

Silver nanoparticle technology in orthopaedic infections

Madhan Jeyaraman, Naveen Jeyaraman, Arulkumar Nallakumarasamy, Karthikeyan P Iyengar, Vijay Kumar Jain, Anish G Potty, Ashim Gupta

Specialty type: Nanoscience and nanotechnology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0

Grade B (Very good): 0

Grade C (Good): C, C, C, C

Grade D (Fair): 0

Grade E (Poor): 0

P-Reviewer: Hadianamrei R, United Kingdom; Koubaa M, Tunisia; Soriano-Ursúa MA, Mexico

Received: April 18, 2023

Peer-review started: April 18, 2023

First decision: July 18, 2023

Revised: July 24, 2023

Accepted: September 4, 2023

Article in press: September 4, 2023

Published online: September 18, 2023



Madhan Jeyaraman, Naveen Jeyaraman, Arulkumar Nallakumarasamy, Department of Orthopaedics, ACS Medical College and Hospital, Dr MGR Educational and Research Institute, Chennai 600077, Tamil Nadu, India

Madhan Jeyaraman, Anish G Potty, Ashim Gupta, Department of Orthopaedics, South Texas Orthopaedic Research Institute, Laredo, TX 78045, United States

Karthikeyan P Iyengar, Department of Trauma and Orthopaedics, Southport & Ormskirk Hosp NHS Trust Southport, Richmond PR8 6PN, Southport, United Kingdom

Vijay Kumar Jain, Department of Orthopaedics, Atal Bihari Vajpayee Institute of Medical Sciences, Dr Ram Manohar Lohia Hospital, Delhi 110001, New Delhi, India

Ashim Gupta, Department of Regenerative Medicine, Regenerative Orthopaedics, Noida 201301, Uttar Pradesh, India

Ashim Gupta, Department of Regenerative Medicine, Future Biologics, Lawrenceville, GA 30043, United States

Ashim Gupta, Department of Regenerative Medicine, BioIntegarte, Lawrenceville, GA 30043, United States

Corresponding author: Madhan Jeyaraman, MS, PhD, Assistant Professor, Research Associate, Department of Orthopaedics, ACS Medical College and Hospital, Dr MGR Educational and Research Institute, Chennai 600077, Tamil Nadu, India. madhanjeyaraman@gmail.com

Abstract

The irrational and prolonged use of antibiotics in orthopaedic infections poses a major threat to the development of antimicrobial resistance. To combat antimicrobial resistance, researchers have implemented various novel and innovative modalities to curb infections. Nanotechnology involves doping ions/metals onto the scaffolds to reach the target site to eradicate the infective foci. In this connotation, we reviewed silver nanoparticle technology in terms of mechanism of action, clinical applications, toxicity, and regulatory guidelines to treat orthopaedic infections.

Key Words: Antimicrobial resistance; Biofilms; Silver; Nanoparticles; Orthopaedics

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Core Tip: To overcome antimicrobial resistance, researchers explored the alternate technology to curb infections in musculo-skeletal disorders. Nanotechnology forms an eye opener in the usage of silver nanoparticles to eradicate infections in osteoarticular system.

Citation: Jeyaraman M, Jeyaraman N, Nallakumarasamy A, Iyengar KP, Jain VK, Potty AG, Gupta A. Silver nanoparticle technology in orthopaedic infections. *World J Orthop* 2023; 14(9): 662-668

URL: <https://www.wjgnet.com/2218-5836/full/v14/i9/662.htm>

DOI: <https://dx.doi.org/10.5312/wjo.v14.i9.662>

INTRODUCTION

Globally, Orthopaedic infections pose a great challenge among orthopaedic surgeons and orthopaedic researchers. In the literature, the incidence of infection after orthopaedic surgeries was 1% to 2% in closed fractures and approximately 30% in open fractures[1]. Universally, there is no accepted classification for infection after orthopaedic surgeries. To overcome these infections, antibiotics were used irrationally which led to antimicrobial resistance (AMR) due to the development of biofilm by the micro-organisms[2,3]. Jefferson reported that biofilm formation is due to (1) protection from host defense; (2) colonization; (3) local environment benefits; and (4) planktonic cultures as *in vitro* artifacts[4]. Aparna *et al*[5] gave 5 stages of the growth cycle of a biofilm namely stage 1 – attachment phase, stage 2 – irreversible binding phase, stage 3 – maturation 1 phase, stage 4 – maturation 2 phase, and stage 5 – cellular dispersion phase.

