

Feng Han et al.

1 **Global brain activity and its coupling with cerebrospinal fluid flow is**
2 **related to tau pathology**

3

4

5 Feng Han^{1*}, JiaQie Lee¹, Xi Chen^{1,2}, Jacob Zientz¹, Tyler Ward¹, Susan M Landau¹,
6 Suzanne L Baker², Theresa M Harrison¹, William J Jagust^{1,2}, for the Alzheimer’s Disease
7 Neuroimaging Initiative[†]

8 **Affiliations:**

9 ¹ Helen Wills Neuroscience Institute, University of California, Berkeley, Berkeley, CA,
10 USA;

11 ² Lawrence Berkeley National Laboratory, Berkeley, CA, USA;

12 *Correspondence to: feng.han@berkeley.edu

13

14 [†] Data used in preparation of this article were obtained from the Alzheimer’s Disease
15 Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the
16 investigators within the ADNI contributed to the design and implementation of ADNI
17 and/or provided data but did not participate in analysis or writing of this report. A
18 complete listing of ADNI investigators can be found at:

19 http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf

Feng Han et al.

1 **Keywords:** cerebrospinal fluid flow; global resting-state fMRI signal; glymphatic system;
2 high-order association regions; tau deposition; propagating wave of brain activity,
3 Alzheimer's disease pathology.

4

5 **Address all correspondence to:**

6 Feng Han

7 250 Earl Warren Hall

8 Helen Wills Neuroscience Institute

9 University of California, Berkeley

10 Berkeley, CA 94720

11 Email: feng.han@berkeley.edu

12 **One Sentence Summary:**

13 Resting-state global brain activity affects tau deposition through the potential
14 involvement of a glymphatic clearance function.

15

1 **ABSTRACT**

2

3 Amyloid- β ($A\beta$) and tau deposition constitute Alzheimer's disease (AD)
4 neuropathology. Cortical tau deposits first in the entorhinal cortex and hippocampus and
5 then propagates to neocortex in an $A\beta$ -dependent manner. Tau also tends to accumulate
6 earlier in higher-order association cortex than in lower-order primary sensory-motor
7 cortex. While previous research has examined the production and spread of tau, little
8 attention has been paid to its clearance. Low-frequency (<0.1 Hz) global brain activity
9 during the resting state is coupled with cerebrospinal fluid (CSF) flow and potentially
10 reflects glymphatic clearance. Here we report that tau deposition in subjects with
11 evaluated $A\beta$, accompanied by cortical thinning and cognitive decline, is strongly
12 associated with decreased coupling between CSF flow and global brain activity.
13 Substantial modulation of global brain activity is also manifested as propagating waves of
14 brain activation between higher- and lower-order regions, resembling tau spreading.
15 Together, the findings suggest an important role of resting-state global brain activity in
16 AD tau pathology.

17

1 INTRODUCTION

2 Alzheimer's disease (AD) is pathologically characterized by the extracellular
3 accumulation of amyloid- β ($A\beta$) plaques and the intracellular accumulation of
4 hyperphosphorylated tau in the form of neurofibrillary tangles (1–4). Converging
5 evidence has shown that $A\beta$ accelerates tau phosphorylation and promotes tau
6 aggregation and oligomerization, while $A\beta$ toxicity is dependent on tau (5–7). Tau plays
7 an especially critical role in cognitive decline (8), brain atrophy, particularly cortical
8 thinning (9, 10) and neuronal and synaptic loss (8, 11). For example, a recent study has
9 shown that tau burden measured with positron emission tomography (PET) predicts the
10 severity and topography of subsequent cortical atrophy in AD patients (12). Recently, tau
11 has received special attention as a potential therapeutic target of AD. The deposition of
12 tau aggregates in AD follows a stereotypical pattern, beginning in the entorhinal cortex
13 and hippocampus and then propagating to distinct regions of interest (ROIs) that have
14 been characterized in postmortem and imaging studies as Braak Stages (13, 14). Recent
15 studies have characterized tau deposition as occurring later in lower-order primary
16 sensory-motor (SM) regions but earlier in higher-order association regions that can be
17 captured through the examination of network, such as the default mode network (DMN)
18 and frontoparietal network (FPN) (15, 16). The neural mechanism underlying such a
19 stereotyped pattern of tau accumulation remains elusive but is likely multifactorial.
20 Current hypotheses have attributed tau spreading to neural activity (17, 18), and a “prion-
21 like” mechanism manifested as abnormal tau seeds transferring through anatomically
22 (19–23) and functionally (24–26) connected brain regions.

Feng Han et al.

1 Beyond the accumulation of tau pathology, its clearance may be anti-correlated
2 with deposition and thus has received increasing attention (27, 28). For example, recent
3 studies have identified the critical role of glymphatic function in clearing brain wastes,
4 including A β and tau, via a pathway involving cerebrospinal fluid (CSF) flow, and the
5 exchange between CSF and interstitial fluid (ISF), as well as ISF solutes (27, 29, 30). In
6 mice, the sleep-wake cycle has been found to regulate ISF tau, and sleep deprivation can
7 significantly increase ISF and CSF tau as well as tau spreading (31), presumably due to
8 inadequate sleep-dependent glymphatic clearance (29). This sleep-dependent feature
9 links glymphatic function with spontaneous low-frequency (< 0.1 Hz) resting-state global
10 brain activity assessed with the global blood-oxygen-level-dependent (global BOLD,
11 gBOLD) signal in functional MRI that increases during light sleep or low arousal states
12 (32–36). More importantly, CSF movement, a key determinant of glymphatic function, is
13 coupled with gBOLD signal during both sleep (37) and wakefulness (38). The coupling
14 strength between CSF inflow and gBOLD fMRI signal has recently been proposed as a
15 method to evaluate glymphatic function, and found to be related to cortical A β in an early
16 AD cohort (38, 39), cognitive decline in AD and Parkinson’s disease patients (38, 40),
17 and aging (41).

18 Of note, a recent study has linked the early accumulation of cortical A β in higher-
19 order association regions to the local brain activity there and its coupling to CSF inflow
20 (39), which also suggests the potential role of both global and regional low-frequency (<
21 0.1 Hz) neural activity in driving glymphatic clearance of A β . Consistent with this
22 notion, the spread of A β and tau aggregates, showing similar although not identical
23 predominance for higher-order cognitive networks over primary sensory-motor networks

Feng Han et al.

1 (15, 16, 42), has been hypothesized to follow the direction of glymphatic inflow (43).
2 Resting-state global brain activity, measured by gBOLD or brain-wide
3 electrophysiological signals, is strongly associated with lower-order sensory-motor
4 activity (44–46), resembling the opposite spatial pattern of tau spreading (42). In
5 addition, global brain activity often takes the form of dynamic propagating waves
6 (around gBOLD peaks) between higher-order association and lower-order sensory-motor
7 regions (39, 47, 48), also showing spatial correspondence with tau spreading. These
8 dynamic propagating waves at gBOLD peaks have also been found to attenuate with the
9 decrease of A β 42 in CSF in the earliest AD stage (39), which further suggests the close
10 link between AD pathology and the gBOLD-related propagating waves, presumably
11 through glymphatic function. Together, these findings point to key questions: Does
12 coupling between the global brain activity and CSF movement (gBOLD-CSF coupling),
13 assessing glymphatic function, explain inter-subject variability of tau? Are propagating
14 waves around gBOLD peaks related to glymphatic function, and do they explain the
15 spatial pattern of tau deposition and spreading?

16 To address these hypothetical questions, we examined multimodal data from the
17 Alzheimer’s Disease Neuroimaging Initiative-3 (ADNI3) to investigate the relationship
18 among the coupling between gBOLD and CSF inflow fMRI signals, tau distribution
19 measured with PET, cortical thickness, cognitive function, and dynamic propagating
20 waves.

21

1 **RESULTS**

2 *Cohort demographics*

3 We analyzed resting-state fMRI (rsfMRI), structural MRI, and PET data from 115
4 participants (72.5 ± 7.8 years; 60 females) in the ADNI-3 project (49, 50). The
5 participants included 6 AD patients, 42 with mild cognitive impairment (MCI), 5 subjects
6 with subjective memory concern (SMC), and 62 healthy controls. These subjects were
7 selected based on the availability of tau-PET, A β -PET, rsfMRI (advanced sequence only;
8 repetition time [TR] = 0.607 sec or s), and cortical thickness. Cortical [18 F]florbetaben
9 (FBB) and [18 F]florbetapir (FBP) standardized uptake value ratios (SUVRs) were used to
10 separate the A β + (FBB > 1.08 SUVR; FBP > 1.11 SUVR) and A β - subjects following
11 previous studies (51, 52). The diagnostic information (cognitively unimpaired: SMC and
12 control; cognitively impaired: AD and MCI) was also employed to further define the sub-
13 groups reflecting AD progression, including unimpaired A β -, unimpaired A β +, and
14 impaired A β + (see **Table 1** for details). In addition, 20 impaired A β - subjects were
15 included to create comparable samples of A β - and A β + individuals for fMRI-based
16 glymphatic comparison measures, but the impaired A β - individuals were not included in
17 our main analyses of glymphatic function-tau associations). In general, A β + and A β -
18 subjects were different ($p < 0.001$) in age and cognitive function, i.e., Montreal Cognitive
19 Assessment (MoCA), but with a similar sex ratio between males and females. The
20 unimpaired A β - subjects were also relatively younger and had higher MoCA scores
21 compared with the impaired A β + subjects (both $p < 3.7 \times 10^{-3}$).

Feng Han et al.

1 ***fMRI-based glymphatic measure is closely related to tau across A β + subjects***

2 CSF inflow rsfMRI signal was negatively correlated with gBOLD signal with a
3 +4.856 sec time-lag (**Fig. S1C**; with a representative example in **Fig. S1, A and B**), which
4 confirmed the strong coupling between the two suggested by previous work (38). The
5 gBOLD-CSF coupling relevant to glymphatic function (38) was positively correlated
6 with tau in most cortical regions across all subjects, i.e., subjects with more cortical tau
7 deposition had weaker (less negative) coupling (**Fig. 1A**), similar to the trend noted
8 previously of reduced coupling strength in subjects with more A β (38). This association
9 was significant for regional tau deposition in the Braak V-VI ROI (isocortical regions),
10 Braak III-IV ROI (limbic area; see ref. (14) for detailed cortical regions), and temporal
11 meta-ROI (53) across the whole cohort (all $r > 0.21$, all $p < 0.026$; $N = 115$; **Fig. 1B**).
12 Results were similar in the Braak I ROI/entorhinal cortex although not significant ($r =$
13 0.14 , $p = 0.12$). The strong coupling-tau relationship was evident for A β + and impaired
14 A β + subjects (**Fig. 1, C-F**), but not for subjects who were A β - or unimpaired A β +. (**Fig.**
15 **S2**). Of note, tau remained strongly correlated with the gBOLD-CSF coupling after
16 adjusting for the subjects' motion level (**Fig. S3**).

17 ***Tau partially mediates the strong association between gBOLD-CSF coupling and***
18 ***cortical thickness***

19 We next examined the coupling-thickness association, as tau is closely associated
20 with cortical atrophy and ultimately leads to cognitive decline (12, 54). Similar to the
21 coupling-tau links above, gBOLD-CSF coupling was also significantly related to cortical
22 thickness in widespread cortical regions, including the Braak V-VI ROI, Braak III-IV

Feng Han et al.

1 ROI, and temporal meta-ROI, across the entire group of subjects, and more specifically
2 among the impaired A β ⁺ subjects (**Fig. 2**), rather than other groups (**Fig. S4**).

3 Cortical tau and thickness show a similar spatial distribution pattern averaged
4 across the entire cohort and when contrasting A β ⁺ and A β ⁻ subjects (both $p < 1.8 \times 10^{-4}$;
5 **Fig. 3, A and B**). Cortical tau and thickness were also closely associated across subjects
6 (all $p < 0.05$; **Fig. S5**). Given the corresponding spatial pattern as well as the predictive
7 role of tau in brain atrophy (12), we used mediation analysis to examine whether tau in
8 Braak V-VI ROI, Braak III-IV ROI, and the temporal meta-ROI mediated the observed
9 relationship between gBOLD-CSF coupling and cortical thickness in the same regions.
10 We found that tau in the three ROIs played a significant role in mediating the coupling-
11 thickness link across the whole cohort and the A β ⁺ group ($p < 0.05$; **Fig. 3, C and D**),
12 although the tau in Braak V-VI ROI was a marginally significant mediator among A β ⁺
13 subjects ($p = 0.09$). The mediation effect was less robust in impaired A β ⁺ subjects (**Fig.**
14 **3E**; only significant in the meta-temporal region, $p = 0.02$), which may be attributed to
15 the limited sample size of this group ($N = 28$). We also examined whether coupling
16 mediates the tau-thickness relationship. However, the effect size of this mediation was
17 smaller and only significant for limited ROIs among all A β ⁺ and impaired A β ⁺ subjects
18 (**Fig. S6**). Together, these results suggest that tau is a mediator of the association between
19 glymphatic function, reflected by gBOLD-CSF coupling, and cortical thickness.

20 Given the close association between cortical thickness and MoCA scores (**Fig.**
21 **S7**), we also found that tau partially mediated the association between the coupling
22 metrics and MoCA (**Fig. S8**), indicating a role of glymphatic clearance of tau in cognitive
23 decline.

1 ***Preferential tau deposition in high-order regions is related to the gBOLD-CSF***
2 ***coupling decrease***

3 In the pathophysiology of AD, tau aggregates earlier in high-order association
4 regions than in lower-order sensory ones (15, 16, 55, 56). We next examined how this
5 preferential tau deposition in high-order regions is related to gBOLD-CSF coupling.
6 Previous studies have examined patterns of functional connectivity covariance and
7 depicted continuous and slow brain hierarchy changes across cortices with the concept of
8 “principal gradient” (PG; **Fig. 4A**) anchored at one end by lower-order primary
9 sensory/motor regions and at the other end by higher-order association areas (47, 57).
10 Consistent with previous studies (15, 16, 55, 56), we demonstrated preferential tau
11 deposition in higher-order regions by spatially correlating the PG pattern with the group-
12 difference in tau between A β ⁺ and A β ⁻ subjects or group-mean tau maps across Desikan-
13 Killiany-Tourville (DKT-68) parcels (58). Both significant associations (both $r > 0.26$, p
14 < 0.032 ; **Fig. 4B**) suggested that tau deposition and progression increases with the
15 hierarchy across the entire cortex.

16 In the PG map, the DMN and frontoparietal network (FPN) best capture regions at
17 the high-order end of the hierarchical gradient and thus were extracted as a higher-order
18 mask (**Fig. 4C**). Tau deposition in these higher-order regions was greater in A β ⁺
19 compared to A β ⁻ subjects. There was also greater tau burden in impaired A β ⁺ individuals
20 compared to unimpaired A β ⁺ individuals, who in turn showed greater tau burden
21 compared to unimpaired A β ⁻ individuals (all $p < 0.05$, two-sample t-test; **Fig. 4D**),
22 suggesting that tau in higher-order regions tracks with AD severity. Consistent with **Fig.**

Feng Han et al.

1 **1**, reduced gBOLD-CSF coupling was associated with higher-order tau across the whole
2 cohort, $A\beta+$, and impaired $A\beta+$ groups (both $r > 0.23$, $p < 0.030$; **Fig. 4E**).

