fMRI data. Next, we cluster these data in time using K-means and visualize the corresponding  
921 co-activation states on the cortical surface. To generate this plot, we include a function called `surface_plot` that  
922 utilizes tools from Nilearn (<https://nilearn.github.io/stable/index.html>):

```
923     1 # extract 5 clusters of activity
924     2 n_clusters = 5
925     3 kmeans = KMeans(n_clusters=n_clusters, random_state=0).fit(rsfmri_concat)
926     4
927     5 # extract cluster centers. These represent dominant patterns of recurrent activity over time
928     6 centroids = kmeans.cluster_centers_
929     7 print(centroids.shape)
930     8
931     9 # plot centroids on brain surface
932    10 lh_annot_file = ("/path/to/schaefer/files/lh.Schaefer2018_200Parcels_7Networks_order.annot")
933    11 rh_annot_file = ("/path/to/schaefer/files/rh.Schaefer2018_200Parcels_7Networks_order.annot")
934    12 fsaverage = datasets.fetch_surf_fsaverage(mesh="fsaverage5")
935    13
936    14 for cluster in np.arange(n_clusters):
937    15     f = surface_plot(
938    16         data=centroids[cluster, :],
939    17         lh_annot_file=lh_annot_file,
940    18         rh_annot_file=rh_annot_file,
941    19         fsaverage=fsaverage,
942    20         order="lr",
943    21         cmap="coolwarm",
944    22     )
```

```
945     1 (5, 200)
```

946 Figure 6 illustrates the centroids for 2 of the 5 clusters we extracted using K-means. As we are focused on illustrating  
947 a single state transition for the purposes of this protocol, the remaining 3 centroids are not shown (see Cornblath *et al.*  
948 [20] for detailed discussion of all 5 clusters). These centroids represent patterns of activity that recur throughout our  
949 concatenated time series. The spatial patterning of these 2 centroids indicate activity concentrated in visual cortex  
950 (Figure 6A) and the default mode system (Figure 6B), respectively. Using the same functions outlined above, we  
951 can extract this pair of centroids as brain states and recompute the control energy:

```
952     1 # extract visual cluster as initial state
953     2 initial_state = centroids[1, :]
954     3 # extract default mode cluster as target state
955     4 target_state = centroids[4, :]
956     5
957     6 # normalize state magnitude
958     7 initial_state = normalize_state(initial_state)
959     8 target_state = normalize_state(target_state)
960     9
961    10 # get the state trajectory and the control signals
962    11 state_trajectory, control_signals, numerical_error = get_control_inputs(
963    12     A_norm=adjacency_norm,
964    13     T=time_horizon,
965    14     B=control_set,
966    15     x0=initial_state,
967    16     xf=target_state,
968    17     system=system,
969    18     rho=rho,
970    19     S=trajectory_constraints,
971    20 )
972    21
973    22 # get energy
974    23 node_energy = integrate_u(control_signals)
975    24 energy = np.sum(node_energy)
```



976 As stated above in step 5, the temporal unfolding of  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  can be visualized using line plots (Figure 6B top,  
977 C top). Alternatively, to illustrate the spatial patterning of the state transition,  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  can be visualized on the  
978 cortical surface for specific time points (Figure 6B bottom, C bottom):

```
979 1 timepoints_to_plot = np.arange(0, state_trajectory.shape[0], int(state_trajectory.shape[0] / 5))  
980 2  
981 3 for timepoint in timepoints_to_plot:  
982 4     f = surface_plot(  
983 5         data=state_trajectory[timepoint, :],  
984 6         lh_annot_file=lh_annot_file,  
985 7         rh_annot_file=rh_annot_file,  
986 8         fsaverage=fsaverage,  
987 9         order="lr",  
988 10        cmap="coolwarm"  
989 11    )
```

990

### Partial and variable control sets

991 In Section IX A, we illustrated a state transition controlled via a *uniform full control set*. Such a control set amounts to  
992 assigning all nodes of the system the same degree of control over system dynamics. Below, we provide examples of using  
993 alternatives to this regime that involve using variable (instead of uniform) and partial (instead of full) control sets.

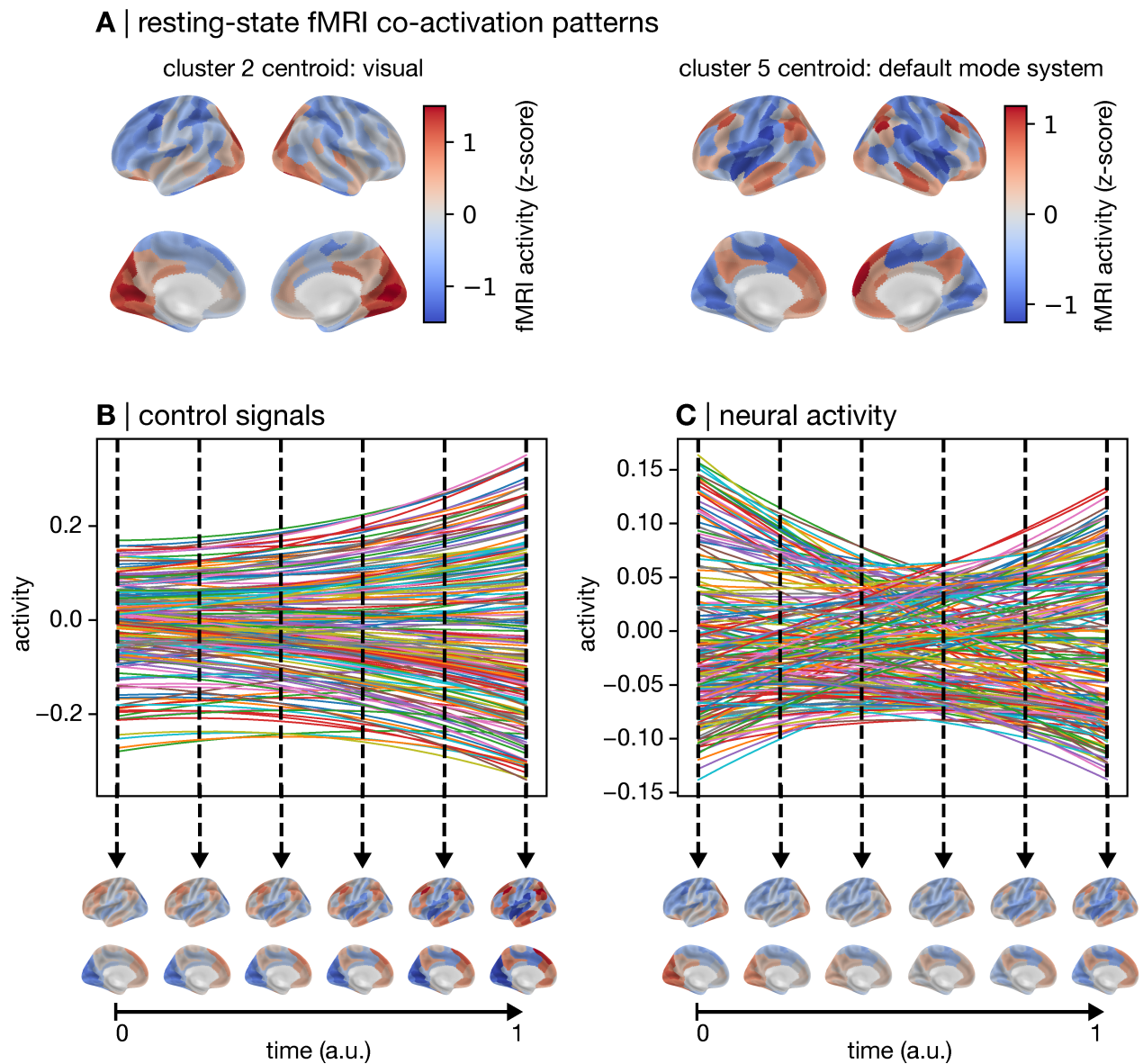
994 3B) *Define a control task: uniform partial control set*. Defining a *uniform partial control set* is straight forward. Instead  
995 of assigning the  $N \times N$  identity matrix to  $B$ , we select specific diagonal elements to assign the value of 1, and assign  
996 0 to the remaining diagonal (and non-diagonal) elements. Here, we illustrate the example of selecting the bystander  
997 regions (see Figure 4E, F) as our control set:

```
998 1 # specify a uniform partial control set: some nodes are control nodes  
999 2 # and all control nodes are assigned equal control weight  
1000 3 bystanders = np.logical_and(  
1001 4     initial_state == 0, target_state == 0  
1002 5 ) # use initial state and final state to find bystanders. note, this only works for binary  
1003     states  
1004 6 control_set = np.zeros((n_nodes, n_nodes)) # initialize control nodes matrix  
1005 7 control_set[bystanders, bystanders] = 1 # set bystanders to control nodes
```

1006 Figure S2 shows the results of generating  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  under the above control set. Note that the above code only  
1007 works for binary brain states, because in this case bystanders can be trivially defined as nodes with no activity in  
1008 either the initial or target states. This fact does not imply that partial control sets cannot be used for non-binary brain  
1009 states. In Figure S2, we observe that the control signals are 0 for both  $x_0$  and  $x_f$ , indicating that they received no  
1010 control signals. Additionally, the control signals for the bystanders, as well as the neural activity of all nodes, has  
1011 changed substantially compared to Figure 4. Notably, the control signals are several orders of magnitude greater  
1012 for this *uniform partial control set* compared to the *uniform full control set* used above. In turn, although this state  
1013 transition completes successfully, the energy we observe here is also several orders of magnitude greater (energy =  
1014  $6.64 \times 10^9$ ). See Figures S3, S4, and S5 for more examples of *uniform partial control sets*, including some for which  
1015 the state transition does not complete. While the code implementation of a *uniform partial control set* is straight  
1016 forward, researchers must ensure that their control set is large enough to achieve numeric stability (see Section VII).

1017 3C) *Define a control task: a priori variable full control set*. Instead of assigning all nodes of the system the same degree of  
1018 control over dynamics, researchers may want to make statements about which nodes should have more or less control  
1019 according to their hypotheses. One way that this can be achieved is by using node-level annotation maps [146, 147] to  
1020 assign control weights. For a single annotation map—which we assume is stored in 'neuromap.npy'—assigning  
1021 weights can be achieved in the following way:

```
1022 1 # helper func for printing descriptive stats  
1023 2 def print_stats(x):  
1024 3     print(  
1025 4         "min={:.2f}; max={:.2f}; mean={:.2f}; std={:.2f}; skew={:.2f}; kurt={:.2f}".format(  
1026 5             np.min(x),  
1027 6             np.max(x),  
1028 7             np.mean(x),  
1029 8             np.std(x),  
1030 9             sp.stats.skew(x),
```



**FIG. 6**

**Control signals and state trajectory for non-binary brain states derived from resting-state fMRI.** We applied K-means clustering to resting-state fMRI time-series to extract patterns of co-activation. These patterns were used as non-binary brain states for network control theory (NCT) analysis. **A**, Resting-state fMRI clusters that represent visual system activity (left) and default mode activity (right). The control signals and the state trajectory were modeled by assigning the visual system to the initial state and the default mode system to the target state. **B**, Control signals visualized using line plots (top) and on the cortical surface for select time points (bottom). **C**, State trajectory (neural activity) visualized using line plots (top) and on the cortical surface for select time points (bottom).

```

1031         sp.stats.kurtosis(x),
1032     )
1033 )
1034
1035 14 neuromap_file = 'neuromap.npy'
1036 15 neuromap = np.load(os.path.join(datadir, neuromap_file))
1037 16 print(neuromap.shape)
1038 17 print_stats(neuromap)
1039 18
1040 19 control_set = np.zeros((n_nodes, n_nodes)) # initialize B matrix
1041 20 control_set[np.diag_indices(n_nodes)] = neuromap # set weights using neuromap

```

1042 1 (200,)

```
1043 2 min=-0.41; max=0.42; mean=-0.04; std=0.20; skew=0.25; kurt=-0.54
```

1044 The above code demonstrates that our annotations, and therefore our control set, include both positive and negative  
1045 values. The sign of a given weight will determine how a positive control signal delivered to that node influences its  
1046 neural state; a positive weight will cause a positive change in neural activity, while a negative weight will cause a  
1047 negative change. As control signals can also carry positive and negative values over time (see Figure 4), this behavior  
1048 has no impact on *control energy*; a positive control signal driven into a node using a positive control weight will cause  
1049 the same change in neural state as a negative control signal driven via a negative control weight. However, it does  
1050 impact the interpretation of  $\mathbf{u}(t)$ . Thus, to simplify the interpretation of the control weights, and of  $\mathbf{u}(t)$ , we suggest  
1051 adding a constant (1) as well as the absolute minimum value to the annotation map:

```
1052 1 # modify neuromap so that its minimum value is 1
1053 2 neuromap += 1 + np.abs(np.min(neuromap))
1054 3 print_stats(neuromap)
1055 4
1056 5 control_set = np.zeros((n_nodes, n_nodes)) # initialize B matrix
1057 6 control_set[np.diag_indices(n_nodes)] = neuromap # set weights using neuromap
```

```
1058 1 min=1.00; max=1.84; mean=1.38; std=0.20; skew=0.25; kurt=-0.54
```

1059 This process will create a set of weights with a minimum value of 1 and, in this case, a maximum value of 1.84 (see  
1060 Figure S10 for a plot  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  derived from this control set). This fact simplifies our interpretation. For example,  
1061 because all weights are positive, we can say that the highest weight node has 1.84 times more control over system  
1062 dynamics compared to the lowest weight node. Additionally, whether a control signal drives a positive or negative  
1063 change in neural state now depends solely on its own sign at a given point in time.

1064 A recent study by Singleton *et al.* [30] utilized an *a priori variable full control set* by assigning control weights  
1065 according to a range of serotonin receptor maps. Singleton *et al.* [30] found that the *control energy* associated with  
1066 their state transition was lowest when using a 5-HT2a receptor map compared to 5-HT1a, 5-HT1b, 5-HT4, and 5-  
1067 HTT maps. This result suggests that, compared to other receptors, the spatial patterning of 5-HT2a receptors yielded  
1068 the most efficient state transition (i.e., by reducing *control energy* the most). The authors subsequently replicated  
1069 their results using N,N-Dimethyltryptamine [31]. However, to draw this conclusion, researchers need to be mindful of  
1070 the following caveat. In our model, increasing the total amount of control necessarily reduces energy. This relation  
1071 exists because the task of completing a given state transition is easier for the model when any node is granted a  
1072 greater degree of control over dynamics than it had previously, leading to smaller amplitude control signals and thus  
1073 lower energy. For example, if we compared energy between our *uniform full control set* and our above *variable full*  
1074 *control set*, energy would be trivially lower for the latter. This difference occurs because all weights on the diagonal  
1075 of  $B$  are 1 in our *uniform full control set*, while all but one of the weights in our *variable full control set* are  $> 1$ . This  
1076 issue is pertinent for researchers who want to compare different *variable full control sets* (as in [30–32]), because any  
1077 comparison of *control energy* across two different annotation maps needs to account for differences in those maps'  
1078 distributions. The simplest solution to this problem is to take the rank of each annotation map and then rescale the  
1079 ranks to be between 1 and 2. `normalize_weights` performs this normalization:

```
1080 1 neuromap_file_1 = 'neuromap.npy'
1081 2 neuromap_1 = np.load(os.path.join(datadir, neuromap_file_1))
1082 3 neuromap_1_norm = normalize_weights(neuromap_1)
1083 4
1084 5 neuromap_file_2 = 'neuromap2.npy'
1085 6 neuromap_2 = np.load(os.path.join(datadir, neuromap_file_2))
1086 7 neuromap_2_norm = normalize_weights(neuromap_2)
1087 8
1088 9 print_stats(neuromap_1)
1089 10 print_stats(neuromap_1_norm)
1090 11 print_stats(neuromap_2)
1091 12 print_stats(neuromap_2_norm)
```

```
1092 1 min=-0.41; max=0.42; mean=-0.04; std=0.20; skew=0.25; kurt=-0.54
1093 2 min=1.00; max=2.00; mean=1.50; std=0.29; skew=-0.00; kurt=-1.20
1094 3 min=-0.10; max=0.16; mean=-0.00; std=0.07; skew=0.59; kurt=-0.78
1095 4 min=1.00; max=2.00; mean=1.50; std=0.29; skew=-0.00; kurt=-1.20
```

1096 Once annotation maps have been normalized in this manner, differences in energy can only be attributed to differ-  
1097 ences in the (rank) spatial patterning between maps. This independence occurs because both annotation maps now  
1098 conform to a uniform distribution. Note, energy derived from this normalization approach will still be lower than  
1099 our *uniform full control set*.