In humans, 80% of microbial infections are due to non-healing chronic wounds, osteomyelitis, prosthesis- and implant-related infections, endocarditis, rhinosinusitis, and cystic fibrosis[6-9]. Literature states that both gram-positive and gram-negative bacteria form biofilms on the surface of medical devices which are *E. faecalis*, *S. aureus*, *S. epidermidis*, *S. viridans*, *E. coli*, *K. pneumoniae*, *P. mirabilis*, and *P. aeruginosa*[10]. Out of all these organisms, *S. aureus* and coagulase-negative staphylococci pose a greater risk of forming biofilms in orthopaedic implants[11,12], whereas *P. aeruginosa* sustain and survive in harsh environments and forms a resistant biofilm[13].

With the rise of AMR, antibiotic prophylaxis has become ineffective in curbing infections. Novel techniques such as nanomedicine, bacteriophage therapy, antimicrobial peptides, sonic therapies, scaffold-loaded nanoparticles, antimicrobial adjuvants in the form of silver, electrical and electromagnetic methods, bioacoustic effects, surface modification of biomaterials, antimicrobial photodynamic therapies, biosensors, hyperbaric oxygen, and fecal microbiota transplantation have been introduced to combat AMR[10,14,15]. With the profound literature evidence of the antimicrobial property of silver (Ag), nanotechnology experts doped nanoparticles (NP) with Ag for targeted drug delivery and enhanced interaction with the surrounding environment to curb biofilm-producing organisms[16]. In this article, the usage of AgNP in curbing orthopaedic infections is narrated.