3 *High-order tau deposition may be attributed to coupling-related brain*
4 *propagations dynamics*

5 Further analyses focused on identifying the brain dynamics underlying the
6 gBOLD-CSF coupling and investigating the association between these dynamics and tau
7 or cognitive measures. Global brain activation often takes the form of infra-slow (< 0.1
8 Hz) propagating waves/events between lower- and higher-order cortical regions (very
9 close to the higher-order tau mask noted above), along the PG direction (47, 48, 57). We
10 thus extracted the dynamic propagating waves of cortical activation around several
11 specific gBOLD peaks following a previously described method (47). The gBOLD
12 propagating waves were manifested as tilted bands in the time-position graphs showing
13 brain activation in the PG-sorted ROIs transiting along the PG direction between lower-
14 order (e.g., sensory-motor [SM] networks) and higher-order (e.g., DMN) regions (**Fig. 5,**
15 **A and B**, first column). The time-position graphs for the lower-order sensory-motor to
16 higher-order association (Low-to-High order or L-H; averaged across $N = 621$ events
17 from 115 subjects) and the High-to-Low order or H-L ($N = 576$; event-based mean)
18 propagations, as well as corresponding spatial patterns (within the -7.2-sec to +7.2-sec
19 windows around gBOLD peaks), are shown in **Fig. 5, A and B**, respectively. In addition
20 to the topographic information about progression of waves from Low-to-High order
21 cortex and the reverse, the bands demonstrate reduced intensity at higher-order cortex,
22 compared to the lower-order end. For each subject, the occurrence rates (the number of
23 propagating events during the rsfMRI scanning within approximately 10 minutes) of the

Feng Han et al.

1 bidirectional propagation were summed. Subjects with more propagation events (larger
2 occurrence rates) had less tau deposition in the higher-order cortices ($r = - 0.23$, $p =$
3 0.015 ; **Fig. 5C**), stronger gBOLD-CSF coupling ($r = - 0.21$, $p = 0.022$; **Fig. 5D**;
4 especially for the A β ⁺ and impaired A β ⁺ groups shown in **Fig. S9**), and higher MoCA
5 scores ($r = 0.20$, $p = 0.040$; **Fig. 5F**; consistent with the strong association between
6 gBOLD-CSF coupling and MoCA in **Fig. S10**). Neither of these close links between
7 propagation occurrence and tau, coupling, or MoCA was affected by the number of
8 gBOLD peaks over the rsfMRI time-course (**Fig. S11**), suggesting the important role of
9 specific gBOLD peaks with propagating waves in tau pathology. We found no significant
10 correlation between the propagation frequency and thickness in higher-order association
11 regions ($r = 0.10$, $p = 0.27$), suggesting a weak link between the brain structure and the
12 dynamic propagating activity.

13 We summarized the multiple associations among gBOLD-CSF coupling,
14 propagation, tau, thickness, and MoCA in **Fig. 5F**. As depicted in this schematic, global
15 brain activity-related coupling metrics, indicative of glymphatic function, and associated
16 dynamic propagating waves modulate cortical tau, thickness, and ultimately affect
17 cognitive performance.

18 ***Relationship between coupling weakening and reduced one-way propagations***
19 ***depends on AD stages***

20 We finally explored the changes of cortical propagations in each of two directions
21 across AD stages and whether the propagation of cortical activity would account for the
22 strength of gBOLD-CSF coupling. The summed occurrence rates of both bidirectional
23 propagations and the L-H ones decreased from A β ⁻ to A β ⁺ stages, and from unimpaired

Feng Han et al.

1 A β - to unimpaired A β + stages (all $p < 0.057$, two-sample t-test), but not from unimpaired
2 A β + to impaired A β + stages (**Fig. S12, A and B**). A similar but weaker trend was also
3 found in the direction of H-L propagation.

4 For the L-H propagation direction, gBOLD-CSF coupling decreased with reduced
5 propagation occurrence across A β - and unimpaired A β - subjects (both $r < - 0.30$, $p <$
6 0.016 ; **Fig. S12C**), while the association was marginally significant across the whole
7 cohort ($r = - 0.17$, $p = 0.071$). In other words, in A β - subjects, the coupling-propagation
8 relationship in **Fig. 5D** may be attributed to the reduced L-H propagation. For the H-L
9 propagation direction, the coupling was strongly associated with decreased propagation
10 frequencies only across A β + and impaired A β + individuals (both $r < - 0.36$, $p < 0.018$;
11 **Fig. S12D** [$r = - 0.30$, $p = 0.040$ and $r = - 0.34$, $p = 0.083$ after excluding the outlier
12 sample with most propagation events in the A β + and impaired A β + group, respectively]),
13 which may be related to the decreased high-order DMN activity in A β + stage (59–62)
14 hindering the propagation initiated from there.

15 **DISCUSSION**

16 We show a close association between resting-state global brain activity and tau
17 pathology, including the spatiotemporal pattern of tau deposition, brain atrophy, and
18 cognitive decline. The coupling between global brain activity, quantified by gBOLD, and
19 CSF movement was attenuated and correlated with tau deposition in widespread
20 neocortical regions characterized by relatively later-involved Braak stage ROIs (V-VI
21 and III-IV). This finding was evident in the whole cohort but more striking for the A β +
22 and impaired A β + subjects, as would be expected since tau burden is likely to be greater
23 for these groups in later-involved neocortical regions. Declining glymphatic function,

Feng Han et al.

1 reflected by weaker gBOLD-CSF coupling, was also strongly associated with reduced
2 cortical thickness in the same regions, likely related to the mediating role of tau. With the
3 significant aggregation of A β , tau deposition is seen earlier in higher-order cortices over
4 lower-order ones (15, 16, 55, 56). ■ This preferential deposition of tau is related to
5 impaired gBOLD-relevant glymphatic function and to the dynamics of propagating
6 waves of brain activation between higher- and lower-order cortical regions, occurring at
7 gBOLD peaks, that resembles the spatiotemporal pattern of tau spreading and explains its
8 inter-subject variability. Together, these results suggest that resting-state global brain
9 activity modulates the stereotyped pattern of tau deposition in neocortex among
10 individuals with elevated A β , presumably via its effect on glymphatic clearance.

11 Pathological tau aggregation has received increasing attention because of its
12 strong relation to brain atrophy and cognitive impairment (8, 11). While a majority of
13 recent studies have attributed the stereotyped pattern of tau accumulation over cortical
14 regions to neural activity and anatomical and functional connectivity (17–26), other data
15 has repeatedly demonstrated that the brain clearance system affects tau pathology (27)
16 through the glymphatic pathway (30, 31). The glymphatic pathway clears brain wastes
17 through CSF movement pushing the exchange between CSF and ISF and its solutes,
18 including A β and tau (30, 31).

19 Several lines of evidence support the link between resting-state global brain
20 activity and glymphatic clearance. First, global brain activity, measured by gBOLD fMRI
21 signal and whole brain electrophysiology signals (44, 45, 63), are coupled with CSF
22 movement (37–41), a key determinant of glymphatic flow (27, 29, 30). This coupling is
23 particularly striking during sleep (37), when glymphatic function can be 20-fold stronger

Feng Han et al.

1 than wakefulness (29). More recently, the coupling between gBOLD and CSF inflow
2 rsfMRI signals has been found to be correlated with various AD risk factors, cortical A β
3 deposition (38, 39), older age (41), and cognitive decline in AD and Parkinson's disease
4 (PD) (38, 40), further supporting the relationship between global brain activity and
5 glymphatic function. Second, neuronal firing cascades that underlie global spontaneous
6 brain events and gBOLD signal are often accompanied by modulation of sympathetic
7 outflow, including cardiac and respiratory pulsations (64–68), heart rate variability (69),
8 and pupil size (70–72). The sympathetic activity could not only facilitate peri-arterial
9 CSF movements via arterial constriction (66, 73) but also arouse slow ($< 0.1\text{Hz}$)
10 modulations of cardiac and respiratory pulsations, considered as the major driving forces
11 of glymphatic CSF movement (74–76). Third, the intrinsic subcortical vasoactive
12 pathways (73), particularly the basalo-cortical projections (77) relevant to the cholinergic
13 system and astrocytes, are involved with modulation of global brain activity on vascular
14 tone (66). Importantly, while a small proportion of perivascular neurons have direct
15 contact with the vessel wall, most abut astrocytic endfeet (73) constituting the astroglial
16 aquaporin-4 (AQP4) channels that facilitate glymphatic flow (30, 78). A recent study
17 suggested that intrinsic large astrocytic Ca^{2+} spikes were coupled with the negative
18 gBOLD peaks (79). In short, global brain activity and specific neural and physiological
19 factors play a critical role in supporting glymphatic flow and thus affect the AD
20 progresses via moderating the “accumulation-removal balance” of toxic proteins, such as
21 A β and tau.

22 Recent studies in animal models have repeatedly demonstrated the role of
23 glymphatic function in tau aggregation (31, 80–82). Using intracortical injection of

Feng Han et al.

1 human tau into mice, a previous study tracked the tau clearance pathway and found that
2 tau can be removed by CSF flow via glymphatic routes, especially for those with
3 traumatic brain injury (TBI), a risk factor for tau aggregation (80). Further investigations
4 on tau and glymphatic function suggested that tau is cleared from brain by an AQP4-
5 dependent mechanism (81, 82). For example, a recent mouse study has suggested that the
6 impaired CSF-ISF exchange and AQP4 polarization, especially using an AQP4 inhibitor,
7 in the glymphatic system could exacerbate or even induce pathogenic accumulation of tau
8 (81). The same study further showed an inverse association between glymphatic function
9 and tau deposition in the healthy mouse cortex (81). In addition, the glymphatic system
10 was hypothesized to affect the cell-to-cell propagation of tau in brain (83), since tau can
11 be secreted and taken up by both neurons and glia (84, 85), and tau secretion to the
12 extracellular space plays an important role in intracellular tau spreading (80, 86, 87).
13 Beyond these mouse studies, a recent human study identified the relationship between
14 reduced whole-brain glymphatic activity, assessed with diffusion tensor image analysis
15 along the perivascular space (DTI-ALPS), and the deposition of tau using PET along with
16 cognitive decline (88). All these studies support our findings showing the strength of
17 gBOLD-CSF coupling reflecting glymphatic function decreased with tau deposition and
18 cognitive impairment.

19 It is not surprising that tau mediates the close association between glymphatic
20 function and cortical thickness. Our results showing a close association between coupling
21 and atrophy is consistent with a recent study showing that the impaired glymphatic
22 function in TDP-43 transgenic mice, mimicking the pathology of amyotrophic lateral
23 sclerosis (ALS), was accompanied by neocortical atrophy (89). More importantly,

Feng Han et al.

1 middle-aged AQP4 knock-out mice showed elevated tau in both CSF and hippocampus,
2 as well as severe brain atrophy with thinner cortices and hippocampus (82). This brain
3 atrophy was attributed to neuronal loss, including the reduction of dentate granule cells
4 and pyramidal cell layer neurons in the piriform cortex, presumably modulated by tau
5 aggregation induced by the AQP4 deficiency (82).

6 AD-related tau predominantly deposits in higher-order cognitive networks over
7 lower-order sensory-motor networks (15), consistent with our finding that tau deposition
8 follows this brain hierarchy. This is compatible with significant evidence for early tau
9 accumulation regions resembling connectivity-defined networks (19–23). Moreover,
10 global brain activity, profoundly affecting glymphatic function, shows a sensory-
11 dominant pattern opposite to early tau deposition (45), and propagates as waves traveling
12 between higher- and lower-order cortical regions (47). Interestingly, the identified
13 propagating waves in our study displayed higher intensity in lower-order SM cortex, in
14 both directions whether “Low-to-High” and “High-to-Low” (dark red elements in the
15 time-position graphs, **Fig.5 A and B**), consistent with the hypothesis that the stronger
16 brain activity during propagating waves in the lower-order regions may prevent tau
17 deposition there. Consistent with this notion, a previous study suggested that ADNI
18 participants in the earliest stages of AD with stronger SM activity during propagation
19 appeared to have higher CSF A β 42 (**Fig.5** in (39)). These notions thus not only shore up
20 the tight link between the glymphatic function, reflected by the gBOLD-CSF coupling,
21 and the propagating waves at gBOLD peaks, but also support the critical role of dynamic
22 factors in the spatiotemporal pattern of tau deposition. It is worth noting that increased
23 neuronal activity stimulates the release of tau and enhances tau accumulation (18).

Feng Han et al.

1 However, it has not been previously shown that dynamic brain activity, i.e., propagating
2 waves, has an impact on the propagation of tau pathology. The findings presented here
3 provide initial evidence that the frequency of propagation waves is correlated with tau
4 burden in higher-order brain regions. In addition, recent dynamic functional connectivity
5 analyses have repeatedly shown that higher order association regions, including the DMN
6 and FPN, play prominent roles in AD progression (90, 91), especially the strong
7 correlation between elevated tau accumulation and the declined dynamic activity in DMN
8 and its posterior regions (91), which further confirms the effect of the brain dynamics in
9 high-order DMN on AD pathology. Our findings imply that elevated tau deposition in
10 higher order cortex may be related to failure of initiation of H-L propagating waves or
11 failure of L-H propagating waves to reach association cortex, and the weaker activation
12 there compared to the lower-order SM network. Our data warrants future follow-up
13 studies to validate and extend the findings. Together, the dynamic global brain activity
14 may affect the spatiotemporal pattern of tau deposition.

15 Exploring dynamic brain activity, particularly the directional propagating waves
16 that drive glymphatic function, is of high scientific interest, but complicated by the fact
17 that glymphatic flow and brain activity are difficult to record simultaneously *in-vivo* in
18 the human brain. Interestingly, the present study not only linked the occurrence of
19 propagation waves to gBOLD-CSF coupling, reflecting glymphatic strength across the
20 entire cohort of subjects, but further identified that propagation in two opposite directions
21 could account for the glymphatic function in distinct populations. Specifically, among
22 (unimpaired) A β - subjects, coupling reductions can be explained by the occurrence of L-
23 H propagation decrease, which is also related to significant increase of cortical A β

Feng Han et al.

1 deposition (i.e., from A β - to A β +). Previous findings from the ADNI cohort have also
2 demonstrated that the reduction of CSF A β -42 is accompanied by both reduced gBOLD-
3 CSF coupling and weaker L-H propagation among A β - subjects with significantly lower
4 CSF A β -42 (39). On the other hand, H-L propagation may account for coupling changes
5 in later AD progression (i.e., across impaired A β + subjects; **Fig. S12D**), when glymphatic
6 function reflected by the coupling metrics was closely associated with tau deposition,
7 suggesting the potential link between H-L propagation and tau pathology (although the
8 frequency of H-L propagation may not be sensitive enough to account for the tau in high-
9 order regions; $r = -0.14$, $p = 0.15$). This might result from the decreased DMN activity
10 often occurring at the A β + stage (59–62) and interfering with the start of H-L
11 propagating waves there. However, these interpretations should be further confirmed with
12 a more precise quantification of dynamic brain activation.

13 ■ Our current study systematically demonstrates that gBOLD-CSF coupling
14 reflecting glymphatic function and underlying propagation dynamics is strongly related to
15 tau deposition and further affects cortical thickness and cognitive function. Previous
16 evidence has repeatedly reported that tau deposition reduces cortical thickness (12, 92)
17 and further leads to cognitive decline (8, 93). The current study first links the dynamic
18 global brain activity and relevant glymphatic clearance to cortical tau and then suggests
19 the effect of glymphatic function on brain atrophy and cognitive change through the
20 mediating role of tau. It is worth noting that ■ a recent single neuron recording study
21 linking global brain activity to the memory system has shown that a spiking cascade of
22 widespread neurons featuring sequential activation, similar to the global brain dynamics,
23 strongly modulated the occurrence of hippocampal sharp-wave ripples (70) that play a

Feng Han et al.

1 critical role in memory consolidation (94, 95). This suggests the close link between
2 resting-state global brain activity and cognitive function, consistent with our finding that
3 the MoCA score is related to both gBOLD-CSF coupling and the propagating waves.

4 There are a few limitations of the present study. First, we used a cross-sectional
5 analysis and evaluated the spatiotemporal pattern of tau spreading through its between-
6 group differences (i.e., A β + vs. A β -). A longitudinal design should be used to test tau
7 accumulation rates linked to fMRI-based coupling or propagating waves. Second, the
8 global propagating waves between higher-order association regions and lower-order
9 sensory-motor networks is one type of propagation event at some gBOLD peaks but do
10 not thoroughly account for the dynamic activations underlying global brain activity or its
11 relevant glymphatic movements. Future studies on an independent dataset with a larger
12 sample size are needed to validate and extend our findings on the global brain dynamics
13 underlying glymphatic flow.