3D) *Define a control task: data-driven variable full control set.* Instead of assigning variable weights *a priori*, researchers may assign them in a data-driven manner. In our recent work, we developed an approach for achieving this goal using gradient descent (see [51] for more details). Briefly, starting with a *uniform full control set*, this approach involves perturbing control nodes' weight one at a time by a constant arbitrary amount and measuring the corresponding change in energy. This process results in  $N$  estimates of *perturbed control energy* for a given state transition. As mentioned above, each of these perturbations will necessarily reduce *control energy* compared to the baseline *uniform full control set*, creating negative  $\Delta$ s. In turn, differences in  $\Delta$  magnitude encode the relative importance of each node to completing a specific state transition; nodes with more negative energy  $\Delta$ s are more important:

```
1108 1 # container for perturbed energies
1109 2 energy_perturbed = np.zeros(n_nodes)
1110 3
1111 4 for node in tqdm(np.arange(n_nodes)):
1112 5     # start with a uniform full control set
1113 6     control_set = np.eye(n_nodes)
1114 7
1115 8     # add arbitrary amount of additional control to node
1116 9     control_set[node, node] += 0.1
1117 10
1118 11     # get perturbed control signals (u_p)
1119 12     _, control_signals, _ = get_control_inputs(
1120 13         A_norm=adjacency_norm,
1121 14         T=time_horizon,
1122 15         B=control_set,
1123 16         x0=initial_state,
1124 17         xf=target_state,
1125 18         system=system,
1126 19         rho=rho,
1127 20         S=trajectory_constraints,
1128 21     )
1129 22
1130 23     # integrate control signals to get control energy
1131 24     node_energy = integrate_u(control_signals)
1132 25
1133 26     # summarize nodal energy
1134 27     energy_perturbed[node] = np.sum(node_energy)
1135 28
1136 29 # check if perturbed energy is lower than original energy. Should print True
1137 30 print(np.all(energy_perturbed < energy))
1138 31
1139 32 # calculate energy delta. these values will all be negative,
1140 33 # indicating reduced energy compared to control_set=np.eye(n_nodes)
1141 34 energy_delta = energy_perturbed - energy
1142
1143 1 True
```

Once estimated, these  $\Delta$ s can be used as weights on  $B$  to obtain an optimized version of *control energy*. Note that here we draw a distinction between *optimal* and *optimized*. The former refers to constraining the magnitudes of  $\mathbf{x}(t)$  and  $\mathbf{u}(t)$  in the optimization problem, whereas the latter refers to finding the weights that create the most efficient transition, irrespective of this constraint:

```
1147 1 # re-compute energy using energy deltas as weights
1148 2 # we do this by taking a single step down the
1149 3 # gradient created by the energy deltas
1150 4 learning_rate = 0.01 # set a learning rate for gradient descent
1151 5 control_set = np.zeros((n_nodes, n_nodes)) # initialize container for optimized weights
1152 6 control_set[np.diag_indices(n_nodes)] = (1 - energy_delta * learning_rate) # step down gradient
1153 7 control_set = (
1154 8     control_set / sp.linalg.norm(control_set) * sp.linalg.norm(control_set)
1155 9 ) # normalize
1156 10 # normalization ensures that the optimized weights have the same
1157 11 # norm as control_set=np.eye(n_nodes)
1158 12
1159 13 # get optimized control signals
1160 14 _, control_signals, _ = get_control_inputs(
1161 15     A_norm=adjacency_norm,
1162 16     T=time_horizon,
1163 17     B=control_set,
1164 18     x0=initial_state,
```

```
1165     19     xf=target_state,
1166     20     system=system,
1167     21     rho=rho,
1168     22     S=trajectory_constraints,
1169     23 )
1170     24
1171     25 # integrate control signals to get control energy
1172     26 node_energy_optimized = integrate_u(control_signals)
1173     27 print(np.round(node_energy_optimized[:5], 2))
1174     28
1175     29 # summarize nodal energy
1176     30 energy_optimized = np.sum(node_energy_optimized)
1177     31 print(np.round(energy_optimized, 2))
```

```
1178     1 [20.56 34.94 22.77 20.92 26.96]
1179     2 2429.68
```

1180 The above code uses `energy_delta` to define a gradient that we step down one time using a learning rate of 0.01. Stepping down this gradient yields a set of optimized weights,  $B_o$ , that we then use to re-estimate *control energy*. As expected, this new estimate of energy (2429.68) is lower than the energy derived from our *uniform full control set* (2604.71). Thus, we have found a set of control weights that optimize (i.e., reduce) our *control energy* in a data-driven way. Additionally, setting up this algorithm using gradient descent allows researchers to optimize energy over multiple steps, wherein each new set of optimized weights is calculated from the previous set. In `nctpy` we include a *Python* class called `ComputeOptimizedControlEnergy` that wraps the above optimization steps and allows researchers to define their own learning rate and number of gradient steps:

```
1188     1 control_task = dict() # initialize dict
1189     2 control_task["x0"] = initial_state # store initial state
1190     3 control_task["xf"] = target_state # store target state
1191     4 control_task["S"] = trajectory_constraints # store state trajectory constraints
1192     5 control_task["rho"] = rho # store rho
1193     6 compute_opt_control_energy = ComputeOptimizedControlEnergy(
1194     7     A=adjacency,
1195     8     control_task=control_task,
1196     9     system,
1197    10     c=1,
1198    11     time_horizon,
1199    12     n_steps=2,
1200    13     lr=learning_rate,
1201    14 )
1202    15 compute_opt_control_energy.run()
```

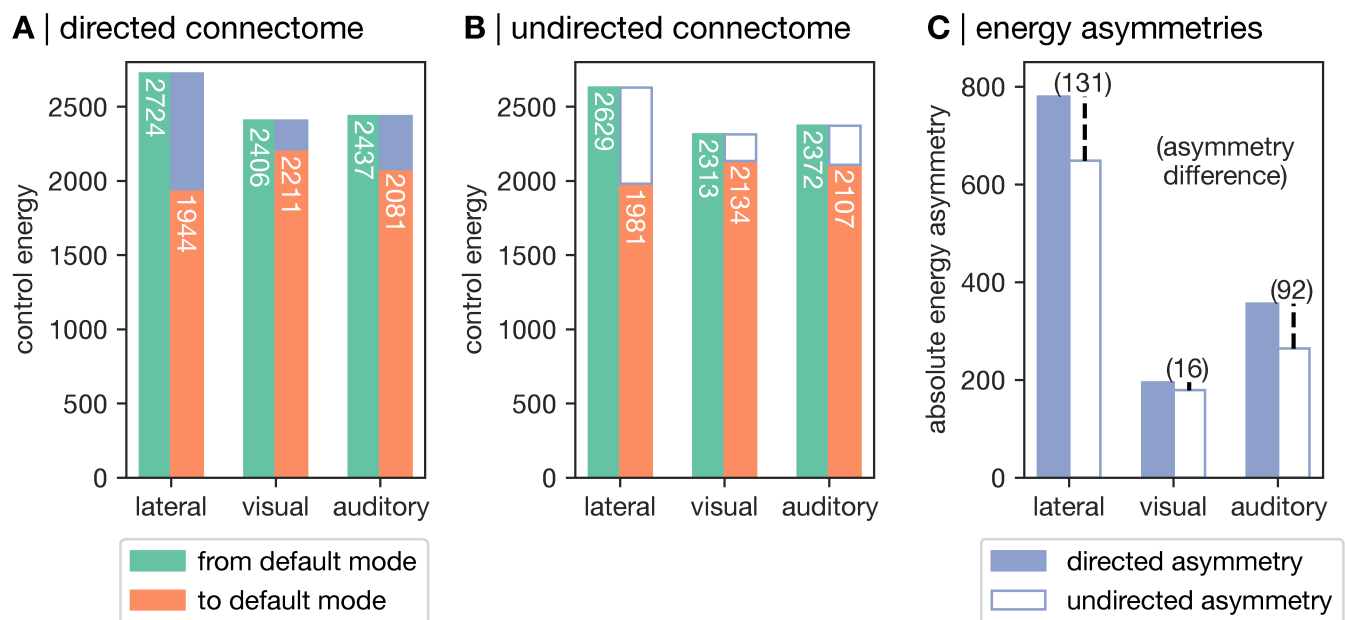
1203 `ComputeOptimizedControlEnergy` is similar to `ComputeControlEnergy` but has some notable differences. Like `ComputeControlEnergy`, `ComputeOptimizedControlEnergy` will perform matrix normalization internally, thus we input  $A$  rather than  $A_{norm}$ . Additionally, `ComputeOptimizedControlEnergy` requires that users specify the time system (`system='continuous'`), normalization constant ( $c=1$ ), and time horizon ( $T=1$ ) as input arguments. The differences are as follows. First, `ComputeOptimizedControlEnergy` only accepts a single control task dictionary, rather than a list of tasks. Second, `ComputeOptimizedControlEnergy` requires that users also specify the number of gradient steps (`n_steps=2`) and the learning rate (`lr=0.01`) as inputs. Once instantiated, `ComputeOptimizedControlEnergy.run()` will run the above optimization steps. Once completed, optimized energy at each gradient step will be stored in `ComputeControlEnergy.E_opt` as a vector of length  $N_s$ , where  $N_s$  is the number of steps. The corresponding optimized control weights will be stored in `ComputeControlEnergy.B_opt` as an  $N_s \times m$  matrix.

#### 1214 *Directed structural connectome*

1215 In Section IX A, we performed NCT analysis using an undirected structural connectome derived from the human brain. However, our protocol is designed to work with directed connectomes as well. Thus, as a final variation on Pathway A, we present results from a directed connectome obtained in the mouse brain. As discussed in Section VIII, our model assumes that  $A_{ij}$  encodes the edge connecting *node j to node i*. Provided that this assumption is met, Pathway A can be run without modification.



Using the Allen Mouse Brain Connectivity Atlas [24, 25, 148], we extracted the ipsilateral directed connectivity from 43 regions in the mouse isocortex. These 43 regions were grouped into 6 systems: auditory, lateral, medial, prefrontal, somatomotor, and visual. Following [148], we combined the prefrontal and medial systems, as well as 3 regions from the somatomotor system, to create the mouse default mode system. Then, using `ComputeControlEnergy`, we estimated the energy required to transition from the default mode to the lateral, visual, and auditory systems and back again. Similar to the undirected human connectome (see Figure 5), this process yielded energy asymmetries (Figure 7A). We repeated this process using a symmetric version of the mouse connectome ( $A_s = \frac{A+A^T}{2}$ ; Figure 7B) and examined how energy asymmetries differed between the directed and undirected cases (Figure 7C); see here for `Python` code. For both the directed (Figure 7A) and undirected connectomes (Figure 7B), we found that *control energy* was lower when transitioning to the default mode compared to from the default mode. Critically, these observed energy asymmetries were larger for the directed connectome compared to the undirected connectome (Figure 7C); the lateral energy asymmetry was larger by 131 units (20% larger), the visual asymmetry was larger by 16 units (9% larger), and the auditory asymmetry was larger by 92 units (35% larger). Thus, the presence of directed edges increased the energy asymmetries observed in our model.



**FIG. 7**

**Energy asymmetries are larger for the directed than the undirected mouse connectome.** Our protocol can be applied to connectomes with directed edges as well as undirected edges. In the directed case, our protocol assumes that  $A_{ij}$  encodes the edge connecting *node j* to *node i*. Here, we estimated *control energy* using the Allen Mouse Brain Connectivity Atlas. **A**, *Control energy* associated with transitioning from the default mode (DMN) to the lateral, visual, and auditory systems (green) and back again (orange) in the directed mouse connectome. *Control energy* associated with transitioning to the default mode was lower than the reverse direction, indicating a clear asymmetry (blue). **B**, *Control energy* estimated in the undirected mouse connectome. We recomputed energy using a symmetrized version of the mouse connectome ( $A_s = \frac{A + A^T}{2}$ ) and observed the same set of energy asymmetries. **C**, Differences in energy asymmetries between the directed and undirected mouse connectome. We observed that the size of the energy asymmetries were larger for the directed than the undirected mouse connectome.

1233

## B. Protocol Pathway B: Average Controllability

Pathway A is the primary component of our protocol. Implementing these steps assumes that researchers are interested in studying a specific set of state transitions defined in accordance with their research questions and hypotheses. In the absence of such hypotheses, researchers may instead wish to examine nodes' general capacity to control a broad range of unspecified state transitions. To support these types of hypotheses, we present a complementary pathway to our protocol that yields estimates of *average controllability*, where higher values indicate that a region is better positioned in the network to control dynamics (see section II):

1. **Compute average controllability** (Timing: discrete time, < 1 second for 200 nodes; continuous time, 10 – 20 seconds for 200 nodes).

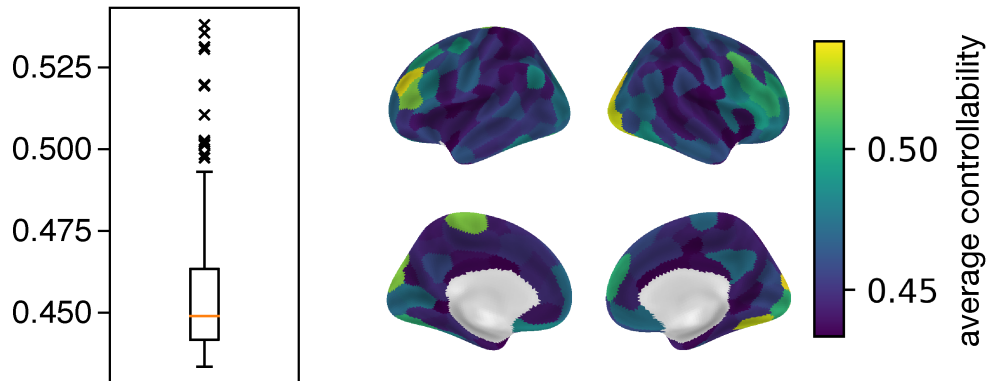
1240  
1241

```
1242 1 # compute average controllability
1243 2 average_controllability = ave_control(adjacency_norm, system)
```

1244 average\_controllability will be a vector containing the *average controllability* of each node of the system.

1245 2. **Visualize average controllability.** As *average controllability* is a regional metric, we can simply plot its distribution

1246 of values on the surface of the cortex (Figure 8).



**FIG. 8**

**Average controllability.** Each system node receives an impulse of equal magnitude. Nodes with higher *average controllability* are able to broadcast that impulse throughout the system to a greater extent compared to nodes with lower *average controllability*. Thus, nodes with high *average controllability* are better positioned within the network to control dynamics. Distribution of *average controllability* values are displayed using a box plot (left) as well as projected onto the cortical surface (right). In the box plot, the orange line represents the median, the box spans the middle 50% of the data, the whiskers span 1.5 times the interquartile range on either side, and the crosses represent outliers beyond this limit.

1247

## X. ANTICIPATED RESULTS

1248 The final outputs of our protocol will depend on whether researchers choose to follow Pathway A (see Section IX A)

1249 or Pathway B (see Section IX B). For the former, the output will be one estimate of *control energy* per control task, or

1250 one estimate per brain region per task if energy was not summarized across regions. This value will be positive and can

1251 be thought of as the amount of effort the model has to exert in order to complete a specific control task; higher energy

1252 corresponds to greater effort. For the latter, output will be one estimate of *average controllability* per brain region; a

1253 regional map of control over system dynamics. These regional values will also be positive. Greater *average controllability*

1254 indicates that regions are better positioned within the network's topology to broadcast an impulse, and as such may better

1255 orchestrate control of brain dynamics.

1256 What can researchers do with these outputs? The answers to this question are diverse and depend heavily on the re-

1257 searcher's goals. As we discussed in Section III, we have used NCT to investigate a range of research questions that

1258 spanned from examining the influence of topology [10, 49, 50], to predicting state transitions observed in functional data

1259 [14, 73], to studying individual differences, including psychosis symptoms [69], executive function [71], and sex effects

1260 [19]. Providing detailed guidance on each of these applications is beyond the scope of this protocol. However, to con-

1261 clude this protocol, we outline the use of null network models as an initial analysis that we believe is an essential step

1262 irrespective of researchers' study goals.

1263

### 1. Null network models

1264 Null models allow researchers to examine the extent to which different aspects of topology explain NCT model outputs.

1265 As discussed in Ref. [149], these null models take different forms, including edge rewiring, generative models of surrogate

1266 networks, and spatially-preserved node permutation. Of these different forms, the appropriate choice will depend on the

1267 research question. Here, in order to understand the extent to which topology informs *control energy*, we focus on null

1268 models that rewire an empirical adjacency matrix (see Ref. [69] for an example of spatially-preserved node permutation

used to compare maps of *average controllability* with other properties of network topology). In this case, a null network model involves randomly swapping the edges of the adjacency matrix  $n$  times subject to certain constraints—for example, while preserving the spatial embedding of the nodes as well as the degree or strength distribution [149, 150]—and recalculating energy upon each rewired matrix. This process generates an empirical null distribution that observed *control energy* can be compared against. For instance, *control energy* that is lower than expected under the null suggests that NCT was able to leverage properties of network topology, beyond those preserved by the null model, to complete a given state transition. In turn, by deploying a range of null models that each preserve different topological properties, researchers can systematically probe the aspects of topology that explain their observed outputs.