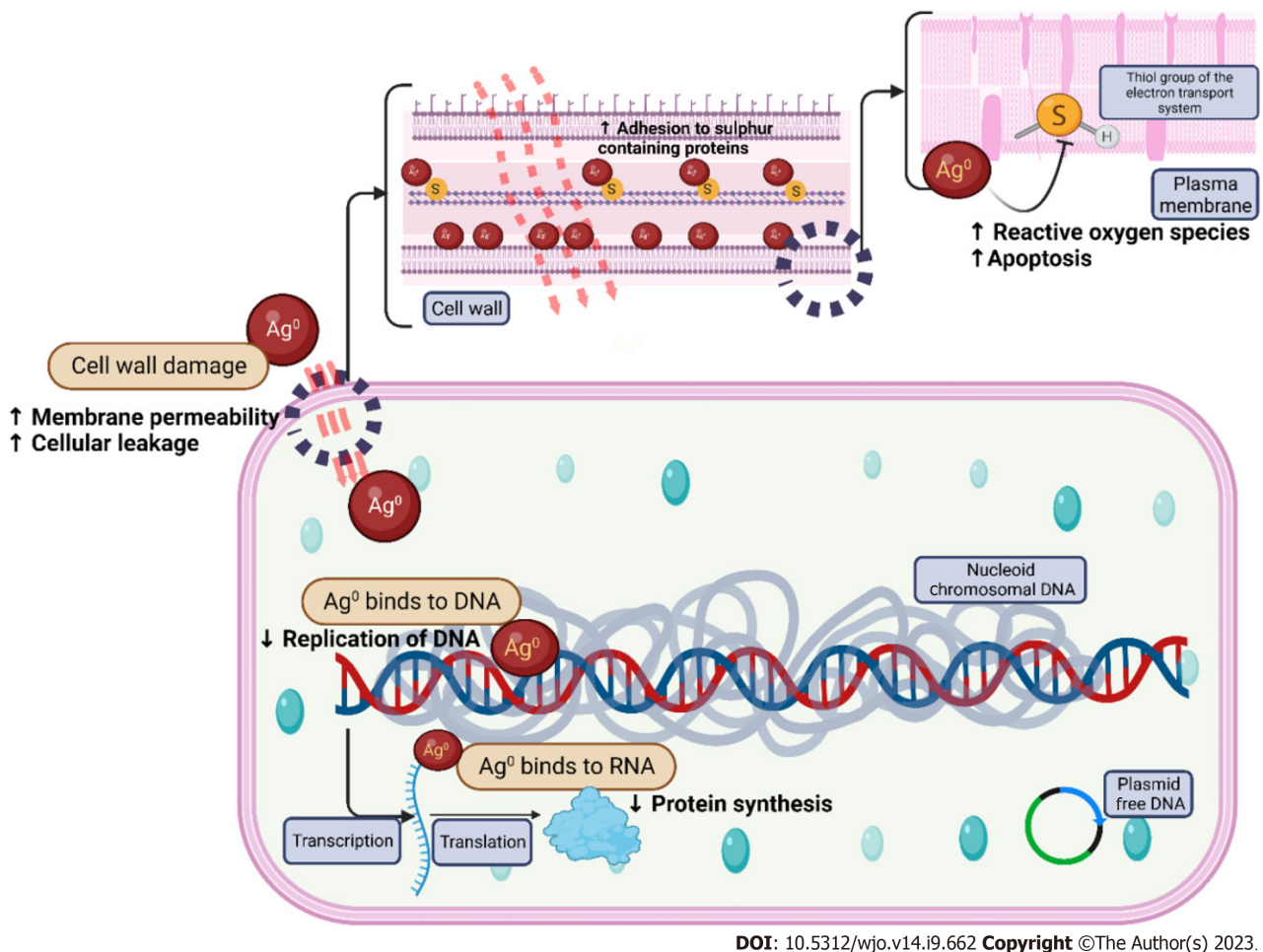
AGNP TECHNOLOGY

In literature, Ag has been identified to have antimicrobial properties[17-19], but in recent years clinicians have demonstrated this property in various clinical conditions. The oligodynamic action of Ag refers to the toxic nature of metal ions on microbes by integrating with microbial deoxyribonucleic acid (DNA)[20-22]. Few studies have emphasized the precipitation of DNA within the microbial cell[23]. Ag exerts antibacterial action by inhibiting cell wall synthesis[24,25].

Recent technologies use NP to load Ag ions which may be used as an antimicrobial agent in a target-specific manner [23]. AgNP permeates into cells and interferes with the enzymes of bacterial respiratory chains to inhibit ATP production and growth of the bacteria[16,26]. The particle size of AgNP determines the bactericidal activity. 10-nm Ag demonstrates the complete bacterial interaction, henceforth AgNP exerts a higher bactericidal effect[27]. The mechanism of action of AgNP in infections is as follows (as depicted in Figure 1).

Direct contact with microbes through leakage of cellular contents and bacterial death due to the damage of cell membrane and higher production of reactive oxygen species and free radical species, release of Ag⁺ ions through interaction with sulfhydryl groups (cysteine) in cell wall proteins and enzymes, induction of bacterial death when Ag⁺ ions in AgNPs by bombarding the electron transport chain in bacterial mitochondria. Entering of Ag⁺ ions into periplasmic space which leads to the separation of the cytosol from the cell membrane, occurrence of cellular pits after the exposure of Ag⁺ ions, inhibition of ribosomal functions leading to enhanced ROS production, malformation of proteins resulting in improper DNA function, antibiofilm activity of AgNPs, and dose-dependent cytotoxic and genotoxic effects of AgNP on osteoblasts and impaired cellular viability of AgNPs at 10 µg/g.

To impart biocompatibility of AgNP in musculoskeletal tissues: (1) Biosynthesis process from bacteria, yeast, and fungi; (2) physical property adjustment; and (3) biomolecule combinations can be tried.



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Figure 1 Mechanism of action of silver nanoparticles. DNA: Deoxyribonucleic acid.