14 In summary, this study provides initial evidence that glymphatic function,
15 reflected by the coupling between global brain activity and the CSF movement signals, is
16 closely associated with tau pathology in people with elevated A β deposition. Global
17 dynamic brain activity is related to glymphatic function and may in part explain the
18 preferential exposure of higher-order cognitive brain regions to tau deposition.

19 **MATERIALS AND METHODS**

20 *Participants and data*

21 We included 115 participants from the ADNI-3 project according to the
22 availability of tau-PET (18F-Flortaucipir [FTP]; AV-1451), A β -PET (either florbetaben

Feng Han et al.

1 [FBB] or florbetapir [FBP]), rsfMRI (TR = 0.607 sec sessions only), and structural MRI
2 (cortical thickness) data. Participants included 4 different disease conditions defined by
3 ADNI (<http://adni.loni.usc.edu/study-design/>): 6 AD patients, 42 with mild cognitive
4 impairment (MCI), 5 subjects with subjective memory concern (SMC), and 62 healthy
5 controls. We further categorized the participants into “impaired” (AD and MCI) and
6 “unimpaired” (SMC and control) groups based on their clinical diagnosis. Two groupings
7 were applied for our analyses: 1) $A\beta+$ and $A\beta-$ subjects; 2) unimpaired $A\beta-$, unimpaired
8 $A\beta+$, and impaired $A\beta+$ subjects (20 impaired $A\beta-$ subjects were used to augment the
9 sample size of the $A\beta-$ group in the comparison of fMRI-based glymphatic or
10 propagation measures with $A\beta+$ group). We also included participant demographics (age
11 and sex) and cognitive assessment, i.e., Montreal Cognitive Assessment (MoCA) scores
12 in our study. All participants provided written informed consent. Ethical approval from
13 the individual institutional review board (IRB; [http://adni.loni.usc.edu/wp-](http://adni.loni.usc.edu/wp-content/uploads/2013/09/DOD-ADNI-IRB-Approved-Final-protocol-08072012.pdf)
14 [content/uploads/2013/09/DOD-ADNI-IRB-Approved-Final-protocol-08072012.pdf](http://adni.loni.usc.edu/wp-content/uploads/2013/09/DOD-ADNI-IRB-Approved-Final-protocol-08072012.pdf)) has
15 been granted to the investigators at each ADNI participating site. All the ADNI data were
16 collected per the principles of the Declaration of Helsinki.

17 Measurement of $A\beta$ -PET, tau-PET, rsfMRI, cortical thickness, and MoCA were
18 obtained from the same study visit (the time interval between pairwise modalities was no
19 more than 183 days [\sim 6 months] (96)). The file “UC Berkeley – AV1451 PVC 8mm Res
20 Analysis [ADNI2,3] (version: 2023-02-17)” was used to provide the tau-PET SUVR (97–
21 100). Both “UC Berkeley – AV45 8mm Res Analysis [ADNIGO,2,3] (version: 2023-02-
22 17)” and “UC Berkeley – FBB 8mm Res Analysis [ADNI3] (version: 2023-02-17)” were
23 used to provide the $A\beta$ -PET SUVR (51, 52, 101). MoCA score was also directly acquired

Feng Han et al.

1 from ADNI as “Montreal Cognitive Assessment (MoCA) [ADNIGO,2,3]”. All these
2 data, as well as the rsfMRI and cortical thickness, are publicly accessible on the ADNI
3 website (<http://adni.loni.usc.edu/>).

4 The use of de-identified data from ADNI has been approved by the University of
5 California Berkeley and also strictly followed the ADNI data use agreements.

6 ***Image acquisition and preprocessing***

7 All rsfMRI was acquired at 3 Tesla MR scanners from multiple ADNI
8 participating sites following a unified protocol
9 (<http://adni.loni.usc.edu/methods/documents/mri-protocols/>). The MRI data used in the
10 current study was collected in Siemens MRI scanners (Siemens Medical Solutions,
11 Siemens, Erlangen, Germany). Each MR imaging session began with a T1-weighted
12 (T1w) MPRAGE sequence (flip angle = 9° , spatial resolution = $1 \times 1 \times 1 \text{ mm}^3$, echo time
13 [TE] = 3.0 ms, repetition time [TR] = 2,300 ms), which was used for cortical thickness,
14 anatomical segmentation, and registration (see details in
15 <http://adni.loni.usc.edu/methods/documents/>) (102). During rsfMRI acquisition, 976
16 fMRI volumes were collected with an echo-planar image (EPI) sequence with
17 TR/TE=607/32 ms (flip angle = 50° , spatial resolution = $2.5 \times 2.5 \times 2.5 \text{ mm}^3$, slice
18 thickness = 2.5 mm; see details at: <http://adni.loni.usc.edu/methods/documents/>).

19 PET imaging was acquired according to standardized protocols at each ADNI site.
20 FTP-PET data were acquired from 75-105 minutes post-injection of 10 mCi tracer. FBP-
21 PET and FBB PET data were acquired from the 50-70 minutes post-injection of 10 mCi
22 tracer and from the 90-110 minutes post-injection of 8.1 mCi tracer (52, 103, 104),

1 respectively ([https://adni.loni.usc.edu/wp-content/uploads/2012/10/ADNI3-Procedures-](https://adni.loni.usc.edu/wp-content/uploads/2012/10/ADNI3-Procedures-Manual_v3.0_20170627.pdf)
2 [Manual_v3.0_20170627.pdf](https://adni.loni.usc.edu/wp-content/uploads/2012/10/ADNI3-Procedures-Manual_v3.0_20170627.pdf)).

3 We preprocessed structural MRI using FreeSurfer v7.1
4 (<https://surfer.nmr.mgh.harvard.edu/fswiki/DownloadAndInstall5.3>) (105) to derive
5 FreeSurfer ROIs (DKT-68 parcel (58)) in participants' native space and extract the
6 parcel-based cortical thickness. Following the previous study (38), we preprocessed the
7 rsfMRI data using FSL (<https://fsl.fmrib.ox.ac.uk/fsl/fslwiki>) (106) and AFNI
8 (<https://afni.nimh.nih.gov/>) (107) with a modification of excluding rsfMRI sessions with
9 large head-motion (session-mean frame-wise displacement [FD] larger than 0.6 mm or
10 the maximal FD larger than 3 mm) (108). The general procedures for rsfMRI
11 preprocessing include motion correction, skull stripping, spatial smoothing (full width at
12 half maximum [FWHM] = 4mm), temporal filtering (bandpass filter, 0.01 to 0.1 Hz), and
13 the co-registration of each fMRI volume to its anatomical MRI and then to the 152-brain
14 Montreal Neurological Institute (MNI-152) space. Referring to the previous studies (38,
15 68), we also excluded the procedure of motion parameter regression to avoid weakening
16 the gBOLD signal, and removed the first 20 and last 20 volumes for each rsfMRI session
17 to reduce the edge effect from the temporal filtering and to ensure a steady
18 magnetization.

19 We used the tau-PET SUVR data summarized in “UC Berkeley – AV1451 PVC
20 8mm Res Analysis [ADNI2,3] (version: 2023-02-17)”, and the A β -PET SUVR from “UC
21 Berkeley – AV45 8mm Res Analysis [ADNIGO,2,3] (version: 2023-02-17)” (for 42
22 subjects) and “UC Berkeley – FBB 8mm Res Analysis [ADNI3] (version: 2023-02-17)”
23 (for 73 subjects) (51, 97–101). To generate the PET data in DKT-68 parcels, several

Feng Han et al.

1 preprocessing steps were performed, including image averaging, spatial smoothing, and
2 registration to the (structural) MRI space to extract the tau or A β intensity in each DKT-
3 68 parcel (58). We then normalized parcel-wise tau with the inferior cerebellar reference
4 region to derive the tau SUVR and further applied partial volume correction (PVC) to
5 reduce the influence from low image resolution and limited tissue sampling (100).
6 Regarding the A β SUVR, we normalized the FBP or FBB intensity in each DKT-68
7 parcel using the whole cerebellum reference region.

8 *The extraction of gBOLD and CSF inflow signals*

9 Following the previous study (38), we derived the gBOLD signal by averaging
10 the rsfMRI (Z-normalized) time-series across all voxels in the gray-matter region (see a
11 representative example in **Fig. S1B, upper, green**; corresponding to the signal in **Fig.**
12 **S1A, green**). Specifically, we used the Harvard-Oxford cortical and subcortical structural
13 atlases (<https://neurovault.org/collections/262/>) to define the gray matter mask, which
14 was then transformed to the original fMRI space of each subject referring to previous
15 studies (37, 38). The rsfMRI in individual original space went through the above
16 preprocessing procedures (not including nuisance regression; particularly to avoid the
17 CSF signal regression attenuating the CSF inflow signal) (38). The preprocessed rsfMRI
18 signal was averaged in individual gray-matter mask to derive the gBOLD signal for each
19 subject.

20 To derive the CSF inflow signal, the preprocessed fMRI in the original individual
21 space was averaged across the CSF voxels at the bottom slice of fMRI acquisition
22 following previous studies (37, 38). All subjects have their individual CSF masks (**Fig.**

1 **S1B, lower**; corresponding signal in **Fig. S1A, purple**) with similar voxel numbers
2 (~14).

3 *The coupling between the gBOLD signal and the CSF signal*

4 We also calculated the cross-correlation functions between the gBOLD signal and
5 the CSF inflow signal (by assessing Pearson's correlation) at the lag of +4.856 sec, where
6 the negative peak of the mean cross-correlation located (see arrow in **Fig. S1C**), to
7 evaluate the gBOLD-CSF coupling for each subject as the previous study (38).

8 *Stage classifications*

9 Two grouping methods were applied for the entire group of subjects. First,
10 subjects were divided into A β ⁺ and A β ⁻ based on the whole cortical SUVR (AV45–A β >
11 1.11 SUVR or FBB–A β > 1.08 SUVR; the whole cerebellum as reference region).
12 Second, we further incorporated the diagnosis information and staged the subjects into
13 (cognitively) unimpaired A β ⁻, unimpaired A β ⁺, and impaired A β ⁺ groups (unimpaired:
14 SMC and control).

15 *Correlating the gBOLD–CSF coupling to cortical A β , tau, thickness, and MoCA*

16 The gBOLD-CSF coupling was evaluated using their cross-correlation at the lag
17 of +4.856 sec. We first compared the gBOLD-CSF coupling (adjusted for age and sex)
18 across two sets of stages above, including the pairwise comparison (two-sample t-test)
19 and the linear trend evaluation for unimpaired A β ⁻ to unimpaired A β ⁺ to impaired A β ⁺
20 (ordinal regression).

Feng Han et al.

1 For each of these 5 stages and the entire cohort, we correlated the gBOLD-CSF
2 coupling (age- and sex-adjusted) with the cortical tau SUVR in each DKT-68 parcel and
3 then mapped to the brain surface (using the WorkBench software [version: 1.5.0;
4 <https://www.humanconnectome.org/software/workbench-command>]) to see the spatial
5 distribution of correlation coefficients. We further tested the coupling-tau correlation in 4
6 ROIs, including Braak V-VI ROI, Braak III-IV ROI, temporal meta-ROI, and Braak I ROI
7 across subjects within each of these 5 different groups.

8 The coupling-tau association was retested after regressing out the subject-wise
9 head motion measure, the mean FD, from the gBOLD-CSF coupling, especially in the
10 Braak V-VI ROI, Braak III-IV ROI, and temporal meta-ROI among the whole cohort, A β +, or
11 impaired A β + subjects.

12 Similar to the above analyses on the coupling-tau association, we also correlated
13 the gBOLD-CSF coupling with the cortical thickness in the same sets of ROIs or MoCA
14 scores (also examined the MoCA-thickness correlation) across the same groups of
15 subjects.

16 ***Determining the mediator role of tau in the coupling-thickness and***
17 ***coupling-MoCA associations***

18 We first averaged tau SUVR and cortical thickness across all subjects and
19 compared their distribution patterns by correlating them across DKT-68 parcels.
20 Similarly, the tau difference between A β + and A β - groups (reflecting the tau spreading
21 with AD progression) was spatially compared with that difference in thickness. We
22 further evaluated the inter-subject similarity between tau and thickness in each of the
23 above 4 ROIs across subjects within each group of these 5 different stages.

Feng Han et al.

1 Given that the gBOLD-CSF coupling may reveal glymphatic clearance for A β
2 and tau, as well as the role of tau in predicting brain atrophy (12, 92), we then examined
3 the hypothesis that tau mediates the association between gBOLD-CSF coupling and
4 cortical thickness using a mediation analysis (109) in the Braak V-VI ROI, Braak III-IV
5 ROI, or temporal meta-ROI among the groups of the whole cohort, A β +, and impaired
6 A β +, respectively, among which the coupling-tau and coupling-thickness associations
7 were significant in **Figs. 1** and **2**. The possibility that coupling mediated the relationship
8 between tau and thickness was also tested in these ROIs.

9 Same mediation analysis was further applied to test if the coupling-MoCA
10 association would be mediated by the tau, since tau pathology may lead to cognitive
11 impairment (8, 110).

12 *Relating the mean tau pattern and its cross-stage changes to brain hierarchy*

13 Due to the earlier accumulation in higher-order brain regions than lower-order
14 sensory ones (15, 16, 55, 56), we tested whether tau and its cross-stage change followed
15 the brain hierarchy. We thus applied the PG map obtained in (57) to quantify the cortical
16 hierarchy and correlated the tau and PG values across cortical parcels. We also assessed
17 such spatial correspondence between PG and tau changes from A β - to A β + groups.

18 *Quantifying tau in higher-order brain regions*

19 We further defined a higher-order cortical mask following a previous study (39)
20 to better quantify the tau in higher-order regions, which may reflect the spreading pattern
21 of tau during AD progression. We employed the DMN and FPN defined from a previous

Feng Han et al.

1 study (*III*) as the higher-order association regions and then identified the DKT-68
2 parcels belonging to the two networks due to the (DKT-68) parcellation of tau.

3 Averaged tau in higher-order brain parcels was then compared between A β - to
4 A β + groups, as well as between the groups of unimpaired A β -, unimpaired A β +, and
5 impaired A β +. We also replicated the analyses in **Fig. 1** and examined the correlation
6 between the gBOLD-CSF coupling and tau in higher-order parcels across subjects in the
7 whole cohort, A β + stage, and impaired A β + stage, in which the coupling-tau
8 relationships in **Fig. 1** were significant.

9 ***Quantifying gBOLD propagating waves and linking it to tau pathology***

10 We followed previous studies (*39, 47*) to project the rsfMRI signal onto the
11 direction of PG (indicative of cortical hierarchy) and derived the time-position graphs
12 over the whole time-series for each subject. We sorted all the cortical voxels based on
13 their PG values and divided them into 70 position bins of equal size. The rsfMRI time-
14 series was then averaged within each bin and thus generated as a time-position graph with
15 the horizontal direction encoding time and the vertical direction representing 70 bins
16 along the PG direction. The time-position graph was cut into multiple segments
17 according to the troughs of the gBOLD signal (*47*). Within the time-position graph for
18 each subject, we identified several continuous and tilted bands representing brain
19 activation transiting from lower-order sensory regions to higher-order regions along the
20 PG direction, as the L-H propagation events; we also detected and extracted the bands
21 showing brain activation transiting along the opposite direction as the H-L propagation
22 events. We followed the previous studies in event detection (see more details in (*39, 47*)).
23 The general procedures included, within each segment of time-position graph, deriving

Feng Han et al.

1 the timing relative to the global (spatial) mean peak (i.e., gBOLD peak), and correlating
2 the relative timings with the position of the corresponding bin (across 70 bins) along the
3 PG direction. We used a correlation threshold of 0.385 (corresponding to the p -value of
4 0.001) to extract the L-H propagations and -0.385 to detect the H-L ones, applied a bin
5 number threshold of 50 to identify these “continuous” bands (i.e., excluding the
6 segments/events with less than 50 local peaks [each bin row could have a “local peak” or
7 not]), and used a (normalized BOLD) threshold of 0.1 to exclude these bin-rows with too
8 weak activation (i.e., regarded as no “local peak”).