Using the undirected human connectome, we provide an example of the above approach using two of our binary state transitions: the default mode to visual transition and the default mode to ventral attention (VAN) transition. We chose these transitions as they represent two control tasks with strong but opposing energy asymmetries (see Figure 5). For each transition, we recompute the *control energy* for each direction under two null models. The first preserves the spatial embedding of the nodes as well as the strength distribution (strength-preserving). The second preserves spatial embedding and the strength sequence of the nodes (sequence-preserving). That is, the sequence-preserving null preserves the strength of each node as it was in the original connectome. By contrast, the strength-preserving null only preserves the distribution of strength across the network; the strength of each node is allowed to change. The sequence-preserving null is a more stringent test than the strength-preserving null as it preserves how strength is embedded in the connectome [150]. We begin by loading the coordinates of our nodes in 3 dimensions:

```
1287 1 # null networks
1288 2 centroids = pd.read_csv(
1289 3     os.path.join(datadir, "pnc_schaefer200_centroids.csv")
1290 4 ) # load coordinates of nodes
1291 5 centroids.set_index("node_names", inplace=True)
1292 6 print(centroids.head())
```

```
1293      1          vox_x  vox_y  vox_z
1294      2 node_names
1295      3 LH_Vis_1      121    149    69
1296      4 LH_Vis_2      123    174    65
1297      5 LH_Vis_3      143    166    70
1298      6 LH_Vis_4      107    164    74
1299      7 LH_Vis_5      124    192    66
```

Then, we use those coordinates to define a distance matrix that encodes the physical distance between node pairs:

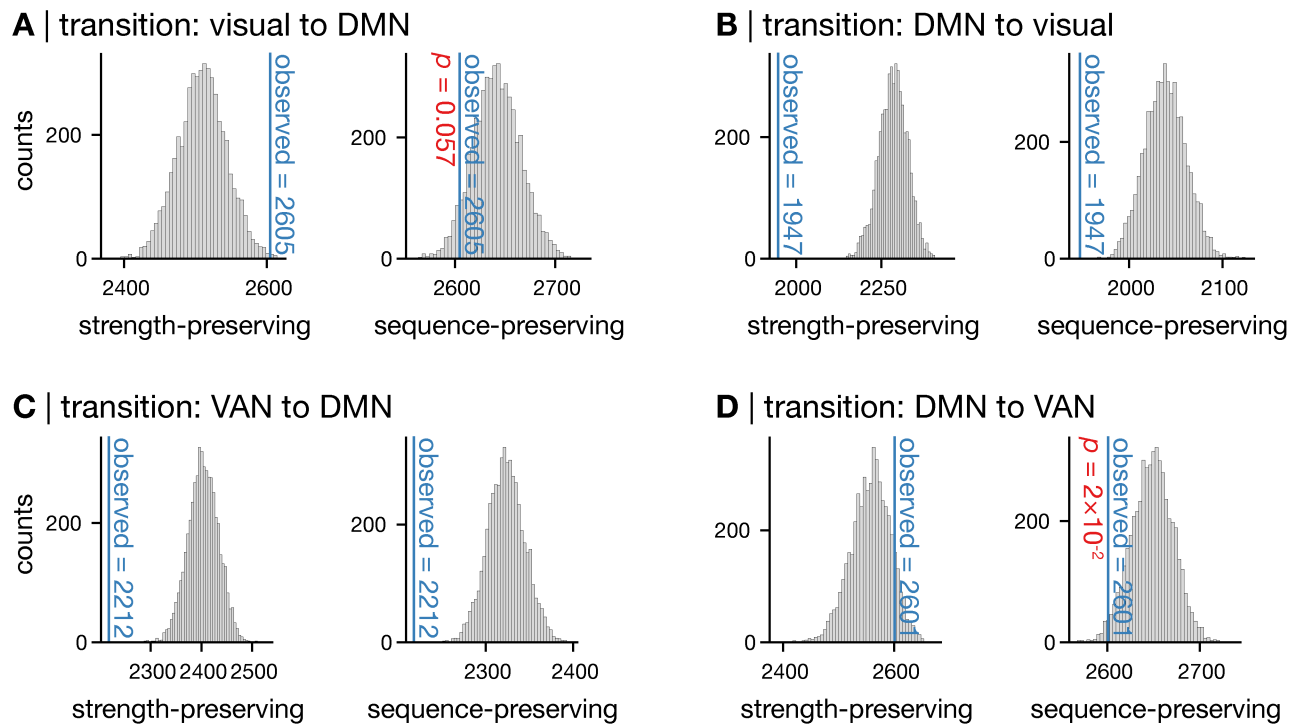
```
1301 1 distance_matrix = distance.pdist(centroids, 'euclidean') # get euclidean distances between nodes
1302 2 distance_matrix = distance.squareform(distance_matrix) # reshape to square matrix
```

Finally, we define our control task and compute our nulls using the included function, `geomsurr` [150]:

```
1304 1 # extract initial state
1305 2 initial_state = states == state_labels.index("Vis") # 'Vis' or 'SalVentAttn'
1306 3 initial_state = normalize_state(initial_state) # normalize
1307 4
1308 5 # extract target state
1309 6 target_state = states == state_labels.index("Default")
1310 7 target_state = normalize_state(target_state) # normalize
1311 8
1312 9 # compute true control energy
1313 10 _, control_signals, _ = get_control_inputs(
1314 11     A_norm=adjacency_norm,
1315 12     T=time_horizon,
1316 13     B=control_set,
1317 14     x0=initial_state,
1318 15     xf=target_state,
1319 16     system=system,
1320 17     rho=rho,
1321 18     S=trajectory_constraints,
1322 19 )
1323 20 node_energy = integrate_u(control_signals) # integrate control signals
1324 21 energy = np.sum(node_energy) # get energy
1325 22
1326 23 # run permutation
1327 24 n_perms = 5000 # number of permutations
1328 25
1329 26 # containers for null distributions
```

```
1330 27 energy_null_sp = np.zeros(n_perms)
1331 28 energy_null_ssp = np.zeros(n_perms)
1332 29
1333 30 for perm in tqdm(np.arange(n_perms)):
1334 31     # rewire adjacency matrix using geomsurr
1335 32     _, Wsp, Wssp = geomsurr(W=adjacency, D=distance_matrix, seed=perm)
1336 33     # Wsp is the adjacency matrix rewired while preserving spatial embedding and the strength
1337 34     # Wssp is the adjacency matrix rewired while preserving spatial embedding and the strength
1338 35     # sequence
1339 36     # this python implementation is included with our toolbox, but if you use these nulls
1340 37     # in your own work, please cite:
1341 38     #     Roberts et al. NeuroImage (2016), doi:10.1016/j.neuroimage.2015.09.009
1342 39
1343 40     # compute control energy for Wsp
1344 41     Wsp = matrix_normalization(Wsp, system)
1345 42     _, control_signals, _ = get_control_inputs(
1346 43         A_norm=Wsp,
1347 44         T=time_horizon,
1348 45         B=control_set,
1349 46         x0=initial_state,
1350 47         xf=target_state,
1351 48         system=system,
1352 49         rho=rho,
1353 50         S=trajectory_constraints,
1354 51     )
1355 52     node_energy = integrate_u(control_signals)
1356 53     energy_null_sp[perm] = np.sum(node_energy)
1357 54
1358 55     # compute control energy for Wssp
1359 56     Wssp = matrix_normalization(Wssp, system)
1360 57     _, control_signals, _ = get_control_inputs(
1361 58         A_norm=Wssp,
1362 59         T=time_horizon,
1363 60         B=control_set,
1364 61         x0=initial_state,
1365 62         xf=target_state,
1366 63         system=system,
1367 64         rho=rho,
1368 65         S=trajectory_constraints,
1369 66     )
1370 67     node_energy = integrate_u(control_signals)
1371 68     energy_null_ssp[perm] = np.sum(node_energy)
1372 69
1373 70 # plot
1374 71 f, ax = plt.subplots(1, 2, figsize=(7, 3))
1375 72 null_plot(
1376 73     observed=energy,
1377 74     null=energy_null_sp,
1378 75     xlabel="strength-preserving",
1379 76     ax=ax[0],
1380 77 )
1381 78 null_plot(
1382 79     observed=energy,
1383 80     null=energy_null_ssp,
1384 81     xlabel="sequence-preserving",
1385 82     ax=ax[1],
1386 83 )
1387 84 f.tight_layout()
1388 85 plt.show()
```

1390 Figure 9 displays the energy associated with transitioning from the visual cortex to the DMN (Figure 9A) and back again  
1391 (Figure 9B), as well from the VAN to the DMN (Figure 9C) and back again (Figure 9D). In each panel of Figure 9, the  
1392 strength-preserving null is shown on the left and the sequence-preserving null is shown on the right. These results provide  
1393 several insights. First, as mentioned above, both transitions show energy asymmetries but in opposite directions. The  
1394 energy associated with transitioning from visual cortex to the DMN is larger (energy = 2605) compared to the reverse  
1395 direction (energy = 1947). By contrast, the energy associated with transitioning from the VAN to the DMN is lower  
1396 (energy = 2218) compared to the reverse direction (energy = 2601). Note that the former result represents an exception to  
1397 the general finding that energy is lower when transitioning to the DMN than from it (see Figure 5). This pattern of findings



**FIG. 9**

**Null network models uncover the topological properties that are important for control energy.** For a given state transition, we recompute *control energy* 5,000 times. Each time we randomly rewired the edges of the adjacency matrix subject to certain constraints. This process generates an empirical null distribution for *control energy*. Here, we generate two null distributions per state transition; one that preserves the spatial embedding of system nodes as well as the strength distribution (strength-preserving), and another that preserves spatial embedding and the strength sequence of the nodes (sequence-preserving). We computed these nulls for the visual-to-DMN transition (A), the DMN-to-visual transition (B), the VAN-to-DMN transition (C), and the DMN-to-VAN transition (D). Collectively, these results demonstrate that *control energy* for some transitions is likely driven by strength (e.g., visual-to-DMN and DMN-to-VAN) while others may be driven by higher-order topology (e.g., DMN-to-visual and VAN-to-DMN). See main text for extended discussion.

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is consistent with what we observe in the mouse connectome (Figure 7).

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Second, when we preserve the strength distribution in our null model, we observe larger-than-expected energy when transitioning from the visual cortex to the DMN (Figure 9A, left). This result appears counter intuitive until we consider the differences in strength between these brain states. In the structural connectome used here, the mean strength of the nodes within the visual state is 67,773, while the mean strength of the nodes in the DMN is 59,455, and the mean strength of the remaining nodes is 53,761. Thus, the connectivity strength between the visual state and the rest of the brain is higher than the connectivity strength between the DMN and the rest of the brain. When we preserve only the strength distribution in the null, this difference in strength is not maintained, which results in relatively high-strength nodes being redistributed throughout the brain. In turn, this redistribution results in reduced *control energy* in the null distribution for the visual-to-DMN transition. This finding suggests that the high strength nodes of the visual system broadcast activity in a way that necessitates high amounts of *control energy* to guide those dynamics toward the DMN. By contrast, when we preserve the strength sequence (Figure 9A, right) we also preserve the between-state difference in strength, which yields a null that is much closer to the observed energy. Together, the results in Figure 9A demonstrate that differences in connectivity strength between brain states drives the energy observed for the visual-to-DMN transition.

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Third, the above line of reasoning does not hold when we consider the transition from DMN back to visual cortex (Figure 9B). Here, we observe lower-than-expected energy under both the strength-preserving and the sequence-preserving nulls. This result demonstrates that the aforementioned difference in strength between the visual state and the DMN is not what drives observed energy. In turn, this result suggests that higher-order topological properties of the connectome may support the efficient completion of the DMN-to-visual transition. Finally, Figure 9C and D show that all of the above results and interpretations vary as a function of states.

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The above findings—that the spatial embedding of node strength drives energy for one transition direction but not the other—underscores the utility of probing NCT outputs using null network models. That is, through comparing a pair of null network models, we obtained evidence for how certain aspects of network topology (i.e., strength) contribute to



1421 different state transitions.

1422 Null network models can also be applied to *average controllability*:

```
1423 1 # run permutation
1424 2 n_perms = 5000 # number of permutations
1425 3
1426 4 # container for null distribution
1427 5 ave_ctrb_null = np.zeros((n_perms, n_nodes))
1428 6
1429 7 for perm in tqdm(np.arange(n_perms)):
1430 8     # rewire adjacency matrix using geomsurr
1431 9     _, _, Wssp = geomsurr(W=adjacency, D=distance_matrix, seed=perm)
1432 10
1433 11     # compute average controllability
1434 12     Wssp = matrix_normalization(Wssp, system)
1435 13     ave_ctrb_null[perm, :] = ave_control(Wssp, system)
```

1436 The above code will generate an empirical null distribution for *average controllability* at each system node. In turn, this  
1437 procedure will yield  $N$  null distributions for a given null network model, the visualization of which is impractical. As  
1438 such, researchers may instead assign  $p$ -values to the observed *average controllability* values using `get_null_p`:

```
1439 1 # calculate p-values
1440 2 p_vals_ssp = np.zeros(n_nodes)
1441 3
1442 4 for node in tqdm(np.arange(n_nodes)):
1443 5     # version='standard' will calculate the number of times the null is larger than the observed
1444 6     # value
1445 7     # version='reverse' will calculate the number of times the null is smaller than the observed
1446 8     # value
1447 9     p_vals_ssp[node] = get_null_p(
1448 10         average_controllability[node], ave_ctrb_null[:, node], version="standard"
1449 11     )
1450 12
1451 13 p_vals_ssp = get_fdr_p(p_vals_ssp) # correct p values for multiple comparisons
```

1452 The above code will yield FDR-corrected  $p$ -values denoting the proportion of times that a nodes' *average controllability*  
1453 was larger than expected under the null. As touched upon above, this result only tells half the story, and there may be  
1454 reasons for *average controllability* to be smaller than expected under the null. As such, we recommend running both  
1455 `version='standard'` and `version='reverse'` to test both tails of the null distribution.

1456

## XI. TIMING

1457 As noted throughout the protocol, the timing of each step is relatively short, often not exceeding 1 second per step. We  
1458 note two clarifications. First, these time estimates are only for a single execution of each step as shown in the protocol.  
1459 In reality, these steps will likely need to be executed many times over to achieve researchers' goals. For example, a given  
1460 study may need to compute control energy for multiple control tasks across multiple subjects, which will increase run  
1461 time. This time will increase further if null network models are used, wherein each step may be run thousands of times  
1462 for a single control task. However, in these instances, protocol steps can be trivially parallelized using High Performance  
1463 Computing (HPC), which will reduce run time. Second, timing will vary as a function of researchers' data processing.  
1464 For example, in this protocol, we performed analysis on a structural connectome comprising 200 nodes. Increasing  
1465 parcellation resolution will increase run time.

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1488  
1489 **Author contributions**

1490 Conceptualization: L.P., J.Z.K., T.D.S., and D.S.B.

1491 Methodology: L.P., J.Z.K., J.S., and D.S.B.

1492 Software: L.P., J.Z.K., and J.S.

1493 Formal analysis: L.P., and J.Z.K.

1494 Visualization: L.P., and J.Z.K.

1495 Data curation: J.K.B., M.C., S.C., R.E.G., R.C.G., R.T.S., D.Z., and T.D.S.

1496 Writing—original draft: L.P., and J.Z.K.

1497 Writing—reviewing and editing: L.P., J.Z.K., J.S., J.K.B., M.C., S.C., R.E.G., R.C.G., F.P., R.T.S., D.Z., T.D.S., and D.S.B.

1498  
1499 **Competing interests**

1500 R.T.S. receives consulting compensation from Octave Bioscience and compensation for reviewership duties from the  
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1502  
1503 **Data availability**

1504 The PNC data are publicly available in the Database of Genotypes and Phenotypes: accession number: phs00607.v3.p2;  
1505 [https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study\\_id=phs00607.v3.p2](https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs00607.v3.p2)  
1506 p2

1507  
1508 **Code availability**

1509 All analysis code is freely available at <https://github.com/BassettLab/nctpy/>

REFERENCES

1510  
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1551  
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1553  
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1555  
1556  
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1561  
1562