AGNP IN ORTHOPAEDIC INFECTIONS

Recently, the modification of orthopaedic implants with the application of AgNPs on the surface leads to the prevention of implant-associated infections. With the availability of AgNP-coated external fixators, mega prostheses, and AgNP-coated bone cement, orthopaedic infections are showing a downtrend. Since AgNP-coated orthopaedic implants demonstrate antimicrobial activity, the specific molecular mechanism of osteogenic-related cells warrants an understanding. Due to dose-dependent cytotoxicity, the effect of AgNP on osteoblast and osteoclast is controversial. Aureore *et al*[28] demonstrated the bactericidal effect of AgNP against non-virulent *E.coli* and virulent MRSA at non-toxic concentrations. Macrophage polarization towards the M1 phenotype enables the cells to kill the engulfed microorganisms. Elevated reactive oxygen species (ROS) responses were found in AgNP-treated osteoclasts. The modification of orthopaedic implants by using AgNPs enhances antimicrobial effects through plasma immersion ion implantation (PIII), magnetron sputtering, plasma electrolytic oxidation, and 3DP-Ag-containing scaffolds[16].

Tumor prosthesis: In orthopaedic oncology, peri-prosthetic infection rates from 9% to 29%. Due to the immunosuppressive environment, this group of patients is more prone to infection than arthroplasty patients. Gosheger *et al*[29] demonstrated a superior antimicrobial effect with a silver-coated mega prosthesis (7% infection rate) than titanium prosthesis (47% infection rate) in a rabbit model. Ag coated group showed fewer signs of inflammation as measured by ESR, CRP, and neutrophil count. In sarcoma patients, Hardes *et al*[30] observed 17.6% of infection in the non-silver coated mega prosthesis group than 5.6% of infection in silver coated mega prosthesis group. About 38.5% of cases of amputation in the non-silver-coated mega prosthesis group were due to deep infections whereas no case of amputation was reported in the silver-coated mega prosthesis group. Hence, silver-coated mega prosthesis reduces the risk of further infection in oncology cases as they are already in an immunocompromised state. However, large-scale blinded controlled trials have to prove the safety and efficacy of infection prevention with the silver-coated prosthesis in orthopaedic oncological cases.

External fixator pins: Pin tract infections mount for 42% of orthopaedic infections which results in loosening of the implant, fracture non-union, and osteomyelitis. An in-vitro study demonstrated a 3-log step reduction of *S. epidermidis* biofilm producers when incubating the stainless steel pins coated with AgNP compared with titanium and copper pins for 20 h (9). Wassall *et al*[31] revealed the antimicrobial effects of Ag in Ag-coated pins with the significant reduction of adhesion of *E. coli*, *P. aeruginosa*, and *S. aureus* when compared with normal stainless steel pins. Loosening of external fixator pins was less frequently found with Ag-coated pins. Hence silver inhibits microbial adhesion by inhibiting the formation of the glycocalyx on the surface of the pins.

Osteomyelitis and infected non-union: Ag ionophoresis act as an adjunct treatment option for osteomyelitis and infection in non-union of fractures. The continuous inflow of electrically driven Ag into the infective foci curbs the infection and promotes the environment for tissue regeneration. At the end of 3 mo follow-up, radiographic and histological analysis revealed neo-osteogenesis in a 6 mm critical bone defect in the femoral diaphysis of rats managed with bone graft with BMP-2 coupled with AgNP and poly lactic-co-glycolic acid (PLGA) scaffold injected with 10 CFUs of vancomycin-resistant MRSA[32]. Microbial elimination with 2% Ag NP coupled with composite bioscaffold resulted in fracture union.

Bone cement: Alt *et al*[33] proved that nanosilver-coated cement produced a high index of antimicrobial activity against *S. epidermidis*, MRSE, and MRSA. Tiopronin, a stabilizing agent, coupled with AgNP in bone cement expressed a good antimicrobial efficacy without displaying any cement-related cytotoxicity[34]. AgNP coated bone cement resulted in antibacterial activity against MRSA and decrease the formation of polymer debris in joint replacement[35].

TOXICITY OF AGNP

Though nanotechnology poses a greater advantage in clinical applications, a considerable note of precautions and toxic effects are observed with AgNP. The interaction of AgNP with biological media leads to Ag agglomeration and dissolution. Doping of Ag into NP results in the induction of toxic responses. The failure of protective coatings on NPs to prevent aggregation in biological fluids leads to AgNP instability. *In vitro* and *in vivo* AgNP studies demonstrated the induction of lung fibroblasts, genotoxicity, chromosomal aberrations, DNA damage, and apoptosis of cells. There is only limited evidence for carcinogenicity in any biological tissues.