9 The time-position graphs for identified L-H propagation events were averaged
10 (time window from -8-sec to 8-sec; aligned based on the timing of gBOLD peak [0 sec])
11 across all instances from all subjects to derive the mean time-position pattern of the
12 propagation at this direction. This mean time-position graph was then traced back and
13 projected onto the brain surface to generate the spatial patterns of such propagation. The
14 same approach was applied to derive the mean pattern of the H-L propagation and
15 corresponding spatial patterns.

16 We also counted the number of identified L-H propagation and H-L propagation
17 events within the full time-position graph (of equal length in the horizontal direction; ~
18 10mins) for each subject as the propagation occurrence rate or frequency for the
19 respective direction. The frequencies of the two distinct directions were further summed
20 to derive the total propagation frequency for each subject.

21 To investigate the role of dynamic activation propagation in tau, gBOLD-CSF
22 coupling reflecting glymphatic function, cortical thickness, and the cognitive level, we
23 correlated the total propagation frequency (age- and sex-adjusted) with the tau deposition

Feng Han et al.

1 and thickness in higher-order association regions, the gBOLD-CSF coupling, and MoCA
2 score across the whole cohort. We also further examined the propagation-coupling
3 association for each of the 5 different (participants) sub-groups.

4 To demonstrate the role of A β or AD pathology in the frequency of all
5 propagation, the summed frequencies of two directions were also compared between A β -
6 and A β + groups, or between unimpaired A β -, unimpaired A β +, and impaired A β +
7 groups. The frequencies of L-H propagation and the H-L propagation were also
8 compared between the above groups.

9 To investigate the contribution/relation of propagation at each direction to
10 gBOLD-CSF coupling, we correlated the propagation frequency of each direction with
11 gBOLD-CSF coupling at all (participants) sub-groups, including whole cohort, A β -, A β +,
12 unimpaired A β -, unimpaired A β +, impaired A β + groups.

13 *Statistical analysis*

14 A two-sample t-test was performed for group comparisons on continuous
15 measures, including age, gBOLD-CSF coupling, tau SUVR in higher-order regions, and
16 propagation frequency. We used Fisher's exact test (112) to compare categorical
17 measures (i.e., sex) between stages of A β pathology progression. An ordinal regression
18 test was applied to evaluate the linear trend of gBOLD-CSF coupling changes across the
19 unimpaired A β -, unimpaired A β +, and impaired A β + groups. Pearson's correlation was
20 used to quantify the relationship between different variables in **Fig. 5F**. The single-level
21 mediation analysis (109) was performed to evaluate the role of tau in the coupling-
22 thickness and the coupling-MoCA links, as well as the role of coupling in the tau-

Feng Han et al.

1 thickness relationship. A p-value no more than 0.05 was considered statistically
2 significant.

3 To test the effect of head-motion on our major results, we regressed out the mean
4 FD (*113*) from gBOLD-CSF coupling and repeated the analyses on coupling-tau
5 association across the groups of subjects in **Fig. 1**.

6 To examine if the gBOLD peak number drove the close association between the
7 frequency of propagation and tau, coupling, or MoCA in **Fig. 5, C-E**, we correlated the
8 gBOLD peak number with the above three measures across the whole cohort of subjects.
9 We further regressed out the gBOLD peak number from the propagation frequency and
10 re-tested the correlations between the adjusted frequency and tau, coupling, as well as
11 MoCA across the whole cohort, respectively.

12 **ACKNOWLEDGEMENTS**

13 **Funding:** This work is supported by fundings from the following: NIH (U19AG024904
14 to W.J.J. and S.M.L., K01AG078443 to T.M.H., F31AG079595 to J.Z.) and BrightFocus
15 Foundation (A2021004F to X.C.).

16
17 **Author contributions:** F.H. and W.J.J. conceptualized and designed the study; F.H.,
18 J.Q.L., J.Z., T.W., S.M.L., and W.J.J. curated the data and performed analyses; F.H.,
19 X.C., S.L.B., and T.M.H. and W.J.J. designed and performed assays; F.H. and W.J.J.
20 wrote the original draft, and J.Q.L.; F.H., X.C., J.Z., T.W., S.M.L., S.L.B., T.M.H. and
21 W.J.J. reviewed and edited the manuscript; W.J.J. supervised the project; and all authors
22 read and approved the manuscript.

Feng Han et al.

1

2 **Competing interests:** W.J.J. has served as consultant for Biogen, Eisai, Lilly, and
3 Bioclinica. The remaining authors declare no competing financial interests.

4

5 Data collection and sharing for this project was funded by the ADNI (National
6 Institutes of Health Grant U01 AG024904; Principal Investigator: Michael Weiner) and
7 DOD ADNI (Department of Defense award number W81XWH-12-2-0012; Principal
8 Investigator: Michael Weiner). ADNI is funded by the National Institute on Aging, the
9 National Institute of Biomedical Imaging and Bioengineering (NIBIB), and through
10 generous contributions from the following: AbbVie, Alzheimer’s Association;
11 Alzheimer’s Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen;
12 Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan
13 Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd
14 and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.;
15 Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson
16 Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co.,
17 Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies;
18 Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda
19 Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of
20 Health Research is providing funds to support ADNI clinical sites in Canada. Private
21 sector contributions are facilitated by the Foundation for the National Institutes of Health
22 (www.fnih.org). The grantee organization is the Northern California Institute for
23 Research and Education, and the study is coordinated by the Alzheimer’s Therapeutic

Feng Han et al.

1 Research Institute at the University of Southern California. ADNI data are disseminated
2 by the Laboratory for Neuro Imaging at the University of Southern California. The
3 funders had no role in study design, data collection and analysis, decision to publish, or
4 preparation of the manuscript.

5 **DATA AND MATERIALS AVAILABILITY**

6 The multimodal data, including subject characteristics, rsfMRI, structural MRI,
7 A β -PET, tau-PET, and MoCA measures are all publicly available at the ADNI website
8 upon the approval of the data use application (<http://adni.loni.usc.edu/>). The ADNI was
9 launched in 2003 as a public-private partnership, led by Principal Investigator Michael
10 W. Weiner, MD. The primary goal of ADNI has been to test whether serial magnetic
11 resonance imaging (MRI), positron emission tomography (PET), other biological
12 markers, and clinical and neuropsychological assessment can be combined to measure the
13 progression of mild cognitive impairment (MCI) and early Alzheimer's disease (AD). For
14 up-to-date information, see www.adni-info.org. All the code used in the present study are
15 available from the corresponding author upon request.

16

17

1 **REFERENCES**

- 2 1. H. Hampel, J. Hardy, K. Blennow, C. Chen, G. Perry, S. H. Kim, V. L. Villemagne, P.
3 Aisen, M. Vendruscolo, T. Iwatsubo, C. L. Masters, M. Cho, L. Lannfelt, J. L. Cummings,
4 A. Vergallo, The Amyloid- β Pathway in Alzheimer's Disease, *Mol. Psychiatry* **26**, 5481–
5 5503 (2021).
- 6 2. A. del C. Alonso, I. Grundke-Iqbal, H. S. Barra, K. Iqbal, Abnormal phosphorylation
7 of tau and the mechanism of Alzheimer neurofibrillary degeneration: Sequestration of
8 microtubule-associated proteins 1 and 2 and the disassembly of microtubules by the
9 abnormal tau, *Proc. Natl. Acad. Sci.* **94**, 298–303 (1997).
- 10 3. A. del C. Alonso, T. Zaidi, M. Novak, I. Grundke-Iqbal, K. Iqbal,
11 Hyperphosphorylation induces self-assembly of τ into tangles of paired helical
12 filaments/straight filaments, *Proc. Natl. Acad. Sci.* **98**, 6923–6928 (2001).
- 13 4. C. R. Jack, D. A. Bennett, K. Blennow, M. C. Carrillo, B. Dunn, S. B. Haeberlein, D.
14 M. Holtzman, W. Jagust, F. Jessen, J. Karlawish, E. Liu, J. L. Molinuevo, T. Montine, C.
15 Phelps, K. P. Rankin, C. C. Rowe, P. Scheltens, E. Siemers, H. M. Snyder, R. Sperling, C.
16 Elliott, E. Masliah, L. Ryan, N. Silverberg, NIA-AA Research Framework: Toward a
17 biological definition of Alzheimer's disease, *Alzheimer's Dement.* **14**, 535–562 (2018).
- 18 5. H. Zhang, W. Wei, M. Zhao, L. Ma, X. Jiang, H. Pei, Y. Cao, H. Li, Interaction
19 between A β and Tau in the Pathogenesis of Alzheimer's Disease, *Int J Biol Sci* **17**, 2181–
20 2192 (2021).
- 21 6. M. A. Busche, B. T. Hyman, Synergy between amyloid- β and tau in Alzheimer's
22 disease, *Nat. Neurosci.* **23**, 1183–1193 (2020).
- 23 7. G. S. Bloom, Amyloid- β and tau: The trigger and bullet in Alzheimer disease
24 pathogenesis, *JAMA Neurol.* (2014), doi:10.1001/jamaneurol.2013.5847.
- 25 8. A. Bejanin, D. R. Schonhaut, R. La Joie, J. H. Kramer, S. L. Baker, N. Sosa, N. Ayakta,
26 A. Cantwell, M. Janabi, M. Lauriola, J. P. O'Neil, M. L. Gorno-Tempini, Z. A. Miller, H.
27 J. Rosen, B. L. Miller, W. J. Jagust, G. D. Rabinovici, Tau pathology and
28 neurodegeneration contribute to cognitive impairment in Alzheimer's disease, *Brain* **140**,
29 3286–3300 (2017).
- 30 9. T. M. Harrison, R. Du, G. Klencklen, S. L. Baker, W. J. Jagust, Distinct effects of
31 beta-amyloid and tau on cortical thickness in cognitively healthy older adults,
32 *Alzheimer's & Dement.* **17**, 1085–1096 (2021).
- 33 10. R. Ossenkoppele, R. Smith, T. Ohlsson, O. Strandberg, N. Mattsson, P. S. Insel, S.
34 Palmqvist, O. Hansson, Associations between tau, A β , and cortical thickness with
35 cognition in Alzheimer disease, *Neurology* **92**, e601--e612 (2019).

- 1 11. S. L. DeVos, R. L. Miller, K. M. Schoch, B. B. Holmes, C. S. Kebodeaux, A. J.
2 Wegener, G. Chen, T. Shen, H. Tran, B. Nichols, T. A. Zanardi, H. B. Kordasiewicz, E. E.
3 Swayze, C. F. Bennett, M. I. Diamond, T. M. Miller, Tau reduction prevents neuronal
4 loss and reverses pathological tau deposition and seeding in mice with tauopathy, *Sci.*
5 *Transl. Med.* **9**, eaag0481 (2017).
- 6 12. R. La Joie, A. V. Visani, S. L. Baker, J. A. Brown, V. Bourakova, J. Cha, K.
7 Chaudhary, L. Edwards, L. Iaccarino, M. Janabi, O. H. Lesman-Segev, Z. A. Miller, D. C.
8 Perry, J. P. O’Neil, J. Pham, J. C. Rojas, H. J. Rosen, W. W. Seeley, R. M. Tsai, B. L.
9 Miller, W. J. Jagust, G. D. Rabinovici, Prospective longitudinal atrophy in Alzheimer’s
10 disease correlates with the intensity and topography of baseline tau-PET, *Sci. Transl.*
11 *Med.* **12**, eaau5732 (2020).
- 12 13. H. Braak, I. Alafuzoff, T. Arzberger, H. Kretschmar, K. Del Tredici, Staging of
13 Alzheimer disease-associated neurofibrillary pathology using paraffin sections and
14 immunocytochemistry, *Acta Neuropathol.* **112**, 389–404 (2006).
- 15 14. M. Schöll, S. N. Lockhart, D. R. Schonhaut, J. P. O’Neil, M. Janabi, R. Ossenkoppele,
16 S. L. Baker, J. W. Vogel, J. Faria, H. D. Schwimmer, G. D. Rabinovici, W. J. Jagust, PET
17 Imaging of Tau Deposition in the Aging Human Brain, *Neuron* **89**, 971–982 (2016).
- 18 15. O. Hansson, M. J. Grothe, T. O. Strandberg, T. Ohlsson, D. Hägerström, J. Jögi, R.
19 Smith, M. Schöll, Tau Pathology Distribution in Alzheimer’s disease Corresponds
20 Differentially to Cognition-Relevant Functional Brain Networks, *Front. Neurosci.* **11**
21 (2017), doi:10.3389/fnins.2017.00167.
- 22 16. K. A. Johnson, A. Schultz, R. A. Betensky, J. A. Becker, J. Sepulcre, D. Rentz, E.
23 Mormino, J. Chhatwal, R. Amariglio, K. Papp, G. Marshall, M. Albers, S. Mauro, L.
24 Pepin, J. Alverio, K. Judge, M. Philiossaint, T. Shoup, D. Yokell, B. Dickerson, T.
25 Gomez-Isla, B. Hyman, N. Vasdev, R. Sperling, Tau positron emission tomographic
26 imaging in aging and early Alzheimer disease, *Ann. Neurol.* **79**, 110–119 (2016).
- 27 17. A. M. Pooler, E. C. Phillips, D. H. W. Lau, W. Noble, D. P. Hanger, Physiological
28 release of endogenous tau is stimulated by neuronal activity, *EMBO Rep.* (2013),
29 doi:10.1038/embor.2013.15.
- 30 18. J. W. Wu, S. A. Hussaini, I. M. Bastille, G. A. Rodriguez, A. Mrejeru, K. Rilett, D. W.
31 Sanders, C. Cook, H. Fu, R. A. C. M. Boonen, M. Herman, E. Nahmani, S. Emrani, Y. H.
32 Figueroa, M. I. Diamond, C. L. Clelland, S. Wray, K. E. Duff, Neuronal activity
33 enhances tau propagation and tau pathology in vivo, *Nat. Neurosci.* **19**, 1085–1092
34 (2016).
- 35 19. A. Mudher, M. Colin, S. Dujardin, M. Medina, I. Dewachter, S. M. Alavi Naini, E.-M.
36 Mandelkow, E. Mandelkow, L. Buée, M. Goedert, J.-P. Brion, What is the evidence that
37 tau pathology spreads through prion-like propagation?, *Acta Neuropathol. Commun.* **5**, 99
38 (2017).

- 1 20. F. Clavaguera, T. Bolmont, R. A. Crowther, D. Abramowski, S. Frank, A. Probst, G.
- 2 Fraser, A. K. Stalder, M. Beibel, M. Staufenbiel, M. Jucker, M. Goedert, M. Tolnay,
- 3 Transmission and spreading of tauopathy in transgenic mouse brain, *Nat. Cell Biol.* **11**,
- 4 909–913 (2009).