1. Bassett, D. S. & Sporns, O. Network neuroscience. en. *Nature Neuroscience* **20**, 353–364. ISSN: 1097-6256, 1546-1726. <http://www.nature.com/articles/nn.4502> (2022-04-12) (Mar. 2017).
2. Bassett, D. S., Zurn, P. & Gold, J. I. On the nature and use of models in network neuroscience. en. *Nature Reviews Neuroscience* **19**, 566–578. ISSN: 1471-003X, 1471-0048. <http://www.nature.com/articles/s41583-018-0038-8> (Sept. 2018).
3. Betzel, R. F. & Bassett, D. S. Multi-scale brain networks. en. *NeuroImage* **160**, 73–83. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811916306152> (Oct. 2017).
4. Fornito, A., Zalesky, A. & Bullmore, E. T. *Fundamentals of Brain Network Analysis* ISBN: 9780124079083 (Academic Press Elsevier, 2016).
5. Menara, T., Katewa, V., Bassett, D. S. & Pasqualetti, F. *The Structured Controllability Radius of Symmetric (Brain) Networks in 2018 Annual American Control Conference (ACC) 2018 Annual American Control Conference (ACC)* (IEEE, Milwaukee, WI, 2018), 2802–2807. ISBN: 978-1-5386-5428-6. <https://ieeexplore.ieee.org/document/8431724/>.
6. Pasqualetti, F., Zampieri, S. & Bullo, F. Controllability Metrics, Limitations and Algorithms for Complex Networks. *IEEE Transactions on Control of Network Systems* **1**, 40–52. ISSN: 2325-5870. <http://ieeexplore.ieee.org/document/6762966/> (2014).
7. Kim, J. Z. & Bassett, D. S. in *Neural Engineering* (ed He, B.) 497–518 (Springer International Publishing, Cham, 2020). ISBN: 978-3-030-43395-6. [https://doi.org/10.1007/978-3-030-43395-6\\_17](https://doi.org/10.1007/978-3-030-43395-6_17).
8. Karrer, T. M. *et al.* A practical guide to methodological considerations in the controllability of structural brain networks. *Journal of Neural Engineering* **17**, 026031. ISSN: 1741-2552. <https://iopscience.iop.org/article/10.1088/1741-2552/ab6e8b> (2020).
9. Seguin, C., Sporns, O. & Zalesky, A. Brain network communication: concepts, models and applications. en. *Nature Reviews Neuroscience*. ISSN: 1471-003X, 1471-0048. <https://www.nature.com/articles/s41583-023-00718-5> (July 2023).
10. Gu, S. *et al.* Controllability of structural brain networks. *Nature Communications* **6**, 8414 (2015).
11. Gu, S. *et al.* Optimal trajectories of brain state transitions. *NeuroImage* **148**, 305–317. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811917300058> (2017).
12. Tang, E. *et al.* Developmental increases in white matter network controllability support a growing diversity of brain dynamics. *Nature Communications* **8**, 1252. ISSN: 2041-1723. <http://www.nature.com/articles/s41467-017-01254-4> (2017).
13. Tang, E. *et al.* Control of brain network dynamics across diverse scales of space and time. en. *Physical Review E* **101**, 062301. ISSN: 2470-0045, 2470-0053. <https://link.aps.org/doi/10.1103/PhysRevE.101.062301> (June 2020).
14. Stiso, J. *et al.* White Matter Network Architecture Guides Direct Electrical Stimulation through Optimal State Transitions. *Cell Reports* **28**, 2554–2566.e7. ISSN: 22111247. <https://linkinghub.elsevier.com/retrieve/pii/S2211124719310411> (2019).
15. Scheid, B. H. *et al.* Time-evolving controllability of effective connectivity networks during seizure progression. *Proceedings of the National Academy of Sciences* **118**, e2006436118. ISSN: 0027-8424, 1091-6490. <https://pnas.org/doi/full/10.1073/pnas.2006436118> (2021).
16. Medaglia, J. D. *et al.* Network Controllability in the Inferior Frontal Gyrus Relates to Controlled Language Variability and Susceptibility to TMS. en. *The Journal of Neuroscience* **38**, 6399–6410. ISSN: 0270-6474, 1529-2401. <https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.0092-17.2018> (July 2018).
17. Medaglia, J. D. *et al.* Language Tasks and the Network Control Role of the Left Inferior Frontal Gyrus. en. *eneuro* **8**, ENEURO.0382–20.2021. ISSN: 2373-2822. <https://www.eneuro.org/lookup/doi/10.1523/ENEURO.0382-20.2021> (Sept. 2021).
18. Muldoon, S. F. *et al.* Stimulation-Based Control of Dynamic Brain Networks. en. *PLOS Computational Biology* **12** (ed Hilgetag, C. C.) e1005076. ISSN: 1553-7358. <https://dx.plos.org/10.1371/journal.pcbi.1005076> (Sept. 2016).
19. Cornblath, E. J. *et al.* Sex differences in network controllability as a predictor of executive function in youth. *NeuroImage* **188**, 122–134. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811918321293> (2019).
20. Cornblath, E. J. *et al.* Temporal sequences of brain activity at rest are constrained by white matter structure and modulated by cognitive demands. *Communications Biology* **3**, 261. ISSN: 2399-3642. <http://www.nature.com/articles/s42003-020-0961-x> (2020).
21. Satterthwaite, T. D. *et al.* Neuroimaging of the Philadelphia Neurodevelopmental Cohort. en. *NeuroImage* **86**, 544–553. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811913008331> (Feb. 2014).

- 1563 22. Satterthwaite, T. D. *et al.* The Philadelphia Neurodevelopmental Cohort: A publicly available resource for the study of normal  
1564 and abnormal brain development in youth. en. *NeuroImage* **124**, 1115–1119. ISSN: 10538119. [https://linkinghub.  
1565 elsevier.com/retrieve/pii/S1053811915002529](https://linkinghub.elsevier.com/retrieve/pii/S1053811915002529) (Jan. 2016).
- 1566 23. Oh, S. W. *et al.* A mesoscale connectome of the mouse brain. *Nature* **508**, 207–214 (2014).
- 1567 24. Hierarchical organization of cortical and thalamic connectivity. en. *Nature* **575**, 195–202. ISSN: 0028-0836, 1476-4687. [https:  
1568 //www.nature.com/articles/s41586-019-1716-z](https://www.nature.com/articles/s41586-019-1716-z) (Nov. 2019).
- 1569 25. Knox, J. E. *et al.* High-resolution data-driven model of the mouse connectome. en. *Network Neuroscience* **3**, 217–236. ISSN:  
1570 2472-1751. <https://direct.mit.edu/netn/article/3/1/217-236/2194> (Jan. 2019).
- 1571 26. Chiêm, B., Crevecoeur, F. & Delvenne, J.-C. Structure-informed functional connectivity driven by identifiable and state-specific  
1572 control regions. en. *Network Neuroscience* **5**, 591–613. ISSN: 2472-1751. [https://direct.mit.edu/netn/article/  
1573 5/2/591/98351/Structure-informed-functional-connectivity-driven](https://direct.mit.edu/netn/article/5/2/591/98351/Structure-informed-functional-connectivity-driven) (June 2021).
- 1574 27. Jeganathan, J. *et al.* Fronto-limbic dysconnectivity leads to impaired brain network controllability in young people with bipolar  
1575 disorder and those at high genetic risk. en. *NeuroImage: Clinical* **19**, 71–81. ISSN: 22131582. [https://linkinghub.  
1576 elsevier.com/retrieve/pii/S2213158218301025](https://linkinghub.elsevier.com/retrieve/pii/S2213158218301025) (2018).
- 1577 28. Kenett, Y. N. *et al.* Driving the brain towards creativity and intelligence: A network control theory analysis. en. *Neuropsychologia*  
1578 **118**, 79–90. ISSN: 00283932. [https://linkinghub.elsevier.com/retrieve/pii/  
1579 S0028393218300010](https://linkinghub.elsevier.com/retrieve/pii/S0028393218300010) (Sept. 2018).
- 1580 29. Yuan, J., Ji, S., Luo, L., Lv, J. & Liu, T. Control energy assessment of spatial interactions among  
1581 macro-scale brain networks. en. *Human Brain Mapping* **43**, 2181–2203. ISSN: 1065-9471, 1097-0193. [https:  
1582 //onlinelibrary.wiley.com/doi/10.1002/hbm.25780](https://onlinelibrary.wiley.com/doi/10.1002/hbm.25780) (May 2022).
- 1583 30. Singleton, S. P. *et al.* Receptor-informed network control theory links LSD and psilocybin to a flattening of the brain’s control  
1584 energy landscape. en. *Nature Communications* **13**, 5812. ISSN: 2041-1723. [https://www.nature.com/articles/  
1585 s41467-022-33578-1](https://www.nature.com/articles/s41467-022-33578-1) (Oct. 2022).
- 1586 31. Singleton, S. P. *et al.* Time-resolved network control analysis links reduced control energy under DMT with the serotonin 2a re-  
1587 ceptor, signal diversity, and subjective experience en. preprint (Neuroscience, May 2023). [http://biorxiv.org/lookup/  
1588 doi/10.1101/2023.05.11.540409](http://biorxiv.org/lookup/doi/10.1101/2023.05.11.540409).
- 1589 32. Luppi, A. I. *et al.* Transitions between cognitive topographies: contributions of network structure, neuromodulation, and disease  
1590 en. preprint (Neuroscience, Mar. 2023). <http://biorxiv.org/lookup/doi/10.1101/2023.03.16.532981>.
- 1591 33. Maxwell, J. C. I. On governors. *Proceedings of the Royal Society of London*, 270–283 (1868).
- 1592 34. Grasser, F., D’arrigo, A., Colombi, S. & Rufer, A. C. JOE: a mobile, inverted pendulum. *IEEE Transactions on industrial elec-  
1593 tronics* **49**, 107–114 (2002).
- 1594 35. Hodgkin, A. L. & Huxley, A. F. A quantitative description of membrane current and its application to conduction and excitation  
1595 in nerve. *The Journal of physiology* **117**, 500 (1952).
- 1596 36. Papadopoulos, L., Kim, J. Z., Kurths, J. & Bassett, D. S. Development of structural correlations and synchronization from  
1597 adaptive rewiring in networks of Kuramoto oscillators. *Chaos: An Interdisciplinary Journal of Nonlinear Science* **27**, 073115  
1598 (2017).
- 1599 37. Wilson, H. R. & Cowan, J. D. Excitatory and inhibitory interactions in localized populations of model neurons. *Biophysical  
1600 journal* **12**, 1–24 (1972).
- 1601 38. Schiff, S. J. *et al.* Controlling chaos in the brain. *Nature* **370**, 615–620 (1994).
- 1602 39. Suárez, L. E., Markello, R. D., Betzel, R. F. & Misic, B. Linking Structure and Function in Macroscale Brain Networks. en.  
1603 *Trends in Cognitive Sciences* **24**, 302–315. ISSN: 13646613. [https://linkinghub.elsevier.com/retrieve/pii/  
1604 S1364661320300267](https://linkinghub.elsevier.com/retrieve/pii/S1364661320300267) (Apr. 2020).
- 1605 40. Vázquez-Rodríguez, B. *et al.* Gradients of structure–function tethering across neocortex. en. *Proceedings of the National  
1606 Academy of Sciences* **116**, 21219–21227. ISSN: 0027-8424, 1091-6490. [https://pnas.org/doi/full/10.1073/  
1607 pnas.1903403116](https://pnas.org/doi/full/10.1073/pnas.1903403116) (Oct. 2019).
- 1608 41. Baum, G. L. *et al.* Development of structure–function coupling in human brain networks during youth. en. *Proceedings of the  
1609 National Academy of Sciences* **117**, 771–778. ISSN: 0027-8424, 1091-6490. [http://www.pnas.org/lookup/doi/10.  
1610 1073/pnas.1912034117](http://www.pnas.org/lookup/doi/10.1073/pnas.1912034117) (Jan. 2020).
- 1611 42. Preti, M. G. & Van De Ville, D. Decoupling of brain function from structure reveals regional behavioral specialization in humans.  
1612 en. *Nature Communications* **10**, 4747. ISSN: 2041-1723. [http://www.nature.com/articles/s41467-019-  
1613 12765-7](http://www.nature.com/articles/s41467-019-12765-7) (Dec. 2019).
- 1614 43. Luo, N. *et al.* Structural Brain Architectures Match Intrinsic Functional Networks and Vary across Domains: A Study from 15  
1615 000+ Individuals. en. *Cerebral Cortex* **30**, 5460–5470. ISSN: 1047-3211, 1460-2199. [https://academic.oup.com/  
1616 cercor/article/30/10/5460/5850539](https://academic.oup.com/cercor/article/30/10/5460/5850539) (Sept. 2020).

- 1617 44. Seguin, C., Tian, Y. & Zalesky, A. Network communication models improve the behavioral and functional predictive utility of  
1618 the human structural connectome. en. *Network Neuroscience* **4**, 980–1006. ISSN: 2472-1751. [https://direct.mit.edu/  
1619 netn/article/4/4/980-1006/95847](https://direct.mit.edu/netn/article/4/4/980-1006/95847) (Jan. 2020).
- 1620 45. Betzel, R. F., Faskowitz, J., Mišić, B., Sporns, O. & Seguin, C. Multi-policy models of interregional communication in the  
1621 human connectome. en. *Network Neuroscience* **22** (2022).
- 1622 46. Fox, P. T. & Friston, K. J. Distributed processing; distributed functions? en. *NeuroImage* **61**, 407–426. ISSN: 10538119. [https://  
1623 //linkinghub.elsevier.com/retrieve/pii/S1053811911014534](https://linkinghub.elsevier.com/retrieve/pii/S1053811911014534) (June 2012).
- 1624 47. Hespanha, J. P. *Linear Systems Theory* ISBN13: 9780691179575 (Princeton Press, Princeton, New Jersey, Feb. 2018).
- 1625 48. Yan, G. *et al.* Network control principles predict neuron function in the *Caenorhabditis elegans* connectome. *Nature* **550**, 519–  
1626 523. ISSN: 0028-0836, 1476-4687. <http://www.nature.com/articles/nature24056> (2017).
- 1627 49. Betzel, R. F., Gu, S., Medaglia, J. D., Pasqualetti, F. & Bassett, D. S. Optimally controlling the human connectome: the role of  
1628 network topology. *Scientific Reports* **6**, 30770. ISSN: 2045-2322. <http://www.nature.com/articles/srep30770>  
1629 (2016).
- 1630 50. Kim, J. Z. *et al.* Role of graph architecture in controlling dynamical networks with applications to neural systems. *Nature Physics*  
1631 **14**, 91–98. ISSN: 1745-2473, 1745-2481. <http://www.nature.com/articles/nphys4268> (2018).
- 1632 51. Parkes, L. *et al.* Asymmetric signaling across the hierarchy of cytoarchitecture within the human connectome. en. *Science Ad-  
1633 vances* **8**, eadd2185. ISSN: 2375-2548. <https://www.science.org/doi/10.1126/sciadv.add2185> (Dec.  
1634 2022).
- 1635 52. Sejnowski, T. J., Churchland, P. S. & Movshon, J. A. Putting big data to good use in neuroscience. en. *Nature Neuroscience* **17**,  
1636 1440–1441. ISSN: 1097-6256, 1546-1726. <http://www.nature.com/articles/nn.3839> (Nov. 2014).
- 1637 53. Tong, F. Primary visual cortex and visual awareness. en. *Nature Reviews Neuroscience* **4**, 219–229. ISSN: 1471-003X, 1471-0048.  
1638 <http://www.nature.com/articles/nrn1055> (Mar. 2003).
- 1639 54. Gordon, E. M. *et al.* A somato-cognitive action network alternates with effector regions in motor cortex. en. *Nature* **617**, 351–359.  
1640 ISSN: 0028-0836, 1476-4687. <https://www.nature.com/articles/s41586-023-05964-2> (2023) (May 2023).
- 1641 55. Bertolero, M. A., Yeo, B. T. T. & D’Esposito, M. The modular and integrative functional architecture of the human brain. en.  
1642 *Proceedings of the National Academy of Sciences* **112**. ISSN: 0027-8424, 1091-6490. [https://pnas.org/doi/full/  
1643 10.1073/pnas.1510619112](https://pnas.org/doi/full/10.1073/pnas.1510619112) (Dec. 2015).
- 1644 56. Bertolero, M. A., Yeo, B. T. T., Bassett, D. S. & D’Esposito, M. A mechanistic model of connector hubs, modularity and  
1645 cognition. en. *Nature Human Behaviour* **2**, 765–777. ISSN: 2397-3374. [http://www.nature.com/articles/s41562-  
1646 018-0420-6](http://www.nature.com/articles/s41562-018-0420-6) (Oct. 2018).
- 1647 57. Van den Heuvel, M. P. & Sporns, O. Network hubs in the human brain. en. *Trends in Cognitive Sciences* **17**, 683–696. ISSN:  
1648 13646613. <https://linkinghub.elsevier.com/retrieve/pii/S1364661313002167> (Dec. 2013).
- 1649 58. Van den Heuvel, M. P. & Sporns, O. Rich-Club Organization of the Human Connectome. en. *Journal of Neuroscience* **31**, 15775–  
1650 15786. ISSN: 0270-6474, 1529-2401. [https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.3539-  
1651 11.2011](https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.3539-11.2011) (Nov. 2011).
- 1652 59. Fornito, A., Zalesky, A. & Breakspear, M. The connectomics of brain disorders. en. *Nature Reviews Neuroscience* **16**, 159–172.  
1653 ISSN: 1471-003X, 1471-0048. <http://www.nature.com/articles/nrn3901> (Mar. 2015).
- 1654 60. Crossley, N. A. *et al.* The hubs of the human connectome are generally implicated in the anatomy of brain disorders. en. *Brain*  
1655 **137**, 2382–2395. ISSN: 1460-2156, 0006-8950. [https://academic.oup.com/brain/article-lookup/doi/10.  
1656 1093/brain/awu132](https://academic.oup.com/brain/article-lookup/doi/10.1093/brain/awu132) (Aug. 2014).
- 1657 61. Van Essen, D. C. *et al.* The WU-Minn Human Connectome Project: An overview. en. *NeuroImage* **80**, 62–79. ISSN: 10538119.  
1658 <https://linkinghub.elsevier.com/retrieve/pii/S1053811913005351> (Oct. 2013).
- 1659 62. Casey, B. *et al.* The Adolescent Brain Cognitive Development (ABCD) study: Imaging acquisition across 21 sites. en. *Develop-  
1660 mental Cognitive Neuroscience* **32**, 43–54. ISSN: 18789293. [https://linkinghub.elsevier.com/retrieve/pii/  
1661 S1878929317301214](https://linkinghub.elsevier.com/retrieve/pii/S1878929317301214) (Aug. 2018).
- 1662 63. Alexander, L. M. *et al.* An open resource for transdiagnostic research in pediatric mental health and learning disorders. en.  
1663 *Scientific Data* **4**, 170181. ISSN: 2052-4463. <http://www.nature.com/articles/sdata2017181> (Dec. 2017).
- 1664 64. Amunts, K. *et al.* BigBrain: An Ultrahigh-Resolution 3D Human Brain Model. en. *Science* **340**, 1472–1475. ISSN: 0036-8075,  
1665 1095-9203. <https://www.science.org/doi/10.1126/science.1235381> (June 2013).
- 1666 65. Hawrylycz, M. J. *et al.* An anatomically comprehensive atlas of the adult human brain transcriptome. en. *Nature* **489**, 391–399.  
1667 ISSN: 0028-0836, 1476-4687. <http://www.nature.com/articles/nature11405> (Sept. 2012).
- 1668 66. Thomas Yeo, B. T. *et al.* The organization of the human cerebral cortex estimated by intrinsic functional connectivity. en. *Journal  
1669 of Neurophysiology* **106**, 1125–1165. ISSN: 0022-3077, 1522-1598. [https://www.physiology.org/doi/10.1152/  
1670 jn.00338.2011](https://www.physiology.org/doi/10.1152/jn.00338.2011) (Sept. 2011).