The interaction of human alveolar basal epithelial cells with AgNP results in the generation of reactive oxygen species, reduction of mitochondrial membrane potential and cellular viability, and enhance cellular apoptosis. Exposure to higher concentrations of AgNP in human cell lines induces not only cellular apoptosis but also induces cellular morphology and genetic mutations. The toxicity of AgNP has been demonstrated in microbes ranging from bacteria, viruses, fungi, and algae. AgNP molecules penetrate the skin and blood tube of zebrafish larvae in aggregate form whereas it induces heat shock, oxidate stress, and DNA damage in *Drosophila melanogaster*.

AgNP technology is associated with developmental abnormalities in zebrafish embryos, cytotoxic, and genotoxic effects with systemic immunosuppression. AgNP-mediated cytotoxicity with 10 µg/g is observed on primary human MSCs and osteoblast cells[36]. In zebrafish, AgNP induced neurotoxicity and persistent abnormal behavior[36]. Cytotoxicity of AgNP depends on the particle size of Ag. Literature has documented the cytotoxic effects of Ag of 24 nm causing an intrinsic pro-inflammatory response and apoptosis of surrounding cells[36].

Drake *et al*[37] and Lansdown[38] have done an extensive review of the exposure-related health effects of silver and its related compounds in health. The evidence of toxic effects and complications of AgNP in human studies is limited. Silver represents occupational health hazards like argyria in long-term exposure[37]. The critical oral dosage of silver varies for every individual. The accumulation of silver and nanosilver particles occur in the liver, spleen, kidney, nails, and mucous membranes[39]. With the topical application of Ag, the risk of percutaneous absorption is very low as the epidermis is a relatively impenetrable barrier[40]. Munger *et al*[41] performed a cross-over time exposure study with oral AgNP (5-10 nm) and demonstrated no change in metabolic, hematologic, physical, or morphological findings. However, the toxicity of AgNP in humans is understudied.

Due to the increase of AgNP usage in the market, governmental regulations in the United States [Environment Protection Agency on the nanomaterials regulation], European Union [European Strategy for Nanotechnology], and Canada [Health Canada and Environment Canada] have been implemented[42]. These agencies mentioned that the size of AgNP must be ranging from 1–100 nm in at least one spatial dimension[42]. No specific occupational exposure limits have been laid by these governmental agencies on AgNPs. Global organizations focus on the safety and sustainability of AgNPs in the market for optimal benefits in the community.

RECENT ADVANCES IN AGNP TECHNOLOGY

With the introduction of new fabricating methods, the toxic effects of AgNP are minimized in the microenvironment[43]. Doping of copolymers and growth factors with AgNPs is more effective in hastening wound healing. Green-synthesized AgNPs are cheaper and eco-friendly for the desired environment. The phytochemicals to be doped with AgNPs have to be characterized and seek appropriate regulatory approvals before commercializing the product for preclinical and clinical studies[44,45]. The addition of electrospun nanofibers along with AgNP offers a great advantage in curbing the infection along with debridement and antibiotics. Once the wound is healed, such electrospun nanofibers provide a naïve extracellular matrix in the newly regenerated tissues[46].

With the evolution of 3-D printing technology, the fabrication of scaffolds with Ag coating prevents infections in complex orthopaedic cases[47,48]. 3D scaffolds with porous structures are ideal for loading biomolecules and ions for targeting the desired site. Doping of AgNP into scaffolds with antimicrobial activity and biocompatibility properties in musculoskeletal tissues aids in curbing infection and promoting tissue regeneration as depicted in Figure 2.