- 5 21. J. W. Vogel, Y. Iturria-Medina, O. T. Strandberg, R. Smith, E. Levitis, A. C. Evans,
- 6 O. Hansson, M. Weiner, P. Aisen, R. Petersen, C. R. Jack, W. Jagust, J. Q. Trojanowki, A.
- 7 W. Toga, L. Beckett, R. C. Green, A. J. Saykin, J. Morris, L. M. Shaw, E. Liu, T.
- 8 Montine, R. G. Thomas, M. Donohue, S. Walter, D. Gessert, T. Sather, G. Jiminez, D.
- 9 Harvey, M. Donohue, M. Bernstein, N. Fox, P. Thompson, N. Schuff, C. DeCarli, B.
- 10 Borowski, J. Gunter, M. Senjem, P. Vemuri, D. Jones, K. Kantarci, C. Ward, R. A.
- 11 Koeppe, N. Foster, E. M. Reiman, K. Chen, C. Mathis, S. Landau, N. J. Cairns, E.
- 12 Householder, L. T. Reinwald, V. Lee, M. Korecka, M. Figurski, K. Crawford, S. Neu, T.
- 13 M. Foroud, S. Potkin, L. Shen, F. Kelley, S. Kim, K. Nho, Z. Kachaturian, R. Frank, P. J.
- 14 Snyder, S. Molchan, J. Kaye, J. Quinn, B. Lind, R. Carter, S. Dolen, L. S. Schneider, S.
- 15 Pawluczyk, M. Beccera, L. Teodoro, B. M. Spann, J. Brewer, H. Vanderswag, A.
- 16 Fleisher, J. L. Heidebrink, J. L. Lord, R. Petersen, S. S. Mason, C. S. Albers, D.
- 17 Knopman, K. Johnson, R. S. Doody, J. V. Meyer, M. Chowdhury, S. Rountree, M. Dang,
- 18 Y. Stern, L. S. Honig, K. L. Bell, B. Ances, J. C. Morris, M. Carroll, S. Leon, E.
- 19 Householder, M. A. Mintun, S. Schneider, A. OliverNG, R. Griffith, D. Clark, D.
- 20 Geldmacher, J. Brockington, E. Roberson, H. Grossman, E. Mitsis, L. de Toledo-Morrell,
- 21 R. C. Shah, R. Duara, D. Varon, M. T. Greig, P. Roberts, M. Albert, C. Onyike, D.
- 22 D’Agostino, S. Kielb, J. E. Galvin, D. M. Pogorelec, B. Cerbone, C. A. Michel, H.
- 23 Rusinek, M. J. de Leon, L. Glodzik, S. De Santi, P. M. Doraiswamy, J. R. Petrella, T. Z.
- 24 Wong, S. E. Arnold, J. H. Karlawish, D. Wolk, C. D. Smith, G. Jicha, P. Hardy, P. Sinha,
- 25 E. Oates, G. Conrad, O. L. Lopez, M. Oakley, D. M. Simpson, A. P. Porsteinsson, B. S.
- 26 Goldstein, K. Martin, K. M. Makino, M. S. Ismail, C. Brand, R. A. Mulnard, G. Thai, C.
- 27 Mc Adams Ortiz, K. Womack, D. Mathews, M. Quiceno, R. D. Arrastia, R. King, M.
- 28 Weiner, K. M. Cook, M. DeVous, A. I. Levey, J. J. Lah, J. S. Cellar, J. M. Burns, H. S.
- 29 Anderson, R. H. Swerdlow, L. Apostolova, K. Tingus, E. Woo, D. H. S. Silverman, P. H.
- 30 Lu, G. Bartzokis, N. R. G. Radford, F. Parfitt, T. Kendall, H. Johnson, M. R. Farlow, A.
- 31 M. Hake, B. R. Matthews, S. Herring, C. Hunt, C. H. van Dyck, R. E. Carson, M. G.
- 32 MacAvoy, H. Chertkow, H. Bergman, C. Hosein, S. Black, B. Stefanovic, C. Caldwell, G.
- 33 Y. R. Hsiung, H. Feldman, B. Mudge, M. A. Past, A. Kertesz, J. Rogers, D. Trost, C.
- 34 Bernick, D. Munic, D. Kerwin, M. M. Mesulam, K. Lipowski, C. K. Wu, N. Johnson, C.
- 35 Sadowsky, W. Martinez, T. Villena, R. S. Turner, K. Johnson, B. Reynolds, R. A.
- 36 Sperling, K. A. Johnson, G. Marshall, M. Frey, J. Yesavage, J. L. Taylor, B. Lane, A.
- 37 Rosen, J. Tinklenberg, M. N. Sabbagh, C. M. Belden, S. A. Jacobson, S. A. Sirrel, N.
- 38 Kowall, R. Killiany, A. E. Budson, A. Norbash, P. L. Johnson, T. O. Obisesan, S.
- 39 Wolday, J. Allard, A. Lerner, P. Ogrocki, L. Hudson, E. Fletcher, O. Carmichael, J.
- 40 Olichney, C. DeCarli, S. Kittur, M. Borrie, T. Y. Lee, R. Bartha, S. Johnson, S. Asthana,
- 41 C. M. Carlsson, S. G. Potkin, A. Preda, D. Nguyen, P. Tariot, A. Fleisher, S. Reeder, V.
- 42 Bates, H. Capote, M. Rainka, D. W. Scharre, M. Kataki, A. Adeli, E. A. Zimmerman, D.
- 43 Celmins, A. D. Brown, G. D. Pearlson, K. Blank, K. Anderson, R. B. Santulli, T. J.
- 44 Kitzmiller, E. S. Schwartz, K. M. SinkS, J. D. Williamson, P. Garg, F. Watkins, B. R. Ott,
- 45 H. Querfurth, G. Tremont, S. Salloway, P. Malloy, S. Correia, H. J. Rosen, B. L. Miller, J.

- 1 Mintzer, K. Spicer, D. Bachman, E. Finger, S. Pasternak, I. Rachinsky, J. Rogers, A.
2 Kertesz, D. Drost, N. Pomara, R. Hernando, A. Sarrael, S. K. Schultz, L. L. Boles Ponto,
3 H. Shim, K. E. Smith, N. Relkin, G. Chaing, L. Raudin, A. Smith, K. Fargher, B. A. Raj,
4 A. D. N. Initiative, the S. B. Study, Spread of pathological tau proteins through
5 communicating neurons in human Alzheimer's disease, *Nat. Commun.* **11**, 2612 (2020).
- 6 22. A. de Calignon, M. Polydoro, M. Suárez-Calvet, C. William, D. H. Adamowicz, K. J.
7 Kopeikina, R. Pitstick, N. Sahara, K. H. Ashe, G. A. Carlson, T. L. Spires-Jones, B. T.
8 Hyman, Propagation of Tau Pathology in a Model of Early Alzheimer's Disease, *Neuron*
9 **73**, 685–697 (2012).
- 10 23. Z. He, J. L. Guo, J. D. McBride, S. Narasimhan, H. Kim, L. Changolkar, B. Zhang, R.
11 J. Gathagan, C. Yue, C. Dengler, A. Stieber, M. Nitla, D. A. Coulter, T. Abel, K. R.
12 Brunden, J. Q. Trojanowski, V. M.-Y. Lee, Amyloid- β plaques enhance Alzheimer's
13 brain tau-seeded pathologies by facilitating neuritic plaque tau aggregation, *Nat. Med.* **24**,
14 29–38 (2018).
- 15 24. R. Ossenkoppele, L. Iaccarino, D. R. Schonhaut, J. A. Brown, R. La Joie, J. P. O'Neil,
16 M. Janabi, S. L. Baker, J. H. Kramer, M.-L. Gorno-Tempini, B. L. Miller, H. J. Rosen, W.
17 W. Seeley, W. J. Jagust, G. D. Rabinovici, Tau covariance patterns in Alzheimer's
18 disease patients match intrinsic connectivity networks in the healthy brain, *NeuroImage*
19 *Clin.* **23**, 101848 (2019).
- 20 25. N. Franzmeier, A. Rubinski, J. Neitzel, Y. Kim, A. Damm, D. L. Na, H. J. Kim, C. H.
21 Lyoo, H. Cho, S. Finsterwalder, M. Duering, S. W. Seo, M. Ewers, Functional
22 connectivity associated with tau levels in ageing, Alzheimer's, and small vessel disease,
23 *Brain* **142**, 1093–1107 (2019).
- 24 26. T. E. Cope, T. Rittman, R. J. Borchert, P. S. Jones, D. Vatansever, K. Allinson, L.
25 Passamonti, P. Vazquez Rodriguez, W. R. Bevan-Jones, J. T. O'Brien, J. B. Rowe, Tau
26 burden and the functional connectome in Alzheimer's disease and progressive
27 supranuclear palsy, *Brain* **141**, 550–567 (2018).
- 28 27. J. M. Tarasoff-Conway, R. O. Carare, R. S. Osorio, L. Glodzik, T. Butler, E.
29 Fieremans, L. Axel, H. Rusinek, C. Nicholson, B. V. Zlokovic, B. Frangione, K.
30 Blennow, J. Ménard, H. Zetterberg, T. Wisniewski, M. J. De Leon, Clearance systems in
31 the brain - Implications for Alzheimer disease *Nat. Rev. Neurol.* (2015),
32 doi:10.1038/nrneurol.2015.119.
- 33 28. J. Kaur, L. M. Fahmy, E. Davoodi-Bojd, L. Zhang, G. Ding, J. Hu, Z. Zhang, M.
34 Chopp, Q. Jiang, Waste Clearance in the Brain, *Front. Neuroanat.* **15** (2021),
35 doi:10.3389/fnana.2021.665803.
- 36 29. L. Xie, H. Kang, Q. Xu, M. J. Chen, Y. Liao, M. Thiyagarajan, J. O'Donnell, D. J.
37 Christensen, C. Nicholson, J. J. Iliff, T. Takano, R. Deane, M. Nedergaard, Sleep drives
38 metabolite clearance from the adult brain, *Science* (80-.). (2013),

- 1 doi:10.1126/science.1241224.
- 2 30. J. J. Iliff, M. Wang, Y. Liao, B. A. Plogg, W. Peng, G. A. Gundersen, H. Benveniste,
3 G. E. Vates, R. Deane, S. A. Goldman, E. A. Nagelhus, M. Nedergaard, A paravascular
4 pathway facilitates CSF flow through the brain parenchyma and the clearance of
5 interstitial solutes, including amyloid β , *Sci. Transl. Med.* (2012),
6 doi:10.1126/scitranslmed.3003748.
- 7 31. J. K. Holth, S. K. Fritschi, C. Wang, N. P. Pedersen, J. R. Cirrito, T. E. Mahan, M. B.
8 Finn, M. Manis, J. C. Geerling, P. M. Fuller, B. P. Lucey, D. M. Holtzman, The sleep-
9 wake cycle regulates brain interstitial fluid tau in mice and CSF tau in humans, *Science*
10 (80-). (2019), doi:10.1126/science.aav2546.
- 11 32. M. L. Schölvinck, A. Maier, F. Q. Ye, J. H. Duyn, D. A. Leopold, M. L. Scholvinck,
12 A. Maier, F. Q. Ye, J. H. Duyn, D. A. Leopold, M. L. Schölvinck, A. Maier, F. Q. Ye, J.
13 H. Duyn, D. A. Leopold, Neural basis of global resting-state fMRI activity, *Proc. Natl.*
14 *Acad. Sci. U. S. A.* **107**, 10238–10243 (2010).
- 15 33. M. Fukunaga, S. G. Horovitz, P. van Gelderen, J. A. de Zwart, J. M. Jansma, V. N.
16 Ikonomidou, R. Chu, R. H. R. Deckers, D. A. Leopold, J. H. Duyn, Large-amplitude,
17 spatially correlated fluctuations in BOLD fMRI signals during extended rest and early
18 sleep stages, *Magn. Reson. Imaging* **24**, 979–992 (2006).
- 19 34. L. J. Larson-Prior, J. M. Zempel, T. S. Nolan, F. W. Prior, A. Snyder, M. E. Raichle,
20 Cortical network functional connectivity in the descent to sleep, *Proc. Natl. Acad. Sci. U.*
21 *S. A.* (2009), doi:10.1073/pnas.0900924106.
- 22 35. C. W. Wong, V. Olafsson, O. Tal, T. T. Liu, The amplitude of the resting-state fMRI
23 global signal is related to EEG vigilance measures, *Neuroimage* **83**, 983–990 (2013).
- 24 36. M. P. McAvoy, E. Tagliazucchi, H. Laufs, M. E. Raichle, Human non-REM sleep
25 and the mean global BOLD signal, *J. Cereb. Blood Flow Metab.* (2019),
26 doi:10.1177/0271678X18791070.
- 27 37. N. E. Fultz, G. Bonmassar, K. Setsompop, R. A. Stickgold, B. R. Rosen, J. R.
28 Polimeni, L. D. Lewis, Coupled electrophysiological, hemodynamic, and cerebrospinal
29 fluid oscillations in human sleep, *Science* (80-). (2019), doi:10.1126/science.aax5440.
- 30 38. F. Han, J. Chen, A. Belkin-Rosen, Y. Gu, L. Luo, O. M. Buxton, X. Liu, Reduced
31 coupling between cerebrospinal fluid flow and global brain activity is linked to
32 Alzheimer disease–related pathology, *PLoS Biol.* **19**, 1–25 (2021).
- 33 39. F. Han, X. Liu, R. B. Mailman, X. Huang, X. Liu, Early β -amyloid accumulation and
34 hypoconnectivity in the default mode network are related to its disengagement from
35 global brain activity, *bioRxiv* (2022), doi:10.1101/2022.07.24.501309.

- 1 40. F. Han, G. L. Brown, Y. Zhu, A. E. Belkin-Rosen, M. M. Lewis, G. Du, Y. Gu, P. J.
2 Eslinger, R. B. Mailman, X. Huang, X. Liu, Decoupling of Global Brain Activity and
3 Cerebrospinal Fluid Flow in Parkinson's Disease Cognitive Decline, *Mov. Disord.* **36**,
4 2066–2076 (2021).
- 5 41. F. Han, X. Liu, Y. Yang, X. Liu, Sex-specific age-related changes in glymphatic
6 function assessed by resting-state functional magnetic resonance imaging, *bioRxiv* (2023),
7 doi:10.1101/2023.04.02.535258.
- 8 42. W. Jagust, Imaging the evolution and pathophysiology of Alzheimer disease *Nat. Rev.*
9 *Neurosci.* (2018), doi:10.1038/s41583-018-0067-3.
- 10 43. M. Nedergaard, S. A. Goldman, Glymphatic failure as a final common pathway to
11 dementia, *Science* (80-.). **370**, 50–56 (2020).
- 12 44. X. Liu, T. Yanagawa, D. A. Leopold, C. Chang, H. Ishida, N. Fujii, J. H. Duyn,
13 Arousal transitions in sleep, wakefulness, and anesthesia are characterized by an orderly
14 sequence of cortical events, *Neuroimage* (2015), doi:10.1016/j.neuroimage.2015.04.003.
- 15 45. X. Liu, J. A. De Zwart, M. L. Schölvinck, C. Chang, F. Q. Ye, D. A. Leopold, J. H.
16 Duyn, Subcortical evidence for a contribution of arousal to fMRI studies of brain activity,
17 *Nat. Commun.* (2018), doi:10.1038/s41467-017-02815-3.
- 18 46. J. Li, T. Bolt, D. Bzdok, J. S. Nomi, B. T. T. Yeo, R. N. Spreng, L. Q. Uddin,
19 Topography and behavioral relevance of the global signal in the human brain, *Sci. Rep.*
20 (2019), doi:10.1038/s41598-019-50750-8.
- 21 47. Y. Gu, L. E. Sainburg, S. Kuang, F. Han, J. W. Williams, Y. Liu, N. Zhang, X. Zhang,
22 D. A. Leopold, X. Liu, Brain Activity Fluctuations Propagate as Waves Traversing the
23 Cortical Hierarchy, *Cereb. Cortex* **31**, 3986–4005 (2021).
- 24 48. R. V. Raut, A. Z. Snyder, A. Mitra, D. Yellin, N. Fujii, R. Malach, M. E. Raichle,
25 Global waves synchronize the brain's functional systems with fluctuating arousal, *Sci.*
26 *Adv.* **7**, 1–16 (2021).
- 27 49. M. W. Weiner, D. P. Veitch, P. S. Aisen, L. A. Beckett, N. J. Cairns, R. C. Green, D.
28 Harvey, C. R. Jack, W. Jagust, E. Liu, J. C. Morris, R. C. Petersen, A. J. Saykin, M. E.
29 Schmidt, L. Shaw, L. Shen, J. A. Siuciak, H. Soares, A. W. Toga, J. Q. Trojanowski, The
30 Alzheimer's Disease Neuroimaging Initiative: A review of papers published since its
31 inception *Alzheimer's Dement.* (2013), doi:10.1016/j.jalz.2013.05.1769.
- 32 50. M. W. Weiner, D. P. Veitch, P. S. Aisen, L. A. Beckett, N. J. Cairns, R. C. Green, D.
33 Harvey, C. R. Jack Jr., W. Jagust, J. C. Morris, R. C. Petersen, J. Salazar, A. J. Saykin, L.
34 M. Shaw, A. W. Toga, J. Q. Trojanowski, A. D. N. Initiative, The Alzheimer's Disease
35 Neuroimaging Initiative 3: Continued innovation for clinical trial improvement,
36 *Alzheimer's Dement.* **13**, 561–571 (2017).