- 1671 67. De Reus, M. A. & van den Heuvel, M. P. Simulated rich club lesioning in brain networks: a scaffold for communication and  
1672 integration? *Frontiers in Human Neuroscience* **8**, ISSN: 1662-5161. [http://journal.frontiersin.org/article/](http://journal.frontiersin.org/article/10.3389/fnhum.2014.00647/abstract)  
1673 [10.3389/fnhum.2014.00647/abstract](http://journal.frontiersin.org/article/10.3389/fnhum.2014.00647/abstract) (Aug. 2014).
- 1674 68. Van den Heuvel, M. P. & Sporns, O. An Anatomical Substrate for Integration among Functional Networks in Human Cortex.  
1675 en. *Journal of Neuroscience* **33**, 14489–14500. ISSN: 0270-6474, 1529-2401. [https://www.jneurosci.org/lookup/](https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.2128-13.2013)  
1676 [doi/10.1523/JNEUROSCI.2128-13.2013](https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.2128-13.2013) (Sept. 2013).
- 1677 69. Parkes, L. *et al.* Network Controllability in Transmodal Cortex Predicts Psychosis Spectrum Symptoms. *Biological Psychiatry*  
1678 **89**, S370–S371 (2021).
- 1679 70. Margulies, D. S. *et al.* Situating the default-mode network along a principal gradient of macroscale cortical organization. en.  
1680 *Proceedings of the National Academy of Sciences* **113**, 12574–12579. ISSN: 0027-8424, 1091-6490. [http://www.pnas.](http://www.pnas.org/lookup/doi/10.1073/pnas.1608282113)  
1681 [org/lookup/doi/10.1073/pnas.1608282113](http://www.pnas.org/lookup/doi/10.1073/pnas.1608282113) (Nov. 2016).
- 1682 71. Cui, Z. *et al.* Optimization of energy state transition trajectory supports the development of executive function during youth. *eLife*  
1683 **9**, e53060. ISSN: 2050-084X. <https://elifesciences.org/articles/53060> (2020).
- 1684 72. Niendam, T. A. *et al.* Meta-analytic evidence for a superordinate cognitive control network subserving diverse executive func-  
1685 tions. en. *Cognitive, Affective, & Behavioral Neuroscience* **12**, 241–268. ISSN: 1530-7026, 1531-135X. [http://link.](http://link.springer.com/10.3758/s13415-011-0083-5)  
1686 [springer.com/10.3758/s13415-011-0083-5](http://link.springer.com/10.3758/s13415-011-0083-5) (June 2012).
- 1687 73. Braun, U. *et al.* Brain network dynamics during working memory are modulated by dopamine and diminished in schizophrenia.  
1688 *Nature Communications* **12**, 3478. ISSN: 2041-1723. <http://www.nature.com/articles/s41467-021-23694-9>  
1689 (2021).
- 1690 74. Parkes, L., Fulcher, B. D., Yücel, M. & Fornito, A. Transcriptional signatures of connectomic subregions of the human striatum.  
1691 en. *Genes, Brain and Behavior* **16**, 647–663. ISSN: 16011848. [https://onlinelibrary.wiley.com/doi/10.1111/](https://onlinelibrary.wiley.com/doi/10.1111/gbb.12386)  
1692 [gbb.12386](https://onlinelibrary.wiley.com/doi/10.1111/gbb.12386) (Sept. 2017).
- 1693 75. Fulcher, B. D., Murray, J. D., Zerbi, V. & Wang, X.-J. Multimodal gradients across mouse cortex. en. *Proceedings of the National*  
1694 *Academy of Sciences* **116**, 4689–4695. ISSN: 0027-8424, 1091-6490. [http://www.pnas.org/lookup/doi/10.1073/](http://www.pnas.org/lookup/doi/10.1073/pnas.1814144116)  
1695 [pnas.1814144116](http://www.pnas.org/lookup/doi/10.1073/pnas.1814144116) (Mar. 2019).
- 1696 76. Fulcher, B. D. & Fornito, A. A transcriptional signature of hub connectivity in the mouse connectome. *Proceedings of the*  
1697 *National Academy of Sciences* **113**, 1435–1440. ISSN: 0027-8424, 1091-6490. [https://pnas.org/doi/full/10.](https://pnas.org/doi/full/10.1073/pnas.1513302113)  
1698 [1073/pnas.1513302113](https://pnas.org/doi/full/10.1073/pnas.1513302113) (2016).
- 1699 77. Larivière, S. *et al.* Microstructure-Informed Connectomics: Enriching Large-Scale Descriptions of Healthy and Diseased Brains.  
1700 en. *Brain Connectivity* **9**, 113–127. ISSN: 2158-0014, 2158-0022. [https://www.liebertpub.com/doi/10.1089/](https://www.liebertpub.com/doi/10.1089/brain.2018.0587)  
1701 [brain.2018.0587](https://www.liebertpub.com/doi/10.1089/brain.2018.0587) (Mar. 2019).
- 1702 78. Arnatkevičiūtė, A., Fulcher, B. D. & Fornito, A. A practical guide to linking brain-wide gene expression and neuroimaging  
1703 data. en. *NeuroImage* **189**, 353–367. ISSN: 10538119. [https://linkinghub.elsevier.com/retrieve/pii/](https://linkinghub.elsevier.com/retrieve/pii/S1053811919300114)  
1704 [S1053811919300114](https://linkinghub.elsevier.com/retrieve/pii/S1053811919300114) (Apr. 2019).
- 1705 79. Arnatkevičiūtė, A. *et al.* Genetic influences on hub connectivity of the human connectome. en. *Nature Communications* **12**, 4237.  
1706 ISSN: 2041-1723. <http://www.nature.com/articles/s41467-021-24306-2> (Dec. 2021).
- 1707 80. Arnatkevičiūtė, A., Fulcher, B. D., Pockock, R. & Fornito, A. Hub connectivity, neuronal diversity, and gene expression in the  
1708 *Caenorhabditis elegans* connectome. en. *PLOS Computational Biology* **14** (ed Inman, C.) e1005989. ISSN: 1553-7358. [https://dx.plos.org/10.1371/](https://dx.plos.org/10.1371/journal.pcbi.1005989)  
1709 [journal.pcbi.1005989](https://dx.plos.org/10.1371/journal.pcbi.1005989) (Feb. 2018).
- 1710 81. Anderson, K. M. *et al.* Gene expression links functional networks across cortex and striatum. en. *Nature Communications* **9**,  
1711 1428. ISSN: 2041-1723. <http://www.nature.com/articles/s41467-018-03811-x> (Dec. 2018).
- 1712 82. Anderson, K. M. *et al.* Convergent molecular, cellular, and cortical neuroimaging signatures of major depressive disorder. en.  
1713 *Proceedings of the National Academy of Sciences* **117**, 25138–25149. ISSN: 0027-8424, 1091-6490. [https://pnas.org/](https://pnas.org/doi/full/10.1073/pnas.2008004117)  
1714 [doi/full/10.1073/pnas.2008004117](https://pnas.org/doi/full/10.1073/pnas.2008004117) (Oct. 2020).
- 1715 83. Paquola, C. *et al.* Microstructural and functional gradients are increasingly dissociated in transmodal cortices. en. *PLOS Biology*  
1716 **17** (ed Kennedy, H.) e3000284. ISSN: 1545-7885. [https://dx.plos.org/10.1371/](https://dx.plos.org/10.1371/journal.pbio.3000284)  
1717 [journal.pbio.3000284](https://dx.plos.org/10.1371/journal.pbio.3000284) (May 2019).
- 1718 84. García-Cabezas, M. Á., Zikopoulos, B. & Barbas, H. The Structural Model: a theory linking connections, plasticity, pathology,  
1719 development and evolution of the cerebral cortex. *Brain Structure and Function* **224**, 985–1008. ISSN: 1863-2653, 1863-2661.  
1720 <http://link.springer.com/10.1007/s00429-019-01841-9> (2019).
- 1721 85. Yarkoni, T., Poldrack, R. A., Nichols, T. E., Van Essen, D. C. & Wager, T. D. Large-scale automated synthesis of human func-  
1722 tional neuroimaging data. en. *Nature Methods* **8**, 665–670. ISSN: 1548-7091, 1548-7105. [https://www.nature.com/](https://www.nature.com/articles/nmeth.1635)  
1723 [articles/nmeth.1635](https://www.nature.com/articles/nmeth.1635) (Aug. 2011).
- 1724 86. Propagation of electrical signals along giant nerve fibres. en. *Proceedings of the Royal Society of London. Series B - Biological*  
1725 *Sciences* **140**, 177–183. ISSN: 2053-9193. [https://royalsocietypublishing.org/doi/10.1098/rspb.1952.](https://royalsocietypublishing.org/doi/10.1098/rspb.1952.0054)  
1726 [0054](https://royalsocietypublishing.org/doi/10.1098/rspb.1952.0054) (Oct. 1952).

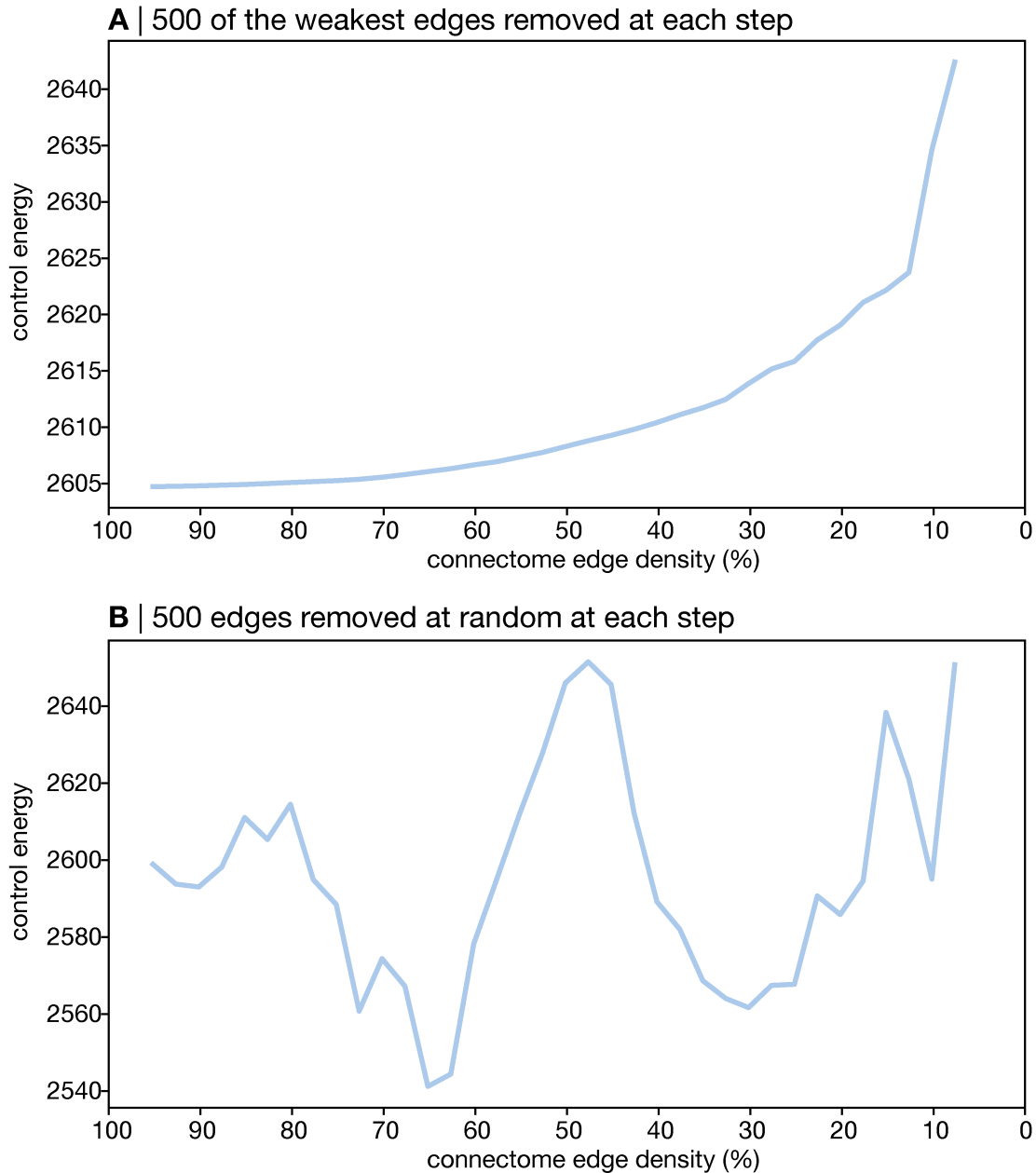
- 1727 87. Breakspear, M. Dynamic models of large-scale brain activity. en. *Nature Neuroscience* **20**, 340–352. ISSN: 1097-6256, 1546-  
1728 1726. <http://www.nature.com/articles/nn.4497> (Mar. 2017).
- 1729 88. Shine, J. M. *et al.* Computational models link cellular mechanisms of neuromodulation to large-scale neural dynamics. en.  
1730 *Nature Neuroscience* **24**, 765–776. ISSN: 1097-6256, 1546-1726. [http://www.nature.com/articles/s41593-](http://www.nature.com/articles/s41593-021-00824-6)  
1731 021-00824-6 (June 2021).
- 1732 89. Papadopoulos, L., Kim, J. Z., Kurths, J. & Bassett, D. S. Development of structural correlations and synchronization from  
1733 adaptive rewiring in networks of Kuramoto oscillators. en. *Chaos: An Interdisciplinary Journal of Nonlinear Science* **27**, 073115.  
1734 ISSN: 1054-1500, 1089-7682. <http://aip.scitation.org/doi/10.1063/1.4994819> (July 2017).
- 1735 90. Lu, Z. & Bassett, D. S. Invertible generalized synchronization: A putative mechanism for implicit learning in neural systems.  
1736 en. *Chaos: An Interdisciplinary Journal of Nonlinear Science* **30**, 063133. ISSN: 1054-1500, 1089-7682. [http://aip.](http://aip.scitation.org/doi/10.1063/5.0004344)  
1737 [scitation.org/doi/10.1063/5.0004344](http://aip.scitation.org/doi/10.1063/5.0004344) (June 2020).
- 1738 91. Suárez, L. E., Richards, B. A., Lajoie, G. & Misisic, B. Learning function from structure in neuromorphic networks. en. *Nature*  
1739 *Machine Intelligence*. ISSN: 2522-5839. <https://www.nature.com/articles/s42256-021-00376-1> (Aug.  
1740 2021).
- 1741 92. Roberts, J. A. *et al.* Metastable brain waves. en. *Nature Communications* **10**, 1056. ISSN: 2041-1723. [http://www.nature.](http://www.nature.com/articles/s41467-019-08999-0)  
1742 [com/articles/s41467-019-08999-0](http://www.nature.com/articles/s41467-019-08999-0) (Dec. 2019).
- 1743 93. Demirtaş, M. *et al.* Hierarchical Heterogeneity across Human Cortex Shapes Large-Scale Neural Dynamics. en. *Neuron* **101**,  
1744 1181–1194.e13. ISSN: 08966273. <https://linkinghub.elsevier.com/retrieve/pii/S0896627319300443>  
1745 (Mar. 2019).
- 1746 94. Deco, G. *et al.* Resting-State Functional Connectivity Emerges from Structurally and Dynamically Shaped Slow Linear Fluctua-  
1747 tions. en. *Journal of Neuroscience* **33**, 11239–11252. ISSN: 0270-6474, 1529-2401. [http://www.jneurosci.org/cgi/](http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.1091-13.2013)  
1748 [doi/10.1523/JNEUROSCI.1091-13.2013](http://www.jneurosci.org/cgi/doi/10.1523/JNEUROSCI.1091-13.2013) (July 2013).
- 1749 95. Deco, G. *et al.* Dynamical consequences of regional heterogeneity in the brain’s transcriptional landscape. en. *Science Advances*  
1750 **7**, eabf4752. ISSN: 2375-2548. <https://www.science.org/doi/10.1126/sciadv.abf4752> (July 2021).
- 1751 96. Monasson, R. & Rosay, S. Transitions between spatial attractors in place-cell models. *Physical review letters* **115**, 098101 (2015).
- 1752 97. Mante, V., Sussillo, D., Shenoy, K. V. & Newsome, W. T. Context-dependent computation by recurrent dynamics in prefrontal  
1753 cortex. *nature* **503**, 78–84 (2013).
- 1754 98. Fornito, A., Zalesky, A. & Breakspear, M. Graph analysis of the human connectome: Promise, progress, and pitfalls. en. *NeuroIm-*  
1755 *age* **80**, 426–444. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811913004345>  
1756 (Oct. 2013).
- 1757 99. Bertolero, M. A. & Bassett, D. S. On the Nature of Explanations Offered by Network Science: A Perspective From and  
1758 for Practicing Neuroscientists. en. *Topics in Cognitive Science* **12**, 1272–1293. ISSN: 1756-8757, 1756-8765. [https://](https://onlinelibrary.wiley.com/doi/10.1111/tops.12504)  
1759 [onlinelibrary.wiley.com/doi/10.1111/tops.12504](https://onlinelibrary.wiley.com/doi/10.1111/tops.12504) (Oct. 2020).
- 1760 100. Vázquez-Rodríguez, B., Liu, Z.-Q., Hagmann, P. & Misisic, B. Signal propagation via cortical hierarchies. en. *Network Neuro-*  
1761 *science* **4**, 1072–1090. ISSN: 2472-1751. <https://direct.mit.edu/netn/article/4/4/1072-1090/95842>  
1762 (Jan. 2020).
- 1763 101. Bazinet, V., Vos de Wael, R., Hagmann, P., Bernhardt, B. C. & Misisic, B. Multiscale communication in cortico-cortical net-  
1764 works. en. *NeuroImage* **243**, 118546. ISSN: 10538119. [https://linkinghub.elsevier.com/retrieve/pii/](https://linkinghub.elsevier.com/retrieve/pii/S1053811921008193)  
1765 [S1053811921008193](https://linkinghub.elsevier.com/retrieve/pii/S1053811921008193) (Nov. 2021).
- 1766 102. Fornito, A., Zalesky, A., Pantelis, C. & Bullmore, E. T. Schizophrenia, neuroimaging and connectomics. en. *NeuroImage* **62**,  
1767 2296–2314. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811912002133>  
1768 (Oct. 2012).
- 1769 103. Bassett, D. S. *et al.* Hierarchical Organization of Human Cortical Networks in Health and Schizophrenia. en. *Journal of Neu-*  
1770 *roscience* **28**, 9239–9248. ISSN: 0270-6474, 1529-2401. [https://www.jneurosci.org/lookup/doi/10.1523/](https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.1929-08.2008)  
1771 [JNEUROSCI.1929-08.2008](https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.1929-08.2008) (Sept. 2008).
- 1772 104. Bassett, D. S., Nelson, B. G., Mueller, B. A., Camchong, J. & Lim, K. O. Altered resting state complexity in schizophre-  
1773 nia. en. *NeuroImage* **59**, 2196–2207. ISSN: 10538119. [https://linkinghub.elsevier.com/retrieve/pii/](https://linkinghub.elsevier.com/retrieve/pii/S1053811911011633)  
1774 [S1053811911011633](https://linkinghub.elsevier.com/retrieve/pii/S1053811911011633) (Feb. 2012).
- 1775 105. Seguin, C., Razi, A. & Zalesky, A. Inferring neural signalling directionality from undirected structural connectomes. en. *Nature*  
1776 *Communications* **10**, 4289. ISSN: 2041-1723. <http://www.nature.com/articles/s41467-019-12201-w> (Dec.  
1777 2019).
- 1778 106. Seguin, C., Mansour L, S., Sporns, O., Zalesky, A. & Calamante, F. Network communication models narrow the gap between  
1779 the modular organization of structural and functional brain networks. en. *NeuroImage* **257**, 119323. ISSN: 10538119. [https://](https://linkinghub.elsevier.com/retrieve/pii/S1053811922004426)  
1780 [linkinghub.elsevier.com/retrieve/pii/S1053811922004426](https://linkinghub.elsevier.com/retrieve/pii/S1053811922004426) (2023) (Aug. 2022).
- 1781 107. Mišić, B. *et al.* Cooperative and Competitive Spreading Dynamics on the Human Connectome. en. *Neuron* **86**, 1518–1529. ISSN:  
1782 08966273. <https://linkinghub.elsevier.com/retrieve/pii/S0896627315004742> (June 2015).