3D bioprinting technology dispenses “bio-inks” which contain cells with regenerative potential, scaffolds doped with metal/ion nanoparticles, and biomimetic molecules in a temporospatial controlled fashion[49,50]. Such bio-inks with antimicrobial properties aim at uprooting the infection and facilitating the regeneration of tissues. Damle *et al*[51] proved the proliferation and differentiation of mesenchymal stromal cells when doped with AgNPs which gave further insights in

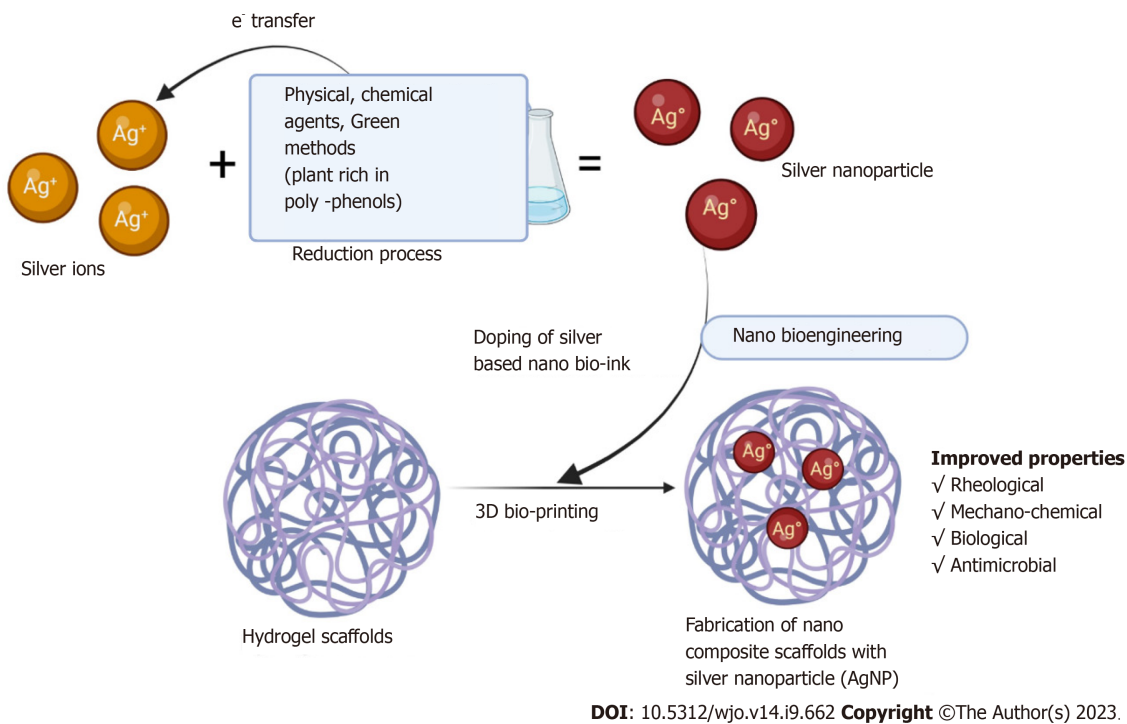


Figure 2 Doping of silver nanoparticles in scaffolds.

osseous tissue engineering. The concept of “Smart Coating” depends on light responsiveness, temperature responsiveness, pH responsiveness, and piezo responsiveness for improving osseous integration, inhibiting biofilm formation, and preventing post-operative complications associated with orthopaedic infections[52].

CONCLUSION

In orthopaedics, AgNP technology has the potential to reduce implant-related orthopaedic infections. Doping AgNP with scaffolds and bio-inks must adhere to the regulatory guidelines to avoid toxicity in clinical applications. With the evidence of preclinical studies, large-scale blinded controlled trials on AgNP in orthopaedic infections have to be assessed for further validation.

FOOTNOTES

Author contributions: Jeyaraman M conceptualized the study; Jeyaraman M, Jeyaraman N, Nallakumarasamy A, Iyengar KP and Jain VK contributed to writing, reviewing, and editing the manuscript; Jeyaraman M, Iyengar KP, Potty AG, and Gupta A supervised the study; All authors contributed to the article and approved the submitted version.

Conflict-of-interest statement: All authors declare no conflict of interest.

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Country/Territory of origin: India

ORCID number: Madhan Jeyaraman 0000-0002-9045-9493; Naveen Jeyaraman 0000-0002-4362-3326; Arulkumar Nallakumarasamy 0000-0002-2445-2883; Karthikeyan P Iyengar 0000-0002-4379-1266; Vijay Kumar Jain 0000-0003-4164-7380; Anish G Potty 0000-0001-9894-1500; Ashim Gupta 0000-0003-1224-2755.

S-Editor: Liu JH

L-Editor: A

P-Editor: Liu JH

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