- 1 51. S. M. Landau, M. A. Mintun, A. D. Joshi, R. A. Koeppe, R. C. Petersen, P. S. Aisen,
2 M. W. Weiner, W. J. Jagust, Amyloid deposition, hypometabolism, and longitudinal
3 cognitive decline, *Ann. Neurol.* (2012), doi:10.1002/ana.23650.
- 4 52. S. K. Royse, D. S. Minhas, B. J. Lopresti, A. Murphy, T. Ward, R. A. Koeppe, S.
5 Bullich, S. DeSanti, W. J. Jagust, S. M. Landau, for the A. D. N. Initiative, Validation of
6 amyloid PET positivity thresholds in centiloids: a multisite PET study approach,
7 *Alzheimers. Res. Ther.* **13**, 99 (2021).
- 8 53. C. R. Jack, H. J. Wiste, S. D. Weigand, T. M. Therneau, V. J. Lowe, D. S. Knopman,
9 J. L. Gunter, M. L. Senjem, D. T. Jones, K. Kantarci, M. M. Machulda, M. M. Mielke, R.
10 O. Roberts, P. Vemuri, D. A. Reyes, R. C. Petersen, Defining imaging biomarker cut
11 points for brain aging and Alzheimer’s disease, *Alzheimer’s Dement.* **13**, 205–216 (2017).
- 12 54. C. Boccalini, F. Ribaldi, I. Hristovska, A. Arnone, D. E. Peretti, L. Mu, M. Scheffler,
13 D. Perani, G. B. Frisoni, V. Garibotto, The impact of tau deposition and hypometabolism
14 on cognitive impairment and longitudinal cognitive decline, *Alzheimer’s & Dement.* **n/a**,
15 doi:<https://doi.org/10.1002/alz.13355>.
- 16 55. J. Sepulcre, M. J. Grothe, M. Sabuncu, J. Chhatwal, A. P. Schultz, B. Hanseeuw, G.
17 El Fakhri, R. Sperling, K. A. Johnson, Hierarchical Organization of Tau and Amyloid
18 Deposits in the Cerebral Cortex, *JAMA Neurol.* **74**, 813–820 (2017).
- 19 56. J. B. Pereira, R. Ossenkoppele, S. Palmqvist, T. O. Strandberg, R. Smith, E. Westman,
20 O. Hansson, M. Irish, T. E. Behrens, Eds. Amyloid and tau accumulate across distinct
21 spatial networks and are differentially associated with brain connectivity, *Elife* **8**, e50830
22 (2019).
- 23 57. D. S. Margulies, S. S. Ghosh, A. Goulas, M. Falkiewicz, J. M. Huntenburg, G. Langs,
24 G. Bezgin, S. B. Eickhoff, F. X. Castellanos, M. Petrides, E. Jefferies, J. Smallwood,
25 Situating the default-mode network along a principal gradient of macroscale cortical
26 organization, *Proc. Natl. Acad. Sci.* **113**, 12574–12579 (2016).
- 27 58. R. S. Desikan, F. Ségonne, B. Fischl, B. T. Quinn, B. C. Dickerson, D. Blacker, R. L.
28 Buckner, A. M. Dale, R. P. Maguire, B. T. Hyman, M. S. Albert, R. J. Killiany, An
29 automated labeling system for subdividing the human cerebral cortex on MRI scans into
30 gyral based regions of interest, *Neuroimage* (2006),
31 doi:10.1016/j.neuroimage.2006.01.021.
- 32 59. R. A. Sperling, P. S. LaViolette, K. O’Keefe, J. O’Brien, D. M. Rentz, M.
33 Pihlajamaki, G. Marshall, B. T. Hyman, D. J. Selkoe, T. Hedden, R. L. Buckner, J. A.
34 Becker, K. A. Johnson, Amyloid Deposition Is Associated with Impaired Default
35 Network Function in Older Persons without Dementia, *Neuron* **63**, 178–188 (2009).
- 36 60. T. Hedden, K. R. A. Van Dijk, J. A. Becker, A. Mehta, R. A. Sperling, K. A. Johnson,
37 R. L. Buckner, Disruption of functional connectivity in clinically normal older adults

- 1 harboring amyloid burden, *J. Neurosci.* **29**, 12686–12694 (2009).
- 2 61. Y. I. Sheline, M. E. Raichle, A. Z. Snyder, J. C. Morris, D. Head, S. Wang, M. A.
3 Mintun, Amyloid Plaques Disrupt Resting State Default Mode Network Connectivity in
4 Cognitively Normal Elderly, *Biol. Psychiatry* **67**, 584–587 (2010).
- 5 62. M. R. Brier, J. B. Thomas, A. M. Fagan, J. Hassenstab, D. M. Holtzman, T. L.
6 Benzinger, J. C. Morris, B. M. Ances, Functional connectivity and graph theory in
7 preclinical Alzheimer’s disease, *Neurobiol. Aging* **35**, 757–768 (2014).
- 8 63. T. T. Liu, A. Nalci, M. Falahpour, The global signal in fMRI: Nuisance or
9 Information?, *Neuroimage* **150**, 213–229 (2017).
- 10 64. P. S. Özbay, C. Chang, D. Picchioni, H. Mandelkow, M. G. Chappel-Farley, P. van
11 Gelderen, J. A. de Zwart, J. Duyn, Sympathetic activity contributes to the fMRI signal,
12 *Commun. Biol.* (2019), doi:10.1038/s42003-019-0659-0.
- 13 65. C. Chang, J. P. Cunningham, G. H. Glover, Influence of heart rate on the BOLD
14 signal: The cardiac response function, *Neuroimage* (2009),
15 doi:10.1016/j.neuroimage.2008.09.029.
- 16 66. P. S. Özbay, C. Chang, D. Picchioni, H. Mandelkow, T. M. Moehlman, M. G.
17 Chappel-Farley, P. van Gelderen, J. A. de Zwart, J. H. Duyn, Contribution of systemic
18 vascular effects to fMRI activity in white matter, *Neuroimage* (2018),
19 doi:10.1016/j.neuroimage.2018.04.045.
- 20 67. R. M. Birn, J. B. Diamond, M. A. Smith, P. A. Bandettini, Separating respiratory-
21 variation-related fluctuations from neuronal-activity-related fluctuations in fMRI,
22 *Neuroimage* (2006), doi:10.1016/j.neuroimage.2006.02.048.
- 23 68. Y. Gu, F. Han, L. E. Sainburg, X. Liu, Transient Arousal Modulations Contribute to
24 Resting-State Functional Connectivity Changes Associated with Head Motion Parameters,
25 *Cereb. Cortex* (2020), doi:10.1093/cercor/bhaa096.
- 26 69. C. Chang, C. D. Metzger, G. H. Glover, J. H. Duyn, H. J. Heinze, M. Walter,
27 Association between heart rate variability and fluctuations in resting-state functional
28 connectivity, *Neuroimage* **68**, 93–104 (2013).
- 29 70. X. Liu, D. A. Leopold, Y. Yang, Single-neuron firing cascades underlie global
30 spontaneous brain events, *Proc. Natl. Acad. Sci. U. S. A.* **118**, 1–10 (2021).
- 31 71. P. Pais-Roldán, K. Takahashi, F. Sobczak, Y. Chen, X. Zhao, H. Zeng, Y. Jiang, X.
32 Yu, Indexing brain state-dependent pupil dynamics with simultaneous fMRI and optical
33 fiber calcium recording, *Proc. Natl. Acad. Sci. U. S. A.* **117**, 6875–6882 (2020).
- 34 72. J. M. Shine, P. G. Bissett, P. T. Bell, O. Koyejo, J. H. Balsters, K. J. Gorgolewski, C.

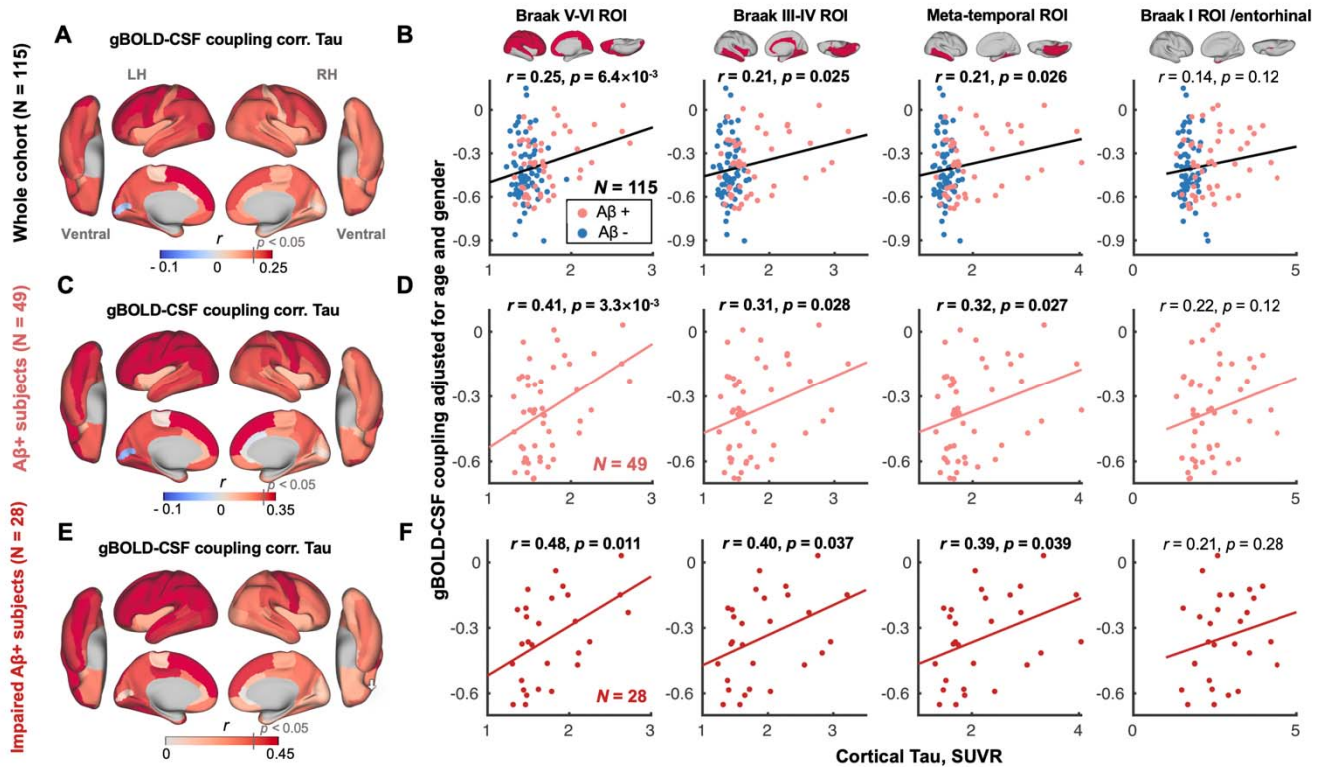
- 1 A. Moodie, R. A. Poldrack, The Dynamics of Functional Brain Networks: Integrated
2 Network States during Cognitive Task Performance, *Neuron* **92**, 544–554 (2016).
- 3 73. E. Hamel, Perivascular nerves and the regulation of cerebrovascular tone, *J. Appl.*
4 *Physiol.* **100**, 1059–1064 (2006).
- 5 74. J. J. Iliff, M. Wang, D. M. Zeppenfeld, A. Venkataraman, B. A. Plog, Y. Liao, R.
6 Deane, M. Nedergaard, Cerebral arterial pulsation drives paravascular CSF-Interstitial
7 fluid exchange in the murine brain, *J. Neurosci.* (2013), doi:10.1523/JNEUROSCI.1592-
8 13.2013.
- 9 75. S. Yamada, M. Miyazaki, Y. Yamashita, C. Ouyang, M. Yui, M. Nakahashi, S.
10 Shimizu, I. Aoki, Y. Morohoshi, J. G. McComb, Influence of respiration on cerebrospinal
11 fluid movement using magnetic resonance spin labeling, *Fluids Barriers CNS* (2013),
12 doi:10.1186/2045-8118-10-36.
- 13 76. D. Picchioni, P. S. Özbay, H. Mandelkow, J. A. de Zwart, Y. Wang, P. van Gelderen,
14 J. H. Duyn, Autonomic arousals contribute to brain fluid pulsations during sleep,
15 *Neuroimage* **249**, 118888 (2022).
- 16 77. E. Hamel, Cholinergic modulation of the cortical microvascular bed, *Prog. Brain Res.*
17 **145**, 171–178 (2004).
- 18 78. N. A. Jessen, A. S. F. Munk, I. Lundgaard, M. Nedergaard, The Glymphatic System:
19 A Beginner’s Guide, *Neurochem. Res.* (2015), doi:10.1007/s11064-015-1581-6.
- 20 79. M. Wang, Y. He, T. J. Sejnowski, X. Yu, Brain-state dependent astrocytic Ca²⁺
21 signals are coupled to both positive and negative BOLD-fMRI signals, *Proc. Natl. Acad.*
22 *Sci. U. S. A.* **115**, E1647–E1656 (2018).
- 23 80. J. J. Iliff, M. J. Chen, B. A. Plog, D. M. Zeppenfeld, M. Soltero, L. Yang, I. Singh, R.
24 Deane, M. Nedergaard, Impairment of glymphatic pathway function promotes tau
25 pathology after traumatic brain injury, *J. Neurosci.* (2014),
26 doi:10.1523/JNEUROSCI.3020-14.2014.
- 27 81. I. F. Harrison, O. Ismail, A. Machhada, N. Colgan, Y. Ohene, P. Nahavandi, Z.
28 Ahmed, A. Fisher, S. Meftah, T. K. Murray, O. P. Ottersen, E. A. Nagelhus, M. J.
29 O’Neill, J. A. Wells, M. F. Lythgoe, Impaired glymphatic function and clearance of tau
30 in an Alzheimer’s disease model, *Brain* (2020), doi:10.1093/brain/awaa179.
- 31 82. K. Ishida, K. Yamada, R. Nishiyama, T. Hashimoto, I. Nishida, Y. Abe, M. Yasui, T.
32 Iwatsubo, Glymphatic system clears extracellular tau and protects from tau aggregation
33 and neurodegeneration, *J. Exp. Med.* **219** (2022), doi:10.1084/jem.20211275.
- 34 83. D. M. Lopes, S. K. Llewellyn, I. F. Harrison, Propagation of tau and α -synuclein in
35 the brain: therapeutic potential of the glymphatic system, *Transl. Neurodegener.* **11**, 19