- 1783 108. Oldham, S. *et al.* The efficacy of different preprocessing steps in reducing motion-related confounds in diffusion MRI connectomics. *NeuroImage* **222**, 117252. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811920307382> (2020).
- 1784
- 1785
- 1786 109. De Reus, M. A. & van den Heuvel, M. P. Estimating false positives and negatives in brain networks. *NeuroImage* **70**, 402–409. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811913000050> (2013).
- 1787
- 1788 110. Yendiki, A., Koldewyn, K., Kakunoori, S., Kanwisher, N. & Fischl, B. Spurious group differences due to head motion in a diffusion MRI study. en. *NeuroImage* **88**, 79–90. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811913011312> (Mar. 2014).
- 1789
- 1790
- 1791 111. Baum, G. L. *et al.* The impact of in-scanner head motion on structural connectivity derived from diffusion MRI. en. *NeuroImage* **173**, 275–286. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811918301381> (June 2018).
- 1792
- 1793
- 1794 112. Cieslak, M. *et al.* QSIPrep: an integrative platform for preprocessing and reconstructing diffusion MRI data. en. *Nature Methods* **18**, 775–778. ISSN: 1548-7091, 1548-7105. <http://www.nature.com/articles/s41592-021-01185-5> (July 2021).
- 1795
- 1796
- 1797 113. Roalf, D. R. *et al.* The impact of quality assurance assessment on diffusion tensor imaging outcomes in a large-scale population-based cohort. en. *NeuroImage* **125**, 903–919. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811915009854> (Jan. 2016).
- 1798
- 1799
- 1800 114. Bargmann, C. I. & Marder, E. From the connectome to brain function. en. *Nature Methods* **10**, 483–490. ISSN: 1548-7091, 1548-7105. <http://www.nature.com/articles/nmeth.2451> (June 2013).
- 1801
- 1802 115. Schaefer, A. *et al.* Local-Global Parcellation of the Human Cerebral Cortex from Intrinsic Functional Connectivity MRI. en. *Cerebral Cortex* **28**, 3095–3114. ISSN: 1047-3211, 1460-2199. <https://academic.oup.com/cercor/article/28/9/3095/3978804> (Sept. 2018).
- 1803
- 1804
- 1805 116. Sarwar, T., Ramamohanarao, K. & Zalesky, A. Mapping connectomes with diffusion MRI: deterministic or probabilistic tractography? en. *Magnetic Resonance in Medicine* **81**, 1368–1384. ISSN: 0740-3194, 1522-2594. <https://onlinelibrary.wiley.com/doi/10.1002/mrm.27471> (Feb. 2019).
- 1806
- 1807
- 1808 117. Robinson, P. *et al.* Eigenmodes of brain activity: Neural field theory predictions and comparison with experiment. en. *NeuroImage* **142**, 79–98. ISSN: 10538119. <https://linkinghub.elsevier.com/retrieve/pii/S1053811916300908> (Nov. 2016).
- 1809
- 1810
- 1811 118. Deco, G., Jirsa, V. K., Robinson, P. A., Breakspear, M. & Friston, K. The Dynamic Brain: From Spiking Neurons to Neural Masses and Cortical Fields. en. *PLoS Computational Biology* **4** (ed Sporns, O.) e1000092. ISSN: 1553-7358. <https://dx.plos.org/10.1371/journal.pcbi.1000092> (Aug. 2008).
- 1812
- 1813
- 1814 119. Shenoy, K. V. & Kao, J. C. Measurement, manipulation and modeling of brain-wide neural population dynamics. en. *Nature Communications* **12**, 633. ISSN: 2041-1723. <http://www.nature.com/articles/s41467-020-20371-1> (Dec. 2021).
- 1815
- 1816
- 1817 120. Nozari, E. *et al.* *Is the brain macroscopically linear? A system identification of resting state dynamics* 2020.
- 1818 121. He, X. *et al.* Uncovering the biological basis of control energy: Structural and metabolic correlates of energy inefficiency in temporal lobe epilepsy. en. *Science Advances* **8**, eabn2293. ISSN: 2375-2548. <https://www.science.org/doi/10.1126/sciadv.abn2293> (2023) (Nov. 2022).
- 1819
- 1820
- 1821 122. McCormick, D. A., Shu, Y. & Yu, Y. Hodgkin and Huxley model—still standing? *Nature* **445**, E1–E2 (2007).
- 1822 123. Durstewitz, D., Seamans, J. K. & Sejnowski, T. J. Neurocomputational models of working memory. *Nature neuroscience* **3**, 1184–1191 (2000).
- 1823
- 1824 124. Hu, S. *et al.* Associative memory realized by a reconfigurable memristive Hopfield neural network. *Nature communications* **6**, 1–8 (2015).
- 1825
- 1826 125. Vaidyanathan, S., Volos, C., *et al.* *Advances and applications in nonlinear control systems* (Springer, 2016).
- 1827 126. Kumar, E. V. & Jerome, J. Robust LQR controller design for stabilizing and trajectory tracking of inverted pendulum. *Procedia Engineering* **64**, 169–178 (2013).
- 1828
- 1829 127. Bastos, A. M. & Schoffelen, J.-M. A tutorial review of functional connectivity analysis methods and their interpretational pitfalls. *Frontiers in systems neuroscience* **9**, 175 (2016).
- 1830
- 1831 128. Schwemmer, M. A. & Lewis, T. J. in *Phase response curves in neuroscience* 3–31 (Springer, 2012).
- 1832 129. Park, Y. & Ermentrout, B. Weakly coupled oscillators in a slowly varying world. *Journal of computational neuroscience* **40**, 269–281 (2016).
- 1833
- 1834 130. Brunton, S. L., Brunton, B. W., Proctor, J. L. & Kutz, J. N. Koopman invariant subspaces and finite linear representations of nonlinear dynamical systems for control. *PloS one* **11**, e0150171 (2016).
- 1835
- 1836 131. Proctor, J. L., Brunton, S. L. & Kutz, J. N. Dynamic mode decomposition with control. *SIAM Journal on Applied Dynamical Systems* **15**, 142–161 (2016).
- 1837

- 1838 132. Zañudo, J. G. T., Yang, G. & Albert, R. Structure-based control of complex networks with nonlinear dynamics. *Proceedings of*  
1839 *the National Academy of Sciences* **114**, 7234–7239 (2017).
- 1840 133. Haynes, G. & Hermes, H. Nonlinear controllability via Lie theory. *SIAM Journal on Control* **8**, 450–460 (1970).
- 1841 134. Towlson, E. K. *et al.* *Caenorhabditis elegans* and the network control framework—FAQs. en. *Philosophical Transactions of the*  
1842 *Royal Society B: Biological Sciences* **373**, 20170372. ISSN: 0962-8436, 1471-2970. <https://royalsocietypublishing.org/doi/10.1098/rstb.2017.0372> (Oct. 2018).
- 1843
- 1844 135. Felleman, D. J. & Van Essen, D. C. Distributed Hierarchical Processing in the Primate Cerebral Cortex. *Cerebral Cortex* **1**,  
1845 1–47. ISSN: 1047-3211, 1460-2199. [https://academic.oup.com/cercor/article-lookup/doi/10.1093/](https://academic.oup.com/cercor/article-lookup/doi/10.1093/cercor/1.1.1)  
1846 [cercor/1.1.1](https://academic.oup.com/cercor/article-lookup/doi/10.1093/cercor/1.1.1) (1991).
- 1847 136. Stiso, J. *et al.* Learning in brain-computer interface control evidenced by joint decomposition of brain and behavior. *Journal of*  
1848 *Neural Engineering* **17**, 046018. ISSN: 1741-2552. [https://iopscience.iop.org/article/10.1088/1741-](https://iopscience.iop.org/article/10.1088/1741-2552/ab9064)  
1849 [2552/ab9064](https://iopscience.iop.org/article/10.1088/1741-2552/ab9064) (July 2020).
- 1850 137. Szymula, K. P., Pasqualetti, F., Graybiel, A. M., Desrochers, T. M. & Bassett, D. S. Habit learning supported by efficiently  
1851 controlled network dynamics in naive macaque monkeys. *arXiv*. <https://arxiv.org/abs/2006.14565> (2020).
- 1852 138. Rosen, A. F. *et al.* Quantitative assessment of structural image quality. en. *NeuroImage* **169**, 407–418. ISSN: 10538119. [https://](https://linkinghub.elsevier.com/retrieve/pii/S1053811917310832)  
1853 [linkinghub.elsevier.com/retrieve/pii/S1053811917310832](https://linkinghub.elsevier.com/retrieve/pii/S1053811917310832) (Apr. 2018).
- 1854 139. Ciric, R. *et al.* Mitigating head motion artifact in functional connectivity MRI. en. *Nature Protocols* **13**, 2801–2826. ISSN: 1754-  
1855 2189, 1750-2799. <http://www.nature.com/articles/s41596-018-0065-y> (Dec. 2018).
- 1856 140. Gorgolewski, K. *et al.* Nipype: A Flexible, Lightweight and Extensible Neuroimaging Data Processing Framework in Python.  
1857 *Frontiers in Neuroinformatics* **5**. ISSN: 1662-5196. [http://journal.frontiersin.org/article/10.3389/](http://journal.frontiersin.org/article/10.3389/fninf.2011.00013/abstract)  
1858 [fninf.2011.00013/abstract](http://journal.frontiersin.org/article/10.3389/fninf.2011.00013/abstract) (2011).
- 1859 141. Satterthwaite, T. D. *et al.* An improved framework for confound regression and filtering for control of motion artifact in  
1860 the preprocessing of resting-state functional connectivity data. en. *NeuroImage* **64**, 240–256. ISSN: 10538119. [https://](https://linkinghub.elsevier.com/retrieve/pii/S1053811912008609)  
1861 [linkinghub.elsevier.com/retrieve/pii/S1053811912008609](https://linkinghub.elsevier.com/retrieve/pii/S1053811912008609) (Jan. 2013).
- 1862 142. Esteban, O. *et al.* fMRIPrep: a robust preprocessing pipeline for functional MRI. en. *Nature Methods* **16**, 111–116. ISSN: 1548-  
1863 7091, 1548-7105. <http://www.nature.com/articles/s41592-018-0235-4> (Jan. 2019).
- 1864 143. Harris, J. A. *et al.* Hierarchical organization of cortical and thalamic connectivity. *Nature* **575**, 195–202 (2019).
- 1865 144. Knox, J. E. *et al.* High-resolution data-driven model of the mouse connectome. *Network Neuroscience* **3**, 217–236 (2018).
- 1866 145. Mahadevan, A. S. *et al.* Alprazolam modulates persistence energy during emotion processing in first-degree relatives of individ-  
1867 uals with schizophrenia: a network control study en. preprint (bioRxiv, Apr. 2021). [http://biorxiv.org/lookup/doi/](http://biorxiv.org/lookup/doi/10.1101/2021.04.22.440935)  
1868 [10.1101/2021.04.22.440935](http://biorxiv.org/lookup/doi/10.1101/2021.04.22.440935).
- 1869 146. Markello, R. D. *et al.* neuromaps: structural and functional interpretation of brain maps. en. *Nature Methods*. ISSN: 1548-7091,  
1870 1548-7105. <https://www.nature.com/articles/s41592-022-01625-w> (Oct. 2022).
- 1871 147. Hansen, J. Y. *et al.* Mapping neurotransmitter systems to the structural and functional organization of the human neocortex. en.  
1872 *Nature Neuroscience*. ISSN: 1097-6256, 1546-1726. <https://www.nature.com/articles/s41593-022-01186-3>  
1873 (Oct. 2022).
- 1874 148. Regional, Layer, and Cell-Type-Specific Connectivity of the Mouse Default Mode Network. en. *Neuron* **109**, 545–559.e8. ISSN:  
1875 08966273. <https://linkinghub.elsevier.com/retrieve/pii/S0896627320308898> (Feb. 2021).
- 1876 149. Váša, F. & Mišić, B. Null models in network neuroscience. en. *Nature Reviews Neuroscience*. ISSN: 1471-003X, 1471-0048.  
1877 <https://www.nature.com/articles/s41583-022-00601-9> (May 2022).
- 1878 150. Roberts, J. A. *et al.* The contribution of geometry to the human connectome. *NeuroImage* **124**, 379–393. ISSN: 10538119.  
1879 <https://linkinghub.elsevier.com/retrieve/pii/S105381191500806X> (2016).