- 1 (2022).
- 2 84. S. Calafate, A. Buist, K. Miskiewicz, V. Vijayan, G. Daneels, B. de Strooper, J.
3 de Wit, P. Verstreken, D. Moechars, Synaptic Contacts Enhance Cell-to-Cell Tau
4 Pathology Propagation, *Cell Rep.* **11**, 1176–1183 (2015).
- 5 85. A. Gómez-Ramos, M. Díaz-Hernández, R. Cuadros, F. Hernández, J. Avila,
6 Extracellular tau is toxic to neuronal cells, *FEBS Lett.* **580**, 4842–4850 (2006).
- 7 86. M. Jucker, L. C. Walker, Self-propagation of pathogenic protein aggregates in
8 neurodegenerative diseases, *Nature* **501**, 45–51 (2013).
- 9 87. M. Jucker, L. C. Walker, Propagation and spread of pathogenic protein assemblies in
10 neurodegenerative diseases, *Nat. Neurosci.* **21**, 1341–1349 (2018).
- 11 88. J.-L. Hsu, Y.-C. Wei, C. H. Toh, I.-T. Hsiao, K.-J. Lin, T.-C. Yen, M.-F. Liao, L.-S.
12 Ro, Magnetic Resonance Images Implicate That Glymphatic Alterations Mediate
13 Cognitive Dysfunction in Alzheimer Disease, *Ann. Neurol.* **93**, 164–174 (2023).
- 14 89. A. Zamani, A. K. Walker, B. Rollo, K. L. Ayers, R. Farah, T. J. O’Brien, D. K.
15 Wright, Impaired glymphatic function in the early stages of disease in a TDP-43 mouse
16 model of amyotrophic lateral sclerosis, *Transl. Neurodegener.* **11**, 17 (2022).
- 17 90. C. Zhao, W.-J. Huang, F. Feng, B. Zhou, H.-X. Yao, Y.-E. Guo, P. Wang, L.-N.
18 Wang, N. Shu, X. Zhang, Abnormal characterization of dynamic functional connectivity
19 in Alzheimer’s disease, *Neural Regen. Res.* **17** (2022) (available at
20 [https://journals.lww.com/nrronline/Fulltext/2022/09000/Abnormal_characterization_of_d](https://journals.lww.com/nrronline/Fulltext/2022/09000/Abnormal_characterization_of_dynamic_functional.33.aspx)
21 [ynamic_functional.33.aspx](https://journals.lww.com/nrronline/Fulltext/2022/09000/Abnormal_characterization_of_dynamic_functional.33.aspx)).
- 22 91. X. Ma, Z. Zhuo, L. Wei, Z. Ma, Z. Li, H. Li, Altered Temporal Organization of Brief
23 Spontaneous Brain Activities in Patients with Alzheimer’s Disease, *Neuroscience* **425**, 1–
24 11 (2020).
- 25 92. A. Schäfer, P. Chaggar, T. B. Thompson, A. Goriely, E. Kuhl, Predicting brain
26 atrophy from tau pathology: a summary of clinical findings and their translation into
27 personalized models, *Brain Multiphysics* **2**, 100039 (2021).
- 28 93. A.-T. Du, N. Schuff, J. H. Kramer, H. J. Rosen, M. L. Gorno-Tempini, K. Rankin, B.
29 L. Miller, M. W. Weiner, Different regional patterns of cortical thinning in Alzheimer’s
30 disease and frontotemporal dementia, *Brain* **130**, 1159–1166 (2007).
- 31 94. G. Girardeau, M. Zugaro, Hippocampal ripples and memory consolidation, *Curr.*
32 *Opin. Neurobiol.* **21**, 452–459 (2011).
- 33 95. H. R. Joo, L. M. Frank, The hippocampal sharp wave–ripple in memory retrieval for
34 immediate use and consolidation, *Nat. Rev. Neurosci.* **19**, 744–757 (2018).

- 1 96. T. M. Harrison, A. Maass, J. N. Adams, R. Du, S. L. Baker, W. J. Jagust, Tau
2 deposition is associated with functional isolation of the hippocampus in aging, *Nat.*
3 *Commun.* **10**, 4900 (2019).
- 4 97. D. P. Veitch, M. W. Weiner, P. S. Aisen, L. A. Beckett, C. DeCarli, R. C. Green, D.
5 Harvey, C. R. Jack Jr., W. Jagust, S. M. Landau, J. C. Morris, O. Okonkwo, R. J. Perrin,
6 R. C. Petersen, M. Rivera-Mindt, A. J. Saykin, L. M. Shaw, A. W. Toga, D. Tosun, J. Q.
7 Trojanowski, A. D. N. Initiative, Using the Alzheimer’s Disease Neuroimaging Initiative
8 to improve early detection, diagnosis, and treatment of Alzheimer’s disease, *Alzheimer’s*
9 *Dement.* **18**, 824–857 (2022).
- 10 98. A. Maass, S. Landau, S. L. Baker, A. Horng, S. N. Lockhart, R. La Joie, G. D.
11 Rabinovici, W. J. Jagust, Comparison of multiple tau-PET measures as biomarkers in
12 aging and Alzheimer’s disease, *Neuroimage* **157**, 448–463 (2017).
- 13 99. S. L. Baker, S. N. Lockhart, J. C. Price, M. He, R. H. Huesman, D. Schonhaut, J.
14 Faria, G. Rabinovici, W. J. Jagust, Reference Tissue-Based Kinetic Evaluation of 18F-
15 AV-1451 for Tau Imaging, *J. Nucl. Med.* **58**, 332–338 (2017).
- 16 100. S. L. Baker, A. Maass, W. J. Jagust, Considerations and code for partial volume
17 correcting [18F]-AV-1451 tau PET data, *Data Br.* **15**, 648–657 (2017).
- 18 101. S. M. Landau, M. Lu, A. D. Joshi, M. Pontecorvo, M. A. Mintun, J. Q. Trojanowski,
19 L. M. Shaw, W. J. Jagust, Comparing positron emission tomography imaging and
20 cerebrospinal fluid measurements of β -amyloid, *Ann. Neurol.* (2013),
21 doi:10.1002/ana.23908.
- 22 102. C. R. Jack, M. A. Bernstein, N. C. Fox, P. Thompson, G. Alexander, D. Harvey, B.
23 Borowski, P. J. Britson, J. L. Whitwell, C. Ward, A. M. Dale, J. P. Felmlee, J. L. Gunter,
24 D. L. G. Hill, R. Killiany, N. Schuff, S. Fox-Bosetti, C. Lin, C. Studholme, C. S. DeCarli,
25 G. Krueger, H. A. Ward, G. J. Metzger, K. T. Scott, R. Mallozzi, D. Blezek, J. Levy, J. P.
26 Debbins, A. S. Fleisher, M. Albert, R. Green, G. Bartzokis, G. Glover, J. Mugler, M. W.
27 Weiner, The Alzheimer’s Disease Neuroimaging Initiative (ADNI): MRI methods, *J.*
28 *Magn. Reson. Imaging* **27**, 685–691 (2008).
- 29 103. J. J. Gomar, G. Tan, J. Halpern, M. L. Gordon, B. Greenwald, J. Koppel, Increased
30 retention of tau PET ligand [18F]-AV1451 in Alzheimer’s Disease Psychosis, *Transl.*
31 *Psychiatry* **12**, 82 (2022).
- 32 104. J. Giorgio, W. J. Jagust, S. Baker, S. M. Landau, P. Tino, Z. Kourtzi, A. D. N.
33 Initiative, A robust and interpretable machine learning approach using multimodal
34 biological data to predict future pathological tau accumulation, *Nat. Commun.* **13**, 1887
35 (2022).
- 36 105. B. Fischl, *FreeSurferNeuroimage* (2012), doi:10.1016/j.neuroimage.2012.01.021.

- 1 106. S. M. Smith, M. Jenkinson, M. W. Woolrich, C. F. Beckmann, T. E. J. Behrens, H.
2 Johansen-Berg, P. R. Bannister, M. De Luca, I. Drobnjak, D. E. Flitney, R. K. Niazy, J.
3 Saunders, J. Vickers, Y. Zhang, N. De Stefano, J. M. Brady, P. M. Matthews, Advances
4 in functional and structural MR image analysis and implementation as FSL, *Neuroimage*
5 **23**, S208–S219 (2004).
- 6 107. R. W. Cox, AFNI: Software for analysis and visualization of functional magnetic
7 resonance neuroimages, *Comput. Biomed. Res.* (1996), doi:10.1006/cbmr.1996.0014.
- 8 108. S. Palmqvist, M. Schöll, O. Strandberg, N. Mattsson, E. Stomrud, H. Zetterberg, K.
9 Blennow, S. Landau, W. Jagust, O. Hansson, Earliest accumulation of β -amyloid occurs
10 within the default-mode network and concurrently affects brain connectivity, *Nat.*
11 *Commun.* (2017), doi:10.1038/s41467-017-01150-x.
- 12 109. P. E. Shrout, N. Bolger, Mediation in experimental and nonexperimental studies:
13 New procedures and recommendations, *Psychol. Methods* **7**, 422–445 (2002).
- 14 110. R. A. Sperling, E. C. Mormino, A. P. Schultz, R. A. Betensky, K. V Papp, R. E.
15 Amariglio, B. J. Hanseeuw, R. Buckley, J. Chhatwal, T. Hedden, G. A. Marshall, Y. T.
16 Quiroz, N. J. Donovan, J. Jackson, J. R. Gatchel, J. S. Rabin, H. Jacobs, H.-S. Yang, M.
17 Properzi, D. R. Kirn, D. M. Rentz, K. A. Johnson, The impact of amyloid-beta and tau on
18 prospective cognitive decline in older individuals, *Ann. Neurol.* **85**, 181–193 (2019).
- 19 111. A. Schaefer, R. Kong, E. M. Gordon, T. O. Laumann, X.-N. Zuo, A. J. Holmes, S. B.
20 Eickhoff, B. T. T. Yeo, Local-Global Parcellation of the Human Cerebral Cortex from
21 Intrinsic Functional Connectivity MRI, *Cereb. Cortex* (2018), doi:10.1093/cercor/bhx179.
- 22 112. R. S. Society, On the Interpretation of χ^2 from Contingency Tables , and the
23 Calculation of P Author (s): R . A . Fisher Source□: Journal of the Royal Statistical
24 Society , Jan ., 1922 , Vol . 85 , No . 1 (Jan ., 1922), Published by□: Wiley for the
25 Royal Statistical, **85**, 87–94 (1922).
- 26 113. J. D. Power, K. A. Barnes, A. Z. Snyder, B. L. Schlaggar, S. E. Petersen, Spurious
27 but systematic correlations in functional connectivity MRI networks arise from subject
28 motion, *Neuroimage* (2012), doi:10.1016/j.neuroimage.2011.10.018.
- 29

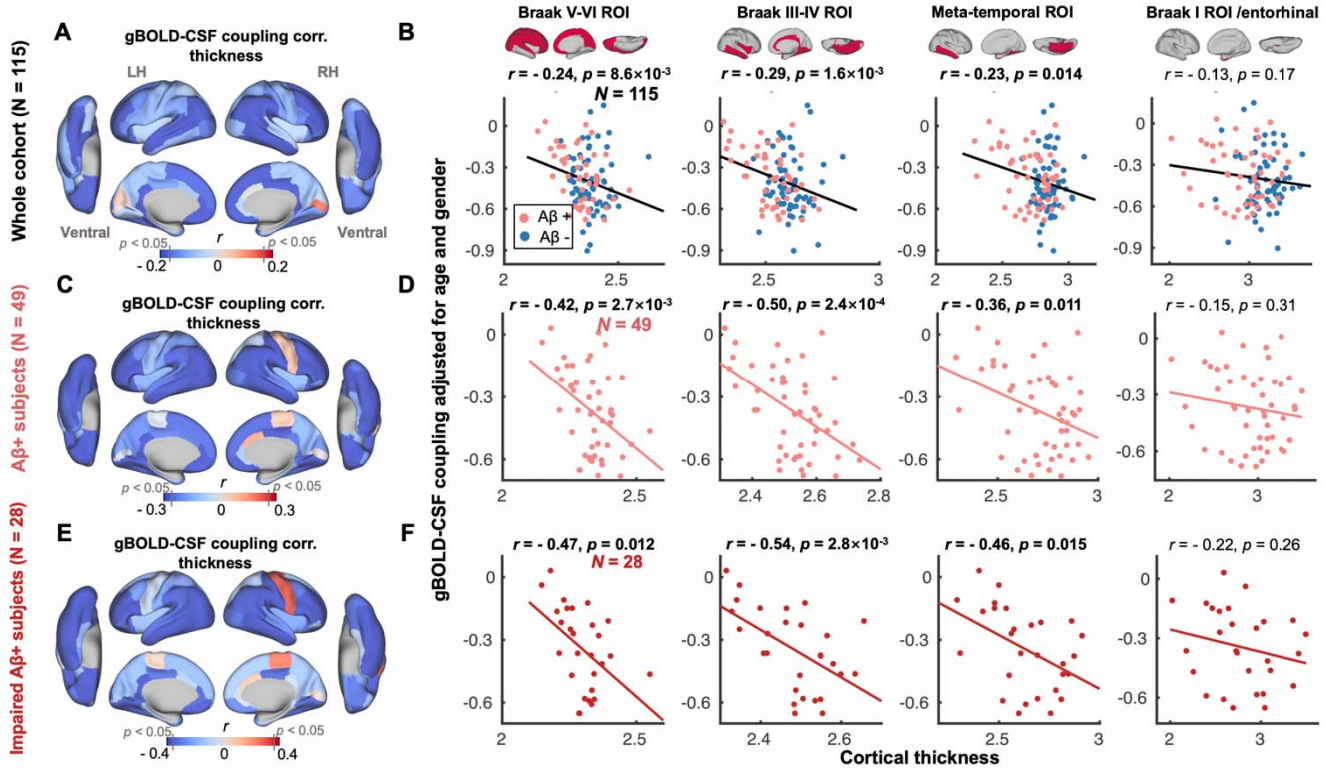
1 **FIGURES**



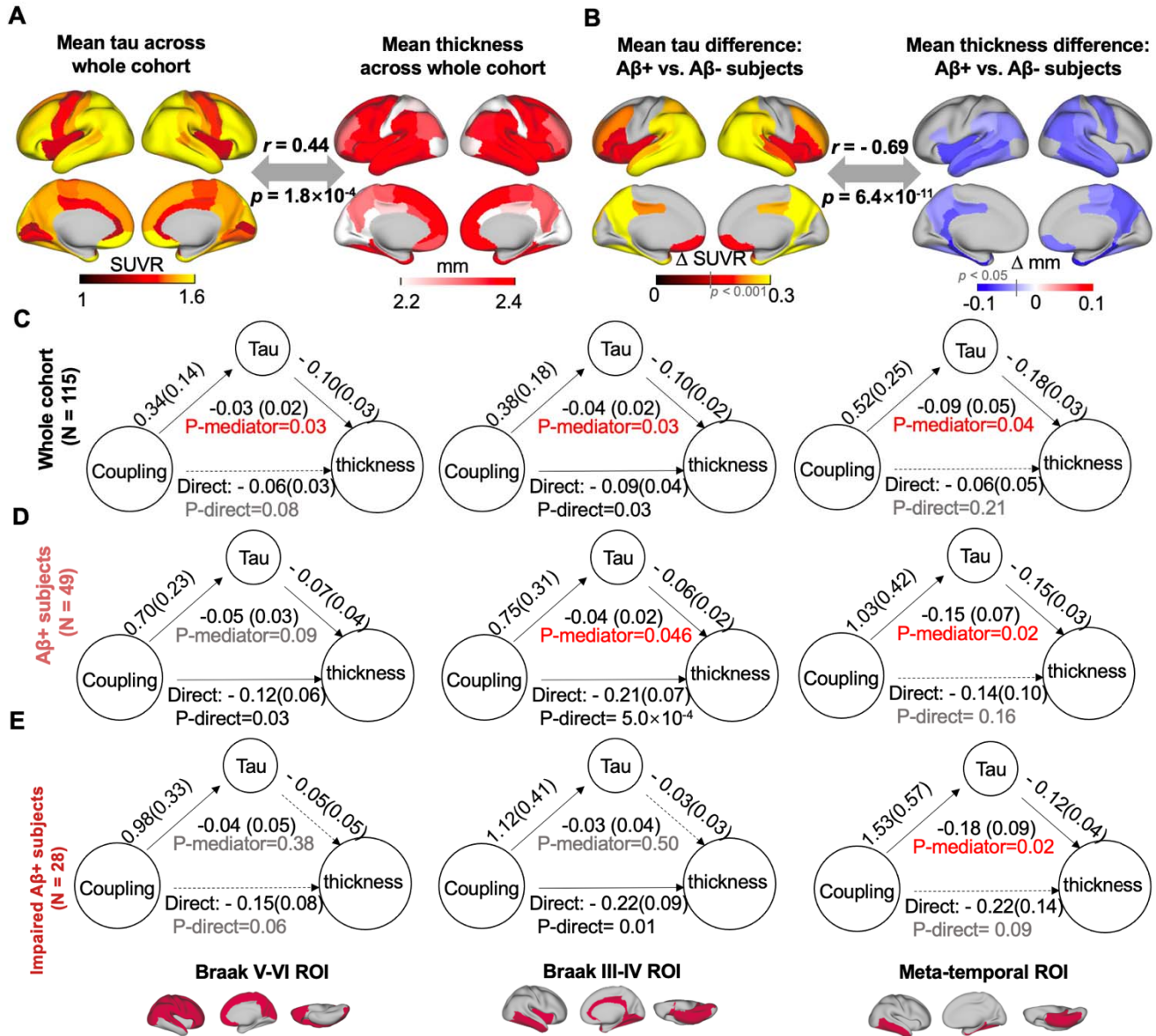
2
3 **Fig. 1. gBOLD-CSF coupling is correlated with the cortical tau across the whole cohort and Aβ+ subjects.**

4 (A) gBOLD-CSF coupling was positively correlated with tau in most cortical regions across all subjects, i.e.,
5 subjects with weaker (less negative) coupling had more tau deposition. (B) gBOLD-CSF coupling (strength)
6 significantly decreased with more tau deposition in Braak V-VI, Braak III-IV, and temporal meta-ROI across
7 the whole cohort (all $r > 0.21$, all $p < 0.026$; $N = 115$). This association was similar, although not significant ($r =$
8 $0.14, p = 0.12$), for tau in Braak I (entorhinal cortex). (C-F) These associations between gBOLD-CSF coupling
9 and regional tau were also evident among Aβ+ and/or specifically impaired Aβ+ subjects (all $r > 0.31$, all $p <$
10 0.039 ; in Braak V-VI, Braak III-IV, and temporal meta-ROI), but not for the rest of subjects (**Fig. S2**). Aβ+:
11 cortical AV45-Aβ > 1.11 SUVR or cortical FBB-Aβ > 1.08 SUVR. Impaired group: AD and MCI subjects.
12 Each point represents one subject. The linear regression line was estimated based on the linear least-squares
13 fitting (the same hereinafter unless noted otherwise).

14
15



1
2 **Fig. 2. gBOLD-CSF coupling is correlated with cortical thickness across the whole cohort and A β +**
3 **subjects. (A)** Subjects with weaker (less negative) gBOLD-CSF coupling had thinner cortices in the majority of
4 brain regions, especially in the frontal, parietal, and temporal lobes, including default mode network (DMN) and
5 fronto-parietal network (FPN). **(B)** The gBOLD-CSF coupling strength significantly decreased with thinner
6 cortices in Braak V-VI, Braak III-IV, and temporal meta-ROIs across the whole cohort (all $r < -0.23$, all $p <$
7 0.014 ; $N = 115$). A similar but not significant coupling-thickness association was found in the entorhinal region
8 ($r = -0.13$, $p = 0.17$). **(C-F)** Among A β + subjects, particularly impaired A β + ones, the coupling-thickness
9 remained striking (all $r < -0.36$, all $p < 0.015$ in Braak V-VI, Braak III-IV, and temporal meta-ROI in **D** and **F**)
10 while this was not the case in other participants (**Fig. S4**). Each point represents one subject.

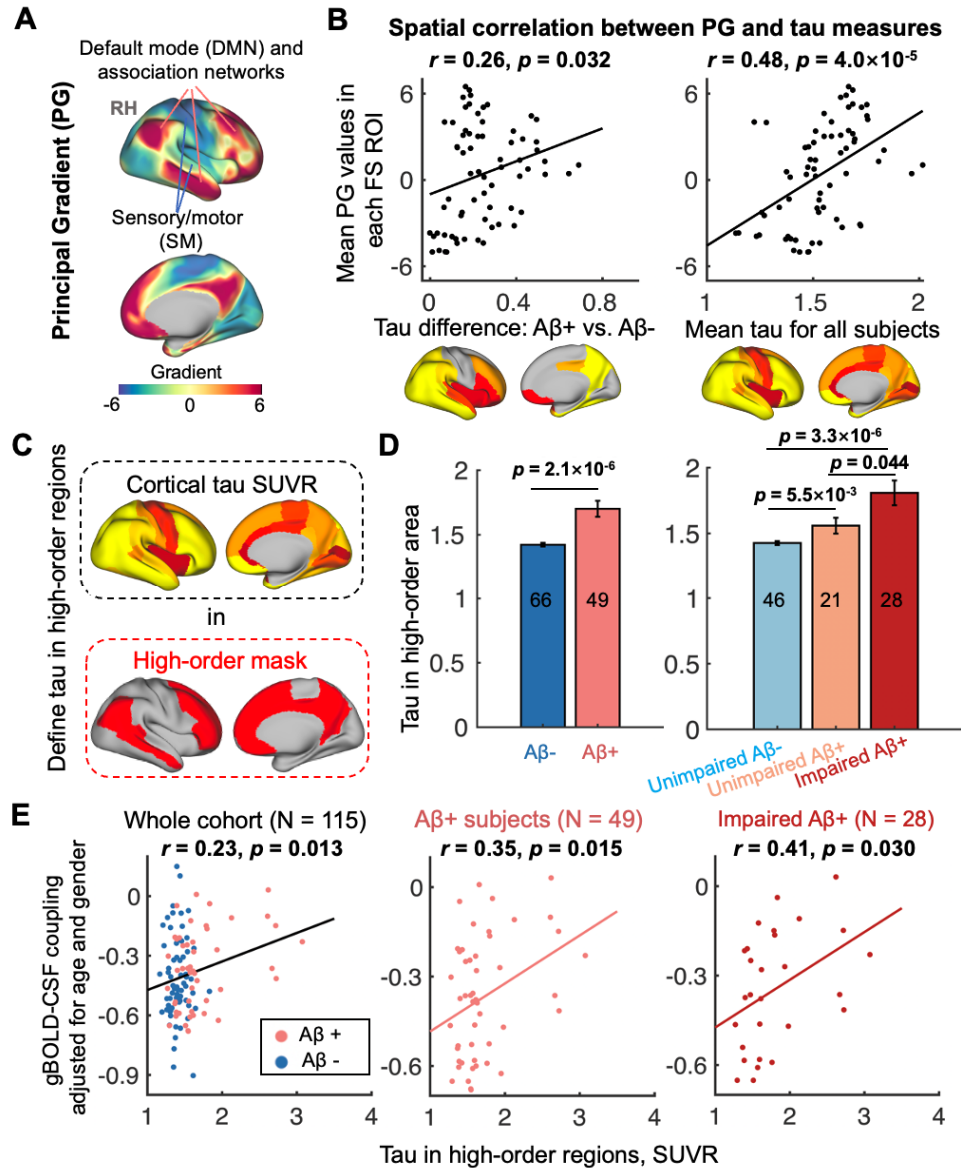


1
2 **Fig. 3. The association between coupling metrics and thickness is mediated by tau.** (A-B) The averaged
3 map of cortical tau and thickness across all subjects ($r = 0.44$, $p = 1.8 \times 10^{-4}$), as well as the difference between
4 A β - and A β + subjects ($r = -0.69$, $p = 6.4 \times 10^{-11}$), indicative of tau and atrophy distributions were similar. This
5 close association between tau pathology and atrophy is also consistent with their significant correlation across
6 subjects (Fig. S5). (C-E) Across the entire cohort and A β + subjects, tau mediated the significant association in
7 Fig. 2 between gBOLD-CSF coupling and thickness throughout cortex, including Braak V-VI, Braak III-IV, and
8 meta-temporal area ($p < 0.046$, except for the marginally significant relationship [$p = 0.090$] in Braak V-VI

Feng Han et al.

1 among the A β + subjects), whereas the meditation effect was weaker for impaired A β + ones, which might be
2 attributed to the limited sample size.

3



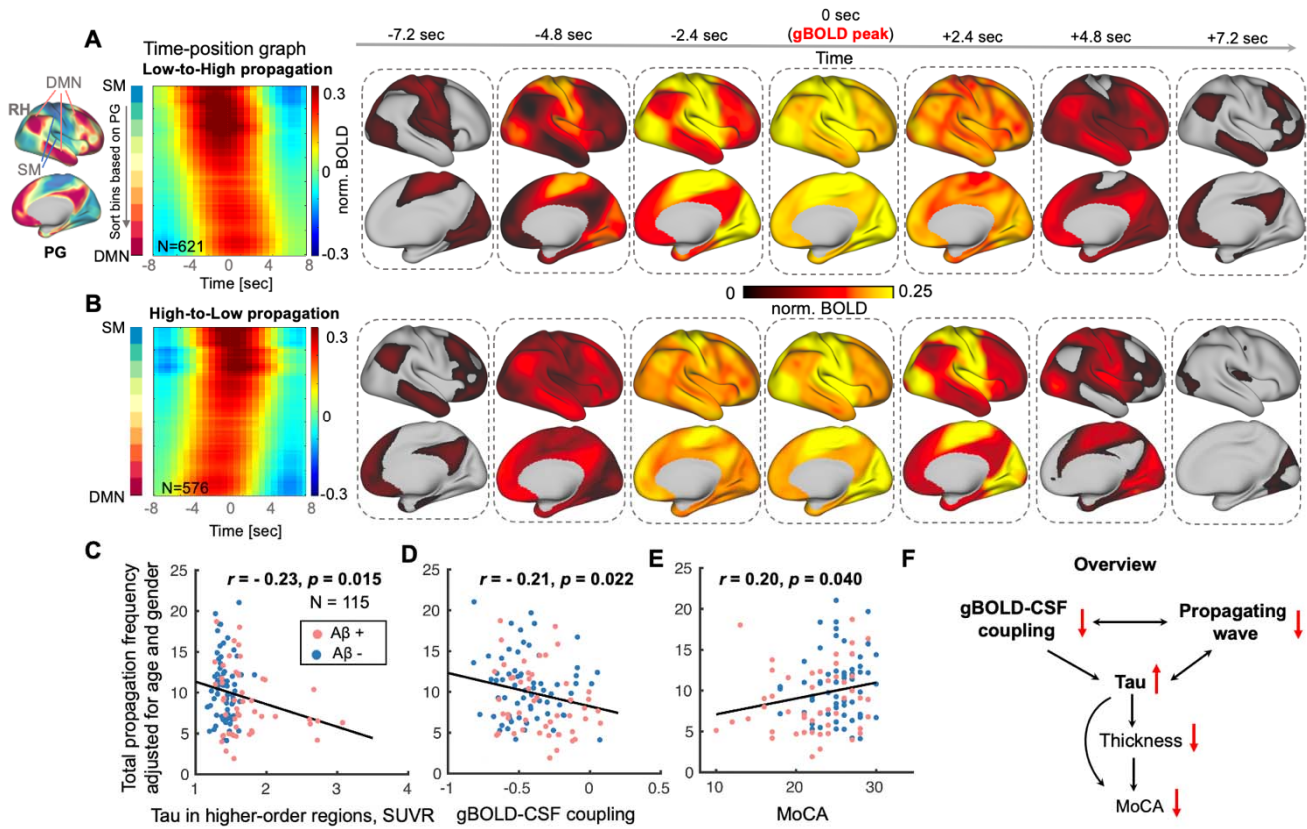
1
2

3 **Fig. 4. Preferential tau deposition in higher-order regions is related to coupling decrease.** (A) A principal
 4 gradient (PG) of functional connectivity identified by a previous study (57) that classified cortical hierarchies by
 5 examining similarity of connectivity was employed to quantify regional cortical hierarchy. (B) Cortical regions
 6 (DKT-parcels) with higher tau burden, comparing A β - and A β + individuals, were correlated with higher cortical
 7 hierarchy ($r = 0.26, p = 0.032$). Similarly, among all subjects, higher order regions had more tau deposition ($r =$
 8 $0.48, p = 4.0 \times 10^{-5}$). (C) Two association networks with the highest PG scores, i.e., DMN and FPN (111), were
 9 combined and served as a higher-order mask (39) and used to extract the tau deposition. (D) Higher-order tau

Feng Han et al.

1 increased from A β - and A β + subjects, or from unimpaired A β - to unimpaired A β + to impaired A β + stages. (**E**)
2 Higher-order tau was significantly correlated with decreased gBOLD-CSF coupling strength in the whole cohort,
3 A β + subjects and impaired A β + subjects, similar to **Fig. 1**. Each error bar represents the standard error of the
4 mean; each point in **B** and **E** represents one DKT parcel and one subject, respectively; the sample sizes of each
5 subgroup are shown by numbers on the bars of the bar plots hereafter.
6

1



2

3

4

5 **Fig. 5. Brain propagation dynamics involved with higher-order cortices are linked to tau, coupling**

6 **metrics, and cognitive measures.** (A-B) Propagations between lower-order sensory-motor (SM) and higher-

7 order association cortex were identified near gBOLD peaks as previously described (47). The two group-mean

8 time-position graphs are shown as two types of continuous titled bands reflecting brain activation gradually

9 transiting from low-to-high (L-H) order or in the opposite direction, which are shown as spatial pattern changes

10 in the right seven columns. (C-E) The frequency of all identified propagation events (sum of the propagation

11 frequency of both low-to-high and high-to-low (H-L); extracted within an equal length of fMRI time-series for

12 each subject; adjusted for age and sex) decreased with the increase of tau deposition in higher-order regions,

13 weaker gBOLD-CSF coupling, and the MoCA decline, across all subjects (all $p < 0.040$). This propagation-

14 coupling relationship was also strong for $A\beta+$ and impaired $A\beta+$ subjects (Fig. S9; both $r < -0.26$, both $p <$

15 0.068), similar to Fig. 1 and 2. (F) An overview summarizes the links among coupling metrics, dynamic

Feng Han et al.

1 propagation, and AD markers including thickness, tau, and MoCA. It shows that the glymphatic function-related
2 gBOLD-CSF coupling and dynamic propagating waves modulate tau deposition, cortical thickness, and the
3 cognitive changes, while the propagation frequency and thickness (also in higher-order association regions) was
4 not significantly correlated ($r = 0.10$, $p = 0.27$). Each point in scatter plots represents one subject.
5

1

2 **Table 1. Participant characteristics**

N=115	Aβ⁻ (N = 66)	Aβ⁺ (N = 49)		P-value		
Age	70.0 (7.6)	75.8 (6.9)		5.5×10⁻⁵		
Sex (M/F)	32/34	23/26		1.0		
Diagnosis (AD:MCI:SMC: Control)	0:20:1:45	6:22:4:17		-		
MoCA	25.3 (2.9) (3 N/A)	22.3 (4.8) (1 N/A)		1.0×10⁻⁴		
	Stage1 (S1): Unimpaired Aβ⁻ (N = 46)	Stage2 (S2): Unimpaired Aβ⁺ (N = 21)	Stage3 (S3): Impaired Aβ⁺ (N = 28)	P-value		
				S2 vs S1	S3 vs S1	S3 vs S2
Age	69.4 (7.8)	76.7 (5.5)	75.1 (7.9)	2.9×10⁻⁴	3.7×10⁻³	0.42
Sex (M/F)	17/29	6/15	17/11	0.59	0.057	0.042
Diagnosis	0:0:1:45	0:0:4:17	6:22:1:45	-	-	-
MoCA	25.1 (2.8) (2 N/A)	25.5 (2.7)	19.8 (4.7) (1 N/A)	0.60	7.2×10⁻⁸	8.0×10⁻⁶

3

4 Data represent the “mean (standard deviation)” unless otherwise indicated. Two-sample t-test was applied to
5 compare the continuous measures, while a Fisher exact test between groups was used for the sex ratio.

6 Aβ⁺: cortical AV45–Aβ > 1.11 SUVR or cortical FBB–Aβ > 1.08 SUVR; SUVR: standardized uptake value
7 ratio referring to whole cerebellum reference region; M/F: male/female; AD, Alzheimer’s disease group; MCI:
8 mild cognitive impairment; SMC: subjective memory concern; Unimpaired diagnosis group: SMC and Control
9 subjects; Impaired group: AD and MCI subjects.

10

Feng Han et al.

1 **List of Supplementary Materials**

2 Fig. S1 to S12 for multiple supplementary figures

3 References (37, 38, 68)

4