XII. EXTENDED DATA

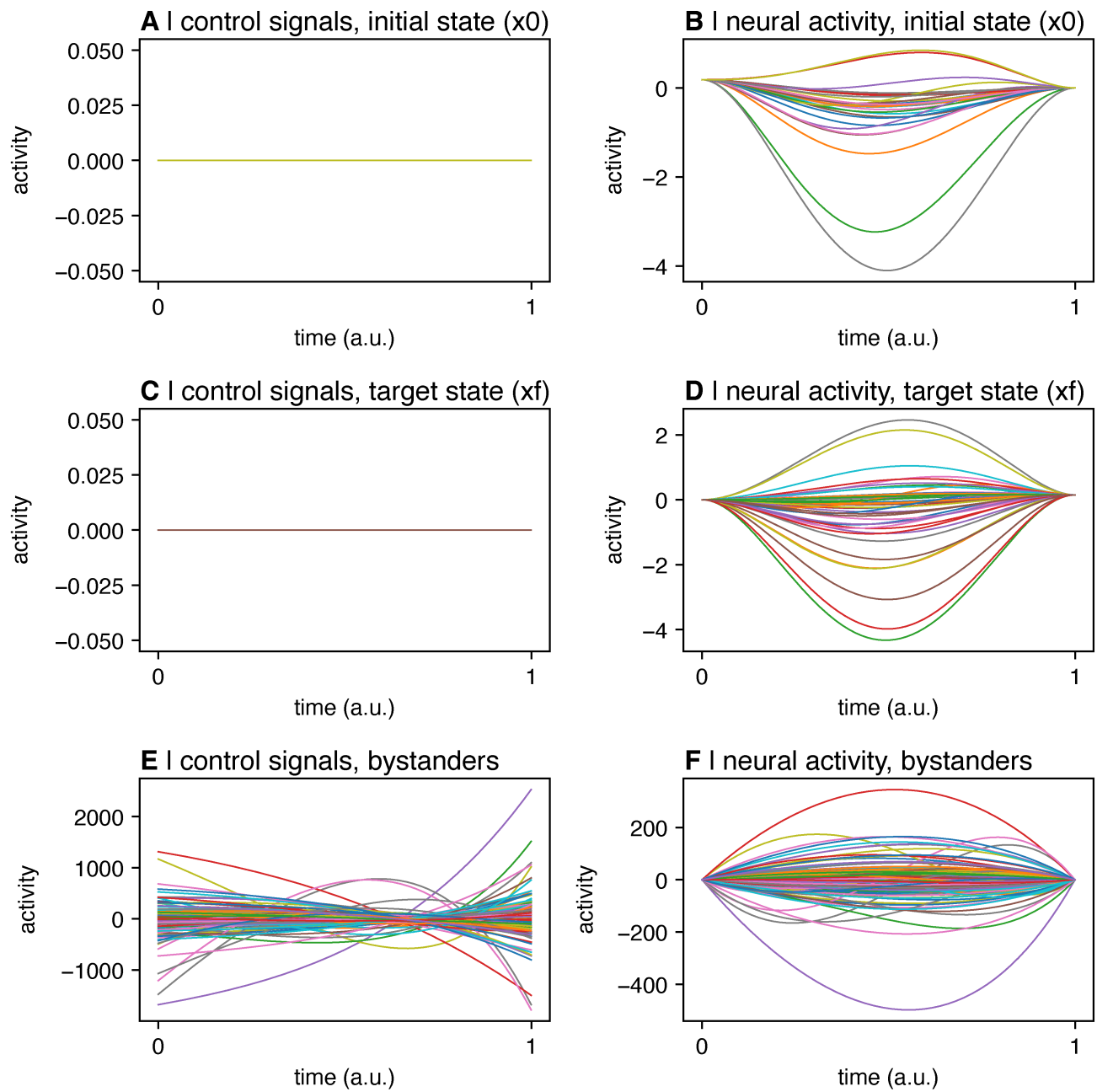
Control energy as a function of connectome edge density



**FIG. S1**

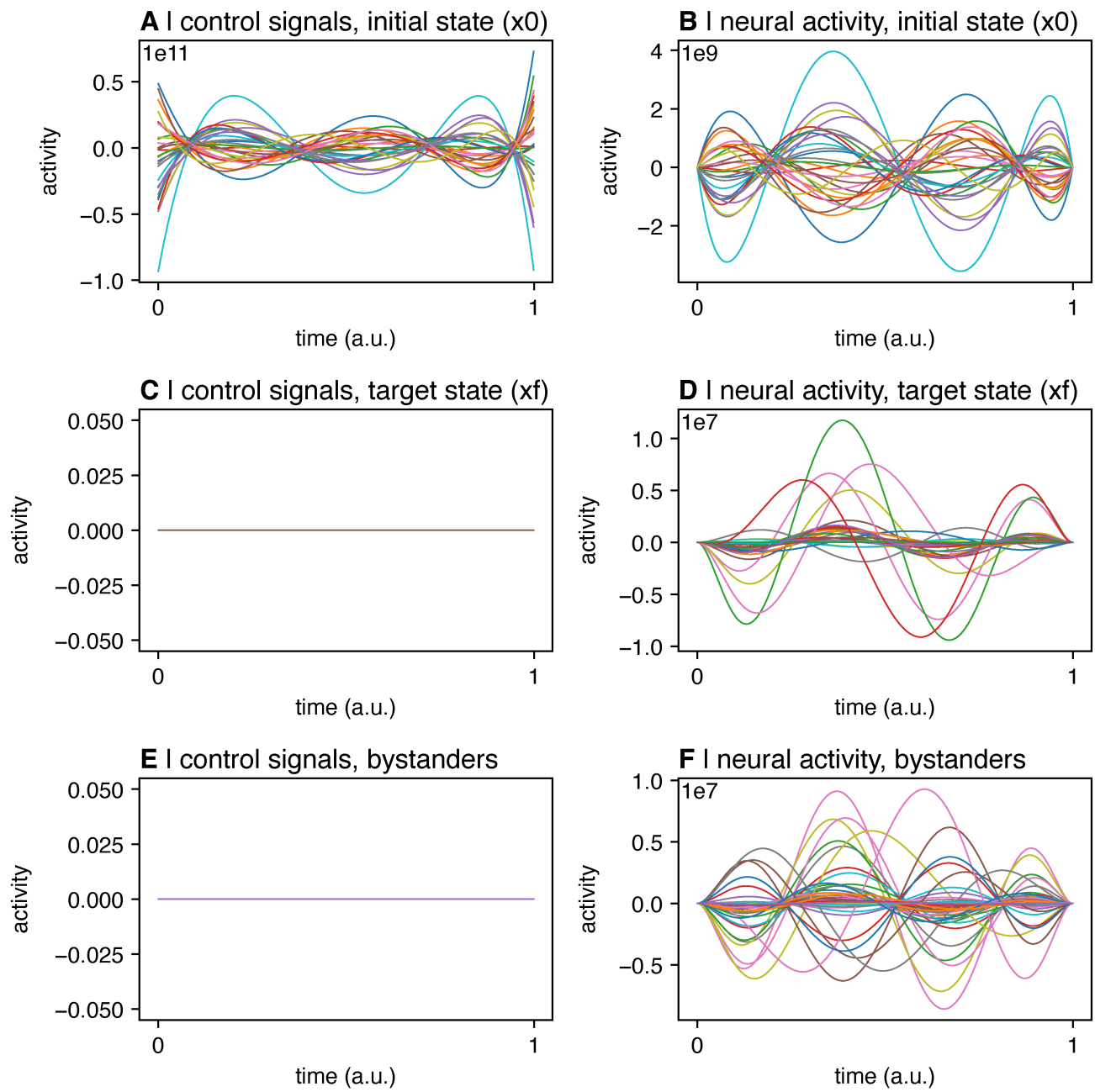
**Control energy as a function of connectome edge density.** In the undirected human connectome, we iteratively set 500 of the weakest edges (A) or 500 random edges (B) to 0, stopping once edge density fell below 10%. *Control energy* was recomputed at each iteration and is shown here. This plot illustrates the fact that energy varies as a function of edge density and is primarily driven by the strongest edges in the network.





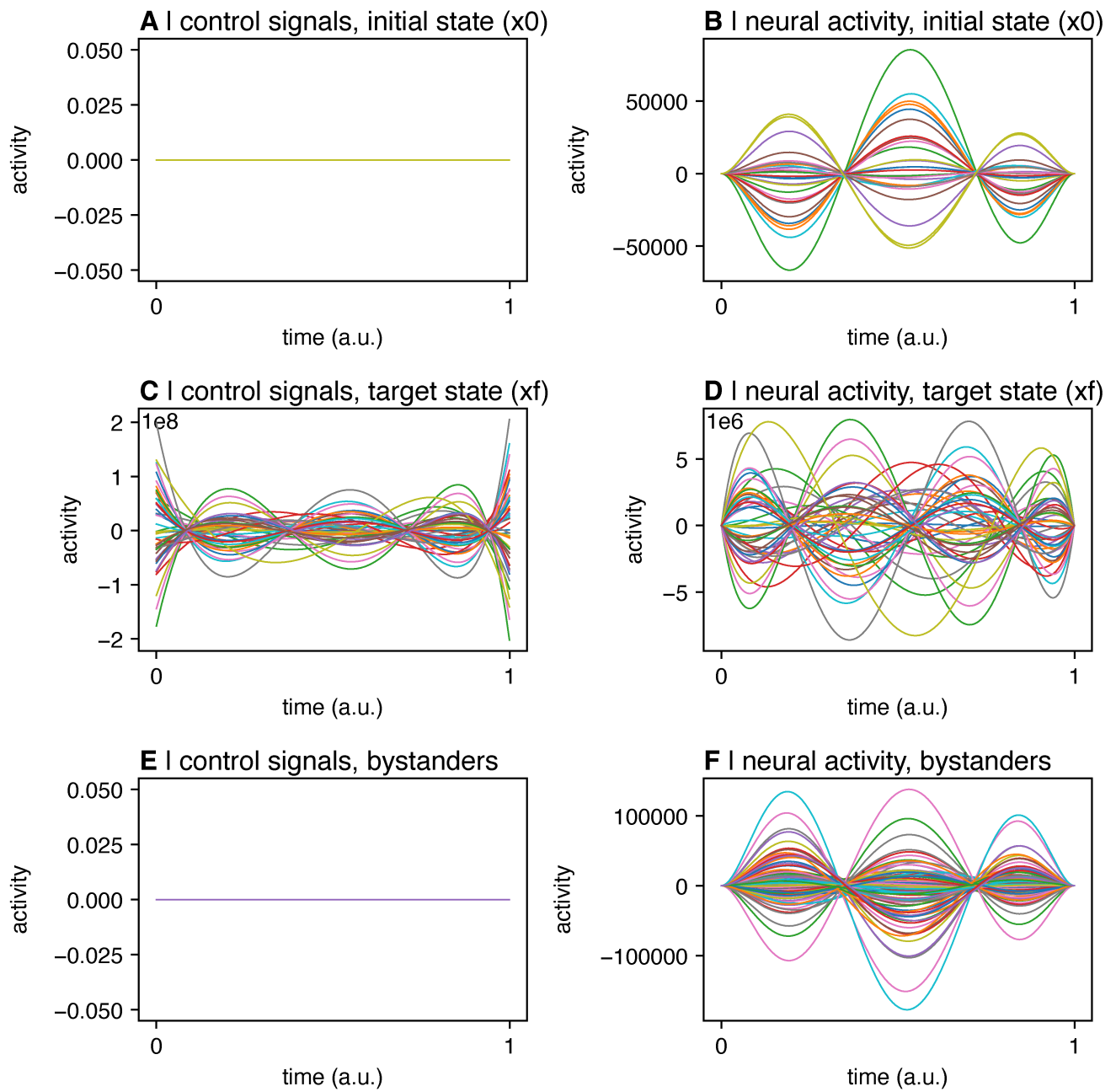
**FIG. S2**

**Control signals and state trajectory.** Only bystanders are set as control nodes.  $T=1$ . Initial state = visual system. Target state = default mode system. Inversion error =  $2.09 \times 10^{-10}$ . Reconstruction error =  $1.75 \times 10^{-8}$ . Energy =  $6.64 \times 10^9$ .



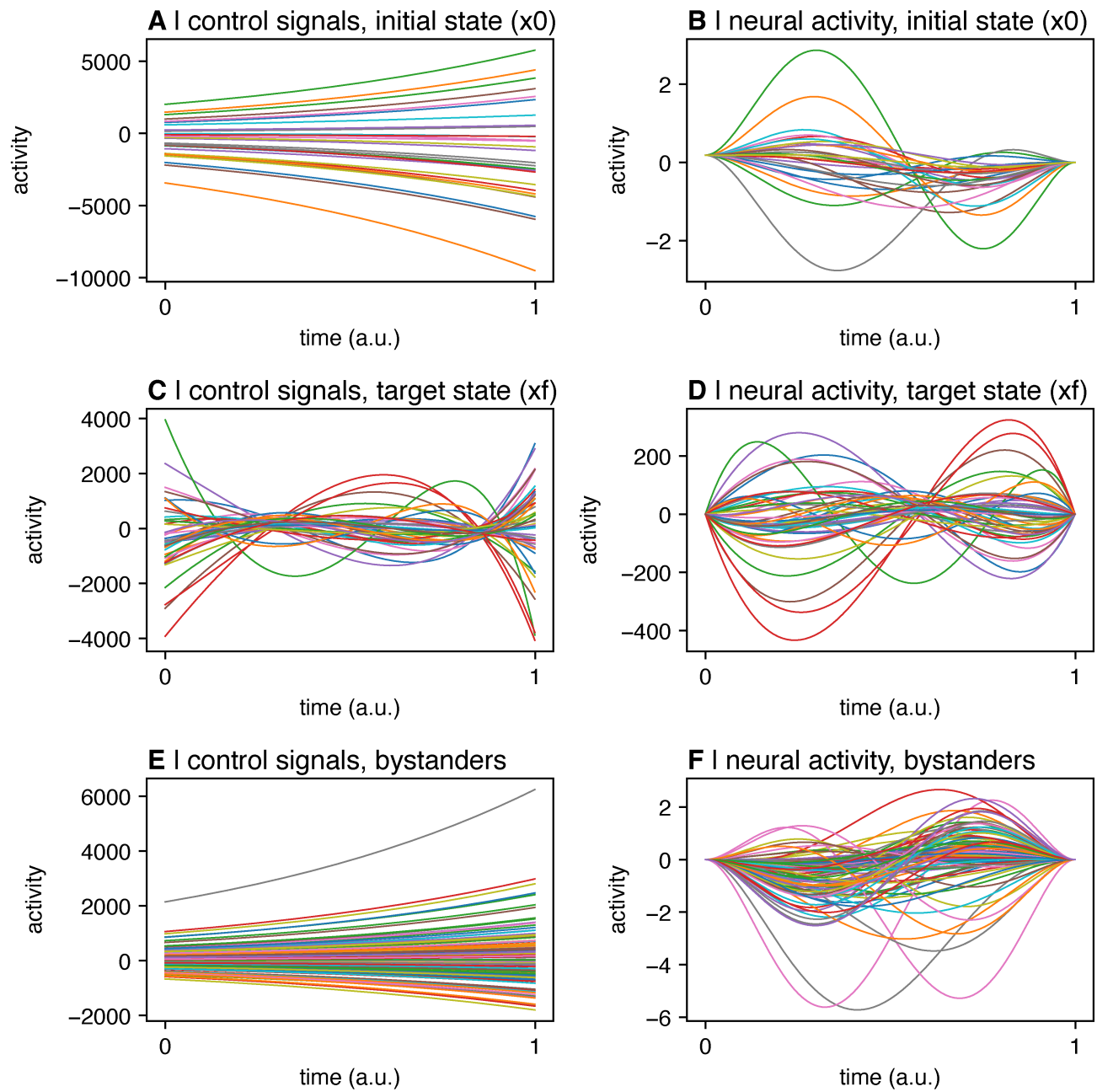
**FIG. S3**

**Control signals and state trajectory.** Only nodes in the initial state are set as control nodes.  $T=1$ . Initial state = visual system. Target state = default mode system. Inversion error =  $1.37 \times 10^3$ . Reconstruction error =  $2.04 \times 10^5$ . Energy =  $3.68 \times 10^{24}$ . Note that this state transition does not complete successfully.



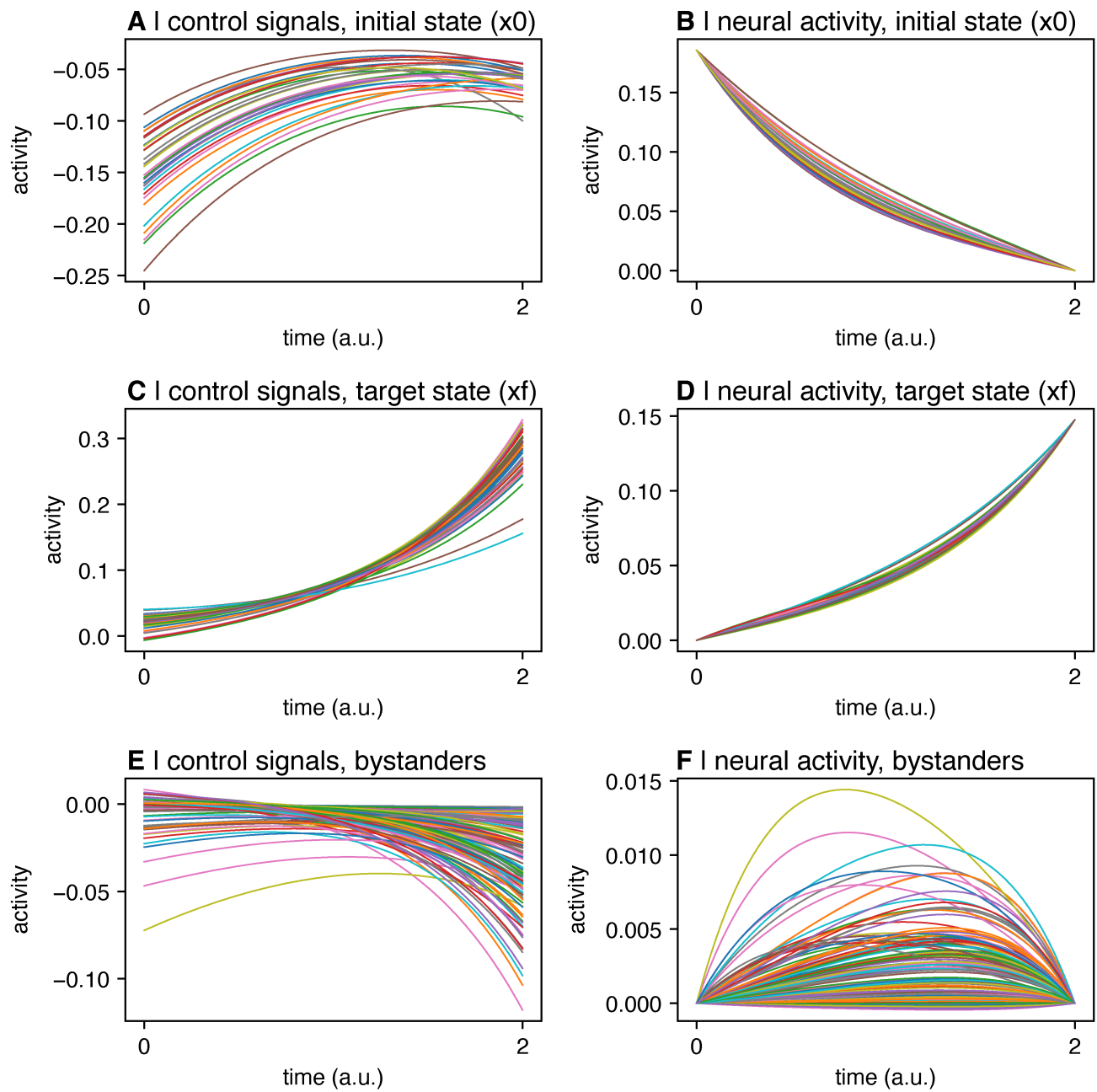
**FIG. S4**

**Control signals and state trajectory.** Only nodes in the target state are set as control nodes.  $T=1$ . Initial state = visual system. Target state = default mode system. Inversion error = 1.50. Reconstruction error =  $1.12 \times 10^2$ . Energy =  $3.38 \times 10^{19}$ . Note that this state transition does not complete successfully.



**FIG. S5**

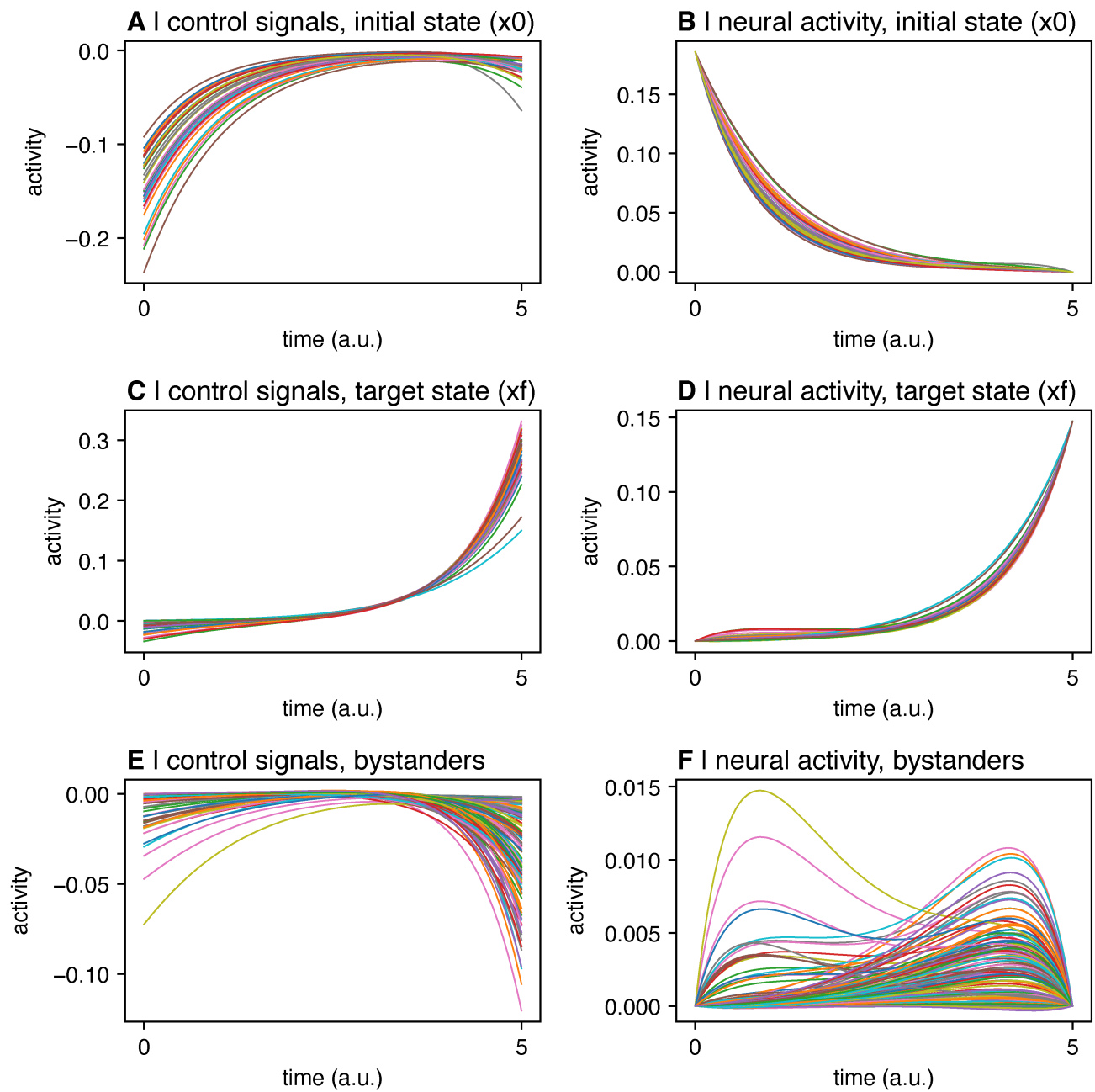
**Control signals and state trajectory.** Nodes in the target state are set as control nodes with control weights of 1, whereas the remaining nodes are given a small amount of control ( $1 \times 10^{-5}$ ).  $T=1$ . Initial state = visual system. Target state = default mode system. Inversion error =  $2.00 \times 10^{-8}$ . Reconstruction error =  $1.16 \times 10^{-6}$ . Energy =  $2.31 \times 10^{11}$ . Unlike the scenario depicted in Figure S4, this state transition completes successfully.



**FIG. S6**

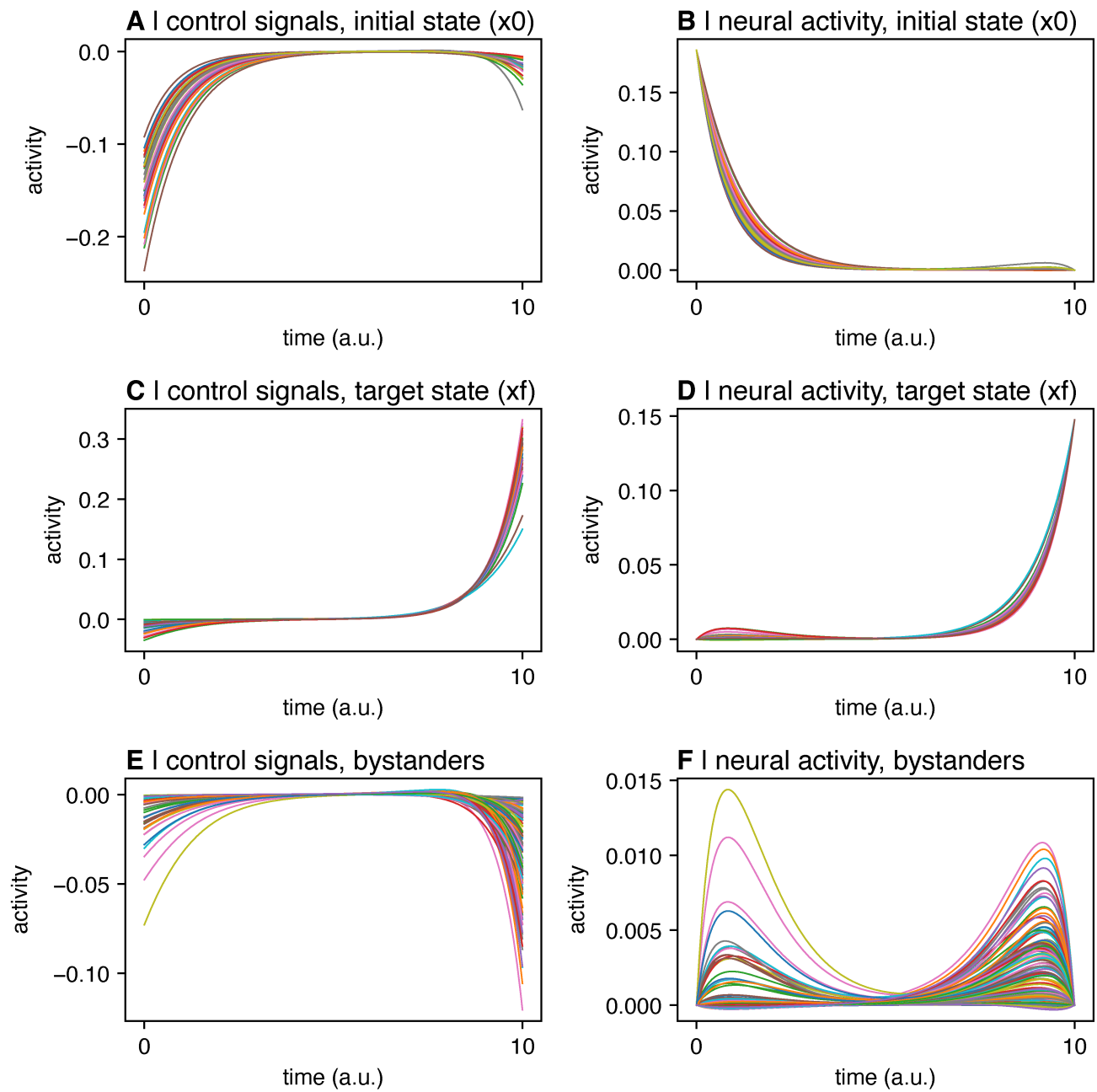
**Control signals and state trajectory.** Uniform full control set.  $T=2$ . Initial state = visual system. Target state = default mode system. Inversion error =  $3.24 \times 10^{-15}$ . Reconstruction error =  $1.55 \times 10^{-13}$ . Energy = 1904.





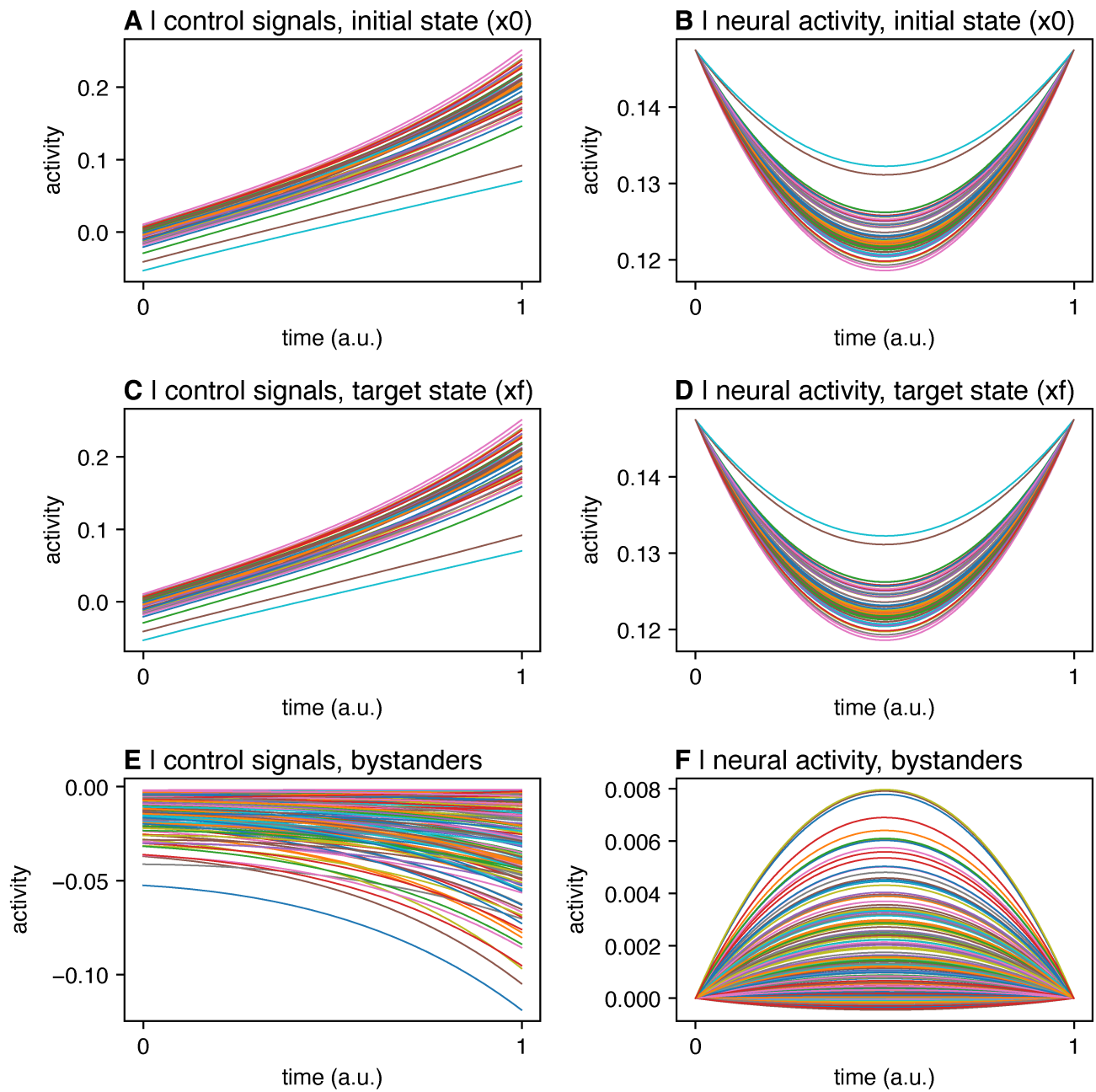
**FIG. S7**

**Control signals and state trajectory.** Uniform full control set.  $T=5$ . Initial state = visual system. Target state = default mode system. Inversion error =  $1.33 \times 10^{-13}$ . Reconstruction error =  $1.03 \times 10^{-11}$ . Energy = 1797.



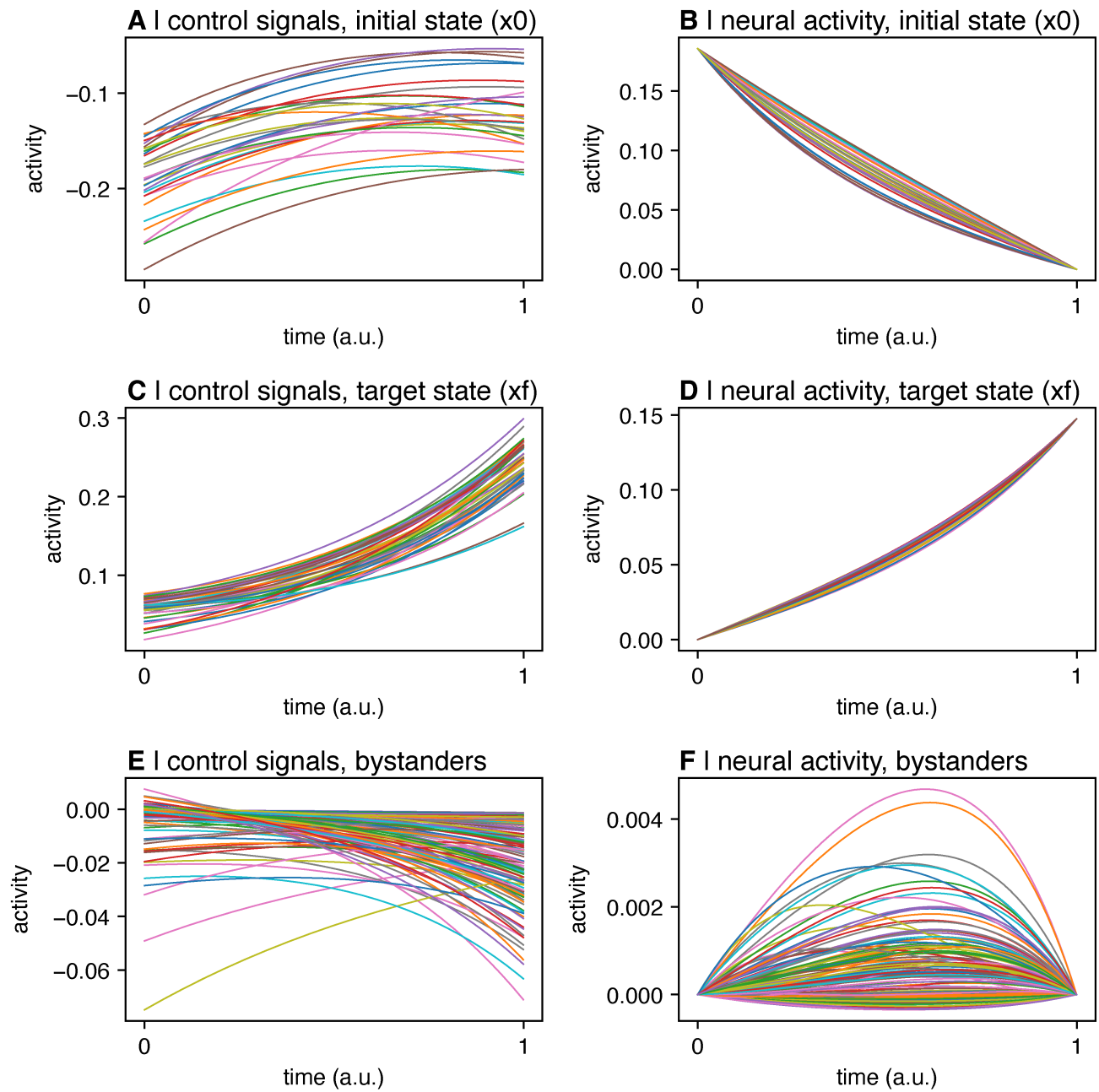
**FIG. S8**

**Control signals and state trajectory.** Uniform full control set.  $T=10$ . Initial state = visual system. Target state = default mode system. Inversion error =  $1.48 \times 10^{-10}$ . Reconstruction error =  $1.71 \times 10^{-8}$ . Energy = 1801.



**FIG. S9**

**Control signals and state trajectory.** Uniform full control set.  $T=1$ . *Initial state = default mode system. Target state = default mode system.* Inversion error =  $2.20 \times 10^{-16}$ . Reconstruction error =  $3.95 \times 10^{-14}$ . Energy = 571.



**FIG. S10**

**Control signals and state trajectory.** *Annotation map control set.*  $T=1$ . Initial state = visual system. Target state = default mode system. Inversion error =  $1.87 \times 10^{-15}$ . Reconstruction error =  $6.07 \times 10^{-14}$ . Energy = 1455.